



HAL
open science

Transcriptional control of physiological and pathological processes by the nuclear receptor PPAR beta/delta

Nguan Soon Tan, Manuel Vázquez-Carrera, Alexandra Montagner, Ming Keat Sng, Hervé Guillou, Walter Wahli

► **To cite this version:**

Nguan Soon Tan, Manuel Vázquez-Carrera, Alexandra Montagner, Ming Keat Sng, Hervé Guillou, et al.. Transcriptional control of physiological and pathological processes by the nuclear receptor PPAR beta/delta. *Progress in Lipid Research*, 2016, 64, pp.98-122. 10.1016/j.plipres.2016.09.001 . hal-01594607

HAL Id: hal-01594607

<https://hal.science/hal-01594607v1>

Submitted on 26 Sep 2017

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Accepted Manuscript

Transcriptional control of physiological and pathological processes by the nuclear receptor PPAR β/δ

Nguan Soon Tan, Manuel Vázquez-Carrera, Alexandra Montagner, Ming Keat Sng, Hervé Guillou, Walter Wahli



PII: S0163-7827(16)30013-3
DOI: doi: [10.1016/j.plipres.2016.09.001](https://doi.org/10.1016/j.plipres.2016.09.001)
Reference: JPLR 925

To appear in:

Received date: 1 April 2016
Revised date: 31 August 2016
Accepted date: 20 September 2016

Please cite this article as: Tan Nguan Soon, Vázquez-Carrera Manuel, Montagner Alexandra, Sng Ming Keat, Guillou Hervé, Wahli Walter, Transcriptional control of physiological and pathological processes by the nuclear receptor PPAR β/δ , (2016), doi: [10.1016/j.plipres.2016.09.001](https://doi.org/10.1016/j.plipres.2016.09.001)

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Comment citer ce document :

Tan, N. S. (Auteur de correspondance), Vázquez-Carrera, M., Montagner, A., Sng, M. K., Guillou, H., Wahli, W. (2016). Transcriptional control of physiological and pathological processes by the nuclear receptor PPAR beta/delta. *Progress in Lipid Research*, 64, 98-122. DOI : [10.1016/j.plipres.2016.09.001](https://doi.org/10.1016/j.plipres.2016.09.001)

Transcriptional control of physiological and pathological processes by the nuclear receptor
PPAR β/δ

Nguan Soon Tan^{1,2,3,4}, Manuel Vázquez-Carrera⁵, Alexandra Montagner⁶, Ming Keat Sng^{1,2},
Hervé Guillou⁶, Walter Wahli^{2,6,7}

¹School of Biological Sciences, Nanyang Technological University, 60 Nanyang Drive,
Singapore 637551.

²Lee Kong Chian School of Medicine, Nanyang Technological University, The Academia,
20 College Road, Singapore 169856.

³Institute of Molecular and Cell Biology, 61 Biopolis Drive, Proteos, Agency for Science
Technology & Research, Singapore 138673.

⁴KK Research Centre, KK Women's and Children's Hospital, 100 Bukit Timah Road, Singapore
229899.

⁵Department of Pharmacology and Therapeutic Chemistry, Faculty of Pharmacy, University of
Barcelona, Institute of Biomedicine of the University of Barcelona (IBUB), Pediatric Research
Institute-Hospital Sant Joan de Déu, and Spanish Biomedical Research Centre in Diabetes and
Associated Metabolic Diseases (CIBERDEM), Barcelona, Spain.

⁶INRA ToxAlim, UMR1331, Chemin de Tournefeuille, Toulouse Cedex 3, France.

⁷Center for Integrative Genomics, University of Lausanne, Le Genopode, CH 1015 Lausanne,
Switzerland.

Correspondence to WW or NST:

Email: walter.wahli@ntu.edu.sg or nstan@ntu.edu.sg

Tel: +65 6576 7336 (WW)

Tel: +65 6316 2941(NST)

Contents

1. Introduction
 2. Molecular mechanisms of transcriptional regulation by PPAR β/δ
 - 2.1 Cellular localization
 - 2.2 Transactivation and repression
 - 2.3 Transrepression
 3. PPAR β/δ in early fetal development
 4. PPAR β/δ in gametes and preimplantation embryos and in placentation
 5. PPAR β/δ in tissue repair
 - 5.1 Inflammatory response during tissue repair
 - 5.2 Skin repair
 - 5.3 Tissue repair in other organs
 6. PPAR β/δ in cardiovascular disease
 - 6.1 Endothelial dysfunction
 - 6.2 Atherosclerosis
 - 6.3 Cardiac disease
 7. PPAR β/δ in insulin resistance and T2DM
 - 7.1 Adipose tissue
 - 7.2 Skeletal muscle
 - 7.3 Liver
 - 7.4 Pancreas
 8. PPAR β/δ and cancer
 - 8.1 Function in the development of colorectal carcinoma (CRC)
 - 8.2 Function in the development of skin carcinoma
 9. PPAR β/δ in neurological diseases
 - 9.1 Neuroprotective role in neurological injuries
 - 9.2 Function in neurological diseases
 10. Conclusion
- Acknowledgements
- References

1. Introduction

Peroxisome proliferator-activated receptors (PPARs) are a class of transcription factors that belong to the nuclear hormone receptor superfamily, and their activities are dependent on their respective ligands. Since the discovery of PPAR α (NR1C1) as a receptor that mediates peroxisome proliferation in rodent hepatocytes in 1990 [1], two other isotypes, PPAR β/δ (NR1C2) and PPAR γ (NR1C3), were subsequently identified and characterized [2,3]. Members of the PPAR family have been extensively linked to numerous systemic and cellular activities that range far beyond simply mediating peroxisome proliferation in rodents [4]. Dependent on isotype-specific or shared tissue expression, all three PPARs regulate different or, in some cases, overlapping functions [5]. PPAR α and PPAR γ have established roles in fatty acid (FA) catabolism and in adipocyte differentiation and lipid storage, respectively, and both are pharmacological targets of FDA-approved drugs for the treatment of numerous metabolic diseases [6,7]. In contrast, no PPAR β/δ ligands are currently used in the treatment of any disease, although small studies on human subjects have used PPAR β/δ ligands to treat metabolic syndrome [8,9]. PPAR β/δ is expressed in numerous tissues [10,11], and many important functions have been attributed to PPAR β/δ in skeletal muscle, adipose tissue, the cardiovascular system, uterine implantation, the gut, the brain, and skin [12]. These factors make PPAR β/δ a very attractive and challenging target, as it is involved in numerous key functions, such as energy metabolism, cellular differentiation and proliferation, tissue repair, and cancer progression.

As is the case for all other members of the nuclear receptor superfamily, PPAR β/δ is structurally organized into functional modular domains. The DNA- and ligand-binding domains (DBD and LBD, respectively) are the most conserved regions among all members of the nuclear receptor superfamily. The DBD consists of α -helical DNA-binding motifs together with two zinc

finger-like motifs. The structure of the DBD allows PPAR β/δ , as a heterodimer with a retinoid X receptor (RXR), to bind to peroxisome proliferator response elements (PPREs) in the regulatory domains of PPAR β/δ target genes. The LBD is composed of 13 α -helices and a 4-stranded β -sheet. As a member of the PPAR family, PPAR β/δ also possesses a relatively large ligand-binding pocket ($\sim 1300 \text{ \AA}^3$), which allows PPAR β/δ to interact with a broad spectrum of ligands, including natural FAs, eicosanoids, and synthetic agonists or antagonists. In addition to the presence of a ligand, transcriptional activity is dependent on and regulated by a tight balance between the levels of binding with coactivators and corepressors. Ligands are able to allosterically alter the conformation of the helical structure (helix 12) at the C-terminal end of the LBD of PPAR β/δ . When no ligand is present, helix 12 exists in its open conformation, which allows for corepressor molecules to bind and repress its transcriptional activities. Ligand binding induces a conformational change at helix 12, exposing a binding domain that allows for coactivator molecules to bind to the LBD of PPAR β/δ and consequently promote the transcription of downstream target genes [13].

In this review, we will discuss studies related to the functions of the less-well known PPAR, PPAR β/δ as well as the status of this nuclear receptor as a potential therapeutic target for many of the metabolic diseases caused by the dysregulation of PPAR β/δ expression and activity.

2. Molecular mechanisms of transcriptional regulation by PPAR β/δ

2.1 Cellular localization

As a nuclear receptor, PPAR β/δ is localized in the nucleus of cells. For ligands to bind to PPAR β/δ , they must be transported from the cytoplasm into the nucleus [14]. The intracellular lipid-binding protein FA-binding protein 5 (FABP5) selectively performs this transport function

to enhance the transcriptional activity of PPAR β/δ [15] (Figure 1). Since the characterization of FABP5 in keratinocytes nearly two decades ago, numerous studies have demonstrated the expression of FABP5 in many tissues and organs, such as the epidermis, adipose tissue, mammary glands, brain, kidneys, liver, lungs, heart, skeletal muscles, and testes, as well as in specific cell types such as macrophages [16]. This expression pattern of FABP5 parallels that of PPAR β/δ in many of these tissues, and the interaction of FABP5 with PPAR β/δ is thought to affect many functions of PPAR β/δ , including those involving cellular glucose and lipid homeostasis, cell differentiation, and apoptotic resistance [15,17,18]. Indeed, cooperative activities between FABP5 and PPAR β/δ have been reported to be involved in neurogenesis [19] and in pathological conditions such as cancer [20,21], metabolic syndrome [22,23], and atherosclerosis [24].

2.2 Transactivation and repression

The transactivation of PPAR β/δ involves the binding of an agonist to the LBD of PPAR β/δ monomers or heterodimers with RXRs and the recruitment of coactivator molecules, such as CBP/p300 and other histone acetyltransferases [25-27] (Figure 1). However, in contrast to PPAR α and PPAR γ , PPAR β/δ functions as a transcriptional repressor in its unliganded state (Figure 1). This function of PPAR β/δ is attributed to two interrelated properties: unliganded PPAR β/δ is able to repress basal transcription as well as PPAR α - and PPAR γ -mediated transcription [28]. In that report, the authors demonstrated that unliganded PPAR β/δ suppressed basal transcription via the recruitment of corepressor molecules. They also demonstrated that PPAR β/δ induced isotype-specific repression of PPAR α and PPAR γ target genes through competition for the PPRE sites of those genes. It has also previously been shown that PPAR β/δ -

mediated repression involves potent corepressor molecules including nuclear receptor corepressor (NCoR) and silencing mediator for retinoid and thyroid hormone receptor (SMRT) [29,30], which can fine-tune receptor activity according to the expression levels of corepressors and coactivators [25,27,31,32] (Figure 1). Interestingly, PPAR β/δ alone is also regulated by another transcription factor, NF- κ B. It has been shown that the p65/RelA subunit of NF- κ B is able to exert a potent repressive effect on the PPAR β/δ -mediated transactivation of target genes in human keratinocytes and that this repression occurs only on PPAR β/δ bound to a PPRE via the recruitment of histone deacetylases (Figure 1) [26].

2.3 *Transrepression*

In addition to regulating cellular functions via the direct induction of target genes, PPAR β/δ is able to perform certain activities in concert with the PPAR β/δ -associated transcriptional repressor B-cell lymphoma-6 (BCL-6) (Figure 1). PPAR β/δ is expressed in vascular smooth muscle cells and endothelial cells together with BCL-6 [28,33], and in these cells, PPAR β/δ plays critical roles in modulating the survival and proliferation of endothelial cells [29,34,35] (Figure 1). In the latter process, the anti-inflammatory activity of PPAR β/δ is triggered by interaction with its ligands, which causes the dissociation of BCL-6 from PPAR β/δ . This dissociation allows BCL-6 to relocate to the promoter regions of pro-inflammatory genes such as VCAM-1 and E-selectin to repress their transcription, thereby protecting the vasculature [35,36]. Such anti-inflammatory effects of PPAR β/δ activation have also been reported for the BCL-6-mediated suppression of monocyte chemoattractant protein-1 and osteopontin, exerting anti-atherosclerotic effects, in angiotensin II-accelerated atherosclerosis [37,38] and in diabetic

nephropathy [39]. In addition, unliganded PPAR β/δ has been found to be associated with BCL-6 in macrophages, where it may play a role in regulating chemokine and cytokine release [2].

2.4 Natural and synthetic ligands of PPAR β/δ

Over the years, many independent groups have reported various findings related to PPAR β/δ and its association with different diseases. Across these studies, controversies regarding the role of PPAR β/δ in certain diseases have been reported, and many of these discrepancies are likely due to differences in the experimental conditions. Despite these controversies, the development of synthetic ligands of PPAR β/δ , including both agonists and antagonists, has greatly contributed to meeting the goal of fully understanding the structure and functions of PPAR β/δ . In this review, we will discuss studies related to less well-known functions of PPAR β/δ as well as the status of this nuclear receptor as a potential therapeutic target for many of the metabolic diseases caused by the dysregulation of its expression and activity.

As a member of the PPAR family, PPAR β/δ is responsive to naturally occurring free FAs and their metabolites, such as monounsaturated and polyunsaturated FAs, carbaprostacyclin, and components of very-low-density lipoprotein (VLDL). Arachidonic acid metabolites, or eicosanoids, are also natural ligands that can activate PPAR β/δ [40–42]. One such example is the lipoxygenation product 8(*S*) hydroxyeicosatetraenoic acid [8(*S*)-HETE] [43,44]. The peroxidation products of polyunsaturated FAs 4-hydroxynonenal (4-HNE) and 4-hydroxydodecadienal (4-HDDE) were reported to activate PPAR β/δ in cultured endothelial cells and pancreatic β -cells [45,46]. However, the mechanism by which these two 4-hydroxyalkenals activate PPAR β/δ remains unclear. It has been proposed that these chemically reactive aldehydes may form covalent bonds with nucleophilic groups of certain amino acids (i.e. histidine, lysine,

arginine or cysteine). However, it has also been reported that steric hindrance prevented covalent interaction of 4-HNE with histidine residues in the ligand binding domain of PPAR β/δ [47]. Very interestingly, it was also previously reported that the vitamin A metabolite all-*trans*-retinoic acid (RA) binds to and activates PPAR β/δ with nanomolar affinity [17,48,49]. As RA is a known ligand for the retinoic acid receptors (RARs), it was uncertain how RA could play a dual, simultaneous role in inhibiting and promoting cellular growth. The same group subsequently reported that the activation of RARs and PPAR β/δ by RA is regulated by a partitioning of RA itself between the two receptors by the intracellular FA transporters CRABP2 (also called CRABP-II) and FABP5, respectively. It has been demonstrated that FABP5 and CRABP2 translocate to the nucleus in response to PPAR β/δ and RAR agonists, respectively, and that these transporters directly bind to their corresponding receptors, enabling direct transfer of their ligands to their cognate receptors [15,17,50–52]. The structural features that allow these proteins to translocate to the nucleus only upon binding of agonists to their cognate receptors and not upon association with other ligands have been identified [51,53]. Moreover, it was demonstrated that the FABP5/CRABP2 expression ratio is critical to the efficient activation of PPAR β/δ and RARs in cultured cells and *in vivo* [15,17,49,54,55]. RAR activation by RA occurs in cells with a low FABP5:CRABP2 ratio, resulting in a pro-apoptotic effect. On the other hand, PPAR β/δ activation by RA was found to promote the survival of cells with higher FABP5 expression [17]. Thus, the relative expression levels of FABP5 and CRABP2 under different physiological circumstances have important functional consequences. For example, numerous reports have documented that the FABP5:CRABP2 ratio is increased in many human tumors as a result of upregulation of FABP5, silencing of CRABP2, or both. These tumor types include glioblastoma, astrocytic gliomas, oral and head-and-neck squamous cell carcinomas, and cancers

of the pancreas, bladder, prostate, breast, skin, colon, and esophagus [56,57]. High levels of FABP5 in tumors are associated with poor patient prognosis in various cancer types (Oncomine, Compendia Bioscience database) [58,59]. Moreover, it has been documented that elevated levels of FABP5 promote cancer cell proliferation, migration, and invasion and tumor development and that in line with the role of FABP5 in enabling the activation of PPAR β/δ , FABP5 exerts these oncogenic effects by enhancing the transcriptional activity of PPAR β/δ , thereby inducing the expression of oncogenic PPAR β/δ target genes [17,20,49,60–62]. However, Rieck et al. (2008) argued against this proposition of PPAR β/δ activation by RA based on a comparative analysis between RA and other PPAR β/δ selective agonists. In their study, RA failed to activate PPAR β/δ , as shown by their reporter assays, and to recruit coactivators to bind to PPAR β/δ [63]. Hence, it is also possible that the effects of RA observed by previous studies may involve a currently undiscovered indirect mechanism. Interestingly, PPAR β/δ and FABP5 are downregulated in adipose tissue of obese subjects [64] but are upregulated in neural stem/progenitor cells during postnatal hippocampal neurogenesis [65]. In addition to the desire to identify naturally occurring ligands, the need to discover therapeutic agents for metabolic diseases resulted in the development of highly specific synthetic PPAR β/δ ligands as potential hypolipidemic drugs. These ligands include agonists and antagonists; the potential benefits of these ligands in the healthcare setting have previously been reviewed and will not be discussed further here [66]. Although none of these compounds have been approved for clinical use, studies of these ligands demonstrated the central role of PPAR β/δ in metabolism. Suitable drug therapies targeting PPAR β/δ could provide multiple benefits against a diverse range of metabolic disorders.

Interestingly, a nucleocytoplasmic shuttling mechanism between FABP4 and PPAR γ similar to that between FABP5 and PPAR β/δ has been proposed [67,68]. Despite numerous

studies, many questions remain to be answered. Using its relatively large ligand-binding pocket, PPAR β/δ interacts with a broad spectrum of ligands, including a variety of natural FAs, eicosanoids, and synthetic agonists or antagonists. However, whether intracellular FA transporters accommodate and shuttle these various ligands to PPAR β/δ remains to be determined, and the selectivity of the various FABPs, such as FABP4 and FABP5, for particular PPAR agonists is uncertain. It is of particular interest that this shuttling mechanism has never been demonstrated for antagonist- or inverse agonist-mediated inhibition of PPAR β/δ . Interestingly, an antagonist of one nuclear receptor family member can function as an agonist of another. For example, Ro 41-5253 is a potent antagonist of RAR α but is able to activate PPAR γ [69]. In this context, the interplay among the various effectors implicated in the selective partitioning of agonists or antagonists of different nuclear receptors further underscores the importance of nucleocytoplasmic shuttling.

3. PPAR β/δ in early fetal development.

In adults, PPAR β/δ regulates cellular processes that are also key for development [70]. PPAR β/δ is highly expressed in early-stage *Xenopus laevis* embryos, in which PPAR α is weakly expressed and PPAR γ is not expressed at detectable levels [2,39]. The PPAR β/δ protein is present in all cell nuclei of the embryo and is strongly upregulated during gastrulation, which suggests a prominent function of this receptor at this developmental stage [38]. In support of this hypothesis, a PPAR β/δ loss-of-function experiment using an antisense morpholino (MO) demonstrated disrupted gastrulation movement. These results are supported by reduced expression of marker genes reflective of defects in muscle (*actc1*, *myod1*) and brain (*krox20*) differentiation at the neurula stage in the absence of PPAR β/δ . By combining the loss-of-

function approach with high-throughput RNA-seq transcript expression analyses and chromatin immunoprecipitation experiments, an important cooperation between epigenetic marks and PPAR β/δ was unveiled. During gastrulation, PPAR β/δ recognizes H3K27me3 marks previously deposited at the pluripotent stage to activate the transcription of early differentiation genes. Importantly, PPAR β/δ is the first identified transcription factor to drive an epigenetic signature of pluripotency *in vivo* during embryonic development [70]. These findings open the door for the development of a greater mechanistic understanding of how the activation of many genes might be coordinated during early development. In mammals, PPAR β/δ also plays a major role later in development during the formation of the placenta, in which PPAR γ is also involved [71–73].

Despite its discovery as early as 1992 and the numerous studies underscoring the important roles of PPAR β/δ in the transcriptional control of energy homeostasis and various pathological processes, surprisingly little is known about its expression during human fetal development. The mRNA and protein expression of PPAR β/δ in human various fetal tissues was examined by a few groups [74,75]. Based on an analysis of nine human fetal tissues, specifically intestinal, liver, lung, heart, kidney, adrenal, thymus, stomach and spleen tissues, ranging in age from embryonic day (ED) 54 to 125, PPAR β/δ mRNA expression in fetal lung, adrenal, stomach (ED54–120), kidney (ED54–125), spleen (ED67–125) and thymus tissues (ED74–120) did not change with age, as determined by real-time PCR using β 2-microglobulin as the internal control gene [75]. Human fetal liver expressed PPAR β/δ mRNA from ED54–125, and its expression increased significantly across the age range; in contrast, its expression decreased in fetal heart (between ED54 and 125) and intestinal tissues (ED54–120). Except for adrenal tissue, in which PPAR β/δ protein expression decreased with fetal age, no tissue showed a significant change in the PPAR β/δ protein level as determined by western blot analysis. The spatiotemporal

distribution of PPAR β/δ during development (from 7 to 22 weeks of gestation, WD; ~ED49-154) of the human fetal digestive tract was further examined via immunohistochemistry using specific polyclonal antibodies [74]. PPAR β/δ was detected throughout the gastric epithelium at all the developmental stages examined. PPAR β/δ expression in the cytoplasm and nucleus of human esophageal epithelial cells was detectable between 7WD and 14WD, peaked at 15WD, and then markedly decreased at 20WD. At later stages, PPAR β/δ staining was more restricted to nuclei. In the small intestine, stronger PPAR β/δ staining was observed in the ileum than in the jejunum at all stages studied. At 22WD, the PPAR β/δ staining signal was stronger in the ileal crypt epithelial cells than in the differentiated villous cells. PPAR β/δ was expressed in the different layers of the human fetal colon at 8-20WD. The PPAR β/δ signal was primarily localized to the nuclei in epithelial and mesenchymal cells. Compared to PPAR β/δ staining in the ileum, stronger PPAR β/δ staining was observed in the crypt regions of the colon. However, PPAR β/δ staining was barely detected in these regions at 14WD [74]. The distinct expression profiles of PPAR β/δ in various human fetal tissues and regions of the fetal digestive tract suggest that PPAR β/δ may play major roles in tissue morphogenesis, development and/or physiology.

4. PPAR β/δ in gametes and preimplantation embryos and in placentation

All three PPAR isotypes are expressed in somatic and germ cells of the testis. In rats, PPAR β/δ is expressed in Leydig cells and Sertoli cells [76]. In mice, PPAR β/δ is expressed in spermatids and spermatocytes [10]. The expression of PPAR β/δ in mouse spermatids and spermatocytes is further supported by the observed expression of *Ssm*, a novel PPAR target gene, in mouse testes [77]. Preimplantation mouse embryos show PPAR β/δ expression, which is already detectable by immunohistochemistry at the two-cell stage [78] or the eight-cell stage [79]

and remains detectable throughout the preimplantation period. Mouse blastocysts also express PPAR β/δ in the inner cell mass and the trophoctoderm [79,80]. These observations suggest that PPAR β/δ plays an important role in early development.

To understand the functions of PPAR β/δ in an *in vivo* and physiologically relevant context in rodents, three independent groups have used different strategies to generate PPAR β/δ knockout mice. The observations from these groups reveal an apparent discrepancy in the role of PPAR β/δ in placentation. In 2000, Peters et al. disrupted the mouse PPAR β/δ gene by inserting a 1.14-kb phosphoribosyltransferase II gene into the last exon in the same direction of transcription in the genomic clone, herein referred to as PPAR $\beta/\delta^{\text{ex8}}$. This insertion truncated only the C-terminal 60 amino acids of PPAR β/δ [81]. Northern blot analysis of RNA from brain, adipose and liver tissues revealed an mRNA transcript 1 kb larger than the wild-type mRNA in the PPAR $\beta/\delta^{\text{ex8}}$ mouse, and the expression of this mutant transcript was lower than that of the wild-type transcript in non-mutant mice. Neither the larger nor a truncated mRNA was detected in the skin of PPAR $\beta/\delta^{\text{ex8}}$ mice, and this result was explained by a later study that revealed that PPAR β/δ was undetectable in adult mouse skin cells [82]. However, despite the presence of these mRNA species, the PPAR β/δ protein was not detected in hepatic nuclear extracts from PPAR $\beta/\delta^{\text{ex8}}$ mice. The breeding of heterozygous offspring of mixed genetic backgrounds (C57BL/6N x Sv/129) produced fewer homozygous PPAR $\beta/\delta^{\text{ex8}}$ mice than expected. An analysis of E10 embryos and E18 fetuses revealed that the absence of PPAR β/δ was not lethal during intrauterine development, and the gross morphology of the concepti appeared normal [81]. Subsequent backcrosses produced PPAR $\beta/\delta^{\text{ex8}}$ mice, ~75% of which were in the C57BL/6N genetic background. The colony of PPAR $\beta/\delta^{\text{ex8}}$ mice reproduced successfully, and normal Mendelian genotype distributions were found in subsequent heterozygous matings.

Barak et al. (2002) used a Cre/lox-mediated recombination strategy to delete exon 4 of the PPAR β/δ gene, which harbors the N-terminal half of the DBD, herein referred to as PPAR $\beta/\delta^{\text{ex4}}$ [72]. In contrast to the approach by Peters et al. (2000), this disruption strategy eliminated nearly the entire PPAR β/δ gene product. PPAR $\beta/\delta^{\text{ex4}}$ pups were rarely obtained from heterozygous crosses, suggesting that the homozygous loss of PPAR β/δ frequently resulted in lethality to embryos, which appeared to occur at E10.5. In contrast to PPAR $\beta/\delta^{\text{ex8}}$ mice, PPAR $\beta/\delta^{\text{ex4}}$ embryos showed no increase in survival rate despite the performance of up to six backcrosses of the original knockout strain (129sv/Jae) with isogenic C57BL/6J breeders. This result indicates that the lethality of PPAR $\beta/\delta^{\text{ex4}}$ is not alleviated by either a heritable component or a specific genetic background [72,81]. Surviving PPAR β/δ -deficient progeny showed markedly stunted growth at term, and most PPAR $\beta/\delta^{\text{ex4}}$ pups were smaller than their wild-type counterparts. Histological examination of PPAR $\beta/\delta^{\text{ex4}}$ concepti at E9.5 revealed abnormally loose placental-decidual connections. By E12.5, three of four PPAR $\beta/\delta^{\text{ex4}}$ embryos surviving the major E10.5 lethality point exhibited extensive maternal hemorrhages into the labyrinthine zone.

Nadra et al. (2006) confirmed that deletion of the PPAR β/δ gene results in a severe failure of the placenta to undergo proper morphogenesis, leading to embryonic lethality between E9.5 and E10.5 [73]. This independent group used a homologous recombination strategy to delete exon 4 and most of exon 5 from the PPAR β/δ gene (i.e., PPAR $\beta/\delta^{\text{ex4}}$). Heterozygote mating (mixed background, C57BL6/Sv129) did not produce the expected Mendelian ratio of mutant animals, and live PPAR $\beta/\delta^{\text{ex4}}$ offspring were rare, although no instances of perinatal death were observed. Further investigation revealed that the trophoblast giant cell layer of the placenta was the most severely affected by this mutation. The authors showed a direct role of PPAR β/δ in promoting trophoblast cell differentiation towards giant cells. This function of

PPAR β/δ depended on phosphatidylinositol 3-kinase (PI3K) and Akt1 (also referred to as protein kinase B) and was at least partly associated with high expression levels of two kinases involved in Akt activation, specifically 3-phosphoinositide-dependent kinase 1 (PDK1; also referred to as PDPK1) and integrin-linked kinase (ILK). In addition, PPAR β/δ caused an increase in the expression of I-mfa, a non-helix-loop-helix (HLH) inhibitor of the myogenic basic HLH (bHLH) subfamily that is involved in the differentiation process. Maternal PPAR β/δ is critical to implantation and decidualization, and embryonic PPAR β/δ is crucial for placentation. Moreover, Wang et al. (2007) showed a reciprocal relationship between the PPAR β/δ -AKT and leukemia inhibitory factor- signal transducer and activator of transcription 3 (STAT3) signaling pathways, which serve as cell lineage sensors to direct trophoblast cell fates during placentation. This finding of stage-specific integration of maternal and embryonic PPAR β/δ signaling provides further evidence that PPAR β/δ is a molecular link that coordinates implantation, decidualization, and placentation, all of which are crucial to a successful pregnancy [83].

In fact, PPAR $\beta/\delta^{\text{ex4}}$ embryos can be lost prior to E9.5 at any stage, including ovulation, fertilization, preimplantation, implantation, and postimplantation [80]. PPAR $\beta/\delta^{\text{ex4}}$ ablation adversely affected embryo development during the preimplantation stage and consequently hampered implantation [79]. Compared with wild-type embryos, PPAR $\beta/\delta^{\text{ex4}}$ embryos show developmental delay as early as 48 hours after harvest of two-cell stage embryos. At 96 hours after harvest of two-cell embryos, only 65% of PPAR $\beta/\delta^{\text{ex4}}$ embryos had reached or passed the blastocyst stage, and 28% subsequently underwent hatching or had hatched completely, compared with 100% and 85% of wild-type embryos, respectively [79]. PPAR $\beta/\delta^{\text{ex4}}$ embryos also exhibit decreased embryonic cell proliferation compared with wild-type embryos. Investigators have proposed that activation of PPAR β/δ by an endogenous PPAR ligand, such as

prostaglandin I₂ (also known as prostacyclin) [84] or retinoic acid [17], may be necessary for biological processes in embryos during preimplantation and subsequent stages of development.

Although genetic approaches support a beneficial role of PPAR β/δ in placentation, pharmacological approaches have yielded varying outcomes at different stages of gestation. PPAR β/δ activation by the synthetic agonist L-165041 or iloprost (a stable PGI₂ analogue) enhanced the development and implantation of cultured embryos in a concentration-dependent manner [85–87]. Embryos preconditioned with L-165041 or iloprost showed higher implantation rates when transferred to gestational carriers [79,86]. Administration of the more selective PPAR β/δ agonist GW501516 to pregnant rats resulted in malformations of the endometrium and placenta. Histopathologically, a single oral administration of GW501516 induced cystic degeneration associated with lysis of glycogen cells [88], unexpectedly leading to significant developmental toxicity. These results suggest that a transrepression mechanism may dominate the activity of PPAR β/δ during development.

This role of PPAR β/δ in preimplantation and placentation clearly warrants further investigation. The reason for the stark contrast in the role of PPAR β/δ in placentation between PPAR $\beta/\delta^{\text{ex8}}$ [81] and PPAR $\beta/\delta^{\text{ex4}}$ [72,73] mice remains unclear, and determining the mechanisms underlying these differences would require comparative analysis of these two models of PPAR β/δ disruption in a congenic background. It has been proposed that the different disruption strategies employed may be responsible for these discrepancies. Disruption of the DBD, as performed by Barak et al. (2002) and Nadra et al. (2006), eliminated nearly the entire PPAR β/δ gene product [72,73], whereas the approach employed by Peters et al. (2000) disrupted only the last 60 amino acids of the LBD of PPAR β/δ [81]. In the latter approach, the limited deletion might not completely abolish PPAR β/δ activity and could theoretically produce a hypomorphic

allele that retains some aporeceptor function [88–90]. Importantly, either $\text{PPAR}\beta/\delta^{\text{ex}4}$ or $\text{PPAR}\beta/\delta^{\text{ex}8}$ mice may be applicable to studies of various diseases, such as tumorigenesis and neurological diseases. As elaborated in subsequent sections of this review, opposing roles of $\text{PPAR}\beta/\delta$ have been reported, particularly in relation to tumorigenesis; however, the conclusions of these studies have been roughly concordant when using $\text{PPAR}\beta/\delta^{\text{ex}4}$ mice derived by two independent laboratories [72,73].

5. Tissue repair

We have previously reviewed the involvement of all three PPARs in tissue injury and wound repair [91]. Below, we focus on recent findings related to the specific roles of $\text{PPAR}\beta/\delta$ in the tissue repair process.

5.1 Inflammatory response during tissue repair

A cardinal feature of poorly healing wounds is a persistent inflammatory response at the wound site [92]. This can lead to unbalanced proteolytic activity, which overwhelms the protective mechanisms of the local tissue and leads to a dysfunctional wound microenvironment that cannot support healing. PPARs are central effectors in the induction and modulation of several inflammatory reactions. These inflammatory processes serve purposes that range from protection against external microbial infections to promotion of pathological inflammation-related disorders such as atherosclerosis and type 2 diabetes [93]. The expression of $\text{PPAR}\alpha$ is transiently elevated after adult mouse injury and has been implicated in the promotion of anti-inflammatory effects [82,94]. In comparison, $\text{PPAR}\gamma$ activation leads to reduced production of pro-inflammatory cytokines, including $\text{TNF}\alpha$, IL-6 and nitric oxide (NO) [95]. In contrast, the role of $\text{PPAR}\beta/\delta$ in the modulation of inflammation during wound repair is poorly understood.

However, one current model exists; this model suggests that the role of PPAR β/δ involves the switching of macrophage functions from pro- to anti-inflammatory. This switching mechanism involves the ligand-mediated activation of PPAR β/δ and the subsequent promotion of the release of BCL-6 to repress the transcriptional activities of pro-inflammatory genes (Figure 1). In agreement with these observations, other reports have suggested that PPAR β/δ agonists are able to downregulate lipopolysaccharide-induced pro-inflammatory genes, including inducible NO synthase (iNOS) and COX2, via NF- κ B [96,97]. Interestingly, PPAR-mediated inflammatory crosstalk has also been implicated in the maintenance of tissue homeostasis, particularly in skin. While PPAR α is important in the early inflammatory phases of wound healing for the recruitment of immune cells to the wound bed, PPAR β/δ plays a crucial role in reducing these inflammatory activities as the wound closes and heals.

5.2 Skin repair

Of all the human tissues in which the role of PPAR β/δ has been studied, skin tissue repair has been particularly well studied in terms of the functions of PPAR β/δ at various stages of tissue repair (Figure 2). Cutaneous wound repair is classified as a high-priority survival response to skin injuries. Upon formation of a cutaneous wound, the primary goal of the body is to reconstruct a new and effective epidermal barrier that covers the wound, thereby preventing tissue dehydration and any opportunistic microbial infections [98,99]. Normal wound healing is composed of three major phases – inflammation, cell proliferation and migration (re-epithelialization), and extracellular matrix (ECM) remodeling. These three phases occur in a partially overlapping pattern via the production of cytokines and growth factors in a temporally and spatially regulated manner [98]. The inflammation phase begins immediately after skin

injury. Damaged blood vessels in the vicinity of the injury release signals that initiate localized blood clotting and influx of inflammatory cells.

PPAR β/δ is the dominant PPAR subtype in the human epidermis. In mice, the expression of PPAR β/δ in the interfollicular epidermis declines as the animal develops from the fetal stage to adulthood. However, upon cutaneous injury, PPAR β/δ expression is rapidly upregulated in the epidermis at the wound edges, and PPAR β/δ -activating ligands are produced [82]. Upon the influx of inflammatory cells during skin injury, the expression levels of pro-inflammatory cytokines, such as TNF α , have been found to be associated with *Ppar* β/δ gene stimulation via the SAPK/AP-1 pathway. Consequently, induction of PPAR β/δ expression leads to increased production of ILK and PDK1 (or PDPK1), both of which have been reported to be direct targets of PPAR β/δ -mediated activation. In addition, PPAR β/δ activation coordinates the downregulation of the expression of phosphatase and tensin homologue deleted on chromosome 10 (PTEN). These activities lead to an increase in AKT1 activation, which suppresses keratinocyte apoptosis by modulating both the death-receptor and mitochondrial apoptotic pathways. This process ensures the recruitment of a sufficient quantity of viable keratinocytes at the wound edge for re-epithelialization to occur [82,99]. Furthermore, *in vivo* wound studies have shown that PPAR β/δ activation exerts an anti-apoptotic effect on keratinocytes in wounded tissues [100]. In addition to its anti-apoptotic effects during wound healing, PPAR β/δ upregulates the activity of NF- κ B, which leads to increased matrix metalloproteinase-9 (MMP-9) production [100]. This increased production of MMP-9 was subsequently found to play a role in the regulation of keratinocyte migration during wound healing [101] (Figure 2).

Wound repair involves epithelial-mesenchymal interactions that require communication among different cell types in a temporal manner. The signals that are communicated within the

wounded tissue are mediated by key effectors such as growth factors secreted by cells in the epithelium and mesenchyme. Therefore, in addition to studying the functions of keratinocytes in the process of wound healing, our group has previously identified a novel and critical role of fibroblast-derived PPAR β/δ in modulating skin homeostasis in a paracrine fashion in the adjacent epithelium [102]. Keratinocyte-derived interleukin-1 (IL-1) promotes the upregulation of PPAR β/δ expression in neighboring fibroblasts [102]. Fibroblast-derived PPAR β/δ subsequently stimulates the production of secreted IL-1 receptor antagonist (sIL-1ra), thereby inhibiting further IL-1-induced production of mitogenic factors by fibroblasts. This regulated decrease in mitogenic factor production leads to tight regulation of keratinocyte proliferation during wound healing, thereby ensuring homeostasis in the epidermal layer of the skin [102] (Figure 2). Such control over homeostasis during normal wound healing highlights the importance of tight regulation of the complex epithelial-mesenchymal interactions. When this tight control becomes dysregulated, mechanisms of epithelial-mesenchymal crosstalk may become defective, leading to disorders such as poor wound healing in diabetes patients.

The importance of PPAR β/δ as a potential therapeutic target for promoting proper wound healing, particularly healing of chronic diabetic wounds, has also been recently demonstrated [103]. Diabetic wounds are imbued with early, excessive and protracted reactive oxygen species (ROS) production. Wang et al. (2015) underscored a novel role of PPAR β/δ in fibroblasts in the modulation of oxidative stress levels in a diabetic wound microenvironment. They demonstrated that *catalase* and *glutathione peroxidase 1 (Gpx1)* were novel target genes of PPAR β/δ in fibroblasts. Both catalase and GPx1 are antioxidants that function to scavenge the excessive hydrogen peroxide (H₂O₂) that accumulates in diabetic wound beds. In addition, these authors showed that excessive levels of H₂O₂ modified the ECM, which subsequently impaired

keratinocyte migration, leading to the formation of poorly healed or unhealed wounds in diabetic patients. To demonstrate the therapeutic potential of fibroblast-derived PPAR β/δ for promoting diabetic wound closure, wounds in diabetic mice were treated with a microencapsulated form of the PPAR β/δ agonist GW501516. The microencapsulation approach provided early and sustained release of GW501516 into the wound bed, increasing its bioavailability. When translated clinically, the concept of drug microencapsulation would help maintain patient compliance with treatment. Importantly, diabetic wounds treated with GW501516 healed much more rapidly than vehicle-treated wounds due to reduced levels of H₂O₂ production by fibroblasts in the diabetic wound microenvironment, leading to enhanced keratinocyte migration and, therefore, improved wound closure [103] (Figure 2). The importance of PPAR β/δ in fibroblasts is also well supported by other previous studies. A separate report demonstrated its importance in downregulating the UVB-induced secretion of MMP-2, along with upregulating elastin and decreasing ROS levels. Such processes may contribute to the maintenance of skin integrity [104]. In another study, the expression of PPAR β/δ in fibroblasts was increased upon heat-induced injury, which subsequently initiated processes that led to cellular protection from heat injury-induced morphological changes and to increased keratinocyte proliferation [105]. Another report has shown that activation of PPAR β/δ in fibroblasts induced the upregulation of *Ski* in a rat model, thereby leading to increased fibroblast proliferation and improved wound repair [106,107]. *Ski* encodes the nuclear proto-oncogene protein homologue of the avian sarcoma viral (v-ski) oncogene. Ski functions as a repressor of TGF β signaling [108]. Recently, PPAR β/δ activation was implicated in α -smooth muscle actin (α -SMA) upregulation [109], which was shown to be caused by a dual binding mechanism involving the interaction of activated PPAR β/δ with a direct repeat-1 (DR1) site in the α -SMA promoter and the interaction

of Smad3 with a Smad-binding element (SBE) in another region of the α -SMA promoter. Elevated α -SMA levels correlate with increased fibroblast-to-myofibroblast differentiation, concomitant with enhanced fibroblast migration and contractility. While this report demonstrated the beneficial role of PPAR β/δ in wound healing via the upregulation of α -SMA in fibroblasts, the study was performed in the presence of serum in the culture. The presence of serum could have contributed to non-specific activation of PPAR β/δ in the cultured fibroblasts. Nonetheless, all of these studies suggest that PPAR β/δ plays crucial roles in promoting appropriate cutaneous wound repair.

In addition to their crosstalk with fibroblasts, keratinocytes communicate with endothelial cells during wound repair. In order for the skin to have proper and efficient nourishment, waste removal, inflammatory responses, and temperature regulation, a functional vascular circulation is essential. In contrast, dysfunctional vascular circulation severely impairs the tissue repair process [106,110–112]. This dysfunction is one of the main impediments to the proper healing of diabetic wounds [113]. Previously, it has been shown that angiopoietin-like protein 4 (ANGPTL4) is a direct target of PPAR β/δ [112,114]. The C-terminal form of the post-processed ANGPTL4 protein (cANGPTL4) was shown to weaken cell-cell tight junctions and to induce NO production through the integrin/JAK/STAT3-mediated upregulation of iNOS in the epithelial layer of the wound. This finding reveals a novel mechanism in which ligand-mediated activation of PPAR β/δ participates in upregulating the expression and secretion of keratinocyte-derived cANGPTL4 to promote the angiogenic process via keratinocyte-endothelial cell crosstalk, leading to accelerated wound healing [115,116] (Figure 2).

In addition to cell-cell communication, PPAR β/δ modulates the interactions between cells and the ECM. Similar to its observed activities in normal keratinocytes, the PPAR β/δ target

cANGPTL4 was found to be primarily produced by wound keratinocytes at the wound site [117,118]. As a matricellular protein, cANGPTL4 can associate with the myriad of proteins that are located in the ECM reservoir and, therefore, with various cell surface receptors to act as a possible mediator of paracrine signaling during the wound healing process and as a component of various regulatory networks [114,119]. Wound keratinocyte-derived cANGPTL4 has previously been shown to modulate cell migration by regulating both integrin-mediated signaling and cell-ECM crosstalk via the control of matrix protein integrity. In addition, cANGPTL4 has been shown to interact with integrin $\alpha 1\beta 5$, thereby activating the integrin-FAK-Src-PAK1 signaling pathway and leading to selective integrin internalization to promote cell migration [120,121] (Figure 2). Considering these observations, it is possible that exogenous administration of PPAR β/δ agonists could aid in wound healing via cANGPTL4-mediated regulation of cell-ECM communication.

5.3 Tissue repair in other organs

In addition to the skin, tissues from other organs are also known to undergo repair via processes involving the activation of PPAR β/δ . Corneal stromal fibrosis, which occurs as a result of poor wound healing in the cornea, is characterized by an increased abundance of myofibroblasts and excess ECM deposition. Corneal scarring occurs naturally following trauma, infection, or refractive surgery and could result in loss of vision [122]. One study showed that activation of PPAR β/δ using GW501516 after excimer laser keratectomy provided antifibrotic effects by targeting different phases of wound healing. For example, GW501516 was shown to promote angiogenesis in the early phase of healing, resulting in increased nutrient supply. However, at the beginning of the remodeling phase of wound healing, GW501516 deregulated

the activation and proliferation of keratinocytes and subsequently prevented their transdifferentiation into myofibroblasts, which caused aberrant ECM deposition and, hence, the appearance of corneal haze [122].

PPAR β/δ is also known to play a role in maintaining endothelial homeostasis. In the presence of GW501516 and its co-activator PPAR γ co-activator 1 α (PGC1 α), PPAR β/δ has been shown to induce the production and activity of heme oxygenase-1, which in turn has a vasculoprotective effect against H₂O₂-induced vascular injury by increasing the production of bilirubin and heavy-chain ferritin and promoting the opening of Fe²⁺ export channels [123]. These end products have significant anti-inflammatory, anti-apoptotic, and anti-oxidant effects.

In the liver, chronic alcohol consumption can impair liver regeneration as a result of attenuated insulin signaling and oxidative injury. In the rat liver, PPAR β/δ agonist administration reduces the severity of ethanol-induced injury and attenuates the effects of ethanol on liver repair by restoring insulin responsiveness. Interestingly, this beneficial effect of PPAR β/δ agonist administration persists even when ethanol consumption is continued at a high level [124].

6. PPAR β/δ in cardiovascular disease

Several reports have shown that PPAR β/δ is involved in vascular processes, suggesting its role in cardiovascular disease (CVD) [125].

6.1 Endothelial dysfunction

Perturbation of endothelial function due to conditions such as hypertension or dyslipidemia can provoke vasospasms, thrombosis, intimal growth, inflammation, and plaque rupture, leading to the development and progression of cardiovascular diseases [126]. PPAR β/δ activation may protect blood vessels through direct and indirect effects. In the vascular

endothelium, PPAR β/δ activation directly causes endothelium-dependent vasodilatation, increased eNOS phosphorylation, and elevated NO production in rat aortae through activation of the PI3K/Akt pathway [127] (Figure 3). Indirect effects of PPAR β/δ agonists include the prevention of dyslipidemia and beneficial effects on insulin resistance and glucose homeostasis, which protect the vascular endothelium from well-established risk factors for the initiation and progression of vascular disease. Both the direct and indirect effects of PPAR β/δ agonists might contribute to restoration of endothelial function in streptozotocin-induced animal models of type 1 [127] and type 2 diabetes [128] by increasing NO bioavailability as a result of reduced NADPH oxidase-mediated superoxide production and preserved PI3K/Akt pathway function. In a more recent study [129], PPAR β/δ activation was confirmed to reduce endothelial dysfunction in aortae from mice fed a high-fat diet (HFD), and this reduction was found to be associated with reduced levels of vascular ROS, decreased activity of NADPH, and upregulated expression of antioxidant genes. Lipids cause endothelial dysfunction through an ROS-mediated reduction in NO bioavailability, and PPAR β/δ activation ameliorates FA-induced endothelial dysfunction by upregulating carnitine palmitoyltransferase 1 (CPT-1), which controls an essential step in mitochondrial FA β -oxidation, thus reducing diacylglycerol (DAG) accumulation and subsequent protein kinase C (PKC)-mediated ROS production as well as causing endothelial NO synthase (eNOS) inhibition. In another animal model of spontaneous hypertension in rats, PPAR β/δ activation has been shown to reduce blood pressure, ameliorate endothelial dysfunction, and reduce the vascular pro-inflammatory and pro-atherogenic status of animals [130]. These changes were associated with increased eNOS protein levels, antioxidant gene upregulation, reduced NADPH oxidase-driven superoxide production and increased expression of regulator of G protein-coupled signaling protein (RGS)4 and RGS5. RGSs play important roles in the

regulation of G protein signaling by binding to active G subunits and stimulating GTP hydrolysis, thus switching off G protein signaling [131]. Therefore, RGS upregulation may attenuate the effects of vasoconstrictors such as endothelin and angiotensin II [132]. Similar findings were obtained in a renin-independent hypertension model induced using deoxycorticosterone acetate (DOCA) salt. Upregulation of RGS5 by PPAR β/δ agonists appears to be an essential underlying their antihypertensive and antioxidant properties and their protective effects on endothelial function. Interestingly, the antidiabetic drug metformin has been shown to reverse impaired endothelium-dependent relaxation through the inhibition of endoplasmic reticulum (ER) stress-induced ROS generation, thereby increasing NO availability in the aortae of diet-induced obese wild-type mice but not their diet-induced obese PPAR $\beta/\delta^{\text{ex8}}$ knockout littermates [133]. These findings suggest that the vasoprotective effects of the commonly used drug metformin require PPAR β/δ . Finally, some evidence suggests that PPAR β/δ activation *in vivo* might increase angiogenesis [30].

6.2 Atherosclerosis

Dyslipidemia and macrophage-mediated inflammation are key factors in the development of atherosclerosis. PPAR β/δ may retard the development and progression of atherosclerosis by modulating lipid metabolism and exerting anti-inflammatory effects on macrophages. Regarding dyslipidemia, one study reported that PPAR $\beta/\delta^{\text{ex8}}$ knockout mice exhibited elevated levels of plasma triglycerides (TGs) compared to wild-type littermates [134], while another report showed no differences in plasma lipid profiles between PPAR $\beta/\delta^{\text{ex4}}$ mice and wild-type mice [35,72]. When PPAR $\beta/\delta^{\text{ex8}}$ knockout mice were fed an HFD, they displayed elevated plasma levels of TGs, primarily in association with increased hepatic VLDL production and reduced lipoprotein

lipase-mediated TG catabolism, although these animals showed no change in cholesterol levels [134]. Treatment of animal models of obesity and diabetes with PPAR β/δ ligands has been shown to have beneficial effects on plasma lipoprotein levels, including reductions in plasma VLDL-TG, low-density lipoprotein (LDL)-cholesterol and non-esterified FA (NEFA) levels as well as an increase in high-density lipoprotein (HDL)-cholesterol levels [135,136] (Figure 3). In addition, PPAR β/δ activators can reduce plasma TG levels by increasing hepatic FA oxidation (FAO), which reduces the availability of FAs for VLDL synthesis, via amplification of the PGC-1 α -lipin 1-PPAR α pathway [137]. The PPAR β/δ -mediated modulation of hepatic expression of genes involved in lipoprotein metabolism, including *ApoA4*, *ApoA5*, *ApoC1* and the VLDL receptor (*Vldlr*), may also contribute to the hypotriglyceridemic effect of PPAR β/δ *in vivo* [138]. The increase in plasma HDL-cholesterol levels following PPAR β/δ activation has been linked to an increase in the hepatic expression of *ApoA1* and *ApoA2* [139]. A recent study demonstrated that PPAR β/δ activation also regulates the expression of the hepatic phospholipid transfer protein (*Pltp*), which contributes to the maintenance of HDL levels in plasma and generates pre β -HDL [140]. Regarding its reduction of LDL-cholesterol levels, PPAR β/δ activation decreases intestinal cholesterol absorption, which has been associated with significant downregulation of the Niemann-Pick C1-like gene [141] and with the stimulation of fecal cholesterol excretion, primarily by increasing transintestinal cholesterol efflux [142]. The findings of pre-clinical studies prompted the examination of PPAR β/δ ligands in small-scale clinical trials, primarily for the treatment of atherogenic dyslipidemia. This type of dyslipidemia is an important risk factor for myocardial infarction and cardiovascular disease and is a typical feature of obesity, insulin resistance, metabolic syndrome and type 2 diabetes mellitus (T2DM). Atherogenic dyslipidemia manifests as three characteristic features: increased blood concentrations of small, dense LDL

particles, decreased levels of HDL particles, and increased levels of TGs. In the first relevant study, performed on healthy volunteers, GW501516 administration increased plasma HDL-cholesterol levels and improved postprandial TG clearance [8]. In abdominally obese men with moderate dyslipidemia, GW501516 reduced plasma TG, ApoB, LDL-cholesterol and insulin levels but did not alter HDL-cholesterol levels [9]. Moreover, treatment with this drug reduced liver fat content by 20%. To elucidate the mechanism of action of GW501516, Ooi et al. (2011) studied the effects of this drug on the kinetics of lipoproteins in subjects with central obesity together with dyslipidemia [143]. GW501516 reduced plasma TG, NEFA, ApoB100, and ApoB48 concentrations. Additionally, treatment with this drug increased the hepatic catabolism of VLDL particles, which may have resulted from decreased ApoC3 levels, and increased plasma HDL-cholesterol and ApoA2 levels. In another study, performed on patients with fasting plasma HDL-cholesterol levels below 45 mg/dl (1.16 mmol/L), GW501516 produced significant increases in the level of HDL-cholesterol and reductions in the levels of NEFAs, ApoA1, ApoB and LDL-cholesterol [144]. Furthermore, plasma LDL particle levels decreased as a result of a reduction in the number of small LDL particles, which are more atherogenic, whereas an increase in the number of large LDL particles was observed. Overall, these changes suggest a transition towards lipoprotein profiles that are less atherogenic following GW501516 treatment. The efficacy of the novel PPAR β/δ agonist MBX-8025 has also been evaluated alone and in combination with atorvastatin to treat overweight patients with mixed dyslipidemia. The effects of MBX-8025 monotherapy were very similar to those reported for GW501516 treatment (significant reductions in plasma LDL-cholesterol, ApoB100, TG and NEFA levels). Administration of MBX-8025 in combination with atorvastatin led to a greater reduction in LDL-cholesterol and ApoB levels than MBX-8025 monotherapy, but the combination of MBX-

8025 and atorvastatin was not more effective in reducing LDL-cholesterol or ApoB levels than atorvastatin alone [145,146]. Similarly, the combination of MBX-8025 and atorvastatin caused a greater reduction in TG levels and increase in HDL-cholesterol levels than atorvastatin monotherapy but not MBX-8025 monotherapy [146]. When lipoprotein subfractions were analyzed, MBX-8025 treatment was observed to provoke a shift in LDLs from smaller to larger particles in approximately 90% of the subjects, and this effect of MBX-8025 was greater than that of atorvastatin [146]. MBX-8025 administration also enhanced insulin sensitivity and increased the levels of markers of liver function [146]. Recently, CER-002, a new PPAR β/δ agonist that has been shown to increase HDL-cholesterol levels and halt the progression of atherosclerosis in pre-clinical studies, has been studied in a completed Phase I clinical trial for the treatment of cardiovascular disease (<http://www.cerenis.com/en/>).

In addition to dyslipidemia, maladaptive immune responses driven by the accumulation of cholesterol-laden macrophages in arterial walls, which can result in vascular inflammation, are central to atherosclerosis. PPAR β/δ agonists have been found to reduce the area of atherosclerotic lesions through anti-inflammatory mechanisms in ApoE^{-/-} and LDLR^{-/-} mice [29,147–149]; however, some studies have reported no such reduction in atherosclerotic lesion size [150]. While PPAR β/δ agonists provoke anti-inflammatory effects in macrophages *in vitro*, surprisingly, selective deletion of PPAR β/δ from macrophages attenuates atherosclerosis in LDLR^{-/-} mice [35]. This observation suggests that PPAR β/δ can also have pro-inflammatory effects. The mechanism underlying these contradictory findings involves the interaction between PPAR β/δ and the inflammatory suppressor protein BCL-6 [144] (Figure 1). This interaction explains the apparent contradiction that both PPAR β/δ deficiency and PPAR β/δ activation exert

anti-inflammatory effects in macrophages due to the release of BCL-6 bound to PPAR β/δ [29,35,148].

6.3 Cardiac disease

Genes involved in FAO are transcriptionally controlled by PPAR β/δ . Consequently, the Cre/loxP-mediated cardiomyocyte-restricted deletion of PPAR β/δ in mice (α MyHC-Cre PPAR $\beta/\delta^{\text{ex4}}$) downregulates the constitutive expression of key FAO genes and decreases basal myocardial FAO, resulting in progressive myocardial lipid accumulation. This is accompanied by cardiac dysfunction, cardiac hypertrophy and congestive heart failure, leading to reduced survival [151]. Another study found that PPAR $\beta/\delta^{\text{ex4}}$ knockout in the adult mouse heart caused a reduction in lipid and glucose oxidation and attenuated the cardiac expression of endogenous antioxidants, and these effects were accompanied by increased oxidative damage to the heart [152]. In addition, expression of the key mitochondrial biogenesis gene *Pgc-1 α* was substantially decreased, together with a decrease in the number of mitochondria. Moreover, the PPAR $\beta/\delta^{\text{ex4}}$ conditional knockout mice showed a decrease in cardiac performance and developed cardiac hypertrophy. The constitutive cardiomyocyte-specific expression of PPAR β/δ (driven by the myosin heavy chain promoter in MHC-PPAR β/δ mice) has been shown to increase the expression of FAO-associated genes but was not found to induce lipid accumulation or cardiac dysfunction; this observation contrasted with the findings in MHC-PPAR α mice [153]. Liu et al. (2008, 2011) generated a transgenic mouse model that expressed a constitutively active form of PPAR β/δ upon tamoxifen administration in a tissue-specific manner [154,155]. In this model, the expression of genes involved in FAO, the levels of endogenous antioxidants, the *Pgc-1 α* levels, the mitochondrial DNA copy numbers, and cardiac performance were increased (Figure 3).

Moreover, activation of PPAR β/δ in the adult heart improved cardiac function and impeded the development of pathological characteristics under conditions of pressure overload [154].

Given the important role of PPAR β/δ in cardiac FAO, a reduction in PPAR β/δ expression or activity might contribute to the exacerbation of the pathological processes involved in cardiac dysfunction. It has been reported that myocardial hypoxia, a characteristic of heart failure, provokes an increase in the expression of the miRNA cluster miR-199a~214, which decreases PPAR β/δ protein levels and mitochondrial FAO [156]. These changes facilitate a metabolic shift from a predominant reliance on FAO in the healthy myocardium towards increased reliance on glucose metabolism at the onset of heart failure. Moreover, cardiomyopathy in streptozotocin-induced diabetic rats was found to be associated with a marked decrease in cardiac PPAR β/δ levels [157]. Activation of PPAR β/δ can ameliorate certain cardiac pathologies linked to reductions in FAO. Thus, PPAR β/δ activation in the adult heart improves cardiac performance and reduces fibrosis and mitochondrial abnormalities in mice subjected to pressure-overload-induced cardiac hypertrophy [155]. Moreover, activation of PPAR β/δ protects the heart from ischemia-reperfusion injury in Zucker diabetic fatty rats through several mechanisms, including attenuation of the lipotoxicity caused by ischemia-reperfusion injury by increasing cardiac FAO and upregulating pro-survival signaling in the heart [155]. Similarly, PPAR β/δ activation might alleviate lipid-induced ER stress and inflammation in the heart [158,159]. In contrast, a recent study showed that treatment with the PPAR β/δ agonist GW610742 had only a temporary effect on early fibrosis in the infarcted myocardium and that the functional and structural beneficial effects of GW610742 might not be sustained [160]. Park et al. (2016) showed that PPAR β/δ expression was highest 3 days after myocardial infarction and began to decrease by day 14 [160]. While investigating the potential therapeutic effects of GW610742 on cardiac healing in rats

after myocardial infarction, the authors showed that the degrees of angiogenesis and fibrosis after myocardial infarction were significantly higher in the GW610742-treated rats than in the untreated rats at 1 week following myocardial infarction, although no significant difference in angiogenesis or fibrosis was observed at 2 weeks after myocardial infarction. Notably, GW610742 increased myofibroblast differentiation and TGF β 2 expression in the infarct zone at 7 days after myocardial infarction. GW610742 also increased bone marrow-derived mesenchymal stem cell recruitment in the whole myocardium and increased serum platelet-derived growth factor B, stromal-derived factor-1 α , and MMP9 levels at day 3 after myocardial infarction [160].

Cardiovascular diseases are a group of disorders of the heart and blood vessels, including events such as hypertension, heart attack, stroke, atherosclerosis, and chronic comorbidities such as kidney disease. It is well established that complications of CVD may develop over a considerable period after onset of the original disease. Most studies of PPAR β/δ in the setting of CVD have focused on the relatively short-term impact of PPAR β/δ ; however, the long-term functions of PPAR β/δ in CVD must also be investigated to determine the therapeutic potential of drugs targeting PPAR β/δ .

7. PPAR β/δ in insulin resistance and T2DM

Over the years, PPAR β/δ has been found to be involved in many processes in insulin-responsive organs and tissues. Its roles in such organs are summarized below.

7.1. Adipose tissue

As adipose tissue expands during obesity, there is an increase in chronic, systemic low-grade inflammation, mainly due to greater macrophage infiltration and polarization towards the

pro-inflammatory M1 phenotype. This chronic inflammatory process is thought to play a major role in the development of insulin resistance and diabetes [161]. PPAR β/δ has been identified as a potential drug target for modulating tissue macrophage activation and insulin sensitivity. Kang et al. (2008) demonstrated in mice that Th2 cytokines secreted by adipocytes, including IL-13 and, to a lesser extent, IL-4, induce PPAR β/δ expression in macrophages through a signal transducer and activator of transcription 6 (STAT6) binding site on the PPAR β/δ promoter, which triggers alternative M2 activation (Figure 4) [162]. Consequently, HFD-fed myeloid-specific PPAR $\beta/\delta^{\text{ex4}}$ knockout mice showed increased levels of M1 markers and decreased levels of markers of the alternative anti-inflammatory M2 phenotype, which is believed to improve insulin sensitivity. Increased adipose tissue inflammation, lipolysis and insulin resistance accompanied these changes.

Moreover, specifically overexpressing PPAR β/δ in white adipose tissue renders mice resistant to both HFD-induced and genetically predisposed obesity, in association with a reduction in triglyceride accumulation in adipocytes and circulating NEFA and TG levels [163]. These effects might be the result of the PPAR β/δ -mediated activation of overall oxidative metabolism in white adipose tissue [164]. In contrast, PPAR $\beta/\delta^{\text{ex4}}$ knockout mice fed an HFD were prone to weight gain [165]. In a different study, the adipose tissue of PPAR $\beta/\delta^{\text{ex4}}$ knockout mice displayed increased STAT3 phosphorylation and elevated suppressor of cytokine signaling (SOCS)3 protein levels than their wild-type littermates [165]. These findings suggest that PPAR β/δ activation might prevent the activation of STAT3 and the subsequent increase in SOCS3 expression, thus inhibiting insulin signaling by preventing the coupling of insulin receptor substrate (IRS)-1 to the insulin receptor and by promoting the proteasome-mediated degradation of IRS-1 [166]. Moreover, adipose tissue inflammation and glucose intolerance were

shown to be exacerbated in fructose-fed PPAR $\beta/\delta^{\text{ex4}}$ knockout mice via nuclear factor E2-related factor 2 (Nrf2), a transcription factor that has been reported to impair insulin signaling [163].

A reduction in dietary salt intake is recommended for many diabetic patients, although the potential benefits of sodium reduction remain debatable. Furthermore, the relationship between sodium intake and glucose homeostasis remains unclear in diabetes. Using adipose-specific PPAR β/δ knockout mice (Fabp4-PPAR $\beta/\delta^{\text{ex4}}$), Zhao et al. (2016) recently showed that a high-salt diet (HSD, high sodium) reduced fasting blood glucose levels in wild-type control mice compared with Fabp4-PPAR $\beta/\delta^{\text{ex4}}$ mice, suggesting that PPAR β/δ in adipocytes improves glucose tolerance and enhances natriuresis [167]. The HSD selectively elevated the expression of PPAR β/δ in perirenal fat by increasing osmotic pressure, leading to an increase in adiponectin production. Adipose-derived adiponectin suppressed the kidney expression of sodium-glucose cotransporter 2 (SGLT2), which plays a crucial role in the regulation of renal glucose transport and sodium reabsorption. Adiponectin reduced the levels of the transcription factors HNF-1 α and SP-1 bound to the promoter of the gene encoding SGLT2 (*SLC5A2*), leading to reduced SGLT2 expression. These findings reveal an interesting mode of communication between adipose tissue and the kidney that links sodium intake to glucose homeostasis. Notably, the expression of PPAR β/δ is lower in morbidly obese patients than in non-obese subjects [64]. Overall, these findings suggest that reduced PPAR β/δ expression in adipose tissue might contribute to metabolic alterations.

7.2 Skeletal muscle

Lipid-induced insulin resistance in skeletal muscle is of paramount interest because myocytes are responsible for approximately 80% of insulin-stimulated glucose uptake and thus

greatly affect the whole-body rate of glucose clearance [168]. In addition, skeletal muscle is an important consumer of FAs, and muscle-specific PPAR β/δ overexpression results in a switch to more oxidative fibers [169], probably through an estrogen-related receptor γ (ERR γ)/miRNA circuit [170]. This alteration is accompanied by an increase in the expression of genes involved in oxidative metabolism, leading to a reduction in body fat mass [169]. Conversely, mice with selective PPAR $\beta/\delta^{\text{ex4}}$ ablation in skeletal muscle myocytes exhibit a shift in skeletal muscle fiber type towards muscle fibers with lower oxidative capacity, leading to obesity and T2DM [171]. The genes involved in FA metabolism that are upregulated by PPAR β/δ *in vivo* include genes involved in cellular FA uptake, such as FAT/CD36; FA activation, such as acyl-CoA synthase (ACS); mitochondrial FA uptake, such as carnitine palmitoyl transferase (CPT); and FA β -oxidation, such as hydroxyacyl-CoA dehydrogenase. PPAR β/δ activation also upregulates the expression of pyruvate dehydrogenase kinase 4 (PDK4), which inhibits the pyruvate dehydrogenase complex, leading to FA oxidation [136,169,172]. The increase in FAO caused by PPAR β/δ appears to be dependent on PGC-1 α [172], which itself is a PPAR β/δ target gene [171]. This evidence reveals that PPAR β/δ in skeletal muscle promotes a metabolic switch favoring FAO, reducing the availability of FAs to be accumulated in skeletal muscle and other tissues such as the liver, ultimately preventing obesity and insulin resistance [173]. Recent findings indicate that PPAR β/δ -mediated increases in skeletal muscle FA uptake are also coupled to diurnal hepatic PPAR β/δ activity through the synthesis of the phosphatidylcholine 18:0/18:1, an endogenous ligand of PPAR α [174].

Altogether, these findings suggest that PPAR β/δ activation increases the oxidation of FAs, reducing their availability to be accumulated as FA derivatives such as diacylglycerol or ceramides, which promote inflammation and insulin resistance. This PPAR β/δ -mediated

elevation in oxidative FA metabolism might explain why ligands of this receptor prevent lipid-induced ER stress, a process that induces inflammation and insulin resistance, and may explain the observed increase in the levels of ER stress markers in the skeletal muscles of PPAR β/δ ^{ex4} knockout mice [175]. Moreover, PPAR β/δ acts as a FA sensor in skeletal muscle by regulating the expression of several genes, including *Angptl4* [176,177], which affects lipid metabolism by inhibiting lipoprotein lipase [178] and activating lipolysis in adipose tissue [112,179,180]. The increase in skeletal muscle ANGPTL4 secretion caused by FAs-activated PPAR β/δ has been proposed to enhance adipose tissue lipolysis, ensuring the supply of FAs to skeletal muscle during states of increased FA β -oxidation, such as fasting and exercise [176]. In contrast, stimulation of cardiac *Angptl4* expression by FAs via PPAR β/δ has been considered part of a feedback mechanism aimed at protecting the heart against lipid overload by reducing lipoprotein lipase activity [177], which is responsible for hydrolyzing plasma TGs to release FAs. Another PPAR β/δ target gene is adipocyte differentiation-related protein (ADRP, also referred to as adipophilin or perilipin2), a lipid droplet-associated protein that binds to and sequesters intracellular FAs. ADRP is involved in intramuscular lipid accumulation and T2DM [173,181]. ADRP protein expression is muscle fiber type specific in humans and rats, with the highest protein content in fibers containing the most intramyocellular lipids. The muscle ADRP protein content was 2- to 3-fold higher in Zucker diabetic fatty rats during the progression of type 2 diabetes than in lean normoglycemic control rats, and this elevation in ADRP protein levels was paralleled by high intramyocellular lipid levels in diabetic animals [182]. In humans, ADRP is highly expressed in the vastus lateralis muscle, the largest and most powerful component of the quadriceps, of obese non-diabetic and obese diabetic patients in the fasting state [183]. Consistent with the proposed role of ADRP in the progression of T2DM, ADRP expression is

upregulated under circumstances of improved glucose tolerance. The upregulation of ADRP may act to sequester FAs as TGs in discrete lipid droplets that could protect muscle from the detrimental effects of FAs on insulin activity and glucose tolerance. Clearly, in addition to controlling muscle FA metabolism, PPAR β/δ is involved in glucose metabolism in muscles. Transgenic mice overexpressing PPAR β/δ specifically in muscle exhibited increased capacity for glucose oxidation by skeletal muscles through the cooperation of AMP kinase (AMPK) and the muscle-enriched transcription factor myocyte enhancer factor 2 (MEF2) [184]. Interestingly, this pattern of changes induced by PPAR β/δ resembles that following exercise. Additional protective effects of PPAR β/δ on fructose-induced insulin resistance may involve increased skeletal muscle expression of fibroblast growth factor 21 (*Fgf21*) [185], a hormone with a wide range of endocrine and autocrine effects on carbohydrate and lipid metabolism. Consistent with this hypothesis, circulating FGF21 levels are increased in humans in response to pharmacological activation of PPAR β/δ [186].

7.3 Liver

Non-alcoholic fatty liver disease (NAFLD) is the most common form of chronic liver disease and is considered to be the hepatic manifestation of metabolic syndrome [187]. The hallmark of NAFLD is persistent and excessive neutral lipid accumulation within hepatocytes. NAFLD can be benign, but it predisposes individuals to non-alcoholic steatohepatitis (NASH), which may result in fibrosis and/or cancer [188]. While the etiology of NASH is not well understood, the transition from NAFLD to NASH is considered a critical point in the progression of NASH pathology. Interestingly, the dual PPAR α/γ agonist Saroglitazar (Lipaglyn) [189],

which was launched in India for the treatment of hypertriglyceridemia and diabetic dyslipidemia in patients with T2DM not controllable by statins, is now also under examination in Phase III clinical trials evaluating its effect in adult NASH patients. Furthermore, recent evidence from pre-clinical [190] and clinical [191] studies has highlighted the potential of the dual-agonistic GFT505 (Elafibranor) targeting of both PPAR α and PPAR β/δ in the treatment of NASH. Lipids that accumulate in association with NAFLD may originate from increased uptake from chylomicrons, *de novo* lipogenesis and/or decreased lipid degradation [192,193]. Therefore, the ability of PPAR β/δ to modulate the activity of components important in lipogenesis and FAO may be responsible for the beneficial effects of PPAR β/δ activation in relation to NAFLD and NASH.

Lipogenesis is an enzymatic pathway that allows the cell to produce saturated and mono-unsaturated FAs, which can be used to replace TG energy stores, to generate lipid-based membranes or to produce lipid-dependent signaling molecules. *De novo* lipogenesis is very active in murine liver tissue after feeding but is repressed in response to fasting [194]. The mechanism by which feeding regulates the activity of lipogenic enzymes is largely transcriptional [195]. Numerous transcription factors have been shown to play a significant role in this process. Among these transcription factors, both SREBP-1c [196] and ChREBP [192] appear to integrate hormonal (insulin) and nutritional (glucose) signals, respectively, that coordinate the transcription of genes encoding lipogenic enzymes. Importantly, both SREBP-1c and ChREBP are regulated by LXR [193], a nuclear receptor that is activated by oxysterols [197]. Interestingly, SREBP-1c [198], ChREBP [199], LXR [200,201] and PPAR β/δ [202] appear to be involved in coordinating gene regulation in response to exogenous FAs (Figure 5). In fact, one article reported that PPAR β/δ suppresses lipogenesis by reducing SREBP-1c levels and

consequently ameliorating hepatic steatosis in obese diabetic *db/db* mice [203]. The mechanism by which PPAR β/δ reduces SREBP-1c activation might involve the induction of the insulin-induced gene (Insig)-1 protein, which blocks SREBP-1c activity by slowing the release of the SREBP cleavage-activating protein (SCAP)/SREBP complex from the ER. In one study, *Ldlr*^{-/-} mice fed a high-fat diet and treated with GW501516 showed reduced lipogenesis and increased insulin sensitivity [204]. However, several other studies have suggested that PPAR β/δ may promote *de novo* lipogenesis. The increase in hepatic steatosis following treatment with a PPAR β/δ agonist has been reported to be strictly time-dependent and evident only after 4 weeks of treatment, whereas longer treatments were ultimately protective against liver steatosis [204,205]. Comparisons of the gene expression profiles of livers between control and GW501516-treated *db/db* mice suggest that PPAR β/δ activation reduces hepatic glucose production by increasing glycolysis and the activity of the pentose phosphate shunt as well as by enhancing lipogenesis in the liver [206]. Moreover, a recent study performing adenovirus-mediated liver-restricted PPAR β/δ activation confirmed that PPAR β/δ activation increases hepatic *de novo* lipogenesis [207]. However, during hepatic lipid accumulation, PPAR β/δ activation has been shown to increase the production of monounsaturated FAs and reduce the formation of saturated FAs, leading to reduced hepatic damage and JNK-mediated stress signaling. These findings suggest that hepatic PPAR β/δ -regulated lipogenesis protects against lipotoxicity. Finally, one unique and recent report describing experiments performed in mice specifically lacking PPAR β/δ in hepatocytes provides evidence that PPAR β/δ is involved in the circadian control of lipogenesis [174]. Importantly, this work also indicated that this diurnal control by PPAR β/δ significantly influences whole-body physiology. Specifically, PPAR β/δ from hepatocytes regulates the production of FAs required to produce phosphatidylcholine

18:0/18:1 PC (18:0/18:1), which acts as a hepatic lipokine for communication with skeletal muscles, thus influencing FAO in myocytes. Administration of GW501516 to wild-type [137] or LDLR^{-/-} mice [204] enhanced hepatic FAO and AMPK activity, probably by increasing the AMP:ATP ratio in the liver [137]. As a result of PPAR β/δ activation, a reduction in substrate availability indirectly activates AMPK, and this effect also contributes to the glucose-lowering effect of this nuclear receptor [207]. Additionally, it has recently been shown that PPAR α is required for the effect of PPAR β/δ agonists on steatosis [205]. A recent report identified PPAR α from hepatocytes as a critical transcription factor for FA degradation in hepatocytes and in protection against NAFLD [208]. How PPAR β/δ may influence PPAR α activity in hepatocytes remains to be established, and whether this process occurs through a cell-autonomous pathway remains to be investigated.

One key issue in understanding the role of PPAR β/δ in the liver is elucidating its respective roles in different cell types because PPAR β/δ is highly expressed in Kupffer and stellate cells. These cell types are involved in the control of inflammation and fibrosis, which develops in the most severe stages of NAFLD. PPAR β/δ controls the expression of the alternative anti-inflammatory phenotype of Kupffer cells, the resident macrophages in the liver; additionally, transplantation of PPAR β/δ ^{-/-} bone marrow into wild-type mice reduces the alternative activation of Kupffer cells, leading to mitochondrial dysfunction and insulin resistance in hepatocytes [209,210]. In wounded areas of the liver, stellate cells are stimulated by factors that promote their proliferation and transition from a quiescent, vitamin A-storing phenotype towards an activated, proliferative myofibroblast-like phenotype. In turn, these cells contribute to the production of abnormally high levels of ECM proteins and remodeling factors, which ultimately results in ECM accumulation [211]. PPAR β/δ is highly expressed in hepatic

stellate cells and contributes to the enhancement of their proliferation [212]. PPAR β/δ is also involved in the control of vitamin A metabolism during the activation of stellate cells [213]. Pre-clinical studies have shown contrasting results regarding the protective [209,214] and detrimental effects [215] of PPAR β/δ agonists on liver fibrosis.

7.4 Pancreas

The clinical onset of T2DM develops when pancreatic β -cells fail to compensate for the decrease in insulin sensitivity due to insulin resistance. Incretins, such as glucose-dependent insulinotropic polypeptide (GIP) and glucagon-like peptide-1 (GLP-1), are intestinal postprandial hormones that stimulate insulin release from the pancreas in response to nutrients. In addition, incretins can preserve the morphology and function of pancreatic β -cells [216]. Treatment of wild-type or *ob/ob* mice with a PPAR β/δ agonist potentiates GLP-1 production by the small intestine, leading to increased plasma levels of this incretin after an oral glucose load and improved glucose tolerance [217]. Moreover, PPAR β/δ is required for the expression of genes involved in mitochondrial function and, thus, the production of ATP, which is required for glucose-stimulated insulin secretion (GSIS). Activation of PPAR β/δ in β -cells is triggered by the lipolytic activity of the major triacylglycerol hydrolase desnutrin [218]. Thus, PPAR β/δ agonists restore GSIS and the expression of mitochondrial oxidative genes in mice in which desnutrin is ablated in β -cells. Because desnutrin levels are reduced in obesity, these findings suggest a mechanism by which prolonged treatment of obese and diabetic *db/db* mice with a PPAR β/δ agonist reduces blood glucose levels in association with improved insulin sensitivity and pancreatic islet function [219]. Interestingly, specific deletion of *Ppar β/δ* (PPAR β/δ ^{ex4} knockout) in the epithelial compartment of the mouse pancreas led to increased numbers of islets and

enhanced insulin secretion, resulting in hyperinsulinemia [220]. The broad repressive role of PPAR β/δ in pancreatic β -cells also affects the vesicular and granular compartment and the actin cytoskeleton. Studies of insulin release from isolated PPAR β/δ -deficient islets revealed an accelerated second phase of GSIS that parallels an increase in filamentous actin (F-actin) disassembly and correlates with elevated protein kinase D activity, resulting in alterations in Golgi organization. This evidence shed new light on the metabolic function of PPAR β/δ by indicating a repressive effect of PPAR β/δ on β -cell mass and insulin exocytosis. Together, these results underscore a possible dual function of PPAR β/δ in β -cells, combining gene activation and gene repression/derepression.

8. PPAR β/δ and cancer

Among the different physiological and pathological functions attributed to PPAR β/δ , its involvement in cancer is certainly the topic in which studies of PPAR β/δ have shown most discrepancies. Overall, PPAR β/δ has been shown to be involved in carcinomas, in which it appears to have opposing effects. In this section, we discuss findings from *in vivo* studies of mouse cancer models and human tumors to characterize the advances in research on the biological functions of PPAR β/δ in cancer.

The *in vivo* involvement of PPAR β/δ in the development and progression of colorectal cancer and skin cancer has been well documented in different studies; however, the precise functions of PPAR β/δ remain controversial.

8.1 Function in colorectal carcinoma (CRC) development

PPAR β/δ is highly expressed in the small and large intestines in rodents and humans, and *in situ* hybridization studies have shown that this nuclear receptor is expressed throughout the intestinal epithelium, with higher expression at the bottom of the crypts. Function of PPAR β/δ has been proposed in Paneth cells, enteroendocrine cells and enterocytes [221]. Two models are classically used for *in vivo* studies of CRC development. One of these models is the multiple intestinal neoplasia (Min) mouse strain, which carries a heterozygous nonsense mutation in the adenomatous polyposis coli (*Apc*) gene (*Apc*^{min/+}). This mutation predisposes mice to developing cancer, and examination of this mouse strain facilitates the identification of factors that regulate the transition between normal and neoplastic cell growth. However, adenoma formation requires the inactivation of both *Apc* alleles. Of note, *Apc* has been found to be mutated in both sporadic CRC and hereditary familial adenomatous polyposis (FAP) in humans [222–224]. The second model involves chemically induced colon cancer using the mutagenic agent azoxymethane (AOM), which exerts colonotropic carcinogenicity. Repeated intraperitoneal administration of AOM results in the spontaneous development of tumors within 30 weeks. As an alternative approach, inflammation-dependent tumor growth can be investigated by administering a combination of AOM and the inflammatory agent dextran sodium sulfate (DSS) in drinking water, which causes rapid growth of multiple colon tumors within 10 weeks [225].

The first study investigating PPAR β/δ function in CRC showed that *Pparb/d* is induced by β -catenin/T-cell factor 4, which in turn induces the well-characterized proto-oncogenes cyclin D1 and c-Myc, suggesting that PPAR β/δ could play a role in CRC development [226]. To investigate PPAR β/δ function in intestinal carcinogenesis, total and intestine-specific KO of PPAR β/δ have been performed. In mice in an *Apc*^{min/+} genetic background, PPAR β/δ ^{ex4} deletion decreased intestinal adenoma growth and inhibited the tumor-promoting effects of the PPAR β/δ

agonist GW501516 [227]. Consistent with this finding, intestine-specific PPAR $\beta/\delta^{\text{ex4}}$ deletion in mice subjected to AOM treatment resulted in resistance to the chemical induction of CRC [89]. In both studies, these effects were attributed to the stimulatory effects of PPAR β/δ on *Vegf* expression, and it was suggested that PPAR β/δ acts as a pro-tumorigenic factor in CRC development [227–229]. Another study conducted using the PPAR $\beta/\delta^{\text{ex4}}$ *Apc^{min/+}* model supports these findings [230]. These observations support earlier findings that PPAR β/δ stimulates *Vegf* expression in bladder cancer cells [229]. Other studies have shown that PPAR β/δ exerts a protective effect against genetic and chemically induced intestinal tumorigenesis. AOM administration to PPAR $\beta/\delta^{\text{ex8}}$ mice, with or without GW0742 co-administration, resulted in an enhanced number of colon polyps compared to AOM administration to wild-type mice [72,227]. When crossed with mice in an *Apc^{min/+}* genetic background, PPAR $\beta/\delta^{\text{ex8}}$ -deleted mice displayed a sex-specific difference in tumor susceptibility. PPAR $\beta/\delta^{\text{ex8}}$ deletion also differentially affected tumor size and location depending on the study, which hinders a clear understanding of the anti-tumor activities of PPAR β/δ . Indeed, some studies have shown that PPAR β/δ deletion enhances the formation of polyps in the colons of male and female mice but that polyps in the small intestine appeared only in female mice [231]. However, other studies have shown that PPAR $\beta/\delta^{\text{ex8}}$ deletion combined with the *Apc^{min/+}* genotype led to an increase in colon polyp count only in females and that the polyp sizes were increased in both sexes. In the small intestine, polyp count did not differ between sexes [231]. This result is in accordance with another study, which showed that PPAR $\beta/\delta^{\text{ex8}}$ specifically leads to an increase in the size and number of small intestinal polyps, without affecting the colon polyp count [229,232]. Taken together, these studies highlight the complexity of the role of PPAR β/δ in CRC formation, in which sex is a determining factor. Highly contrasting results emerged from studies using two different specific

pharmacological agonists of PPAR β/δ . While *Apc*^{min/+} mice treated with GW501516 for 6 weeks displayed an increase in the total number of polyps and in polyp size [227,233], long-term (22 weeks) activation of PPAR β/δ with its ligand GW0742 attenuated chemically induced colon carcinogenesis, and no effects on polyp number and size were observed in *Apc*^{min/+} mice treated with GW0742 for 6 weeks [229,232].

Consistent with a pro-tumorigenic role of PPAR β/δ , a recent elegant study by Beyaz et al. (2016) showed that dietary fat (60% fat) activated PPAR β/δ and promoted the proliferation of intestinal Lgr5⁺ stem and progenitor cells [230]. Treating mice with the PPAR β/δ agonist GW501516 produced similar tissue remodeling and regeneration. The mechanism underlying these effects involved the Wnt- β -catenin signaling pathways. Importantly, HFD- and agonist-activated PPAR β/δ signaling endowed an organoid-initiating capacity to progenitor cells, and enforced PPAR β/δ signaling permitted these progenitor cells to form *in vivo* tumors after loss of the tumor suppressor *Apc*. These findings highlight how diet-modulated PPAR β/δ activation alters not only the function of intestinal stem and progenitor cells but also their capacity to initiate tumors [230,234].

In humans, PPAR β/δ mRNA expression was found to be strongly upregulated in colorectal tumor samples compared to paired, adjacent, normal tissue samples in some studies [226,235], although other studies showed only a modest [236] or no difference in PPAR β/δ gene expression between CRC and normal tissue [237,238]. However, there appears to be massive heterogeneity between CRC samples [239], which could be due to the heterogeneity of tumor tissues, the tumor stage analyzed, and treatments applied to the tissues after surgical ablation.

Immunohistological and protein quantification analyses have been also performed to examine the involvement of PPAR β/δ in human CRC; these studies have been reviewed

extensively by Neels and Grimaldi [240]. The results obtained from these analyses should be considered with caution due to the lack of specificity of commercially available anti-PPAR β/δ antibodies [241]. Moreover, the activity level of PPAR β/δ based on the mRNA expression levels of its target genes could be a better marker of PPAR β/δ activity in CRC and, more generally, in cancer development.

Altogether, the studies of animal cancer models or human tumors conducted to date have failed to clearly characterize the function of PPAR β/δ in CRC development. Several hypotheses have been proposed to explain the disparities between studies [242]. Interestingly, the genetic background of the *Apc*^{min/+} PPAR β/δ mice used in various studies can greatly impact CRC development [243]. Indeed, some studies have been performed using PPAR β/δ -deficient mice in a mixed genetic background, while others used mice in the pure C57Bl/6 background. These differences become important when the animals are crossed with C57Bl/6 *Apc*^{min/+} mice, as the genetic background could strongly influence the penetrance of the Min phenotype due to strain-specific impacts of the loci, termed modifiers of the Min phenotype (Moms). Indeed, *Apc*^{min/+} mice in the C57Bl/6 background are more prone to develop intestinal tumors than *Apc*^{min/+} mice in a mixed genetic background [243,245]. Moms are likely the cause of this variation, and there is considerable evidence that the genetic background is responsible for major differences in adenoma counts. The different strategies used to ablate PPAR β/δ could be another major reason for the discrepancies in the findings between studies (see **PPAR β/δ in placentation**). In addition, animal housing conditions might influence other processes involved in intestinal tumorigenesis, such as inflammatory status or the COX2 enzymatic pathway [246,247], which could also explain the disparities among the diverse studies using these animal models. Moreover, recent results from studies of CRC tumorigenesis showed that both the gut microbiome, which differs

between animal facilities and diets administered, and the circadian rhythm, which could influence the expression of genes encoding proteins involved in DNA repair and the cell cycle such as the proto-oncogene c-Myc, are critical for the initiation and progression of CRC, as supported by clinical and epidemiological data [247–252]. Although the evidence from many studies favors a pro-tumorigenic role of PPAR β/δ , its precise role may be stage- and tumor-specific and requires further investigation. We propose that future reports also indicate the genetic models of PPAR β/δ used, such as PPAR $\beta/\delta^{\text{ex8}}$ or PPAR $\beta/\delta^{\text{ex4}}$, as well as the genetic background, housing conditions, and dietary intake in which these mutations are studied.

8.2 PPAR β/δ function in skin carcinoma development

Squamous cell carcinomas (SCCs) and basal cell carcinomas (BCCs), both of which are derived from keratinocytes, are the most common types of non-melanoma skin cancer (NMSC). Although NMSCs are rarely fatal, they can be highly disfiguring and can result in a heavy personal burden and an enormous cost to society. Approximately 90% of NMSC cases are associated with excessive and/or chronic exposure to ultraviolet (UV) radiation from the sun; in particular, UVB radiation (290–320 nm, sunburn rays) shows low skin penetration and therefore predominantly affects keratinocytes [253,254]. The role of PPAR β/δ in skin carcinoma development has been investigated *in vivo* using two different experimental models: chronic UVB radiation and a two-stage model based on the chemical induction of tumors via topical application of 7,12-dimethylbenz[*a*]anthracene/12-O-tetradecanoylphorbol 13-acetate (DMBA/TPA), in which DMBA is used as a carcinogen to generate *H-Ras* mutations, followed by 20 weekly applications of the nonmutagenic agent TPA, which provides a microenvironment and proliferative stimuli that favors the clonal outgrowth of initiated keratinocytes. These two

methodological approaches have shown contradictory results. Chronic UV exposure to wild-type and PPAR β/δ ^{ex4} mice showed that PPAR β/δ promotes UV-induced skin tumorigenesis *in vivo* by upregulating the Src gene and protein expression as well as Src activity, which in turn enhances the EGFR/Erk1/2 signaling pathway in response to UVB radiation [249,255]. In this mouse model, PPAR β/δ sensitizes keratinocytes to UV radiation and promotes the development of actinic keratosis and progression of tumors by stimulating the expression of epithelial-mesenchymal transition (EMT) markers that affect basement membrane integrity. Interestingly, PPAR β/δ expression directly correlates with *SRC*, *MMP19* and *SNAIL* expression in SCC samples from human skin, and these relationships suggest a direct effect of PPAR β/δ on EMT initiation in human SCC. Furthermore, a meta-analysis of PPAR β/δ activity in various epithelial tumors, including human SCC, revealed a positive interaction between the expression levels of PPAR β/δ and those of two of its targets, *SRC* and *TGFBI*. The identification of these relationships in large, independent datasets of various tumor types, particularly in lung, colon and ovarian cancers known to have high levels of Src expression and activity, strongly supports a pro-carcinogenic role of PPAR β/δ in human carcinomas [255]. One can speculate that early in the process of UV-induced skin actinic keratosis formation, PPAR β/δ regulates Src and may act in concert with genetic mutations resulting from UV exposure to promote tumor progression by increasing the migratory and proliferative properties of keratinocytes. These data help to define the pro-tumorigenic properties of PPAR β/δ in skin carcinoma and most likely in carcinomas originating from other tissues. Inflammation is increasingly recognized as an essential component of tumor development [256,257]. Recently, a novel PPAR β/δ -miR-21-3p axis was reported to be involved in UV radiation-induced skin inflammation [258]. This PPAR β/δ -dependent molecular cascade involving TGF β 1 and the passenger strand of the miRNA miR-21-

3p was activated in the epidermis in response to UV exposure. miR-21-3p performs a pro-inflammatory function in keratinocytes, and a high level of miR-21-3p expression in human skin was associated with psoriasis and SCC. Importantly, inhibiting miR-21-3p reduced UV radiation-induced cutaneous inflammation in *ex vivo* human skin biopsies, underscoring the clinical relevance of this signaling axis to inflammatory cutaneous disorders [258]. In contrast, Kim et al. (2004) showed anti-cancer properties of PPAR β/δ in skin carcinoma using a DMBA/TPA-induced model of carcinogenesis, in which tumor development was more severe in PPAR $\beta/\delta^{\text{ex8}}$ mice than in wild-type animals. Their results suggest that PPAR β/δ attenuates the development of chemically induced skin cancer by inhibiting keratinocyte proliferation *via* PKC α [259]. However, in mice, susceptibility for DMBA/TPA-induced skin tumorigenesis was not affected by transgenic overexpression of PKC α [250,251]. Using the same model of DMBA/TPA-induced tumorigenesis, topical treatment of wild-type and PPAR $\beta/\delta^{\text{ex8}}$ mice with a PPAR β/δ agonist (GW0742) inhibited chemically induced skin tumorigenesis only in wild-type mice by reducing skin tumor formation and multiplicity [260].

The discrepancies in the results obtained using these two models could be explained by the fact that induction of skin cancer using DMBA/TPA involves different molecular mechanisms and creates different genetic mutations than those associated with skin cancer caused by chronic UV radiation exposure [261,262]. Notably, tumor initiation via administration of a single topical sub-carcinogenic dose of a genotoxic carcinogen (DMBA) primarily induces activating mutations in *H-Ras*. The frequency of *H-Ras* mutations of only 10–20% in human SCC and BCC reflects a minor contribution of this oncogene to the development of human skin carcinomas [263–265]. In this context, the pro-carcinogenic function of PPAR β/δ via the EGFR/Src pathway could be masked by activating *H-Ras* mutations, rendering activation of the

downstream MAPK Erk1/2 independent of Src/EGFR pathway activation. These differences in results between studies also highlight that the mutational status of the tumor cell and the tumor environment could modulate the activities of PPAR β/δ in cancer development. This dual function of PPAR β/δ as a pro- or anti-carcinogenic effector parallels the dual function of TGF β 1 in skin carcinoma, which has been defined as a PPAR β/δ target gene [266]. Indeed, TGF β 1 acts as both a tumor suppressor and an oncogene in skin carcinoma, a dual specificity that appears to depend on the stage of the cancer [267,268]. The generally accepted paradigm for TGF β 1 is that it acts as a tumor suppressor during the early stage of tumor formation but as a promoter of malignant conversion and metastasis in the later stages of skin carcinoma development in the DMBA/TPA model [269,270]. However, pharmacological inhibition of TGF β 1 signaling during the tumor promotion phase (TPA application) leads to a reduction in benign tumor formation. However, once formed, these tumors display an increased frequency of conversion to SCC [267]. Moreover, in a model of UVB radiation-induced skin carcinoma, TGF β 1 has been shown to exert a pro-tumorigenic effect that enforces PPAR β/δ function [249,271]. Regarding TGF β 1, it has been proposed that the conversion from its tumor-suppressing activities to its oncogenic activities reflects genetic or epigenetic alterations in the signaling pathways in tumor cells that alter the downstream effects of the TGF β 1 pathway [267,272,273]. This evidence clearly highlights the importance of the genetic configuration of the tumor cell in managing the TGF β 1 response, which could also be the case for PPAR β/δ . Intriguingly, studies from independent laboratories have clearly demonstrated PPAR β/δ as a pro-differentiating factor in keratinocytes after skin injury [274–276]. Similarly, a pro-differentiation function has been attributed to TGF β 1 [277] and the MAPK Erk1/2 pathway in keratinocytes during skin renewal, which is a pathway that is clearly pro-carcinogenic during skin tumor formation [278,279]

Together, these studies highlight the dual function of PPAR β/δ in tumor development, as its function is likely modulated by the genetic alterations and epigenetic modifications that occur during tumor formation. However, this hypothesis must be tested to clearly address the putative cellular context-dependent specificities of the effects of PPAR β/δ on tumor cell responses. These findings also illustrate the crucial role of the tumor environment, which could regulate and/or be regulated by PPAR β/δ in tumor cells, as PPAR β/δ is able to drive the expression of circulating factors that can affect tumor development, such as VEGF, ANGPTL4 and TGF β 1.

9. PPAR β/δ in neurological diseases

The spatial expression of PPAR β/δ has been examined in the central nervous system (CNS) of rats and mice. All the PPAR isotypes are coexpressed in the nervous system during rat embryogenesis, and PPAR β/δ is the predominant isotype [76,280]. PPAR β/δ can be detected in the rat CNS beginning from E13.5 of gestation and remains highly expressed in the CNS postnatally [76]. Immunolocalization studies showed high levels of PPAR β/δ in the hippocampus, telencephalic cortex and cerebellar cortex of rats. In the rat spinal cord, PPAR β/δ is the most abundant PPAR isotype and is expressed in every cell layer [281]. A detailed analysis of mouse brains revealed that PPAR β/δ mRNA and protein are expressed throughout the brain, with particularly high levels in the entorhinal cortex, hypothalamus and hippocampus and lower levels in the corpus callosum and caudate-putamen. At the cellular level, PPAR β/δ was found to be expressed in oligodendrocytes and neurons but not in astrocytes [282]. However, PPAR β/δ expression has been reported in *in vitro* cultures of rat and mouse astrocytes [281,283]. These observations suggest a role of PPAR β/δ in neuronal function within the CNS. Despite the comparatively low expression of PPAR β/δ in the corpus callosum, histological examinations of

PPAR $\beta/\delta^{\text{ex8}}$ knockout mice have revealed alterations in the extent of myelination [81]. This deficiency in myelination was not observed in other brain regions, including the cerebellum and brainstem. However, the levels of mRNAs encoding for proteins important for myelination, such as myelin basic protein (MBP), proteolipid protein (PLP), and two brain-specific acyl-CoA synthases (ASC2 and ASC3), in the corpus callosum were similar between PPAR $\beta/\delta^{\text{ex8}}$ mice and their wild-type counterparts. Whether this region-specific defect in myelination also occurs in PPAR $\beta/\delta^{\text{ex4}}$ mice remains to be investigated [72,73]. Neuron-specific deletion of PPAR β/δ (Nestin-PPAR $\beta/\delta^{\text{ex4}}$) in mice did not cause any apparent developmental abnormalities in comparison to floxed littermate control mice. Nissl staining of coronal sections of the hippocampus and hypothalamus revealed no evident differences in the structure of nuclei in the hippocampus, hypothalamus, or any other forebrain structure between Nestin-PPAR $\beta/\delta^{\text{ex4}}$ mice and corresponding control mice [279]. In summary, although these observations suggest that deletion of PPAR β/δ from neurons does not cause gross anatomical malformations, functional alterations have been reported in mice deficient in PPAR β/δ [81,283–285].

9.1 Neuroprotective role in neurological injuries

Cerebral ischemia (also known as cerebrovascular ischemia) is a condition in which there is insufficient blood flow to the brain to meet metabolic demand. This leads to poor oxygen supply, or cerebral hypoxia, and thus results in the death of brain tissue or cerebral infarction/ischemic stroke. In mice and rats, agonist-activated PPAR β/δ has been shown to reduce ischemic brain injury [280,286]. In a model of ischemic brain injury based on middle cerebral artery occlusion (MCAO) in rats, intracerebroventricular infusion of the PPAR β/δ agonist L-165041 or GW501516 significantly attenuated ischemic brain damage. Lending

support for a neuroprotective role of PPAR β/δ , the results from this model showed that ischemic damage was doubled and the hyperphagic response was absent in PPAR $\beta/\delta^{\text{ex4}}$ mice compared with wild-type mice [287]. Hyperphagia is the abnormal increase in appetite associated with hypothalamic injury. Indeed, increased malondialdehyde (MDA) and interferon γ (IFN- γ) levels as well as reduced glutathione and manganese superoxide dismutase (MnSOD) levels after MCAO in PPAR $\beta/\delta^{\text{ex4}}$ knockout mice are indicative of hypothalamic injury. Oxidative stress in the brain was dramatically enhanced in these PPAR $\beta/\delta^{\text{ex4}}$ mice. Enhanced oxidative stress and inflammatory responses in the ischemic brain contribute to neuronal death. The neuroprotective role of PPAR β/δ was associated with its anti-inflammatory and antioxidant properties, as demonstrated by an increased content of MDA and decreased expression levels of antioxidant genes, such as glutathione, MnSOD and catalase, in PPAR $\beta/\delta^{\text{ex4}}$ mice [280,281,286].

Apoptosis in neurons and oligodendrocytes is a prominent feature after spinal cord injury (SCI) and may greatly contribute to patient paralysis [288]. *In vivo* animal studies have shown that administration of the PPAR β/δ agonist GW0742 to mice after SCI improves their limb functions. Conversely, the PPAR β/δ antagonist GW0660 has been shown to block the effect of GW0742 on motor function, suggesting that the neuroprotective effects of GW0742 are dependent on ligand-activated PPAR β/δ [283,289]. PPAR β/δ is also involved in the neuroprotective activities of palmitoylethanolamide (PEA) after SCI. PEA is an endogenous FA amide that displays anti-inflammatory and analgesic activities [283,289]. Similarly, the effects of GW0742 are associated with reduced oxidative stress and pro-inflammatory responses. Ligand-activated PPAR β/δ has been shown to reduce neutrophil infiltration, pro-inflammatory cytokine expression, and oxidative damage, including nitrotyrosine formation and lipid peroxidation [283].

Multiple sclerosis (MS) is a potentially disabling disease of the brain and spinal cord. In MS, the immune system attacks the myelin that covers nerve fibers and causes communication deficiencies between the brain and the remainder of the body. Ultimately, MS causes the nerves themselves to deteriorate or become permanently damaged. There is currently no cure for MS. However, treatments can help speed recovery from attacks and modify the course of the disease. Experimental autoimmune encephalomyelitis (EAE), in which mice are immunized with the encephalitogenic peptide myelin oligodendrocyte glycoprotein, is a well-recognized model of MS. Interestingly, treatment with the selective PPAR β/δ agonist GW0742 exerts a notable effect on clinical outcome [290]. Oral administration of GW0742 in this mouse model of EAE only modestly attenuated clinical symptoms when the drug was administered simultaneously with immunization. A further reduction in clinical symptoms accompanied by a reduction in the appearance of new cortical lesions was observed when GW0742 was administered during EAE progression. Additionally, Polak et al. (2005) showed that IFN- γ production by T cells was not affected by GW0742, although GW0742 reduced astroglial and microglial inflammatory activation and IL-1 β levels in the brains of mice subjected to EAE [290]. This result is interesting because numerous studies using this animal model of MS have provided convincing evidence that T cells specific for self-antigens mediate the disease pathology. T helper type 1 (Th1) cells were previously thought to be the main effector T cells responsible for autoimmune inflammation [291]. However, recent studies have highlighted an important pathogenic role of CD4(+) T cells that secrete interleukin (IL)-17, termed Th17 cells, and IL-17-secreting $\gamma\delta$ T cells in EAE as well as other autoimmune and chronic inflammatory conditions. In contrast to the findings of Polak et al. (2005), two studies have shown that PPAR β/δ ameliorates EAE in C57BL/6 mice by blocking IFN- γ and IL-17 production by T helper type 1 (Th1) and Th17 cells

[286,292]. The inhibition of EAE by the PPAR β/δ agonists GW501516 and L165041 was also associated with the reduction of IL-12 and IL-23 levels in antigen-presenting cells and the inhibition of Th1 and Th17 polarization [292]. Independently, Dunn et al. (2010) showed that PPAR $\beta/\delta^{\text{ex8}}$ mice exhibited a more severe course of classical EAE, characterized by persistent hindlimb weakness and a reduced frequency of recovery after the initial acute phase of EAE [286]. The more severe EAE that developed in PPAR $\beta/\delta^{\text{ex8}}$ mice was due to immune-mediated myelin and axon damage. PPAR $\beta/\delta^{\text{ex8}}$ mice also displayed more inflammatory foci in the CNS during the course of EAE, in association with preferential expansion of CD4(+) and CD19(+) cells [286]. The molecular mechanisms involved in the regulation of Th1 and Th17 responses in EAE by PPAR β/δ agonists remain unclear. In brief, analogous to the anti-apoptotic, anti-inflammatory and anti-oxidative activities of PPAR β/δ during skin wound repair (see **Tissue repair**), PPAR β/δ activation in the CNS has been shown to confer neuroprotective effects, thus attenuating neuro-inflammation, in many neurological injuries.

9.2 Function in neurological diseases

Many research groups have begun to explore the possible benefit of targeting PPAR β/δ in neurological diseases, such as Alzheimer's disease (AD). AD is a neurodegenerative disorder characterized by cognitive deterioration, memory loss, progressive functional impairment, and multiple behavioral and psychological disturbances. The formation of amyloid fibrils and neurofibrillary tangles is thought to contribute to the degradation of neurons in the brain and the subsequent symptoms of AD [293]. Amyloid plaques primarily composed of β -amyloid (A β) peptide progressively form in the brains of AD patients, and mutations in three genes (amyloid precursor protein [APP], presenilin 1 and presenilin 2) cause early-onset familial AD by directly

increasing production of the toxic, plaque-promoting A β peptide [293]. A β peptide, the product of the large type 1 trans-membrane protein APP, is produced in a two-step proteolytic process initiated by the β -secretase BACE1 and followed by the γ -secretase presenilin 1 or 2. Due to its apparent rate-limiting activity, BACE1 appears to be a prime target for prevent A β peptide generation in AD [294,295]. In the healthy brain, these protein fragments are degraded and eliminated. In AD, these fragments aggregate to form insoluble plaques. The receptor for advanced glycation end-products (RAGE) is a multiligand receptor that transports circulating A β peptides across the blood-brain barrier (BBB) into the brain. The RAGE-A β peptide interaction at the BBB leads to oxidative stress, inflammatory responses and reduced cerebral blood flow. Thus, regulating RAGE activity at the BBB and/or within the brain could be beneficial to AD patients [296–298]. A β peptide interacts with the signaling pathways that regulate the phosphorylation of the microtubule-associated protein tau. Hyperphosphorylation of tau disrupts its normal function in regulating axonal transport and leads to the accumulation of insoluble, twisted fibers termed neurofibrillary tangles within neurons. Furthermore, degradation of hyperphosphorylated tau by the proteasome is inhibited by the actions of A β peptide. These two proteins and their associated signaling pathways therefore represent important therapeutic targets for AD [295,298].

PPAR β / δ ^{ex4} mice showed cognitive impairment in an object recognition task in association with increased IL-6 expression, tau phosphorylation, and levels of BACE1 and RAGE, which are involved in A β peptide synthesis and deposition. The protein levels of glial fibrillary acidic protein (GFAP) were increased in the cortex of PPAR β / δ ^{ex4} mice, suggesting the presence of reactive astrogliosis [299,300]. Astrocyte activation and the release of pro-inflammatory cytokines that regulate astrocytic hypertrophy and proliferation, collectively referred to as

reactive astrogliosis, is a ubiquitous but poorly understood hallmark of all CNS pathologies, including AD. The increases in the levels of GFAP, inflammatory cytokines such as IL-6 and TNF α , BACE1 and RAGE were proposed to increase NF- κ B DNA-binding activity in the cortex of PPAR β/δ ^{ex4} mice [300]. The expression of RAGE and BACE1 is regulated by the transcription factor NF- κ B. The ability of the PPAR β/δ agonist GW0742 to reduce the amyloid burden and suppress neuroinflammation in 5XFAD mice, a transgenic mouse model of AD, has been reported by two independent groups [299,301]. PPAR β/δ activation decreased A β peptide levels in the subiculum, hippocampus and cortical layer V. Notably, Malm et al. (2015) showed that these reductions were specific to extracellular deposits because the number of APP/A β peptide-positive neurons remained unchanged by PPAR β/δ activation [301]. Different mechanisms of action of PPAR β/δ have been proposed. In a study by Kalinin et al. (2009), the investigators found that the GW0742-mediated change in the plaque burden was accompanied by increased expression of the amyloid-degrading enzymes neprilysin and insulin-degrading enzyme [299]. GW0742 also reduced astrocyte activation, suggesting its anti-inflammatory effects on glia. In contrast, Malm et al. (2015) proposed that the A β reduction was due to enhanced microglial-mediated clearance of A β peptides. The authors also suggested that differences in GW0742 treatment regimens and in the age of the mice used between the two studies might have resulted in the disparate outcomes [301]. Clearly, additional investigations are needed to clarify the precise reason for these discrepancies, and such clarity would help in evaluating PPAR β/δ ligands as potential treatments for AD.

AD has been described as “Type 3 diabetes” that selectively involves the brain, as AD has molecular and biochemical features that overlap with both type 1 diabetes mellitus and T2DM [302–304]. Insulin and IGF-1 receptors in the brain begin to disappear early in Alzheimer's

disease and continue to be downregulated as the disease progresses. The progressive breakdown of insulin signaling results in increased oxidative stress, impaired metabolism and cell death, all of which lead to neurodegeneration. Intracerebral administration of streptozotocin to rats or mice depletes the brain of insulin and mimics the neurodegeneration observed in AD (i.e., plaque deposits, neurofibrillary tangles, diminished brain size, impaired cognitive function, cell loss and overall brain deterioration) [302,303,305]. Thus, it has been proposed that AD may be treatable, preventable and curable using anti-diabetic drugs. In phase I/II trials, the PPAR β/δ and PPAR γ agonist T3D-959 (www.t3dtherapeutics.com) has been shown to possess striking activities in preserving the hypothalamus and the temporal lobes, areas of the brain responsible for memory, learning, and behavior [306,307]. These activities were found to be mediated by increased binding of insulin to insulin receptors, reduced levels of oxidative stress and tau phosphorylation, and increased choline acetyltransferase expression in the brain [308]. AD appears to be caused by parallel abnormalities, impaired insulin signaling and oxidative stress, which is regulated by the NOS and NOX protein families [309]. PPAR β/δ showed the greatest function in reducing oxidative stress and improving learning and memory.

Parkinson's disease (PD) is a degenerative disorder of the CNS that mainly affects the motor system. The pathologic hallmark of PD is the loss of dopaminergic neurons in the *substantia nigra*, a region of the midbrain, and the presence of intracytoplasmic inclusions termed Lewy bodies, which are aggregates of α -synuclein and ubiquitin [310]. The causes of cell death in PD are poorly understood. The progression of PD has been attributed to the enzymatic and non-enzymatic oxidation of dopamine, which generates ROS that induce apoptotic cell death in dopaminergic neurons. Experimental models for dopaminergic neurodegeneration associated with PD can be established using neurotoxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine

(MPTP) [311,312]. The connection between PD and MPTP was traced to the illicit use of an analog of the narcotic meperidine tainted with MPTP by several drug abusers [309]. MPTP induces most of the biochemical, pathological and clinical features related to PD in nonhuman primates. MPTP, which is lipid soluble, readily penetrates the BBB and enters brain cells. MPTP itself is not toxic, but its oxidation by monoamine oxidase B in astrocytes and serotonergic neurons produces a toxic cation 1-methyl-4-phenylpyridinium (MPP⁺). MPP⁺ primarily kills dopamine-producing neurons by interfering with complex I of the electron transport chain, a component of mitochondrial metabolism, which leads to cell death and causes the buildup of free radicals, toxic molecules that further contribute to cell destruction [280,311,313,314]. In this model, the PPAR β/δ levels in mice were shown to be increased immediately after MPTP treatment compared with saline treatment [280,314], and heterozygous PPAR $\beta/\delta^{\text{ex4}}$ mice were not protected from MPTP toxicity. However, intra-striatal infusion of the PPAR β/δ agonist GW0742 reduced the MPTP-induced loss of dopaminergic neurons in mice compared to vehicle infusion [314]. In contrast, a recent study using another PD animal model established via injection of 6-hydroxydopamine (6-OHDA) into the *substantia nigra* to selectively cause ROS-mediated destruction of dopaminergic neurons suggested that PPAR β/δ activation leads to a decrease in expression of full-length tropomyosin receptor kinase B, which consequently increases neuronal death, thereby contributing to the progression of PD [304]. The levels of PPAR β/δ and the oxidative stress mediator 4-hydroxynonenal, an endogenous ligand of PPAR β/δ , were increased in the damaged *substantia nigra* [47]. One reason for this discrepancy in the involvement of PPAR β/δ in PD may be due to the use of different animal models of PD. Particularly, distinct 6-OHDA models of PD have been developed by injecting the toxin into different parts of the nigrostriatal pathway to induce degeneration of dopaminergic neurons,

which is a hallmark characteristic of PD. Although injection of 6-OHDA into the *substantia nigra pars compacta* leads to more specific and moderate DA depletion, subsequent evaluation of DA depletion and behavioral deficits that mimic PD indicated that specific DA depletion in the subregions of the caudate–putamen complex was the most appropriate model for studying the early and late stages of PD [315]. Clearly, further mechanistic investigation of the involvement of PPAR β/δ in the development and progression of PD is required.

To date, the biological roles of PPAR β/δ have been attributed to its mechanistic functions as a ligand-activated transcriptional factor (see **Introduction**). Transrepression, involving the interaction of PPAR β/δ with the transcriptional repressor BCL-6, is thought to underlie the anti-inflammatory effects of PPAR β/δ . Recently, PPAR β/δ was implicated in Huntington's disease (HD). HD is an autosomal dominant neurodegenerative disorder that causes the progressive breakdown of neurons in the brain, affecting muscle coordination and resulting in mental decline. HD is caused by a CAG trinucleotide repeat expansion in the huntingtin (HTT) gene, which encodes a polyglutamine tract in the HTT protein. Notably, PPAR β/δ can directly interact with the HTT protein, and mutant HTT represses PPAR β/δ -mediated transactivation [316]. Consistent with a neuroprotective role of PPAR β/δ , increased PPAR β/δ activity is associated with reduced mitochondrial dysfunction and increased survival of neurons in mouse models of HD. Neuron-specific expression of the dominant-negative mutant PPAR $\beta/\delta^{\text{E411P}}$ (Nestin-Cre:PPAR $\beta/\delta^{\text{E411P}}$) protein in mice results in a markedly reduced adult brain size and a decreased neuronal count compared with the wild-type genotype. PPAR $\beta/\delta^{\text{E411P}}$ expression in the CNS of mice was sufficient to induce motor dysfunction, neurodegeneration, mitochondrial abnormalities and transcriptional alterations that recapitulated HD-like phenotypes. In mouse models of HD, pharmacologic activation of PPAR β/δ using the agonist KD3010 has been shown to improve

motor function, reduce neurodegeneration and increase survival. Thus, these observations suggest that PPAR β/δ activation may be beneficial in HD and related disorders [316].

All the above studies using *in vivo* models underscore the relevance of the biological functions of PPAR β/δ to the development of therapies for multiple neurodegenerative diseases and injuries. Additionally, further investigation of the mechanisms of PPAR β/δ function and the use of other experimental animal models of disease are clearly needed to realize the therapeutic potential of PPAR β/δ modulators.

10. Conclusions

Since the discovery of PPAR β/δ in 1992 [2,39], its wide expression along with its promiscuous ligand binding capability has hampered our understanding of the pathophysiological roles of this nuclear receptor. Over the past two decades, numerous studies have revealed pleotropic and unexpected roles of PPAR β/δ in various diseases, underscoring PPAR β/δ as a potential therapeutic target. As summarized in this review, PPAR β/δ has been implicated in various physiological and pathophysiological conditions. It is clear that PPAR β/δ , similar to the other two PPARs, regulates numerous cellular and physiological functions. As a ligand-activated transcription factor, PPAR β/δ is capable of altering gene expression in response to the metabolic status of the cellular environment. This function enables PPAR β/δ to be involved in the coordination of tissue development, tissue repair, and metabolism-related conditions. Global knockout animal models have greatly increased our understanding of the biological roles of PPAR β/δ under normal physiological conditions and in the context of diseases.

Looking forward, it is clear that much more information is needed to understand the molecular mechanisms by which PPAR β/δ participates in the etiology of various diseases. In

many of the processes summarized in this review, the mechanism of PPAR β/δ function remains unclear. Although PPAR β/δ primarily regulates gene expression in a ligand-dependent manner, increasing evidence suggests additional mechanisms of PPAR β/δ function, such as transrepression. Mechanistically, the activity of PPAR β/δ involves dynamic switching between co-activator and corepressor molecules in the presence or absence of its cognate ligands. A major knowledge gap concerning PPAR β/δ function is our lack of understanding of the nature of endogenous PPAR β/δ ligands, including their origin, types and levels, at sites of pathology. Despite the diverse nature and size of PPAR β/δ ligands (agonists and antagonists), the transcriptional activation or repression activity of PPAR β/δ may depend on critical interactions between its ligands and specific amino side chains of its LBD, which subsequently facilitate the recruitment of coregulators. To date, no crystal structure of the entire monomeric PPAR β/δ protein or its heterodimer with an RXR has been reported. With advances in protein expression, instrumentation and methods for protein structure resolution, such as single-particle cryoelectron microscopy [317–320], we envisage that future effort may be directed to solve the protein structure of nuclear hormone receptors, a clinically important group of transcription factors. Moreover, new computational tools may be useful in the search for natural and synthetic agonists of PPAR β/δ . Guided by 16 crystal structures of synthetic PPAR β/δ ligands bound to the PPAR β/δ LBD, Kahremany et al. (2015) recently created an *in silico* model that helped in analyzing the binding pocket in the LBD and in determining the obligatory interactions between various synthetic PPAR β/δ agonists and amino acid side chains in the LBD of PPAR β/δ [321]. This model also tests the assumption that both large (e.g., long FAs and their metabolites) and small PPAR β/δ ligands (e.g., hydroxyalkenals and RA) may enter and interact with the bulky cavity in the LBD of PPAR β/δ . Although *in vitro* investigations have suggested that unliganded

PPAR β/δ is bound to chromatin, whether this interaction occurs *in vivo* remains unclear. Importantly, the transcriptional potential of unliganded PPAR β/δ , if present on chromatin, and the effect of the summation of the post-translational modifications of PPAR β/δ on its transactivation activity remain to be determined.

A theme that has continually surfaced in this review is the effect of different PPAR β/δ treatment regimens on disease prognosis. In diabetic wound healing, early controlled release of the PPAR β/δ agonist GW501516 influenced the oxidative wound microenvironment and consequently impacted the wound healing rate. Similarly, in EAE, an experimental model of MS, improved clinical outcomes were observed when oral treatment of the PPAR β/δ agonist GW0742 was administered during disease progression compared with upon disease initiation. Differential effects of PPAR β/δ agonists on the clearance of A β peptide, a pathological hallmark of AD, were also reported. From a different perspective, determining the effects of PPAR β/δ agonists in a particular disease requires us to understand the dynamic host response and the pharmacokinetic profile of the agonist. For instance, the relative PPAR β/δ expression pattern in the major cellular components and the relative contributions of cellular and non-cellular PPAR β/δ to the initiation, development and progression of disease are uncertain. Although recent studies have begun to explore the function of PPAR β/δ during wound healing and atherosclerosis, relatively little is known about the roles of PPAR β/δ in other diseases. In addition to a better understanding of PPAR β/δ in disease development, one will need to appreciate the pharmacokinetic profile, including the bioavailability and stability, of the various PPAR β/δ drugs. Along these lines, it is noteworthy that very few studies exploring various controlled drug delivery systems have focused on our understanding of the PPAR β/δ expression profile and PPAR β/δ drug pharmacokinetics.

The role of PPAR β/δ has been well established for some conditions, such as wound healing and lipid metabolism, while its role in others conditions, such as cancer and neurological diseases, remains controversial. Currently published studies have used different mouse models (e.g., PPAR $\beta/\delta^{\text{ex4}}$ or PPAR $\beta/\delta^{\text{ex8}}$) and experimental regimes, which may confound our understanding of the role of PPAR β/δ . We do not believe that it is currently possible to declare any of these models to be most relevant, as each model fails to faithfully recapitulate all aspects of human disease development, particularly tumorigenesis. For example, *APC^{min}* mice, the most widely used mouse model of colon cancer, only harbor polyps that do not progress to colon tumors. Furthermore, most studies did not examine the role of PPAR β/δ in tumor stromal cells. The tumor microenvironment contains numerous different cell types, such as the predominant cancer-associated fibroblasts (CAFs), tumor associated macrophages (TAMs), tumor-infiltrating lymphocytes (TILs) and pericytes. Despite the well-established impact of these cell types on tumorigenesis, the role of PPAR β/δ in these tumor-associated stromal cells remains unclear and conceivably may be specific to the tumor type, tissue, and cancer stage. Nonetheless, knowledge gathered from all disease models has been useful to some extent, in that the results provide mechanistic insights that have collectively advanced our understanding of different aspects of tumorigenesis. Combining the results from different murine models with data obtained from human tumors will help clarify the role of PPAR β/δ and its associated machinery in CRC and other cancers.

We predict that the use of genetic and pharmacological approaches, such as tissue- and/or cell-type specific conditional knockout animal models along with specific PPAR β/δ agonists and antagonists, will prove to be useful in clarifying the role of PPAR β/δ in various diseases. Such approaches may also reveal novel roles of PPAR β/δ in the initiation and progression of particular

diseases, and such studies could aid in the future design of novel and efficient pharmacological agents that target PPAR β/δ for the treatment of many diseases. Realizing this goal will require multidisciplinary collaborative efforts of various scientists and clinicians.

ACCEPTED MANUSCRIPT

Acknowledgements

The authors thank Eeswari Paramalingam for her help in preparing the figures. The research in the authors' laboratories described in this review was supported by grants from the organizations indicated below. NST received grants from the A*STAR-NTU-NHG Skin Research Grant (SRG/14003), the Academic Research Council Tier 2 of the Singapore Ministry of Education (MOE2014-T2-1-012), and the National Medical Research Council of the Singapore Ministry of Health (NMRC/1280/2010). MV-C received grants from the *Ministerio de Economía y Competitividad* of the Spanish Government [SAF2012-30708 and SAF2015-64146-R] and *CIBER de Diabetes y Enfermedades Metabólicas Asociadas* (CIBERDEM). The research of AM and HG was supported by grants from the Région Midi-Pyrénées, ANR and Société Francophone du Diabète. WW received a Start-Up Grant from the Lee Kong Chian School of Medicine, Nanyang Technological University, the Swiss National Science Foundation, the Human Frontier Science Program, the EUMORPHIA programs, the European Mouse Disease Clinic, TORNADO, the Région Midi-Pyrénées (Chaire d'Excellence Pierre de Fermat), the Bonizzi-Theler-Stiftung, and the Etat de Vaud.

CIBERDEM is an initiative of the *Instituto de Salud Carlos III (ISCIII) - Ministerio de Economía y Competitividad*.

References

- [1] I. Issemann, S. Green, Activation of a member of the steroid hormone receptor superfamily by peroxisome proliferators, *Nature* 347 (1990) 645–650.
- [2] C. Dreyer, G. Krey, H. Keller, F. Givel, G. Helftenbein, W. Wahli, Control of the peroxisomal β -oxidation pathway by a novel family of nuclear hormone receptors, *Cell* 68 (1992) 879–887.
- [3] S.A. Kliewer, B.M. Forman, B. Blumberg, E.S. Ong, U. Borgmeyer, D.J. Mangelsdorf, K. Umesono, R.M. Evans, Differential expression and activation of a family of murine peroxisome proliferator-activated receptors, *Proc. Natl. Acad. Sci. U S A* 91 (1994) 7355–7359.
- [4] W. Wahli, L. Michalik, PPARs at the crossroads of lipid signaling and inflammation, *Trends Endocrinol. Metab.* 23 (2012) 351–363.
- [5] B. Desvergne, W. Wahli, Peroxisome proliferator-activated receptors: nuclear control of metabolism, *Endocr. Rev.* 20 (1999) 649–688.
- [6] F. Lalloyer, B. Staels, Fibrates, glitazones, and peroxisome proliferator-activated receptors, *Arterioscler. Thromb. Vasc. Biol.* 30 (2010) 894–899.
- [7] R.E. Soccio, E.R. Chen, M.A. Lazar, Thiazolidinediones and the promise of insulin sensitization in type 2 diabetes, *Cell Metab.* 20 (2014) 573–591.
- [8] D.L. Sprecher, C. Massien, G. Pearce, A.N. Billin, I. Perlstein, T.M. Willson, D.G. Hassall, N. Ancellin, S.D. Patterson, D.C. Lobe, T.G. Johnson, Triglyceride:high-density lipoprotein cholesterol effects in healthy subjects administered a peroxisome proliferator activated receptor agonist, *Arterioscler. Thromb. Vasc. Biol.* 27 (2007) 359–365.

- [9] U. Riserus, D. Sprecher, T. Johnson, E. Olson, S. Hirschberg, A. Liu, Z. Fang, P. Hegde, D. Richards, L. Sarov-Blat, J.C. Strum, S. Basu, J. Cheeseman, B.A. Fielding, S.M. Humphreys, T. Danoff, N.R. Moore, P. Murgatroyd, S. O'Rahilly, P. Sutton, T. Willson, D. Hassall, K.N. Frayn, F. Karpe, Activation of peroxisome proliferator-activated receptor (PPAR) promotes reversal of multiple metabolic abnormalities, reduces oxidative stress, and increases fatty acid oxidation in moderately obese men, *Diabetes* 57 (2008) 332–339.
- [10] H. Higashiyama, A.N. Billin, Y. Okamoto, M. Kinoshita, S. Asano, Expression profiling of peroxisome proliferator-activated receptor-delta (PPAR-delta) in mouse tissues using tissue microarray, *Histochem. Cell Biol.* 127 (2007) 485–494.
- [11] O. Braissant, F. Fougelle, C. Scotto, M. Dauça, W. Wahli, Differential expression of peroxisome proliferator-activated receptors (PPARs): tissue distribution of PPAR-alpha, -beta, and -gamma in the adult rat, *Endocrinology* 137 (1996) 354–366.
- [12] A. Yessoufou, W. Wahli, Multifaceted roles of peroxisome proliferator-activated receptors (PPARs) at the cellular and whole organism levels, *Swiss. Med. Wkly.* 140 (2010) w13071.
- [13] R. Sever, C.K. Glass, Signaling by nuclear receptors, *Cold Spring Harb. Perspect. Biol.* 5 (2013) a016709.
- [14] C. Wolfrum, C.M. Borrmann, T. Borchers, F. Spener, Fatty acids and hypolipidemic drugs regulate peroxisome proliferator-activated receptors alpha - and gamma-mediated gene expression via liver fatty acid binding protein: a signaling path to the nucleus, *Proc. Natl. Acad. Sci. U S A* 98 (2001) 2323–2328.

- [15] N.S. Tan, N.S. Shaw, N. Vinckenbosch, P. Liu, R. Yasmin, B. Desvergne, W. Wahli, N. Noy, Selective cooperation between fatty acid binding proteins and peroxisome proliferator-activated receptors in regulating transcription, *Mol. Cell. Biol.* 22 (2002) 5114–5127.
- [16] R.L. Smathers, D.R. Petersen, The human fatty acid-binding protein family: evolutionary divergences and functions, *Human Genom.* 5 (2011) 170–191.
- [17] T.T. Schug, D.C. Berry, N.S. Shaw, S.N. Travis, N. Noy, Opposing effects of retinoic acid on cell growth result from alternate activation of two different nuclear receptors, *Cell* 129 (2007) 723–733.
- [18] Y. Bando, M. Yamamoto, K. Sakiyama, K. Inoue, S. Takizawa, Y. Owada, S. Iseki, H. Kondo, O. Amano, Expression of epidermal fatty acid binding protein (E-FABP) in septoclasts in the growth plate cartilage of mice, *J. Mol. Histol.* 45 (2014) 507–518.
- [19] S. Yu, L. Levi, G. Casadesus, G. Kunos, N. Noy, Fatty acid-binding protein 5 (FABP5) regulates cognitive function both by decreasing anandamide levels and by activating the nuclear receptor peroxisome proliferator-activated receptor β/δ (PPAR β/δ) in the brain, *J. Biol. Chem.* 289 (2014) 12748–12758.
- [20] E. Morgan, P. Kannan-Thulasiraman, N. Noy, Involvement of fatty acid binding protein 5 and PPAR β/δ in prostate cancer cell growth, *PPAR Res.* 2010 (2010) 1–9.
- [21] N.-N. Chen, Y. Li, M.-L. Wu, Z.-L. Liu, Y.-S. Fu, Q.-Y. Kong, X.-Y. Chen, H. Li, J. Liu, CRABP-II- and FABP5-independent all-trans retinoic acid resistance in COLO 16 human cutaneous squamous cancer cells, *Exp. Dermatol.* 21 (2012) 13–18.

- [22] K. Maeda, K.T. Uysal, L. Makowski, C.Z. Gorgun, G. Atsumi, R.A. Parker, J. Bruning, A.V. Hertzler, D.A. Bernlohr, G.S. Hotamisligil, Role of the fatty acid binding protein *mab1* in obesity and insulin resistance, *Diabetes* 52 (2003) 300–307.
- [23] D.C. Berry, N. Noy, All-trans-retinoic acid represses obesity and insulin resistance by activating both peroxisome proliferation-activated receptor beta/delta and retinoic acid receptor, *Mol. Cell. Biol.* 29 (2009) 3286–3296.
- [24] Y. Shi, M. Hon, R.M. Evans, The peroxisome proliferator-activated receptor delta, an integrator of transcriptional repression and nuclear receptor signaling, *Proc. Natl. Acad. Sci. U S A* 99 (2002) 2613–2618.
- [25] A.-M. Krogsdam, C.A.F. Nielsen, S. Neve, D. Holst, T. Helledie, B. Thomsen, C. Bendixen, S. Mandrup, K. Kristiansen, Nuclear receptor corepressor-dependent repression of peroxisome-proliferator-activated receptor δ -mediated transactivation, *Biochem. J.* 363 (2002) 157–165.
- [26] L. Aarenstrup, E.N. Flindt, K. Otkjaer, M. Kirkegaard, J.S. Andersen, K. Kristiansen, HDAC activity is required for p65/RelA-dependent repression of PPAR δ -mediated transactivation in human keratinocytes, *J. Investig. Dermatol.* 128 (2008) 1095–1106.
- [27] P. Germain, W. Bourguet, Dimerization of nuclear receptors, *Methods Cell Biol.* 117 (2013) 21–41.
- [28] Y. Fan, Y. Wang, Z. Tang, H. Zhang, X. Qin, Y. Zhu, Y. Guan, X. Wang, B. Staels, S. Chien, N. Wang, Suppression of pro-inflammatory adhesion molecules by PPAR-delta in human vascular endothelial cells, *Arterioscler. Thromb. Vasc. Biol.* 28 (2008) 315–321.
- [29] Y. Takata, J. Liu, F. Yin, A.R. Collins, C.J. Lyon, C.H. Lee, A.R. Atkins, M. Downes, G.D. Barish, R.M. Evans, W.A. Hsueh, R.K. Tangirala, PPARdelta-mediated

- antiinflammatory mechanisms inhibit angiotensin II-accelerated atherosclerosis, *Proc. Natl. Acad. Sci. U S A* 105 (2008) 4277–4282.
- [30] L. Piqueras, A.R. Reynolds, K.M. Hodivala-Dilke, A. Alfranca, J.M. Redondo, T. Hatae, T. Tanabe, T.D. Warner, D. Bishop-Bailey, Activation of PPARbeta/delta induces endothelial cell proliferation and angiogenesis, *Arterioscler. Thromb. Vasc. Biol.* 27 (2007) 63–69.
- [31] V. Perissi, K. Jepsen, C.K. Glass, M.G. Rosenfeld, Deconstructing repression: evolving models of co-repressor action, *Nat. Rev. Genet.* 11 (2010) 109–123.
- [32] L. Mouchiroud, L.J. Eichner, R.J. Shaw, J. Auwerx, Transcriptional coregulators: fine-tuning metabolism, *Cell Metab.* 20 (2014) 26–40.
- [33] J. Zhang, M. Fu, X. Zhu, Y. Xiao, Y. Mou, H. Zheng, M.A. Akinbami, Q. Wang, Y.E. Chen, Peroxisome proliferator-activated receptor delta is up-regulated during vascular lesion formation and promotes post-confluent cell proliferation in vascular smooth muscle cells, *J. Biol. Chem.* 277 (2002) 11505–11512.
- [34] L. Ravaux, C. Denoyelle, C. Monne, I. Limon, M. Raymondjean, K. El Hadri, Inhibition of interleukin-1beta-induced group IIA secretory phospholipase A2 expression by peroxisome proliferator-activated receptors (PPARs) in rat vascular smooth muscle cells: cooperation between PPARbeta and the proto-oncogene BCL-6, *Mol. Cell. Biol.* 27 (2007) 8374–8387.
- [35] C.H. Lee, A. Chawla, N. Urbiztondo, D. Liao, W.A. Boisvert, R.M. Evans, L.K. Curtiss, Transcriptional repression of atherogenic inflammation: modulation by PPAR delta, *Science* 302 (2003) 453–457.

- [36] Y. Matsushita, D. Ogawa, J. Wada, N. Yamamoto, K. Shikata, C. Sato, H. Tachibana, N. Toyota, H. Makino, Activation of peroxisome proliferator-activated receptor δ inhibits streptozotocin-induced diabetic nephropathy through anti-inflammatory mechanisms in mice, *Diabetes* 60 (2011) 960–968.
- [37] N.S. Tan, G. Icre, A. Montagner, B.B.T. Heggeler, W. Wahli, L. Michalik, The nuclear hormone receptor peroxisome proliferator-activated receptor beta/delta potentiates cell chemotactism, polarization, and migration, *Mol. Cell. Biol.* 27 (2007) 7161–7175.
- [38] N. Rotman, L. Michalik, B. Desvergne, W. Wahli, PPARs in fetal and early postnatal development, in: R. Taneja (Ed.), *Nuclear Receptors in Development*, Elsevier, Amsterdam, 2006, pp. 430.
- [39] G. Krey, H. Keller, A. Mahfoudi, J. Medin, K. Ozato, C. Dreyer, W. Wahli, Xenopus peroxisome proliferator activated receptors: genomic organization, response element recognition, heterodimer formation with retinoid X receptor and activation by fatty acids, *J. Steroid Biochem. Mol. Biol.* 47 (1993) 65–73.
- [40] W. Wahli, P.R. Devchand, A.I. Jpenberg, B. Desvergne, Fatty acids, eicosanoids, and hypolipidemic agents regulate gene expression through direct binding to peroxisome proliferator-activated receptors, *Adv. Exp. Med. Biol.* 447 (1999) 199–209.
- [41] E. Bell, F. Ponthan, C. Whitworth, F. Westermann, H. Thomas, C.P.F. Redfern, Cell survival signalling through PPAR δ and arachidonic acid metabolites in neuroblastoma, *PLoS One* 8 (2013) e68859.
- [42] J. Zhang, X. Liu, X.-B. Xie, X.-C. Cheng, R.-L. Wang, Multitargeted bioactive ligands for PPARs discovered in the last decade, *Chem. Biol. Drug Des.* (2016).

- [43] G. Krey, O. Braissant, F. L'Horset, E. Kalkhoven, M. Perroud, M.G. Parker, W. Wahli, Fatty acids, eicosanoids, and hypolipidemic agents identified as ligands of peroxisome proliferator-activated receptors by coactivator-dependent receptor ligand assay, *Mol. Endocrinol.* 11 (1997) 779–791.
- [44] B. Grygiel-Górniak, Peroxisome proliferator-activated receptors and their ligands: nutritional and clinical implications - a review, *Nutr. J.* 13 (2014) 17.
- [45] Y. Riahi, Y. Sin-Malia, G. Cohen, E. Alpert, A. Gruzman, J. Eckel, B. Staels, M. Guichardant, S. Sasson, The natural protective mechanism against hyperglycemia in vascular endothelial cells: roles of the lipid peroxidation product 4-hydroxydodecadienal and peroxisome proliferator-activated receptor delta, *Diabetes* 59 (2010) 808–818.
- [46] G. Cohen, Y. Riahi, V. Sunda, S. Deplano, C. Chatgililoglu, C. Ferreri, N. Kaiser, S. Sasson, Signaling properties of 4-hydroxyalkenals formed by lipid peroxidation in diabetes, *Free Radic. Biol. Med.* 65 (2013) 978–987.
- [47] J.D. Coleman, K.S. Prabhu, J.T. Thompson, P.S. Reddy, J.M. Peters, B.R. Peterson, C.C. Reddy, J.P. Vanden Heuvel, The oxidative stress mediator 4-hydroxynonenal is an intracellular agonist of the nuclear receptor peroxisome proliferator-activated receptor- β/δ (PPAR β/δ), *Free Radic. Biol. Med.* 42 (2007) 1155–1164.
- [48] N. Shaw, M. Elholm, N. Noy, Retinoic acid is a high affinity selective ligand for the peroxisome proliferator-activated receptor beta/delta, *J. Biol. Chem.* 278 (2003) 41589–41592.
- [49] L. Levi, Z. Wang, M.K. Doud, S.L. Hazen, N. Noy, Saturated fatty acids regulate retinoic acid signalling and suppress tumorigenesis by targeting fatty acid-binding protein 5, *Nat. Commun.* 6 (2015) 8794.

- [50] D. Dong, S.E. Ruuska, D.J. Levinthal, N. Noy, Distinct roles for cellular retinoic acid-binding proteins I and II in regulating signaling by retinoic acid, *J. Biol. Chem.* 274 (1999) 23695–23698.
- [51] A.S. Budhu, N. Noy, Direct channeling of retinoic acid between cellular retinoic acid-binding protein II and retinoic acid receptor sensitizes mammary carcinoma cells to retinoic acid-induced growth arrest, *Mol. Cell. Biol.* 22 (2002) 2632–2641.
- [52] R.J. Sessler, N. Noy, A ligand-activated nuclear localization signal in cellular retinoic acid binding protein-II, *Mol. Cell* 18 (2005) 343–353.
- [53] E.H. Armstrong, D. Goswami, P.R. Griffin, N. Noy, E.A. Ortlund, Structural basis for ligand regulation of the fatty acid-binding protein 5, peroxisome proliferator-activated receptor β/δ (FABP5-PPAR β/δ) signaling pathway, *J. Biol. Chem.* 289 (2014) 14941–14954.
- [54] L.J. Donato, Suppression of mammary carcinoma growth by retinoic acid: proapoptotic genes are targets for retinoic acid receptor and cellular retinoic acid-binding protein II signaling, *Cancer Res.* 65 (2005) 8193–8199.
- [55] L.J. Donato, J.H. Suh, N. Noy, Suppression of mammary carcinoma cell growth by retinoic acid: the cell cycle control gene *Btg2* is a direct target for retinoic acid receptor signaling, *Cancer Res.* 67 (2007) 609–615.
- [56] J. Adamson, E.A. Morgan, C. Beesley, Y. Mei, C.S. Foster, H. Fujii, P.S. Rudland, P.H. Smith, Y. Ke, High-level expression of cutaneous fatty acid-binding protein in prostatic carcinomas and its effect on tumorigenicity, *Oncogene* 22 (2003) 2739–2749.

- [57] K. Kawaguchi, S. Senga, C. Kubota, Y. Kawamura, Y. Ke, H. Fujii, High expression of fatty acid-binding protein 5 promotes cell growth and metastatic potential of colorectal cancer cells, *FEBS Open Bio.* 6 (2016) 190–199.
- [58] D.R. Rhodes, S. Kalyana-Sundaram, V. Mahavisno, R. Varambally, J. Yu, B.B. Briggs, T.R. Barrette, M.J. Anstet, C. Kincaid-Beal, P. Kulkarni, S. Varambally, D. Ghosh, A.M. Chinnaiyan, Oncomine 3.0: genes, pathways, and networks in a collection of 18,000 cancer gene expression profiles, *Neoplasia* 9 (2007) 166–180.
- [59] Compendia goes clinical, *Cancer Disc.* 2 (2012) OF2. doi: 10.1158/2159-8290.CD-NB2012-128.
- [60] T.T. Schug, D.C. Berry, I.A. Toshkov, L. Cheng, A.Y. Nikitin, N. Noy, Overcoming retinoic acid-resistance of mammary carcinomas by diverting retinoic acid from PPARbeta/delta to RAR, *Proc. Natl. Acad. Sci. U S A* 105 (2008) 7546–7551.
- [61] P. Kannan-Thulasiraman, D.D. Seachrist, G.H. Mahabeleshwar, M.K. Jain, N. Noy, Fatty acid-binding protein 5 and PPARbeta/delta are critical mediators of epidermal growth factor receptor-induced carcinoma cell growth, *J. Biol. Chem.* 285 (2010) 19106–19115.
- [62] L. Levi, G. Lobo, M.K. Doud, J. von Lintig, D. Seachrist, G.P. Tochtrop, N. Noy, Genetic ablation of the fatty acid-binding protein FABP5 suppresses HER2-induced mammary tumorigenesis, *Cancer Res.* 73 (2013) 4770–4780.
- [63] M. Rieck, W. Meissner, S. Ries, S. Muller-Brusselbach, R. Muller, Ligand-mediated regulation of peroxisome proliferator-activated receptor (PPAR) beta/delta: a comparative analysis of PPAR-selective agonists and all-trans retinoic acid, *Mol. Pharmacol.* 74 (2008) 1269–1277.

- [64] J.W. Bortolotto, R. Margis, Â.C.B. Ferreira, A.V. Padoin, C.C. Mottin, R.M. Guaragna, Adipose tissue distribution and quantification of PPARbeta/delta and PPARgamma1-3 mRNAs: discordant gene expression in subcutaneous, retroperitoneal and visceral adipose tissue of morbidly obese patients, *Obes. Surg.* 17 (2007) 934–940.
- [65] S. Yu, L. Levi, R. Siegel, N. Noy, Retinoic acid induces neurogenesis by activating both retinoic acid receptors (RARs) and peroxisome proliferator-activated receptor β/δ (PPAR β/δ), *J. Biol. Chem.* 287 (2012) 42195–42205.
- [66] L.S. Mackenzie, L. Lione, Harnessing the benefits of PPAR β/δ agonists, *Life Sci.* 93 (2013) 963–967.
- [67] R.E. Gillilan, S.D. Ayers, N. Noy, Structural basis for activation of fatty acid-binding protein 4, *J. Mol. Biol.* 372 (2007) 1246–1260.
- [68] S.D. Ayers, K.L. Nedrow, R.E. Gillilan, N. Noy, Continuous nucleocytoplasmic shuttling underlies transcriptional activation of PPARgamma by FABP4, *Biochemistry* 46 (2007) 6744–6752.
- [69] M. Schupp, J.C. Curtin, R.J. Kim, A.N. Billin, M.A. Lazar, A widely used retinoic acid receptor antagonist induces peroxisome proliferator-activated receptor-gamma activity, *Mol. Pharmacol.* 71 (2007) 1251–1257.
- [70] N. Rotman, N. Guex, E. Gouranton, W. Wahli, PPAR β interprets a chromatin signature of pluripotency to promote embryonic differentiation at gastrulation, *PLoS One* 8 (2013) e83300.
- [71] Y. Barak, M.C. Nelson, E.S. Ong, Y.Z. Jones, P. Ruiz-Lozano, K.R. Chien, A. Koder, R.M. Evans, PPAR gamma is required for placental, cardiac, and adipose tissue development, *Mol. Cell* 4 (1999) 585–595.

- [72] Y. Barak, D. Liao, W. He, E.S. Ong, M.C. Nelson, J.M. Olefsky, R. Boland, R.M. Evans, Effects of peroxisome proliferator-activated receptor delta on placentation, adiposity, and colorectal cancer, *Proc. Natl. Acad. Sci. U S A* 99 (2002) 303–308.
- [73] K. Nadra, S.I. Anghel, E. Joye, N.S. Tan, S. Basu-Modak, D. Trono, W. Wahli, B. Desvergne, Differentiation of trophoblast giant cells and their metabolic functions are dependent on peroxisome proliferator-activated receptor beta/delta, *Mol. Cell. Biol.* 26 (2006) 3266–3281.
- [74] C. Huin, L. Corriveau, A. Bianchi, J.M. Keller, P. Collet, P. Kremarik-Bouillaud, L. Domenjoud, P. Becuwe, H. Schohn, D. Menard, M. Dauca, Differential expression of peroxisome proliferator-activated receptors (PPARs) in the developing human fetal digestive tract, *J. Histochem. Cytochem.* 48 (2000) 603–611.
- [75] B.D. Abbott, C.R. Wood, A.M. Watkins, K.P. Das, C.S. Lau, Peroxisome proliferator-activated receptors alpha, beta, and gamma mRNA and protein expression in human fetal tissues, *PPAR Res.* 2010 (2010) 1–2.
- [76] O. Braissant, W. Wahli, Differential expression of peroxisome proliferator-activated receptor-alpha, -beta, and -gamma during rat embryonic development, *Endocrinology* 139 (1998) 2748–2754.
- [77] K. Han, H. Song, I. Moon, R. Augustin, K. Moley, M. Rogers, H. Lim, Utilization of DR1 as true RARE in regulating the Ssm, a novel retinoic acid-target gene in the mouse testis, *J. Endocrinol.* 192 (2007) 539–551.
- [78] P.L. Pakrasi, A.K. Jain, Evaluation of cyclooxygenase 2 derived endogenous prostacyclin in mouse preimplantation embryo development *in vitro*, *Life Sci.* 80 (2007) 1503–1507.

- [79] J.C. Huang, W.S.A. Wun, J.S. Goldsby, I.C. Wun, D. Noorhasan, K.K. Wu, Stimulation of embryo hatching and implantation by prostacyclin and peroxisome proliferator-activated receptor delta activation: implication in IVF, *Human Reprod.* 22 (2007) 807–814.
- [80] J.-C. Huang, The role of peroxisome proliferator-activated receptors in the development and physiology of gametes and preimplantation embryos, *PPAR Res.* 2008 (2008) 732303.
- [81] J.M. Peters, S.S.T. Lee, W. Li, J.M. Ward, O. Gavrilova, C. Everett, M.L. Reitman, L.D. Hudson, F.J. Gonzalez, Growth, adipose, brain, and skin alterations resulting from targeted disruption of the mouse peroxisome proliferator-activated receptor beta(delta), *Mol. Cell. Biol.* 20 (2000) 5119–5128.
- [82] L. Michalik, B. Desvergne, N.S. Tan, S. Basu-Modak, P. Escher, J. Rieusset, J.M. Peters, G. Kaya, F.J. Gonzalez, J. Zakany, D. Metzger, P. Chambon, D. Duboule, W. Wahli, Impaired skin wound healing in peroxisome proliferator-activated receptor (PPAR) α and PPAR β mutant mice, *J. Cell. Biol.* 154 (2001) 799–814.
- [83] H. Wang, H. Xie, X. Sun, S. Tranguch, H. Zhang, X. Jia, D. Wang, S.K. Das, B. Desvergne, W. Wahli, R.N. DuBois, S.K. Dey, Stage-specific integration of maternal and embryonic peroxisome proliferator-activated receptor delta signaling is critical to pregnancy success, *J. Biol. Chem.* 282 (2007) 37770–37782.
- [84] J.C. Huang, W.S.A. Wun, J.S. Goldsby, N. Matijevic-Aleksic, K.K. Wu, Cyclooxygenase-2-derived endogenous prostacyclin enhances mouse embryo hatching, *Human Reprod.* 19 (2004) 2900–2906.

- [85] J.C. Huang, J.S. Goldsby, W.S.A. Wun, Prostacyclin enhances the implantation and live birth potentials of mouse embryos, *Human Reprod.* 19 (2004) 1856–1860.
- [86] C. Liu, M. Lee, C. Hsieh, C. Huang, H. Tsao, Y. Hsieh, Prostacyclin enhances mouse embryo development and hatching but not increased embryonic cell number and volume, *Fertil. Steril.* 86 (2006) 1047–1052.
- [87] H.J. Kang, S.J. Hwang, J.A. Yoon, J.H. Jun, H.J. Lim, T.K. Yoon, H. Song, Activation of peroxisome proliferators-activated receptor δ (PPAR δ) promotes blastocyst hatching in mice, *Mol. Human Reprod.* 17 (2011) 653–660.
- [88] K. Nishimura, N. Nakano, V.S. Chowdhury, M. Kaneto, M. Torii, M.-A. Hattori, N. Yamauchi, M. Kawai, Effect of PPAR β/δ agonist on the placentation and embryo-fetal development in rats, *Birth Defects Res. B Dev. Reprod. Toxicol.* 98 (2013) 164–169.
- [89] X. Zuo, Z. Peng, M.J. Moussalli, J.S. Morris, R.R. Broaddus, S.M. Fischer, I. Shureiqi, Targeted genetic disruption of peroxisome proliferator-activated receptor-delta and colonic tumorigenesis, *J. Natl. Cancer Inst.* 101 (2009) 762–767.
- [90] D. Wang, L. Fu, W. Ning, L. Guo, X. Sun, S.K. Dey, R. Chaturvedi, K.T. Wilson, R.N. DuBois, Peroxisome proliferator-activated receptor δ promotes colonic inflammation and tumor growth, *Proc. Natl. Acad. Sci. U S A* 111 (2014) 7084–7089.
- [91] L. Michalik, W. Wahli, Involvement of PPAR nuclear receptors in tissue injury and wound repair, *J. Clin. Investig.* 116 (2006) 598–606.
- [92] P. Martin, R. Nunan, Cellular and molecular mechanisms of repair in acute and chronic wound healing, *Br. J. Dermatol.* 173 (2015) 370–378.
- [93] S.J. Bensinger, P. Tontonoz, Integration of metabolism and inflammation by lipid-activated nuclear receptors, *Nature* 454 (2008) 470–477.

- [94] P.R. Devchand, H. Keller, J.M. Peters, M. Vazquez, F.J. Gonzalez, W. Wahli, The PPAR α -leukotriene B₄ pathway to inflammation control, *Nature* 384 (1996) 39–43.
- [95] D.G. Alleva, E.B. Johnson, F.M. Lio, S.A. Boehme, P.J. Conlon, P.D. Crowe, Regulation of murine macrophage proinflammatory and anti-inflammatory cytokines by ligands for peroxisome proliferator-activated receptor-gamma: counter-regulatory activity by IFN-gamma, *J. Leukoc. Biol.* 71 (2002) 677–685.
- [96] J.S. Welch, M. Ricote, T.E. Akiyama, F.J. Gonzalez, C.K. Glass, PPARgamma and PPARdelta negatively regulate specific subsets of lipopolysaccharide and IFN-gamma target genes in macrophages, *Proc. Natl. Acad. Sci. U S A* 100 (2003) 6712–6717.
- [97] K.S. Kilgore, A.N. Billin, PPARbeta/delta ligands as modulators of the inflammatory response, *Curr. Opin. Investig. Drugs* 9 (2008) 463–469.
- [98] N.S. Tan, L. Michalik, B. Desvergne, W. Wahli, Peroxisome proliferator-activated receptor- β as a target for wound healing drugs, *Expert Opin. Therap. Targ.* 8 (2004) 39–48.
- [99] N.S. Tan, L. Michalik, N. Di-Poi, B. Desvergne, W. Wahli, Critical roles of the nuclear receptor PPAR β (peroxisome-proliferator-activated receptor β) in skin wound healing, *Biochem. Soc. Trans.* 32 (2004) 97–102.
- [100] N. Di-Poi, N.S. Tan, L. Michalik, W. Wahli, B. Desvergne, Antiapoptotic role of PPAR β in keratinocytes via transcriptional control of the Akt1 signaling pathway, *Mol. Cell* 10 (2002) 721–733.
- [101] M.P. Caley, V.L.C. Martins, E.A. O'Toole, Metalloproteinases and Wound Healing, *Adv. Wound Care* 4 (2015) 225–234.

- [102] H.C. Chong, M.J. Tan, V. Philippe, S.H. Tan, C.K. Tan, C.W. Ku, Y.Y. Goh, W. Wahli, L. Michalik, N.S. Tan, Regulation of epithelial–mesenchymal IL-1 signaling by PPAR β/δ is essential for skin homeostasis and wound healing, *J Cell Biol* 184 (2009) 817–831.
- [103] X. Wang, M.K. Sng, S. Foo, H.C. Chong, W.L. Lee, M.B.Y. Tang, K.W. Ng, B. Luo, C. Choong, M.T.C. Wong, B.M.K. Tong, S. Chiba, S.C.J. Loo, P. Zhu, N.S. Tan, Early controlled release of peroxisome proliferator-activated receptor β/δ agonist GW501516 improves diabetic wound healing through redox modulation of wound microenvironment, *J. Control. Release* 197 (2015) 138–147.
- [104] S.A. Ham, T. Yoo, J.S. Hwang, E.S. Kang, K.S. Paek, C. Park, J.-H. Kim, J.T. Do, H.G. Seo, Peroxisome proliferator-activated receptor δ modulates MMP-2 secretion and elastin expression in human dermal fibroblasts exposed to ultraviolet B radiation, *J. Dermatol. Sci.* 76 (2014) 44–50.
- [105] C. Zuo, P. Liang, X. Huang, Role of PPARbeta in fibroblast response to heat injury, *Indian J. Biochem. Biophys.* 49 (2012) 219–227.
- [106] M.S. Wietecha, L.A. DiPietro, Therapeutic approaches to the regulation of wound angiogenesis, *Adv. Wound Care* 2 (2013) 81–86.
- [107] J. Li, P. Li, Y. Zhang, G.-B. Li, F.-T. He, Y.-G. Zhou, K. Yang, S.-S. Dai, Upregulation of ski in fibroblast is implicated in the peroxisome proliferator- activated receptor d-mediated wound healing, *Cell. Physiol. Biochem.* 30 (2012) 1059–1071.
- [108] J. Deheuninck, K. Luo, Ski and SnoN, potent negative regulators of TGF- β signaling, *Cell Res.* 19 (2009) 47–57.
- [109] S.A. Ham, J.S. Hwang, T. Yoo, W.J. Lee, K.S. Paek, J.-W. Oh, C.-K. Park, J.-H. Kim, J.T. Do, J.-H. Kim, H.G. Seo, Ligand-activated PPAR δ upregulates α -smooth muscle

- actin expression in human dermal fibroblasts: a potential role for PPAR δ in wound healing, *J. Dermatol. Sci.* 80 (2015) 186–195.
- [110] A. Martin, M.R. Komada, D.C. Sane, Abnormal angiogenesis in diabetes mellitus, *Med. Res. Rev.* 23 (2003) 117–145.
- [111] N.S. Greaves, K.J. Ashcroft, M. Baguneid, A. Bayat, Current understanding of molecular and cellular mechanisms in fibroplasia and angiogenesis during acute wound healing, *J. Dermatol. Sci.* 72 (2013) 206–217.
- [112] S. Kersten, S. Mandard, N.S. Tan, P. Escher, D. Metzger, P. Chambon, F.J. Gonzalez, B. Desvergne, W. Wahli, Characterization of the fasting-induced adipose factor FIAF, a novel peroxisome proliferator-activated receptor target gene, *J. Biol. Chem.* 275 (2000) 28488–28493.
- [113] K.A. Ekmektzoglou, G.C. Zografos, A concomitant review of the effects of diabetes mellitus and hypothyroidism in wound healing, *World J. Gastroenterol.* 12 (2006) 2721–2729.
- [114] P. Zhu, Yan Y. Goh, Hwee Fang A. Chin, S. Kersten, Nguan S. Tan, Angiopoietin-like 4: a decade of research, *Biosci. Rep.* 32 (2012) 211–219.
- [115] R.L. Huang, Z. Teo, H.C. Chong, P. Zhu, M.J. Tan, C.K. Tan, C.R.I. Lam, M.K. Sng, D.T.W. Leong, S.M. Tan, S. Kersten, J.L. Ding, H.Y. Li, N.S. Tan, ANGPTL4 modulates vascular junction integrity by integrin signaling and disruption of intercellular VE-cadherin and claudin-5 clusters, *Blood* 118 (2011) 3990–4002.
- [116] H.C. Chong, J.S.K. Chan, C.Q. Goh, N.V. Gounko, B. Luo, X. Wang, S. Foo, M.T.C. Wong, C. Choong, S. Kersten, N.S. Tan, Angiopoietin-like 4 stimulates STAT3-mediated

- iNOS expression and enhances angiogenesis to accelerate wound healing in diabetic mice, *Mol. Ther.* 22 (2014) 1593–1604.
- [117] Y.Y. Goh, M. Pal, H.C. Chong, P. Zhu, M.J. Tan, L. Punugu, C.K. Tan, R.L. Huang, S.K. Sze, M.B.Y. Tang, J.L. Ding, S. Kersten, N.S. Tan, Angiopoietin-like 4 interacts with matrix proteins to modulate wound healing, *J. Biol. Chem.* 285 (2010) 32999–33009.
- [118] M. Pal, M.J. Tan, R.-L. Huang, Y.Y. Goh, X.L. Wang, M.B.Y. Tang, N.S. Tan, Angiopoietin-like 4 regulates epidermal differentiation, *PLoS One* 6 (2011) e25377.
- [119] P. Bornstein, E.H. Sage, Matricellular proteins: extracellular modulators of cell function, *Curr. Opin. Cell Biol.* 14 (2002) 608–616.
- [120] Y.Y. Goh, M. Pal, H.C. Chong, P. Zhu, M.J. Tan, L. Punugu, C.R.I. Lam, Y.H. Yau, C.K. Tan, R.-L. Huang, S.M. Tan, M.B.Y. Tang, J.L. Ding, S. Kersten, N.S. Tan, Angiopoietin-like 4 interacts with integrins $\beta 1$ and $\beta 5$ to modulate keratinocyte migration, *Am. J. Pathol.* 177 (2010) 2791–2803.
- [121] J.S. Chan, Z. Teo, M.K. Sng, N.S. Tan, Probing for protein-protein interactions during cell migration: limitations and challenges, *Histol. Histopathol.* 29 (2014) 965–976.
- [122] Y. Gu, X. Li, T. He, Z. Jiang, P. Hao, X. Tang, The antifibrosis effects of peroxisome proliferator-activated receptor δ on rat corneal wound healing after excimer laser keratectomy, *PPAR Res.* 2014 (2014) 464935.
- [123] F. Ali, N.S. Ali, A. Bauer, J.J. Boyle, S.S. Hamdulay, D.O. Haskard, A.M. Randi, J.C. Mason, PPAR δ and PGC1 α act cooperatively to induce haem oxygenase-1 and enhance vascular endothelial cell resistance to stress, *Cardiovasc. Res.* 85 (2010) 701–710.

- [124] M. Pang, S.M. de la Monte, L. Longato, M. Tong, J. He, R. Chaudhry, K. Duan, J. Ouh, J.R. Wands, PPAR δ agonist attenuates alcohol-induced hepatic insulin resistance and improves liver injury and repair, *J. Hepatol.* 50 (2009) 1192–1201.
- [125] E. Ehrenborg, J. Skogsberg, Peroxisome proliferator-activated receptor delta and cardiovascular disease, *Atherosclerosis* 231 (2013) 95–106.
- [126] R. Jimenez, M. Sanchez, M.J. Zarzuelo, M. Romero, A.M. Quintela, R. Lopez-Sepulveda, P. Galindo, M. Gomez-Guzman, J.M. Haro, A. Zarzuelo, F. Perez-Vizcaino, J. Duarte, Endothelium-dependent vasodilator effects of peroxisome proliferator-activated receptor beta agonists via the phosphatidyl-inositol-3 kinase-Akt pathway, *J. Pharmacol. Exp. Ther.* 332 (2010) 554–561.
- [127] A.M. Quintela, R. Jiménez, M. Gómez-Guzmán, M.J. Zarzuelo, P. Galindo, M. Sánchez, F. Vargas, Á. Cogolludo, J. Tamargo, F. Pérez-Vizcaíno, J. Duarte, Activation of peroxisome proliferator-activated receptor- β/δ (PPAR β/δ) prevents endothelial dysfunction in type 1 diabetic rats, *Free Radic. Biol. Med.* 53 (2012) 730–741.
- [128] X.Y. Tian, W.T. Wong, N. Wang, Y. Lu, W.S. Cheang, J. Liu, L. Liu, Y. Liu, S.S.T. Lee, Z.Y. Chen, J.P. Cooke, X. Yao, Y. Huang, PPAR δ activation protects endothelial function in diabetic mice, *Diabetes* 61 (2012) 3285–3293.
- [129] M. Toral, M. Romero, R. Jiménez, A.M. Mahmoud, E. Barroso, M. Gómez-Guzmán, M. Sánchez, Á. Cogolludo, A.B. García-Redondo, A.M. Briones, M. Vázquez-Carrera, F. Pérez-Vizcaíno, J. Duarte, Carnitine palmitoyltransferase-1 up-regulation by PPAR- β/δ prevents lipid-induced endothelial dysfunction, *Clin. Sci.* 129 (2015) 823–837.
- [130] M.J. Zarzuelo, R. Jimenez, P. Galindo, M. Sanchez, A. Nieto, M. Romero, A.M. Quintela, R. Lopez-Sepulveda, M. Gomez-Guzman, E. Bailon, I. Rodriguez-Gomez, A. Zarzuelo, J.

- Galvez, J. Tamargo, F. Perez-Vizcaino, J. Duarte, Antihypertensive effects of peroxisome proliferator-activated receptor- β activation in spontaneously hypertensive rats, *Hypertension* 58 (2011) 733–743.
- [131] G. Bansal, K.M. Druey, Z. Xie, R4 RGS proteins: regulation of G-protein signaling and beyond, *Pharmacol. Ther.* 116 (2007) 473–495.
- [132] S.P. Heximer, R.H. Knutsen, X. Sun, K.M. Kaltenbronn, M.-H. Rhee, N. Peng, A. Oliveira-dos-Santos, J.M. Penninger, A.J. Muslin, T.H. Steinberg, J.M. Wyss, R.P. Mecham, K.J. Blumer, Hypertension and prolonged vasoconstrictor signaling in RGS2-deficient mice, *J. Clin. Investig.* 111 (2003) 445–452.
- [133] W.S. Cheang, X.Y. Tian, W.T. Wong, C.W. Lau, S.S.T. Lee, Z.Y. Chen, X. Yao, N. Wang, Y. Huang, Metformin protects endothelial function in diet-induced obese mice by inhibition of endoplasmic reticulum stress through 5' adenosine monophosphate-activated protein kinase-peroxisome proliferator-activated receptor δ pathway, *Arterioscler. Thromb. Vasc. Biol.* 34 (2014) 830–836.
- [134] T.E. Akiyama, G. Lambert, C.J. Nicol, K. Matsusue, J.M. Peters, H.B. Brewer, F.J. Gonzalez, Peroxisome proliferator-activated receptor beta/delta regulates very low density lipoprotein production and catabolism in mice on a Western diet, *J. Biol. Chem.* 279 (2004) 20874–20881.
- [135] M.D. Leibowitz, C. Fiévet, N. Hennuyer, J. Peinado-Onsurbe, H. Duez, J. Berger, C.A. Cullinan, C.P. Sparrow, J. Baffic, G.D. Berger, C. Santini, R.W. Marquis, R.L. Tolman, R.G. Smith, D.E. Moller, J. Auwerx, Activation of PPAR δ alters lipid metabolism in db/db mice, *FEBS Lett.* 473 (2000) 333–336.

- [136] T. Tanaka, J. Yamamoto, S. Iwasaki, H. Asaba, H. Hamura, Y. Ikeda, M. Watanabe, K. Magoori, R.X. Ioka, K. Tachibana, Y. Watanabe, Y. Uchiyama, K. Sumi, H. Iguchi, S. Ito, T. Doi, T. Hamakubo, M. Naito, J. Auwerx, M. Yanagisawa, T. Kodama, J. Sakai, Activation of peroxisome proliferator-activated receptor delta induces fatty acid beta-oxidation in skeletal muscle and attenuates metabolic syndrome, *Proc. Natl. Acad. Sci. U S A* 100 (2003) 15924–15929.
- [137] E. Barroso, R. Rodríguez-Calvo, L. Serrano-Marco, A.M. Astudillo, J. Balsinde, X. Palomer, M. Vázquez-Carrera, The PPAR β/δ activator GW501516 prevents the down-regulation of AMPK Caused by a high-fat diet in liver and amplifies the PGC-1 α -lipin 1-PPAR α pathway leading to increased fatty acid oxidation, *Endocrinology* 152 (2011) 1848–1859.
- [138] L.M. Sanderson, M.V. Boekschoten, B. Desvergne, M. Muller, S. Kersten, Transcriptional profiling reveals divergent roles of PPAR α and PPAR β/δ in regulation of gene expression in mouse liver, *Physiol. Genom.* 41 (2010) 42–52.
- [139] W.R. Oliver, J.L. Shenk, M.R. Snaith, C.S. Russell, K.D. Plunket, N.L. Bodkin, M.C. Lewis, D.A. Winegar, M.L. Sznaidman, M.H. Lambert, H.E. Xu, D.D. Sternbach, S.A. Kliewer, B.C. Hansen, T.M. Willson, A selective peroxisome proliferator-activated receptor delta agonist promotes reverse cholesterol transport, *Proc. Natl. Acad. Sci. U S A* 98 (2001) 5306–5311.
- [140] K. Chehaibi, L. Cedó, J. Metso, X. Palomer, D. Santos, H. Quesada, M. Naceur Slimane, W. Wahli, J. Julve, M. Vázquez-Carrera, M. Jauhiainen, F. Blanco-Vaca, J.C. Escolà-Gil, PPAR- β/δ activation promotes phospholipid transfer protein expression, *Biochem. Pharmacol.* 94 (2015) 101–108.

- [141] J.N. van der Veen, J.K. Kruit, R. Havinga, J.F. Baller, G. Chimini, S. Lestavel, B. Staels, P.H. Groot, A.K. Groen, F. Kuipers, Reduced cholesterol absorption upon PPARdelta activation coincides with decreased intestinal expression of NPC1L1, *J. Lipid Res.* 46 (2005) 526–534.
- [142] C.L.J. Vrans, A.E. van der Velde, K. van den Oever, J.H.M. Levels, S. Huet, R.P.J. Oude Elferink, F. Kuipers, A.K. Groen, Peroxisome proliferator-activated receptor delta activation leads to increased transintestinal cholesterol efflux, *J. Lipid Res.* 50 (2009) 2046–2054.
- [143] E.M.M. Ooi, G.F. Watts, D.L. Sprecher, D.C. Chan, P.H.R. Barrett, Mechanism of action of a peroxisome proliferator-activated receptor (PPAR)- δ agonist on lipoprotein metabolism in dyslipidemic subjects with central obesity, *J. Clin. Endocrinol. Metab.* 96 (2011) E1568–E1576.
- [144] E.J. Olson, G.L. Pearce, N.P. Jones, D.L. Sprecher, Lipid effects of peroxisome proliferator-activated receptor- δ agonist GW501516 in subjects with low high-density lipoprotein cholesterol: characteristics of metabolic syndrome, *Arterioscler. Thromb. Vasc. Biol.* 32 (2012) 2289–2294.
- [145] Y.-J. Choi, B.K. Roberts, X. Wang, J.C. Geaney, S. Naim, K. Wojnoonski, D.B. Karpf, R.M. Krauss, Effects of the PPAR- δ agonist MBX-8025 on atherogenic dyslipidemia, *Atherosclerosis* 220 (2012) 470–476.
- [146] H.E. Bays, S. Schwartz, T. Littlejohn, B. Kerzner, R.M. Krauss, D.B. Karpf, Y.-J. Choi, X. Wang, S. Naim, B.K. Roberts, MBX-8025, a novel peroxisome proliferator receptor-delta agonist: lipid and other metabolic effects in dyslipidemic overweight patients treated with and without atorvastatin, *J. Clin. Endocrinol. Metab.* 96 (2011) 2889–2897.

- [147] T.L. Graham, C. Mookherjee, K.E. Suckling, C.N. Palmer, L. Patel, The PPARdelta agonist GW0742X reduces atherosclerosis in LDLR(-/-) mice, *Atherosclerosis* 181 (2005) 29–37.
- [148] G.D. Barish, A.R. Atkins, M. Downes, P. Olson, L.W. Chong, M. Nelson, Y. Zou, H. Hwang, H. Kang, L. Curtiss, R.M. Evans, C.H. Lee, PPARdelta regulates multiple proinflammatory pathways to suppress atherosclerosis, *Proc. Natl. Acad. Sci. U S A* 105 (2008) 4271–4276.
- [149] L.A. Bojic, A.C. Burke, S.S. Chhoker, D.E. Telford, B.G. Sutherland, J.Y. Edwards, C.G. Sawyez, R.G. Tirona, H. Yin, J.G. Pickering, M.W. Huff, Peroxisome proliferator-activated receptor δ agonist GW1516 attenuates diet-induced aortic inflammation, insulin resistance, and atherosclerosis in low-density lipoprotein receptor knockout mice, *Arterioscler. Thromb. Vasc. Biol.* 34 (2014) 52–60.
- [150] A.C. Li, C.J. Binder, A. Gutierrez, K.K. Brown, C.R. Plotkin, J.W. Pattison, A.F. Valledor, R.A. Davis, T.M. Willson, J.L. Witztum, W. Palinski, C.K. Glass, Differential inhibition of macrophage foam-cell formation and atherosclerosis in mice by PPAR α , β/δ , and γ , *J. Clin. Investig.* 114 (2004) 1564–1576.
- [151] L. Cheng, G. Ding, Q. Qin, Y. Huang, W. Lewis, N. He, R.M. Evans, M.D. Schneider, F.A. Brako, Y. Xiao, Y.E. Chen, Q. Yang, Cardiomyocyte-restricted peroxisome proliferator-activated receptor- δ deletion perturbs myocardial fatty acid oxidation and leads to cardiomyopathy, *Nat. Med.* 10 (2004) 1245–1250.
- [152] P. Wang, J. Liu, Y. Li, S. Wu, J. Luo, H. Yang, R. Subbiah, J. Chatham, O. Zhelyabovska, Q. Yang, Peroxisome proliferator-activated receptor δ is an essential

- transcriptional regulator for mitochondrial protection and biogenesis in adult heart, *Circ. Res.* 106 (2010) 911–919.
- [153] E.M. Burkart, N. Sambandam, X. Han, R.W. Gross, M. Courtois, C.M. Gierasch, K. Shoghi, M.J. Welch, D.P. Kelly, Nuclear receptors PPAR β/δ and PPAR α direct distinct metabolic regulatory programs in the mouse heart, *J. Clin. Investig.* 117 (2007) 3930–3939.
- [154] J. Liu, P. Wang, J. Luo, Y. Huang, L. He, H. Yang, Q. Li, S. Wu, O. Zhelyabovska, Q. Yang, Peroxisome proliferator-activated receptor β/δ activation in adult hearts facilitates mitochondrial function and cardiac performance under pressure-overload condition, *Hypertension* 57 (2011) 223–230.
- [155] T.L. Yue, S.S. Nerurkar, W. Bao, B.M. Jucker, L. Sarov-Blat, K. Steplewski, E.H. Ohlstein, R.N. Willette, *In vivo* activation of peroxisome proliferator-activated receptor-delta protects the heart from ischemia/reperfusion injury in Zucker fatty rats, *J. Pharmacol. Exp. Ther.* 325 (2008) 466–474.
- [156] H. el Azzouzi, S. Leptidis, E. Dirx, J. Hoeks, B. van Bree, K. Brand, E.A. McClellan, E. Poels, J.C. Sluimer, M.M.G. van den Hoogenhof, A.-S. Armand, X. Yin, S. Langley, M. Bourajjaj, S. Olieslagers, J. Krishnan, M. Vooijs, H. Kurihara, A. Stubbs, Y.M. Pinto, W. Krek, M. Mayr, P.A. da C. Martins, P. Schrauwen, L.J. De Windt, The hypoxia-inducible microRNA cluster miR-199a~214 targets myocardial PPAR δ and impairs mitochondrial fatty acid oxidation, *Cell Metab.* 18 (2013) 341–354.
- [157] B.C. Yu, C.K. Chang, H.Y. Ou, K.C. Cheng, J.T. Cheng, Decrease of peroxisome proliferator-activated receptor delta expression in cardiomyopathy of streptozotocin-induced diabetic rats, *Cardiovasc. Res.* 80 (2008) 78–87.

- [158] D. Álvarez-Guardia, X. Palomer, T. Coll, L. Serrano, R. Rodríguez-Calvo, M.M. Davidson, M. Merlos, I. El Kochairi, L. Michalik, W. Wahli, M. Vázquez-Carrera, PPAR β/δ activation blocks lipid-induced inflammatory pathways in mouse heart and human cardiac cells, *Biochim. Biophys. Acta* 1811 (2011) 59–67.
- [159] X. Palomer, E. Capdevila-Busquets, G. Botteri, L. Salvadó, E. Barroso, M.M. Davidson, L. Michalik, W. Wahli, M. Vázquez-Carrera, PPAR β/δ attenuates palmitate-induced endoplasmic reticulum stress and induces autophagic markers in human cardiac cells, *Int. J. Cardiol.* 174 (2014) 110–118.
- [160] J.R. Park, J.H. Ahn, M.H. Jung, J.-S. Koh, Y. Park, S.-J. Hwang, Y.-H. Jeong, C.H. Kwak, Y.S. Lee, H.G. Seo, J.H. Kim, J.-Y. Hwang, Effects of peroxisome proliferator-activated receptor- δ agonist on cardiac healing after myocardial infarction, *PLoS One* 11 (2016) e0148510.
- [161] C.K. Glass, J.M. Olefsky, Inflammation and lipid signaling in the etiology of insulin resistance, *Cell Metab.* 15 (2012) 635–645.
- [162] K. Kang, S.M. Reilly, V. Karabacak, M.R. Gangl, K. Fitzgerald, B. Hatano, C.-H. Lee, Adipocyte-derived Th2 cytokines and myeloid PPAR δ regulate macrophage polarization and insulin sensitivity, *Cell Metab.* 7 (2008) 485–495.
- [163] E. Barroso, R. Rodríguez-Rodríguez, M.R. Chacón, E. Maymó-Masip, L. Ferrer, L. Salvadó, E. Salmerón, M. Wabistch, X. Palomer, J. Vendrell, W. Wahli, M. Vázquez-Carrera, PPAR β/δ ameliorates fructose-induced insulin resistance in adipocytes by preventing Nrf2 activation, *Biochim. Biophys. Acta* 1852 (2015) 1049–1058.

- [164] Y.-X. Wang, C.-H. Lee, S. Tiep, R.T. Yu, J. Ham, H. Kang, R.M. Evans, Peroxisome-proliferator-activated receptor δ activates fat metabolism to prevent obesity, *Cell* 113 (2003) 159–170.
- [165] L. Serrano-Marco, R. Rodriguez-Calvo, I. El Kochairi, X. Palomer, L. Michalik, W. Wahli, M. Vazquez-Carrera, Activation of peroxisome proliferator-activated receptor- β / δ (PPAR- β / δ) ameliorates insulin signaling and reduces SOCS3 levels by inhibiting STAT3 in interleukin-6-stimulated adipocytes, *Diabetes* 60 (2011) 1990–1999.
- [166] D. Qu, J. Liu, C.W. Lau, Y. Huang, IL-6 in diabetes and cardiovascular complications, *Br. J. Pharmacol.* 171 (2014) 3595–3603.
- [167] Y. Zhao, P. Gao, F. Sun, Q. Li, J. Chen, H. Yu, L. Li, X. Wei, H. He, Z. Lu, X. Wei, B. Wang, Y. Cui, S. Xiong, Q. Shang, A. Xu, Y. Huang, D. Liu, Z. Zhu, Sodium intake regulates glucose homeostasis through the PPAR δ /adiponectin-mediated SGLT2 pathway, *Cell Metab.* 23 (2016) 699–711.
- [168] R.A. DeFronzo, D. Tripathy, Skeletal muscle insulin resistance is the primary defect in type 2 diabetes, *Diabetes Care* 32 (2009) S157-S163.
- [169] S. Luquet, J. Lopez-Soriano, D. Holst, A. Fredenrich, J. Melki, M. Rassoulzadegan, P.A. Grimaldi, Peroxisome proliferator-activated receptor delta controls muscle development and oxidative capability, *FASEB J.* 17 (2003) 2299–2301.
- [170] Z. Gan, J. Rumsey, B.C. Hazen, L. Lai, T.C. Leone, R.B. Vega, H. Xie, K.E. Conley, J. Auwerx, S.R. Smith, E.N. Olson, A. Kralli, D.P. Kelly, Nuclear receptor/microRNA circuitry links muscle fiber type to energy metabolism, *J. Clin. Investig.* 123 (2013) 2564–2575.

- [171] M. Schuler, F. Ali, C. Chambon, D. Duteil, J.-M. Bornert, A. Tardivel, B. Desvergne, W. Wahli, P. Chambon, D. Metzger, PGC1 α expression is controlled in skeletal muscles by PPAR β , whose ablation results in fiber-type switching, obesity, and type 2 diabetes, *Cell Metab.* 4 (2006) 407–414.
- [172] S. Kleiner, V. Nguyen-Tran, O. Bare, X. Huang, B. Spiegelman, Z. Wu, PPAR{delta} agonism activates fatty acid oxidation via PGC-1{alpha} but does not increase mitochondrial gene expression and function, *J. Biol. Chem.* 284 (2009) 18624–18633.
- [173] E. Ehrenborg, A. Krook, Regulation of skeletal muscle physiology and metabolism by peroxisome proliferator-activated receptor delta, *Pharmacol. Rev.* 61 (2009) 373–393.
- [174] S. Liu, J.D. Brown, K.J. Stanya, E. Homan, M. Leidl, K. Inouye, P. Bhargava, M.R. Gangl, L. Dai, B. Hatano, G.S. Hotamisligil, A. Saghatelian, J. Plutzky, C.-H. Lee, A diurnal serum lipid integrates hepatic lipogenesis and peripheral fatty acid use, *Nature* 502 (2013) 550–554.
- [175] L. Salvadó, E. Barroso, A.M. Gómez-Foix, X. Palomer, L. Michalik, W. Wahli, M. Vázquez-Carrera, PPAR β/δ prevents endoplasmic reticulum stress-associated inflammation and insulin resistance in skeletal muscle cells through an AMPK-dependent mechanism, *Diabetologia* 57 (2014) 2126–2135.
- [176] H. Staiger, C. Haas, J. Machann, R. Werner, M. Weisser, F. Schick, F. Machicao, N. Stefan, A. Fritsche, H.U. Haring, Muscle-derived angiotensin-like protein 4 is induced by fatty acids via peroxisome proliferator-activated receptor (PPAR)-delta and is of metabolic relevance in humans, *Diabetes* 58 (2009) 579–589.
- [177] A. Georgiadi, L. Lichtenstein, T. Degenhardt, M.V. Boekschoten, M. van Bilsen, B. Desvergne, M. Muller, S. Kersten, Induction of cardiac Angptl4 by dietary fatty acids is

- mediated by peroxisome proliferator-activated receptor beta/delta and protects against fatty acid-induced oxidative stress, *Circ. Res.* 106 (2010) 1712–1721.
- [178] K. Yoshida, T. Shimizugawa, M. Ono, H. Furukawa, Angiotensin-like protein 4 is a potent hyperlipidemia-inducing factor in mice and inhibitor of lipoprotein lipase, *J. Lipid Res.* 43 (2002) 1770–1772.
- [179] S. Mandard, F. Zandbergen, E. van Straten, W. Wahli, F. Kuipers, M. Muller, S. Kersten, The fasting-induced adipose factor/angiotensin-like protein 4 is physically associated with lipoproteins and governs plasma lipid levels and adiposity, *J. Biol. Chem.* 281 (2006) 934–944.
- [180] W. Dijk, A.P. Beigneux, M. Larsson, A. Bensadoun, S.G. Young, S. Kersten, Angiotensin-like 4 (ANGPTL4) promotes intracellular degradation of lipoprotein lipase in adipocytes, *J. Lipid Res.* (2016) pii: jlr.M067363.
- [181] S.M. Reilly, C.H. Lee, PPAR delta as a therapeutic target in metabolic disease, *FEBS Lett.* 582 (2008) 26–31.
- [182] R. Minnaard, P. Schrauwen, G. Schaart, J.A. Jorgensen, E. Lenaers, M. Mensink, M.K.C. Hesselink, Adipocyte differentiation-related protein and OXPAT in rat and human skeletal muscle: involvement in lipid accumulation and type 2 diabetes mellitus, *J. Clin. Endocrinol. Metab.* 94 (2009) 4077–4085.
- [183] S.A. Phillips, C.C. Choe, T.P. Ciaraldi, A.S. Greenberg, A.P.S. Kong, S.C. Baxi, L. Christiansen, S.R. Mudaliar, R.R. Henry, Adipocyte differentiation-related protein in human skeletal muscle: relationship to insulin sensitivity, *Obes. Res.* 13 (2005) 1321–1329.

- [184] Z. Gan, E.M. Burkart-Hartman, D.H. Han, B. Finck, T.C. Leone, E.Y. Smith, J.E. Ayala, J. Holloszy, D.P. Kelly, The nuclear receptor PPAR β/δ programs muscle glucose metabolism in cooperation with AMPK and MEF2, *Genes Dev.* 25 (2011) 2619–2630.
- [185] E. Benetti, R. Mastrocola, M. Rogazzo, F. Chiazza, M. Aragno, R. Fantozzi, M. Collino, M.A. Minetto, High sugar intake and development of skeletal muscle insulin resistance and inflammation in mice: a protective role for PPAR- δ agonism, *Mediators Inflamm.* 2013 (2013) 509502.
- [186] C. Christodoulides, P. Dyson, D. Sprecher, K. Tsintzas, F. Karpe, Circulating fibroblast growth factor 21 is induced by peroxisome proliferator-activated receptor agonists but not ketosis in man, *J. Clin. Endocrinol. Metab.* 94 (2009) 3594–3601.
- [187] K. Ray, NAFLD—the next global epidemic, *Nat. Rev. Gastroenterol. Hepatol.* 10 (2013) 621.
- [188] G.A. Michelotti, M.V. Machado, A.M. Diehl, NAFLD, NASH and liver cancer, *Nat. Rev. Gastroenterol. Hepatol.* 10 (2013) 656–665.
- [189] S.R. Joshi, Saroglitazar for the treatment of dyslipidemia in diabetic patients, *Expert Opin. Pharmacother.* 16 (2015) 597–606.
- [190] B. Staels, A. Rubenstrunk, B. Noel, G. Rigou, P. Delataille, L.J. Millatt, M. Baron, A. Lucas, A. Tailleux, D.W. Hum, V. Ratziu, B. Cariou, R. Hanf, Hepatoprotective effects of the dual peroxisome proliferator-activated receptor alpha/delta agonist, GFT505, in rodent models of nonalcoholic fatty liver disease/nonalcoholic steatohepatitis, *Hepatology* 58 (2013) 1941–1952.
- [191] V. Ratziu, S.A. Harrison, S. Francque, P. Bedossa, P. Lehert, L. Serfaty, M. Romero-Gomez, J. Boursier, M. Abdelmalek, S. Caldwell, J. Drenth, Q.M. Anstee, D. Hum, R.

- Hanf, A. Roudot, S. Megnien, B. Staels, A. Sanyal, Elafibranor, an Agonist of the Peroxisome proliferator-activated receptor- α and - δ , induces resolution of nonalcoholic steatohepatitis without fibrosis worsening, *Gastroenterology* 150 (2016) 1147–1159.e1145.
- [192] F. Baraille, J. Planchais, R. Dentin, S. Guilmeau, C. Postic, Integration of ChREBP-mediated glucose sensing into whole body metabolism, *Physiology* 30 (2015) 428–437.
- [193] J.Y. Cha, J.J. Repa, The liver X receptor (LXR) and hepatic lipogenesis: the carbohydrate-response element-binding protein is a target gene of LXR, *J. Biol. Chem.* 282 (2007) 743–751.
- [194] H. Guillou, P.G.P. Martin, T. Pineau, Transcriptional regulation of hepatic fatty acid metabolism, *Subcell. Biochem.* 49 (2008) 3–47.
- [195] Y. Wang, J. Viscarra, S.-J. Kim, H.S. Sul, Transcriptional regulation of hepatic lipogenesis, *Nat. Rev. Mol. Cell Biol.* 16 (2015) 678–689.
- [196] P. Ferré, F. Fofelle, Hepatic steatosis: a role for de novo lipogenesis and the transcription factor SREBP-1c, *Diabetes Obes. Metab.* 12 (2010) 83–92.
- [197] D.J. Peet, B.A. Janowski, D.J. Mangelsdorf, The LXRs: a new class of oxysterol receptors, *Curr. Opin. Genet. Dev.* 8 (1998) 571–575.
- [198] J. Xu, M.T. Nakamura, H.P. Cho, S.D. Clarke, Sterol Regulatory Element binding protein-1 expression is suppressed by dietary polyunsaturated fatty acids: a mechanism for the coordinate suppression of lipogenic genes by polyunsaturated fats, *J. Biol. Chem.* 274 (1999) 23577–23583.
- [199] R. Dentin, F. Benhamed, J.P. Pegorier, F. Fofelle, B. Viollet, S. Vaulont, J. Girard, C. Postic, Polyunsaturated fatty acids suppress glycolytic and lipogenic genes through the

- inhibition of ChREBP nuclear protein translocation, *J. Clin. Invest.* 115 (2005) 2843–2854.
- [200] T. Yoshikawa, H. Shimano, N. Yahagi, T. Ide, M. Amemiya-Kudo, T. Matsuzaka, M. Nakakuki, S. Tomita, H. Okazaki, Y. Tamura, Y. Iizuka, K. Ohashi, A. Takahashi, H. Sone, J.-I. Osuga, T. Gotoda, S. Ishibashi, N. Yamada, Polyunsaturated fatty acids suppress sterol regulatory element-binding protein 1c promoter activity by inhibition of liver X receptor (LXR) binding to LXR response elements, *J. Biol. Chem.* 277 (2002) 1705–1711.
- [201] S. Ducheix, A. Montagner, A. Polizzi, F. Lasserre, A. Marmugi, J. Bertrand-Michel, N. Podechard, T. Al Saati, M. Chétiveaux, S. Baron, J. Boué, G. Dietrich, L. Mselli-Lakhal, P. Costet, J.-M.A. Lobaccaro, T. Pineau, V. Theodorou, C. Postic, P.G.P. Martin, H. Guillou, Essential fatty acids deficiency promotes lipogenic gene expression and hepatic steatosis through the liver X receptor, *J. Hepatol.* 58 (2013) 984–992.
- [202] L.M. Sanderson, T. Degenhardt, A. Koppen, E. Kalkhoven, B. Desvergne, M. Muller, S. Kersten, Peroxisome proliferator-activated receptor beta/delta (PPARbeta/delta) but not PPARalpha serves as a plasma free fatty acid sensor in liver, *Mol. Cell. Biol.* 29 (2009) 6257–6267.
- [203] X. Qin, X. Xie, Y. Fan, J. Tian, Y. Guan, X. Wang, Y. Zhu, N. Wang, Peroxisome proliferator-activated receptor-delta induces insulin-induced gene-1 and suppresses hepatic lipogenesis in obese diabetic mice, *Hepatology* 48 (2008) 432–441.
- [204] L.A. Bojic, D.E. Telford, M.D. Fullerton, R.J. Ford, B.G. Sutherland, J.Y. Edwards, C.G. Sawyez, R. Gros, B.E. Kemp, G.R. Steinberg, M.W. Huff, PPAR δ activation attenuates

- hepatic steatosis in *Ldlr*^{-/-} mice by enhanced fat oxidation, reduced lipogenesis, and improved insulin sensitivity, *J. Lipid Res.* 55 (2014) 1254–1266.
- [205] W.G. Garbacz, J.T.J. Huang, L.G. Higgins, W. Wahli, C.N.A. Palmer, PPAR α is required for PPAR δ action in regulation of body weight and hepatic steatosis in mice, *PPAR Res.* 2015 (2015) 927057.
- [206] C.H. Lee, P. Olson, A. Hevener, I. Mehl, L.W. Chong, J.M. Olefsky, F.J. Gonzalez, J. Ham, H. Kang, J.M. Peters, R.M. Evans, PPAR δ regulates glucose metabolism and insulin sensitivity, *Proc. Natl. Acad. Sci. U S A* 103 (2006) 3444–3449.
- [207] S. Liu, B. Hatano, M. Zhao, C.C. Yen, K. Kang, S.M. Reilly, M.R. Gangl, C. Gorgun, J.A. Balschi, J.M. Ntambi, C.H. Lee, Role of peroxisome proliferator-activated receptor $\{\delta\}/\{\beta\}$ in hepatic metabolic regulation, *J. Biol. Chem.* 286 (2011) 1237–1247.
- [208] A. Montagner, A. Polizzi, E. Fouché, S. Ducheix, Y. Lippi, F. Lasserre, V. Barquissau, M. Régnier, C. Lukowicz, F. Benhamed, A. Iroz, J. Bertrand-Michel, T. Al Saati, P. Cano, L. Mselli-Lakhal, G. Mithieux, F. Rajas, S. Lagarrigue, T. Pineau, N. Loiseau, C. Postic, D. Langin, W. Wahli, H. Guillou, Liver PPAR α is crucial for whole-body fatty acid homeostasis and is protective against NAFLD, *Gut* 65 (2016) 1202–1214.
- [209] W. Shan, C.J. Nicol, S. Ito, M.T. Bility, M.J. Kennett, J.M. Ward, F.J. Gonzalez, J.M. Peters, Peroxisome proliferator-activated receptor- β/δ protects against chemically induced liver toxicity in mice, *Hepatology* 47 (2008) 225–235.
- [210] J.I. Odegaard, R.R. Ricardo-Gonzalez, A. Red Eagle, D. Vats, C.R. Morel, M.H. Goforth, V. Subramanian, L. Mukundan, A.W. Ferrante, A. Chawla, Alternative M2 activation of Kupffer cells by PPAR δ ameliorates obesity-induced insulin resistance, *Cell Metab.* 7 (2008) 496–507.

- [211] S.L. Friedman, Molecular regulation of hepatic fibrosis, an integrated cellular response to tissue injury, *J. Biol. Chem.* 275 (2000) 2247–2250.
- [212] K. Hellemans, L. Michalik, A. Dittie, A. Knorr, K. Rombouts, J. de Jong, C. Heirman, E. Quartier, F. Schuit, W. Wahli, A. Geerts, Peroxisome proliferator-activated receptor- β signaling contributes to enhanced proliferation of hepatic stellate cells, *Gastroenterology* 124 (2003) 184–201.
- [213] K. Hellemans, K. Rombouts, E. Quartier, A.S. Dittie, A. Knorr, L. Michalik, V. Rogiers, F. Schuit, W. Wahli, A. Geerts, PPAR β regulates vitamin A metabolism-related gene expression in hepatic stellate cells undergoing activation, *J. Lipid Res.* 44 (2003) 280–295.
- [214] K. Iwaisako, M. Haimerl, Y.H. Paik, K. Taura, Y. Kodama, C. Sirlin, E. Yu, R.T. Yu, M. Downes, R.M. Evans, D.A. Brenner, B. Schnabl, Protection from liver fibrosis by a peroxisome proliferator-activated receptor δ agonist, *Proc. Natl. Acad. Sci. U S A* 109 (2012) E1369–E1376.
- [215] R. Kostadinova, A. Montagner, E. Gouranton, S. Fleury, H. Guillou, D. Dombrowicz, P. Desreumaux, W. Wahli, GW501516-activated PPAR β/δ promotes liver fibrosis via p38-JNK MAPK-induced hepatic stellate cell proliferation, *Cell. Biosci.* 2 (2012) 34.
- [216] Y.M. Cho, Y. Fujita, T.J. Kieffer, Glucagon-like peptide-1: glucose homeostasis and beyond, *Annu. Rev. Physiol.* 76 (2014) 535–559.
- [217] M. Daoudi, N. Hennuyer, M.G. Borland, V. Touche, C. Duhem, B. Gross, R. Caiazza, J. Kerr-Conte, F. Pattou, J.M. Peters, B. Staels, S. Lestavel, PPAR β/δ activation induces enteroendocrine L Cell GLP-1 production, *Gastroenterology* 140 (2011) 1564–1574.

- [218] T. Tang, M.J. Abbott, M. Ahmadian, A.B. Lopes, Y. Wang, H.S. Sul, Desnutrin/ATGL activates PPAR δ to promote mitochondrial function for insulin secretion in islet β Cells, *Cell Metab.* 18 (2013) 883–895.
- [219] M.S. Winzell, E.M. Wulff, G.S. Olsen, P. Sauerberg, C.F. Gotfredsen, B. Ahrén, Improved insulin sensitivity and islet function after PPAR δ activation in diabetic db/db mice, *Eur. J. Pharmacol.* 626 (2010) 297–305.
- [220] J. Iglesias, S. Barg, D. Vallois, S. Lahiri, C. Roger, A. Yessoufou, S. Pradevand, A. McDonald, C. Bonal, F. Reimann, F. Gribble, M.-B. Debril, D. Metzger, P. Chambon, P. Herrera, G.A. Rutter, M. Prentki, B. Thorens, W. Wahli, PPAR β/δ affects pancreatic β cell mass and insulin secretion in mice, *J. Clin. Investig.* 122 (2012) 4105–4117.
- [221] F. Varnat, B.B.T. Heggeler, P. Grisel, N. Boucard, I. Corthésy–Theulaz, W. Wahli, B. Desvergne, PPAR β/δ regulates paneth cell differentiation via controlling the Hedgehog signaling pathway, *Gastroenterology* 131 (2006) 538–553.
- [222] E.R. Fearon, B. Vogelstein, A genetic model for colorectal tumorigenesis, *Cell* 61 (1990) 759–767.
- [223] J. Groden, A. Thliveris, W. Samowitz, M. Carlson, L. Gelbert, H. Albertsen, G. Joslyn, J. Stevens, L. Spirio, M. Robertson, L. Sargeant, K. Krapcho, E. Wolff, R. Burt, J.P. Hughes, J. Warrington, J. McPherson, J. Wasmuth, Identification and characterization of the familial adenomatous polyposis coli gene, *Cell* 66 (1991) 589–600.
- [224] I. Nishisho, Y. Nakamura, Y. Miyoshi, Y. Miki, H. Ando, A. Horii, K. Koyama, J. Utsunomiya, S. Baba, P. Hedge, Mutations of chromosome 5q21 genes in FAP and colorectal cancer patients, *Science* 253 (1991) 665–669.

- [225] C. Neufert, C. Becker, M.F. Neurath, An inducible mouse model of colon carcinogenesis for the analysis of sporadic and inflammation-driven tumor progression, *Nat. Protoc.* 2 (2007) 1998–2004.
- [226] T.-C. He, T.A. Chan, B. Vogelstein, K.W. Kinzler, PPAR δ is an APC-regulated target of nonsteroidal anti-inflammatory drugs, *Cell* 99 (1999) 335–345.
- [227] D. Wang, H. Wang, Y. Guo, W. Ning, S. Katkuri, W. Wahli, B. Desvergne, S.K. Dey, R.N. DuBois, Crosstalk between peroxisome proliferator-activated receptor delta and VEGF stimulates cancer progression, *Proc. Natl. Acad. Sci. U S A* 103 (2006) 19069–19074.
- [228] F.S. Harman, C.J. Nicol, H.E. Marin, J.M. Ward, F.J. Gonzalez, J.M. Peters, Peroxisome proliferator-activated receptor- δ attenuates colon carcinogenesis, *Nat. Med.* 10 (2004) 481–483.
- [229] H.E. Marin, M.A. Peraza, A.N. Billin, T.M. Willson, J.M. Ward, M.J. Kennett, F.J. Gonzalez, J.M. Peters, Ligand activation of peroxisome proliferator-activated receptor beta inhibits colon carcinogenesis, *Cancer Res.* 66 (2006) 4394–4401.
- [230] S. Beyaz, M.D. Mana, J. Roper, D. Kedrin, A. Saadatpour, S.-J. Hong, K.E. Bauer-Rowe, M.E. Xifaras, A. Akkad, E. Arias, L. Pinello, Y. Katz, S. Shinagare, M. Abu-Remaileh, M.M. Mihaylova, D.W. Lamming, R. Dogum, G. Guo, G.W. Bell, M. Selig, G.P. Nielsen, N. Gupta, C.R. Ferrone, V. Deshpande, G.-C. Yuan, S.H. Orkin, D.M. Sabatini, Ö.H. Yilmaz, High-fat diet enhances stemness and tumorigenicity of intestinal progenitors, *Nature* 531 (2016) 53–58.

- [231] K.R. Reed, O.J. Sansom, A.J. Hayes, A.J. Gescher, D.J. Winton, J.M. Peters, A.R. Clarke, PPAR δ status and Apc-mediated tumorigenesis in the mouse intestine, *Oncogene* 23 (2004) 8992–8996.
- [232] H.E. Hollingshead, M.G. Borland, A.N. Billin, T.M. Willson, F.J. Gonzalez, J.M. Peters, Ligand activation of peroxisome proliferator-activated receptor-beta/delta (PPARbeta/delta) and inhibition of cyclooxygenase 2 (COX2) attenuate colon carcinogenesis through independent signaling mechanisms, *Carcinogenesis* 29 (2008) 169–176.
- [233] R.A. Gupta, D. Wang, S. Katkuri, H. Wang, S.K. Dey, R.N. DuBois, Activation of nuclear hormone receptor peroxisome proliferator-activated receptor- δ accelerates intestinal adenoma growth, *Nat. Med.* 10 (2004) 245–247.
- [234] C. Luo, P. Puigserver, Stem cells: dietary fat promotes intestinal dysregulation, *Nature* 531 (2016) 42–43.
- [235] R.A. Gupta, J. Tan, W.F. Krause, M.W. Geraci, T.M. Willson, S.K. Dey, R.N. DuBois, Prostacyclin-mediated activation of peroxisome proliferator-activated receptor delta in colorectal cancer, *Proc. Natl. Acad. Sci. U S A* 97 (2000) 13275–13280.
- [236] B. Delage, A. Rullier, M. Capdepon, E. Rullier, P. Cassand, The effect of body weight on altered expression of nuclear receptors and cyclooxygenase-2 in human colorectal cancers, *Nutr. J.* 6 (2007) 20.
- [237] L. Yang, Z.G. Zhou, H.Z. Luo, B. Zhou, Q.J. Xia, C. Tian, Quantitative analysis of PPAR δ mRNA by real-time RT-PCR in 86 rectal cancer tissues, *Eur. J. Surg. Oncol.* 32 (2006) 181–185.

- [238] J.E. Foreman, W.-C.L. Chang, P.S. Palkar, B. Zhu, M.G. Borland, J.L. Williams, L.R. Kramer, M.L. Clapper, F.J. Gonzalez, J.M. Peters, Functional characterization of peroxisome proliferator-activated receptor- β/δ expression in colon cancer, *Mol. Carcinog.* 50 (2011) 884–900.
- [239] J. Feilchenfeldt, M.-A. Bründler, C. Soravia, M. Tötsch, C.A. Meier, Peroxisome proliferator-activated receptors (PPARs) and associated transcription factors in colon cancer: reduced expression of PPAR γ -coactivator 1 (PGC-1), *Cancer Lett.* 203 (2004) 25–33.
- [240] J.G. Neels, P.A. Grimaldi, Physiological functions of peroxisome proliferator-activated receptor β , *Physiol. Rev.* 94 (2014) 795–858.
- [241] E.E. Girroir, H.E. Hollingshead, P. He, B. Zhu, G.H. Perdew, J.M. Peters, Quantitative expression patterns of peroxisome proliferator-activated receptor- β/δ (PPAR β/δ) protein in mice, *Biochem. Biophys. Res. Commun.* 371 (2008) 456–461.
- [242] J.M. Peters, H.E. Hollingshead, F.J. Gonzalez, Role of peroxisome-proliferator-activated receptor β/δ (PPAR β/δ) in gastrointestinal tract function and disease, *Clin. Sci.* 115 (2008) 107–127.
- [243] A.E. McCart, N.K. Vickaryous, A. Silver, Apc mice: models, modifiers and mutants, *Pathol. Res. Pract.* 204 (2008) 479–490.
- [244] W.F. Dove, R.T. Cormier, K.A. Gould, R.B. Halberg, A.J. Merritt, M.A. Newton, A.R. Shoemaker, The intestinal epithelium and its neoplasms: genetic, cellular and tissue interactions, *Philos. Trans. Royal Soc. B: Biol. Sci.* 353 (1998) 915–923.
- [245] A.R. Shoemaker, A.R. Moser, C.A. Midgley, L. Clipson, M.A. Newton, W.F. Dove, A resistant genetic background leading to incomplete penetrance of intestinal neoplasia and

- reduced loss of heterozygosity in *ApcMin/+* mice, *Proc. Natl. Acad. Sci. U S A* 95 (1998) 10826–10831.
- [246] T. Karantanos, G. Theodoropoulos, D. Pektasides, M. Gazouli, Clock genes: their role in colorectal cancer, *World J. Gastroenterol.* 20 (2014) 1986–1992.
- [247] T. Karantanos, G. Theodoropoulos, M. Gazouli, A. Vaiopoulou, C. Karantanou, D.J. Stravopodis, K. Bramis, M. Lymperi, D. Pektasidis, Association of the clock genes polymorphisms with colorectal cancer susceptibility, *J. Surg. Oncol.* 108 (2013) 563–567.
- [248] C.H. Johnson, C.M. Dejea, D. Edler, L.T. Hoang, A.F. Santidrian, B.H. Felding, J. Ivanisevic, K. Cho, E.C. Wick, E.M. Hechenbleikner, W. Uritboonthai, L. Goetz, R.A. Casero, D.M. Pardoll, J.R. White, G.J. Patti, C.L. Sears, G. Siuzdak, Metabolism links bacterial biofilms and colon carcinogenesis, *Cell Metab.* 21 (2015) 891–897.
- [249] A. Montagner, W. Wahli, N.S. Tan, Nuclear receptor peroxisome proliferator activated receptor (PPAR) beta/delta in skin wound healing and cancer, *Eur. J. Dermatol.* 25 (2015) 4–11.
- [250] C. Cataisson, E. Joseloff, R. Murillas, A. Wang, C. Atwell, S. Torgerson, M. Gerdes, J. Subleski, J.L. Gao, P.M. Murphy, R.H. Wiltout, C. Vinson, S.H. Yuspa, Activation of cutaneous protein kinase C alpha induces keratinocyte apoptosis and intraepidermal inflammation by independent signaling pathways, *J. Immunol.* 171 (2003) 2703–2713.
- [251] H.Q. Wang, R.C. Smart, Overexpression of protein kinase C-alpha in the epidermis of transgenic mice results in striking alterations in phorbol ester-induced inflammation and COX-2, MIP-2 and TNF-alpha expression but not tumor promotion, *J. Cell Sci.* 112 (1999) 3497–3506.

- [252] G. Nakatsu, X. Li, H. Zhou, J. Sheng, S.H. Wong, W.K.K. Wu, S.C. Ng, H. Tsoi, Y. Dong, N. Zhang, Y. He, Q. Kang, L. Cao, K. Wang, J. Zhang, Q. Liang, J. Yu, J.J.Y. Sung, Gut mucosal microbiome across stages of colorectal carcinogenesis, *Nat. Commun.* 6 (2015) 8727.
- [253] J.-C. Ehrhart, F.P. Gosselet, R.L.M. Culerrier, A. Sarasin, UVB-induced mutations in human key gatekeeper genes governing signalling pathways and consequences for skin tumourigenesis, *Photochem. Photobiol. Sci.* 2 (2003) 825–834.
- [254] Y. Matsumura, H.N. Ananthaswamy, Toxic effects of ultraviolet radiation on the skin, *Toxicol. Appl. Pharmacol.* 195 (2004) 298–308.
- [255] A. Montagner, M.B. Delgado, C. Tallichet-Blanc, J.S.K. Chan, M.K. Sng, H. Mottaz, G. Degueurce, Y. Lippi, C. Moret, M. Baruchet, M. Antsiferova, S. Werner, D. Hohl, T. Al Saati, P.J. Farmer, N.S. Tan, L. Michalik, W. Wahli, Src is activated by the nuclear receptor peroxisome proliferator-activated receptor β/δ in ultraviolet radiation-induced skin cancer, *EMBO Mol. Med.* 6 (2014) 80–98.
- [256] S.I. Grivennikov, F.R. Greten, M. Karin, Immunity, inflammation, and cancer, *Cell* 140 (2010) 883–899.
- [257] T. Bondar, R. Medzhitov, The origins of tumor-promoting inflammation, *Cancer Cell* 24 (2013) 143–144.
- [258] G. Degueurce, I. D'Errico, C. Pich, M. Ibberson, F. Schütz, A. Montagner, M. Sgandurra, L. Mury, P. Jafari, A. Boda, J. Meunier, R. Rezzonico, N.C. Brembilla, D. Hohl, A. Kolios, G. Hofbauer, I. Xenarios, L. Michalik, Identification of a novel PPAR β/δ /miR-21-3p axis in UV-induced skin inflammation, *EMBO Mol. Med.* 8 (2016) 919–936.

- [259] D.J. Kim, T.E. Akiyama, F.S. Harman, A.M. Burns, W. Shan, J.M. Ward, M.J. Kennett, F.J. Gonzalez, J.M. Peters, Peroxisome proliferator-activated receptor beta (delta)-dependent regulation of ubiquitin C expression contributes to attenuation of skin carcinogenesis, *J. Biol. Chem.* 279 (2004) 23719–23727.
- [260] M.T. Bility, M.K. Devlin-Durante, N. Blazanin, A.B. Glick, J.M. Ward, B.H. Kang, M.J. Kennett, F.J. Gonzalez, J.M. Peters, Ligand activation of peroxisome proliferator-activated receptor beta/delta (PPAR beta/delta) inhibits chemically induced skin tumorigenesis, *Carcinogenesis* 29 (2008) 2406–2414.
- [261] P. Boukamp, Non-melanoma skin cancer: what drives tumor development and progression? *Carcinogenesis* 26 (2005) 1657–1667.
- [262] E.L. Abel, J.M. Angel, K. Kiguchi, J. DiGiovanni, Multi-stage chemical carcinogenesis in mouse skin: fundamentals and applications, *Nat. Protoc.* 4 (2009) 1350–1362.
- [263] F.M. Lieu, K. Yamanishi, K. Konishi, S. Kishimoto, H. Yasuno, Low incidence of Ha-ras oncogene mutations in human epidermal tumors, *Cancer Lett.* 59 (1991) 231–235.
- [264] W.E. Pierceall, L.H. Goldberg, M.A. Tainsky, T. Mukhopadhyay, H.N. Ananthaswamy, Ras gene mutation and amplification in human nonmelanoma skin cancers, *Mol. Carcinog.* 4 (1991) 196–202.
- [265] C. Campbell, A.G. Quinn, J.L. Rees, Codon 12 Harvey-ras mutations are rare events in non-melanoma human skin cancer, *Br. J. Dermatol.* 128 (1993) 111–114.
- [266] H.J. Kim, S.A. Ham, S.U. Kim, J.Y. Hwang, J.H. Kim, K.C. Chang, C. Yabe-Nishimura, J.H. Kim, H.G. Seo, Transforming growth factor-beta1 is a molecular target for the peroxisome proliferator-activated receptor delta, *Circ. Res.* 102 (2008) 193–200.

- [267] A.B. Glick, TGF β ? Back to the future: revisiting its role as a transforming growth factor, *Cancer Biol. Ther.* 3 (2004) 276–283.
- [268] L. Mordasky Markell, R. Perez-Lorenzo, K.E. Masiuk, M.J. Kennett, A.B. Glick, Use of a TGF β type I receptor inhibitor in mouse skin carcinogenesis reveals a dual role for TGF β signaling in tumor promotion and progression, *Carcinogenesis* 31 (2010) 2127–2135.
- [269] W. Cui, D.J. Fowles, S. Bryson, E. Duffie, H. Ireland, A. Balmain, R.J. Akhurst, TGF β 1 inhibits the formation of benign skin tumors, but enhances progression to invasive spindle carcinomas in transgenic mice, *Cell* 86 (1996) 531–542.
- [270] A.B. Glick, The role of TGF β signaling in squamous cell cancer: lessons from mouse models, *J. Skin Cancer* 2012 (2012) 249063.
- [271] A. Ravindran, J. Mohammed, A.J. Gunderson, X. Cui, A.B. Glick, Tumor-promoting role of TGF β 1 signaling in ultraviolet B-induced skin carcinogenesis is associated with cutaneous inflammation and lymph node migration of dermal dendritic cells, *Carcinogenesis* 35 (2014) 959–966.
- [272] H. Ikushima, K. Miyazono, Cellular context-dependent “colors” of transforming growth factor- β signaling, *Cancer Sci.* 101 (2010) 306–312.
- [273] P. Papageorgis, TGF β signaling in tumor initiation, epithelial-to-mesenchymal transition, and metastasis, *J. Oncol.* 2015 (2015) 587193.
- [274] N.S. Tan, L. Michalik, N. Noy, R. Yasmin, C. Pacot, M. Heim, B. Fluhmann, B. Desvergne, W. Wahli, Critical roles of PPAR beta/delta in keratinocyte response to inflammation, *Genes Dev.* 15 (2001) 3263–3277.

- [275] A.D. Burdick, M.T. Bility, E.E. Girroir, A.N. Billin, T.M. Willson, F.J. Gonzalez, J.M. Peters, Ligand activation of peroxisome proliferator-activated receptor- β/δ (PPAR β/δ) inhibits cell growth of human N/TERT-1 keratinocytes, *Cell. Signal.* 19 (2007) 1163–1171.
- [276] J.M. Peters, F.J. Gonzalez, Sorting out the functional role(s) of peroxisome proliferator-activated receptor- β/δ (PPAR β/δ) in cell proliferation and cancer, *Biochem. Biophys. Res. Commun.* 1796 (2009) 230–241.
- [277] E. Rognoni, M. Widmaier, M. Jakobson, R. Ruppert, S. Ussar, D. Katsougkri, R.T. Böttcher, J.E. Lai-Cheong, D.B. Rifkin, J.A. McGrath, R. Fässler, Kindlin-1 controls Wnt and TGF- β availability to regulate cutaneous stem cell proliferation, *Nat. Med.* 20 (2014) 350–359.
- [278] H.R. Seo, Y.-W. Kwan, C.-K. Cho, S. Bae, S.-J. Lee, J.-W. Soh, H.-Y. Chung, Y.-S. Lee, PKC α induces differentiation through ERK1/2 phosphorylation in mouse keratinocytes, *Exp. Mol. Med.* 36 (2004) 292–299.
- [279] H.E. Kocalis, M.K. Turney, R.L. Printz, G.N. Laryea, L.J. Muglia, S.S. Davies, G.D. Stanwood, O.P. McGuinness, K.D. Niswender, Neuron-specific deletion of peroxisome proliferator-activated receptor delta (PPAR δ) in mice leads to increased susceptibility to diet-induced obesity, *PLoS One* 7 (2012) e42981.
- [280] A. Iwashita, Y. Muramatsu, T. Yamazaki, M. Muramoto, Y. Kita, S. Yamazaki, K. Mihara, A. Moriguchi, N. Matsuoka, Neuroprotective efficacy of the peroxisome proliferator-activated receptor delta-selective agonists *in vitro* and *in vivo*, *J. Pharmacol. Exp. Ther.* 320 (2007) 1087–1096.

- [281] C.I. Schnegg, M.E. Robbins, Neuroprotective mechanisms of PPAR δ : modulation of oxidative stress and inflammatory processes, *PPAR Res.* 2011 (2011) 373560.
- [282] J.W. Woods, M. Tanen, D.J. Figueroa, C. Biswas, E. Zycband, D.E. Moller, C.P. Austin, J.P. Berger, Localization of PPAR δ in murine central nervous system: expression in oligodendrocytes and neurons, *Brain Res.* 975 (2003) 10–21.
- [283] I. Paterniti, E. Esposito, E. Mazzon, M. Galuppo, R. Di Paola, P. Bramanti, A. Kapoor, C. Thiemermann, S. Cuzzocrea, Evidence for the role of peroxisome proliferator-activated receptor-beta/delta in the development of spinal cord injury, *J. Pharmacol. Exp. Ther.* 333 (2010) 465–477.
- [284] M.G. Hall, L. Quignodon, B. Desvergne, Peroxisome proliferator-activated receptor β/δ in the brain: facts and hypothesis, *PPAR Res.* 2008 (2008) 780452.
- [285] S. Aleshin, M. Strokin, M. Sergeeva, G. Reiser, Peroxisome proliferator-activated receptor (PPAR) β/δ , a possible nexus of PPAR α - and PPAR γ -dependent molecular pathways in neurodegenerative diseases: review and novel hypotheses, *Neurochem. Int.* 63 (2013) 322–330.
- [286] S.E. Dunn, R. Bhat, D.S. Straus, R.A. Sobel, R. Axtell, A. Johnson, K. Nguyen, L. Mukundan, M. Moshkova, J.C. Dugas, A. Chawla, L. Steinman, Peroxisome proliferator-activated receptor δ limits the expansion of pathogenic Th cells during central nervous system autoimmunity, *J. Exp. Med.* 207 (2010) 1599–1608.
- [287] D. Arsenijevic, F. de Bilbao, J. Plamondon, E. Paradis, P. Vallet, D. Richard, W. Langhans, P. Giannakopoulos, Increased infarct size and lack of hyperphagic response after focal cerebral ischemia in peroxisome proliferator-activated receptor β -deficient mice, *J. Cereb. Blood Flow Metab.* 26 (2006) 433–445.

- [288] N. Zhang, Y. Yin, S.J. Xu, Y.P. Wu, W.S. Chen, Inflammation & apoptosis in spinal cord injury, *Indian J. Med. Res.* 135 (2012) 287–296.
- [289] I. Paterniti, D. Impellizzeri, R. Crupi, R. Morabito, M. Campolo, E. Esposito, S. Cuzzocrea, Molecular evidence for the involvement of PPAR- δ and PPAR- γ in anti-inflammatory and neuroprotective activities of palmitoylethanolamide after spinal cord trauma, *J. Neuroinflamm.* 10 (2013) 20.
- [290] P.E. Polak, S. Kalinin, C. Dello Russo, V. Gavrilyuk, A. Sharp, J.M. Peters, J. Richardson, T.M. Willson, G. Weinberg, D.L. Feinstein, Protective effects of a peroxisome proliferator-activated receptor- β/δ agonist in experimental autoimmune encephalomyelitis, *J. Neuroimmunol.* 168 (2005) 65–75.
- [291] J.M. Fletcher, S.J. Lalor, C.M. Sweeney, N. Tubridy, K.H.G. Mills, T cells in multiple sclerosis and experimental autoimmune encephalomyelitis, *Clin. Exp. Immunol.* 162 (2010) 1–11.
- [292] S. Kanakasabai, W. Chearwae, C.C. Walline, W. Iams, S.M. Adams, J.J. Bright, Peroxisome proliferator-activated receptor δ agonists inhibit T helper type 1 (Th1) and Th17 responses in experimental allergic encephalomyelitis, *Immunology* 130 (2010) 572–588.
- [293] E.J. Mufson, M.D. Ikonovic, S.E. Counts, S.E. Perez, M. Malek-Ahmadi, S.W. Scheff, S.D. Ginsberg, Molecular and cellular pathophysiology of preclinical Alzheimer's disease, *Behav. Brain Res.* 311 (2016) 54–69.
- [294] P. Kandalepas, R. Vassar, The normal and pathologic roles of the Alzheimer's β -secretase, BACE1, *Curr. Alzheimer Res.* 11 (2014) 441–449.

- [295] M. Ohno, Alzheimer's therapy targeting the β -secretase enzyme BACE1: benefits and potential limitations from the perspective of animal model studies, *Brain Res. Bull.* (2016).
- [296] A.M. Schmidt, B. Sahagan, R.B. Nelson, J. Selmer, R. Rothlein, J.M. Bell, The role of RAGE in amyloid-beta peptide-mediated pathology in Alzheimer's disease, *Curr. Opin. Investig. Drugs* 10 (2009) 672–680.
- [297] S.S. Khan, G.S. Bloom, Tau: the center of a signaling nexus in Alzheimer's disease, *Front. Neurosci.* 10 (2016) 31.
- [298] Z. Cai, N. Liu, C. Wang, B. Qin, Y. Zhou, M. Xiao, L. Chang, L.-J. Yan, B. Zhao, Role of RAGE in Alzheimer's disease, *Cell. Mol. Neurobiol.* 36 (2016) 483–495.
- [299] S. Kalinin, J. Richardson, D. Feinstein, A PPARdelta agonist reduces amyloid burden and brain inflammation in a transgenic mouse model of Alzheimer's disease, *Curr. Alzheimer Res.* 6 (2009) 431–437.
- [300] E. Barroso, J. del Valle, D. Porquet, A.M. Vieira Santos, L. Salvadó, R. Rodríguez-Rodríguez, P. Gutiérrez, M. Anglada-Huguet, J. Alberch, A. Camins, X. Palomer, M. Pallàs, L. Michalik, W. Wahli, M. Vázquez-Carrera, Tau hyperphosphorylation and increased BACE1 and RAGE levels in the cortex of PPAR β/δ -null mice, *Biochim. Biophys. Acta* 1832 (2013) 1241–1248.
- [301] T. Malm, M. Mariani, L.J. Donovan, L. Neilson, G.E. Landreth, Activation of the nuclear receptor PPAR δ is neuroprotective in a transgenic mouse model of Alzheimer's disease through inhibition of inflammation, *J. Neuroinflamm.* 12 (2015) 7.

- [302] N. Lester-Coll, E.J. Rivera, S.J. Soscia, K. Doiron, J.R. Wands, S.M. de la Monte, Intracerebral streptozotocin model of type 3 diabetes: relevance to sporadic Alzheimer's disease, *J. Alzheimers Dis.* 9 (2006) 13–33.
- [303] S.M. de la Monte, J.R. Wands, Alzheimer's disease is type 3 diabetes--evidence reviewed, *J. Diabet. Sci. Technol.* 2 (2008) 1101–1113.
- [304] R. Falcone, T. Marilena Florio, E.D. Giacomo, E. Benedetti, L. Cristiano, A. Antonosante, A. Fidoamore, M. Massimi, M. Alecci, R. Ippoliti, A. Giordano, A. Cimini, PPAR β/δ and γ in a rat model of Parkinson's disease: possible involvement in PD symptoms, *J. Cell. Biochem.* 116 (2015) 844–855.
- [305] S.M. de la Monte, M. Tong, N. Lester-Coll, M. Plater Jr., J.R. Wands, Therapeutic rescue of neurodegeneration in experimental type 3 diabetes: relevance to Alzheimer's disease, *J. Alzheimers Dis.* 10 (2006) 89–109.
- [306] M. Tong, C. Dominguez, Targeting Alzheimer's disease neuro-metabolic dysfunction with a small molecule nuclear receptor agonist (T3D-959) reverses disease pathologies, *J. Alzheimers Dis. Parkinsonism* 6 (2016) 238.
- [307] M. Tong, C. Deochand, J. Didsbury, S.M. de la Monte, T3D-959: a multi-faceted disease remedial drug candidate for the treatment of Alzheimer's disease, *J. Alzheimers Dis.* 51 (2016) 123–138.
- [308] S.M. de la Monte, J.R. Wands, Molecular indices of oxidative stress and mitochondrial dysfunction occur early and often progress with severity of Alzheimer's disease, *J. Alzheimers Dis.* 9 (2006) 167–181.
- [309] H.L. Weingarten, 1-Methyl-4-Phenyl-1,2,3,6-Tetrahydropyridine (MPTP): one designer drug and serendipity, *J. Foren. Sci.* 33 (1988) 588–595.

- [310] C.A. Davie, A review of Parkinson's disease, *Br. Med. Bull.* 86 (2008) 109–127.
- [311] I.J. Kopin, S.P. Markey, MPTP toxicity: implications for research in Parkinson's disease, *Annu. Rev. Neurosci.* 11 (1988) 81–96.
- [312] V. Jackson-Lewis, S. Przedborski, Protocol for the MPTP mouse model of Parkinson's disease, *Nat. Protoc.* 2 (2007) 141–151.
- [313] J. Sian, M. Youdim, P. Riederer, M. Gerlach, MPTP-induced Parkinsonian syndrome, in: G.J. Siegel, B.W. Agranoff, R.W. Albers, S.K. Fisher, M.D. Uhler (Eds.), *Basic Neurochemistry: Molecular, Cellular and Medical Aspects*, Lippincott Williams & Wilkins, Philadelphia, 1998.
- [314] H.L. Martin, R.B. Mounsey, K. Sathe, S. Mustafa, M.C. Nelson, R.M. Evans, P. Teismann, A peroxisome proliferator-activated receptor- δ agonist provides neuroprotection in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine model of Parkinson's disease, *Neuroscience* 240 (2013) 191–203.
- [315] R. Deumens, A. Blokland, J. Prickaerts, Modeling Parkinson's disease in rats: an evaluation of 6-OHDA lesions of the nigrostriatal pathway, *Exp. Neurol.* 175 (2002) 303–317.
- [316] A.S. Dickey, V.V. Pineda, T. Tsunemi, P.P. Liu, H.C. Miranda, S.K. Gilmore-Hall, N. Lomas, K.R. Sampat, A. Buttgerit, M.-J.M. Torres, A.L. Flores, M. Arreola, N. Arbez, S.S. Akimov, T. Gaasterland, E.R. Lazarowski, C.A. Ross, G.W. Yeo, B.L. Sopher, G.K. Magnuson, A.B. Pinkerton, E. Masliah, A.R. La Spada, PPAR- δ is repressed in Huntington's disease, is required for normal neuronal function and can be targeted therapeutically, *Nat. Med.* 22 (2016) 37–45.

- [317] Y. Cheng, Single-particle Cryo-EM at crystallographic resolution, *Cell* 161 (2015) 450–457.
- [318] R. Henderson, Overview and future of single particle electron cryomicroscopy, *Arch. Biochem. Biophys.* 581 (2015) 19–24.
- [319] E. Nogales, S.H.W. Scheres, Cryo-EM: a unique tool for the visualization of macromolecular complexity, *Mol. Cell* 58 (2015) 677–689.
- [320] J.A. Lyons, A. Shahsavar, P.A. Paulsen, B.P. Pedersen, P. Nissen, Expression strategies for structural studies of eukaryotic membrane proteins, *Curr. Opin. Structural Biol.* 38 (2016) 137–144.
- [321] S. Kahremany, A. Livne, A. Gruzman, H. Senderowitz, S. Sasson, Activation of PPAR δ : from computer modelling to biological effects, *Br. J. Pharmacol.* 172 (2015) 754–770.

Figure 1. Overall mechanism of PPAR β/δ function. Similar to other nuclear receptors, PPAR β/δ is structurally composed of an N-terminal domain (NTD), a DNA-binding domain (DBD), and a ligand-binding domain (LBD). PPAR β/δ ligands are currently understood to be transported from the cytoplasm into the nucleus by fatty-acid binding protein 5 (FABP5). In macrophages, the absence of a PPAR β/δ agonist allows for the scavenging of BCL-6, a PPAR β/δ -associated transcriptional repressor. This allows for transcription factors (TFs) to bind to their respective TF-binding sites (TFBSs) and activate the transcription of BCL-6-repressed genes (*b6rg*) encoding for macrophage-associated chemokines or cytokines. In the presence of a PPAR β/δ ligand, an anti-inflammatory effect of PPAR β/δ activation is observed, as BCL-6 is released from PPAR β/δ , leading to transrepression of *b6rg* with the aid of a corepressor complex. In other tissues, the presence of either a PPAR β/δ agonist or antagonist leads to heterodimerization of PPAR β/δ with a retinoid X receptor (RXR), another type of nuclear receptor, as well as the binding of the heterodimer complex to PPAR response elements (PPREs) found in the promoter regions of PPAR β/δ target genes. Depending on whether an agonist or antagonist is bound to PPAR β/δ , either a co-activator or corepressor complex is recruited, thus leading to the transactivation or transrepression of PPAR β/δ target genes, respectively.

Figure 2. PPAR β/δ in wound repair. Skin injury-induced inflammatory signals (e.g., TNF α) activate PPAR β/δ via the SAPK/AP-1 pathway. PPAR β/δ activation concurrently upregulates the transcriptional activation of direct targets, such as integrin-linked kinase (ILK) and 3-phosphoinositide-dependent kinase-1 (PDK1, also known as PDPK1), and downregulates phosphatase and tensin homologue deleted on chromosome 10 (PTEN). These events increase protein kinase B (AKT1) activation, which suppresses keratinocyte apoptosis via modulation of

the death receptor and mitochondrial apoptosis pathways. Keratinocyte-derived interleukin-1 (IL-1) also upregulates PPAR β/δ in fibroblasts, stimulating the production of secreted IL-1 receptor antagonist (sIL-1ra). sIL-1ra inhibits further IL-1-induced production of mitogenic factors by fibroblasts, thereby tightly regulating keratinocyte proliferation during wound healing. PPAR β/δ -enhanced, and subsequently cleaved, angiopoietin-like protein 4 (cANGPTL4) also plays a role in the modulation of vascular leakiness via the JAK/STAT/iNOS pathway. Keratinocyte migration is coregulated by MMP-9 production and the cANGPTL4/JAK/STAT/iNOS pathway. In diabetic patients, PPAR β/δ activation in fibroblasts upregulates the expression of H₂O₂-scavenging catalase and glutathione peroxidase 1 (GPx1), thereby rescuing oxidatively modified ECM proteins to enhance keratinocyte migration during wound healing.

Figure 3. PPAR β/δ in cardiovascular disease. In the vascular endothelium, PPAR β/δ activation directly causes endothelium-dependent vasodilatation by increasing NO production through the PI3K/Akt pathway. PPAR β/δ activation also upregulates the expression of antioxidant genes and reduces NADPH activity, leading to reduced vascular levels of ROS and increased NO bioavailability. PPAR β/δ activation also reduces DAG-PKC-mediated ROS production by enhancing mitochondrial FA oxidation. PPAR β/δ ligands have beneficial effects on plasma lipoproteins, causing reductions in plasma VLDL-TG and LDL-cholesterol levels and increases in HDL-cholesterol levels. In addition, PPAR β/δ activation decreases the abundance of small, dense LDL particles, which are more atherogenic than larger LDL particles. In macrophages, PPAR β/δ agonists elicit anti-inflammatory effects by modulating the activity of the inflammatory suppressor protein BCL-6. In cardiomyocytes, PPAR β/δ controls the

expression of antioxidant genes, genes involved in FA oxidation and the transcriptional co-activator PGC-1 α . As a result of these activities, PPAR β/δ plays a key role in many physiological and pathological processes in the heart. BCL-6: B-cell lymphoma-6. CPT-1: carnitine palmitoyltransferase 1. DAG: diacylglycerol. HDL: high-density lipoprotein. LDL: low-density lipoprotein. NO: nitric oxide. PGC-1 α : PPAR γ co-activator 1 α . PI3K: phosphatidylinositol 3-kinase. PKC: protein kinase C. ROS: reactive oxygen species. TG: triglyceride. VLDL: very-low-density lipoprotein.

Figure 4. Effects of PPAR β/δ on adipocytes and macrophages in adipose tissue. The Th2 cytokines IL-13 and IL-4 secreted by adipocytes induce the expression of PPAR β/δ in macrophages, which in turn polarizes macrophages towards the alternatively activated, anti-inflammatory M2 phenotype, promoting insulin sensitivity in adipose tissue. PPAR β/δ may also increase insulin sensitivity by preventing cytokine-induced STAT3 activation and the subsequent upregulation of SOCS3, which inhibits insulin signaling by interfering with IRS function. PPAR β/δ also activates the overall level of oxidative metabolism in white adipose tissue, and this effect may reduce triglyceride accumulation in adipocytes and the circulating NEFA levels. IRS: insulin receptor substrate. JAK: Janus kinase. NEFAs: non-esterified FAs. SOCS3: suppressor of cytokine signaling 3. STAT3: signal transducer and activator of transcription 3.

Figure 5. PPAR β/δ in hepatocytes influences the diurnal expression of lipogenic genes in the liver and thereby regulates inter-organ communication between the liver and muscle. In contrast to hepatic PPAR α , which is active during fasting and which controls FA degradation in response to adipose tissue lipolysis, PPAR β/δ shows diurnal expression that peaks during the

night, when lipogenesis is high, in fed mice. Disruption of PPAR β/δ in hepatocytes influences the diurnal expression of acetyl-CoA carboxylase (ACC), a key lipogenic enzyme, via a mechanism that remains unknown. This effect shifts the lipogenic flux and influences the rhythmic synthesis of FAs such as stearic acid (C18:0) and oleic acid (C18:1n-9), which are essential for the synthesis of phosphatidylcholine 18:0/18:1 (PC(18:0/18:1)). PC(18:0/18:1) activates PPAR α and thereby increases FA uptake in muscle, reducing circulating free FA and TG levels.

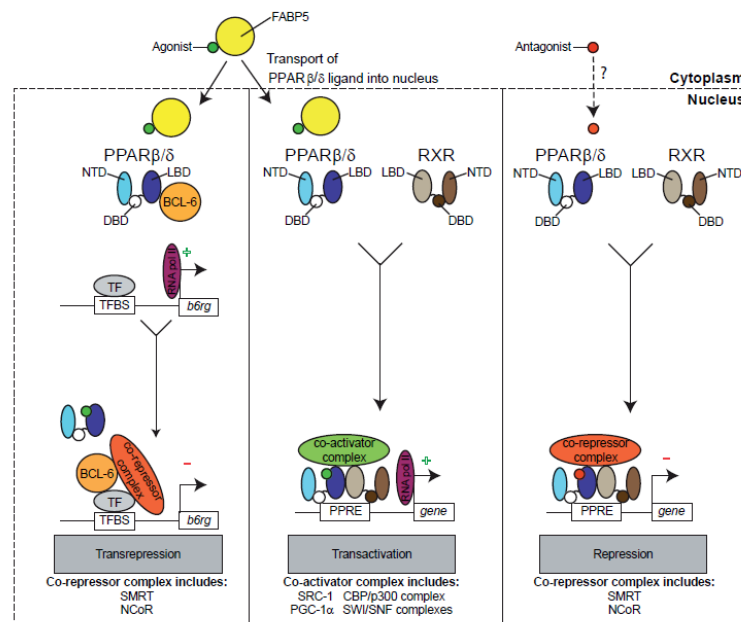


Figure 1

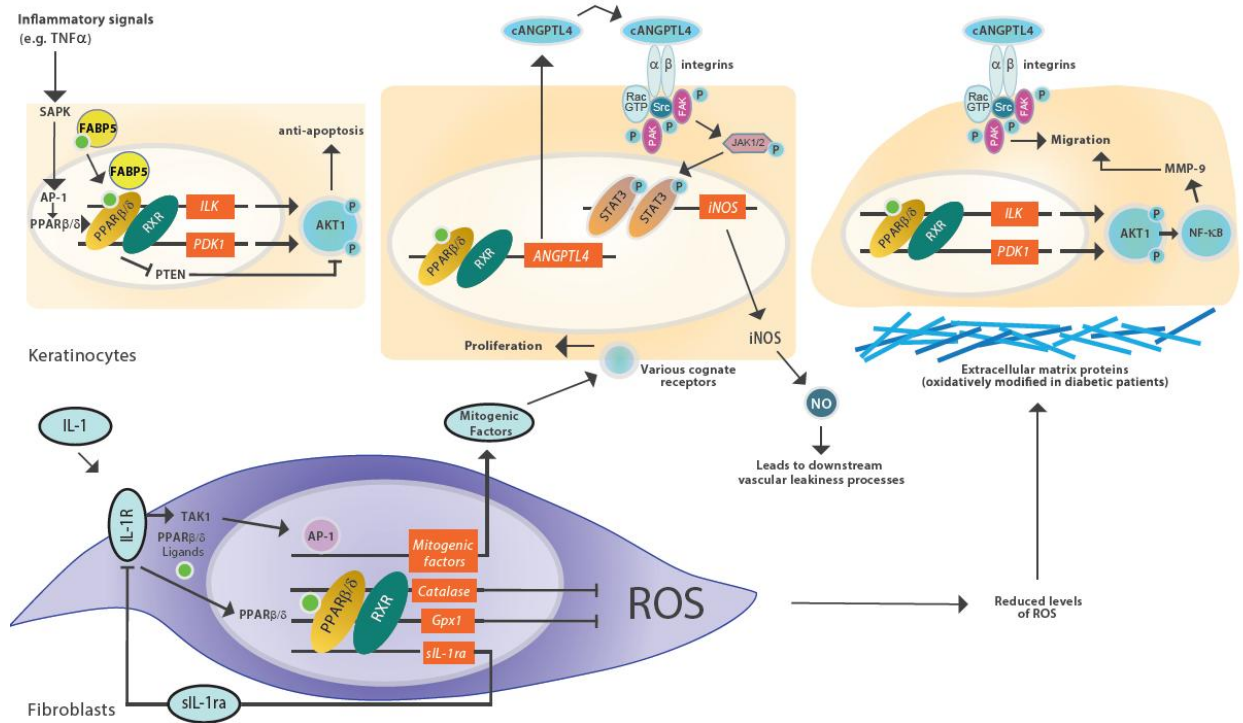


Figure 2

ACCEPTED

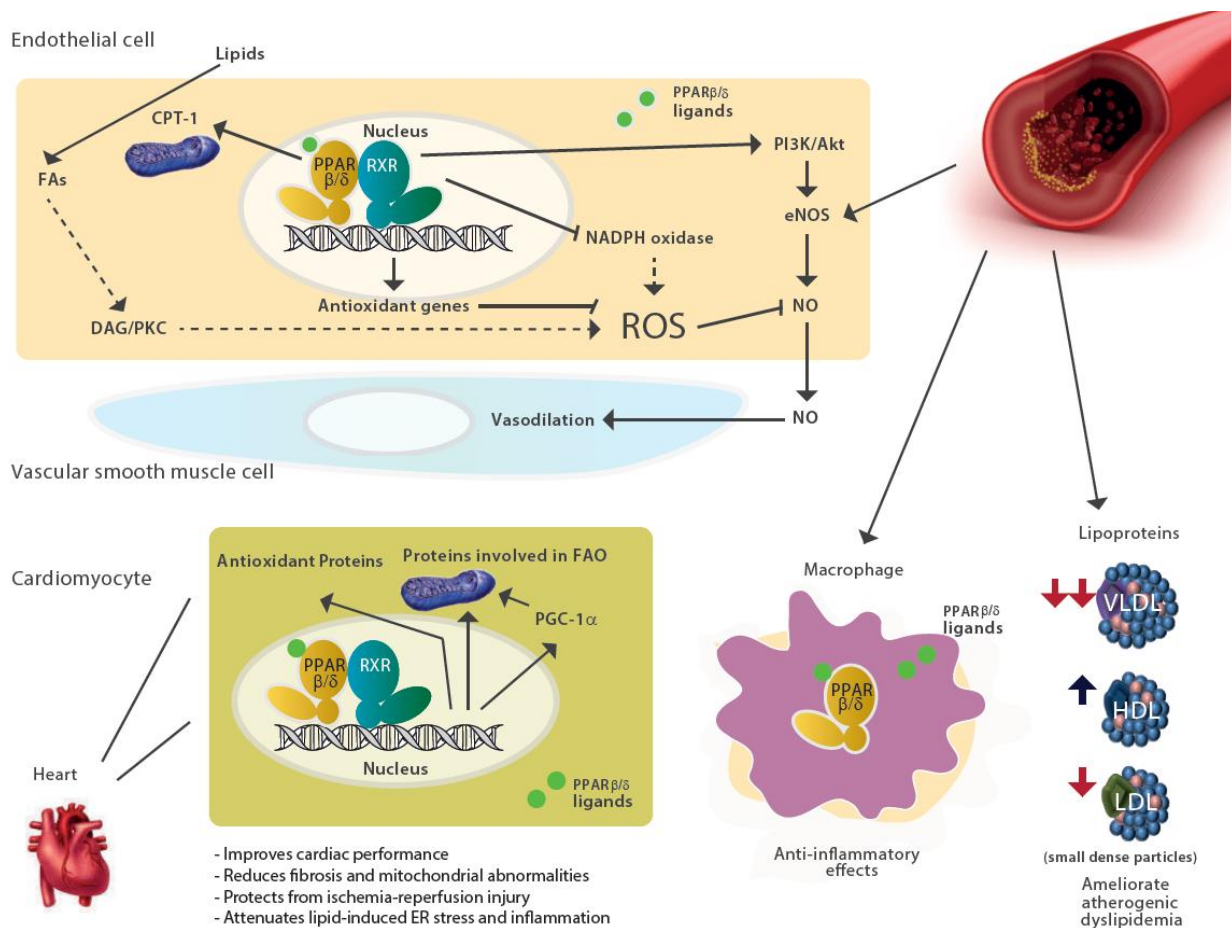


Figure 3

ACCEPTED

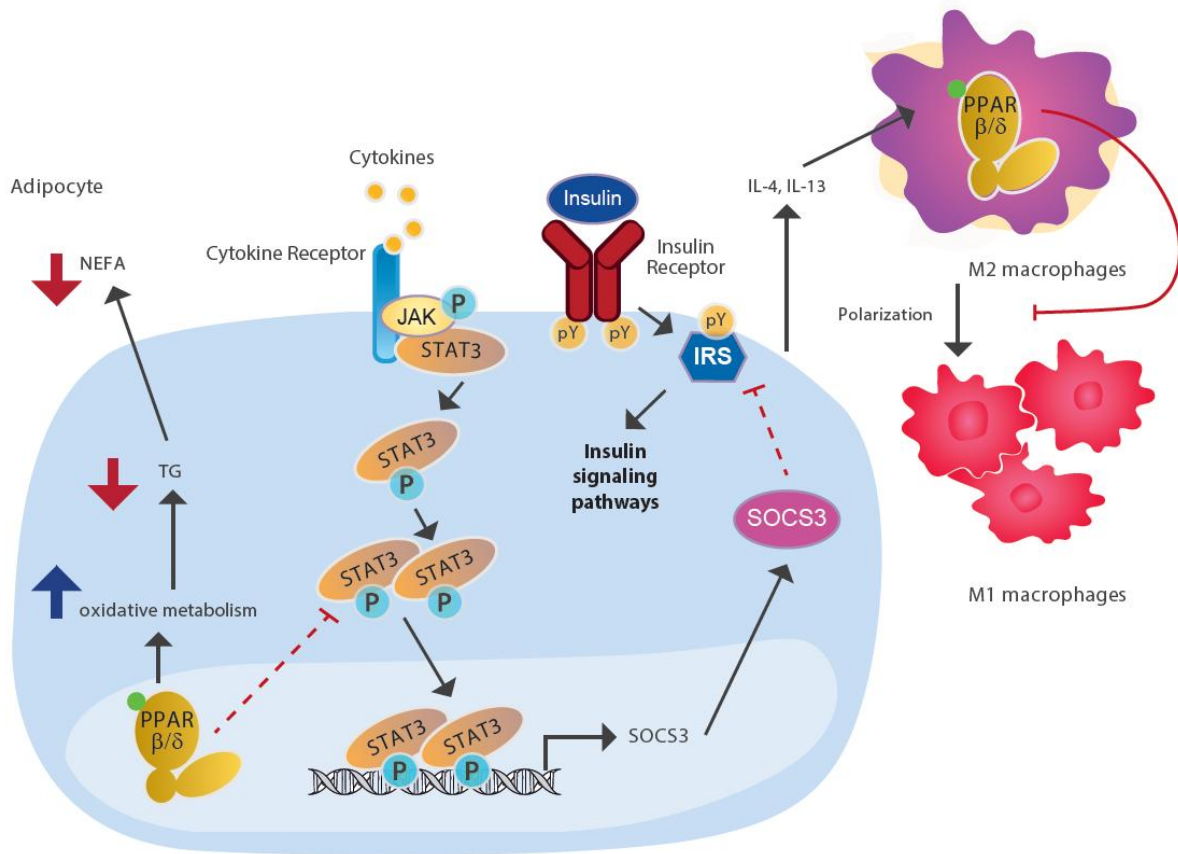


Figure 4

ACCE

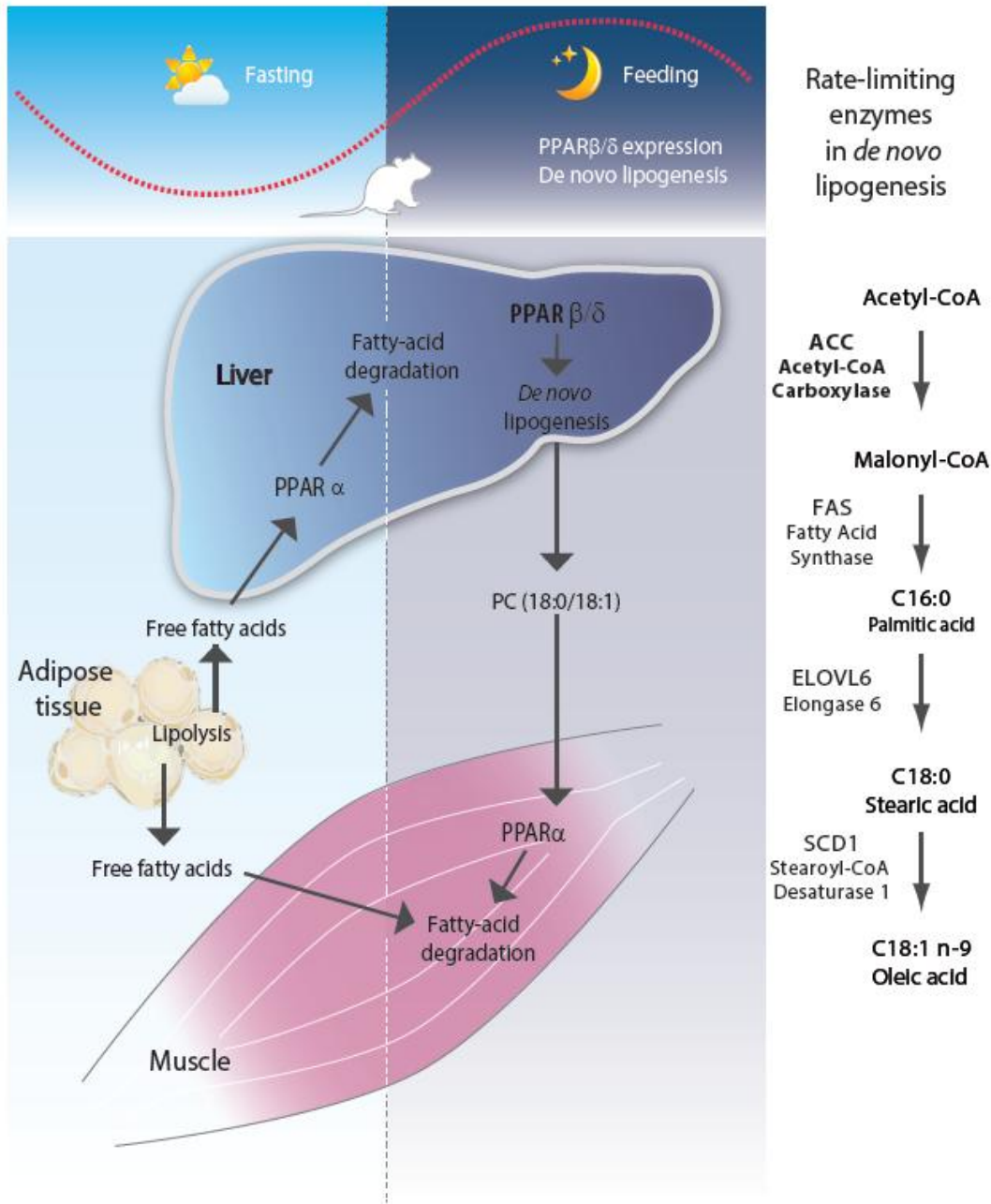


Figure 5