



HAL
open science

Coxiella burnetii Induces Inflammatory Interferon-Like Signature in α Plasmacytoid Dendritic Cells: A New Feature of Immune Response in Q β Fever

Mignane B. Ka, Soraya Mezouar, Amira Ben Amara, Didier Raoult, Eric Ghigo, Daniel Olive, Jean-Louis Mege

► **To cite this version:**

Mignane B. Ka, Soraya Mezouar, Amira Ben Amara, Didier Raoult, Eric Ghigo, et al.. Coxiella burnetii Induces Inflammatory Interferon-Like Signature in α Plasmacytoid Dendritic Cells: A New Feature of Immune Response in Q β Fever. *Frontiers in Cellular and Infection Microbiology*, 2016, 6, pp.70. 10.3389/fcimb.2016.00070 . hal-01466866

HAL Id: hal-01466866

<https://hal.science/hal-01466866>

Submitted on 12 Oct 2018

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License



Coxiella burnetii Induces Inflammatory Interferon-Like Signature in Plasmacytoid Dendritic Cells: A New Feature of Immune Response in Q Fever

Mignane B. Ka^{1,2}, Soraya Mezouar¹, Amira Ben Amara¹, Didier Raoult¹, Eric Ghigo¹, Daniel Olive² and Jean-Louis Mege^{1*}

¹ Unité de Recherche sur les Maladies Infectieuses Tropicales et Emergentes, UMR 63, Centre National de la Recherche Scientifique 7278, INSERM U1095, IRD 198, Aix-Marseille Université, Marseille, France, ² INSERM UMR 1068, Centre de Recherche en Cancérologie de Marseille, Marseille, France

OPEN ACCESS

Edited by:

Damien F. Meyer,
CIRAD, France

Reviewed by:

Janakiram Seshu,
The University of Texas at
San Antonio, USA
Matteo Bonazzi,
Centre National de la Recherche
Scientifique, France

*Correspondence:

Jean-Louis Mege
jean-louis.mege@univ-amu.fr

Received: 30 March 2016

Accepted: 11 June 2016

Published: 27 June 2016

Citation:

Ka MB, Mezouar S, Ben Amara A, Raoult D, Ghigo E, Olive D and Mege J-L (2016) *Coxiella burnetii* Induces Inflammatory Interferon-Like Signature in Plasmacytoid Dendritic Cells: A New Feature of Immune Response in Q Fever. *Front. Cell. Infect. Microbiol.* 6:70. doi: 10.3389/fcimb.2016.00070

Plasmacytoid dendritic cells (pDCs) play a major role in antiviral immunity via the production of type I interferons (IFNs). There is some evidence that pDCs interact with bacteria but it is not yet clear whether they are protective or contribute to bacterial pathogenicity. We wished to investigate whether *Coxiella burnetii*, the agent of Q fever, interacts with pDCs. The stimulation of pDCs with *C. burnetii* increased the expression of activation and migratory markers (CD86 and CCR7) as determined by flow cytometry and modulated gene expression program as revealed by a microarray approach. Indeed, genes encoding for pro-inflammatory cytokines, chemokines, and type I INF were up-regulated. The up-regulation of type I IFN was correlated with an increase in IFN- α release by *C. burnetii*-stimulated pDCs. We also investigated pDCs in patients with Q fever endocarditis. Using flow cytometry and a specific gating strategy, we found that the number of circulating pDCs was significantly lower in patients with Q fever endocarditis as compared to healthy donors. In addition, the remaining circulating pDCs expressed activation and migratory markers. As a whole, our study identified non-previously reported activation of pDCs by *C. burnetii* and their modulation during Q fever.

Keywords: plasmacytoid dendritic cells, *Coxiella burnetii*, Q fever, interferon, infection

INTRODUCTION

Plasmacytoid dendritic cells (pDCs) are a subset of DCs present in the blood and lymphoid organs and are well-documented as the major type I interferon (IFN)-producing cells (Cella et al., 1999; Colonna et al., 2002). Their resting phenotype is characterized by the expression of chemokine receptors such as CCR5, CXCR4, and microbe sensors including C-type lectins, TLR7, and TLR9. Once activated, they overexpress two specific markers, CD86 (activation) and CCR7 (migratory marker), and produce high levels of type I IFN and inflammatory cytokines (Siegal et al., 1999; Facchetti et al., 2003; Liu, 2005; Martinelli et al., 2007; Gilliet et al., 2008). Plasmacytoid dendritic cells specialize in innate antiviral immunity and are also involved in adaptive immunity (Liu, 2005; Fiorentini et al., 2008). Type I IFNs are produced transiently by pDCs in acute viral infections

and have a limited amplitude (Swiecki and Colonna, 2015). Although produced by pDCs in chronic viral infections such as those due to Human Immunodeficiency Virus (HIV; Bowie and Unterholzner, 2008; Sachdeva et al., 2008), hepatitis C virus (Mengshol et al., 2009), and hepatitis B virus (Xie et al., 2009), the role of type I IFNs is more complex. Indeed, pDCs may contribute to chronicity *via* the dysregulation of type I INF production (Swiecki and Colonna, 2015).

In contrast to viral infections, the role of pDCs in the defense against bacteria is poorly understood. There are few examples of pDC maturation in response to bacteria *in vitro*. *Streptococcus pyogenes* (Veckman and Julkunen, 2008) and *Mycobacterium tuberculosis* (Lozza et al., 2014) increase the maturation of human pDCs, leading to the activation of naive CD4⁺ T cells and Th1 polarization. The ability of pDCs to produce type I IFNs in response to bacterial infections depends on the bacterial strain. While *Borrelia burgdorferi* and *Staphylococcus aureus* induce the production of type I IFNs by pDCs, *S. pyogenes* and *Legionella pneumophila* do not (Veckman and Julkunen, 2008; Eberle et al., 2009; Bekeredjian-Ding et al., 2014). In addition, pDCs may be protective against bacterial infections. Hence, the depletion of pDCs in a murine model of *Chlamydia pneumoniae* infection results in severe and prolonged chronic inflammation (Crother et al., 2012).

The presence of *Coxiella burnetii*, the bacterium responsible for Q fever, within pDCs was recently reported in the lymph nodes of patients with lymphomas (Melenotte et al., 2016). This surprising observation led us to investigate the role of pDCs in Q fever. Q fever is an infectious disease characterized by a primary-infection which may become chronic in patients with an altered immune response. The chronic expression of the disease is dominated by endocarditis or vascular infection (Raoult et al., 2005). Despite numerous studies, the mechanisms of Q fever endocarditis and vascular infection remain partially understood. We, and other teams, have reported the modulation of circulating leukocytes, including monocyte and T cell subsets (Ka et al., 2014, 2015), a non-protective inflammatory response (Honstetter et al., 2003; Schoffelen et al., 2013) and the prominent role of IL-10 in bacterial persistence and defective microbicidal activity (Amara et al., 2012). Although, it is largely admitted that monocytes and macrophages are the targets of *C. burnetii*, there is evidence that other cell types, including DCs, may host the microorganisms (Shannon et al., 2005). We recently showed that *C. burnetii* activates human monocyte-derived dendritic cells (moDCs), inducing a transcriptional inflammatory program in which the type I IFN pathway is impaired (Gorvel et al., 2014). However, the interaction of *C. burnetii* with other DC populations has not yet been reported.

In this report, we show that *C. burnetii* induces a migratory phenotype and a specific inflammatory signature in pDCs. *Coxiella burnetii* also stimulates the release of type I IFNs. In addition, the number of circulating pDCs was lower in patients with Q fever endocarditis. Taken as a whole, the present study shows that pDCs are involved in *C. burnetii* infection and identifies a new feature of the immune response in Q fever.

MATERIALS AND METHODS

Patients with Q Fever Endocarditis

The study of 17 patients with Q fever endocarditis (consisting of six women and eleven men aged between 22 and 79 years old) and their controls was conducted with the approval of the Ethics Committee of Aix-Marseille University (Marseille, France) and with the written consent of each participant. The characteristics of the patients were previously described (Ka et al., 2014). Seventeen patients with Q fever endocarditis were included on the basis of the presence of endocarditis, a positive echocardiogram and blood culture, high titers of IgG specific for phase I *C. burnetii* and data scoring (Raoult, 2012). Ten age- and sex-matched individuals were included as healthy controls.

Bacteria Production and Preparation

Coxiella burnetii (Nine Mile strain, RSA496) was cultured as previously described (Gorvel et al., 2014). The L929 cells were infected for 8 days and were sonicated and centrifuged at 300 × *g* for 10 min. Supernatants were collected and centrifuged at 10,000 × *g* for 10 min. Bacteria were then washed and stored at −80°C. The concentration of organisms was determined by Gimenez staining and bacterial viability was assessed using the LIVE/DEAD BacLight bacterial viability kit (Molecular Probes, Life Technologies).

pDC Isolation and Stimulation

Leukopacks were obtained from the Etablissement Français du Sang. Peripheral blood mononuclear cells (PBMCs) were recovered using density gradient centrifugation. The pDCs were isolated using magnetic beads on AutoMacs (Miltenyi Biotech) as previously described (Ka et al., 2014). Briefly, pDCs were isolated by depletion of non-pDCs that were retained in the column, while unlabeled pDCs with high purity (90%) were collected in the flow-through. Plasmacytoid dendritic cells were then suspended in RPMI 1640, supplemented with 20 mM HEPES, 10% fetal calf serum, 2 mM L-glutamine, 100 U penicillin/ml, 50 µg/ml streptomycin (Life Technologies), and 10 ng/ml recombinant IL-3 (R&D Systems), as described previously (Dental et al., 2012) and were stimulated with heat-inactivated (100°C for 30 min) *C. burnetii* organisms (bacterium-to-cell ratio of 50:1), or CpG-A 10 µg/mL (ODN 2216; InvivoGen) for 8 or 24 h.

Flow Cytometry

Flow cytometry was used to study isolated pDCs within PBMCs. First, isolated pDCs were analyzed according to the expression of activation (CD86) and maturation (CCR7) markers. In brief, after stimulation, isolated-pDCs were washed and then incubated with CD86 or CCR7 antibodies or isotypic controls for 20 min. After fixation with 4% paraformaldehyde, the expression of markers was evaluated by flow cytometry. Second, PBMCs from patients with Q fever endocarditis or healthy controls were incubated with specific antibodies or isotypic controls and Aqua-Fluorescent Reactive dye (Aqua-vid), a viability dye, for 20 min (**Supplementary Table 1**). PBMCs were then washed and fixed in 4% paraformaldehyde. Flow cytometry was performed using an LSRII-SORP cytometer (Becton Dickinson). After 500,000 events had been completed and debris on the forward/side scatter dot

plot and dead cells had been excluded with Aqua-Fluorescent Reactive dye, data were exported and analyzed with FlowJo Software (version 9.2, Tree Star Ashland) (Ka et al., 2014). The proportion of pDCs is expressed as the ratio of living cells expressing the fluorescent marker to the total number of analyzed PBMCs. The phenotypic expression of HLA-DR and PD-1 is expressed as the mean fluorescence intensity (MFI).

Microarrays

Isolated pDCs were stimulated with heat-inactivated *C. burnetii* for 8 h. RNA was extracted using RNeasy Mini Kits (Qiagen) with a DNase I step to eliminate DNA contaminants, as previously described (Gorvel et al., 2014). The quantity and quality of the RNA was assessed using a Nanodrop spectrophotometer (Thermo Science) and a 2100 Bioanalyzer (Agilent Technologies), respectively. The microarray study was performed using the technology provided by Agilent Technologies, consisting of microarray chips including 45,000 probes (4x44K Whole Human Genome G4112F) and One-Color Microarray-Based Gene Expression Analysis. In brief, 400 ng RNA were labeled with cyanine-3 CTP using a Low RNA Input Fluorescent Amplification kit. Three biological replicates per group were labeled using a QIAamp labeling kit, deposited on microarray slides, and hybridized for 17 h at 65°C. The microarray slides were scanned with a pixel size of 5 μm using a DNA Microarray scanner G2505B. The scanned images were analyzed with Feature Extraction Software 10.5.1. The data processing and analysis were performed using the R software package (version 2.15). The raw data were filtered and normalized using the Agi4x44 PreProcess library. Unsupervised and supervised analyses were performed using hierarchical clustering and a principal component analysis. Genes were considered to be differentially expressed if the False Discovery Rate was below 0.1% and the absolute fold change was above 2.0 (Gorvel et al., 2014).

Quantitative Real Time-Polymerase Chain Reaction (qRT-PCR)

Specific genes of stimulated pDCs that were found to be highly modulated by microarray were selected and validated by qRT-PCR, as previously described (Ben Amara et al., 2013). Briefly, first-strand cDNA was obtained using oligo(dT) primers and a reverse transcription of 150 ng RNA using a Moloney murine leukemia virus-reverse transcriptase kit (Life Technologies), a ABI7900 Fast Real-Time PCR System, and a SYBR Green Fast Master Mix (Roche Diagnostics). The results were normalized to the housekeeping gene β-actin (ACTB) and were expressed as the median of fold change = $2^{-\Delta\Delta Ct}$, where $\Delta\Delta Ct = (Ct_{Target} - Ct_{Actin})_{assay} - (Ct_{Target} - Ct_{Actin})_{control}$, as previously described (Ben Amara et al., 2010).

Immunoassays

Isolated pDCs were stimulated with heat-inactivated *C. burnetii* organisms for 24 h, and the supernatants were centrifuged at $1000 \times g$ for 10 min and frozen at -80°C . In parallel, PBMCs from healthy controls and Q fever patients were cultivated for

24 h and supernatants were treated as above. The release of IFN-α, IL-6, TNF-α, and IL-10 was determined using immunoassay kits. The concentration of IL-10 and IFN-α in patients was also analyzed by immunoassay.

Statistical Analysis

Data were analyzed using the Student's *t*-test to analyze *in vitro* data and the Mann-Whitney *U*-test to study Q fever patients. The results are presented as the median or the mean \pm SD of three independent experiments, and a $p < 0.05$ was considered statistically significant.

RESULTS

Coxiella burnetii Stimulates Human pDCs

We wanted to investigate whether *C. burnetii* interacts with human pDCs. Living pDCs were isolated from the PBMCs of healthy donors and the expression of BDCA2 and CD123, two specific markers of pDCs, was determined by flow cytometry (Figure 1A). After 24 h of stimulation by *C. burnetii*, the expression of the migratory marker CCR7 and the activation marker CD86 significantly increased as compared to unstimulated pDCs although their expression remained lower than that of pDCs activated with CpG-A used as positive control (Figure 1B), demonstrating that pDCs responded to *C. burnetii* stimulation. We completed the investigation of the response of pDCs to *C. burnetii* using an unbiased approach based on transcriptome study with three biological replicates per group. *Coxiella burnetii* induced a gene expression program which was clearly distinct from that of resting pDCs, as shown by principal component analysis (Figure 1C, left panel) and hierarchical clustering (Figure 1C, right panel). Using multiclass analysis and a fold change of 2.0, we found that 1109 probes were up-regulated and 665 were down-regulated after 8 h of stimulation with *C. burnetii*. The functional annotation enabled keywords enriched in *C. burnetii*-stimulated pDCs to be identified (Table 1). They included an inflammatory response (enrichment score of 17.83), pathogen sensing (enrichment score of 9.93), and a regulation of cell proliferation (enrichment score of 5.63) in up-regulated genes. We selected the genes encoding for IL-6, IL-1β, IL-15, and CD40 because they were highly up-regulated in microarray and we confirmed their over-expression by qRT-PCR (Figure 1D). Taken together, these results show that *C. burnetii* highly stimulates human pDCs.

Coxiella burnetii Stimulates Inflammatory Response in pDCs

As inflammatory genes were modulated in *C. burnetii*-stimulated pDCs, we investigated the secretion by pDCs of TNF-α and IL-6, two cytokines considered as inflammatory cytokines. *Coxiella burnetii* induced the release of TNF-α but not that of IL-6. Surprisingly, *C. burnetii* stimulated the release of IL-10, an anti-inflammatory cytokine (Figure 2A), demonstrating subtle changes in the inflammatory response of pDCs to *C. burnetii*. In the microarray signature, two keywords, namely “inflammatory response” and “pathogen sensing” were enriched in *C. burnetii*-stimulated pDCs; they included genes encoding cytokines and

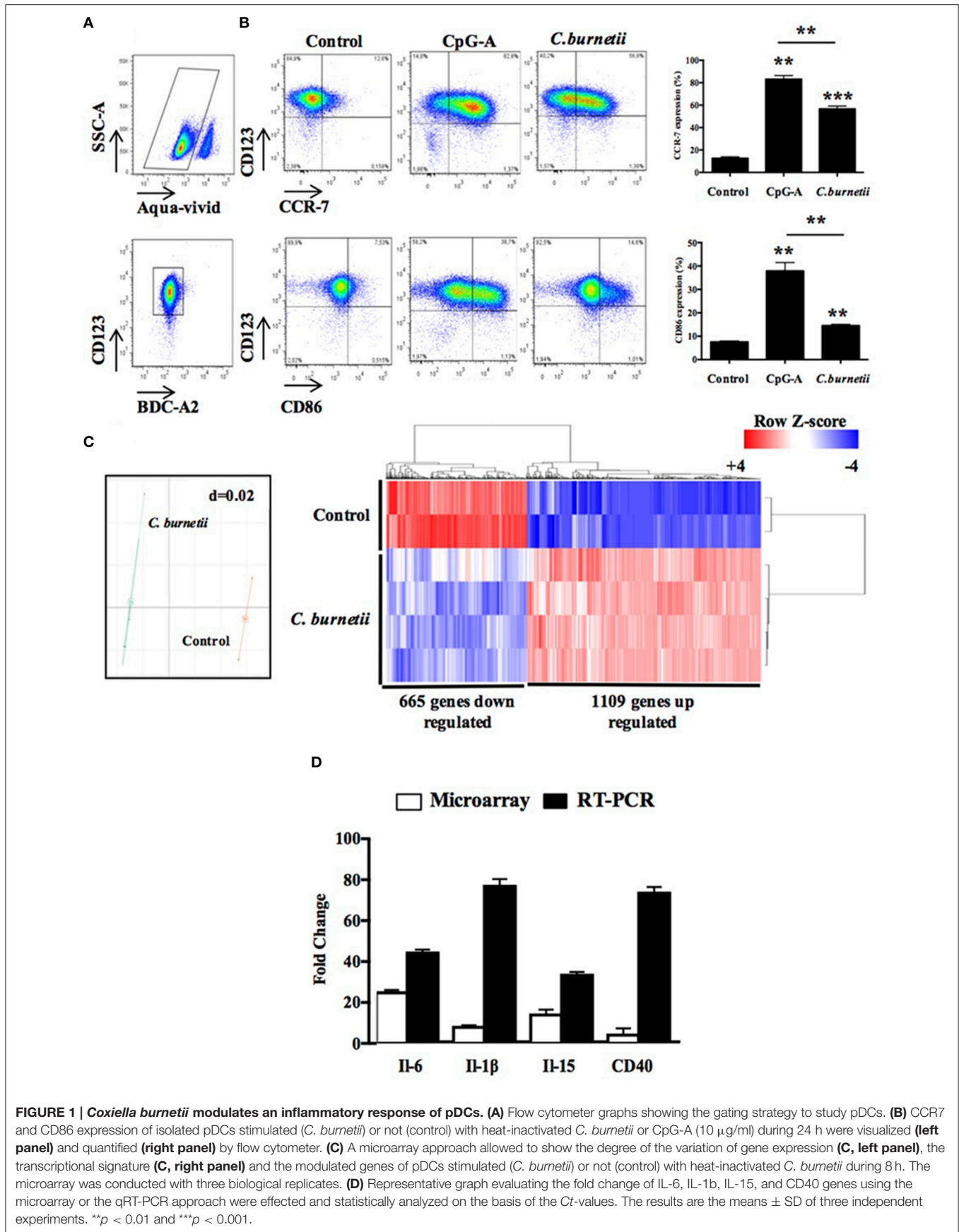


FIGURE 1 | *Coxiella burnetii* modulates an inflammatory response of pDCs. (A) Flow cytometer graphs showing the gating strategy to study pDCs. **(B)** CCR7 and CD86 expression of isolated pDCs stimulated (*C. burnetii*) or not (control) with heat-inactivated *C. burnetii* or CpG-A (10 μg/ml) during 24 h were visualized (**left panel**) and quantified (**right panel**) by flow cytometer. **(C)** A microarray approach allowed to show the degree of the variation of gene expression (**C, left panel**), the transcriptional signature (**C, right panel**) and the modulated genes of pDCs stimulated (*C. burnetii*) or not (control) with heat-inactivated *C. burnetii* during 8 h. The microarray was conducted with three biological replicates. **(D)** Representative graph evaluating the fold change of IL-6, IL-1β, IL-15, and CD40 genes using the microarray or the qRT-PCR approach were effected and statistically analyzed on the basis of the Ct-values. The results are the means ± SD of three independent experiments. ***p* < 0.01 and ****p* < 0.001.

TABLE 1 | Up-regulated genes of pDCs-stimulated by *C. burnetii*.

Keywords	P-values
INFLAMMATORY RESPONSE (ES: 17.83-463 genes; INFA, INFB1, INFE, IL1A, IL1B, TNF, IL-6, IL28A, IL28B, IL-29, IL12A, IL12B, IL13, IL15, IL8, CCL2, CCL8, CCL8, CXCL2, CXCL9)	
Cytokine activity	1.2×10^{-37}
Cytokine cytokine receptor interaction	4.0×10^{-28}
Immune response	1.9×10^{-24}
Defense response	7.8×10^{-21}
Inflammatory response	1.1×10^{-12}
PATHOGEN SENSING (ES: 9.93-80 genes; INFA, INFB, INFE, IL1B, IL28A, IL28B, IL-29, IL12A, IL12B, IL-6, IL8, IL13, IL15, TNF, CCL4, CCL10, CCL10, CXCL9, CXCL11)	
RIG-I-like receptor signaling pathway	5.5×10^{-18}
Toll-like receptor signaling pathway	2.4×10^{-17}
Jak-STAT signaling pathway	1.6×10^{-15}
Cytosolic DNA-sensing pathway	8.4×10^{-12}
REGULATION OF CELL PROLIFERATION (ES: 5.63-88 genes; CD38, CD40, TNF, IL1B, IL4, IL6, IL13, IL15, IL12A, IL12B, IL2R)	
Positive regulation of lymphocytes proliferation	4.9×10^{-10}
Positive regulation of leukocytes proliferation	6.1×10^{-10}
Positive regulation of mononuclear cells proliferation	6.1×10^{-10}
Positive regulation of lymphocytes proliferation	3.7×10^{-9}
Regulation of lymphocytes proliferation	4.0×10^{-9}
Regulation of lymphocytes proliferation	4.6×10^{-9}
Regulation of apoptosis	1.5×10^{-7}
Regulation of programmed death	1.9×10^{-7}
Regulation of death cells	2.0×10^{-7}
Anti-apoptosis	1.5×10^{-5}
Negative regulation of apoptosis	9.9×10^{-5}
Negative regulation of programmed death	1.1×10^{-4}

Identification of three clusters defined as: Inflammatory response, Pathogens sensing, and regulation of cell proliferation.

chemokines and type I and III IFNs (IFN- α , IFN- β 1, and IFN- ϵ) with a fold change higher than 15 (Table 1). As IFN- α was documented as a cytokine specifically expressed by pDCs, we measured its release in supernatants from pDCs stimulated (or not) with *C. burnetii* or CpG-A, a potent agonist of pDCs. While resting pDCs did not release IFN- α , *C. burnetii*-stimulated pDCs released ~ 1.5 ng/ml IFN- α (Figure 2B), demonstrating that *C. burnetii* stimulated type I IFN expression at both transcriptional and transductional levels. Taken together, these results showed that *C. burnetii* affected the inflammatory response of pDCs and particularly the IFN- α response.

Circulating pDCs Are Affected in Patients with Q Fever Endocarditis

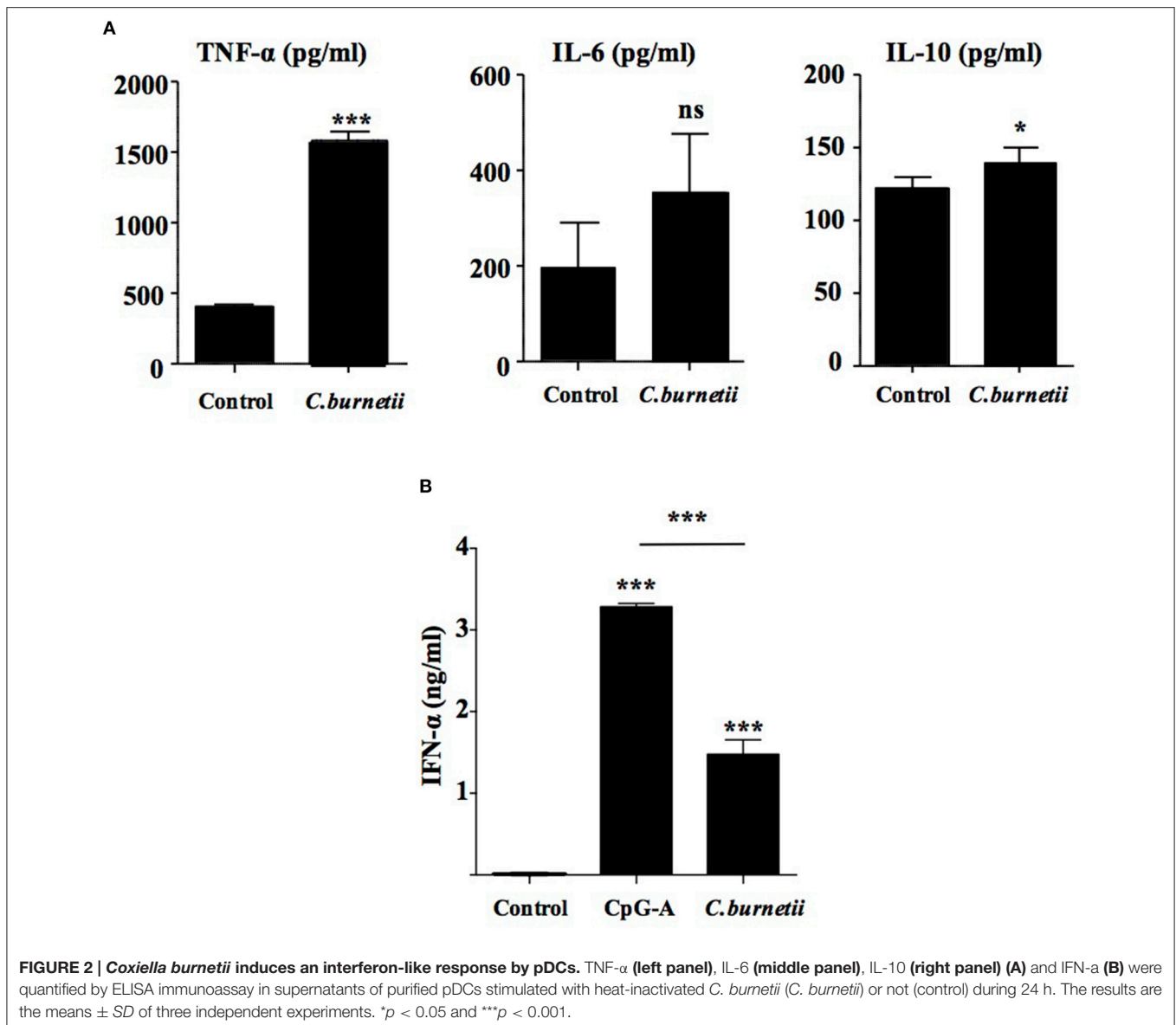
As pDCs are activated *in vitro* by *C. burnetii*, we wanted to investigate whether they are modulated in patients during Q fever endocarditis. We investigated the pDC population in blood samples from healthy donors ($n = 10$) and Q fever endocarditis patients ($n = 17$). Using flow cytometry and a gating strategy with specific antibodies (Supplementary Table 1), we

were able to discriminate between living pDCs and mDC2 within PBMCs (Supplementary Figure 1). The number of pDCs decreased by 30% in patients with Q fever endocarditis as compared to controls (Figure 3A, up panel). In contrast, the percentage of mDC2 was higher in patients with Q fever endocarditis (Figure 3A, down panel), demonstrating that pDCs were specifically affected by *C. burnetii* infection. We also found that the phenotypic characteristics of pDCs were affected in Q fever endocarditis. Indeed, the expression of activation markers HLA-DR (Figure 3B, up panel) and PD-1 (Figure 3C, up panel) was increased in pDCs from patients with Q fever endocarditis. It should be noted that only PD-1 increased in mDC2 from the same patients (Figures 3B,C, down panels). Finally, we studied the spontaneous release of IFN- α by PBMCs from patients with Q fever endocarditis. We found that the release of IFN- α was markedly depressed in patients compared with healthy controls (Figure 3D, right panel), which is likely to be because the number of circulating pDCs was low. The release of IL-10 was increased in patients with Q fever endocarditis (Figure 3D, left panel), in accordance with previous results (Capo et al., 1996). Taken together, these results showed that the number of pDCs was depressed in patients with Q fever endocarditis and that the remaining pDCs exhibited specific features of activation.

DISCUSSION

In this study, we report, for the first time, the ability of *C. burnetii* to activate human pDCs *in vitro*. Indeed, they expressed CCR7 and CD86, migratory and activation markers, respectively. CCR7 plays a role in the migration of pDCs from blood to T cell areas of the lymph nodes and the white pulp of the spleen (Swiecki and Colonna, 2015). Using a high throughput microarray approach, we showed that *C. burnetii* modulated the expression of a large number of genes in which keywords including “inflammatory response,” “pathogen sensing,” and “regulation of cell proliferation” were enriched. We showed that *C. burnetii* stimulated the production of IFN- α associated with the release of TNF- α and IL-10, suggesting a modulation of inflammatory response to the pathogen.

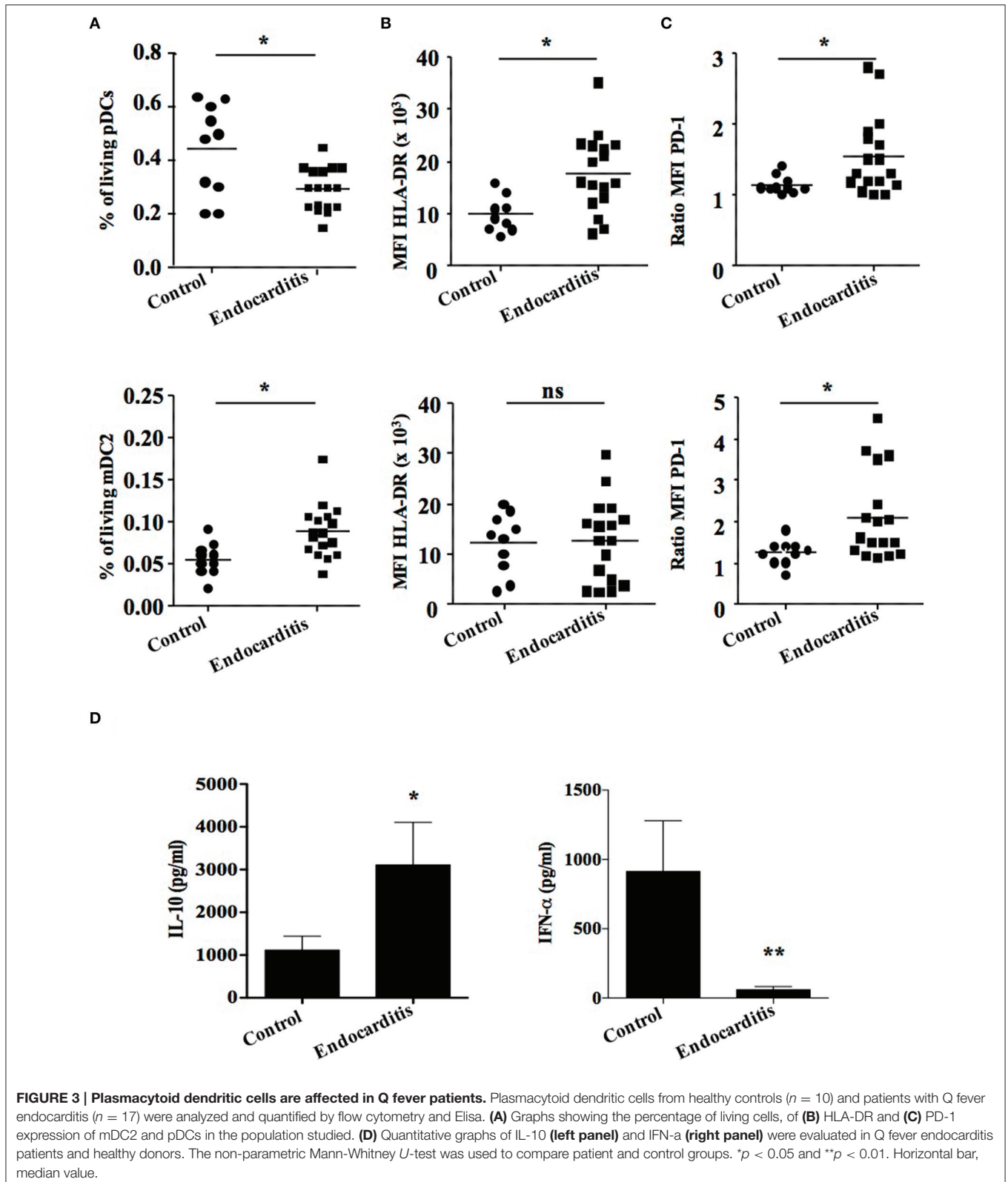
Interestingly, transcriptomic analysis revealed the up-regulation of genes that could be involved in the type I IFN pathway. First, genes enriched in the keyword “pathogen sensing,” such as *TLR7* and signaling molecules such as *MyD88* and *IRF7*, were up-regulated in *C. burnetii*-stimulated pDCs. The overexpression of *TLR7* and *TLR9* with the *IRF7* gene are necessary for type I IFN production by pDCs, suggesting that this pathway may play a role in pDCs’ response to *C. burnetii* (Kawai and Akira, 2010; Trinchieri, 2010). Secondly, the RNA helicase retinoic acid-inducible gene I (RIG-I) was up-regulated in *C. burnetii*-stimulated pDCs. RIG-I is a cytoplasmic receptor known to be responsible for triggering type I IFN secretion (Pandey et al., 2009; Kawai and Akira, 2010). Thirdly, *C. burnetii* up-regulated the expression of the genes encoding the nucleotide-binding oligomerization domain-containing protein (NOD)-1 and NOD2 receptors, which may be involved in the production of type I IFN (Pandey et al., 2009; Watanabe et al.,



2010). Indeed, *M. tuberculosis* induces type I IFN production through the activation of NOD2 in macrophages (Pandey et al., 2009) and *Helicobacter pylori* through NOD1 in epithelial cells (Watanabe et al., 2010). In addition, *Neisseria meningitidis*, *Hemophilus influenzae*, and *S. aureus* were found to stimulate the secretion of IFN- α and to increase the expression of CD86 on pDCs (Michea et al., 2013). The production of IFN- α by *C. burnetii*-stimulated pDCs associated with the activation phenotype is distinct from the response to *S. pyogenes* in which IFN- α is not produced (Veckman and Julkunen, 2008) and from the *M. tuberculosis* response in which the activation markers are lacking (Lozza et al., 2014).

As *C. burnetii* induced a specific response in pDCs *in vitro*, we questioned whether circulating pDCs are affected in Q fever. We showed that pDCs were decreased in patients with Q fever endocarditis. We studied patients with Q fever endocarditis and

not patients with chronic Q fever since this concept is debated among teams involved in Q fever investigation. In addition, preliminary results showed that patients with primary infection also exhibited decreased amounts of circulating pDCs (data not shown). To our knowledge, this is the first report demonstrating the modulation of the number of circulating pDCs in Q fever. We do not know whether the decrease in circulating pDCs reflects their migration to tissues nor the nature of their role in resistance or susceptibility to *C. burnetii*. The role of pDCs in the defense against bacteria is only supported by a report in which the depletion of pDCs increases the virulence of *C. pneumoniae* in mice (Crother et al., 2012). Plasmacytoid dendritic cells are protective in viral infections. The number of circulating pDCs is reduced in patients who are chronically infected with hepatitis B virus, hepatitis C virus and HIV (Goutagny et al., 2004; Ulsenheimer et al., 2005; Meyers et al., 2007). This decrease is



correlated with a high viral load (Soumelis et al., 2001; Gurney et al., 2004). Interestingly, patients treated with highly active anti-retroviral therapy are marked by a decreased viral load

and an increased number of circulating pDCs, suggesting this cell population is involved in the control of HIV infection (Pacanowski et al., 2001; Keir et al., 2002).

We also showed that circulating pDCs were likely to be activated in Q fever. Indeed, they over-expressed HLA-DR and PD-1. This pattern was not specific since it was also found in mDC2 from patients with Q fever endocarditis. The activated phenotype of pDCs in patients with Q fever may account for the increased traffic of pDCs from blood to peripheral tissues. The increased *in vitro* expression of a migratory marker such as CCR7 in response to *C. burnetii* may account for greater traffic to tissues. We recently showed that pDCs are present in the lymph nodes from Q fever lymphomas and are infected with *C. burnetii* (Melenotte et al., 2016). This suggests that *C. burnetii* infection may encourage the traffic of pDCs to lymph nodes. We also observed that patients with Q fever endocarditis exhibited a decrease in IFN- α production by PBMCs. We hypothesized that the decrease in IFN- α production is related to decrease levels of circulating pDCs. Whether IFN- α is involved in the pathophysiology of Q fever will require additional experiments.

In conclusion, we demonstrated that *C. burnetii* induced a strong inflammatory response in pDCs in which type I IFN was specifically enriched. The present study described

a new feature of the immune response in Q fever and also suggested the importance of pDCs in chronic bacterial infections.

AUTHOR CONTRIBUTIONS

Performed the experiments: MK, SM, and AB. Analyzed the data and wrote the paper: DR, JM, DO, EG, and AB.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <http://journal.frontiersin.org/article/10.3389/fcimb.2016.00070>

Supplementary Figure 1 | Gating strategy to study pDC population (A) Representative graphs showing the gating strategy of pDCs identification by flow cytometry.

Supplementary Table 1 | List of fluorescent reagents used in this study.

(AF, Alexa fluor; APC, Allophycocyanin; ECD, Phycoerythrin-texas red, FITC, Fluorescein isothiocyanate; PB, Pacific blue; PC, Phycoerythrin; PerCP-Cy5.5, Peridinin chlorophyll protein-cyanin 5.5; and PE, Phycoerythrin).

REFERENCES

- Amara, A. B., Bechah, Y., and Mege, J.-L. (2012). Immune response and *Coxiella burnetii* invasion. *Adv. Exp. Med. Biol.* 984, 287–298. doi: 10.1007/978-94-007-4315-1_15
- Bekeredjian-Ding, I., Greil, J., Ammann, S., and Parcina, M. (2014). Plasmacytoid dendritic cells: neglected regulators of the immune response to *Staphylococcus aureus*. *Front. Immunol.* 5:238. doi: 10.3389/fimmu.2014.00238
- Ben Amara, A., Ghigo, E., Le Priol, Y., Lépolard, C., Salcedo, S. P., Lemichez, E., et al. (2010). *Coxiella burnetii*, the agent of Q fever, replicates within trophoblasts and induces a unique transcriptional response. *PLoS ONE* 5:e15315. doi: 10.1371/journal.pone.0015315
- Ben Amara, A., Gorvel, L., Baulan, K., Derain-Court, J., Buffat, C., Vérolet, C., et al. (2013). Placental macrophages are impaired in chorioamnionitis, an infectious pathology of the placenta. *J. Immunol.* 191, 5501–5514. doi: 10.4049/jimmunol.1300988
- Bowie, A. G., and Unterholzner, L. (2008). Viral evasion and subversion of pattern-recognition receptor signalling. *Nat. Rev. Immunol.* 8, 911–922. doi: 10.1038/nri2436
- Capo, C., Zaffran, Y., Zugun, F., Houpiqian, P., Raoult, D., and Mege, J. L. (1996). Production of interleukin-10 and transforming growth factor beta by peripheral blood mononuclear cells in Q fever endocarditis. *Infect. Immun.* 64, 4143–4147.
- Cella, M., Jarrossay, D., Facchetti, F., Aleardi, O., Nakajima, H., Lanzavecchia, A., et al. (1999). Plasmacytoid monocytes migrate to inflamed lymph nodes and produce large amounts of type I interferon. *Nat. Med.* 5, 919–923. doi: 10.1038/11360
- Colonna, M., Krug, A., and Cella, M. (2002). Interferon-producing cells: on the front line in immune responses against pathogens. *Curr. Opin. Immunol.* 14, 373–379. doi: 10.1016/S0952-7915(02)00349-7
- Crother, T. R., Ma, J., Jupelli, M., Chiba, N., Chen, S., Slepkin, A., et al. (2012). Plasmacytoid dendritic cells play a role for effective innate immune responses during *Chlamydia pneumoniae* infection in mice. *PLoS ONE* 7:e48655. doi: 10.1371/journal.pone.0048655
- Dental, C., Florentin, J., Aouar, B., Gondois-Rey, F., Durantel, D., Baumert, T. F., et al. (2012). Hepatitis C virus fails to activate NF- κ B signaling in plasmacytoid dendritic cells. *J. Virol.* 86, 1090–1096. doi: 10.1128/JVI.05444-11
- Eberle, F., Sirin, M., Binder, M., and Dalpke, A. H. (2009). Bacterial RNA is recognized by different sets of immunoreceptors. *Eur. J. Immunol.* 39, 2537–2547. doi: 10.1002/eji.200838978
- Facchetti, F., Vermi, W., Mason, D., and Colonna, M. (2003). The plasmacytoid monocyte/interferon producing cells. *Virchows Arch. Int. J. Pathol.* 443, 703–717. doi: 10.1007/s00428-003-0918-8
- Fiorentini, S., Riboldi, E., Facchetti, F., Avolio, M., Fabbri, M., Tosti, G., et al. (2008). HIV-1 matrix protein p17 induces human plasmacytoid dendritic cells to acquire a migratory immature cell phenotype. *Proc. Natl. Acad. Sci. U.S.A.* 105, 3867–3872. doi: 10.1073/pnas.0800370105
- Gilliet, M., Cao, W., and Liu, Y.-J. (2008). Plasmacytoid dendritic cells: sensing nucleic acids in viral infection and autoimmune diseases. *Nat. Rev. Immunol.* 8, 594–606. doi: 10.1038/nri2358
- Gorvel, L., Textoris, J., Banchereau, R., Ben Amara, A., Tantibhedhyangkul, W., von Bargen, K., et al. (2014). Intracellular bacteria interfere with dendritic cell functions: role of the type I interferon pathway. *PLoS ONE* 9:e99420. doi: 10.1371/journal.pone.0099420
- Goutagny, N., Vieux, C., Decullier, E., Ligeois, B., Epstein, A., Trepo, C., et al. (2004). Quantification and functional analysis of plasmacytoid dendritic cells in patients with chronic hepatitis C virus infection. *J. Infect. Dis.* 189, 1646–1655. doi: 10.1086/383248
- Gurney, K. B., Colantonio, A. D., Blom, B., Spits, H., and Uittenbogaart, C. H. (2004). Endogenous IFN-alpha production by plasmacytoid dendritic cells exerts an antiviral effect on thymic HIV-1 infection. *J. Immunol.* 173, 7269–7276. doi: 10.4049/jimmunol.173.12.7269
- Honstetter, A., Imbert, G., Ghigo, E., Gouriet, F., Capo, C., Raoult, D., et al. (2003). Dysregulation of cytokines in acute Q fever: role of interleukin-10 and tumor necrosis factor in chronic evolution of Q fever. *J. Infect. Dis.* 187, 956–962. doi: 10.1086/368129
- Ka, M. B., Gondois-Rey, F., Capo, C., Textoris, J., Million, M., Raoult, D., et al. (2014). Imbalance of circulating monocyte subsets and PD-1 dysregulation in Q fever endocarditis: the role of IL-10 in PD-1 modulation. *PLoS ONE* 9:e107533. doi: 10.1371/journal.pone.0107533
- Ka, M. B., Gondois-Rey, F., Ghigo, E., Raoult, D., Olive, D., and Mege, J.-L. (2015). Imbalance of circulating lymphoid cells in Q fever endocarditis. *Pathog. Dis.* 73, 1–3. doi: 10.1093/femspd/ftu010
- Kawai, T., and Akira, S. (2010). The role of pattern-recognition receptors in innate immunity: update on Toll-like receptors. *Nat. Immunol.* 11, 373–384. doi: 10.1038/ni.1863
- Keir, M. E., Stoddart, C. A., Linquist-Stepps, V., Moreno, M. E., and McCune, J. M. (2002). IFN-alpha secretion by type 2 predendritic cells up-regulates MHC class I in the HIV-1-infected thymus. *J. Immunol.* 168, 325–331. doi: 10.4049/jimmunol.168.1.325

- Liu, Y.-J. (2005). IPC: professional type 1 interferon-producing cells and plasmacytoid dendritic cell precursors. *Annu. Rev. Immunol.* 23, 275–306. doi: 10.1146/annurev.immunol.23.021704.115633
- Lozza, L., Farinacci, M., Faé, K., Bechtle, M., Stäber, M., Dorhoi, A., et al. (2014). Crosstalk between human DC subsets promotes antibacterial activity and CD8+ T-cell stimulation in response to bacille Calmette-Guérin. *Eur. J. Immunol.* 44, 80–92. doi: 10.1002/eji.201343797
- Martinelli, E., Cicala, C., Van Ryk, D., Goode, D. J., Macleod, K., Arthos, J., et al. (2007). HIV-1 gp120 inhibits TLR9-mediated activation and IFN- α secretion in plasmacytoid dendritic cells. *Proc. Natl. Acad. Sci. U.S.A.* 104, 3396–3401. doi: 10.1073/pnas.0611353104
- Melenotte, C., Million, M., Audoly, G., Gorse, A., Dutronc, H., Roland, G., et al. (2016). B-cell non-Hodgkin lymphoma linked to *Coxiella burnetii*. *Blood* 127, 113–121. doi: 10.1182/blood-2015-04-639617
- Mengshol, J. A., Golden-Mason, L., Castelblanco, N., Im, K. A., Dillon, S. M., Wilson, C. C., et al. (2009). Impaired plasmacytoid dendritic cell maturation and differential chemotaxis in chronic hepatitis C virus: associations with antiviral treatment outcomes. *Gut* 58, 964–973. doi: 10.1136/gut.2008.168948
- Meyers, J. H., Justement, J. S., Hallahan, C. W., Blair, E. T., Sun, Y. A., O'Shea, M. A., et al. (2007). Impact of HIV on cell survival and antiviral activity of plasmacytoid dendritic cells. *PLoS ONE* 2:e458. doi: 10.1371/journal.pone.0000458
- Michea, P., Vargas, P., Donnadieu, M.-H., Roseblatt, M., Bono, M. R., Duménil, G., et al. (2013). Epithelial control of the human pDC response to extracellular bacteria. *Eur. J. Immunol.* 43, 1264–1273. doi: 10.1002/eji.201242990
- Pacanowski, J., Kahi, S., Baillet, M., Lebon, P., Deveau, C., Goujard, C., et al. (2001). Reduced blood CD123+ (lymphoid) and CD11c+ (myeloid) dendritic cell numbers in primary HIV-1 infection. *Blood* 98, 3016–3021. doi: 10.1182/blood.V98.10.3016
- Pandey, A. K., Yang, Y., Jiang, Z., Fortune, S. M., Coulombe, F., Behr, M. A., et al. (2009). NOD2, RIP2 and IRF5 play a critical role in the type I interferon response to *Mycobacterium tuberculosis*. *PLoS Pathog.* 5:e1000500. doi: 10.1371/journal.ppat.1000500
- Raoult, D. (2012). Chronic Q fever: expert opinion versus literature analysis and consensus. *J. Infect.* 65, 102–108. doi: 10.1016/j.jinf.2012.04.006
- Raoult, D., Marrie, T., and Mege, J. (2005). Natural history and pathophysiology of Q fever. *Lancet Infect. Dis.* 5, 219–226. doi: 10.1016/S1473-3099(05)70052-9
- Sachdeva, N., Asthana, V., Brewer, T. H., Garcia, D., and Asthana, D. (2008). Impaired restoration of plasmacytoid dendritic cells in HIV-1-infected patients with poor CD4 T cell reconstitution is associated with decrease in capacity to produce IFN- α but not proinflammatory cytokines. *J. Immunol.* 181, 2887–2897. doi: 10.4049/jimmunol.181.4.2887
- Schoffelen, T., Joosten, L. A. B., Herremans, T., de Haan, A. F. J., Ammerdorffer, A., Rümke, H. C., et al. (2013). Specific interferon γ detection for the diagnosis of previous Q fever. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* 56, 1742–1751. doi: 10.1093/cid/cit129
- Shannon, J. G., Howe, D., and Heinzen, R. A. (2005). Virulent *Coxiella burnetii* does not activate human dendritic cells: role of lipopolysaccharide as a shielding molecule. *Proc. Natl. Acad. Sci. U.S.A.* 102, 8722–8727. doi: 10.1073/pnas.0501863102
- Siegal, F. P., Kadowaki, N., Shodell, M., Fitzgerald-Bocarsly, P. A., Shah, K., Ho, S., et al. (1999). The nature of the principal type 1 interferon-producing cells in human blood. *Science* 284, 1835–1837. doi: 10.1126/science.284.5421.1835
- Soumelis, V., Scott, I., Gheyas, F., Bouhour, D., Cozon, G., Cotte, L., et al. (2001). Depletion of circulating natural type 1 interferon-producing cells in HIV-infected AIDS patients. *Blood* 98, 906–912. doi: 10.1182/blood.V98.4.906
- Swiecki, M., and Colonna, M. (2015). The multifaceted biology of plasmacytoid dendritic cells. *Nat. Rev. Immunol.* 15, 471–485. doi: 10.1038/nri3865
- Trinchieri, G. (2010). Type I interferon: friend or foe? *J. Exp. Med.* 207, 2053–2063. doi: 10.1084/jem.20101664
- Ulsenheimer, A., Gerlach, J. T., Jung, M.-C., Gruener, N., Wächter, M., Backmund, M., et al. (2005). Plasmacytoid dendritic cells in acute and chronic hepatitis C virus infection. *Hepatology* 41, 643–651. doi: 10.1002/hep.20592
- Veckman, V., and Julkunen, I. (2008). *Streptococcus pyogenes* activates human plasmacytoid and myeloid dendritic cells. *J. Leukoc. Biol.* 83, 296–304. doi: 10.1189/jlb.0707457
- Watanabe, T., Asano, N., Fichtner-Feigl, S., Gorelick, P. L., Tsuji, Y., Matsumoto, Y., et al. (2010). NOD1 contributes to mouse host defense against *Helicobacter pylori* via induction of type I IFN and activation of the ISGF3 signaling pathway. *J. Clin. Invest.* 120, 1645–1662. doi: 10.1172/JCI39481
- Xie, Q., Shen, H.-C., Jia, N.-N., Wang, H., Lin, L.-Y., An, B.-Y., et al. (2009). Patients with chronic hepatitis B infection display deficiency of plasmacytoid dendritic cells with reduced expression of TLR9. *Microbes Infect. Inst. Pasteur* 11, 515–523. doi: 10.1016/j.micinf.2009.02.008

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2016 Ka, Mezouar, Ben Amara, Raoult, Ghigo, Olive and Mege. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.