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1	Assessment of background concentrations of organometallic compounds
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26 ABSTRACT

27 The aim of this work is to estimate background concentrations of organometallic compounds, such as tributyltin (TBT), dibutyltin (DBT), monobutyltin (MBT), triphenyltin (TPhT), 28 diphenvltin (DPhT), monophenvltin (MPhT), methylmercury (MeHg), inorganic mercury 29 (iHg) and diethyllead (Et₂Pb) in the aquatic environment at the French national scale. 30 Both water and sediment samples were collected all over the country, resulting in 152 water 31 samples and 123 sediment samples collected at 181 sampling points. Three types of surface 32 water bodies were investigated: rivers (140 sites), lakes (19 sites) and coastal water (42 sites), 33 spread along the 11 French river basins. The choice of sites was made on the basis of previous 34 investigation results and the following target criteria: reference, urban sites, agricultural and 35 industrial areas. The analytical method was properly validated for both matrices prior to 36 analysis, resulting in low limits of quantification (LOQ), good precision and linearity in 37 38 agreement with the Water Framework Directive demands. The results were first evaluated as a function of their river basins, type of surrounding pressure and water bodies. Later, 39 40 background concentrations at the French national scale were established for both water and sediment matrices, as well as their threshold, i.e., the concentration that distinguishes 41 background from anomalies or contaminations. Background concentrations in water are 42 43 8.43 ng Pb. L^{-1} for Et₂Pb and 0.49 – 151 ng Sn. L^{-1} , <0.15 – 3.04 ng Sn. L^{-1} and <0.25 ng Sn. 44 L^{-1} for MBT, DBT and TBT, respectively. For sediments, background concentrations were 45 set as <0.09 - 1.11 ng Hg. g⁻¹ for MeHg, <0.06 - 24.3 ng Pb. g⁻¹ for Et₂Pb and <2.0 - 13.4 ng 46 Sn. g^{-1} , <1.6 – 8.54 ng Sn. g^{-1} , <0.6 – 1.16 ng Sn. g^{-1} and <0.02 – 0.61 ng Sn. g^{-1} for MBT, 47 DBT, TBT and DPhT, respectively. TBT occurs in higher concentrations than the available 48 49 environmental protection values in 24 and 38 sampling sites for both water and sediment samples, respectively. Other phenyltins (MPhT and TPhT) did not occur above their LOQ and 50

51	therefore no background was possible to establish. Throughout this work, which is the first
52	assessment of background concentrations for organometallic compounds at the French
53	national level ever being published, it was possible to conclude that over the last 10 to 20
54	years organotin concentrations in French river basins have decreased while MeHg
55	concentration remained stable.
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58	Keywords: organometallic compounds; speciation analysis; screening study; background
59	concentration; aquatic environment
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62	1. INTRODUCTION
63	The Water Framework Directive (WFD) requires Member States of the European Union (EU)
64	to monitor at river basin level an established list of substances. Despite these provisions, it is
65	widely recognized by the scientific community that several substances of emerging concern
66	are currently overlooked and as a result they are not adequately monitored by national
67	authorities (Dulio and Andres, 2012). As part of the implementation of the National Action
68	Plan on Micropollutants in the Aquatic Environment, the French Ministry of Ecology decided
69	to implement an innovative and comprehensive approach to improve national monitoring
70	programs (Dulio and Andres, 2012). This includes the setting up of a watch list of substances
71	to be investigated at the national level in order to acquire missing information about the level
72	of exposure of emerging contaminants in the aquatic environment and allow regular updating
73	of the list of River Basin-Specific Pollutants to be regularly monitored. As part of this action
74	plan, a large national screening study took place in 2012 in France, where organometallic
75	compounds, among others, were monitored (ONEMA, 2012).

Organotin compounds (OTC) were extensively used as powerful biocides in anti-fouling 76 paints and pesticide formulations (Graceli et al., 2013; Hoch, 2001) particularly tributyltin 77 (TBT) and triphenyltin (TPhT), until their complete prohibition in January 1st, 2008 78 79 (Comission, 2003; Dafforn et al., 2011). These compounds enter the aquatic ecosystems through landfill leaching (Pinel-Raffaitin et al., 2008; Yi et al., 2012), the constant leach rate 80 of the antifouling paint, its scrap off due to ship grounding or ice-breakers activities, or even 81 due to cleaning activities in shipyards (Du et al., 2014), and therefore most occurrence studies 82 refer to marine and coastal environments, where concentrations range from non-detected to 83 197 ng Sn.L⁻¹ for water samples and from non-detected to 7673 ng Sn.g⁻¹ dw for sediments 84 85 from European harbors (Antizar-Ladislao, 2008). Studies about their volatilization and atmospheric deposition (Amouroux et al., 2000; Huang et al., 2004) suggest that these 86 compounds should also be monitored upstream, in lakes and rivers, where they are expected 87 88 to occur in considerably lower concentrations. Mercury is also widely spread in aquatic ecosystems, not only as a result of anthropogenic 89 activities, but also by natural earth processes (Hsu-Kim et al., 2013). Inorganic mercury (iHg) 90 91 is biomethylated by microorganisms originating methyl mercury (MeHg) which is known to

92 be extremely toxic and bioaccumulate along the food web to potentially harmful

93 concentrations, while, on the other hand, the toxicological role of iHg is still under debate

94 (Clarkson, 2002). When lacking anthropogenic and geological sources, total mercury

95 concentrations in water samples range from 0.3 to 8 ng Hg.L⁻¹ and, in the presence of

96 industrial pollution and gold mining activities, concentrations can vary between 10 and 40 ng

97 Hg.L⁻¹ (Wiener et al., 2003).

Organolead compounds (OLC) were introduced in the early 1920s as antiknocking agents for
gasoline engines and remained in use for 55 years before the beginning of their phase-out in
Japan and USA due to their neurotoxic effects (Craig, 2003). Today, and according to data

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101	from April 2015 available on the United Nation Environment Programme (UNEP) website,
102	there are still 6 countries in the world that use leaded gasoline (UNEP, 2015). As the most
103	important source of lead in urban environments is highway runoff water, due to the gasoline
104	lead deposited onto roads (Pattee and Pain, 2003), concentrations of triethylead (Et ₃ Pb) ranges
105	from non-detects to 400 ng Pb.L ⁻¹ in these environments, and its degradation product,
106	diethyllead (Et ₂ Pb), ranges from non-detected to 1800 ng Pb.L ⁻¹ (Craig, 2003), being the two
107	most frequent organolead species to be detected and quantified in water samples. In
108	sediments, ethylated species are only sporadically detected in concentrations lower than 1 ng
109	Pb.L ⁻¹ (Craig, 2003).
110	REACH (Registration, Evaluation, Authorization and Restriction of Chemicals) is the
111	European Union regulation made to spread awareness about the safe use of chemicals, in
112	order to protect the environment and human health from risks posed by those chemicals. The
113	Predicted No Effect Concentration (PNEC) is the international environmental protection
114	values developed in the context of REACH legislation and can be defined as the concentration
115	below which a specified percentage of species in an ecosystem is expected to be protected
116	(Pennington, 2003), usually 95 %. In risk assessment studies, if the measured concentrations
117	are higher than the environmental protection values, the ecosystem may be classified as at risk
118	of environmental damage. A completely different approach to assess the environmental
119	quality of a given area is by comparing with established background concentration levels.
120	This approach is supported by some authors who claim that the transposition of
121	ecotoxicological results to regulatory levels is difficult, as it might set these regulatory levels
122	down to natural background concentrations and consequently lead to unnecessary clean-up to
123	levels below the natural occurring concentrations, resulting in severe financial and
124	environmental consequences (Reimann and Garrett, 2005).

The term "background" has not yet been defined precisely although this subject is object of 125 several discussions (Ander et al., 2013; Reimann and Garrett, 2005). The US Environmental 126 Protection Agency (USEPA) defines background levels as it is interpreted in this study: "the 127 concentration of a hazardous substance that provides a defensible reference point that can be 128 used to evaluate whether or not a release from the site has occurred" and which do not have to 129 reflect pristine conditions (EPA, 1995). Several authors found more useful to present 130 background levels as a concentration range (minimum, 25th percentile, median, 75th 131 percentile, maximum), and not as a single value usually called "baseline" (Reimann et al., 132 2005; Vestenius et al., 2011). As important as establishing background concentrations it is to 133 determine its threshold, i.e., the upper limit which differentiates background variability from 134 anomalies and defines when action should be taken (Reimann et al., 2005). 135 The aim of this work is to establish background concentrations of several organometallic 136 137 substances with a potential risk for the aquatic environment in the French territory and study the occurrence. For this reason, the substances included in this screening study were 138 monitored at least at the PNEC (predicted no-effect concentration) levels and the background 139 concentrations of the occurring target substances were defined, as well as their threshold. It is 140 also important to highlight that this is the first assessment of organometallic compounds' 141 occurrence in French watersheds at the national level. 142

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- 145 2. EXPERIMENTAL
- 146 2.1. Water and sediment sampling strategy

147 Three types of water bodies were investigated in Spring of 2012 in order to reflect the

structure of the permanent monitoring network: rivers (140 sites), lakes (20 sites) and coastal

sites (40 sites). However, coastal water samples were not sampled due to unsteady physical-

chemical conditions typical in estuaries and coastal environments and related sampling 150 logistic limitations. The sampling sites, some of which are illustrated on Figure 1, were 151 selected by French Water Agencies and selection criteria were defined by the INERIS at 152 national level, not only in metropolitan water basins (Adour-Garonne, Artois-Picardie, Loire-153 Bretagne, Rhône-Méditerranée and Corsica, Rhin-Meuse, Seine-Normandie), but also in 154 overseas territories (Guadeloupe, French Guiana, Mayotte, Martinique and Réunion). 155 Agricultural, urban and industrial pressures were considered for the selection of the sampling 156 sites; additionally, reference sites (no anthropogenic pressures) and some sites classified as 157 "poor ecological status" according to the definitions of the Water Framework Directive, were 158 also included in the study. The choice of sites was made on the basis of previous investigation 159 results and the following target criteria: at least 20 % of reference sites and 16 % of sites 160 where conditions for good ecological status are not achieved. The remaining 64 % were 161 162 chosen based on land use: urban sites, sites for agricultural activities or breeding and industrial areas. The effect of seasonal variability was not addressed and will not be 163 discussed. 164

Surface river bed sediment samples (1-2 cm) were collected within an area close to the river bank, pooled in an hermetic glass jar to avoid aeration and stored at 4 °C until further treatment (see Sample preparation section). The wet river bed sediment samples were sieved at 2 mm, then freeze-dried and sieved at 250 μ m, grinded, homogenized and stored in amber flasks in a cold room (4 °C).

Water sampling was performed directly into 250 mL amber borosilicate bottles (VWR,
Øxh:64x119 mm, Ø int. col: 38) or with the help of a pump or an integrating water-sampler,
and shipped directly for analysis by fast courier within 24 hours (maximum 48 h) in
thermostatic boxes (4 °C). For overseas territories, samples arrived by fast courier all within
48 hours (maximum 96 h). Every water sample was filtered upon arrival at the laboratory and

175 further acidified to 2 % with pure glacial acetic acid and placed in the storage cold room at 4 176 °C until sample preparation. Filtration was performed under a laminar flow hood (Class 100) 177 in an acid–cleaned polysulfonate vacuum filtration unit from Sartorius (Aubagne, France) and 178 under pyrolyzed 0.7 μ m GF/F Whatmann filters (from Sigma-Aldrich in Lyon, France). The 179 250-mL borosilicate bottles were not cleaned prior to sampling because a previous blank 180 storage test was performed and the obtained blank were similar to regular blank values.

181

182 2.2. Speciation analysis of organometallic compounds

183 <u>2.2.1. Chemicals</u>

TBT chloride (96%), DBT dichloride (97%), MBT trichloride (95%) and iHg (II) chloride 184 (≥99.5%) salts were obtained from Sigma-Aldrich, whereas TPhT chloride (98%), DPhT 185 dichloride (96%), MPhT trichloride (95%) and the MeHg chloride standard (98.4%) were 186 187 obtained from Strem Chemicals (Newburyport, MA, USA). Triethyllead chloride (Et₃PbCl, 82.3%) was purchased from Dr. Ehrenstorfer (LGC Standards, Molheim, France). No other 188 organolead standards (such as Et₂Pb) are commercially available, therefore all validation was 189 performed for Et₃Pb and extrapolated to Et₂Pb. The isotopically enriched species used were 190 purchased from ISC Science (Oviedo, Spain): iHg enriched in ¹⁹⁹ Hg (¹⁹⁹iHg, 91%) at 10 µg.g⁻ 191 ¹, MeHg enriched in ²⁰¹ Hg (²⁰¹MeHg, 96.5%) at 5.494 μ g.g⁻¹ and a mix of MBT, DBT and 192 TBT enriched in ¹¹⁹ Sn (82.4%, ¹¹⁹MBT, ¹¹⁹DBT, ¹¹⁹TBT) at 0.110, 0.691 and 1.046 µg.g⁻¹, 193 respectively. 194

A mixture of MPhT, DPhT and TPhT enriched in ¹¹⁶Sn (¹¹⁶MPhT, ¹¹⁶DPhT, ¹¹⁶TPhT) was
synthesized by phenylation of metallic tin obtained from Oak Ridge National Laboratory
(Oak Ridge, TN, USA) according to a method described elsewhere (Kumar et al., 2004).
Purity and concentrations in this methanol solution were the following: ¹¹⁶MPhT (90.2%) at
4.2 µg.L⁻¹, ¹¹⁶DPhT (97.4%) at 48.5 µg.L⁻¹ and ¹¹⁶TPhT (97.5%) at 677 µg.L⁻¹. In order to

200	control any possible degradation or interconversion of species, this solution was kept in the				
201	freezer and its concentration regularly determined by reverse isotope dilution.				
202	Hydrochloric acid (HCl, 33–36%, ultrexII ultrapure reagent) and glacial acetic acid (HOAc,				
203	Instra-analyzed) were purchased from J.T. Baker (Phillipsburg, NJ, USA). Methanol (MeOH,				
204	\geq 99.9% Chromasolv) and isooctane, or 2,2,4-trimethylpentane (ACS reagent, \geq 99.0%) were				
205	obtained from Sigma-Aldrich (Seelze, Germany). Ammonium hydroxide (NH ₄ OH, \geq 25% in				
206	H ₂ O, <i>TraceSELECT[®]</i> Ultra, for ultratrace analysis) was purchased from Fluka (Steinheim,				
207	Germany) and sodium acetate trihydrate (NaOAc, puriss p.a.) from Riedel-de-Haën (Seelze,				
208	Germany). Sodium tetrapropylborate (NaBPr ₄) (98%) was purchased from Merseburger				
209	Spezialchemikalien (Schkopau, Germany).				
210	All stock solutions (1000 μ g.g ⁻¹ as Sn, Hg or Pb) were prepared by dissolving the				
211	corresponding salt either in a 3:1 mixture of acetic acid/methanol for OTC, methanol for				
212	MeHg and Et ₃ Pb, 1% HCl in ultrapure water for iHg and were kept in the dark at 4 $^{\circ}$ C until				
213	use. Working solutions of the organo-tin, -mercury and -lead compounds were prepared daily				
214	before analysis by dilution of the stock solutions with 1% HCl in ultrapure water.				

215

216 <u>2.2.2. Water sample preparation</u>

Water samples were treated according to Monperrus et al. (Monperrus et al., 2005). Briefly, 5 217 mL of HOAc/NaOAc buffer (pH 4.8, 0.1 mol.L⁻¹) was added to a flask containing 100 mL of 218 water sample as well as 5 µL (gravimetrically controlled) of the isotopically enriched spike 219 solution (102.8 µg Hg.L⁻¹ for ¹⁹⁹iHg, 92.4 µg.L⁻¹ for ²⁰¹MeHg, 9.3 µg Sn.L⁻¹ for ¹¹⁹MBT, 34.6 220 μ g Sn.L⁻¹ for ¹¹⁹DBT, 35.6 μ g Sn.L⁻¹ for ¹¹⁹TBT, 4.2 μ g Sn.L⁻¹ for ¹¹⁶MPhT, 48.5 μ g Sn.L⁻¹ 221 for ¹¹⁶DPhT and 677 μ g Sn.L⁻¹ for ¹¹⁶TPhT) for the simultaneous analysis of organometallic 222 species. The pH was then re-adjusted with ultrapure NH₄OH to pH 4.8 and 1 mL of isooctane 223 and 1 mL of 1% (w/v) NaBPr₄ were added. Propylation occurred during vigorous shaking of 224

- the sample for 5 min and the organic phase was then transferred into a 1.5-mL vial kept at -20
 °C until analysis.
- 227
- 228 <u>2.2.3. Sediment sample preparation</u>

The extraction of the organometallic species from the sediments was carried out following the 229 guidelines in Pacheco-Arjona et al. (Pacheco-Arjona et al., 2008) with a few changes. Briefly, 230 an Explorer focused microwave system from CEM Corporation (Mathews, NC, USA) was 231 used to extract the organometallic species from the sediment by adding 5 mL of the mixture 232 of HOAc: methanol (3:1, v/v) to 0.25 g of sediment and stirring for 3 min at 70 °C. The 233 extracts were then centrifuged at 2300 rpm for 3 minutes and 1 mL of the extracted solution 234 was added to 22-mL glass vials containing 5 mL of HOAc/NaOAc buffer (pH 4.8, 0.1 mol.L⁻ 235 ¹). Ten μ L (gravimetrically controlled) of the isotopically enriched spike solution (192.0 μ g 236 Hg.L⁻¹ for ¹⁹⁹iHg, 0.94 µg.L⁻¹ for ²⁰¹MeHg, 27.4 µg Sn.L⁻¹ for ¹¹⁹MBT, 23.5 µg Sn.L⁻¹ for 237 ¹¹⁹DBT, 25.2 μg Sn.L⁻¹ for ¹¹⁹TBT, 16.6 μg Sn.L⁻¹ for ¹¹⁶MPhT, 77.7 μg Sn.L⁻¹ for ¹¹⁶DPhT 238 and 377.5 μ g Sn.L⁻¹ for ¹¹⁶TPhT) was added, the pH was re-adjusted with ultrapure NH₄OH to 239 pH 4.8 and 1 mL of isooctane and 1 mL of 1% (w/v) NaBPr₄ were added to each vial. The 240 mixture was then manually and vigorously shaken for 5 min to allow propylation to occur. 241 The organic phase was retrieved and transferred into a 1.5 mL vial which was kept at -20 °C 242 until analysis. 243

244

245 2.3. GC-ICP/MS analysis

A Thermo X Series 2 inductively coupled plasma-mass spectrometer (ICP/MS) coupled to a
gas chromatograph (GC) (Trace GC Ultra, Thermo Fisher, Waltham, MA, USA equipped
with a TriPlus RSH autosampler) was used with a commercial GC-ICP/MS interface
(SilcoSteel, 0.5 m length, outer i.d. 1.0 mm and o.d. 1.6 mm, inner i.d. 0.28 mm and o.d. 0.53

250	mm, Thermo Fisher). A MXT I column (30 m x 0.28 μ m i.d., 1 μ m, Restek Corporation,
251	Bellefonte, PA, USA) was used to perform the chromatographic separation and Table S1
252	(Supplementary Material) lists the operating conditions. The optimization of the instrument's
253	performance was made with liquid standards and tin (m/z 116, 117, 118, 119, 120), mercury
254	(m/z 199, 200, 201, 202) and lead $(m/z 206, 208)$ isotopes where measured with a 30ms dwell
255	time. The isotopes of Tl (m/z 203, 205 during 5 ms) and Sb (m/z 121, 123 during 5 ms) were
256	measured to check the mass bias in each chromatographic run.

257

258 2.4. Quantification methods

259 Diethyllead was determined by interpolation on the external calibration curve of Et_3Pb as no 260 commercial standard for diethyllead, neither isotopic tracers, are available.

261 The use of a mixed isotopic spike solution containing ¹⁹⁹iHg, ²⁰¹MeHg, ¹¹⁹MBT, ¹¹⁹DBT,

262 ¹¹⁹TBT, ¹¹⁶MPhT, ¹¹⁶DPhT and ¹¹⁶TPhT allowed to quantify organotin and organomercury

263 compounds by speciated isotope dilution mass spectrometry (SIDMS) which takes into

account the cross-species transformations (only for Hg species, in this particular case), the

loss of analytes during sample preparation and the signal drift. The isotopic spike was added

to sediment samples after the microwave extraction, as it is well known that no artifacts are

formed during this procedural step (Rodriguez Martin-Doimeadios et al., 2003; Monperrus et

al., 2003), and method validation through the use of certified reference materials was

- 269 performed.
- 270

271 2.5. QA/QC for sediment samples

272 CRM-462 (coastal sediment, certified for DBT and TBT species), the CRM-646 (freshwater

sediment, certified for Butyl-Sn and Phenyl-Sn species) and IAEA 405 (estuarine sediment,

274 certified for MeHg) were the certified reference materials used for validating the methodology

and were purchased from the Institute for Reference Materials and Measurements (IRMM,
Geel, Belgium).
For organolead species, no CRM is available; therefore, recovery tests were performed on
both CRM-462 and CRM-646 by adding different concentrations of these species (10xLOQ,

279 100xLOQ and 1000xLOQ, where LOQ is the limit of quantification) before microwave

280 extraction. Analyses proceeded as described for sediment samples.

281

282 2.6. QA/QC for water samples

In what concerns water samples, no CRM is available for organotin, organomercury and 283 organolead species. In order to verify the accuracy of the water sample analysis and the 284 stability of the equipment during the whole analytical protocol, several control samples were 285 regularly prepared. Recoveries were determined for mineral water (EvianTM mineral water) 286 287 samples spiked with natural compounds at a concentration level equivalent to the LOQ and 10 times the LOQ. Recoveries were also determined for tap water spiked with 10 times the LOQ 288 during one week of storage without acidification. These quality control samples followed the 289 290 same preparation procedure and analysis as water samples.

291

292 2.7. Data Analysis

For statistical purposes, samples with concentrations between LOQ and LOD were given their calculated value. For samples with concentrations below LOD, the concentration was set on LOD/ $\sqrt{2}$ as it is advised in Croghan et al. (Croghan and Egeghy, 2003). A significant level (p<0.05) was employed for all statistical analysis which were performed using R software from The R Project for Statistical Computing.

298

299

300 **3. RESULTS AND DISCUSSION**

301 3.1. Analytical Performances and Protocol Validation

The figures of merit of the analytical methods were evaluated and are represented in Table S2 302 (Supplementary Information). Limits of detection (LOD) and limits of quantification (LOO) 303 where calculated as 3 and 10 times the standard deviation of twelve procedural blank 304 measurements, respectively, and LOQ ranged between $0.03 - 1.0 \text{ ng.L}^{-1}$ for water samples 305 and between $0.09 - 16.6 \text{ ng.g}^{-1}$ for sediments. The European Directive 2009/90/EC requires 306 that all analytical methods applied in the context of the WFD must present a limit of 307 quantification equal or below 30 % of the relevant EQS. This analytical technique is highly 308 sensitive and efficient for the monitoring of organometallic compounds in water and 309 sediments while complying with the 2009/90/EC Directive requirements (Monperrus et al., 310 2005). However, it was found that the plastic cover of the flasks used for this monitoring 311 caused a slightly higher blanks in particular for MBT and DBT. Additionally, in this work 8 312 different species are studied and 7 isotopically enriched spike solutions are mixed in order to 313 314 quantify all the species simultaneously. As these spike solutions have isotopic abundances different from 100% (in particular the home-made ¹¹⁶phenyltin mix solution), this results in 315 degradation of the method and its analytical performances due to the compromise that a multi-316 contaminant analysis requires. Even so, these limits comply with the WFD demands as they 317 are lower than the established environmental quality standards $(1.2 \mu g.L^{-1} as annual average)$ 318 (AA) value for Pb, 0.07 μ g.L⁻¹ as maximum allowable concentration (MAC) for Hg and 319 $0.0002 \ \mu g.L^{-1}$ as AA and $0.0015 \ \mu g.L^{-1}$ as MAC for TBT). 320

321

Table S3 (Supplementary Information) shows the certified values for organotin species in
CRM-462 and CRM-646 and for MeHg in the CRM 405. The experimental results obtained
for each CRM are also represented in the Table S3.
Good agreement was found between certified values and experimentally obtained results for
organotin and organomercury species, except for MBT which usually bonds more strongly

328 with the matrix and therefore presents lower extraction recoveries in samples with high

organic matter content (Inagaki et al., 2003; Kumar et al., 2003; Ruiz Encinar et al., 2002). In

order to conclude the validation of the sediment analysis, recoveries were also determined for

both these CRM-462 and CRM-646 spiked with 10 times, 100 times and 1000 times the LOQ

for Et₃Pb because there are no CRMs available for the analysis of OLC in sediment matrices.

333 Results are showed in Table S4 (Supplementary Information). All recoveries obtained are

between 92 % and 111 %, which assures the quantitative extraction of the organolead speciesfrom the sediment matrix.

336

337 3.3. Stability tests on water samples

In order to test stability of water samples when stored in cold room, recoveries were
calculated for acidified mineral water (Evian) samples spiked at LOQ concentration level and
10 times the LOQ and results are shown in Tables 1, 2 and 3, as well as the recoveries
obtained for a non acidified tap water sample stored in cold room (4 °C) also spiked with 10
times the LOQ concentration.

343 For all species, a significant decrease in concentration is observed for non-acidified tap water

samples over time and might be due to its adsorption into the walls of the bottle (especially

345 when the species are ionic), while acidified mineral water samples keep their

346 representativeness throughout the 7 days of storage.

Even if the decreases due to adsorption in non-acidifed water are not noteworthy on the first 72 h (which is longer than the maximum delay between sample collection and sample arrival to the laboratory for metropolitan locations), water samples were filtered, transferred into the sampling recipient again and then acidified and stored in a cold room (4° C) in order to recover the maximum amount of analytes possibly adsorbed into the walls of the sampling recipients.

353

354 3.4. Occurrence of organo-tin -mercury and -lead compounds: influence of watersheds,

355 types of pressure and water bodies

The results of water and sediment samples were gathered by watershed, type of surrounding pressure and type of water body, and were represented as boxplots where the LOQs are also indicated. Data outside the 1.5 times the interquartile range were considered as outliers and were removed as recommended by the "Statistical Guidance for Determining Background Ground Water Quality and Degradation" published by the Idaho Department of Environmental Quality (Dai and Welhan, 2014) for the establishment of background

362 concentrations.

MeHg occurred (>LOD) in 86% of water samples, iHg in 99%, Et₂Pb in 79%, MBT in 100%,

364 DBT in 95%, TBT in 40%, MPhT in 13%, while DPhT and TPhT did not occur in any water

samples. In the case of sediment samples, frequency of occurrence was 90% for MeHg, 99%

- for Et₂Pb, 92% for MBT, 72% for DBT, 67% for TBT, 4% for MPhT, 54% for DPhT and
 finally 9% for TPhT.
- 368

369 <u>3.4.1. Differences in French watersheds</u>

370 The results obtained for water and sediments were displayed according to the watershed they

371 were collected from (Figure S1, Supplementary Information). Watersheds were labeled from

372	A to K, in which French metropolitan watersheds are represented from A to F and French
373	overseas departments are labeled from G to K. When observing the boxplot representation on
374	Figure S2, sediments from the departments overseas seem to be consistently lower than
375	metropolitan areas. MeHg ranged from $<0.09 - 7$ ng Hg.g ⁻¹ in metropolitan areas, while
376	remained below the LOQ in the overseas samples. A similar trend was registered for Et_2Pb
377	(metropolitan: $<0.60 - 60$ ng Pb.g ⁻¹ ; overseas: <0.60 ng Pb.g ⁻¹) and for the butyltin species
378	(MBT metropolis: $<2.0 - 50$ ng Sn.g ⁻¹ , overseas territories: $<2.0 - 8$ ng Sn.g ⁻¹ ; DBT
379	metropolis: $<1.6 - 60$ ng Sn.g ⁻¹ , overseas: $<1.6 - 7$ ng Sn.g ⁻¹ ; TBT metropolis: $<0.60 - 1.4$ ng
380	Sn.g ⁻¹ , overseas: <0.60 ng Sn.g ⁻¹). The 3 phenyltin species were only present simultaneously
381	in watershed A. For all the others, MPhT and TPhT were below their LOQ and only DPhT
382	presented data ranging between <0.20 ng Sn.g ⁻¹ – 6.0 ng Sn.g ⁻¹ .
383	This tendency is not observed in the corresponding water samples where MeHg and iHg
384	ranged from $<0.04 - 0.60$ ng Hg.L ⁻¹ and $<0.14 - 2.5$ ng Hg.L ⁻¹ , respectively, Et ₂ Pb from <1.0
385	- 18 ng Pb.L ⁻¹ , MBT from <0.14 $-$ 400 ng Sn.L ⁻¹ , DBT from <0.15 $-$ 5 ng Sn.L ⁻¹ and TBT
386	from $<0.19 - 0.50$ ng Sn.L ⁻¹ . These differences were statistically evaluated through a Kruskal-
387	Wallis Test (p<0.05, α =0.95), because data did not exhibit simultaneously a normal
388	distribution (Shapiro-Will normality test, p<0.05, α =0.95) neither homogeneity of variances
389	(Bartlett test, p<0.05, α =0.95), so ANOVA could not be performed. For sediments,
390	watersheds were significantly different for MeHg, iHg, Et ₂ Pb, MBT, DPhT and TPhT
391	(p<0.05), as for water samples only MBT did not show a significant difference (p=0.11)
392	between the watersheds. A Mann-Whitney test for between-groups comparisons with
393	Bonferroni correction for multiple comparisons (for 11 watersheds, $\alpha = 0.95/11 = 0.09$ and p<1-
394	0.09 or p<0.91) was further performed and confirmed significant differences between
395	metropolitan watersheds and overseas watersheds for mercury species and Et ₂ Pb (p<0.91).

396	As the higher amount of MBT might be due to the TBT and DBT decay over the years,				
397	correlation was searched between these species. In water, no correlation is observed for MBT				
398	and DBT in relation to TBT (r = -0.19 and r = 0.28, respectively, p<0.05), suggesting that				
399	organotin species concentration is not influenced by any particular pollution source. In				
400	sediments, however, the correlation between TBT and its degradation products is stronger (r =				
401	0.42 and $r = 0.68$, respectively, p<0.05) as a record of TBT point sources already decaying.				
402	Older data on butyltins in French watersheds are available for comparison due to the TBT				
403	issue discovered in the Arcachon Bay (France) in the early 80s, which boosted research on				
404	organotin species all over the country, mostly in costal environments at a local scale.				
405	Unfortunately, national surveys are scarce. A 1998 report from the French National				
406	Observation Network (RNO, today named as ROCCH) discloses butyltins and TPT				
407	concentrations in 237 coastal water samples from 36 different coastal areas (Michel and				
408	Averty, 1998), where concentrations in water ranged between $<0.2 - 26.9$ ng Sn.L ⁻¹ for MBT,				
409	<0.2 - 125.0 ng Sn.L ⁻¹ for DBT, $<0.2 - 114.6$ ng Sn.L ⁻¹ for TBT and $<0.32 - 11.1$ ng Sn.L ⁻¹				
410	for TPT. Between 1998 and today, no other national survey data was found, but the				
411	comparison of both allows to conclude that the concentrations of TBT, DBT and TPT in				
412	French river basins are decreasing.				
413	Environmental monitoring programs were also searched in other countries for comparison as				
414	data from ports, shipyards and other coastal environments with high boating activities are				
415	very frequent on the literature, but not adequate to compare with the data from this national				
416	survey. In Canada, a national survey took place in 1994-95 and revealed butyltin				
417	concentrations up to 27.2 ng Sn.L ⁻¹ in fresh and seawater, while phenyltin concentrations				

- 418 were below their limits of quantification. In Portugal, a national survey in 1999-2000
- 419 registered butyltin concentrations from <3.0 30 ng.L⁻¹ (Díez et al., 2005) and in Slovenia

- 420 concentrations varied from <1.1-718 ng.L⁻¹ for butyltins and up to 16 ng.L⁻¹ for phenyltins,
- 421 in the period 2000-2006 (Nemanič et al., 2008).
- 422 Sediments from Mediterranean ponds were studied by the French National Observation
- 423 Network in 1999 and revealed TBT concentrations ranging between <0.3 52.0 ng Sn.g⁻¹ but
- 424 other organotin species were not addressed (RNO, 1999). Two years later, in 2001, eleven
- rivers from the Adour-Garonne river basin were studied and concentrations found in
- sediments for butyltins ranged from <0.03 125 ng Sn.g⁻¹, while phenyltins ranged from
- 427 <0.03 40 ng Sn.g⁻¹ (Bancon-Montigny et al., 2004). In 2007/2008 another study in the
- 428 Adour estuary and in the Arcachon Bay showed sediment concentrations between 1.28 5.44
- 429 ng $Sn.g^{-1}$ (Rodriguez-Gonzalez et al., 2012).
- 430 As for international data, the Canada survey registered butyltin concentrations up to 2525
- 431 $ng.g^{-1}$ and phenyltin concentrations up to 19.6 $ng.g^{-1}$. In Portugal, butyltin concentrations
- 432 ranged from <3.8 78 ng.g⁻¹ while in Slovenia sediments registered concentrations up to
- 433 1995 ng.g⁻¹.
- The organotins concentrations obtained in the sediments collected in the present study are
 consistently lower, supporting the argument that organotins concentrations in the French
 aquatic environment are decreasing. The same decreasing trend for TBT and TPT was found
 by the German Environmental Specimen Bank in biological samples collected during 21
 years, from 1985 to 2006 (Rüdel et al., 2009, 2007).
- 439

440 There are several French local data available which report MeHg concentration in water and 441 sediments. MeHg in the Adour estuary ranged from 0.02 to 0.95 ng Hg.L⁻¹ between 2004 442 (Stoichev et al., 2004) and 2010 (Sharif et al., 2014), the industrialized Seine estuary 443 presented MeHg concentrations between 0.012 - 0.021 ng Hg.L⁻¹ in 2007 while the adjacent

- 444 coastal waters ranged between 0.010 0.014 ng Hg.L⁻¹ (Laurier et al., 2007). These are in the 445 same range of concentrations found in this national survey.
- 446 There are some international data available, although surveys are mostly carried out in
- seafood and fish. Lakes and rivers in California in 1988/1989 exhibited MeHg concentrations
- 448 between 0.04 97 ng.L⁻¹ and 0.17 3.0 ng.L⁻¹, respectively (Gill and Bruland, 1990). In 1994
- the average MeHg concentration was 0.013 ng.L^{-1} in Lake Michigan (Mason and Sullivan,
- 450 1997) and later in 2010 was up to 0.66 ng. L^{-1} on Indiana watersheds (Risch et al., 2010),
- 451 which are again in the same range of concentrations found in this national survey.
- 452 Concerning sediments, Coquery found average MeHg concentrations of 0.18 ng Hg.g⁻¹ in
- 453 sediments from the Loire river in 1994 (Coquery et al., 1997). Ten years later, sediments from
- the Adour estuary exhibited MeHg concentrations between 0.1 1.6 ng Hg.g⁻¹ (Stoichev et
- al., 2004). There are also different studies about Seine river sediments from 1999 to 2011 in
- 456 which the MeHg concentrations varied from $0.1 6 \text{ ng Hg.g}^{-1}$ in 1999 (Mikac et al., 1999),
- 457 $0.37 1.08 \text{ ng Hg.g}^{-1}$ in 2002 (Foucher, 2002), $0.6 3 \text{ ng Hg.g}^{-1}$ in 2008 (Ouddane et al.,
- 458 2008) and finally 0.862 2.55 ng Hg.g⁻¹ in 2011 (Ramond et al., 2011). Today, and similarly
- to what was observed for water concentrations, MeHg levels in France stand in the same
- 460 range of concentration, although seasonal variations are known to occur (Rodriguez-Gonzalez
- 461 et al., 2012; Sharif et al., 2014; Stoichev et al., 2004).
- 462 Data from other countries are available at a local scale, sometimes due to known Hg pollution
 463 sources, and present great variability making it difficult to compare with the present study.
 464
- The Et₂Pb is a result of the natural degradation of the tetraethyllead additives used in gasoline until its phase-out in France during the 90s. Taking into account the short time of half-life of the organolead species (between 0.6h - 8h for Et₄Pb and 1.5 days for Et₃Pb (Harrison and Allen, 1989)) the occurrence of Et₂Pb was expected to be very low. Unfortunately, the

469 presence of Et_2Pb in environmental matrices in not well documented. Most studies on 470 organolead compounds, published between 1984 – 2000, focus on the evaluation of the ratio 471 between the sum of organolead species and total lead (probably due to the performances of 472 the analytical techniques used at the time), which means no data on the occurrence of Et_2Pb 473 alone is available. However, the results obtained in this survey are far below the WFD-EQS 474 which establish an annual average concentration of 1.3 µg.L⁻¹ for lead and its compounds.

475

476 <u>3.4.2. No significant influence of anthropogenic pressures</u>

The influence of types of pressure was evaluated and data was represented as boxplots for 477 each compound as a function of their surrounding pressure: agricultural, bad ecological status 478 (BES), industry, mixed, reference and urban (Figure S2, Supplementary Information). The 479 label "mixed" for surrounding pressure includes sites with urban and agricultural pressure, as 480 481 well as both industrial and agricultural activities or even urban and industrial areas. MeHg in sediments exhibit significant lower concentration for mixed surrounding pressure 482 and significant higher concentration for sites labeled as reference (for 6 types of surrounding 483 pressure, $\alpha = 0.95/6 = 0.16$ and p<1-0.16 or p<0.84), ranging between <0.09 and 1.5 ng Hg.g⁻¹, 484 but no data on grain size is available to further interpret these results. The corresponding 485 water samples do not follow the same tendency for the Hg species, neither for organotin 486 species. All results for MPhT and TPhT for sediments and water samples were below LOO. 487 while for DPhT values were below LOQ only in water samples. 488

489

For sediments, Kruskal-Wallis Test (p<0.05) and Mann-Whitney test for multiple groups (α =0.16 and p<0.84) showed that mixed surrounding pressure was significantly lower than other pressures for MeHg and Et₂Pb in sediments (p<0.84) and water (p<0.84). Higher concentrations of phenyltins were expected for agricultural pressures as a result of the use of

20

494	fungicides (Donard et al., 2001), but MPhT and TPhT were below LOQs for both sediments
495	and water samples and only DPhT occurred more frequently although with concentrations
496	between $<0.2 - 0.6$ ng Sn.g ⁻¹ , very close to LOQ. The lack of information on the occurrence
497	of these compounds in others aquatic environments rather than coastal areas is quite important
498	because the marine impact of organotins was so deep that community was distracted and
499	uninterested in other possible sources of these compounds in the environment (Donard et al.,
500	2001).
501	
502	3.4.3. Butyltin species consistently higher in coastal water bodies
503	When categorized by type of water body (Figure S3, Supplementary Information) – coastal
504	water, lakes and rivers – river concentrations found for DBT and Et_2Pb where significantly
505	higher (p<0.05) than lakes. In sediments, however, Kruskal-Wallis Test (p<0.05) and Mann-
506	Whitney test (for 3 types of water bodies, $\alpha = 0.95/3 = 0.32$ and p<1-0.32 or p<0.68) showed
507	that MeHg concentration is significantly lower (p<0.68) for samples collected in lakes (almost
508	below the related LOQ), while costal sediments had a concentration range between <0.09 and
509	3 ng Hg.g ⁻¹ and river sediments ranged from <0.09 to 1.3 ng Hg.g ⁻¹ .
510	As expected, butyltin species showed higher concentrations in sediments from coastal waters
511	and lower concentrations in sediments from rivers: DBT concentrations in coastal sediments
512	are significantly higher (p<0.68) and TBT concentrations are significantly lower (p<0.68) in
513	rivers (Figure 2). TBT in costal sediments ranged from $<0.6 - 55$ ng Sn.g ⁻¹ , while in lakes and
514	rivers was no higher than 3 ng Sn.g ⁻¹ .
C1C	

515

516 3.5. Background concentrations and thresholds of organometallic compounds in water and
517 sediment samples

In order to define background concentrations, all data were gathered in Figure S4 518 (Supplementary Information) where the results are represented as boxplots. Table 4a and 4b 519 are a brief summary of the results presented, such as the range of concentrations obtained 520 (minimum and maximal concentration), the median concentration, the 25th and 75th 521 percentiles, the number of samples below the LOD and the calculated threshold, i.e., upper 522 limit which differentiates background from anomalies. According to the guidelines from the 523 Idaho Geological Survey (Dai and Welhan, 2014) for non-normal distributed data, which do 524 not meet the steady state condition and for which possible seasonal effects were not corrected, 525 this threshold should be calculated as the (median + 1.65 x (interquartile range)) but the 526 compounds must have a valid median, or, on other words, at least 50 % of the occurrence data 527 for these compounds must be above LOQ. 528 Background concentrations for iHg in water at the French national level are between <0.14 529

and 2.1 ng Hg. L⁻¹, and for MeHg are between <0.04 and 0.14 ng Hg. L⁻¹, which is much lower than the estimated environmental protection values. Et₂Pb background concentration ranged from <0.1 to 8.43 ng Pb. L⁻¹ but no environmental protection values for this specie in particular is established. MBT and DBT estimated background concentrations are 0.49 - 151ng Sn. L⁻¹ and <0.15 - 3.04 ng Sn. L⁻¹, respectively. It is not possible to estimate background range of concentrations for TBT and phenyltin species due to the high number of samples below the LOQ.

In what concerns sediments, background concentration was set as <0.09 - 1.11 ng Hg. g⁻¹ for MeHg and <0.06 - 24.3 ng Pb. g⁻¹ for Et₂Pb, while MBT, DBT and TBT backgrounds are <2.0 - 13.4 ng Sn. g⁻¹, <1.6 - 8.54 ng Sn. g⁻¹ and <0.6 - 1.16 ng Sn. g⁻¹, respectively. Again, phenyltins did not occur above their LOQ, except DPhT, for which it is possible to establish a background concentrations in the <0.02 - 0.61 ng Sn. g⁻¹ range. When compared with the environmental protection values s available (EQS and PNEC), background concentrations

reveal that only TBT occurs at higher concentrations in both water and sediment samples
(EQS: 0.2 ng.L⁻¹ and PNEC: 0.024 ng.g⁻¹), reinforcing the need to monitor this substance and
the correspondent decay products in order to assure its decreasing occurrence.

546

547 **4. CONCLUSIONS**

The analytical method for the determination of organotin (MBT, DBT, TBT, MPhT, DPhT and TPhT), organomercury (MeHg and iHg) and organolead (Et₂Pb) compounds was properly validated before analysis and data treatment. LOQs ranged from 0.03 - 1.0 ng.L⁻¹ for water samples and from 0.09 - 16.6 ng.g⁻¹ for sediment samples, which are below EQS and therefore in agreement to the quality demands of the EU WFD. Additionally, linearity was higher than 0.985 at the concentration range LOQ – 6 ng.L⁻¹ (water) and LOQ – 30 ng.g⁻¹ (sediments), and precision below 10%.

555 This sampling campaign, with 152 water samples and 123 sediment samples, allowed the assessment of the organometallic concentrations of as a function of the watersheds, types of 556 557 surrounding pressure and water bodies. Results obtained for sediments from the overseas departments seemed to be consistently lower than those obtained for metropolitan areas, but 558 significant differences were only found for Et₂Pb, generally correlated with urban 559 environments. When types of pressure were compared, only the samples label as "mixed" 560 pressure showed significantly lower concentrations for water and sediment samples. Finally, 561 coastal water sediments presented the highest concentrations of butyltin species and river 562 sediments presented the lowest. These results reveal an overall decreasing on organotin 563 concentrations in French river basins, when compared to data from the French National 564 Observation Network in 1998, while concentrations for mercury species remain stable over 565 time. 566

567 Updated background concentrations at the French national level were subsequently calculated
568 for the target organometallic compounds in both matrices except phenyltins, which occurred
569 below their LOQ. These data also revealed that TBT background concentration is higher than
570 the available quality standards in both sediments and water samples.
571 The results of this 2012 monitoring campaign will contribute to the regular update of the list
572 of substances to be integrated in the monitoring programs of the Water Agencies in the

573 various river basins.

574

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Table 1. Recoveries (%) obtained for butyltin and phenyltin species in acidified mineral water samples spiked with 1xLOQ concentration, 10xLOQ and for a non acidified tap water sample spiked with 10xLOQ

		MBT			DBT			TBT	
,	Mineral water	Mineral water	Tap water	Mineral water	Mineral water	Tap water	Mineral water	Mineral water	Tap water
days	(1xLOQ)	(10xLOQ)	(10xLOQ)	(1xLOQ)	(10xLOQ)	(10xLOQ)	(1xLOQ)	(10xLOQ)	(10xLOQ)
t = 0	100	85	101	96	95	116	103	105	93
t = 3	-	101	48	-	96	97		108	80
t = 5	-	89	49	-	97	83		94	93
t = 7	-	88	20	-	95	64	<u> </u>	103	60
		MPhT			DPhT			TPhT	
dava	Mineral water	Mineral water	Tap water	Mineral water	Mineral water	Tap water	Mineral water	Mineral water	Tap water
uays	(1xLOQ)	(10xLOQ)	(10xLOQ)	(1xLOQ)	(10xLOQ)	(10xLOQ)	(1xLOQ)	(10xLOQ)	(10xLOQ)
t = 0	99	102	85	101	107	85	93	98	81
t = 3	-	79	67	-	96	17	-	105	75
t = 5	-	94	4	-	108	2	-	84	83
t = 7	-	77	3	-	70	1	-	102	55

Table 2. Recoveries (%) obtained for Et₃Pb in acidified mineral water samples spiked with 1xLOQ concentration, 10xLOQ and for a non acidified tap water sample spiked with 10xLOQ

		Et ₃ Pb	
4	Mineral water	Mineral water	Tap water
days	(1xLOQ)	(10xLOQ)	(10xLOQ)
t = 0	137	108	127
t = 3	-	89	109
t = 7	-	105	47
5			

Table 3. Recoveries (%) obtained for organomercury species in acidified mineral water samples spiked with 1xLOQ concentration, 10xLOQ and for a non acidified tap water sample spiked with 10xLOQ

		MeHg	iHg				
1	Mineral water	Mineral water	Tap water	Mineral water	Mineral water	Tap water	
days	(1xLOQ)	(10xLOQ)	(10xLOQ)	(1xLOQ)	(10xLOQ)	(10xLOQ)	
t = 0	98	110	114	96	117	81	
t = 3	-	n.d.	126	-	n.d.	83	
t = 5	-	n.d.	122	- , C-	n.d.	81	
t = 7	-	n.d.	70	-	n.d.	40	

n.d. not determined

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Table 4a. Background concentrations for water matrices (expressed as metal concentration) defined as minimum, 25^{th} percentile, median, 75^{th} percentile and maximum concentration, as well as the calculated threshold and the Environmental Quality Standard (EQS) regulated for each substance (n = 152 samples)

	MeHg	iHg	Et ₂ Pb	MBT	DBT	TBT	MPhT	DPhT	TPhT
Min Concentration (ng.L ⁻¹)	< 0.04	<0.14	<1.0	0.49	< 0.08	< 0.08			-
25 th percentile (ng.L ⁻¹)	< 0.04	0.30	<1.0	7.89	0.26	<0.08		-	-
Median Concentration (ng.L ⁻¹)	0.04	0.50	2.16	16.1	0.57	<0.08	17.6*	-	-
75 th percentile (ng.L ⁻¹)	0.06	0.83	4.52	46.0	1.33	0.09) -	-	-
Max Concentration (ng.L ⁻¹)	0.14	2.10	8.43	151	3.04	0.25	-	-	-
Nr of samples below LD	21	2	32	0	7	91	132	152	152
Calculated threshold (ng.L ⁻¹)	0.10	1.38	8.60	79.0	2.33	n.c.	n.c.	n.c.	n.c.
Quality Standards (ng.L ⁻¹)	50 °; 10 ^d	$636 - 3180^{a}$	n.a.	n.a.	n.a.	0.08 ^c	n.a.	n.a.	0.64 ^b
*only one sample was quantified. ^a PNEC from (Rodrigues et al., 2013) ^b PNEC from (Yi et al., 2012)							° EU-	WFD	

^d PNEC from (Vincent et al., 2010)

n.c. not calculated n.a. not attributed

Table 4b. Background concentrations for sediments (expressed as metal concentration) defined as minimum, 25^{th} percentile, median, 75^{th} percentile and maximum concentration, as well as the calculated threshold and the Environmental Quality Standard (EQS) regulated for each substance (n = 123 samples)

	MeHg	Et ₂ Pb	MBT	DBT	TBT	MPhT	DPhT	TPhT
Min Concentration (ng.g ⁻¹)	< 0.09	<0.6	<1.4	< 0.82	< 0.25	<0.93	<0.08	- 7
25 th percentile (ng.g ⁻¹)	< 0.09	<0.6	<1.4	< 0.82	< 0.25	<0.93	<0.08	-
Median Concentration (ng.g ⁻¹)	< 0.09	1.66	2.80	1.84	0.25	<0.93	<0.08	3.2*
75 th percentile (ng.g ⁻¹)	0.29	5.79	5.11	3.20	0.52	<0.93	0.21	-
Max Concentration (ng.g ⁻¹)	1.11	24.3	13.4	8.54	1.16	6.1†	0.61	-
Nr of samples below LD	12	1	10	35	40	118	57	112
Calculated threshold (ng.g ⁻¹)	n.c.	10.9	9.96	6.54	0.90	n.c.	n.c.	n.c.
Quality Standards (ng.g ⁻¹)	1100 ^b	n.a.	n.a.	n.a.	0.024 ^a	n.a.	n.a.	n.a.

*only one sample was quantified. †only two samples were quantified. ^a PNEC from (Silva et al., 2014)

^d PNEC from (Vincent et al., 2010) n.

n.c. not calculated n.a. not attributed

Figure 1. Illustration of the sampling sites in the French metropolitan area, in which green, pink, blue, purple and dark red color dots represent watercourses from urban, agricultural, reference, industrial and "poor ecological status", respectively, and black squares refer to water bodies.

Figure 2. Boxplot representations of MeHg, DBT and TBT concentrations in sediments from different water bodies

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HIGHLIGHTS

First assessment of background concentrations for organometallic compounds in France;

Water (152 samples) and sediment samples (123) were collected all over the country;

The analytical method was properly validated for both matrices prior to analysis;

The results reveal an overall decreasing on organotin concentrations in French river basins;

Concentrations for mercury species remain stable over time.