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## **A set of PCRs for rapid identification and characterization of *Pseudomonas syringae* phylogroups**

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### **Abbreviated running headline**

PCR for *Pseudomonas syringae* phylogroups

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### **Abstract**

**Aims:** The aim of this study was to develop a rapid PCR-based method for the specific detection of individual phylogroups of the *Pseudomonas syringae* complex.

**Methods and Results:** Seven primer pairs were developed by analyzing whole genomes of 54 *P. syringae* strains. The specificity and sensitivity of these primer pairs were assessed on 236 strains from a large and comprehensive *P. syringae* collection. The method was also validated by

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characterizing the phylogenetic diversity of 174 putative *P. syringae* isolates from kiwifruit and apricot orchards of southeastern France.

**Conclusion:** Our PCR-based method allows for the detection and characterization of 9 of the 13 *P. syringae* phylogroups (phylogroups 1, 2, 3, 4, 7, 8, 9, 10 and 13).

**Significance and Impact of the Study:** To date, phylogenetic affiliation within the *P. syringae* complex was only possible by sequencing housekeeping genes. Here we propose a rapid PCR-based method for the detection of specific phylogroups of the *P. syringae* complex. Furthermore, for the first time we reveal the presence of *P. syringae* strains belonging to phylogroups 10 and 13 as epiphytes on plants whereas they had previously only been observed in aquatic habitats.

## Keywords

Plant pathogen, microbial ecology, pathobiome, multiplex PCR, orchards

## Introduction

In plant pathology, competitive or synergistic interactions among microorganisms have been disregarded in disease etiology. The emerging pathobiome concept pointed out that etiological agents can work in concert with indigenous microflora to induce disease (Vayssier-Taussat et al. 2014). The impact on disease of infra-specific diversity of microorganisms in individual hosts has been ignored. In the case of plant pathogenic bacteria, analyses of the structure of the pathogenic populations generally concern the metapopulation/population at large in a region (Scortichini 2005), but rarely addresses the structure of pathogen populations in individual plants. Phenomena of gain of fitness, as well as synergistic interactions, are not uncommon within strains of different genetic lines belonging to the same bacterial species and could enhance diseases (Kamoun and Kado 1990; Hirano et al. 1999; Scortichini 2005; Lamichhane and Venturi 2015). On the other hand, competition between strains of the same ecological niche could attenuate disease symptoms (Kerr and Htay 1974; Moss et al. 2007; Kawaguchi et al. 2014). In spite of these observations, conventional diagnosis of plant diseases has

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focused on the characterization of the dominant organisms in infected tissue. To address the issue of microbial complexes responsible for disease occurrence, detection methods that facilitate the assessment of the population structure in individual plants are needed. In addition, metagenomics approaches are in part unable to characterize the structure of different microbial communities at species level (Handelsman 2004; Walker et al. 2014; Barret et al. 2015). In light of these facts, we have developed a set of PCRs to rapidly identify the phylogenetic structure of the plant pathogenic bacterium *Pseudomonas syringae* (Berge et al. 2014). This method requires strain isolation, thereby allowing subsequent evaluation of the pathogenicity of the isolated strains.

*P. syringae* is a common environmental bacterium that is ubiquitous in both aquatic and agricultural habitats where it can have both saprophytic and pathogenic behavior. *P. syringae* is also an important plant pathogen causing diseases such as kiwifruit canker reported in all kiwi-producing regions: Australasia, Europe and South America (Hirano and Upper 1990; Gaignard and Luisetti 1993; Vanneste 2013; Donati et al. 2014). Since the beginning of the 20<sup>th</sup> century, 55 cases of bacterial diseases in 25 countries and on 25 species of trees of agronomic importance were declared as being caused by this bacterium (Lamichhane et al. 2014). Likewise, in this same period *P. syringae* has been identified as the cause of nearly 70 new disease outbreaks affecting over 40 annual crop species (Lamichhane et al. 2015).

*P. syringae* is now recognized as a complex of at least 13 phylogroups (Berge et al. 2014). Each phylogroup displays unique phenotypes and it can be distinguished according to the sequence of one (for example citrate synthase) or a few (Multilocus Sequence Typing) housekeeping genes (Berge et al. 2014). Studies on the non-agricultural habitats (water, litter, wild plants, etc) showed that multiple *P. syringae* phylogroups can co-exist in the same sample (Morris et al. 2007; Monteil et al. 2013). However, there is a paucity of information about such co-existence in crops and about the possible role of co-existing strains in disease occurrence. In light of the current well-defined *P. syringae* classification and the numerous whole genome sequences that are available (Berge et al. 2014), we have sought to develop PCR primers that target as many of the phylogroups as possible to facilitate analysis of the *P. syringae* populations associated with diseased plants as well as with a range of environmental substrates. These molecular markers are more encompassing than those that have been

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developed to detect genetic lines that have pathovar names such as *P. syringae* pv. *phaseolicola* (Mosqueda-Cano and Herrera-Estrella 1997), *P. syringae* pv. *tomato* (Zaccardelli et al. 2005), *P. syringae* pv. *aesculi* (Green et al. 2009) and *P. syringae* pv. *actinidiae* (Sawada et al. 1997; Sawada et al. 1999; Koh and Nou 2002; Rees-George et al. 2010; Gallelli et al. 2011; Balestra et al. 2013; Gallelli et al. 2014; Loreti et al. 2014). Pathovar-specific markers could nevertheless be used in conjunction with phylogroup markers and also with the "universal" marker of all members of the *P. syringae* complex (Guilbaud et al., in press). This latter primer pair, called "Psy", allows for the identification of strains in the *P. syringae* complex with over 97% accuracy. This allows a real gain in time when identifying strains of *P. syringae*. However

, this latter method does not identify the phylogroup affiliation of *P. syringae*.

Here, we propose a set of molecular markers identifying each of 9 major *P. syringae* phylogroups based on the current phylogenetic classification and the genomes of *P. syringae* accessible in public databases. We also describe a method to deploy these markers in a multiplex-PCR with the Psy primer set as a universal control for reliable detection and identification of *P. syringae* phylogroups. An example of the use of this method to characterize communities of *P. syringae* associated with kiwifruit and apricot plant samples from southeastern France is also reported.

## Materials and Methods

### Strains and criteria used for primer design

During this study 439 strains were used for 3 different objectives: i) to develop the primer pairs, ii) to validate the designed primers (specificity/sensitivity), and iii) to perform a "blind test" for the multiplex-PCR method on putative *P. syringae* isolates obtained from kiwifruit and apricot orchards from southeastern France. For primer development, DNA sequences were identified in whole genomes of 54 *P. syringae* strains (Table S1) available on the Joint Genome Institute (JGI) platform

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(<http://jgi.doe.gov/>). The distribution of the corresponding strains within the various phylogroups is uneven and reflects the availability of the sequences.

To evaluate the specificity and sensitivity of the primer pairs, 236 strains whose phylogroups were previously determined by citrate synthase (*cts*) gene sequencing, were used (Table S1). These 236 strains are part of the *P. syringae* collection of the Plant Pathology research Unit of the INRA PACA Avignon center. This collection is composed by over 7000 *P. syringae* strains isolated from crops and non-agricultural habitats.

Strains were maintained at -80°C in phosphate buffer containing 20% glycerol and were cultured on King's B medium (King et al. 1954) and incubated at 25°C for 48 hours prior each analysis. The strains used for primer development and for the specificity and sensitivity tests were pure culture strains.

Putative *P. syringae* isolates, not pure culture strains, were used for the “blind test” described below. These putative *P. syringae* isolates came from field sampling aiming to characterize *P. syringae* populations associated with kiwifruit and apricot orchards in southeastern France (Drôme county). Overall 174 putative *P. syringae* isolates were used. These isolates were collected from different substrates including leaves, buds, twigs, ground-cover, soil and irrigation waters (Table S1). For bacterial isolation, leaf, bud, twig and ground cover samples were homogenized with a stomacher in a phosphate buffer solution for 4 minutes. Soil samples were directly suspended with sterilized distilled water. Bacterial isolation from water samples was carried out according to the method previously described (Morris et al. 2010). For all the obtained suspensions, we performed dilution-plateing on a modified King's B medium (KBC) (Mohan and Schaad 1987). KBC as used in this study was composed of 9 volumes of King's B medium with a smaller quantity of proteose peptone than the original recipe (1.5g K<sub>2</sub>HPO<sub>4</sub>, 1.5g MgSO<sub>4</sub> 7H<sub>2</sub>O, 15g proteose-peptone, 10 ml glycerol and 16g agar in 900 ml distilled water), and 1 volume of boric acid (1.5g H<sub>3</sub>BO<sub>3</sub> in 100 ml distilled water), and contains cycloheximide (200 mg l<sup>-1</sup>) and cephalixin (80 mg l<sup>-1</sup>). The plates were incubated at 25°C for 48-72 hours. Putative *P. syringae* isolates were identified by the morphology of the colony (size, color, reflectance of light and roughness of the colony border). These isolates were stored in a phosphate buffer. No previous data about their phylogroup affiliation were available for these isolates.

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Phylogroup affiliation of these isolates was verified by sequencing the *cts* gene of the corresponding purified strain of each isolate. The resulting *cts* sequences were aligned with the *cts* sequences of the strains from the *P. syringae* collection described above (1 or 2 representatives per phylogroup) by using clustalW2 software. A phylogenetic tree was constructed with the Neighbor Joining Model and 3000 bootstrap replicates by using MEGA6 software (Tamura et al. 2013).

Fifty-four *P. syringae* whole genomes available on the JGI platform and representative of 10 phylogroups were used to identify genes specific for each phylogroup with the IMG/M system (<http://img.jgi.doe.gov>). The Abundance Profile Overview tool was used in order to identify unique genes specific for each phylogroup. This tool provides an overview of the relative abundance of protein families (COGs and Pfams) across the 54 selected genomes used in this study. Genes selected for primer designing are listed in Table 1. All gene sequences used in this study are available in Table S2. For Phylogroups 1, 3 and 10 no unique open reading frames or unique DNA sequences were observed. For these phylogroups, DNA regions of some genes (Table 1) were used to design primers based on single nucleotide polymorphisms (SNPs). The SNP containing region was chosen based on a high G/C content. The size of the SNPs on the forward (Fw) and reverse (Rv) primers varied from 4 to 17 bp as reported in Table 1.

### PCR conditions and analysis of the amplicons for detection of *P. syringae* phylogroups

All primer pairs were tested individually. After verification of the specificity and the size of the amplicons, primer pairs were combined in multiplex PCRs in order to reduce the number of PCRs required to identify the phylogroups. Reaction mixes for the individual PCR were composed of 5.0  $\mu\text{l}$  of GoTaq green 5X PCR buffer (Promega, France), 1.5  $\mu\text{l}$  of a solution of 25  $\text{mmol l}^{-1}$   $\text{MgCl}_2$ , 0.2  $\mu\text{l}$  of a mix of 25  $\text{mmol l}^{-1}$  dNTPs, 2  $\mu\text{l}$  of a mix of primers consisting of 10  $\text{mmol l}^{-1}$  Fw and Rv primers, 0.13  $\mu\text{l}$  of GoTaq G2 DNA polymerase (Promega, France) with 5 units  $\mu\text{l}^{-1}$ , and 2.0  $\mu\text{l}$  of bacterial suspension at  $10^8$  cells  $\text{ml}^{-1}$ . The final volume of the PCR reaction was 25  $\mu\text{l}$  and milliQ water was

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used to mix dilution. All PCR amplifications were performed for 30 cycles; primer sequences and PCR conditions are listed in Table 2. Reaction mixes for the multiplex PCRs were composed of 6.25  $\mu\text{l}$  of 2X QIAGEN Multiplex PCR Master Mix (Qiagen® Multiplex PCR kit), 1.25  $\mu\text{l}$  of primer mix with a final concentration of 2  $\text{mmol l}^{-1}$  per primer, 1.25  $\mu\text{l}$  of Q solution, 2.75  $\mu\text{l}$  of milliQ water and 2.0  $\mu\text{l}$  of bacterial suspension at  $10^8$  cells  $\text{ml}^{-1}$ . The final volume reaction for the multiplex PCR was 11.5  $\mu\text{l}$ . Taking into account the amplicon sizes (Table 2), two multiplex PCR reactions were developed: "Multiplex 1" containing the universal primer pair Psy and the primers specific for phylogroup 1 (PG1), phylogroup 3 (PG3) and phylogroup 9 (PG9) and "Multiplex 2" containing the primer pair Psy and those specific for phylogroup 2 (PG2), phylogroup 4 (PG4) and phylogroup 13 (PG13). Amplification was performed for 30 cycles and denaturation and elongation temperatures were 95°C and 72°C, respectively for all reactions. Annealing temperatures are reported in Table 2. PCR products were migrated at 80V in a 1.5% agarose gel containing ethidium bromide (BET) (Euromedex, EU0070) at a concentration of 0.5  $\mu\text{g ml}^{-1}$ . Migration times varied depending on the size of PCR amplicons. After migration, PCR products were visualized under ultraviolet light (320 nm).

### Sensitivity and performance criteria

To test the sensitivity of the multiplex PCR, different bacterial concentrations were used during PCR amplifications. These concentrations varied from  $10^8$  to  $10^4$  bacterial cells  $\text{ml}^{-1}$ .

To test the performance of the PCRs we used criteria described in previous studies (EPPO 2010; 2014; Loreti et al. 2014). These performance criteria are derived from those used in the development of a PCR based detection method for *P. syringae* pv. *actinidiae* (Loreti et al. 2014). Briefly, these criteria include i) relative accuracy (A) which corresponds to the similarity of the observed results compared to expected results, ii) diagnostic sensitivity (SE) which is the ability of the method to detect the target strain in a contaminated reference sample, and iii) diagnostic specificity (SP) which corresponds to the capacity of the method to not yield a positive result in an uncontaminated sample reference.

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## Results

Eight primer pairs are sufficient to identify nine *P. syringae* phylogroups

In this work, we developed primer pairs for the identification of 9 *P. syringae* phylogroups (1, 2, 3, 4, 9, 10 and 13). A primer pair for the detection of the *P. syringae* phylogroups 7 and 8 was developed and evaluated previously (Bartoli et al, 2014).

To detect PG1, a primer pair was developed based on the inter-phylogroup variability found in the *gyrase B* (*gyrB*) housekeeping gene. The primer pair and the SNPs used to discriminate the PG1 strains among all the rest of the *P. syringae* phylogroups are reported in Table 1 and 2. Except for this primer pair all other primers were designed on non - housekeeping genes. The primer pair for PG1 (Table 2) was designed in the 387-754 bp region of the *gyrB* gene and amplify a fragment of 321 bp. The corresponding SNP localizations are reported in Table 1.

For PG2, a unique gene candidate (DUF421) was found in the 16 genomes used during this study. This gene codes for a domain with unknown function, according to the database of conserved protein domain families (Pfam04239). The PG2 primer pair (Table 2) was designed on the 21-404 bp region of the gene, and amplifies a fragment of 341 bp.

For PG3, a gene coding for a DUF1835 domain was found in all the available PG3 genomes and in some strains belonging to the phylogroups 1, 7, 9 and 10. This gene codes for a protein of unknown function and its domain is classed under the number pfam08874. Gene sequences for these four *P. syringae* phylogroups displayed a high degree of polymorphism. This polymorphism was used to design a specific primer pair that amplifies only PG3. The localizations of the SNPs used to design the specific PG3 primers are reported in Table 1. PG3 specific primer pair (Table 2) was developed on the 61-937 bp region of the gene and amplifies a fragment of 837 bp.

For PG4 one unique gene was found, coding for a lysine-tRNA ligase (pfam01921). The domain of this protein was only found in this *P. syringae* phylogroup. The sequences of the primer pair (Table 2) developed on the 605-1489 bp region of the gene amplify a fragment of 845 bp.

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For PG9 a gene involved in bacterial chemotaxis (pfam04509) was found to be specific for all the strains in this phylogroup. The primer pair (Table 2) for this *P. syringae* phylogroup was developed on the 41-469 bp region of the gene and amplifies a fragment of 389 bp.

For PG10 strains we did not identify a specific candidate gene. Therefore, we selected a gene coding for a DUF2599 domain (pfam10783) present in the genome of the 3 genomes available for the PG10 as well in one strain of the *P. syringae* PG1. The PG10 specific primer pair (Table 2) was developed on the 435-799 bp region of the gene and amplifies a fragment of 324 bp. Localizations of the SNPs that were used to discriminate between the two phylogroups (PG10 and PG1) are reported in Table 1.

For PG13 several specific genes were found in the one available genome of a strain belonging to this phylogroup. The most accurate and specific primer pair targeted a gene encoding an alpha-glucosidase (pfam01055). The primer pair (Table 2) developed on the 599-1220 bp region of the gene amplifies a fragment of 582 bp.

The primer pairs for detection of the nine *P. syringae* phylogroups are specific with a high level of sensitivity.

Results for the performance criteria demonstrated that the primer pairs we developed here for the detection of the nine *P. syringae* phylogroups are highly specific and sensitive (Table 3). In fact, we reported that SP is higher than 99% for all the primer pairs, indicating a high specificity level. Only one false positive PCR result was reported with the PG2 and PG13 primer pairs. All PCR results are available in Table S1. A 100% SE was obtained for the phylogroups 2, 3, 9 and 13 primer pairs, indicating a good diagnostic sensitivity. Only 1 false negative PCR result was observed for the PG1 and PG4 primer pairs. The lowest diagnostic sensitivity was observed for the PG10 primer pair, only 8 of the 18 PG10 strains were amplified.

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Concerning sensitivity, i.e. the amount of bacterial cells  $\text{ml}^{-1}$  needed for positive PCR detection, clear bands were obtained in single and multiplex PCR on suspensions with at least  $10^6$  bacterial cells  $\text{ml}^{-1}$ , which corresponds to  $2 \cdot 10^3$  cells in the  $2\mu\text{l}$  used for PCR reaction. Weak bands were obtained for bacterial suspensions at  $10^5$  bacterial cells  $\text{ml}^{-1}$ .

A blind test on putative *P. syringae* isolates from kiwifruit and apricot orchards showed the efficiency of the primer pairs in identifying PGs in natural samples

Phylogroup affiliation of 174 putative *P. syringae* isolates from kiwifruit and apricot orchards was determined in parallel by *cts* sequencing and with the PCR method. Results based on the primer pair detection method showed the following phylogroup affiliation: i) 31% of the isolates were in *P. syringae* PG1, ii) 14% of the isolates clustered with strains of PG2, iii) 8% of the isolates were located in PG7 and 8, iv) 29% and 7% of the isolates clustered in PG13 and with different bacterial species respectively (Figure 1a). Furthermore for 5% of the isolates the phylogroup affiliation could not be determined (Ni) and in 6% of cases a multiple phylogroup identification (Mi) occurred (identification of 2 or more phylogroups for a same isolate) (Figure 1a). Through *cts* sequencing similar results were observed: i) 34% of the isolates were in the *P. syringae* PG1, ii) 14% of the isolates clustered with strains of PG2, iii) 1% of the isolates were in PG3, iv) 7% and 1% of the isolates were in PG7 and 8 respectively, v) 2% of the isolates were in PG9, vi) 4% of the isolates were in PG 10, vii) 29% and 7% of the isolates clustered with PG13 and with different bacterial species respectively, and viii) 1% of the isolates remain without phylogroup identification (Figure 1b). For the isolates whose results did not match between the primer pair detection method and the *cts* sequencing, the *cts* sequences revealed that 4%, 2%, 3% and 2% of the strains for PG1, PG2, PG7 and PG13 respectively, were misidentified. None of the 7 isolates of PG10 were detected by the PCR method, they were misidentified as belonging to PG7-8 or to PG1. PCR characterization of 10 isolates revealed that they contained mixtures of 2 or more phylogroups. Purification led to the isolation of the predicted components of the mixture. Among the 9 isolates that could not be identified with PCR, based on the

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*cts* sequence of the purified strains six represented PG1, two represented PG9 and one could not be identified with the current phylogenetic classification. Twelve isolates did not belong to the species *P. syringae* according to the PCR results, and this was confirmed by sequencing the *cts* gene.

## Discussion

The PCR markers developed in this study allow rapid classification and identification of the infra-specific diversity for at least 8 *P. syringae* phylogroups (PG1, 2, 3, 4, 7, 8, 9 and 13) with a high level of accuracy and for some of the strains in PG10. By coupling phylogroup-specific primer pairs to the *P. syringae* species-specific primer pair (Guilbaud et al. in press) and to a sensitive selective medium (Mohan and Schaad 1987), the structure of *P. syringae* populations associated with plants and other substrates can be quantified. Quantification can be easily performed by counting the number of colonies present on the *P. syringae* selective medium. From the bacterial isolates obtained by plating, classification of the strains into the corresponding *P. syringae* phylogroups is then possible by using the primer sets proposed here.

Identifying phylogroups, with a sequencing-free molecular technique, will lead to more efficient sequencing that targets only strains for which phylogroup affiliation is not possible. Targeted sequencing could therefore be focused on the more rare phylogroups such as the phylogroup 5, 6, 11 and 12 as well as any currently unknown phylogroups that occurred in the samples. Understanding phylogroup affiliation of isolates is an important step for the identification of *P. syringae* lines implicated in a range of diseases. The method proposed here could be then employed for epidemiological studies aiming at characterizing *P. syringae* diversity associated with diseased plants. To date, the kiwifruit epidemic is known to be caused by strains of the *P. syringae* complex that belong to PG1 (Bartoli et al. 2015). By using the primers developed here, one can attempt to detect strains in the *P. syringae* PG1 associated with disease on kiwifruit plants and then to confirm with more specific primers their identity with the current quasi-clonal epidemic line. In this way association of other *P. syringae* phylogroups as an important factor for disease occurrence on kiwifruit could be revealed. For example, here we revealed the presence of strains belonging to PG10

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and PG13 as epiphytes on ground cover plants and in buds of apricot and kiwifruit orchards. Previously these phylogroups were only observed in water and other non-plant substrates associated with the water cycle (Berge et al. 2014). The potential synergistic or antagonistic roles of these phylogroups in symptom manifestation is unknown and can now be explored.

Furthermore the molecular technique described here leads to considerable time savings in light of the effort required for strain purification and preparation of DNA for sequencing. In fact, when the method is applied to isolates that have not been purified it is possible to identify mixtures of several phylogroups. However, it is not possible to determine the number of isolates of each phylogroup constituting these mixtures nor to identify mixtures of several strains of the same phylogroup.

The cases of misallocation of the phylogroups by PCR can be explained by the extreme genetic variability that exists within the *P. syringae* complex (Berge et al. 2014). Indeed some strains can lose or gain genes as a result of mutation, deletion or insertion. Developing primers to identify species of the same genus or genera sharing the same functions can sometimes be difficult, and differentiating strains within a bacterial species is often even more difficult (Romero et al. 1995; Wang et al. 1997; Hallin and Lindgren 1999; Kidd et al. 2009; Rees-George et al. 2010; Gallelli et al. 2014). The primer pairs described here should be used in two separate PCR protocols: two multiplexes - one to identify PG1, 3 and 9 and the second for identification of PG2, 4 and 13 - and two independent single PCRs. The first single PCRs will allow for the identification of strains in PG 7 and 8 with the specific primer pair (shcF) previously described (Bartoli et al. 2014). The primer pair shcF could not be incorporated into the multiplex due to incompatibility of the annealing temperatures. The second single PCR will allow the identification of strains in PG10. It was not possible to couple this last primer pair because the size of the amplicon, 324 bp, is similar to amplicons obtained for PG1 (321bp) and PG2 (341 bp). The method described here will facilitate exploration of population structures of *P. syringae* assemblages and in particular in samples where different genetic lines co-exist and potentially interact. It will also contribute to the discovery and characterization of unknown *P. syringae* phylogroups in natural samples. With the increase in the number of available *P. syringae* genomes, improving molecular markers for *P. syringae* phylogroup detection is facilitated. The approach

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proposed here will open new perspectives on the importance of assemblages of genotypes of this bacterium in its survival, fitness and pathogenicity.

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### Conflict of Interest

The authors declare no conflict of interest.

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**Table 1.** Genes used for primer development

Strain	Genome Accession Number	Gene ID	Genomic Range (bp) *	Gene length (bp)	Protein Annotation	Presence in phylogroups †	Target Phylogroups ‡	Forward primer localization (bp)	Reverse primer localization (bp)	SNP localisation (bp), Fw primer §	SNP localisation (bp), Rv primer §
NCPPB 1108	GCA_000177495.1	648421386	1935-4352	2418	DNA gyrase/topoisomerase IV, subunit B	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13	1	387-410	732-754	387, 390, 391, 393, 394, 397, 399, 400, 405, 408	732, 733, 735, 738, 741, 744, 747, 750, 751, 753
CC1458	GCA_000452805.2	2506895564	4542-4985	444	Protein of unknown function. DUF421 domain	2	2	21-43	385-404	na	na
NCPPB 3335	GCA_000164015.2	648877554	38103-39338	1236	Protein of unknown function. DUF1835 domain	1, 3, 7, 9, 10	3	61-80	918-937	66, 69, 72, 73, 74, 75, 78, 79	918, 919, 920, 921, 922, 923, 924, 926, 927, 928, 929, 930, 931, 933, 934, 935, 936
CC1513	GCA_000452765.2	2506907413	8421-11348	2928	Lysyl-tRNA synthetase, class I	4	4	605-624	1470-1489	na	na
CC1417	GCA_000452825.2	2506965122	7504-8115	612	Chemotaxis like protein	9	9	41-60	450-469	na	na
CC1466	GCA_000452785.2	2506901715	17733-18980	1248	Protein of unknown function. DUF2599 domain	1, 10	10	435-458	780-799	435, 443, 447, 448, 456, 458	780, 786, 795, 798
UB246	GCA_000452865.1	2507270264	7514-9898	2385	Alpha-glucosidase, glycosyl hydrolase family GH31	13	13	599-618	1201-1220	na	na

\* Range of the gene that was used to design the primer pairs.

† Phylogroup in which the gene was present as single copy.

‡ The given gene was used to develop specific primers for the phylogroups listed in the table.

§ SNP localization refers to those genes that were present in more than one phylogroup.

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**Table 2.** Primers and PCR conditions used for identification of *Pseudomonas syringae* phylogroups

<b>Simplex PCR</b>			
<b>Primer Pairs</b>	<b>Annealing temperatures (°C)</b>	<b>Expected size (bp)</b>	<b>References</b>
<b>Psy generic <i>P. syringae</i> primers</b>			
Fcb43 ATC ATG GGA GAC GCG CTT AAG			
Rcb43 GCT GCA GAG AGC ACT	61	<b>144</b>	Guilbaud et al. in press
<b>PG1</b>			
PG1-Fw AGA ACT TCT GTT GCT GAC AGT ACG			
PG1-Rv CAA TAC CGT CAT CGC GCT GGA CA	63	<b>321</b>	This study
<b>PG2</b>			
PG2-Fw AGC AGC GAT TTA TTT TGT CTT GA			
PG2-Rv ATG ATC GCG TAC CTG ATG TC	60	<b>341</b>	This study
<b>PG3</b>			
PG3-Fw AAG CGT GCC AAA GAG CTT CT			
PG3-Rv CTT GGC CGA AGG TCA TTG AC	61	<b>837</b>	This study
<b>PG4</b>			
PG4-Fw CAC CTG ACT TAC TGA TCA AT			
PG4-Rv ACA GGC ACG CAT ATA TCG TT	60	<b>845</b>	This study
<b>PG7 and 8 (shcF)</b>			
shcF-Fw CGA CCT GCT TTC GAT CA			
shcF-Rv TCA ATA CTC TGG AGA TCA G	54	<b>850</b>	Bartoli et al. 2014
<b>PG9</b>			
PG9-Fw CAT TGC AGG AAC TGA TGA AC			
PG9-Rv GGT TCA ACT CTT CGA GAC AA	58	<b>389</b>	This study
<b>PG10</b>			
PG10-Fw CAC GAC GAC CTA TAT TGA AAA AAC			
PG10-Rv CAG GGA CCC ATT TGC CGG TC	60	<b>324</b>	This study
<b>PG13</b>			
PG13-Fw GGA TGT TCT ACG ACA ACC TG			
PG13-Rv AGC TGC GAG GTG ACG TTT GA	60	<b>582</b>	This study
<b>Multiplex PCR</b>			
<b>Multiplex 1 and 2*</b>	61	see above	This study

\*The multiplex 1 PCR contains primers for PG1, PG3 and PG9, while multiplex 2 PCR contains primers for PG2, PG4 and PG13. The two multiplex PCR primer sets included the Psy primers for the detection of the whole *P. syringae* complex. The amplicon sizes for multiplex PCRs are described in the single PCR table section.

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**Table 3.** Specificity and sensitivity of the primers tested with individual primer pairs

Target Phylogroups	Number of strains tested	PA	PD	ND	NA	Relative accuracy, A (%)	Diagnostic sensitivity, SE (%)	Diagnostic specificity, SP (%)
PG 1	131	24	0	1	106	99	96	100
PG 2	157	38	1	0	119	99	100	99
PG 3	132	17	0	0	115	100	100	100
PG 4	167	7	0	1	159	99	87	100
PG 9	138	14	0	0	124	100	100	100
PG 10	131	8	0	10	113	92	44	100
PG 13	191	34	1	0	157	99	100	99

PA= positive agreement, or the number of positive results obtained with a sample containing the targeted phylogroup

PD = positive deviation (false positive), or the number of positive results obtained with a sample that did not contain the targeted phylogroup

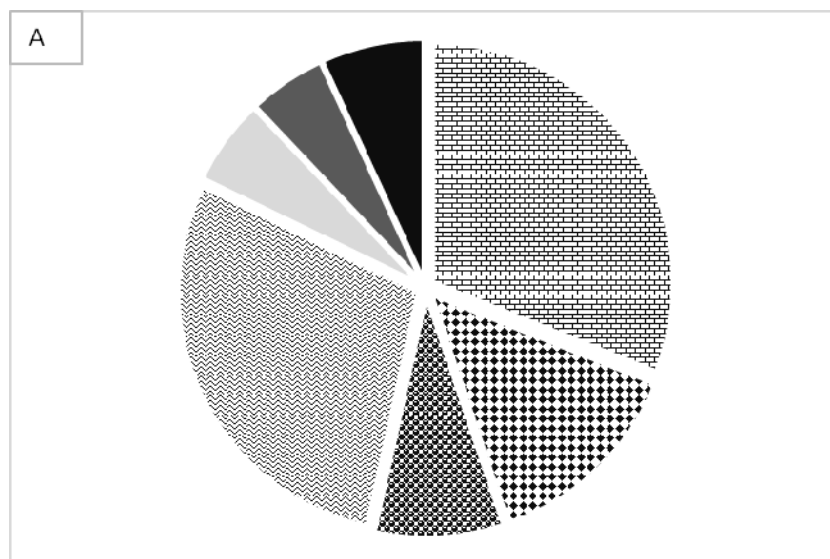
ND = negative deviation (false negative), or the number of negative results obtained with a sample that contained the targeted phylogroup

NA = negative agreement, or the number of negative results obtained with a sample that did not contain the targeted phylogroup

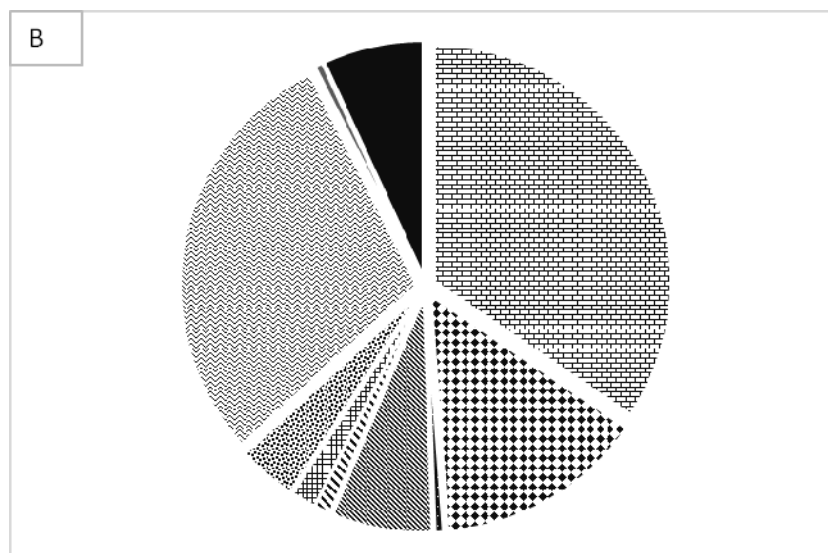
$A = 100\% * [(PA + NA) / (PA + PD + ND + NA)]$













$SE = 100\% [PA / (PA + ND)]$

$SP = 100\% * [NA / (NA + PD)]$



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**Figure 1.** Relative proportion of *Pseudomonas syringae* phylogroups (PG1 , PG2 , PG3 , PG7 , PG8 , PG7/8 , PG9 , PG10 , PG13  ) in kiwifruit and apricot samples from Southeastern France. Results based on the PCR primers described in this work (A) were compared with those obtained by phylogenetic analysis based on sequences the *cts* gene (B). For some isolates the phylogroup could not be determined (Ni ) or they were assigned to multiple phylogroups because they were composed of several strains (Mi ). Some of the isolates tested didn't belong to the *P. syringae* species (NP ).

### Table S1

List of strains used during this study and related information.

### Table S2

List of the gene sequences used during this study.