

## SCCS OPINION ON Ethylzingerone – 'Hydroxyethoxyphenyl Butanone' (HEPB) (Cosmetics Europe No P98)

Ulrike Bernauer, Laurent Bodin, Leonardo Celleno, Qasim Mohammad Chaudhry, Pieter-Jan Coenraads, Maria Dusinska, Jeanne Duus-Johansen, Janine Ezendam, Eric Gaffet, Lodovico Corrado Galli, et al.

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1 2 3 4 5 6 7 8	European Commission
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10 11	Scientific Committee on Consumer Safety
11	SCCS
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	OPINION ON
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	Ethylzingorono –
20	Ethylzingerone –
21	'Hydroxyethoxyphenyl Butanone' (HEPB)
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	(Competing Furgers No DOR)
24 25	(Cosmetics Europe No P98)
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27 28	
29	The SCCS adopted this Opinion by written procedure
30 21	On 22 December 2016
31 32	On 23 December 2016

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20 21 22 23 24 25 26 27	Scientific Committee members Ulrike Bernauer, Laurent Bodin, Leonardo Celleno, Qasim Chaudhry, Pieter Jan Coenraads, Maria Dusinska, Jeanne Duus-Johansen, Janine Ezendam, Eric Gaffet, Corrado Lodovico Galli, Berit Granum, Eirini Panteri, Vera Rogiers, Christophe Rousselle, Maciej Stepnik, Tamara Vanhaecke, Susan Wijnhoven.
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35 36 37 38 39 40		(Scientific Committee on Consumer Safety), Opinion on kyphenyl Butanone' (HEPB) - Cosmetics Europe No P98, ion of 23 December 2016

1	TABLE OF CONT	TENTS
2		
3		
4	ACKNOWLEDGME	ENTS
5	1. BACKGROU	JND5
6	2. Terms of re	eference5
7	3. OPINION	
8	3.1 Chen	nical and Physical Specifications6
9 10 11 12 13 14 15 16 17 18	$\begin{array}{c} 3.1.1 \\ 3.1.2 \\ 3.1.3 \\ 3.1.4 \\ 3.1.5 \\ 3.1.6 \\ 3.1.7 \\ 3.1.8 \\ 3.1.9 \end{array}$	Chemical identity6Physical form6Molecular weight7Purity, composition and substance codes7Impurities / accompanying contaminants7Solubility9Partition coefficient (Log Pow)9Additional physical and chemical specifications9Stability10
19	3.2 Fund	tion and uses 10
20	3.3 Toxic	cological Evaluation 10
21 22 23 24 25 26 27 28 29 30 31 32 33	3.3.1 3.3.2 3.3.3 3.3.4 3.3.5 3.3.6 3.3.7 3.3.8 3.3.9 3.3.10 3.3.11 3.4 Expo	Acute toxicity10Irritation and corrosivity11Skin sensitisation13Toxicokinetics14Repeated dose toxicity17Reproductive toxicity20Mutagenicity / Genotoxicity21Carcinogenicity25Photo-induced toxicity25Human data26Special investigations26sure assessment27
34	3.5 Safet	cy evaluation (including calculation of the MoS)27
35	3.6 Discu	ıssion 29
36	4. CONCLUSI	ON
37	5. MINORITY	OPINION
38 39 40	6. REFERENCI	ES

## 2 1. BACKGROUND

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Ethylzingerone with INCI name 'Hydroxyethoxyphenyl Butanone' (HEPB) Cosmetics Europe
No P98 - CAS No 569646-79-3 is a cosmetic ingredient not regulated under Cosmetic
Regulation No 1223/2009 with the function as skin conditioning agent.

8 In September 2015, the Commission' services received a dossier from industry to support 9 the safe use of Hydroxyethoxyphenyl Butanone (HEPB) when used as preservative in 10 rinse-off, oral care and leave-on cosmetic products.

According to the above-mentioned dossier the safety profile of HEPB has been studied in preclinical studies conducted according to Good Laboratory Practices and following OECD guidelines whenever available. They were conducted using HEPB from batches which were well characterized analytically with a minimum purity of 95.9%.

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## 17 2. Terms of reference

- 18
- 19 (1) In light of the data provided, does the SCCS consider safe the use of
   20 Hydroxyethoxyphenyl Butanone (HEPB) as a cosmetic preservative in rinse-off, oral care
   21 and leave-on cosmetic products with a maximum concentration of 2.0 % under an
   22 aggregate exposure scenario for cosmetics?
- (2) Alternatively, what is according to the SCCS, the maximum target concentration
   considered safe for use of Hydroxyethoxyphenyl Butanone (HEPB) as a cosmetic
   preservative in rinse-off, oral care and leave-on cosmetic products under an aggregate
   exposure scenario for cosmetics?
- 27 (3) Does the SCCS have any further scientific concerns regarding the use of
   28 Hydroxyethoxyphenyl Butanone (HEPB) in cosmetic products?
- 29

1			
2	3. OPINION		
3 4 5	3.1 Chemical and Physical Specifications		
6	3.1.1 Chemical identity		
7 8 9	3.1.1.1 Primary name and/or INCI name		
9 10 11	Hydroxyethoxyphenyl butanone (INCI)		
12	3.1.1.2 Chemical names		
13 14	4-(3-ethoxy-4-hydroxyphenyl)butan-2-one		
15 16	3.1.1.3 Trade names and abbreviations		
17 18	Ethylzingerone		
19 20	R0069279A		
21 22	НЕРВ		
23 24 25 26	<b>SCCS comment</b> For convenience, the abbreviation HEPB will be used throughout the Opinion.		
27	3.1.1.4 CAS / EC number		
28 29 30 31	CAS: 569646-79-3 EC: not assigned		
32	3.1.1.5 Structural formula		
33 34	CH <sub>3</sub>		
	HO		
35 36 37	CH <sub>3</sub>		
38	3.1.1.6 Empirical formula		
39 40 41	C12H16O3		
42	3.1.2 Physical form		
43			

Depending on the temperature, hydroxyethoxyphenyl butanone may appear as a solid
 (white powder or crystals) or as a pale yellow liquid form.

## 3.1.3 Molecular weight

Molecular weight: 208.25 g/mol

## 3.1.4 Purity, composition and substance codes

10 Chemical characterisation was performed using IR, <sup>1</sup>H-NMR, <sup>13</sup>C-NMR, mass spectrometry 11 and UV-Vis spectroscopy for the batches 019 D-004, 019 P-001 and 020-P001 with the 12 following results:

- IR Spectra of the 3 batches were comparable.
- The <sup>1</sup>H-NMR and <sup>13</sup>C-NMR spectra of the three batches were comparable. Presence of an impurity corresponding to the alcoholic form (R0070073A) of the expected structure, (estimated value: 0.04 M/Mole in Batch 019D-004). The impurity (R0070073A) is detected in the three batches.
- UV / Vis spectra of the 3 batches were comparable.
- Mass spectrometry: hydroxyethoxyphenyl butanone batch 019D-004 was analysed 19 20 by infusing a diluted hydro-alcoholic solution of the sample into a Thermo Fischer 21 Scientific LTQ-Orbitrap mass spectrometer operating in the negative ion mode. The  $[M-H]^{-}$  ion of the expected molecule  $C_{12}H_{16}O_3$  was mainly observed in the mass 22 23 spectrum. The MS/MS fragmentation pattern of this ion confirmed the expected 24 structure of the molecule. The mass spectra of the two other batches 019P-001 and 25 020P-001 recorded using the same analytical conditions were comparable to the 26 spectrum of batch 019D-004.

Purity of hydroxyethoxyphenyl butanone was determined by HPLC-PDA ( $\lambda$ =281 nm) against hydroxyethoxyphenyl butanone R0069279A batch 008L-001 primary reference standard considered as pure (100%) for batches 019D-004 and 019P-001 and against hydroxyethoxyphenyl butanone R0069279A batch 018L-002 secondary reference standard considered as 98.9% pure for batch 020P-001.

33 Hydroxyethoxyphenyl butanone purity:

34 Batch 019D-004: 95.9 ± 0.5% [w/w]

- 35 Batch 019P-001: 96.3 ± 0.8% [w/w]
- 36 Batch 020P-001: 96.2 ± 0.8% [w/w]
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#### 3.1.5 Impurities / accompanying contaminants

## 41 Impurities by HPLC-PDA

42 Separation was achieved by a linear gradient reversed phase liquid chromatography (LC) 43 method with a photodiode array detector, Maxplot  $\lambda = 210-700$ nm.

44 The retention time of hydroxyethoxyphenyl butanone was around 8.5 minutes. One impurity 45 (R0070073A, corresponding alcohol) was detected (Retention time  $R_t = 9.7$  min) in the three batches, its content was determined using R0070073A 001L002 as reference standard 46 47 (100%). One impurity (R0070359A, ethylvanillin) was detected ( $R_t = 6.3$  min) in the three batches with a relative purity <0.1 %: its content was determined using R0070359A 48 002L001 as reference standard (100%). One impurity was detected ( $R_t = 7.4$  min) in 49 50 hydroxyethoxyphenyl butanone Batches 019D004 and 019P001 with a relative UV purity <0.1%. Three impurities were detected ( $R_t = 12.8 \text{ min}$ , 14.9 min and 15.9 min) in 51 hydroxyethoxyphenyl butanone Batch 020P001 with a relative UV purity < 0.1 %. One 52 53 impurity was detected ( $R_t = 3.6 \text{ min}$ ) in hydroxyethoxyphenyl butanone batches 019D004 and 019P001 with a relative UV purity < 0.1%. For this compound, the chemical structure 54 55 3-ethoxybenzaldehyde was proposed. 56

- 1 The HPLC profiles of the three batches were comparable. 2
  - According to the HPLC profile, the following impurities were identified:
- 3 4

In the different batches, these impurities were detected in the following amounts:

R0070359A (Ethylvanillin) [µg/g]:

< 1000 in batches 019D004, 019P001 and 020P001.

R0070073A (corresponding alcohol) 2-ethoxy-4-(3-hydroxybutyl) phenol [% w/w]:

 $4.8 \pm 0.05$  in batch 019D004;  $4.0 \pm 0.05$  in batch 019P001;  $3.7 \pm 0.02$  in batch 020D004

## Residual solvents by GC:

Acetone and ethyl acetate used in the manufacturing process of the three batches: < 0.1 % (w/w)

### Heavy Metals [mg/kg]:

```
Batch 019D004:
             As, Cd, Hq, Pb, Pd: each < 1
             Al, Ba, Co, Cr, Cu, Mn, Mo, Ni, Se, Sn, Ti, V, Zn: each <5
             Fe: 9
      Batch 019P001:
             As, Cd, Hg, Pb, Pd: each < 1
             Al, Ba, Co, Cr, Cu, Mn, Mo, Ni, Se, Sn, Ti, V, Zn: each <5
             Fe: 5
31
      Batch 020 P 001:
32
             Al, Ba, Co, Cr, Cu, Mn, Mo, Ni, Se, Sn, Ti, V, Zn : each < 1
             Ba, Co, Cr, Cu, Fe, Mn, Mo, Ni, Se, Sn, Ti,V, Zn : each < 5
33
34
             AI: 6
35
```

```
36 Ash [% w/w]:
```

7

- Batch 019D004: < 0.1
- Batch 019P001: 0.1
- 4 Batch 020P001: 0.1 5 6

## Elemental analysis:

	Theoretical values [% w/w]	, , , ,, ,		
		Batch 019D004	Batch 019P001	Batch 020P001
Carbon	69.2	69.1	69.3	68.8
Hydrogen	7.7	7.7	7.6	7.8
Oxygen	23.0	23.2	23.1	23.3

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#### 3.1.6 Solubility

- 12 Water solubility (batch 019 P001): 7.59 g/L at 20°C ± 0.5 °C
- (EEC method A6 OECD method 105) 13 14
- Solubility (at 21°C) (batch 006 L001 and 007 L001 from batch 019D004) 15
- Water: < 0.1 mg/mL (method not stated) 16
- Ethanol:  $\geq 1000 \text{ mg/mL}$ 17
- 18 - DMSO: ≥1000 mg/mL
- Corn oil: <0.1 mg/mL 19 20

#### **SCCS** Comment 21

The SCCS notes that the analytical file gives considerably differing values for water 22 solubility. The issue has been clarified by the applicant and the correct value for water 23 solubility is 7.59 g/L at 20°C  $\pm$  0.5 °C. 24

25 26

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#### Partition coefficient (Log Pow) 3.1.7

29 Log Pow: 1.46 at 22.8°C ± 1 °C (EEC method A8 - OECD method 107) 30 31

## 3.1.8 Additional physical and chemical specifications

33		
34	Melting point:	$41 \pm 2^{\circ}C$ (DSC method)
35	Boiling point:	328 ± 2°C at 100 to 102 kPa
36	Flash point:	/
37	Vapour pressure:	8.7 x 10 <sup>-3</sup> Pa at 25 °C
38	Density:	/
39	Viscosity:	
40	pKa:	10.03 (25°C and ionic strength 0.15M) for an equilibrium
41		HO/O <sup>-</sup> (GLpKa Sirius)
42	Refractive index:	
43	pH:	
	•	

1UV-Vis spectrum (200-500 nm):The UV-Vis absorption spectrum of a 0.004 g/100 ml2solution of hydroxyethoxyphenyl butanone exhibited a<br/>maximum at 282nm ± 1nm; the absorbance at 282 nm4was ~ 0.6.

## 3.1.9 Stability

8
9 The R0069279A is stable over 2 months at 45°C in hydroalcoholic solution at 0.5 g/100 ml.
10 It is sensitive to photostress but resistant to oxidative, heat, acid or basic stresses.
11 Determination was performed for the three batches using batch R0069279A 001L-001 as
12 reference.

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## 14 SCCS General Comments to physico-chemical characterisation

16 Chemical characterisation of P98 was performed using IR, <sup>1</sup>H-NMR, <sup>13</sup>C-NMR, mass 17 spectrometry and UV-Vis spectroscopy. Purity of P98 was determined by HPLC-PDA at  $\lambda$ max 18 281 nm using reference standard or secondary reference standard and it is accepted.

19 Impurity studies have been performed using HPLC-PDA detection at  $\lambda$ max of P98. Two 20 impurities (2-ethoxy-4-(3-hydroxybutyl) phenol and ethylvanillin) have been quantified 21 using reference standards. Four other impurities have been detected with a relative UV 22 purity < 0.1%. In batches 019D004 and 019P001 an impurity with a relative UV purity < 23 0.1% with a proposed chemical structure of 3-ethoxybenzaldehyde was reported.

24 Chemical identification and characterisation of impurities is considered acceptable.

## 25 3.2 Function and uses

The ingredient HEPB is intended to be used specifically as a preservative in rinse-off, oral care and leave-on cosmetic products up to 2%.

## 29 3.3 Toxicological Evaluation

31	3.3.1 Acute toxicity		
32			
33	3.3.1.1 Acute oral toxicity		
34 35 36	No data provided.		
37	3.3.1.2 Acute dermal toxicity		
38 39 40	No data provided.		
41	3.3.1.3 Acute inhalation toxicity		
42 43 44	No data provided.		
45	3.3.1.4 Acute intraperitoneal toxicity		
46 47 48 49	No data provided. Summary on acute toxicity		
47	סטווווומו ציטוו מכענב נטגוכונצ		

51 No acute toxicity study on any route is available for HEPB. In 14- and 90-day oral repeat 52 dose studies using dose levels of 100, 300 and 1000 mg/kg bw/d no deaths occurred.

1 Hence, it can be assumed that the oral  $LD_{50}$  would be higher than 1 000 mg/kg/d (i.e. the 2 substance is of low acute oral toxicity).

## 2 3 4 5 6 7 8 9 10

## 3.3.2 Irritation and corrosivity

## 3.3.2.1 Skin irritation

### In Vitro Acute Skin Irritation: Reconstructed Human Epidermis Test Method

10		
11	Guideline:	In vitro Episkin <sup>™</sup> Skin Irritation Test
12	Test system:	EPISKIN <sup>SM</sup> Reconstructed Human Epidermis (RHE) small
13		(0.38 cm <sup>2</sup> ) model (Skin Ethic Laboratories)
14	Replicates:	3 tissue batches, 3 replicates per batch
15	Test substance:	R0069279A
16	Test batch:	R0069279A 019 D 004
17	Purity:	95.9 % (HPLC)
18	Test items:	neat (100% powder)
19	Dose level:	10 mg
20	Treatment period:	15 minutes
21	Post-treatment incubation time:	42 hour $\pm$ 1 hour
22	Positive control:	5% (w/v) aqueous solution of sodium dodecyl sulphate
23		(SDS)
24	Negative control:	PBS+
25	Direct interaction with MTT:	Negative
26	Colouring of epidermis:	No information provided in the study report
27	GLP:	In compliance
28	Study period:	13.12.2011 - 17.1.2012
29		

30  $10 \pm 2$  mg of undiluted HEPB and 10 µl of each of the controls were applied onto the slightly wetted reconstructed human epidermis. After 15 min incubation at room temperature, the 31 32 tissues were rinsed with PBS+ and cultivated in fresh medium for 42 hours at 37° C. At the end of the 42-hour post-treatment incubation period, the epidermis was prepared for cell 33 viability measurement (spectrophotometric MTT conversion test). Culture medium was kept 34 35 frozen for IL-1g measurement. The amounts of IL-1g released in the culture medium were determined by a classic quantitative sandwich enzyme immunoassay technique (ELISA). The 36 37 amounts of IL-1a released were not determined for the positive control.

38 39 Results

When compared to the negative control, there were no changes in viability rates and IL-1a concentrations following application of HEPB. Mean cell viability values obtained for HEPB, negative and positive controls were 100%, 100% and 8.3%, respectively. The mean value of final IL-1a release was 10 pg/mL for HEPB.

44

45 Based on the results of this study, the authors concluded that undiluted HEPB can be 46 considered as potentially non-irritant to the skin.

47

48 49 Ref.: 2

#### 50 SCCS Comment

51 An ECVAM validated protocol was used and certificates of analysis with  $IC_{50}$  values (for SDS) 52 for each tissue batch were presented in annex of the study report. Barrier function has been 53 checked by determination of the concentration at which SDS reduces the viability of the 54 tissues by 50 % and the obtained values fell within the defined acceptance range.

3

## 3.3.2.2 Mucous membrane irritation / Eye irritation

#### Bovine Corneal Opacity and Permeability (BCOP) Test Method

4			
5	Guideline:	OECD 437 (September 2009)	
6	Test system:	Isolated bovine eyes (corneas)	
7	Replicates:	3 corneas per condition	
8	Test substance:	R0069279A	
9	Batch:	R0068279A 019 D 004	
10	Purity:	95.9 ± 0.5% (HPLC)	
11 12	Test item:	20 % (w/w) suspension of test substance in vehicle (yielding a pH of 5.5)	
13	Vehicle:	polyethylene glycol (8 OE)	
14	Dose level:	750 ± 8 μl	
15	Treatment period:	4 hours $\pm$ 10 min	
16	Positive control:	20% (w/w) imidazole in NaCl 0.9%	
17	Negative control:	NaCl 0.9% in sterile water	
18	GLP:	In compliance	
19	Study period:	Nov 2011 – Feb 2012	
20			
21			
22		cattle less than 12 months old) were collected at slaughterhouses and	
23	• •	hours of collection. Eyes that were too big or were presenting defects	
24	were rejected. After the pre-incubation and equilibration period of the corneas of at least 1		
25	hour at $32 \pm 1^{\circ}$ C, 750 µl of the test item, positive, negative and vehicle control were		
26	applied onto the corneas for 4 hours. At the end of the contact period, corneas were rinsed		
27	and prepared to measure opacity (changes in light transmission) and permeability		
28	•	fer of 5 mg/ml fluorescein through the cornea by measuring the optical	
29		of the media in the ocular posterior compartment). The in vitro irritancy	
30 21		is the combination of opacity and permeability, was then calculated.	

- 31 Negative, vehicle and positive controls were tested according to the same experimental 32 conditions.
- 3334 Results
- After 4 hours of contact, the mean IVIS was  $13.9 \pm 0.6$  for a HEPB suspension at 20 % in polyethylene glycol. Detachment of the epithelium and edema were observed. The mean IVIS for the vehicle polyethylene glycol and the positive controls were 6.7 ± 0.7 and 182.1  $\pm$  2.7, respectively.
- 39

Ref.: 3

40 Conclusion

The study authors concluded that based on the conditions of this test HEPB is not classified as corrosive or severe eye irritant.

43

## 44 SCCS comment

The SCCS notes that the permeability (OD) values of the vehicle control are much higher (20-fold) than for the negative control, suggesting that the vehicle polyethylene glycol itself might have an adverse effect on the test system. Therefore, the results of the BCOP assay are inconclusive. An appropriate vehicle should be used for which it can be demonstrated that it has no adverse effect on the test system.

50 51

## 52 **Bovine Corneal Opacity and Permeability (BCOP) Test Method**

53		
54	Guideline:	OECD 437 (September 2009) with some deviations
55	Test system:	Isolated bovine eyes (corneas)
56	Replicates:	3 corneas per condition
57	Test substance:	R0069279A

1	Batch:	R0068279A 019 D 004
2	Purity:	95.9 ± 0.5% (HPLC)
3	Test item:	2 % (w/w) suspension in vehicle (yielding a pH of 5.5)
4	Vehicle:	polyethylene glycol (8 OE)
5	Dose level:	750 ± 8 μl
6	Treatment period:	30 min and 4 hours
7	Positive control:	0.5 % (w/w) Cetyl Trimethylammonium Bromide (CTAB) in distilled
8		water
9	Negative control:	nutritive medium
10	GLP:	In compliance
11	Study period:	Jan 2012 – Feb 2012

13 Bovine eyes (from cattle less than 12 months old) were collected at slaughterhouses and prepared within 4 hours of collection. Eyes that were too big or that had defects were 14 rejected. After the pre-incubation and equilibration period of the corneas of at least 1 hour 15 16 at 32  $\pm$  1°C, 750  $\mu$ l of the test item, positive, negative and vehicle control were applied onto the corneas for 30 minutes or 4 hours. At the end of each contact period, corneas were 17 18 rinsed and prepared for measurement of opacity (changes in light transmission) and permeability (evaluation of transfer of 5 mg/ml fluorescein through the cornea by 19 20 measuring the optical density at 490 nm of the media in the ocular posterior compartment). The corneal (IVIS) score, which is the combination of opacification and permeability, was 21 22 then calculated. Negative, vehicle and positive controls were tested according to the same 23 experimental conditions except that the 4-hour treatment period was not performed on the 24 positive control. According to the study authors, this method uses a prediction model that 25 allows a classification of the products into four categories of irritation potential, i.e. from the lowest to the highest ocular irritation potential: slightly irritant, moderately irritant, 26 27 moderately irritant to irritant and irritant to severely irritant 28

29 Results:

The mean corneal (IVIS) scores obtained for 30-min contact time were:  $4.7 \pm 0.6$  for the 2 % HEPB suspension,  $3.9 \pm 0.6$  for the vehicle control, and  $96.0 \pm 2.8$  for the positive control, respectively. The mean corneal (IVIS) scores obtained for 4-hour contact time were:  $5.9 \pm 0.1$  for the 2 % HEPB suspension and  $4.5 \pm 0.0$  for the vehicle control.

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The study authors concluded that under the conditions of this study HEPB has at most a slight ocular irritation potential when tested at 2 %.

Ref.: 4

#### 40 SCCS Comment

The SCCS notes that the permeability (OD) values of the vehicle control are much higher (8-fold for the 4-hour treatment period) than the negative control, suggesting that the vehicle polyethylene glycol itself might have an adverse effect on the test system. Therefore, the results of the BCOP assay are inconclusive. An appropriate vehicle should be used for which it can be demonstrated that it has no adverse effect on the test system.

The SCCS further notes that the BCOP assay (OECD 437, 2009) is only able to identify severe eye irritants and cannot discriminate between slightly irritant, moderately irritant and severely irritant.

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## 3.3.3 Skin sensitisation

#### 53 Local Lymph Node Assay (LLNA)

54 55 Guideline: OECD 429; EC B.42

- 56 Species/strain/sex: mouse, CBA/J, female
- 57 Group size: 4

- 1 Test substance: R0069279A 2 R0069279A 019 D 004 Batch: 3 Purity:  $95.9 \pm 0.5\%$ 4 Vehicle: acetone: olive oil (4:1; v/v) 5 1, 10, 25, 50 and 75 % Concentration: 6 Vehicle control: acetone:olive oil (4:1; v/v) (AOO) a-hexylcinnamaldehyde (HCA) at 25 % (v/v) in AOO 7 Positive control: 8 GLP: In compliance 9 March 2011 – July 2011 Study period: 10 11 Concentrations for the main study were based on findings from a preceding irritation test, in 12 which no increase in skin thickness was noted at test item concentrations of 5.0, 25.0, 50.0 13 and 75.0 % (w/v). Seven treatment groups each comprising 4 mice received either the test substance at 1, 10, 14 25, 50 or 75% (w/v) in AOO, AOO alone as negative control or HCA at 25 % in vehicle as 15 16 positive control. The test item or controls were applied on the dorsum of both ears for three 17 consecutive days. After a 2-day rest lymphocyte proliferation was assessed in the pooled auricular lymph nodes by measuring the incorporation of 3H-TdR. The values were used to 18 19 calculate stimulation indices (SI). 20 Results: The positive control HCA generated a SI value of 3.38. The test substance induced a small 21 22 increase in lymphoproliferation. SI values of 1.16, 1.24, 1.43, 1.53 and 1.78 were obtained at test substance concentrations of 1%, 10%, 25%, 50% and 75%, respectively. 23 24 Therefore the study authors concluded that R0069279A did not induce skin sensitisation in 25 the LLNA. Ref.: 23 26 27 28 SCCS comment HEPB induced a dose-dependent increase of lymphocyte proliferation in the LLNA that 29 30 resulted in a SI value of 1.78 at the highest dose (75%) tested. HEPB is not considered to 31 be a skin sensitiser under the conditions of the test. 32 33 **3.3.4 Toxicokinetics** 34 35 36 3.3.4.1 Dermal / percutaneous absorption 37 38 39 Guideline: OECD 428 (November 2004); SCCS 1358/10 40 Split-thickness human abdomen skin from 4 donors aged 30 -Species/strain: 41 63 years and one split-thickness breast skin sample from a 42 female donor aged 58 years 43 sample exhibiting a resistance  $< 4 \text{ k}\Omega$  excluded Membrane integrity: 44 Replicates: 12 skin samples from 12 different donors 45 Method: Dermatomed thawed skin mounted on static diffusion cells and 46 exposed to radiodiluted test item 47 R0069279A (non-labelled); [<sup>14</sup>C]-R0069279A (labelled) Test substance: R0069279A 019 P 001 (non-labelled material) 48 Batch: 49 CFQ41364 (labelled material)
- 50 Purity:
- 51 52
- Test item:
- 53 54 Exposed membrane area:
- 55 Dose applied:
- 2.11 mg/cm<sup>2</sup> 56 Sampling period: 24 hours
- 14

 $96.0\% \pm 1.3\%$  (non-labelled material)

104.2 % of target (2.0 % (w/w)

3.14 cm<sup>2</sup>

98.0 % (labelled material; radiochemical purity)

radiodiluted [<sup>14</sup>C]-R0069279A in a cosmetic formulation;

- 1 Receptor fluid: calcium- and magnesium-free phosphate buffered saline (PBS) 2 containing bovine serum albumin (5%, w/v)3 Mass balance analysis: Provided 4 Tape stripping: Yes (20) 5 Liquid Scintillation Counting (LSC) Method of Analysis: 6 GLP: In compliance 7 June – July 2012; draft report Nov 2012 Study period:
- 8

9 The in vitro percutaneous absorption of  $[^{14}C]$ -HEPB by using a typical lipophilic skin care 10 formulation containing HEPB at the concentration of 2% was determined in human 11 dermatomed skin. Any split-thickness human skin sample exhibiting a resistance less than 4 12 k $\Omega$  was excluded from subsequent absorption measurements.

Human dermatomed skin samples were mounted onto diffusion cells. Calcium- and magnesium-free PBS was used as the receptor fluid. A quantity of 2 mg/cm<sup>2</sup> of skin care formulation was applied to the skin surface. After 24 hours of exposure, the skin surface was rinsed-off, simulating in-use conditions. 24 hours after application, the stratum corneum was removed by 20 tape strippings and the penetration, mass balance and distribution of [<sup>14</sup>C]-HEPB were determined by measuring its concentration in relevant compartments (e.g. skin wash, epidermis, dermis, and receptor fluid) by using LSC.

20 21 Results

22 Mean recovery of the applied test material (mass balance) was 82.4% of the applied dose. 23 The mean systemically available dose of HEPB sum of amounts measured in living 24 epidermis/dermis and receptor fluid after application of a typical skin care formulation 25 containing this preservative ingredient at 2% was estimated to be  $36.8\pm17\%$  of the applied 26 dose ( $16.0\pm7.4 \mu g equiv/cm^2$ ).

27 28

					Cell N	umber an	d Donor	Number						
	Cell 1	Cell 2	Cell 3	Cell 4	Cell 5	Cell 6	Cell 7	Cell 8	Cell 9	Cell 10	Cell 11	Cell 12		
	0317	0317	0394	0394	0394	0378	0313	0313	0313	0348	0348	0348	Mean	SD
Skin Wash	10.37	16.54	31.34	28.97	26.68	13.31	22.44	12.21	26.67	14.93	29.88	17.12	20.87	7.61
Tissue Swabs	9.50	18.33	25.45	35.76	29.13	14.54	18.61	22.18	31.21	19.79	20.23	20.49	22.10	7.29
Pipette Tips	1.15	0.14	0.14	0.26	0.63	0.12	0.19	0.11	0.37	0.41	0.25	0.16	0.33	0.30
Donor Chamber Wash	9.31	0.14	0.73	0.23	0.52	0.38	1.02	0.29	4.90	0.64	1.13	0.44	1.64	2.74
Dislodgeable Dose 24 h	30.33	35.15	57. <b>6</b> 5	65.22	56.97	28.34	42.27	34.80	63.14	35.78	51.48	38.21	44.95	13.20
Stratum Corneum 1-5	0.20	0.10	0.18	0.57	0.51	0.10	0.12	0.57	0.24	0.12	0.14	0.10	0.24	0.19
Stratum Corneum 6-10	0.09	0.06	0.13	0.35	0.29	0.05	0.09	0.22	0.18	0.09	0.14	0.07	0.15	0.10
Stratum Corneum 11-15	0.06	0.07	0.14	0.26	0.33	0.03	0.08	0.14	0.12	0.08	0.12	0.06	0.12	0.09
Stratum Corneum 16-20	0.04	0.06	0.12	0.29	0.13	0.04	0.07	0.08	0.12	0.06	0.13	0.04	0.10	0.07
Stratum Corneum	0.39	0.28	0.57	1.48	1.27	0.21	0.35	1.01	0.66	0.35	0.53	0.27	0.61	0.42
Unexposed Skin	0.22	0.11	0.04	0.02	0.08	0.08	0.11	0.15	0.04	0.10	0.01	0.05	0.08	0.06
Total Unabsorbed	30.95	35.54	58.26	66.72	58.32	28.64	42.72	35.96	63.84	36.23	52.02	38.53	45.64	13.43
Epidermis	0.50	0.46	0.53	1.15	0.97	0.41	0.30	0.47	0.42	0.55	0.53	0.36	0.55	0.25
Dermis	3.47	2.32	2.13	1.53	1.70	2.98	1.16	1.99	1.03	2.45	1.87	2.04	2.06	0.70
Receptor Fluid	65.33	52.62	*26.12	*16.33	*20.70	43.96	29.26	44.77	*15.73	40.34	*13.97	34.53	°33.64	°16.21
Receptor Chamber Wash	1.35	0.51	0.31	N.C.	0.46	0.72	0.46	0.70	0.24	0.48	0.21	0.74	0.56	0.32
Total Absorbed	66.68	53.12	26.43	16.33	21.16	44.68	29.71	45.47	15.97	40.81	14.18	35.27	34.15	16.51
Dermal Delivery	70.65	55.90	29.09	19.01	23.84	48.07	31.17	47.92	17.42	43.81	16.58	37.66	36.76	16.98
Mass Balance	101.60	91.44	87.35	85.73	82.16	76.71	73.89	83.88	81.25	80.04	68.59	76.19	82.40	8.68

## Table 1 Distribution of Radioactivity (% Applied Dose) at 24 h Post Dose Following Topical Application of [<sup>14</sup>C]-R0069279A in Test Preparation 819193 (2%, w/w) to Human Split-Thickness Skin

\*=Results calculated from data less than 30 d.p.m. above background; °=Mean includes results calculated from data less than 30 d.p.m above background

1														
					Cell 1	Number ar	d Donor	Number					ļ	
	Cell 1	Cell 2	Cell 3	Cell 4	Cell 5	Cell 6	Cell 7	Cell 8	Cell 9	Cell 10	Cell 11	Cell 12		
	0317	0317	0394	0394	0394	0378	0313	0313	0313	0348	0348	0348	Mean	SD
Skin Wash	4.53	7.22	13.68	12.65	11.65	5.81	9.80	5.33	11.64	6.52	13.04	7.48	9.11	3.32
Tissue Swabs	4.15	8.00	11.11	15.62	12.72	6.35	8.13	9.68	13.62	8.64	8.83	8.95	9.65	3.18
Pipette Tips	0.50	0.06	0.06	0.11	0.28	0.05	0.08	0.05	0.16	0.18	0.11	0.07	0.14	0.13
Donor Chamber Wash	4.07	0.06	0.32	0.10	0.23	0.16	0.44	0.13	2.14	0.28	0.49	0.19	0.72	1.19
Dislodgeable Dose 24 h	13.24	15.35	25.17	28.48	24.87	12.38	18.45	15.19	27.57	15.62	22.48	16.68	19.62	5.76
Stratum Corneum 1-5	0.09	0.04	0.08	0.25	0.22	0.04	0.05	0.25	0.10	0.05	0.06	0.04	0.11	0.08
Stratum Corneum 6-10	0.04	0.03	0.06	0.15	0.13	0.02	0.04	0.10	0.08	0.04	0.06	0.03	0.06	0.04
Stratum Corneum 11-15	0.03	0.03	0.06	0.12	0.15	0.01	0.03	0.06	0.05	0.04	0.05	0.02	0.05	0.04
Stratum Corneum 16-20	0.02	0.03	0.05	0.13	0.06	0.02	0.03	0.03	0.05	0.03	0.06	0.02	0.04	0.03
Stratum Corneum	0.17	0.12	0.25	0.65	0.56	0.09	0.15	0.44	0.29	0.15	0.23	0.12	0.27	0.18
Unexposed Skin	0.10	0.05	0.02	0.01	0.03	0.03	0.05	0.06	0.02	0.04	0.00	0.02	0.04	0.03
Total Unabsorbed	13.51	15.52	25.44	29.13	25.46	12.50	18.65	15.70	27.87	15.82	22.71	16.82	19.93	5.86
Epidermis	0.22	0.20	0.23	0.50	0.42	0.18	0.13	0.20	0.18	0.24	0.23	0.16	0.24	0.11
Dermis	1.51	1.01	0.93	0.67	0.74	1.30	0.51	0.87	0.45	1.07	0.82	0.89	0.90	0.31
Receptor Fluid	28.53	22.98	*11.41	*7.13	*9.04	19.20	12.78	19.55	*6.87	17.61	*6.10	15.08	°14.69	°7.08
Receptor Chamber Wash	0.59	0.22	0.14	N.S.	0.20	0.31	0.20	0.31	0.10	0.21	0.09	0.32	0.25	0.14
Total Absorbed	29.12	23.20	11.54	7.13	9.24	19.51	12.98	19.86	6.97	17.82	6.19	15.40	14.91	7.21
Dermal Delivery	30.85	24.41	12.70	8.30	10.41	20.99	13.61	20.93	7.61	19.13	7.24	16.45	16.05	7.42
Mass Balance	44.36	39.93	38.14	37.43	35.87	33.50	32.26	36.63	35.48	34.95	29.95	33.27	35.98	3.79

Table 3Distribution of [14C]-R0069279A (µg equiv./cm²) at 24 h Post Dose Following Topical Application of<br/>[14C]-R0069279A in Test Preparation 819193 (2%, w/w) to Human Split-Thickness Skin

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The applicant concluded that the mean recovery rate of 82.4 % of the applied dose obtained in this study was a bit low. However, similar to greater losses of radioactivity were obtained in a dedicated study evaluating the recovery rate of [<sup>14</sup>C]-HEBP 24 hours after simple spiking (Craig, 2014b, the study is described in the appendix). Therefore it was concluded that the low recovery rate obtained in the in vitro percutaneous absorption study would not put the scientific validity of this study into question.

Ref.: 6; 7

## 12 SCCS Comment

The dedicated study evaluating the recovery rate did not give any explanation for the low recovery. As HEPB possesses a low vapour pressure, evaporation appears unlikely. Nevertheless, in order to avoid loss, occlusive conditions could be used in dermal absorption studies.

According to SCCS 1358/10, recovery should be at least 85 %. A recovery of at least 85 %
was only obtained for 4 cells evaluated. Thus, eliminating all other cells leads to only 4
evaluable cells from 2 different donors, which is not in line with SCCS 1358/10.

20 The chosen cut-off value of 4 k $\Omega$  for barrier function assessment is not of common practice 21 (threshold of 10 k $\Omega$  is normally used) and may lead to an overestimation of the results.

The SCCS notes that dermal penetration was only investigated using a cosmetic formulation
consisting of 93 % of an oily phase and 7 % of water phase. However, appropriate studies
should be performed for the intended uses in representative cosmetic formulations.

26

Based on significant deviations from requirements, the SCCS considers the study
unacceptable. Instead, in the absence of adequate experimental data, a default value of 50
% dermal absorption is taken for MOS calculation according to SCCS 1564/15.

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32 33

2242	Tovical/instica in laboratory animals	
5.5.4.2	Toxicokinetics in laboratory animals	

No data provided.

Results from a structurally similar compound (zingerone) are indicative of almost complete and rapid absorption and excretion (see section 3.3.11). Based on this information and on physico-chemical properties of HEPB (molecular weight: 208.25 g/mol; water solubility of 7.59 g/mol and log  $P_{ow}$  of 1.46), 100 % oral absorption is taken for MoS calculation.

6	3.3.4.3 Toxicokinetics in humans					
7 8	No data provided.					
9	No data provided.					
10	3.3.4.4 Toxicokir	netics in vitro				
11						
12	Guideline:	/				
13	Test system:	Human intestinal epithelial Caco-2 cells				
14	Test substance:	R0069279A				
15	Batch:	R0069279A 019 P 001				
16	Purity:	not mentioned				
17	GLP:	in compliance				
18	Study period:	June – August 2014				
19						
20		ent and if applicable, the respective reference compounds (propranolol:				
21		e; labetalol: moderately permeable; ranitidine: poorly permeable;				
22		oprotein substrate) were tested concurrently with HEPB, and the data were				
23		storical values determined at the testing facility. Only limited information				
24 25		n the experimental setting. In short, the apical-basolateral (A-B) EPB in Caco-2 cells (pH 6.5/7.4) was determined and compared to that of				
25 26		pounds. A test concentration of 10 $\mu$ M HEPB was used.				
20		ipounds. A test concentration of 10 µM HEPB was used.				
28	Results:					
29		a mean apparent permeability coefficient of 62.2 x $10^{-6}$ cm/s. Based on				
30		test item and the reference compounds, the test item should be highly				
31	permeable.	test terr and the relevance compounds, the test terr should be highly				
32	p 00	Ref.: 22				
33						
34	SCCS comment					
35	The study report	is extremely poor. However, it can be inferred from the results of the				
36	study, that penetration of HEPB through the Caco-2 monolayer is high.					
37						
38						
39						
40	3.3.5 Repeated	l dose toxicity				
41						

- 42 3.3.5.1 Repeated Dose (14 days) oral / dermal / inhalation toxicity
- 44 Oral

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46 A 14-day oral repeat dose study was performed as a preliminary (range-finder) study for a 90-day oral toxicity study. The study was performed in August 2011, the final report dates 47 48 from March 2013. Four groups of male and female Wistar rats, respectively, each 49 comprising 5 animals, received dose levels of 100, 300 and 1000 mg/kg bw/d of HEPB (test item R0069279A, batch R0069279A 019 D 004) in 0.5 % (w/v) carboxymethylcellulose 50 (CMC) in water and 2 % Tween® 80 (v/v) by oral gavage. Control groups received vehicle 51 only. Animals were observed twice daily for clinical signs and body weight and food 52 consumption were monitored. At the end of the treatment period, blood and urine samples 53 54 were taken, animals were subjected to gross examination and specified organs were

collected. Histopathological examination was carried out on heart, kidneys, liver and spleen
 from all rats of the vehicle control and high dose group.

3 There were no clinical signs or mortalities in any of the groups during the treatment period. 4 There were no test item-related changes in any of the parameters evaluated in 5 haematology, coagulation, clinical chemistry and urine. An increase in absolute and relative 6 liver weight was observed in high dose males. There were no test item-related gross 7 pathological findings in test-item treated animals. In high-dose males, a 6 % increase in 8 mean platelet volume not accompanied by any significant changes in platelet count was also 9 observed. A dilatation of the uterus was observed in one female of the mid-dose and one 10 female of the high-dose.

11 Based on the results of this 14-d oral repeat dose study, 100, 300 and 1000 mg/kg bw/d 12 were chosen for the 90-day oral toxicity study.

Ref.: 20

15 16 3.3.5.2 Sub-chronic (90 days) toxicity oral/ dermal / inhalation toxicity 17 18 Oral 19 20 Guideline: OECD TG 408 (Sep 1998) Rat, Wistar rats-HsdHan<sup>™</sup>:WIST 21 Species/strain: 22 Group size: 10 / sex / dose 23 Test substance: R0069279A 24 R0069279A 019 D 004 Batch: 25 95.9 ± 0.5 % Purity: 26 Vehicle: 0.5 % (w/v) carboxymethylcellulose (CMC) in water and 2 % Tween® 27 80 (v/v) 0, 100, 300 and 1000 mg/kg bw/d 28 Dose levels: 29 Dose volume: 10 ml/ka 30 Route: oral 31 Administration: gavage 32 GLP: In compliance 33 Study period: Sep 2011 - Oct 2012 34 The test item was stable for 8 days in vehicle suspension; thus dosing formulations were 35

prepared accordingly. Animals received daily oral gavage doses for 90 consecutive days. Detailed clinical examination was performed prior to test item administration and weekly thereafter. Animals were observed regularly for mortality, morbidity and clinical signs. Ophthalmological examination was performed in control and high dose animals prior to and at the end of the treatment. Functional observational battery (FOB) tests were performed between treatment day 82 and 83. Food consumption and body weight were determined on a more or less regular basis throughout the administration period.

43 At the end of the treatment period (day 91), blood was taken for haematology and clinical 44 chemistry and urine was taken for microscopic examination and investigation of selected 45 parameters. All animals were subjected to gross necropsy and selected organs were 46 weighed and preserved. Histopathological examination was performed on all preserved 47 organs from the control and high-dose group and in addition all gross lesions from all rats 48 were examined microscopically. The stomach was examined in all dose groups.

49 50 Results

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No mortalities were observed throughout the study period. No ocular abnormalities were observed in the ophthalmological examinations. Transient clinical signs (slight to moderate salivation, dragging gait, hypoactivity, splayed hind-limbs) appeared shortly (5-15 min) after gavage in both sexes and persisted up to 30 min in animals dosed at 300 mg/kg bw/d. At the highest dose, these signs appeared approximately 5-15 min post dose and persisted up to 1 hour (60 min) post dose in most rats. For 2 rats, hypoactivity and splayed hindlimbs with dragging gait persisted for more than an hour. In FOB, grip strength of hindlimbs was

1 significantly reduced in mid- and high-dose females. Landing hindlimbs foot splay values 2 were significantly lower in males of all dose levels and in low dose females. At 300 and 1000 3 mg/kg bw/d, hindlimb grip strength in females was statistically significantly (p < 0.0001) 4 different from controls. Motor activity was statistically significantly different from controls in 5 males and females at 1000 mg/kg bw/d lower values of stereotypic time (interval 6) and 6 total stereotypic time in males and lower values of stereotypic time, ambulatory time and 7 horizontal and ambulatory counts (interval 6) in females, respectively. The study authors 8 interpreted statistical variations in motor activity as incidental as there were no 9 accompanying changes in muscle tone. Food consumption was statistically significantly 10 reduced in high-dose males and females (about -12 % and -16%, respectively, compared to controls). In high-dose males, statistically significant (about -10 % compared to controls) 11 12 lower body weights were observed. There were no test-item related alterations in 13 haematology, coagulation and clinical chemistry or urine parameters in any of the treated 14 dose groups in both sexes. Gross pathology revealed no changes in rats up to 300 mg/kg bw/d. At the highest dose level, thickening and white multiple raised foci of nonglandular 15 16 mucosa in the stomach was observed in 5/10 males and 3/10 females. The macroscopic 17 lesions were associated with mild to moderate epithelial hyperplasia in the non-glandular 18 stomach (8/10 males and 6/10 females) and with mild to moderate leucocytic infiltration of 19 the mucosa (2/10 males and 1/10 females). In addition, mild mucosal erosion was observed 20 in the glandular stomach (1/10 males). 21

22 Conclusion

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The study authors considered clinical (neurological) signs observed at the mid- and highdose as non-adverse treatment-related effects due to their transient nature. However, the observed macroscopic lesions and microscopic changes in the stomach of high-dose animals were considered test-item related and associated with lower body weight (males) and food consumption (males and females). Thus, an NOAEL of 300 mg/kg bw/d was derived from this study.

Ref.: 24

#### 32 SCCS comment

Macroscopic lesions and microscopic changes in the stomach were observed at the highest
 dose tested (1000 mg/kg bw/d) in a 90-day oral repeat-dose toxicity study in rats.
 Therefore a local NOAEL of 300 mg/kg bw/d is taken as NOAEL for the oral uptake route.

- Concerning systemic effects which are of higher relevance for dermal uptake of cosmetics,
  the SCCS considers 100 mg/kg bw/d as NOAEL based on clinical signs, motor activity scores
  and significantly reduced hindlimb grip strength in female animals at 300 and 1000 mg/kg
- 40 bw/d.
- 41 Clinical signs, mainly hypoactivity and splayed hindlimbs with dragging gait, as observed at
- 42 the mid-dose were more pronounced in the high-dose group, and indicate acute treatment-43 related effects. To which extent these effects are fully reversible (between daily dosing) was
- 44 very poorly documented.
- 45 Concerning motor activity, some statistically significant variations compared to controls 46 were observed at the highest dose, further motor activity scores with a trend for lower 47 values were observed at the highest dose (not statistically significant); thus, observations 48 from motor activity support the findings from clinical observations.
- In female animals, hindlimb grip strength exhibited a dose-dependent decrease with increasing concentration reaching statistical significance (p<0.0001) at 300 and 1000 mg/kg bw/d. The effect can be considered strong (Cohen's d: -3.066 at 300 mg/kg). Although histopathology of femoral muscles, brain or sciatic nerves did not indicate treatment-related changes, clinical signs as well as the results from FOB testing indicate neurotoxicity. Reductions in hindlimb grip strength are considered adverse due to the following reasons:
- 56 (1) the effect is accompanied by other neurological findings (e.g. decreased motor activity 57 (with dose-dependent tendency), hypoactivity, effects on hindlimb foot splay);

(2) Although the observed clinical signs such as hypoactivity and splayed hindlimbs might
 be considered reversible, reversibility of findings from FOB has not been demonstrated

3 (3) the histopathological procedure as performed in the study (H&E staining) might not be
 adequate to reliably detect effects on the nerves and furthermore, nerves other than sciatic
 5 nerves might be of relevance

6 (4) possible manifestations in the form of neurotransmitter changes have not been
7 assessed; (5) similar observations pointing to neurotoxicity (e.g. hypoactivity, spread-out
8 hindlimbs and dragging gait) were also described in an independently performed prenatal
9 developmental toxicity study in rats.

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25 26

27 28 By using Benchmark calculation for reduced hindlimb grip strength in female animals, a  $BMD_{10\%}$  of 220 mg/kg bw/d and a  $BMDL_{10\%}$  of 110 mg/kg bw/d was calculated.

3.3.5.3 Chronic (> 12 months) toxicity

No data provided.

## **3.3.6 Reproductive toxicity**

3.3.6.1 Fertility and reproduction toxicity

No data provided.

3.3.6.2 Developmental toxicity

29		
30	Guideline:	OECD TG 414 (2001)
31	Species/strain:	Rat, Wistar
32	Group size:	24
33	Test substance:	R0069279A
34	Batch:	R0069279A 019 D 004
35	Purity:	95.9 ± 0.5 %
36	Vehicle:	$0.5 \ \% \ (w/v)$ carboxymethylcellulose (CMC) in water and $2 \ \% \ Tween \ @$
37		80 (v/v)
38	Dose levels:	0, 100, 300 or 1000 mg/kg bw/day
39	Dose volume:	10 ml/kg
40	Controls:	Vehicle
41	Route:	oral
42	Administration:	gavage on gestation days (GD) 5 – 19.
43	GLP:	in compliance
44	Study period:	October 2011 – Sep 2013
45		

46 Dose selection was based on the outcome of a GLP-compliant preliminary developmental 47 toxicity study according to OECD TG 414, repeatedly administering three oral gavage 48 dosages of HEPB to pregnant Wistar rats on gestation days (GD) 5-19.

In the definitive study, HEPB was administered by daily oral (gavage) doses of 0, 100, 300 49 50 or 1000 mg/kg/day to mated Wistar female rats (24/group) during the sensitive period of 51 organogenesis (day 5 through day 19 of gestation, the day of detection of sperm positive 52 vaginal smear/vaginal plug being designated as gestation day 0 (GD 0)). Maternal 53 evaluations and measurements included daily clinical signs and body weight/food intake 54 measured at designated intervals. The dams were killed on GD 20 and subjected to 55 macroscopic examination. All required litter parameters were recorded and fetuses were 56 sexed, weighed and submitted to external examination. About one half of the fetuses were

Ref.: 15; 16

1 also examined for soft tissue anomalies, and remaining fetuses were examined for skeletal

- 2 anomalies.
- 3 Results:

4 No deaths were observed. Animals given HEPB at 100 mg/kg bw/d did not show any clinical 5 signs while transient clinical signs of hypoactivity were observed on a single occasion in 6 6 out of 24 rats given 300 mg/kg bw/d. At 1000 mg/kg bw/d, most animals showed clinical 7 signs of hypoactivity within 10 to 45 min of dosing, particularly splayed hindlimbs and 8 dragging gait. The study authors therefore considered these effects as non-adverse due to 9 their transient nature. Dosing with HEPB did not produce any changes in maternal body weight, maternal body weight gain and food consumption. There was a statistically 10 significant increase in dams with resorptions in mid- and high-dose groups when compared 11 12 to the vehicle control group (16.67, 38.1, 43.48 and 47.84 % animals with any resorption at 0, 100, 300 and 1000 mg/kg bw/d). Nevertheless, there were no changes in mean early 13 and late resorptions in these groups when compared to the vehicle control group. Therefore, 14 this increase was considered as incidental. No other adverse changes in maternal and litter 15 16 parameters occurred- and there were no fetuses with external malformations. The visceral 17 and skeletal variations observed were of the type and incidence commonly observed in rats 18 of this strain and age.

19 The study authors concluded that HEPB had no teratogenic potential based on the outcome 20 of this study. The study authors set the maternal and developmental NOAEL at 1000 mg/kg 21 bw/d.

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23

#### 24 SCCS Comment

Hypoactivity and effects on hindlimbs were also observed in a 90-day oral repeated-dose
toxicity study (see section 3.3.5.2).

Using Fisher's exact test instead of the Chi2, the number of dams with any resorptions is only significant compared to control animals at 1000 mg/kg. In Chi2, the significant difference was observed at 300 and 1000 mg/kg. The number of resorptions and implantation losses calculated with the Kruskal-Wallis test did not show any significant differences compared to the controls. All means analysed were within historical control ranges. Altogether, this led the SCCS to conclude that no toxicologically meaningful changes were observed.

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## 3.3.7 Mutagenicity / Genotoxicity

3.3.7.1 Mutagenicity / Genotoxicity in vitro

## **Bacterial Reverse Mutation Test**

41		
42	Guideline:	OECD TG 471 (1997)
43	Test system:	Salmonella typhimurium strains TA98, TA100, TA102, TA1535, and
44		TA1537
45	Replicates:	2 experiments
46	Test substance:	R0069279A
47	Batch:	R0069279A 019 D004
48	Purity:	95.9 ± 0.5 %
49	Concentrations:	Experiment I: with and without S9-mix, 0.32, 1.6, 8, 40, 200, 1000
50		and 5000 µg/plate
51		Experiment II: with and without S9-mix, 39.06 <sup>*)</sup> , 78.13 <sup>*)</sup> , 156.3,
52		312.5, 625, 1250, 2500, 5000 μg/plate
53	Treatment:	Experiment I: direct plate incorporation, with and without S9-mix
54		Experiment II: direct plate incorporation without S9-mix and pre-
55		incubation with S9-mix
56	Vehicle:	DMSO
57	Negative Control:	DMSO

1 2 3 4	Positive Control:	without S9: 2-nitrofluorene (TA98); sodium azide (TA 100; TA1535); 9-aminoacridine (TA1537), mitomycin C (TA102) with S9: benzo[a]pyrene (TA98); 2-aminoanthracene (TA100, TA102; TA1535; TA 1537)					
5 6 7	GLP: Study period:	In compliance March – June 2011					
8 9 10	<sup>(*)</sup> Concentration use 0.05 ml additions	d for strains TA1537 and TA102 in the presence of S9-mix only using					
11 12 13 14 15 16 17	The test item was evaluated in two independent experiments in the absence and presence of metabolic activation (S9-mix from livers of Aroclor 1254 treated rats). Experiment I was performed according to the direct plating incorporation method, experiment II with S9-mix was performed according to the pre-incubation method. HEPB was soluble in DMSO. Known recommended mutagens were used as positive controls; three plates per treatment condition were used.						
18 19		ompletely soluble in the aqueous assay system at all concentrations					
20 21 22 23 24 25 26 27 28 29 30 31 32 33	In experiment I, ev bacterial lawn and/or test bacteria was obs the absence and pre- slight thinning in the numbers to a comple in the presence of S 5000 µg/plate and in Concerning mutation statistically significan of S9-mix. The positiv	tested and in each of the experiments performed. In experiment I, evidence of toxicity ranging from a slight thinning of the background bacterial lawn and/or a marked reduction in revertant numbers to a complete killing of the test bacteria was observed at 1000 and/or 5000 $\mu$ g/plate in strains TA1537 and TA102 in the absence and presence of S9-mix. In experiment II evidence of toxicity ranging from a slight thinning in the background bacterial lawn and/or a marked reduction in revertant numbers to a complete killing of the test bacteria was observed in strains TA98 and TA1537 in the presence of S9-mix and TA102 in the absence of S9-mix at 2500 $\mu$ g/plate and/or 5000 $\mu$ g/plate and in strain TA102 at 1250 $\mu$ g/plate and above in the presence of S9-mix. Concerning mutation, no increases in revertant numbers were observed that were statistically significant following treatment of all the test strains in the presence or absence of S9-mix. The positive control chemicals all induced large increases in revertant numbers in the appropriate strains.					
34 35 36	Conclusion HEPB was not mutage	enic in this bacterial gene mutation test. Ref.: 1					
37 38 39	Mammalian Cell Ge	ne Mutation Test in Mouse Lymphoma Cells (hprt locus)					
40	Guideline:	OECD TG 476 (1997)					
41	Test system:	Mouse lymphoma cell line L5178Y [hprt locus]					
42	Replicates:	duplicate					
43	Test substance:	R0069279A					
44 45	Batch: Purity:	R0069279A 019 D004 95.9 ± 0.5 %					
46	Concentrations:	Experiment I:					
47	concentrations.	without S9-mix: 300, 600, 900, 1100, 1300, 1500, 1900 and 2083					
48		µg/ml					
49		with S9-mix: 300, 600, 1300, 1500 and 1900 µg/ml					
50		Experiment II:					
51		400, 800, 1200, 1400, 1600, 1700, 1800, 1900, 2000 and 2083					
52	Treatment	µg/ml (with and without S9-mix)					
53 54	Treatment:	Experiment I and II: 3 hour treatment without and with S9-mix; an expression period of 7 days and a selection period of 12-13 days					
55	Vehicle:	DMSO					
56	Negative Control:	DMSO, diluted 100-fold in treatment medium					
57	Positive Control:	4-nitroquinoline 1-oxide (without S9-mix)					

- benzo[a]pyrene (with S9-mix)
   GLP: In compliance
   Study period: July August 2011
   Concentrations selected for the mutation experiments were based on the results of a
   cytotoxicity range-finder experiment using 2083 µg/ml as highest concentration; at this
- 7 concentration, precipitate was observed, which did not persist during incubation. No marked 8 changes in osmolality or pH were observed at 2083  $\mu$ g/ml, but excessive toxicity was 9 observed at this dose level.
- 10 Both experiments used a 3-hour treatment. Known mutagens in the presence 11 (Benzo(a)pyrene, BP) or absence of S9-mix (4-nitroquinoline 1-oxide, NQO) were tested at 12 two different concentrations and served as positive controls. Negative controls consisted of 13 cultures treated with the vehicle DMSO diluted 100-fold in the treatment medium
- 13 cultures treated with the vehicle DMSO diluted 100-fold in the treatment medium.
  14
- 15 Results
- In experiment I, the highest concentrations analysed were 2083  $\mu$ g/ml in the absence of S9-mix and 1900  $\mu$ g/ml in the presence of S9-mix, which gave relative survival rates of 14 % and 12 %, respectively. In experiment II, the highest concentration analysed was 2083  $\mu$ g/ml both in the absence and in the presence of S9-mix, which resulted in relative survival rates of 49 % and 27 %, respectively.
- In both experiments no statistically significant increases in mutant frequency were observed following treatment with any concentration of HEPB tested in the absence or presence of
- 23 S9-mix. Two concentrations of the respective positive controls revealed a clear response.
- 24
- 25 Conclusion
- 26 Under the test conditions used HEPB was not mutagenic in this gene mutation test in
  27 mammalian cells.
  28
  - Ref.: 18

#### 31 SCCS comment

The required level of toxicity (10-20% relative survival after the highest concentration) was not reached in experiment II.

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#### In Vitro Micronucleus Test in Cultured Human Lymphocytes

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38	Guideline:	OECD TG 487 (2010)
39	Test system:	lymphocyte cultures from the pooled blood of two male donors
40	Replicates:	duplicate (four replicates for vehicle)
41	Test substance:	R0069279A
42	Batch:	R0069279A 019 D 004
43	Purity:	95.9 ± 0.5 %
44	Concentrations:	Experiment I: without S9-mix 1500, 1900, 1980 and 2020 $\mu$ g/ml
45		Experiment II: with S9-mix 1500, 1780, 1860 and 1900 $\mu$ g/ml
46		Experiment III: without S9-mix 100, 250, 300 and 350 µg/ml
47	Treatment:	Experiment I: 3 h treatment
48		Experiment II: 3 h treatment
49		Experiment III: 24 h treatment
50	Vehicle:	DMSO
51	Negative control:	Vehicle
52	Positive controls:	Mitomycin C (MMC) (without S9-mix)
53		Vinblastin (VIN) (without S9-mix)
54		Cyclophosphamide (CPA) (with S9-mix)
55	GLP:	In compliance
56	Study period:	March - April 2011
57		

HEPB was assayed for the induction of micronuclei in cultured human peripheral blood lymphocytes from the pooled blood from two male volunteers in the absence and presence of metabolic activation (S9-mix prepared from the livers of Aroclor 1254-treated rats). The highest concentration in each test condition was selected on the basis of solubility and cytotoxicity criteria.

Duplicate cultures were treated with each concentration of HEPB or with known clastogens
in the presence (cyclophosphamide) or absence of S9-mix (mitomycin C and vinblastine).
Vehicle-treated cultures (DMSO, four replicates) were used as negative controls.

Blood cultures were incubated in the presence of the mitogen phytohaemagglutinin (PHA) 10 for 48 hours and then treated for 24 or 3 hours in the absence or presence of S9-mix, 11 12 respectively. Cells were harvested 72 hours after the beginning of incubation. Cytochalasin B was added after the 3-hour treatments or before the 24-hour treatments. Lymphocyte 13 preparations were stained and examined microscopically for determining the replication 14 15 index (RI) and the proportion of micronucleated binucleated (MNBN) cells when selected. 16 Where possible, 2000 binucleate cells per concentration (one thousand from each replicate) 17 were analysed blindly.

## 19 Results

18

No marked changes in osmolality or pH were observed at the highest concentration tested
 in the range-finder (2083 μg/ml), compared to the concurrent vehicle controls.

22 When compared to concurrent solvent controls, treatment of cultures with positive controls 23 CPA, MMC and VIN resulted in consistent significant increases in MNBN frequencies, thus 24 validating the sensitivity of the test system and procedure used. The MNBN cell frequencies 25 in all treated cultures under both treatment conditions without S9-mix fell within the normal ranges with the exception of one culture at 1980  $\mu$ g/ml following the 3 hour treatment, 26 27 which fell slightly below the normal range and was considered of no biological relevance. In 28 the experiment with S9-mix, the MNBN cell frequency exceeded the normal range in one of 29 the cultures where a 64% reduction in replication index was observed; therefore the study 30 authors considered this observation in a single culture at high toxicity of little or no 31 biological relevance.

- 32
- 33 Conclusion

34 Under the test conditions used HEPB did not induce biologically relevant increases in the 35 number of cells with micronuclei in cultured human peripheral blood lymphocytes and 36 consequently was not mutagenic (clastogenic/aneugenic) in this micronucleus test.

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#### 39 SCCS comment

Ref.: 17

Results for the 3-hour incubation experiment were from a repeat experiment. In the initial trial, steep concentration-related toxicity was observed under these two treatment conditions and a concentration giving the required level of toxicity could not be identified. The experiment was therefore repeated using more closely spaced concentrations at the upper end of the concentration range.

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## 46 SCCS conclusion on mutagenicity

HEPB was investigated in genotoxicity tests for the 3 endpoints of genotoxicity: gene mutations, structural and numerical chromosome aberrations. HEPB did not induce gene mutations in bacteria nor in mammalian cells when evaluated at the hprt locus. Exposure of human lymphocytes with HEPB did not result in an increase of micronucleated binucleated cells. Based on the present available tests, HEPB can be considered to have no genotoxic potential and additional tests are unnecessary.

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55	3.3.6.2	Mutagenicity / Genotoxicity in vivo
56		

57 No data provided.

#### 3.3.8 Carcinogenicity

No data provided.

#### 3.3.9 **Photo-induced toxicity**

3.3.9.1 Phototoxicity / photo-irritation and photosensitisation 12 13 OECD TG 432 (2004) Guideline: 14 Test system: Balb/c 3T3 mouse fibroblasts, clone 31 15 Replicates: 2 irradiated and 2 non-irradiated microplates (6 wells per 16 concentration) 17 Test substance: R0069279A 18 Batch: R0069279A 019 D 004 95.9 ± 0.5 % (HPLC) 19 Purity: Test concentrations: 20 3.91, 7.81, 15.625, 31.25, 62.5, 125, 250, 500 and 1000 µg/ml 21 Vehicle: PBS<sup>+</sup> 22 Source of light: Atlas CPS+ equipped with 2 anti-UVB filters 23 UVA irradiance: about 1.7 mW/ cm<sup>2</sup> 5 J/cm<sup>2</sup> 24 Irradiation dose: 25 Irradiation duration: about 50 minutes 26 Incubation time:  $60 \pm 10$  minutes (after test substance addition) 27  $20 \pm 2$  hours (after irradiation) chlorpromazine (0.781 – 200 µg/ml without irradiation; 28 Positive control: 29  $0.039 - 10 \,\mu\text{g/ml}$  with irradiation) 30 Negative control: non-irradiated cells in PBS<sup>+</sup> 31 GLP: in compliance 32 Study period: Feb – March 2014

- 33 34
- 35 Sensitivity of the cells to UVA irradiation was investigated using irradiation doses of 1, 5, 9 36 and 11  $J/cm^2$ . In a preliminary cytotoxicity experiment, cytotoxicity was investigated using 37 test item concentrations from 3.906 – 1000  $\mu$ g/ml and an incubation time of 24 ± 3 hours.
- 38 Irradiation doses and test item concentrations for the definitive irradiation experiment were 39 based on the outcome of these investigations.

40 In the definitive experiments, 2 microplates each were used for irradiated and non-41 irradiated conditions. Cells were seeded in microplates and incubated for  $24 \pm 3$  hours. Test 42 substance or positive controls were then applied onto the cells, followed by a  $60 \pm 10$  min 43 incubation period. Irradiation plates were then irradiated for 48 (positive control) or 47 (test 44 item) minutes, while non-irradiated plates were kept in the dark at room temperature. After 45 an incubation period of  $20 \pm 2$  hours, cells were checked for morphology and a neutral red 46 uptake viability assay was subsequently conducted. The photo irritation factor (PIF) and the 47 mean photo effect (MPE) were calculated by "Phototox 2.0" software. 48

49 Results

Sensitivity of cells to light was demonstrated as cells irradiated with 5 and 9 J/cm<sup>2</sup> displayed 50 90 % and 75 % viability, respectively, compared to non-irradiated cells. In the preliminary 51 52 cytotoxicity test, no concentration leading to 50 % mortality (IC<sub>50</sub>) could be established. 53 Thus, a PIF could not be calculated for the test item and the phototoxic potential was 54 evaluated by MPE. For HEPB, an MPE of  $0.016 \pm 0.022$  was calculated.

1 Conclusion

2 A test substance is considered not phototoxic when the MPE is < 0.1. The positive control 3 chlorpromazine revealed a PIF of 40.933 (i.e. >6 indicating phototoxic potential). Based on 4 these results the study authors concluded that HEPB is not phototoxic. 5

Ref.: 5

### **SCCS** comment

In view of the absence of indications of phototoxicity at 1000  $\mu$ g/ml, the SCCS agrees that under the conditions of the test, phototoxicity of HEPB is unlikely.

3.3.9.2 Phototoxicity / photomutagenicity / photoclastogenicity

No data provided.

#### 3.3.10 Human data

No data provided.

22 3.3.11 Special investigations

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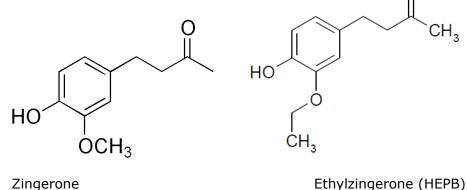
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24 Excretion and metabolism of zingerone, a structurally similar compound to HEPB was 25 described in a publication from the open literature. The structural difference between 26 zingerone (4-(4-Hydroxy-3-methoxyphenyl)butan-2-one; CAS 122-48-5) and 27 ethylzingerone (HEPB) consists in a methylene group in the alkoxy-chain at the aromatic 28 ring of the molecules. Therefore, results obtained from zingerone may allow conclusions to 29 be drawn on the excretion and metabolism of HEPB. 30

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After oral administration of a single dose of 100 mg/kg bw of zingerone to male albino 37 38 (Wistar-derived) rats, urine and faeces were collected in 24 hr periods. For identification of 39 metabolites, free and total fraction (after hydrolysis) were analysed. Within 24 hours, a mean of 95 % of the applied dose had been excreted via urine. A mean of 56 % of the 40 applied dose was excreted unmetabolised (i.e. as parent compound), the remainder was 41 excreted as metabolites (most prominent identified metabolites: zingerol (11 % of the 42 43 applied dose; 4-(3,4-dihydroxyphenyl)butan-2-one (6%) and homovanillic acid (5%). In 24 44 hr – 48 hr urine samples, only traces of zingerone and zingerol were detected. These results

1 2 3 4	demonstrate that zingerone is almost completely absorbed, excreted after oral intake.	metabolised and readily Ref.: 19
5	3.4 Exposure assessment	
6		
7	No data provided.	
8		
9	3.5 Safety evaluation (including calculation of the N	MoS)
10	5.5 Safety evaluation (including calculation of the r	105)
11		
12		
13	CALCULATION OF THE MARGIN OF SAF	ЕТҮ
14		
15		
16	3.5.1 Aggregate exposure from the use as a cosmetic prese	ervative in rinse-off,
17	oral care and leave-on cosmetic products	
18		50.0/
19	Absorption through the skin (default) DAp (%)	= 50 %
20 21	Amount of cosmetic product applied daily A (g/d) Concentration of ingredient in finished product C (%)	= 17.4 g/d = 2 %
22	Typical body weight of human	= 2 % = 60 kg
23	Systemic exposure dose (SED) =	- 00 kg
24	A $(g/d) \times 1000 \text{ mg/g} \times C (\%)/100 \times \text{Dap} (\%)/100 /60$	= 2.9 mg/kg
25	bw/d	
26		
27	No adverse observed effect level NOAEL	= 100 mg/kg bw/d
28	(systemic NOAEL, 90d oral repeat dose toxicity study, rat)	
29	No adjustment, 100 % oral absorption	
30		
31	MOS NOAEL/SED = 34.5	
32		

## 3.5.2 Oral care cosmetics

For oral hygiene products, the relative calculated daily exposure to cosmetic product is 34.7
mg/kg bw/d (2.16 mg/kg bw/d for toothpaste (adults) and 32.54 mg/kg bw/d for
mouthwash), according to the 9<sup>th</sup> revision of the SCCS NoG (SCCS 1564/15).

Product type	Calculated relative daily exposure [mg/kg bw/d]	NOAEL [mg/kg bw/d]	MoS based on NOAEL
Toothpaste (adults) and mouthwash combined	0.694	100	144.1
Toothpaste (adults)	0.043	100	2326
Mouthwash	0.651	100	153.6

39 No adjustment, 100 % oral absorption

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3.5.3	Dermally	applied	cosmetic	products
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Product type	SED	NOAEL	MoS
	[mg/kg bw/d]	[mg/kg bw/d]	
All types of products	2.52	100	39.7
Rinse-off	0.09	100	1111
skin & hair cleansing products			
Leave-on skin and hair care products	2.24	100	44.6
Make-up products	0.19	100	526.3
Leave-on skin and hair care products and make-up products combined	2.42	100	41.3

3 4 5

## The following parameters were used to determine SED for dermally applied cosmetic products:

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7		
8	Concentration of ingredient in finished product C (%)	= 2 %
9	Typical body weight of human	= 60 kg
10	Systemic exposure dose (SED) =	-
11	A (g/d) x 1000 mg/g x C (%)/100 x Dap (%)/100 /60	
12	bw/d	
13		
14	No adverse observed effect level NOAEL	= 100 mg/kg bw/d
15	(systemic NOAEL, 90d oral repeat dose toxicity study, rat)	
16	No adjustment, 100 % oral absorption	
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50		

1 Exposure calculated according to Table of the SCCS NoG, 9<sup>th</sup> revision:

Type of exposure	Product	g/d	mg/kg bw/d
Rinse-off skin & hair cleansing products	Shower gel	0.19	2.79
	Hand wash soap	0.20	3.33
	Shampoo	0.11	1.51
	Hair conditioner	0.04	0.67
	Body lotion	7.82	123.20
	Face cream	1.54	24.14
Leave-on	Hand cream	2.16	32.70
skin & hair care products	Deo non-spray	1.50	22.08
	Hair styling	0.40	5.74
Make-up products	Liquid foundation	0.51	7.90
	Make-up remover	0.50	8.33
	Eye make-up	0.02	0.33
	Mascara	0.025	0.42
	Lipstick	0.06	0.90
	Eyeliner	0.005	0.08
Oral care cosmetics	Toothpaste	0.14	2.16
Oral care cosmetics	Mouthwash	2.16	32.54
TOTAL		± 17.4	269

3 4

## 5 3.6 Discussion

# 67 *Physico-chemical properties*

8 9 Depending on the temperature, hydroxyethoxyphenyl butanone may appear as a solid (white powder or crystals) or as a pale yellow liquid form. Reported purities in the different 10 batches of HEPB were 95.9 % and higher. Two impurities (2-ethoxy-4-(3-hydroxybutyl) 11 12 phenol (up to 4.8 % in different batches reported) and ethylvanillin (< 1000 mg/g)) have 13 been quantified using reference standards. Four other impurities have been detected with a 14 relative UV purity < 0.1%. Water solubility is 7.59 g/L at 20°C, Log Pow is 1.46 at 22.8°C and the vapour pressure is  $8.7 \times 10^{-3}$  Pa at 25 °C. The substance is stable over 2 months at 15 16 45°C in hydroalcoholic solution at 0.5g/100ml. It is sensitive to photostress but resistant to 17 oxidative, heat, acid or basic stresses.

# 19 Function and uses20

The ingredient HEPB is intended to be used specifically as a preservative in rinse-off, oral care and leave-on cosmetic products up to 2%.

## 24 **Toxicological Evaluation**

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# 26 Acute toxicity27

No acute toxicity study on any route is available for HEPB. In 14- and 90-day oral repeat dose studies using dose levels of 100, 300 and 1000 mg/kg bw/d, no deaths occurred. Hence, it can be assumed that the oral LD50 would be higher than 1 000 mg/kg/d (i.e. the substance is of low acute oral toxicity).

# 3233 Local toxicity

- 34
- Based on an in vitro Episkin<sup>SM</sup> Skin Irritation Test performed with the neat substance, HEPB
   can be considered as potentially non-irritant to the skin.

Data on mucous membrane irritation based on the Bovine Corneal Opacity and Permeability
 (BCOP) Test Method are inconclusive.
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### Skin Sensitisation

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HEPB is not considered to be a skin sensitiser.

## 8 Dermal absorption

10 The in vitro percutaneous absorption of HEPB by using a typical lipophilic skin care 11 formulation containing HEPB at the concentration of 2% was determined in human 12 dermatomed skin. A recovery of at least 85 % was only obtained for 4 cells evaluated. The 13 number of 4 evaluable cells is not in line with the SCCS criteria. Further, the in vitro 14 percutaneous absorption has not been determined in aqueous cosmetic formulation.

Based on significant deviations from requirements, the SCCS considers the study not
acceptable. Instead, in the absence of adequate experimental data, a default value of 50 %
dermal absorption is taken for MOS calculation according to SCCS 1564/15.

## 19 **Repeated dose toxicity**

Macroscopic lesions and microscopic changes in the stomach were observed at the highest
dose tested (1000 mg/kg bw/d) in a 90-day oral repeat-dose toxicity study in rats.
Therefore a local NOAEL of 300 mg/kg bw/d is taken as the NOAEL for the oral uptake
route.

Concerning systemic effects which are of higher relevance for dermal uptake of cosmetics, the SCCS considers 100 mg/kg bw/d as the NOAEL based on clinical findings indicating an acute neurotoxic effect and based on FOB findings (statistically significantly reduced hindlimb grip strength observed in females at 300 and 1000 mg/kg bw/d and statistically significant and non-significant findings in some motor activity scores). It is of note that indications for neurotoxicity were not only observed in a 90-day oral repeat dose study in rats but also in a developmental toxicity study performed in rats.

## 33 Mutagenicity

34

HEPB was investigated in genotoxicity tests for the 3 endpoints of genotoxicity: gene mutations, structural and numerical chromosome aberrations. HEPB did not induce gene mutations in bacteria nor in mammalian cells when evaluated at the hprt locus. Exposure of human lymphocytes with HEPB did not result in an increase of micronucleated binucleated cells. Based on the present available tests, HEPB can be considered to have no genotoxic potential and additional tests are unnecessary.

## 42 *Carcinogenicity*

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# 46 *Reproductive toxicity*47

Based on the findings of a developmental toxicity study, it can be concluded that HEPB is
not a developmental toxicant. A separate study addressing fertility has not been performed.

## 51 *Toxicokinetics*

53 Physico-chemical properties and in vivo experimental findings obtained from a structurally 54 similar compound (zingerone) are indicative of almost complete and rapid absorption and 55 excretion. 100 % oral absorption is therefore assumed for MOS calculation.

56 57

## Phototoxicity

In view of the absence of indications of phototoxicity at 1000  $\mu$ g/ml, the SCCS considers that phototoxicity of HEPB is unlikely.

## Human data

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## 12 4. CONCLUSION

- 13
- 14 I. In light of the data provided, does the SCCS consider safe the use of Hydroxyethoxyphenyl Butanone (HEPB) as a cosmetic preservative in rinse-off, oral care and leave-on cosmetic products with a maximum concentration of 2.0 % under an aggregate exposure scenario for cosmetics?
- Based on the information provided by the applicant, the use of Hydroxyethoxyphenyl Butanone (HEPB) as a cosmetic preservative in rinse-off, oral care and leave-on cosmetic products with a maximum concentration of 2.0 % under an aggregate exposure scenario for cosmetics is not considered safe.
- It is of note that a maximum concentration of 2.0 % under an aggregate exposure scenario for cosmetics could be considered safe if more reliable and supporting data on *in vitro* dermal penetration was provided.
- 25
   2. Alternatively, what is, according to the SCCS, the maximum target concentration considered safe for use of Hydroxyethoxyphenyl Butanone (HEPB) as a cosmetic preservative in rinse-off, oral care and leave-on cosmetic products under an aggregate exposure scenario for cosmetics?
- Based on the information provided by the applicant, a maximum concentration of 0.7
  of Hydroxyethoxyphenyl Butanone (HEPB) as a cosmetic preservative in rinse-off,
  oral care and leave-on cosmetic products can be considered safe.
- 32 3. Does the SCCS have any further scientific concerns regarding the use of
   33 Hydroxyethoxyphenyl Butanone (HEPB) in cosmetic products?
- 34 The SCCS does not have any other concerns.
- 35 36
- 37 5. MINORITY OPINION
- 38 39
- 39 / 40
- 41

## 2 6. REFERENCES

4 Toxicity References *in italics* are not submitted as full reports in the present dossier. They 5 consist of reports of dose-range finding studies that can be provided upon request and of 6 documents/guidelines that can be downloaded from the internet (URL provided). 

1. Bowen R (2011). Hydroxyethoxyphenyl Butanone (R0069279A, Batch R0069279A 019 D 004): Reverse Mutation in five histidine-requiring strains of *Salmonella typhimurium*. Covance Study No. 8244263

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## 2 Appendix

6.1.1.1 3 4 Assessment of Recovery of Radiolabelled R0069279A in a Skin Care Formulation 5 This study has been performed in order to explain the low recovery rate in the dermal 6 absorption study. 7 8 Guideline: 9 Determination of recovery of test item when applied to Method: 10 aluminium foil 11 Test substance: R0069279A (non-labelled);  $[^{14}C]$ -R0069279A (labelled) R0069279A 019 P 001 (non-labelled material) 12 Batch: CFO41708 (labelled material) 13 14 Purity:  $96.0\% \pm 1.3\%$  (non-labelled material) 92.2 % (labelled material; HPLC purity) 15 radiodiluted [<sup>14</sup>C]-R0069279A in a cosmetic formulation; 16 Test item: 17 105.85 % of target (2.0 % (w/w) Replicates: 18 8 19 Exposed membrane area:  $3.14 \text{ cm}^2$ 20 Dose applied: ca. 2.0 mg/cm<sup>2</sup> 21 Sampling period: 24 hours 22 Receptor fluid: calcium- and magnesium free phosphate buffered saline (PBS) containing bovine serum albumin (5%, w/v)23 24 Mass balance analysis: Provided 25 not applicable Tape stripping: 26 Method of Analysis: Liquid Scintillation Counting (LSC) In compliance 27 GLP: experimental phase: Feb 2013, Report dated23 October 2014; 28 Study period: 29 draft report Nov 2012 30 Sections of aluminium foil, ca 3 x 3 cm, were cut out, positioned on the receptor chamber of 31 the diffusion cells already connected to the waterbath (heated to maintain a surface 32 temperature of 32°C ± 1°C) in a fume hood. The receptor chambers were filled with 33 34 phosphate buffered saline and a magnetic stirring bar was introduced in each cell. The test 35 item (radiodiluted test item in a combination of one oily and two water-based preformulations) was applied over the exposed surface of 8 samples of aluminium foil. The 36 donor chambers were left open to the atmosphere. To accurately quantify the radioactivity 37 applied to the aluminium foil, seven weighed aliquots of the test preparation were taken 38 39 throughout and analysed by liquid scintillation counting (LSC). At 0 h post dose, the 40 exposure was terminated for 4 samples by placing the aluminium foil and the donor chamber into pre-weighed pots containing ethanol (40 mL). The same procedure was 41 42 carried out for a further 4 samples at 24 h post dose. 43 The radiochemical was then extracted from the test system and each sample was sonicated 44 for ca 10 min. The donor chambers were removed from the pots and duplicate aliquots were 45 analysed by LSC. 46 47 48 49 50 51 52 53

#### 1 Results:

Test Preparation	819181	
Target R0069279A Concentration Actual R0069279A Concentration by Radioactivity		
Application Rate of Test Preparation (mg/cm <sup>2</sup> ) Application Rate of R0069279A (µg equiv./cm <sup>2</sup> )	1.89 mg/cm <sup>2</sup> 39.94 μg equiv./cm <sup>2</sup>	
Distribution	% Applied Dose	
Termination Timepoint (h)	0	24
Mass Balance	94.20	64.01

2 3

The mean mass balance for the aluminium foil samples terminated at 0 h post dose was 4 94.20% of the applied dose and this decreased to 64.01% at 24 h post dose. The results 5 showed a loss of [14C]-R0069279A when applied in Test Preparation 819181 (2%, w/w) to aluminium foil. Recovery at 24 h post dose was variable ranging from 54.54% to 74.57% of 6 7 the applied dose. Recovery was less variable at 0 h post dose ranging from 86.92% to 8 98.48% of the applied dose. This confirmed that there were losses during the processing of

9 the recovery samples.