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# Massive horizontal transfer of transposable elements in insects

Jean Peccoud<sup>a,1</sup>, Vincent Loiseau<sup>a</sup>, Richard Cordaux<sup>a</sup>, and Clément Gilbert<sup>a,1</sup>

<sup>a</sup>UMR CNRS 7267 Ecologie et Biologie des Interactions, Equipe Ecologie Evolution Symbiose, Université de Poitiers, Poitiers F-86073, France

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Horizontal transfer (HT) of genetic material is central to the architecture and evolution of prokaryote genomes. Within eukaryotes, the majority of HTs reported so far are transfers of transposable elements (TEs). These reports essentially come from studies focusing on specific lineages or types of TEs. Because of the lack of large-scale survey, the amount and impact of HT of TEs (HTT) in eukaryote evolution, as well as the trends and factors shaping these transfers, are poorly known. Here, we report a comprehensive analysis of HTT in 195 insect genomes, representing 123 genera and 13 of the 28 insect orders. We found that these insects were involved in at least 2,248 HTT events that essentially occurred during the last 10 My. We show that DNA transposons transfer horizontally more often than retrotransposons, and unveil phylogenetic relatedness and geographical proximity as major factors facilitating HTT in insects. Even though our study is restricted to a small fraction of insect biodiversity and to a recent evolutionary timeframe, the TEs we found to be horizontally transferred generated up to 24% (2.08% on average) of all nucleotides of insect genomes. Together, our results establish HTT as a major force shaping insect genome evolution.

horizontal transfer | transposable elements | insects | genome evolution | biogeography

Horizontal transfer (HT) is the transmission of genetic material between organisms through a mechanism other than reproduction. In prokaryotes, HT is pervasive, its mechanisms are well understood, and it is now viewed as one of the main forces shaping genome architecture and evolution (1, 2). In contrast, the study of HT in eukaryotes is less documented, but has been increasingly investigated. The majority of genes horizontally acquired by eukaryotes come from bacteria, but the extent to which these transfers have contributed to eukaryote evolution is still unclear (3, 4). Gene transfers from eukaryote to eukaryote appear to be largely limited to filamentous organisms, such as oomycetes and fungi (5, 6).

In animals and plants, very few cases of such horizontal gene transfers (HGTs) have been reported so far (7, 8). In fact, most of the genetic material that is horizontally transferred in animals and plants consists of transposable elements (TEs) (9–11), which are pieces of DNA able to move from a chromosomal locus to another (12). The greater ability of TEs to move between organisms certainly relates to their intrinsic ability to transpose within genomes, which genes cannot do. HT of TEs (HTT) may allow these elements to enter naive genomes, which they invade by making copies of themselves, and then escape before they become fully silenced by anti-TE defenses (13). A growing number of studies have identified such HTT (11, 14–16). However, a common drawback of these studies has been the inclusion of a limited set of TEs (11) or organisms (16), which hampers our understanding of the breadth of HTT, its contribution to genome evolution, and of the factors and barriers shaping these transfers in eukaryotes (13). In this study, we overcame these limitations by performing a large-scale, comprehensive analysis of HTT in insects. We focused on insects because a large number of whole-genome sequences are publicly available for this group and because insect genomes are known to harbor diverse and highly dynamic TE landscapes (17).

## Results and Discussion

To detect HTT in insects, we de novo characterized TEs in all reference genome assemblies available in GenBank as of May 2016 ( $n = 195$  species; Dataset S1 and Fig. S1) which represent 123 genera and 13 of the 28 insect orders. To minimize detection biases, we did not rely on established genome annotations that are available for only a subset of the species included in our study, and instead treated every species' genome equally. This automatic characterization was performed with the Repeat-Modeler pipeline (18) and led to the identification of 53,452 TE families assigned to 98 superfamilies (Dataset S2). These exclude 3,417 families whose consensus sequences were found to include genes that may not belong to TEs (*SI Materials and Methods*), as well as all short interspersed element (SINE) consensus sequences, which might correspond to RNA pseudogenes (19). For each species, the consensus sequences of TE families were used to locate TE copies in genomic contigs. TE copies >100 base pairs were compared by pairwise reciprocal homology searches between every two species. After filtering out short and low-score alignments, and alignments between TEs from different superfamilies, we retained a total of ~5.9 million hits, each of which indicated TE homology between two genomes.

TEs inherited from a recent common ancestor by descendent species, rather than horizontally transferred between these species, may present homology passing our filters. We identified clades of related insect species for which this situation may happen, by relying on the common assumption that inherited TEs evolve neutrally and similarly to synonymous sites of protein coding genes (20). This assumption implies that TEs showing higher interspecific homology than the synonymous sites of orthologous genes should share a more recent ancestor than the host species, and hence be the result of HT (16, 21). Conversely,

## Significance

Eukaryotes normally receive their genetic material from their parents but may occasionally, like prokaryotes do, acquire DNA from unrelated organisms through horizontal transfer (HT). In animals and plants, HT mostly concerns transposable elements (TEs), probably because these pieces of DNA can move within genomes. Assessing the impact of HTs on eukaryote evolution and the factors shaping the dynamics of these HTs requires large-scale systematic studies. We have analyzed the genomes from 195 insect species and found that no fewer than 2,248 events of HT of TEs occurred during the last 10 My, particularly between insects that were closely related and geographically close. These results suggest that HT of TEs plays a major role in insect genome evolution.

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<sup>1</sup>To whom correspondence may be addressed. Email: clement.gilbert30@gmail.com or jean.peccoud@univ-poitiers.fr.

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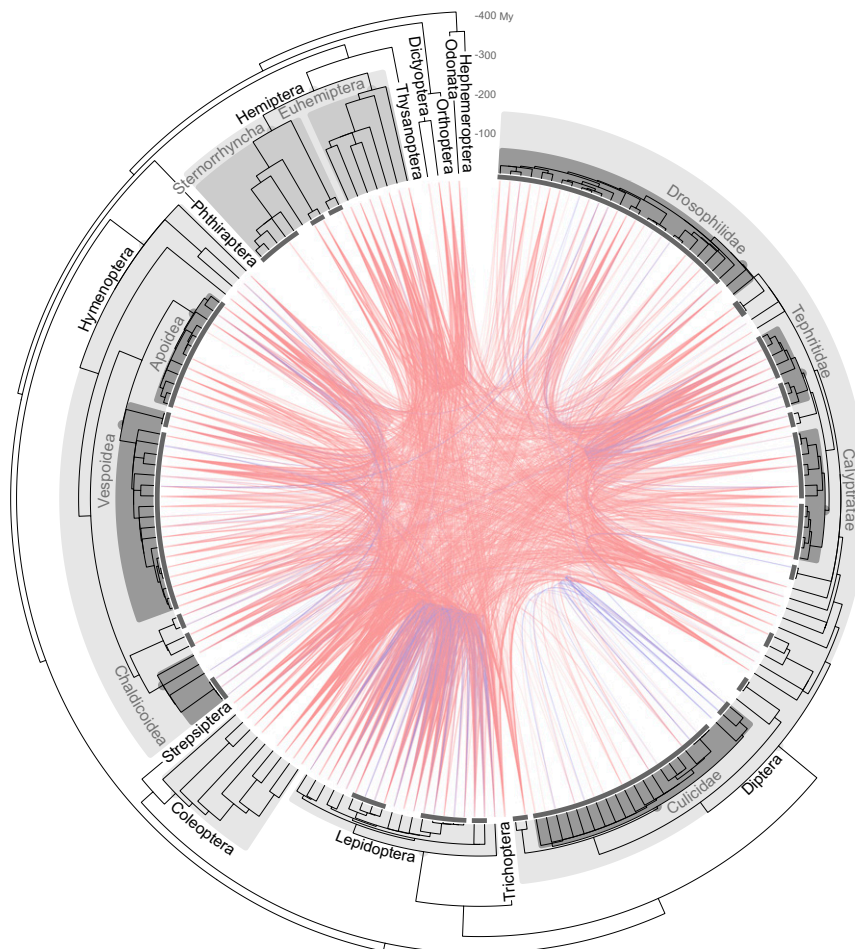
TEs whose divergence is similar to the synonymous divergence of orthologous genes may be vertically inherited. We thus collapsed a clade of insect species into a lineage if (i) a fraction ( $>0.3\%$ ) of its core orthologous genes showed lower synonymous divergence than the highest nucleotide divergence of TEs or (ii) these species diverged in the last 40 My (*Materials and Methods* and Fig. S2). This collapsing resulted in the delineation of 81 lineages. We ignored all homologies within these lineages and considered the  $\sim 1.46$  million hits between species from distinct lineages to essentially result from HT.

To translate these hits into a minimum number of HTT events, it must be considered that many hits may result from a single TE that transferred between two lineages and transposed into multiple similar copies. The resulting hits have a distinct feature: The copies they involve diverged within the recipient lineage after the transfer; hence, they should be more similar to each other than to copies from the other lineage. We applied a heuristic approach based on this principle (*Materials and Methods*) to cluster hits within each pair of lineages and TE superfamily into candidate HTT events. We also considered that a transferred TE might have degraded into nonoverlapping parts that share no sequence homology. To avoid counting these parts as several independently transferred elements, we further clustered hits on the basis of insufficient nucleotide or protein homology between

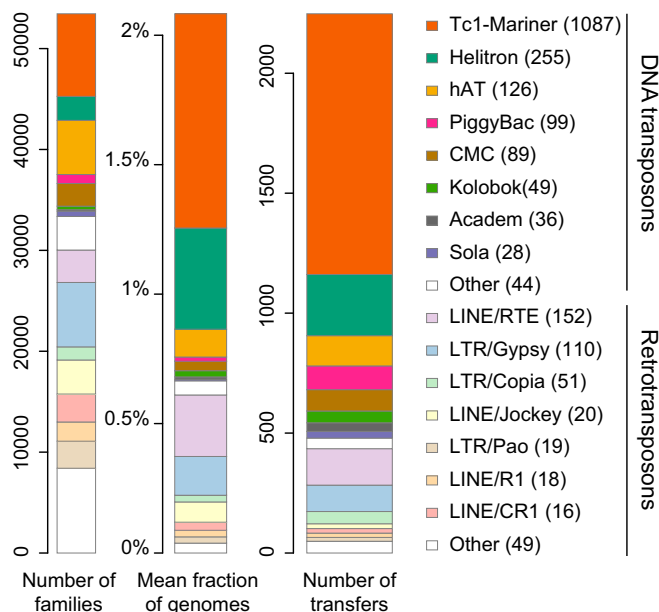
copies. These two rounds of clustering yielded 4,499 HTT events for all pairs of insect lineages (Fig. 1), after discarding transfers that involved fewer than five TE copies per lineage. Taking pairs of lineages separately, each of these transfers represents an independent HTT event.

To infer the minimum number of HTT events across all 81 insect lineages, we considered that any HTT inferred in a pair of lineages might not reflect a direct transfer from one to the other, but separate acquisitions of TEs from other sources, which may already be counted as HTTs in other lineage pairs. There is no method that can discriminate between direct transfer and such “indirect” transfers. However, it is possible to avoid counting indirect HTTs, by constructing networks of insect species sharing similar TEs, in which the minimum number of HTT events is the number of lineages minus one. This parsimonious count resulted in 2,248 HTT events among the 81 insect lineages considered in our study (Fig. 2 and Dataset S2). This unprecedented figure is more than four times higher than the total number of HTT events reported so far in metazoans, plants, and fungi combined (10). Such a high number is still not unexpected, given that studies focusing on one or a few TEs often found one or more HTT events between multiple, distantly related taxa (11, 14–16, 22).

Nonetheless, the actual number of HTT events that occurred in insects must largely exceed our inference, for at least four



**Fig. 1.** HTTs among insects. The concave tree, based on ref. 23, shows the phylogeny of the 195 insect species used in this study. Each dark segment next to tips encompasses related species between which HTT was not inferred, due to the risk of confusion with vertical inheritance, and which were collapsed into a lineage. Each inferred HTT between two lineages, either direct or indirect (4,499 HTTs total), is shown by a curve. If several species of a lineage were found to share TEs due to the same HTT, we connected the two species (one per lineage) that presented the highest level of sequence identity for the transferred element. Blue curves represent transfers of retrotransposons (class I TEs), and red curves transfers of DNA transposons (class II TEs).



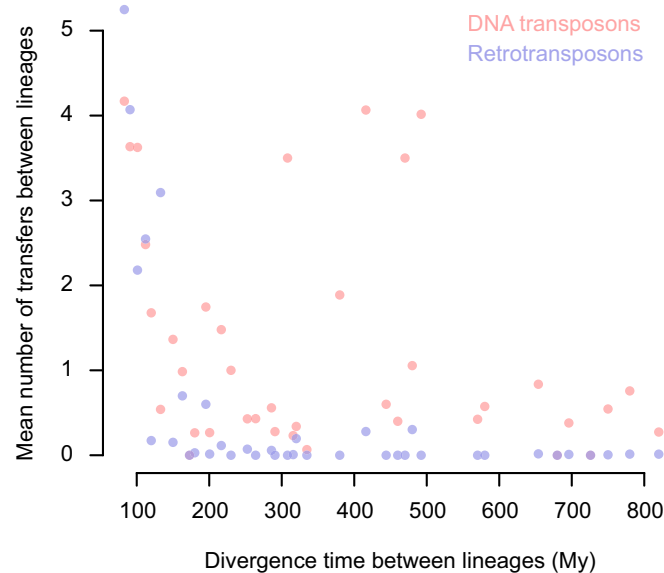
**Fig. 2.** Characteristics of the TEs horizontally transferred between insects. Minimum numbers of transfer events are given in parentheses. The “Other” categories comprise all superfamilies of a given class of TEs [DNA transposons (class II) or retrotransposons (class I)] involved in <15 HTTs each. TE families (Left) correspond to those retrieved using RepeatModeler (Dataset S2). Center represents the mean fraction of an insect genome that is constituted by TEs from these families, which belong to a given super family.

reasons. First, the inherent difficulty in resolving highly similar TE copies as distinct loci during genome assembly may have made some TE families undetectable or unable to pass our selection filters. Second, homology can only be found between recently diverged TEs, putting a maximum date on the HTT events that initiated the divergences. We estimate that the HTT events we uncovered mainly occurred in the last 10 My (assuming that TEs evolve similarly to synonymous sites for which we inferred an overall mutation rate; Fig. S2). Third, we conservatively collapsed many species into lineages within which HTT events have been characterized in previous studies (9). Fourth, the available insect genome sequences represent a very small fraction of the known diversity of insects. However, we found that 72 of the 81 lineages and 176 of the 195 species were involved in at least one HTT event. Therefore, HTT is not only widespread in insects, but the true number of HTT events is likely to be several orders of magnitude larger than the number we report.

The tremendous amount of HTT events we uncovered in insects enabled us to statistically assess the impact of two main factors on this process: phylogenetic and geographic distance. For the former, we relied on recently estimated divergence times (23), and the latter involved assigning as many insect species as possible ( $n = 179$ ) to the six main biogeographic realms. The effects of the two factors may not be confounded, because there is no positive correlation ( $R = -0.01$ ) between species relatedness and geographic cooccurrence (originating from the same realm) among the 81 lineages we defined. We first tested whether HTT events were more likely to occur between less divergent lineages, as observed in bacteria (24, 25). Our analysis revealed a significant negative correlation between the number of HTT events and the divergence times of the lineages (Fig. 3). This pattern could, in theory, reflect vertical inheritance mistakenly inferred as HTT. Indeed, the number of vertically inherited TEs for which we can detect sequence similarities between lineages decreases with their divergence time. If that was the cause for the pattern shown in Fig. 3, the fraction of species

per lineage that are found to share TEs with another lineage, due to an apparent HTT, should also decrease with the divergence of these lineages (SI Discussion). We did not observe such a decrease (Table S1), and therefore saw no evidence that homology between vertically inherited TEs was frequently counted as HTT. The phylogenetic effect we observed is also unlikely to be explained by reduced opportunities of HTT between phylogenetically distant species, because these are not more geographically distant on average. Instead, as proposed for HGT and HTT in bacteria (1, 25, 26), compatibility between TEs and recipient host cells may decrease as genetic distance from source lineages increases. Transposition is known to involve a number of interactions between TEs and host cellular factors (e.g., transcription factors and chromatin), which, depending on the type of TE, may be limited or very intricate (27–30). For example, some DNA transposons from the Tc1-Mariner superfamily only require their transposase to transpose in vitro (27). Weak dependence on host factors may explain why Tc1-Mariner TEs were found to be transferred between more or less distantly related taxa (31). Our systematic search supports this hypothesis as the highest number of HTT in insects was found for members of the Tc1-Mariner superfamily (Fig. 2). Interestingly, retrotransposons show lower transfer rates between distantly related lineages than DNA transposons (Fig. 3). Compared with DNA transposons, retrotransposons may require host factors that: (i) are more numerous and diverse, and/or (ii) tend to be less conserved between taxa. Such differences may explain the overall higher numbers of HTT events of DNA transposons than retrotransposons reported in our study (1,813 vs. 435 transfers; Fig. 2) and others (13, 32).

Finally, we assessed how geographic distance affects the likelihood of HTT at the global scale, by estimating the average number of transfers per pair of species assigned to biogeographic realms. The resulting map (Fig. 4A) shows that all pairs of realms comprise species involved in HTT at varying intensities. To test whether HTT was constrained by geographical distance, we estimated the extent to which two species that share TEs due to a



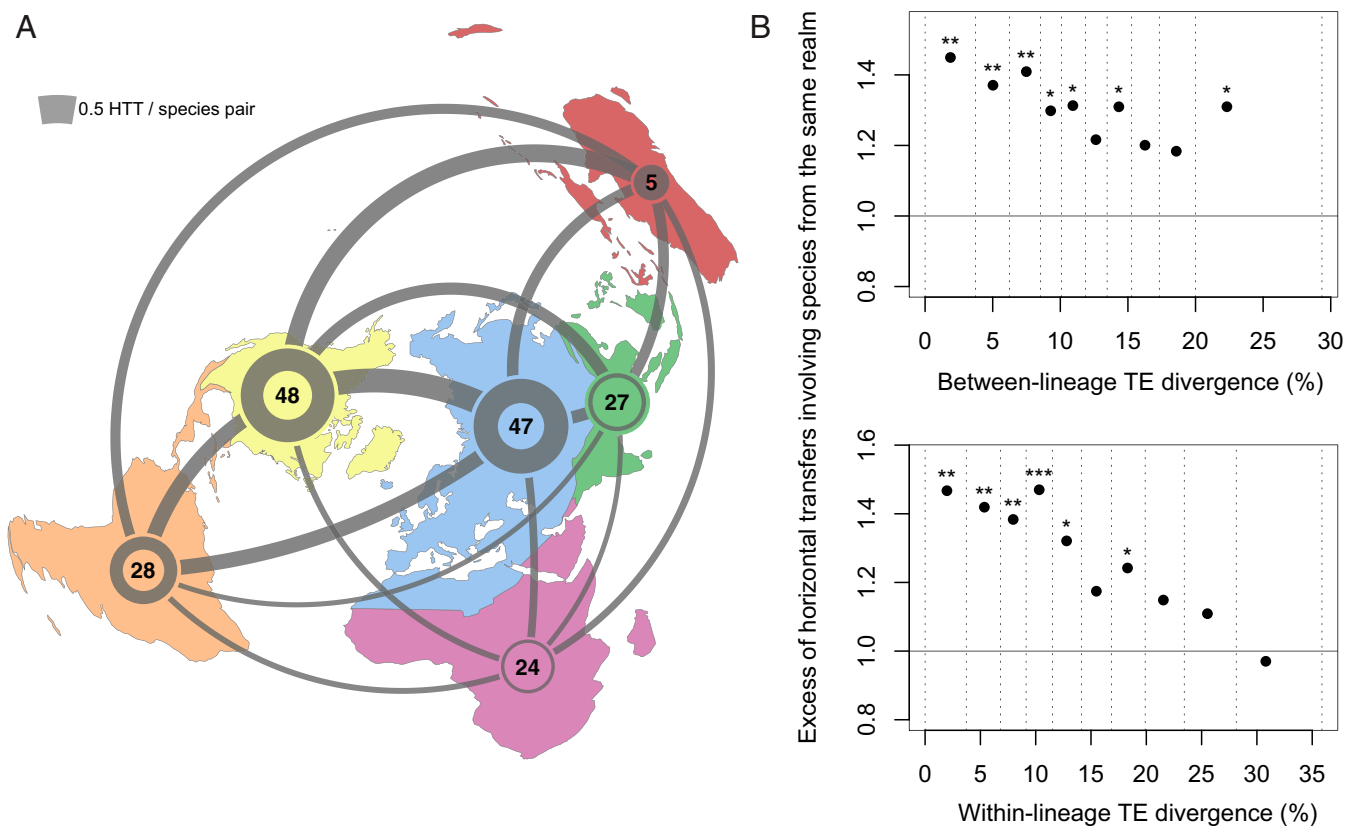
**Fig. 3.** The number of HTTs between insects negatively correlates with insect divergence time. This trend is significant for both DNA transposons (red points) and retrotransposons (blue points) ( $P \sim 4 \times 10^{-3}$  and  $P < 10^{-4}$ , respectively; two-sided Mantel tests). Number of transfers were averaged over pairs of lineages assigned to each 10 My-wide class of divergence time. To avoid bias due to differences in the number of species per lineage, we randomly sampled one species per lineage and computed averages over  $10^4$  samplings.

HTT event preferentially originate from the same native realm. We also accounted for the fact that insect lineages may have moved to different geographic realms after exchanging TEs. This analysis required estimating the time since a transfer event. As proxies for this time, we took two independent measures of nucleotide divergence between TE copies originating from a HTT event: between- and within-lineage divergence. If insect lineages tend to move after exchanging TEs, the geographic cooccurrence of species that share TEs due to HTT should decrease with the estimated age of the transfer. Observations followed these predictions (Fig. 4B). Not only did HTT preferentially involve species belonging to the same realms overall, this tendency was more pronounced for the most recent transfers. These results confirm that geographic proximity generally favors HTT in insects. To some extent, this observation is reminiscent of that found in bacteria, where HGT occurs more frequently between lineages sharing the same habitat (25). Furthermore, our analysis likely underestimates the extent to which transfer events preferentially occur within geographic realms because sampled species may be more or less distantly related to, and geographically distant from, those that directly exchanged TEs.

Strikingly, the geographic cooccurrence of species sharing horizontally transferred TEs negatively correlated with the estimated time since the transfers (Fig. 4B). The correlation with the time inferred from within-lineage divergence of TEs is more

significant ( $P < 0.004$ , permutation test) than with the time inferred from between-lineage divergence ( $p \sim 0.06$ ), possibly due to better accuracy of the former estimate (*Materials and Methods*). This correlation strongly suggests that older HTT events involve lineages whose descendants are more likely to be found in different realms, as expected if insect lineages had more time to migrate after exchanging TEs. Hence, historical and geographical patterns of HTs offer a rare illustration of the global movement of lineages across biogeographic realms over the past few million years.

Despite the conservative constraints imposed by our approach to detect HTT, we found that 2.08% of the nucleotides contained in insect genomes on average, and up to 24.6% in some species (*Dataset S1*), derived from the activity of TEs that were horizontally transferred within just the last  $\sim 10$  My. Extrapolating our estimates over the  $\sim 480$  My of insect evolution and the whole insect biodiversity points toward millions of HTT events generating substantial fractions of insect genomes. These inferences, combined with the pronounced impact TEs have on genome structure and dynamics (33, 34), establish HTT as a major factor driving insect molecular evolution. Our results call for further assessments of the influence of HTT on other taxonomic groups and of the ecological factors and relationships affecting HTT dynamics (35).



**Fig. 4.** Geographic proximity facilitates HTTs among insects. (A) Biogeographic realms to which 179 of the 195 studied insect species could be assigned, excluding Antarctica. The area of a disk is proportional to the number of species assigned to the corresponding realm, which is indicated inside the disk. Transfers involving species from different realms are represented by curves, and transfers involving species from the same realms are presented by circles, the thickness of which is proportional to the mean number of transfer per species pair. (B) Transfers more frequently involve species from the same realm, and significantly so for more recent transfers. Time since transfers is approximated by two measures of nucleotide divergence of TEs. For 10 defined classes of TE divergence (x axis), we plotted the fraction of transfers involving species from the same realms, divided by the mean of its null distribution obtained by permuting species across realms  $10^4$  times. Asterisks represent the portion ( $P$ ) of permutations that yielded higher values than observations. \* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.001$ . Class boundaries are shown by dashed lines and were defined so that classes encompass similar numbers of species pairs, to provide more comparable statistics.

## Materials and Methods

**Source Data and Time-Tree Construction.** We used the latest genome assemblies of 195 insect species (Dataset S1) at the contig level. These assemblies constitute all of the publicly available reference genome sequences of insects (Insecta) as of May 2016, excluding species for which the assembly size appeared too short. A time tree of these species (Fig. S1) was manually constructed by setting node ages to match divergence times obtained from [timetree.org/](http://timetree.org/) (36), using dates established by Misof et al. (23) when available.

The following steps were implemented in R scripts (37) calling other programs. Unless specified otherwise, program and function arguments were left at their default values, and homology searches used blast+ (38) algorithms, retaining only the best alignment per query.

**Extraction of TEs from Genomes and Homology Search.** TE family consensus sequences were generated by RepeatModeler (18), setting "ncbi" as the search engine, and were provided as a custom library to RepeatMasker (39) to locate associated TE copies in each species' genome, ignoring low complexity regions (option "-nolow"). Copies >100 bp were extracted from genomic contigs by using the Biostrings R package (40).

Each homology search was performed with the megablast algorithm. It used a given species' TE copies as query and another species' copies as target. This represented 37,830 searches (195<sup>2</sup> – 195; that is, avoiding self-comparisons). In the following, a "hit" refers to an alignment (or high-scoring segment pair; HSP) resulting from this initial megablast search.

**Defining Insect Lineages Among Which Hits Should Not Result from Vertical Inheritance of TEs.** To compare interspecific divergence at TEs to synonymous divergence of genes, we located core genes in each genome using the BUSCO pipeline (41) and its database of ancestral arthropod proteins. We concatenated exons into coding sequences (CDSs) based on coordinates reported for each complete gene. We used Megan 6 (42) to select translated CDSs that had the best homology to known arthropod proteins, and among this selection, we excluded proteins that had homologies to TEs (with an *e*-value of at most 10<sup>-3</sup>). These homologies were established by Diamond blastp searches (43) against the nonredundant protein database of National Center for Biotechnology Information (NCBI) and the TE database RepBase (44), respectively.

Protein sequences were compared between every two species by using reciprocal blastp searches, with an *e*-value threshold of 10<sup>-4</sup>. Proteins involved in reciprocal best hits and corresponding to the same ancestral protein (same BUSCO identifier) were considered orthologous. Alignments of <100 amino acids and between nonorthologs were discarded. We realigned the pair of protein regions covered by each hit with the pairwiseAlignment() function of Biostrings (40) and translated the resulting alignment into a nucleotide alignment with a custom R function. Rates of synonymous substitution (*dS*) between orthologous CDS were computed with Li's method (45), as implemented by the kaks() function of package seqinr (46).

The distribution of *dS* for each insect clade was established on values obtained from all pairs of species spanning its two immediate subclades. To avoid pseudoreplication in *dS* values between a given CDS and all orthologs from the other subclade, we only used the *dS* value corresponding to the longest alignment of each CDS. A clade was collapsed (all TE homologies within it were ignored) if >0.3% of the *dS* values of orthologous core genes were lower than the highest divergence between TEs that we computed as described below.

Nucleotide divergence at horizontally transferred TEs was established on a random sample of 400,000 megablast hits obtained from pairs of species that diverged in the last 40 My (hence likely representing nonvertical transfers). We realigned TE regions based on the HSP coordinates using Biostrings and computed the distance between copies according to Kimura's two-parameter model (47), which is the model of substitution used by Li's method (45).

**Identification of Independent HTT Events.** Candidate transfer events were identified by clustering hits involving a given pair of insect lineages and TEs from the same superfamily, because hits between TEs from different superfamilies were discarded. See the *SI Discussion* and Fig. S3 for more detail on the clustering approach we used.

We first reduced the number of hits to obtain a manageable number of pairwise comparisons (*SI Materials and Methods*). Every two hits were "connected" if identity of TE copies within one lineage was equal to or higher than at least one of the two between-lineage identities associated with the hits. Within-lineage identity was assessed by a blastn homology search of all TE copies from the same lineage against themselves (i.e., set as both query and target) authorizing all hits for a given query. Alignments <100 bp were not recorded, and identity was considered as zero in that case.

The resulting connections produced an undirected graph of hits, in which clusters were delineated by the algorithm (48) implemented in the cluster\_fast\_greedy() function of the igraph package (49), which maximizes within-cluster connectivity and minimizes between-cluster connectivity (*SI Materials and Methods* and Fig. S4 and S5). Across all TE superfamilies and lineage pairs, this yielded 8,713 clusters of hits.

To test whether any two clusters *i* and *j* represented the retention of nonoverlapping parts of an ancestral TE instead of separate transfer events, we compared protein regions identified in the TEs they involved (*SI Materials and Methods*). Clusters *i* and *j* were considered to represent separate HTTs if they had low connectivity (*SI Materials and Methods* and Fig. S5) and if protein regions overlapped by at least 100 amino acids (Fig. S6). Otherwise, these clusters were "connected". Applying connections to every pair of clusters yielded an undirected graph of clusters where every HTT event would constitute either an unconnected cluster or a "clique" of clusters (Fig. S4B). A clique is a network whose elements (here, clusters of hits) are all directly connected (adjacent) to each other. Cliques were delineated by an algorithm (*SI Materials and Methods* and Dataset S3) implemented in an R function. This clustering resulted in 1,535 cliques and 5,340 unconnected clusters. We collectively refer to those as transfers or HTTs below.

To reduce the risk of cross-contamination of DNA between species seen as HTT, we imposed that the TE families involved in a transfer be represented, in each lineage, by at least five TE copies measuring at least half the length of their respective consensus. We further imposed that at least two of these copies, for each lineage, be present in the retained megablast hits.

**Minimum Number of HTT Events.** The minimum number of HTT events, considering all insect lineages, was counted by establishing networks of lineages connected by transfers of similar TEs (Fig. S7). In such a network, every apparent transfer between two lineages may result from two acquisitions of TEs from (an)other lineage(s), which, according to parsimony, are already represented by transfers in the network. However, two transfers that were previously identified as independent, and involving the same pair of lineages, cannot both result from the same two acquisitions, and should be in different networks. To establish networks, every two transfers were connected if at least one given TE family was involved in both transfers and if these were not previously characterized as independent (*SI Materials and Methods* and Fig. S7). From the resulting graph, networks were delineated by single-linkage clustering. To avoid considering independent transfers in the same network, we split the network into cliques that cannot contain independent transfers (*SI Materials and Methods*).

**Dating HTT Events.** We approximated the time since a transfer by the minimum between-lineage nucleotide divergence of copies resulting from a HTT. This proxy may overestimate the age of the transfer under a scenario where the two lineages considered have not directly exchanged TEs (and acquired these from a third party) or where the sampled species diverged from the donor of TEs before the transfer.

We thus used another proxy, based on the divergence of TEs within the supposed recipient lineage. This measure may underestimate the age of TE acquisition, but is less influenced by the species pair used to date the transfer. Within-lineage divergence associated to a transfer was taken, for each of the two lineages involved, as the ninth decile of the raw nucleotide divergence between TE copies included in the corresponding cluster of hits, which we previously estimated by blastn searches. We used the ninth decile rather than the maximum, because the latter would have put high weight on the two copies that diverged the most. To consider that TEs may have diverged within one of the two lineages (the donor) before the transfer, we used the lower decile value among the two. If the transfer was a clique of several cluster of hits, we used the value obtained from the cluster that comprised the hit having the highest identity, for consistency with the estimate based on between-lineage divergence.

**Analysis of Biogeographic Data.** Native biogeographic realms of 179 insect species (Dataset S1) were obtained from several Internet sources. Within a lineage, closely related species occupying distinct realms may appear involved in the same HTT, due to speciation before or after the transfer. The two realms we associated to each HTT were those of the two species (one per lineage) that yielded the hit of highest identity, which we used to date the transfer (as described above). To avoid counting the same HTT several times, all other species were considered not involved in the transfer. This selection is equivalent to a random draw among the species descending from an ancestor that acquired or emitted TEs, should speciation have occurred after the transfer.

Correlation between time since a transfer and cooccurrence of the species involved (defined as originating from the same realm and encoded as a binary

value) was estimated by Pearson's  $R$  ( $n = 3,863$  transfers between located species). To test its significance, we computed  $R$   $10^4$  times after randomly permuting realms across species and compared it to the value obtained from real data.

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1. Soucy SM, Huang J, Gogarten JP (2015) Horizontal gene transfer: Building the web of life. *Nat Rev Genet* 16:472–482.
2. Frost LS, Leplae R, Summers AO, Toussaint A (2005) Mobile genetic elements: The agents of open source evolution. *Nat Rev Microbiol* 3:722–732.
3. Ku C, Martin WF (2016) A natural barrier to lateral gene transfer from prokaryotes to eukaryotes revealed from genomes: The 70 % rule. *BMC Biol* 14:89–100.
4. Keeling PJ (2009) Functional and ecological impacts of horizontal gene transfer in eukaryotes. *Curr Opin Genet Dev* 19:613–619.
5. Richards TA, et al. (2011) Horizontal gene transfer facilitated the evolution of plant parasitic mechanisms in the oomycetes. *Proc Natl Acad Sci USA* 108:15258–15263.
6. Szöllösi GJ, Davin AA, Tannier E, Daubin V, Boussau B (2015) Genome-scale phylogenetic analysis finds extensive gene transfer among fungi. *Philos Trans R Soc Lond Ser B Biol Sci* 370:20140335.
7. Graham LA, Li J, Davidson WS, Davies PL (2012) Smelt was the likely beneficiary of an antifreeze gene laterally transferred between fishes. *BMC Evol Biol* 12:190–202.
8. Christin PA, et al. (2012) Adaptive evolution of C(4) photosynthesis through recurrent lateral gene transfer. *Curr Biol* 22:445–449.
9. Wallau GL, Ortiz MF, Loreto EL (2012) Horizontal transposon transfer in eukarya: detection, bias, and perspectives. *Genome Biol Evol* 4:689–699.
10. Dotto BR, et al. (2015) HTT-DB: Horizontally transferred transposable elements database. *Bioinformatics* 31:2915–2917.
11. El Baidouri M, et al. (2014) Widespread and frequent horizontal transfers of transposable elements in plants. *Genome Res* 24:831–838.
12. Craig NL, Craigie R, Gellert M, Lambowitz AM (2002) *Mobile DNA II* (AMS Press, Washington, DC), p 1204.
13. Schaack S, Gilbert C, Feschotte C (2010) Promiscuous DNA: Horizontal transfer of transposable elements and why it matters for eukaryotic evolution. *Trends Ecol Evol* 25:537–546.
14. Ivancevic AM, Walsh AM, Kortschak RD, Adelson DL (2013) Jumping the fine LINE between species: Horizontal transfer of transposable elements in animals catalyses genome evolution. *BioEssays* 35:1071–1082.
15. Dupeyron M, Leclercq S, Cerveau N, Bouchon D, Gilbert C (2014) Horizontal transfer of transposons between and within crustaceans and insects. *Mob DNA* 5:4–13.
16. Wallau GL, Capy P, Loreto E, Le Rouzic A, Hua-Van A (2016) VHICA, a new method to discriminate between vertical and horizontal transposon transfer: Application to the mariner family within *Drosophila*. *Mol Biol Evol* 33:1094–1109.
17. Maumus F, Fiston-Lavier AS, Quesneville H (2015) Impact of transposable elements on insect genomes and biology. *Curr Opin Insect Sci* 7:30–36.
18. Smit AFA, Hubley R (2015) RepeatMasker Open-1.0. Available at [www.repeatmasker.org/](http://www.repeatmasker.org/).
19. Vassetzky NS, Kramerov DA (2013) SINEBase: A database and tool for SINE analysis. *Nucleic Acids Res* 41:D83–D89.
20. Lampe DJ, Witherspoon DJ, Soto-Adames FN, Robertson HM (2003) Recent horizontal transfer of mellifera subfamily mariner transposons into insect lineages representing four different orders shows that selection acts only during horizontal transfer. *Mol Biol Evol* 20:554–562.
21. Bartolomé C, Bello X, Maside X (2009) Widespread evidence for horizontal transfer of transposable elements across *Drosophila* genomes. *Genome Biol* 10:R22.
22. Robertson HM (2002) Evolution of DNA transposons in eukaryotes. *Mobile DNA II*, ed Craig NLea (ASM, Washington, DC), pp 1093–1110.
23. Misof B, et al. (2014) Phylogenomics resolves the timing and pattern of insect evolution. *Science* 346:763–767.
24. Hooper SD, Mavromatis K, Kyrpides NC (2009) Microbial co-habitation and lateral gene transfer: What transposases can tell us. *Genome Biol* 10:R45–44.
25. Popa O, Dagan T (2011) Trends and barriers to lateral gene transfer in prokaryotes. *Curr Opin Microbiol* 14:615–623.
26. Wagner A, de la Chaux N (2008) Distant horizontal gene transfer is rare for multiple families of prokaryotic insertion sequences. *Mol Genet Genomics* 280:397–408.
27. Lampe DJ, Churchill ME, Robertson HM (1996) A purified mariner transposase is sufficient to mediate transposition in vitro. *EMBO J* 15:5470–5479.
28. Ivics Z, Izsvak Z (2015) Sleeping beauty transposition. *Microbiol Spectrum* 3:MDNA3-0042-2014.
29. Levin HL, Moran JV (2011) Dynamic interactions between transposable elements and their hosts. *Nat Rev Genet* 12:615–627.
30. Peddigari S, Li PW, Rabe JL, Martin SL (2013) hnRNP and nucleolin bind LINE-1 RNA and function as host factors to modulate retrotransposition. *Nucleic Acids Res* 41:575–585.
31. Hartl DL, Lohe AR, Lovozkaya ER (1997) Modern thoughts on an ancient mariner: Function, evolution, regulation. *Annu Rev Genet* 31:337–358.
32. Silva JC, Loreto EL, Clark JB (2004) Factors that affect the horizontal transfer of transposable elements. *Curr Issues Mol Biol* 6:57–71.
33. Cordaux R, Batzer MA (2009) The impact of retrotransposons on human genome evolution. *Nat Rev Genet* 10:691–703.
34. Feschotte C, Pritham EJ (2007) DNA transposons and the evolution of eukaryotic genomes. *Annu Rev Genet* 41:331–368.
35. Venner S, et al. (2017) Ecological networks to unravel the routes to horizontal transposon transfers. *PLoS Biol* 15:e2001536.
36. Hedges SB, Marin J, Suleski M, Paymer M, Kumar S (2015) Tree of life reveals clock-like speciation and diversification. *Mol Biol Evol* 32:835–845.
37. R Development Core Team (2016) *R: A Language and Environment for Statistical Computing* (R Foundation for Statistical Computing, Vienna, VA).
38. Camacho C, et al. (2009) BLAST+: Architecture and applications. *BMC Bioinformatics* 10:421–429.
39. Smit AFA, Hubley R, Green P (2015) RepeatMasker Open-4.0. Version 4.0. Available at [www.repeatmasker.org/](http://www.repeatmasker.org/).
40. Pagès H, Aboyoun P, Gentleman R, DebRoy S (2016) Biostings: String objects representing biological sequences, and matching algorithms. R package version 2.40.0.
41. Simão FA, Waterhouse RM, Ioannidis P, Kriventseva EV, Zdobnov EM (2015) BUSCO: Assessing genome assembly and annotation completeness with single-copy orthologs. *Bioinformatics* 31:3210–3212.
42. Huson DH, Auch AF, Qi J, Schuster SC (2007) MEGAN analysis of metagenomic data. *Genome Res* 17:377–386.
43. Buchfink B, Xie C, Huson DH (2015) Fast and sensitive protein alignment using DIAMOND. *Nat Methods* 12:59–60.
44. Bao W, Kojima KK, Kohany O (2015) Repbase Update, a database of repetitive elements in eukaryotic genomes. *Mob DNA* 6:11–16.
45. Li WH (1993) Unbiased estimation of the rates of synonymous and nonsynonymous substitution. *J Mol Evol* 36:96–99.
46. Charif D, Lobry JR (2007) Seqin(R) 1.0-2: A contributed package to the {R} project for statistical computing devoted to biological sequences retrieval and analysis. Structural Approaches to Sequence Evolution: Molecules, Networks, Populations, eds Bastolla U, Porto M, Roman HE, Vendruscolo M (Springer, New York), pp 207–232.
47. Kimura M (1980) A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *J Mol Evol* 16:111–120.
48. Clauset A, Newman MEJ, Moore C (2004) Finding community structure in very large networks. *Phys Rev E Stat Nonlin Soft Matter Phys* 70:066111.
49. Csardi G, Nepusz T (2006) The igraph software package for complex network research. *InterJournal Complex Systems*:1695.