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Improved racemateresolution of pentan-2-ol and trans-(Z)-cyclooct-5-ene-1,2-diol by lipase catalysis.

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Abstract

Lipases are important catalysts in chiral synthesis due to their wide substrate recognition combined with a high stereoselectivity. We demonstrate here that the state, free or immobilized, of *Candida antarctica* lipase B (CaLB) affects enantioselectivity and also alters the temperature dependency of the enzyme. This indicates that CaLB undergoes various conformations induced by its interaction with the different immobilization supports studied. Molecular imprinting experiments, using immobilized enzyme co-dried with mimic substrate molecules, enhanced the enantiomeric ratio two-fold or three-fold, depending on the immobilization support. The structure of the acyl donor has a pronounced effect on CaLB catalyzed resolution, due to the proximity of the acyl and alcohol moieties during catalysis. When the acylation of pentan-2-ol was examined, we found that the 3C methyl propanoate donor afforded the highest resolution. Trans-(Z)-cyclooct-5-en-1,2-diol was used as

a model racemic substrate to study the ability of lipase to catalyze the resolution of difunctionalized compounds. There was a clear enhancement in the enantiomer selectivity of the biotransformation of the diol when vinyl butanoate is used as the acyl donor. The conversion and enantiomeric excess of (1*R*,2*R*)-monoacetates were enhanced, using immobilized CaLB, when the chain length of the donors increased from C2 to C4.

Keywords: Biocatalysis; Kinetic Resolution; Lipase; Immobilized Enzymes; Substrate Engineering; Bioreactors.

1. Introduction

Chiral synthesis is a key process in synthetic organic chemistry and, in particular, in medicinal chemistry because the different enantiomers or diastereomers of a molecule frequently have substantially different biological activity. Enzymes with very high regio- and stereo- selectivity are catalysts of choice for chiral resolution and asymmetric synthesis. The requirement for compounds with high stereochemical purity is increasing and there is a growing need for enzymes and reaction conditions with improved and precise performance. Lipases are often used because they are available from various sources, they do not need cofactors and they are relatively thermostable. In addition, they also accept a broad range of substrates, yet exhibit high selectivity (Bornscheuer and Kazlauskas, 2005). *Candida antarctica* Lipase B (CaLB) (Noël et al., 2004; Léonard et al., 2004) and *Burkholderia cepacia* Lipase (BC) are relatively thermostable enzymes that are often used as catalysts in various biotransformations in non-aqueous media and demonstrate high enantioselectivity toward many chiral substrates.

Catalytic activity, selectivity, specificity and enzyme stability are key factors affecting the efficiency of biocatalysts and enzyme immobilization may improve these properties. In addition, immobilization of lipases confers good thermal and operational stabilities, enabling the development of continuous processes. Immobilization of lipases by sol-gel encapsulation is an easy and effective method that can also increase enzyme activity and selectivity (Reetz et al., 2003; Hellner et al., 2011). Another successful strategy for enhancing the activity and selectivity of lipases involves the tuning of the enzyme active site by molecular imprinting. The “molecular imprinting effect” theory was proposed by several authors to explain the modification of enzyme selectivity or activity in the presence of molecules that mimic substrates (Hellner et al., 2011; Yan et al., 2009; Yang et al., 2010, Chaput et al., 2012). The theory is that the enzyme slightly adjusts its active site structure around the imprint molecule and remains “trapped” in this conformation until the substrate enters. For example, pretreatment of a lipase with a chiral substrate such as (*R*)-2-octanol results in enantioselective activation (Furukawa et al., 2002). The dual use of sol-gel entrapment and imprinting has been successfully performed with lipases, to enhance their activity or selectivity (Reetz et al., 2003; Hellner et al., 2011; Cao et al., 2009). In these studies the enzyme is brought together with an “imprinting” molecule and then co-immobilized by sol-gel encapsulation.

The reaction efficiency can be enhanced by modifying the biocatalysts and also by changing the reaction conditions. The influence of temperature on selectivity is one of the major concerns in enzymatic reactions; in most cases, stereoselectivity decreases with increasing temperature. Very few groups have studied the temperature effect on lipase-catalyzed kinetic resolutions in continuous-flow mode; most data use batch mode (Abahazi et al., 2014).

Acyl donors also have the ability to change lipase activity and selectivity during kinetic resolution of chiral molecules through acylation (Devendran and Yadav, 2014). By changing the acyl group of acylation agents, the lipase is transformed into various acyl-enzyme

intermediates, with structurally different active sites. This exerts a variety of chiral discrimination abilities during the enantioselective acylation of chiral molecules (Ema et al., 1996).

In the present study, different strategies -engineering of the substrate molecule, reaction temperature, use of imprinting molecules and immobilization of the enzyme- have been investigated to explore lipase activity and enantioselectivity toward pentan-2-ol, through the acylation reaction. (*S*)-pentan-2-ol is a chiral intermediate in the synthesis of several potential anti-Alzheimer drugs that inhibit β -amyloid peptide release and/or its synthesis (Audia et al., 1998). In addition, lipase-mediated desymmetrization of trans-(*Z*)-cyclooct-5-ene-1,2-diols (Rouillard et al., 2014) was examined. Indeed, enantiomerically pure vicinal diols are interesting building blocks for the production of flavours and fragrances (Rouillard et al., 2014). Resolution of pentan-2-ol experiments were performed in a continuous solid/gas reactor that allows the control and independent variation of the thermodynamic activity of substrates and other components. The solid/gas reactor offers the possibility to modulate, and to study independently, the effect of each component present in the reaction medium or the effect of each reaction parameter, all other things being equal (Lamare et al., 2004).

2. Materials and Methods

2.1. Materials

Lipases from *Candida antarctica* (immobilized on acrylic resin 077K1155, 10 000 PLU⁻¹ Novozym 435[®]) and from *Burkholderia cepacia* (free and immobilized on diatom MKBB3465, 500 PLU⁻¹) were purchased from Sigma Aldrich (Sigma-Aldrich Chemie S.a.r.l., Saint Quentin Fallavier, France). For pentan-2-ol resolution, free CaLB (Lipozyme[®]) was used. It was a gift from Novozymes A/S (Bagsværd, Denmark) and was used after purification by gel filtration

on PD 10 column (Health Care, Denmark), using ultra-pure water as elutant and then freeze-dried. For diol resolution, free CaLB was purchased from Sigma Aldrich. All enzymes were stored in a vacuum dessicator containing P_2O_5 before use.

All other chemicals were purchased from Sigma Aldrich and were used without further purification except in the case of vinyl acetate which was used after fresh distillation.

2.2. Preparation of lipase co-dried with imprinting molecules

In order to prepare Novozym435® co-dried with imprinting molecules, 0.5g of immobilized biocatalyst was mixed in 3mL of hexane at room temperature during two hours with (*R*)-2-aminohexane, (*S*)-2-aminohexane or 1-aminohexane at different concentrations. The aim was to obtain a ratio of 1, 10, 100 or 1000 moles of amine per mole of enzyme for each amine, assuming that the amount of active lipase on the carrier was equal to 1000nmole/g carrier (Hedfors et al., 2010).

2.3. General procedure for Sol-Gel Immobilization of Lipase

Sol-gel immobilized CaLB in presence of imprinting molecules or not, was prepared according to previous works (Hellner et al., 2011; Weiser et al., 2012). The following compounds were mixed in a 5mL glass vial: 390µL Tris-HCl buffer (0.1M, pH 7.5), 200µL propan-2-ol, possibly the imprinting molecule ((*R*)-2-aminohexane, (*S*)-2-aminohexane or 1-aminohexane at a ratio of 1 or 1000 moles of amine per mole of enzyme, assuming a quantity of enzyme of 50mg), 100µL NaF 1M, 200µL polyethylene glycol 1000(4% w/v) at room temperature for 10 min. The silane precursors TEOS(tetraethoxysilane):OcTEOS(octyltriethoxysilane):PhTEOS(phenyltriethoxysilane)(10:7:3mmol) (in the given molar ratio) and 50mg of purified free CaLB were then added under stirring, submitted to sonication during 15 min and agitation during 12 hours at room temperature in order to obtain complete polymerization. The resulting solid was washed with

propan-2-ol (7mL), distilled water (5mL), propan-2-ol (5mL) and hexane (5mL). The residual white powder was dried in a vacuum desiccator.

2.4. Instrumentation

IR spectra were performed with a Perkin–Elmer Spectrum 100 IRFT-ATR instrument. ^1H and ^{13}C NMR were obtained with a JEOL JNM LA400 (400 MHz) spectrometer. Chemical shifts (δ) are given in parts per million (ppm) downfield from internal standard tetramethylsilane (TMS). Coupling constants J are given in Hz. The high resolution mass spectra (HRMS) were obtained with a Varian MAT311 spectrometer in the Centre Régional de Mesures Physiques de l’Ouest (CRMPO, University of Rennes, France).

2.5. General procedure for enantioselective acetylation of racemic pentan-2-ol using lipase

For pentan-2-ol resolution reactions, initial rates of reaction were calculated at different temperatures in a solid-gas reactor (Scheme 1). In this type of reactor, the enzyme in the solid state, is packed between two layers of glass wool, to form a packed bed, which is percolated by a carried gas, transporting gaseous substrates. A full description of this type of reactor was given previously (Lamare et al., 2004). Thermodynamic activities for ester and alcohol substrates were respectively $a_{\text{ester}}=0.1$ and $a_{\text{alcohol}}=0.05$. Reactions were carried out under anhydrous conditions. Pentan-2-ol commercial solutions was dried by refluxing with magnesium and then distilled. Methyl esters commercial solutions were dried with Na_2CO_3 and distilled from P_2O_5 . The dried substrates were stored under argon atmosphere and over molecular sieves. The amount of enzyme in the fixed-bed of the reactor was equal to 2.5 mg of Novozym 435[®] or 0.15 mg of free CaLB. The total gaseous flow was equal to 2000 $\mu\text{mol}\cdot\text{min}^{-1}$.

For experiments with Novozym435[®] co-dried with imprinting molecules, the mass of enzyme used was 1 mg and for sol-gel immobilized CaLB it was equal to 2mg, with a total flow rate

equal to $1400\mu\text{mol}\cdot\text{min}^{-1}$. In these conditions, conversion level of substrates was comprised between 2 and 5% and allowed initial reaction rates to be calculated.

For the solid-gas system analysis, the vapor phase leaving the bioreactor was injected in a gas chromatograph (Agilent model 6890N Series) with the column Chirasil-Dex CB ($25\text{m}\times 0.25\text{mm i.d.}\times 0.25\mu\text{m}$ β -cyclodextrin; Chrompack, France), as previously described (Létisse et al., 2003). The column temperature was programmed to hold between 5 and 20 min at 55°C then to increase at different rates from 55 to 85 to 160°C , depending on the ester used as substrate. The external calibration of the two substrates (pentan-2-ol and methyl esters) was carried out by programming a range of their partial pressures in the bioreactor and by analyzing with the gas chromatograph. For the products pent-2-yl esters, an external calibration was carried out with pure esters synthesized as described below, by using gas chromatography (Agilent 7890A) equipped with an auto-sampler (7688B), in the same analytical conditions as described above. For accurate determination of E values the vapor phase leaving the bioreactor was re-condensed if necessary and was then partially evaporated in order to enhance pent-2-yl esters detection and quantification. The enantiomeric ratio was calculated using the equation from Wescott and Klivanov (Wescott and Klivanov, 1993): $E = V_i^R / V_i^S$, V_i^R and V_i^S are the initial rates of (R)-pent-2-yl ester and (S)-pent-2-yl ester synthesis respectively. The enantiomeric ratios were calculated by taking the average of three different experiments. The associated standard deviation was calculated for each result.

2.6. General procedure for the synthesis of pent-2-yl esters (1-5)

To a solution of pentan-2-ol (1 eq) in pyridine (0.5 eq), anhydride (1.1 eq) was added and the mixture was kept at room temperature for 16 h. The mixture was poured into ice cold water and extracted with ethyl acetate ($3\times 20\text{ mL}$). The ethyl acetate extract was washed with water ($2\times 10\text{ mL}$), 1 N HCl ($2\times 10\text{ mL}$), and finally with water ($3\times 10\text{ mL}$), dried over MgSO_4 ,

filtered, and the solvent was removed under vacuum. The product was purified by column chromatography (silica gel 200–400 mesh, usually using 9:1 petrol ether/ethyl acetate).

Pent-2-yl propanoate(1)

Compound (**1**) was obtained as colorless oil. ν_{\max} (cm^{-1}): 2961, 2937, 1733, 1463, 1377, 1190.

δ_{H} NMR (400 MHz, CDCl_3): 4.80-5.03 (m, 1H, CH-O), 2.12-2.32 (m, 2H, CH_2), 1.19-1.67 (m, 10H, $2\times\text{CH}_3$, $2\times\text{CH}_2$), 0.92 (m, 3H, CH_3).

Pent-2-yl butanoate (2)

Compound (**2**) was obtained as colorless oil. ν_{\max} (cm^{-1}): 2962, 2935, 1731, 1458, 1379, 1184,

1121. δ_{H} NMR (400 MHz, CDCl_3): 4.81-5.00 (m, 1H, CH-O), 2.26 (t, $J = 7.4$ Hz, 2H, CH_2), 1.28-1.77 (m, 6H, $3\times\text{CH}_2$), 1.29 (d, $J=6.4$ Hz, 3H, CH_3), 0.98-0.86 (m, 6H, $2\times\text{CH}_3$).

Pent-2-yl hexanoate(3)

Compound (**3**) was obtained as colorless oil. ν_{\max} (cm^{-1}): 2958, 2933, 1732, 1459, 1377, 1245,

1176. δ_{H} NMR (400 MHz, CDCl_3): 4.85-5.01 (m, 1H, CH-O), 2.27 (t, $J = 7.4$ Hz, 2H, CH_2), 1.67-1.19 (m, 13H, CH_3 , $6\times\text{CH}_2$), 0.93-0.88 (m, 6H, $2\times\text{CH}_3$).

Pent-2-yl isobutanoate(4)

Compound (**4**) was obtained as colorless oil. ν_{\max} (cm^{-1}): 2964, 2935, 1730, 1468, 1384, 1260,

1194, 1160. δ_{H} NMR (400 MHz, CDCl_3): 4.80-4.92 (m, 1H, CH-O), 2.35-2.50 (m, 2H, CH_2), 1.53-1.04 (m, 14H, $4\times\text{CH}_3$, CH_2), 0.85(t, $J=7.2$ Hz, 3H, CH_3).

Pent-2-yl decanoate(5)

Compound (**5**) was obtained as colorless oil. ν_{\max} (cm^{-1}): 2924, 2854, 1732, 1458, 1376, 1175,

1121. δ_{H} NMR (400 MHz, CDCl_3): 4.79-4.95 (m, 1H, CH-O), 2.26 (t, $J = 7.4$ Hz, 2H, CH_2), 1.63-1.20 (m, 21H, CH_3 , $9\times\text{CH}_2$), 0.93-0.86 (m, 6H, $2\times\text{CH}_3$).

2.7. General procedure for the synthesis of diol (**7**) using mesoepoxyde(**6**) as intermediate:

(*Z*)-(1*S*,8*R*)-9-oxa-bicyclo[6.1.0]non-4-ene (**6**). This compound(**6**) was synthesized and analysed by IR and NMR as previously described (Rouillard et al., 2014). Trans-(*Z*)-cyclooct-5-en-1,2-diol (**7**) was then obtained from(**6**) and analyzed as previously described (Rouillard et al., 2014).

2.8. General procedure for enantioselective acetylation of racemic trans-(*Z*)-cyclooct-5-en-1,2-diol (**7**) using lipase

To a solution of racemic diol (**7**) (0.2 g, 1.41 mmol) solubilized in 2.5 mL of THF and under inert atmosphere, the corresponding vinyl ester (ethanoate, propanoate, butanoate, 14 mmol) and free or immobilized lipase were added. The mixture was stirred at room temperature and then filtrated, extracted and purified as previously described (Rouillard et al., 2014) to give the esters: (1*R*,2*R*)-(Z)-1-hydroxy-cyclooct-4-enylethanoate(**8a**), (1*S*,2*S*)-(Z)-1-hydroxy-cyclooct-4-enyl ethanoate(**8b**), (1*R*,2*R*)-(Z)-1-hydroxy-cyclooct-4-enyl propanoate(**9a**), (1*S*,2*S*)-(Z)-1-hydroxy-cyclooct-4-enyl propanoate(**9b**), (1*R*,2*R*)-(Z)-1-hydroxy-cyclooct-4-enyl butanoate(**10a**) and (1*S*,2*S*)-(Z)-1-hydroxy-cyclooct-4-enyl butanoate(**10b**).

Compounds(**8a**) and (**8b**) were analysed by IR and NMR as previously described (Rouillard et al., 2014).

(*Z*)-1-hydroxy-cyclooct-4-enyl propanoate(**9a** and **9b**)

Compounds(**9a** and **9b**) were obtained as colorless oil. ν_{\max} (cm⁻¹): 3413, 2942, 1718, 1356, 1272, 1183, 1028. δ_{H} NMR (400 MHz, CDCl₃): 5.54 – 5.81 (2H, m, $\text{CH}=\text{CH}$), 4.96 (1H, td, $J = 8.7, 3.8$ Hz, $\text{CH}-\text{CO}$), 3.90 (1H, td, $J = 8.4, 3.8$ Hz, $\text{CH}-\text{COH}$), 2.33 – 2.46 (4H, m, 2x CH_2), 2.01 – 2.26 (5H, m, 2x CH_2 , C-OH), 1.66 – 1.79 (2H, m, CH_2), 1.34 – 1.03 (3H, m, CH_3).

(*Z*)-1-hydroxy-cyclooct-4-enyl butanoate (**10a** and **10b**)

Compounds(**10a** and **10b**) were obtained as colorless oil. ν_{\max} (cm⁻¹): 3414, 2962, 1718, 1255, 1177, 1042. δ_{H} NMR (400 MHz, CDCl₃): 5.60-5.73 (1H, m, $\text{CH}=\text{CH}$), 4.97 (1H, td, $J =$

8.7, 3.8 Hz, $\underline{\text{C}}\text{H}-\text{CO}$), 3.91 (1H, td, $J = 8.5, 3.9$ Hz, $\underline{\text{C}}\text{H}-\text{COH}$), 2.30-2.48 (4H, m, $2 \times \underline{\text{C}}\text{H}_2$) 2.03-2.22 (5H, m $4 \times \underline{\text{C}}\text{H}_2$, C-OH), 1.60 – 1.83 (m, 4H, $2 \times \underline{\text{C}}\text{H}_2$), 0.96 (3H, t, $J = 8.0$ Hz, $\underline{\text{C}}\text{H}_3$).

For the determination of enantiomeric excess (*ee*) of monoesters gas chromatography (Agilent 7890A) was used as previously described (Rouillard et al., 2014). The enantiomeric excesses and yields were calculated by taking the average of three different experiments. The associated standard deviation was calculated for each result.

For the determination of enantiomeric excess of remaining diols, high performance liquid chromatography was carried out in a Waters 600s combined with an auto-sampler (Waters 717 plus) as previously described (Rouillard et al., 2014).

3. Results and Discussion

3.1. Enantioselective acylation of pentan-2-ol using free and immobilized CaLB

Novozym 435® (CaLB immobilized on acrylic resin) was used here as a biocatalyst. The effect of acyl donor chain length on resolution was studied by measuring the enantiomeric ratio (*E*) in the kinetic resolution of racemic pentan-2-ol through acylation with different methyl esters (Figure 1), at different temperatures ranging from 50 to 100°C, in a continuous solid-gas reactor.

When CaLB is immobilized on acrylic resin, the highest *E* values are obtained with methyl propanoate as an acyl donor, at each temperature tested. *E* values decrease when the chain length of the acyl donor increases from C3 to C4 and to C6, at each temperature tested. *E* value is very low when methyl isobutanoate is used as an acyl donor (Figure 2A). The same trend was obtained for enzyme activity at 60°C, but the values were divided by 3 and 5 going

from a C3 acyl chain to C4 and C6 acyl chains, respectively, and divided by 10 when methyl isobutanoate is used as an acyl donor.

The enantiomeric ratios and enzyme activities were then measured with free CaLB instead of immobilized lipase, under the same conditions and using a longer acyl donor (C10 instead of C6). The highest enzymatic activities were obtained with the shortest acyl chain tested (C3), both at 60 and 80°C. Three fold higher values of enantiomeric ratio *E* were obtained compared to immobilized enzyme on acrylic resin (Figure 2A and 2B). Interestingly, in previous studies, it was shown by Jacobson *et al.* that free and immobilized CaLB on acrylic resin presented the same enantioselectivity toward other secondary alcohols such as 1-phenoxy-propan-2-ol or 3-chloro-1-phenoxy-propan-2-ol, through acylation with vinyl butanoate in hexane (Jacobsen *et al.*, 2005).

The effect of acyl donor length on the resolution of pentan-2-ol with free CaLB demonstrates that the highest *E* value is obtained at 50°C with a C3 acyl chain, and at 80°C with a C10 acyl chain (Figure 2B). CaLB has a deep narrow active site into which the substrate ester binds in a hairpin structure (Uppenberg *et al.*, 1995). The acyl and pentan-2-ol moieties are thus brought spatially close together during catalysis; this could explain lipase sensitivity in relation to the length of the acyl chain. Each different acyl-enzyme intermediate formed leads to different active site structures, with different enantioselectivity toward the secondary alcohol. Furthermore CaLB exhibits a funnel-like scissile fatty acid binding site, inside which fatty acids with chain lengths as long as 13 carbons can bind completely (Pleiss *et al.*, 1998); this means that amino acid residues in the acyl binding site that are at some distance from the point of chirality also affect enantioselectivity (Haeffner *et al.*, 1998). Finally, it is known that immobilization of a protein may produce distortion of the active site, therefore, it is not surprising that enantioselectivity of CaLB varies both with the acyl chain length of the donor and with immobilization. Recently two different conformations, open and closed, were

observed for crystal structure of CaLB at very high resolution (Stauch et al., 2015). The shift from the open form to the closed form involves the unfolding of the α -helix 5, that allows the formation of a salt bridge between Asp145 and Lys290, thus closing the catalytic cavity. Consequently, changes in both dielectric constant and ionic strength of the medium are able to affect interfacial activation of CaLB (Stauch et al., 2015). These recent findings give new insights into the possible effects of substrates, media and immobilization on CaLB fold and activity. These features make it extremely difficult to predict *a priori* and explain CaLBenantioselectivity.

The resolution of 3-methyl-2-butanol through acylation with vinyl esters was studied by Ottosson and Hult and it was found that long alkyl chains like vinyl octanoate afforded the highest resolution. A decrease in resolution was obtained with shorter chain lengths for vinyl hexanoate and vinyl butanoate. Shortening the acyl chain to vinyl propanoate gave a slight increase in resolution (Ottosson and Hult, 2001). There is no general rule concerning the effect of acyl chain length on CaLB-catalyzed resolution of secondary alcohols as it differs from one combination of acyl donor/alcohol to another. In conclusion, the choice of an achiral acyl donor strongly influences the efficiency of the resolution of a chiral alcohol in acylation reactions with CaLB.

We then wished to ascertain the conditions necessary to obtain a high enantioselectivity in acylation reactions of pentan-2-ol using immobilized lipase, compared with free CaLB. Methyl propanoate was chosen as acyl donor as it gives the best resolution compared to other acyl donors and also has the highest enzyme activity. 70°C was chosen as reaction temperature as it gives the best compromise between high enzyme activity and long-term stability.

The effect of co-drying CaLBNovozym 435 with imprinting molecules in hexane, before performing the acylation reaction on immobilized enzyme activity and enantioselectivity was first studied.

The different amines ((*R*)-, (*S*)-2-aminohexane and 1-aminohexane) have a similar effect on enantioselectivity: *E* doubles for 1 mole of amine per mole of enzyme but the increase of *E* is lower at higher concentrations of amines (Figure 3A). Enzymatic activity is increased 1.4 times to 1.8 times for 1 mole of amine per mole of enzyme and by lower amounts at higher concentrations of amines (1.5 to 1.7 times for 10 moles of amine per mole of enzyme, 1.3 to 1.6 times for 100 moles of amine per mole of enzyme and 1.15 to 1.5 times for 1000 moles of amine per mole of enzyme; Figure 3B). In all cases no amides were detectable in the reaction product. This is in agreement with the fact that alcohols deacylate the acyl-enzyme intermediate faster than amines (Öhrner et al., 1996).

The analysis of these different results is complex. The fact that the amines have a significant effect on enantioselectivity at a ratio 1/1 with the enzyme supports the hypothesis of an imprinting effect, but this is called into question by the fact that the three amines, with different structures, have an almost similar effect. The decrease in both enantioselectivity and activity using 100 or 1000 moles of amine per mole of enzyme instead of 1 or 10 moles could arise from the formation of non-reactive enzyme-amine complexes, as was hypothesized by Garcia-Urdiales et al., when studying the role of amines in the CaLB-catalyzed alcohol resolution (Garcia-Urdiales et al., 2001).

The effect of encapsulation prior to the acylation reaction of free CaLB, in the absence or presence of imprinting molecules, on enzyme activity and enantioselectivity toward pentan-2-ol was then studied.

Encapsulation of free CaLB leads to an increase of *E* from 109.0 ± 2.7 for free enzyme (lipozyme) to 154 ± 1 for CaLB encapsulated in sol/gel matrix (Figure 4). The value

corresponding to the activity of the encapsulated CaLBs multiplied by a factor of 1.7 compared to free enzyme. A substantially improved enantioselectivity and activity of sol-gel entrapped CaLB compared to non-immobilized lipase was also obtained by Ursoiu *et al.* in the acylation reaction of octan-2-ol (Ursoiu *et al.*, 2012).

Enantioselectivity remains almost identical with or without addition of amines in the sol/gel matrix, at 1 or 1000 moles per mole of enzyme and with all amines tested. Enzymatic activity remained constant with or without amines added in sol/gel matrix for all the amines tested.

The enantioselectivity of CaLB toward pentan-2-ol varies substantially between the free form ($E=109.0 \pm 2.7$), Novozym 435 ($E=51.2 \pm 1.0$) and the encapsulated enzyme ($E=154 \pm 1$). It has already been shown that Novozym 435, that corresponds to CaLB immobilized on Lewatit VP OC 1600, a macroporous hydrophobic acrylic polymer resin, displays very different selectivity compared to free CaLB or CaLB immobilized on other supports (Cabrera *et al.*, 2009; Mateo *et al.*, 2007). Recently, it was shown by Zisis *et al.* by combining experiment and simulation, that CalB is an interfacially activated enzyme and that the hydrophobicity of the support used for its immobilization influences both enzyme conformation and substrate specificity (Zisis *et al.*, 2015).

For enzyme encapsulated in sol/gel matrix, neither the enantioselectivity nor the activity were improved by the addition of chiral amines or by the primary amine. On the contrary, these parameters were significantly improved with the same amines when co-dried in hexane with the enzyme immobilized on acrylic resin. In a recent study, docking experiments into CaLB of various additives with imprinting effects, were performed, using the two different enzyme conformations (open and closed) observed by Stauch *et al.* in 2015 (Weiser *et al.*, 2016). These studies revealed synergistic effects including the stabilization of the active conformation of CaLB and provision of extra space for the mobile active site-covering lids after their removal. These types of effects were caused by large additives only, whereas smaller additives were unable to significantly stabilize the active conformation of the active site (Weiser *et al.*, 2016).

On that basis, it is probable that the “imprinting effect” for the investigated amines is rather based on allosteric effects. Further studies are required to fully explain these data.

3.2. Enantioselective acylation of diols using CaLB and BC lipases

Trans-(Z)-cyclooct-5-en-1,2-diol (**7**) was used as a model racemic substrate to study the ability of lipase to catalyze the resolution of these difunctionalized compounds through enantioselective acylation. The comparison of acyl donors and enzyme, together with the effects of immobilization of the enzyme, were carried out as outlined below.

The racemic diol (**7**) was synthesized from the commercial cycloocta-1,2-diene in two steps: epoxidation followed by ring opening in acidic conditions (Figure 5), as previously described (Rouillard et al., 2014).

The preparation of an optically enriched monoester was first performed using free or immobilized CaLB (Novozym 435) to desymmetrize *meso*-symmetric diol (**7**) using vinyl ethanoate, vinyl propanoate and vinyl butanoate as the acylating agents in THF solvent, at room temperature (Figure 6A and Table 1).

When vinyl ethanoate is used as the acyl donor, immobilized CaLB gives a higher conversion than free CaLB (16% compared to 8%) and also demonstrates a higher enantioselectivity for the (1*R*,2*R*) diol. It increased the enantiomeric excess (ee) of (1*R*,2*R*)-monoethanoate (**8a**) from 50% to 55% and the ee of (1*S*,2*S*)-diol (**7b**) increased from 4% to 9%. The immobilized CaLB was then chosen for reactions with three acyl donors containing different chain lengths.

The conversion and the enantiomeric excess of (1*R*,2*R*)-monoacetate were enhanced when the chain length increased from C2 to C4.

Conversion increased from 16% using vinyl ethanoate to 33% using vinyl propanoate and 48%, using vinyl butanoate (Table 1). The enantiomeric excess of the monoester (**8a**) increased

from 55% with a vinyl ethanoate donor to 73% or 96% using vinyl propanoate or vinyl butanoate, respectively. In all cases, the (1*R*,2*R*) diol was the preferred substrate for monoacetylation. The enantiomeric excess of the remaining (1*S*,2*S*)-diol (**7b**) increased from 9% using a vinyl ethanoate donor to 55% using vinyl propanoate and more than 95% using vinyl butanoate.

The same reaction was then performed using BC Lipase (Figure 6B and Table 2).

When vinyl ethanoate was used as an acyl donor, BC lipase resulted in a higher conversion than CaLB lipase, and interestingly, under the same conditions showed an inversion of enantioselectivity, with a preference for (1*S*,2*S*)-diol, and the formation of monoester (**8b**). Enzyme immobilization slightly enhanced conversion (26% compared to 17%) but not selectivity (Table 2). No reaction was observed when an acyl donor with longer chain length was used with immobilized BC Lipase.

Conclusion

In this study, it was shown that for pentan-2-ol resolution in a continuous solid/gas reactor enantioselectivity and temperature dependence of CaLB vary and depend on the conditions of immobilization of the enzyme. This indicates that the enzyme may take on different conformations induced by interactions with the immobilization supports. The acyl donor has a pronounced effect on CaLB-catalyzed resolution through acylation, due to the fact that the acyl and alcohol moieties are brought spatially close together during catalysis. A significant enhancement of resolution of the trans-(*Z*)-cyclooct-5-en-1,2-diol using CaLB Novozym 435 and vinyl butanoate, rather than vinyl ethanoate, as acyl donor, was also noticed. The data presented here contribute to the research into improving the stereochemical purity of

compounds obtained by biocatalysis as well as other possibilities such as the optimisation of solvent, temperature, water activity, choice of enzyme and enzyme immobilization methods.

Author Contributions

M.G. and V.T. designed the research. The experimental work was performed by N.F., B.B. and L.D. for the enantioselective acylation of pentan-2-ol using free and immobilized CaLB, by H.R. and R.D. for the enantioselective acylation of diols using CaLB and BC lipases and by A.B. for NMR spectra. M.G. wrote the manuscript with the cooperation of H.R. All authors discussed, edited and approved the submitted version.

Conflict of Interests

The authors declare that there is no conflict of interest regarding the publication of this paper.

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References

Abaházi, E., Boros, Z., Poppe, L., 2014. Additives Enhancing the Catalytic Properties of Lipase from *Burkholderia cepacia* Immobilized on Mixed-Function-Grafted Mesoporous Silica Gel, *Molecules*, 19, 9818-9837.

Audia, J.E., Britton, T.C., Droste, J.J., Folmer, B.K., Huffman, G.W., John, V., Latimer, L.H., Mabry, T.E., Nissen, J.S., Porter, W.J., Reel, J.K., Thorsett, E.D., Tung, J.S., Wu, J., Eid, C.N., Scott, W.L., 1998. Patent WO 98/22494.

Bornscheuer, U.T., Kazlauskas, R.J., 2005. *Hydrolases in Organic Synthesis: Regio and Stereoselective Transformations*, Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim.

Cabrera, Z., Fernandez-Lorente, G., Fernandez-Lafuente, R., Palomo, J. M., Guisan, J. M., 2009. Novozym 435 displays very different selectivity compared to lipase from *Candida antarctica* B adsorbed on other hydrophobic supports. *J. Mol. Catal. B: Enzym.* 57, 171–176.

Cao, X., Yang, J., Shu, L., Yu, B., Yan, Y., 2009. Improving esterification activity of *Burkholderiacepacia* lipase encapsulated in silica by bioimprinting with substrate analogues, *Proc. Biochem.*, 44, 177-182.

Chaput, L., Marton, Z., Pineau, P., Domon, L., Tran, V., Graber, M., 2012. Enhancing the enantioselectivity of CALB by substrate imprinting: a combined experimental and molecular dynamics simulation model study, *J. Mol. Catal. B: Enzym.*, 84, 55-61.

Devendran, S., Yadav, G.D., 2014. Microwave Assisted Enzymatic Kinetic Resolution of (\pm)-1-Phenyl-2-propyn-1-ol in Nonaqueous Media, *BioMed Research International*, 2014, 1-9.

Ema, T., Maeno, S., Takaya, Y., Sakai, T., Utaka, M., 1996. Significant Effect of Acyl Groups on Enantioselectivity in Lipase-Catalyzed Transesterifications, *Tetrahedron: Asymmetry*, 7, 625-628.

Furukawa, S., Ono, T., Ijima, H., Kawakami, K., 2002. Effect of imprinting sol-gel immobilized lipase with chiral template substrates in esterification of (*R*)-(+)- and (*S*)-(-)-glycidol, *J. Mol. Catal. B-Enzym.*, 17, 23-28.

Garcia-Udiales, E., Rebolledo, F., Gotor, V., 2001. Kinetic resolution of (\pm)-1-Phenylbutan-1-ol by Means of CALB-catalyzed Aminolyses: A study on the Role of the Amine in the Alcohol Resolution, *Adv. Synth. Catal.*, 343, 646-654.

Haefner, F., Norin, T., Hult, K., 1998. Molecular Modeling of the Enantioselectivity in Lipase-Catalyzed Transesterification Reactions, *Biophys. J.*, 74, 1251-1262.

Hedfors, C., Hult, K., Martinelle, M., 2010. Lipase chemoselectivity towards alcohol and thiol acyl acceptors in a transacylation reaction, *J. Mol. Catal. B-Enzym.*, 66, 120-123.

Hellner, G., Boros, Z., Tomin, A., Poppe, L., 2011. Novel Sol-Gel Lipases by Designed Bioimprinting for Continuous-Flow Kinetic Resolutions, *Adv. Synth. Catal.*, 353, 2481-2491.

Jacobsen, E.E., Andresen, L.S., Anthonsen, T., 2005. Immobilization does not influence the enantioselectivity of CAL-B catalyzed kinetic resolution of secondary alcohols, *Tetrahedron: Asymmetry*, 16, 847-850.

Lamare, S., Legoy, M.-D., Graber, M., 2004. Solid/gas bioreactors: powerful tools for fundamental research and efficient technology for industrial applications, *Green Chem.*, 6, 445.

Léonard, V., Lamare, S., Legoy, M.-D., Graber, M., 2004. Enantioselective acylation of R-2-pentanol in a solid/gas reactor catalysed by lipase B from *Candida antarctica*, *J. Mol. Catal. B-Enzym.*, 32, 53-59.

Létisse, F., Lamare, S., Legoy, M.-D., Graber, M., 2003. Solid/gas biocatalysis: an appropriate tool to study the influence of organic compounds on kinetics of lipase catalyzed alcoholysis, *BBA-Proteins and Proteomics*, 1652, 27–34.

Mateo, C., Palomo, J. M., Fernandez-Lorente, G., Guisan, J. M., Fernandez-Lafuente, R., 2007. Improvement of enzyme activity, stability and selectivity via immobilization techniques, *Enz. Microbial Technol.* 40, 1451–1463.

Pleiss, J., Fischer, M., Schmid, R.D., 1998. Anatomy of lipase binding sites: the scissile fatty acid binding site, *Chem. Phys. Lipids*, 93, 67–80.

Noël, M., Lozano, P., Vaultier, M., Iborra, J.L., 2004. Kinetic resolution of rac-2-pentanol catalyzed by *Candida antarctica* lipase B in the ionic liquid, 1-butyl-3-methylimidazolium bis[(trifluoromethyl)sulfonyl]amide. *Biotechnol. Lett.*, 26, 301–306.

Öhrner, N., Orrenius, C., Mattson, A., Norin, T., Hult, K., 1996. Kinetic resolutions of amine and thiol analogues of secondary alcohols catalyzed by the *Candida antarctica* lipase B, *Enzyme Microb. Technol.*, 19, 328–331.

Ottosson, J., Hult, K., 2001. Influence of acyl chain length on the enantioselectivity of *Candida antarctica* lipase B and its thermodynamic components in kinetic resolution of secondary alcohols, *J. Mol. Catal. B-Enzym.*, 11, 1025–1028.

Reetz, M.T., Tielmann, P., Wiesenhöfer, W., Könen, W., Zonta, A., 2003. Second Generation Sol-Gel Encapsulated Lipases: Robust Heterogeneous Biocatalysts, *Adv. Synth. Catal.*, 345, 717–728.

Rouillard, H., Deau, E., Domon, L., Chérouvrier, J.-R., Graber, M., Thiéry, V., 2014. Microwave-Assisted Kinetic Resolution of Homochiral (Z)-Cyclooct-5-ene-1,2-diol and (Z)-2-Acetoxyoct-4-enyl Acetate Using Lipases, *Molecules*, 19, 9215-9227.

Stauch, B., Fisher, S.J., Cianci M. 2015. Open and closed states of *Candida antarctica* lipase B: protonation and the mechanism of interfacial activation, *J. Lipid Res.*, 56, 2348-2358.

Uppenberg, J., Oehrner, N., Norin, T., Hult, K., Kleywegt, G.J., Patkar, S., Jones, T.A., 1995. Crystallographic and molecular-modeling studies of lipase B from *Candida antarctica* reveal a stereospecificity pocket for secondary alcohols, *Biochemistry*, 34, 16838-16851.

Ursoiu, A., Paul, C., Kurtán, T., Péter, F., 2012. Sol-gel Entrapped *Candida antarctica* lipase B - A Biocatalyst with Excellent Stability for Kinetic Resolution of Secondary Alcohols, *Molecules*, 17, 13045-13061.

Weiser, D., Boros, Z., Hornyánszky, G., Tóth, A., Poppe, L., 2012. Disubstituted dialkoxysilane precursors in binary and ternary sol-gel systems for lipase immobilization, *Proc. Biochem.*, 47, 428-434.

Weiser, D., Sóti, P.L., Bánóczy, G., Bódai, V., Kiss, B., Gellért, A., Nagy, Z.K., Koczka, B., Szilágyi, A., Marosi, G., Poppe, L., 2016. Bioimprinted lipases in PVA nanofibers as efficient immobilized biocatalysts. *Tetrahedron*, doi:10.1016/j.tet.2016.06.027 (in press)

Wescott, C.R., Klivanov, A.M., 1993. Predicting the solvent dependence of enzymic substrate specificity using semiempirical thermodynamic calculations, *J. Am. Chem. Soc.*, 115, 10362-10363.

Yan, J.Y., Yan, Y.J., Yang, J.K., Xu, L., Liu, Y., 2009. Combined strategy for preparation of a bioimprinted *Geotrichum sp.* lipase biocatalyst effective in non-aqueous media, *Proc. Biochem.*, 44, 1128-1132.

Yang, J., Liu, L., Cao, X., 2010. Combination of bioimprinting and silane precursor alkyls improved the activity of sol-gel-encapsulated lipase, *Enzyme Microb. Technol.*, 46, 57-261.

Zisis, T., Freddolino, P.L., Turunen, P., van Teeseling, M.C.F., Rowan, A.E., Blank, K.G., 2015. Interfacial Activation of *Candida antarctica* Lipase B: Combined Evidence from Experiment and Simulation, *Biochemistry* 54, 5969-5979.

Table 1

Effect of immobilization and acyl donor for *Candida antarctica* lipase B catalyzed enantioselective acylation of diol (7). Standard deviations were calculated with three values of ee and of yield.

Acyl donor	Enzyme	Conversion (%)	Monoester (8a, 9a or 10a) ee (%)	Diol (7b) ee (%)
Vinyl ethanoate	free CaLB	8±2	50±2 (1R,2R)	4±2 (1S,2S)
Vinyl ethanoate	CaLB immobilized on acrylic resin	16±1	55±3 (1R,2R)	9±2 (1S,2S)
Vinylpropanoate	CaLB immobilized on acrylic resin	33±2	73±2 (1R,2R)	55±2 (1S,2S)
Vinylbutanoate	CaLB immobilized on acrylic resin	48±1	96±1 (1R,2R)	>95 (1S,2S)

Table 2

Effect of immobilization and acyl donor for *Burkholderiacepalipase* catalyzed enantioselective acylation of diol (7). Standard deviations were calculated with three values of ee and of yield.

Acyl donor	Enzyme	Conversion (%)	Monoester (8b) ee (%)	Diol (7a) ee (%)
Vinyl ethanoate	free BC	17±2	60±2 (<i>1S,2S</i>)	12±2 (<i>1R,2R</i>)
Vinyl ethanoate	BC immobilized on diatomite	26±1	57±3 (<i>1S,2S</i>)	12±2 (<i>1R,2R</i>)
Vinylpropanoate	BC immobilized on diatomite	-NR	-	-
Vinylbutanoate	BC immobilized on diatomite	- NR	-	-