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# IN VIVO IDENTIFICATION OF THE PASSIVE MECHANICAL PROPERTIES OF DEEP SOFT TISSUES IN THE HUMAN LEG

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1 **IN VIVO IDENTIFICATION OF THE PASSIVE MECHANICAL**  
2 **PROPERTIES OF DEEP SOFT TISSUES IN THE HUMAN LEG**

3  
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**ABSTRACT**

Goal: A non-invasive method is proposed to identify *in vivo* the passive mechanical properties of deep soft tissues in the human leg.

How: Force-displacement curves in response to a localized compression of the calf are measured with a custom made experimental setup. The material parameters of a finite element model are then calibrated against the experimental curves using a genetic algorithm. A thorough investigation of the efficacy of this method to identify such mechanical properties is conducted through a design of experiments analysis and mixed numerical - experimental validations.

Results: It is the first time that a thorough analysis is conducted to really separate the contribution of deep and superficial tissues in the response to compression tests and this permits to estimate the parameters of deep soft tissues on four subjects independently of the response of their other tissues. Two strain energy density functions are compared. It is shown that a 2<sup>nd</sup> order reduced polynomial better describes the passive mechanical behavior of the deep soft tissues of the leg than the neo-Hookean model.

**KEYWORDS**

Calf, Finite element model, Inverse method, Soft tissues

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# 1. INTRODUCTION

## 1. Motivations

Different draining mechanisms prevent blood from stagnating in the legs of healthy subjects, such as the effect of muscular contraction, the activity of the sympathetic nervous system or the venous valve system [1]. In case of default in one of these mechanisms, chronic venous insufficiency (CVI) may appear. Symptoms of CVI range from “heavy leg” feelings to development of ulcers. CVI affects about thirty percent of the French population [2]. Compression of the limb using medical compression stockings or bandages is a widely used treatment and is efficient for almost all forms of CVI [3]. The benefits of these treatments are not to be proven, but still, for some patients, the therapeutic goal is not reached and generally, there is a need to improve their comfort. To understand the mechanisms of this treatment and to be able to better design and size the stockings or bandages, biomechanical numerical models have been developed [4]–[7].

A challenging step to develop these models is to identify the material properties of soft tissues. To that extent, several characterization methods have been established. A first approach to characterize the mechanical properties of soft tissues is to perform *in vitro* tensile or compression tests on tissue sample [8], [9]. A second and more recent approach is by direct measurements of the mechanical properties of soft tissues *in vivo* with different techniques. For example, ultrasound elastography [10] and magnetic resonance elastography [11] techniques both map the elasticity of soft tissues. A third approach is to combine experimental data acquisitions with finite element (FE) model updating through an inverse method. This approach can address experimental data that could not be analyzed directly to identify material parameters of different constitutive models. Previously, FE model updating has been used *in vivo* on arterial tissue [12], brain tissue [13], skin [14] and adipose tissues [15] or musculo-skeletal tissues [4], [5], [16],[17].

## 2. Problem definition and proposed approach

The leg is composed of several compartments made of soft tissues (Figure 1):

- the compartment composed of adipose tissue and skin which, in this study, is referred to as superficial soft tissues;
- the *Fascia cruris* which is a collagenous membrane separating both compartments;
- the compartment of soft tissues surrounded by the fascia cruris, mainly composed of muscular tissue which, in this study, is referred to as deep soft tissue compartment.

It is believed that the transmission of pressure through soft tissues plays an important role in the treatment of venous insufficiency by elastic compression [4]. Therefore, modelling the biomechanical effects of elastic compression requires a precise

1 identification of the constitutive behavior of this medium through which the pressure is  
2 transmitted.

3 Also, a large inter-subject variability in the efficacy of the compressive treatments has  
4 been observed previously [18]. Therefore, the identification of mechanical properties  
5 must be patient-specific.

6 Thus, a non-invasive and fast methodology is required to permit further clinical studies  
7 with large cohorts.

8  
9 In this context, a fast and non-invasive method to identify passive mechanical properties  
10 of deep soft tissues is proposed and evaluated. Using an inverse method, subject-  
11 specific FE models and experimental localized compression data sets are combined to  
12 identify material parameters on four healthy subjects. The related assumptions are  
13 discussed to assess the accuracy of this approach.

14

### 15 **3. General outline of the paper**

16 The material and methods section details the methodology used to retrieve subject-  
17 specific material properties by acquiring localized compression data on the leg and  
18 implementing an inverse method. Then, in the results section, accuracy of the  
19 methodology is assessed and the material parameters of each subject are reported.  
20 Finally, in the discussion section, the experimental set-up and the assumptions of the FE  
21 model are thoroughly discussed to present their domain of validity and limitations.

22

## 2. MATERIAL AND METHODS

### 1. *Data acquisition and processing*

Four volunteers (3 males, 1 female, mean age: 27) underwent MRI scans, ultrasound acquisitions and localized compression data acquisition on the calf of the right hand-side of the leg, following informed consent. The ultrasound acquisitions (Figure 2 - A) and the MRI scans (Figure 2 - B) provide the subject-specific geometry used for the implementation of a FE model (Figure 2 - C). An ultrasound elastographic device (Aixplorer®; Super-Sonic Imagine, Aix-en-Provence, France) was used in a previous study [19] to reconstruct the fields of elastic modulus in the superficial tissues. B-mode scans acquired all around the leg were stitched together to reconstruct the anatomy of superficial soft tissues of the subject.

In the present study, an experimental setup has been designed to identify the passive mechanical properties of deep soft tissues. More specifically, displacements and reaction forces were measured after compressing the calf with a 30 mm diameter cylinder (Figure 3). Since these experimental data were aimed at being combined to a bi-dimensional finite-element (FE) model, a cylinder was chosen rather than a sphere. More, it is worth noting that with such geometrical arrangement, material with linear elastic behavior would give a linear force/displacement curve..

During data acquisition, the subject was seated in a chair allowing a relaxed state of the calf muscles. The subject's foot was fixed with straps and the knee was supported by rubber buffers avoiding motions of the leg during the acquisition. Double-sided adhesive tape was used at the leg-cylinder contact in order to avoid sliding. This adaptation reduces the number of unknowns in the inverse approach (*i.e.* no friction coefficient to identify), lowering the computation time.

An in-house LabVIEW® (National Instruments, Austin, TX, USA) code was used to display and record the reaction forces given by the 100 N load cell (LCMKD-100N, Omega engineering INC) with an accuracy of  $\pm 0.25\%$  and the displacements given by the 50 mm displacement sensor (MTN/EUG025-10, Monitran) with an accuracy of  $\pm 0.25$  mm. Three sets of localized compression were acquired per subject at a rate of 0.4 mm/s. The compression was increased until a displacement of 30 mm of the cylinder was reached. Only the loading part of the curve was further analyzed for the inverse analysis. This choice will be commented in the discussion section.

### 2. *Implementation of a subject-specific FE model*

The aim of the paper is to identify the deep soft tissue material parameters using an inverse FE analysis through an iterative process. Two imaging modalities were used to create the subject-specific FE model: ultrasound (Figure 2 - A) and MRI (Figure 2 - B). To register both images, the bones edges visible in both images were segmented and

1 used to perform a rigid body registration. The external geometry of the leg was  
 2 segmented from the ultrasound images [20] as well as the position of the *fascia cruris*.  
 3 Since the thickness of the *fascia cruris* shows high inter-individual variability [21] and  
 4 cannot be precisely determined from ultrasound images, a sensitivity analysis was  
 5 performed about this parameter.

6  
 7 In a **first model**, the behavior of soft tissues was described with a neo-Hookean hyper-  
 8 elastic constitutive equation. The strain energy density function was defined such as  
 9 [22]:

$$10 \quad U = C_{10}^H (\bar{I}_1 - 3) + \frac{1}{D_1} (J^{el} - 1)^2 \quad 11 \quad \text{Equation 1}$$

12 where  $\bar{I}_1$  is the first invariant of the isochoric deformation;  $J^{el}$  the elastic volume ratio;  $C_{10}^H$   
 13 and  $D_1$  the material parameters to identify.

14  
 15 In a **second model**, the behavior of deep soft tissues (*i.e.* below the *fascia cruris*) was  
 16 modelled with a 2<sup>nd</sup> order reduced polynomial strain energy density function defined  
 17 such as [22]:

$$U = C_{10}^P (\bar{I}_1 - 3) + C_{20} (\bar{I}_1 - 3)^2 \quad 18 \\ + \frac{1}{D_1} (J^{el} - 1)^2 + \frac{1}{D_2} (J^{el} - 1)^4 \quad 19 \quad \text{Equation 2}$$

20 where  $C_{10}^P$ ,  $C_{20}$ ,  $D_1$  and  $D_2$  are the material parameters to identify.

21  
 22 Material parameters are summarized in Table 1.  $D_1$  and  $D_2$  were not identified in this  
 23 study, common values were assigned to them [16]. This choice is discussed in the  
 24 discussion section.

25  
 26 The iterative process to identify the material parameters requires a low computation time  
 27 of the FE simulation. This is why it was chosen, in accordance with previous studies [4],  
 28 [6], [16], to implement a bi-dimensional FE model to mimic the 3D experiments. The  
 29 construction of the bi-dimensional FE model whose purpose is to replicate the 3D  
 30 experiment required several assumptions. Considering the chosen geometrical  
 31 arrangement, the bi-dimensional FE model is based on a classical plane-strain model  
 32 combined with correction functions in order to take into account the 3D effects.

33  
 34 As the soft tissues compression test was performed thanks to a 50 mm long cylinder,  
 35 edge effects may occur due to a non-uniform pressure distribution on the leg (Figure 4 –  
 36 B).

37 In order to assess the influence of the change from a 3D experiment to a plane-strain FE  
 38 model, a 3D model representing the experimental 3D problem was implemented and  
 39 compared to a plane-strain model. Idealized leg geometry was used to create the 3D FE  
 40 mesh, made of first order tetrahedral elements, using the meshing tools available in

1 Abaqus<sup>®</sup>. The material behavior was described by a neo-Hookean constitutive equation  
 2 (Equation 1).  
 3 Two computations were performed. In the first one, named 3D simulation, the  
 4 compression of the idealized leg is modelled in 3D with a 30 mm diameter and 50 mm  
 5 long cylinder (Figure 4 – A – 1). The external surface of the idealized bone was pinned  
 6 and the top and bottom faces of the idealized leg segment were unconstrained. In the  
 7 second one, named plane-strain simulation, the compression of the idealized leg is  
 8 modelled in plane strain with a 30 mm diameter circle (Figure 4 – A – 2). Plane-strain  
 9 results being calculated for a unit transverse length, reaction forces were multiplied by  
 10 the length of the compression cylinder (*i.e.* 50 mm). In order to evaluate the error due to  
 11 edge effects (Figure 4 – B ) made with a plane strain FE model, a correction function, P,  
 12 was defined as the difference between force/displacement curves from the two previous  
 13 computations as follows:

$$P(d) = 1 - \frac{\frac{RF^{3D}(d)}{L^{cylinder}}}{RF^{2D}(d)} \quad \text{Equation 3}$$

16 with,  $RF^{3D}(d)$  and  $RF^{2D}(d)$  being the reaction forces obtained respectively from the 3D  
 17 and 2D simulations, as a function of the displacement of the cylinder  $d$ ,  $L^{cylinder}$ , the  
 18 length of the cylinder. The results were used to apply a correction function (Figure 4 – C)  
 19 on the force/displacement curves in the bi-dimensional FE models for the 4 subjects.

21 Eventually, the bi-dimensional reconstruction comprising the bones, the compartment of  
 22 superficial soft tissues and the compartment of deep soft tissues, separated by the  
 23 *fascia cruris* (Figure 2 - C) was meshed with 3-node and 4-node linear plane strain finite  
 24 elements with hybrid formulation, using the meshing tools available in Abaqus<sup>®</sup>.

25 The compression cylinder was modelled as a 30 mm diameter circle (Figure 2 – C). Its  
 26 initial position and its translation direction were defined in agreement with each  
 27 experiment. The two bones and the cylinder were implemented as rigid bodies. The  
 28 bones were pinned and a 30 mm radial displacement was assigned as boundary  
 29 condition of the FE model for the cylinder.

30 In this study, the interaction between the leg and the cylinder was managed using  
 31 contact algorithms from those available in the Abaqus<sup>®</sup> library. In the tangential  
 32 direction, a rough behavior was chosen (*i.e.* infinite coefficient of friction) to satisfy the  
 33 experimental conditions (*i.e.* double-sided adhesive tape at the cylinder/leg interface). In  
 34 the normal direction, an exponential contact pressure/over-closure relationship was  
 35 used.

### 36 **3. Evaluation of the FE model sensitivity**

37 A sensitivity analysis of the FE model response to parameters of interest has been  
 38 performed for one subject through a design of experiments approach [23]. This  
 39 methodology allowed determining the sensitivity of the FE model to each parameter  
 40



(material parameters or thickness of the fascia cruris for example) and defining whether this bi-dimensional FE model can be used to identify the material parameters of deep soft tissues. For this sensitivity analysis, the behavior of deep soft tissues was modelled with a 2<sup>nd</sup> order reduced polynomial strain energy density function (Equation 2), as it was observed that this constitutive law better fits the experimental results.

### **Parameter selection**

5 parameters were taken into consideration, the  $C_{10}^P$  and  $C_{20}$  material parameters of deep soft tissues, the thickness of the *fascia cruris*, the  $C_{10}$  material parameter of the fascia cruris and the  $C_{10}^H$  material parameter of the superficial soft tissues (Figure 1). To compare the sensitivities, the parameters were scaled and centered from -1 to 1 in their respected domain as reported in Table 2.

### **Sampling method**

The set of numerical simulations performed was defined using a full factorial design with interactions and resulted in 33 simulations ( $n = 2^5 + 1$ ).

### **Selection of metrics from the FE analysis**

Two types of metrics were studied in order to capture the non-linearity of the response. The first one is the mean slope of the force/displacement curve. The second one is the mean curvature of the force/displacement curve. For each metrics, 3 parts of the force/displacement curve were analyzed to study the influence of the parameters at different stages of the compression test: from 0 mm to 5 mm; from 5 mm to 20 mm; from 20 mm to 30 mm (Figure 5 - A).

### **Response surface definition**

A response surface was then fitted on the metrics values obtained from the 33 numerical simulations. This surface response was modelled with a first order polynomial (Equation 4) including interactions (5 studied parameters coefficients + 10 interaction coefficients + 1 constant term).

$$Y = \beta_0 + \sum_{i=1}^p \beta_i \times x_i + \sum_{\substack{i,j=1 \\ j>i}}^p \beta_{ij} \times x_i \times x_j \quad \text{Equation 4}$$

where  $p = 5$  is the number of parameters;  $Y$  is the output of the model and  $x$  the parameter value in its coded space. Except for  $\beta_0$ , the absolute value of the coefficients  $\beta$  will reflect the sensitivity of the FE model to the corresponding parameters.

### **Statistical analysis of the response surface**

To assess the quality of the fitted response surface, a statistical analysis was used. After ensuring that the outputs of the 33 simulations followed a Normal law, an analysis of

1 variance (ANOVA) was performed [23]. The sum of squares due to the error (SSE) and  
 2 the sum of squares due to the regression (SSR) were calculated as follows:

$$SSE = \sum_{i=1}^n (Y_i - \hat{Y}_i)^2 \quad \begin{array}{l} 3 \\ 4 \end{array} \quad \text{Equation 5}$$

$$SSR = \sum_{i=1}^n (\hat{Y}_i - \bar{Y}_i)^2 \quad \begin{array}{l} 5 \\ 6 \end{array} \quad \text{Equation 6}$$

7 where  $n = 33$  is the number of simulations;  $Y_i$  are the outputs of the simulation;  $\bar{Y}_i$  are  
 8 the means of the outputs and  $\hat{Y}_i$  are the estimated outputs computed from the fitted  
 9 model.

10 From these values the mean sum of squares due to the error (MSE) and the mean sum  
 11 of squares due to the regression (MSR) were deduced. Then, the linear coefficient of  
 12 regression was calculated.  
 13

$$MSE = \frac{SSE}{n - p} \quad \begin{array}{l} 14 \\ 15 \end{array} \quad \text{Equation 7}$$

$$MSR = \frac{SSR}{p - 1} \quad \begin{array}{l} 16 \\ 17 \end{array} \quad \text{Equation 8}$$

$$R^2 = \frac{SSR}{SST} \quad \begin{array}{l} 18 \\ 19 \end{array} \quad \text{Equation 9}$$

20 where  $n = 33$  is the number of simulations and  $p = 5$  the number of parameters.

21  
 22 The global validity of the model was then assessed by applying a Fisher test with  $(p - 1)$   
 23 and  $(n - p)$  degrees of freedom to the following quantity:

$$T^{model} = \frac{MSR}{MSE} \quad 25 \quad \text{Equation 10}$$

24

25 Finally, the significance of the sensitivity of each parameters with respect to the  
 26 experimental variance,  $\hat{\beta}_i$ , was assessed by applying a Student test with  $(n - p)$  degrees  
 27 of freedom to the quantity:  
 28

$$T^{parameter} = \frac{\hat{\beta}_i}{\sqrt{Var(\beta_{experimental})}} \quad \begin{array}{l} 29 \\ 30 \end{array} \quad \text{Equation 11}$$

31 The results of this design of experiments applied to the FE model are presented in the  
 32 results section.

33

#### 34 **4. Identification method**

35 To identify the material parameters of both constitutive models ( $C_{10}^H$  for Equation 1;  $C_{10}^P$   
 36 and  $C_{20}$  for Equation 2), an inverse method was implemented.

37

38 Several steps were necessary:

- 1 1. Extraction of force-displacement curves from the bi-dimensional model.
- 2 2. Definition of a cost function quantifying the mismatch between numerical and
- 3 experimental force-displacement curves.
- 4 3. Minimization of the cost function with the genetic algorithm implemented in
- 5 Matlab<sup>®</sup>.

6  
7 To estimate the mismatch between numerical and experimental force-displacement  
8 curves, the reaction forces from the numerical simulations and the experiments were  
9 sampled on an identical displacement range and the cost function was defined in the  
10 least square sense:

$$E = \sum_{i=1}^l (Y_{sim}(i) - Y_{exp}(i))^2 \quad \text{Equation 12}$$

14 where  $l$  is the number of points,  $Y_{sim}$ , the reaction forces predicted by the numerical  
15 simulation and  $Y_{exp}$ , the obtained experimental reaction forces.

16  
17 The stopping criterion of the genetic algorithm was the tolerance on the cost function,  
18 which was set to  $10^{-3}$ . Reaction forces were of the order of magnitude  
19  $10^0$ .

### 3. Results

#### 1. FE Model sensitivity

A design of experiments approach was used to evaluate the sensitivity of the FE model to different parameters. The aim was to ensure that this simulation can be used to identify deep soft tissue mechanical parameters (*i.e.* that the response of the simulation is sensitive enough to deep soft tissue mechanical parameters).

#### **Statistical analysis of the response surface**

The results of the ANOVA test concerning the surface response evaluation (Equation 5 to Equation 11) are reported in Table 3. The coefficients of determination,  $R^2$ , are above 99% for all tested metrics except for the mean slope for depth of compression in the 20-30 mm range. This indicates that the first order surface response (Equation 4) provides a very good agreement with the simulation sets.

#### **Parameters sensitivity**

The 1<sup>st</sup> order polynomial coefficients (Equation 4) calibrated against the mean curvature values are shown in Figure 5 – B. They represent the sensitivity of the FE model response to each parameter: the higher the absolute value of the coefficient, the more sensitive the FE model response to a change in this parameter. These coefficients were derived from simulations at 3 different stages of the force/displacement curves. Those adjusted on the mean slope are shown in Figure 5 – C. Coefficients characterizing the combined effects of the different parameters ( $\beta_{ij}$  - Equation 4) were derived but they are not significant, so they are not reported here.

The values of the sum of squares due to the regression are higher for the second and third parts of the force/displacement curves, for all studied metrics. This indicates that the identification of the material parameters of the constitutive equations for deep soft tissues is more relevant as the compression progresses.

For both types of analyzed metrics (*i.e.* mean curvature and mean slope of the force/displacement curves), the sensitivities are significantly higher for the material properties of the deep soft tissues, which means that they have a greater impact on the response.

This result confirms that the proposed combination of experimental and numerical approaches permits identifying the mechanical properties of *deep* soft tissues.

1 **2. Identified material properties of 4 subjects**

2 **Experimental data**

3 For the four subjects, the responses to a 25 mm deep localized compression with a 30  
4 mm diameter cylinder were acquired according to the methodology described before.  
5 The results are shown in Figure 6.

6

7 **Identification: neo-Hookean strain energy density function**

8 An example of the experimental and numerical force/displacement curves after  
9 identification of the  $C_{10}^H$  material parameter (Equation 1) is shown in Figure 7 – A.

10 The results of the identified parameter  $C_{10}$  for 4 subjects and the related cost function  
11 values are reported in Table 4.

12

13 **Identification: 2nd order reduced polynomial strain energy density function**

14 An example of the experimental and numerical force/displacement curves after  
15 identification of  $C_{10}^P$  and  $C_{20}$  (Equation 2) is shown in  
16 Figure 7 – B.

17 The  $C_{10}$  and  $C_{20}$  values for four subjects and the related cost function values are  
18 reported in Table 5.

19

## 4. Discussion

### 1. *Contribution of the study*

The methodology developed in this study is a fast and efficient tool to identify the passive mechanical properties of deep soft tissues in the leg of several subjects, taking into account the non-linear behavior of these tissues. In the literature, only a few non-invasive methods have already been developed to identify the passive mechanical properties of deep soft tissues and it is the first time that a thorough analysis is conducted to really separate the contribution of deep and superficial tissues in the response to compression tests or indentation tests.

A method combining an axisymmetric multi-layer (*i.e.* skin, adipose tissue) FE model with an indentation experiment was used to identify the mechanical properties of the forearm soft tissues [15]. A small indenter allowed recruiting the behavior of superficial soft tissues only. An interesting point of this technique was to identify separately each type of soft tissues.

Also, the characterization of the soft tissues of the leg has been performed for several subjects [5], [16]. FE models of the leg in a relaxed initial state and a compressed state with stocking were compared to MRI scans of the legs in the same two states. The identification was performed between two loading states and could not capture the non-linear behavior of soft tissues. However, the FE model was composed of two compartments of soft tissues, allowing a distinction between the mechanical properties of adipose tissue and muscular tissue.

Finally, a study similar to the one presented here identified mechanical properties of soft tissues of the calf [24]. An inverse method combined a subject-specific FE model with the indentation of the leg at the calf location by 8 mm or 19 mm diameters cylinders. In this case, the cylinders were applied perpendicularly to the leg axis. Nevertheless, this FE model was composed of only one compartment of soft tissues, without any distinction between skin, fat and muscle.

### 2. *Limitations of the experiments*

The leg geometry in the FE model was defined from the ultrasound reconstruction and might differ from the actual leg geometry during the acquisition of the compression data. Indeed, both data sets were not acquired on the same day but also, the leg position differed between both data acquisitions: prone position for the ultrasound image acquisition; sitting position for the compression test.

The effects of these geometrical differences and their impact on the identification of mechanical properties still need to be investigated. This matter could be prevented if the leg geometry acquisition was performed right before the compression of the calf soft

1 tissues, with a 3D optical scanner for example. A 3D scanner will provide the external  
2 geometry of the leg, but also the exact location of the cylinder before the compression  
3 as well as the direction of the cylinder displacement. Nevertheless, the bones location  
4 and the different soft tissue compartments are not available with this imaging modality.  
5 As the aim of the study was to identify deep soft tissues mechanical properties, it would  
6 have been impossible to use the 3D scanner to implement the simulation, as it only  
7 provides the external geometry of the leg.

### 8 9 **3. Numerical assumptions**

10 The subject-specific FE model of the leg implemented in this study is based on several  
11 assumptions discussed here.

#### 12 13 ***Bi-dimensional FE model***

14 The identification process compares the 3D experimental curves with that of the bi-  
15 dimensional FE model. The bi-dimensional FE model is built on a classical plane-strain  
16 model combined with correction factors taking into account 3D effects. As a matter of  
17 fact, a plane-strain model would be insufficient, the 3D effects corresponding to about  
18 15% of the reaction force (Figure 4 – C). This reduction of the 3D model may be  
19 questionable for small displacements. Indeed, in the first millimeters of the localized  
20 compression, the contact length is smaller than the length of the applied cylinder  
21 because the leg is not a perfect cylinder. Not surprisingly, the design of experiments  
22 showed that the identification of material parameters is more significant for a  
23 displacement larger than 5 mm.

#### 24 25 ***Material compressibility***

26 It is fairly common to assume soft tissues as nearly incompressible *in vivo* because they  
27 are mainly composed of water [8]. However, a preliminary study showed that a fully  
28 incompressible strain energy density function raised numerical issues. Consequently, a  
29 slightly compressible model was chosen and D values for both constitutive equations  
30 (Equation 1 and Equation 2) were selected from a previous study [16]. In the latter study,  
31 a difference of leg cross-section area of about 5% was observed in MRI transverse  
32 scans between wearing and not wearing compression stockings (Class II delivering  
33 averagely a 20 mmHg pressure to the calf).

34 Two phenomena could explain this observation of tissue loss of volume. First, when the  
35 leg is under compression, the venous system is drained compared to the uncompressed  
36 state and second, soft tissues can move upwards when a compression stocking is worn.  
37 Thus, the loss of area occupied by the leg in the MRI transverse planes could be the  
38 combined result of blood draining and 3D deformations. Assigning the D values reported  
39 in Table 1 was sufficient to take these effects into account in the models. An

1 improvement could be to identify a subject-specific compressibility parameter, but this  
2 would require precise volume measurements of soft tissues during the test.

### 3 4 **Viscoelasticity**

5 The methodology described in this paper presents the identification of soft tissues elastic  
6 properties. However, human soft tissues are often considered as viscoelastic materials  
7 [25]. The experimental setup was designed to compress soft tissue with a very low  
8 displacement rate. It was checked that displacement rate variation (within the range  
9 being possible with this setup) had negligible influence on the response. This confirms  
10 that for the timescales of the experiments, viscoelastic effects remain negligible and  
11 hence an identification on the loading curve is convenient.

## 12 13 **4. Design of experiments**

14 Several studies raised questions about the mechanical role of the *fascia cruris* in the  
15 biomechanics of elastic compression, one of them being the contention of muscular  
16 tissues [26], [27]. However, the sensitivity analysis performed here showed no significant  
17 effect of its mechanical properties or of its thickness. It is still possible that the *fascia*  
18 *cruris* plays a biomechanical role but that this role is marginally involved in the localized  
19 compression applied here. Other investigations should be proposed in the future to  
20 answer the questions regarding the biomechanical role of this tissue and how it  
21 participates to the response in other modes of loading.

## 22 23 **5. Identified material parameters**

24 Material parameters of a neo-Hookean strain energy density function have already been  
25 identified for the soft tissues of the human leg [5], [16]. In those studies,  $C_{10}$  was  
26 identified between 1 kPa and 15 kPa. Here, the  $C_{10}^H$  are between 0.5 kPa and 2.1 kPa.

27 Although material properties of the *fascia cruris* have no significant impact on the  
28 response of the current FE model, the presence of this tissue seems to stiffen the leg,  
29 likely due to a structural effect. This may explain why the identified values of  $C_{10}^H$   
30 identified here are in the lower range of the values found in the literature [5], [16].

31  
32 Due to the large deformations of the leg during localized compression, the neo-Hookean  
33 strain energy density function appeared limited since it could not capture the non-linear  
34 behavior observed experimentally (Figure 7 – A). However, a 2<sup>nd</sup> order reduced  
35 polynomial equation described well the behavior of deep soft tissues of the leg (Figure 7  
36 – B).

37  
38 In this bi-dimensional FE model, muscular tissues were considered as transversally  
39 isotropic, which is fairly common when assuming muscular fibers to be distributed  
40 perpendicular to the transversal plane. In the future, it would be interesting to investigate



1 localized compression in several directions to study the anisotropic behavior of muscular  
2 tissue.  
3 Also, the identified mechanical properties here were those of a relaxed muscular state.  
4 Other investigations with a controlled muscular contraction would allow identifying the  
5 mechanical properties at an active muscular state.  
6 Finally, this methodology applied to a larger population could shed a light on inter-  
7 subject variability.

8

## 9 **5. Conclusion**

10 This paper presented a fast and non-invasive novel methodology to identify the  
11 mechanical properties of deep soft tissues of the human leg, using conventional  
12 ultrasound imaging and a simple mechanical test.

13 To do so, an inverse method was used, combining a subject-specific bi-dimensional FE  
14 model with localized compression tests. After the evaluation of this method on one  
15 healthy subject, it was performed on three other subjects to identify the material  
16 parameters of two constitutive equations of the deep soft tissues. The methodology  
17 takes into account the non-linear behavior of deep soft tissues by identifying the material  
18 properties on a large range of deformations rather than comparing two states (*i.e.* initial  
19 and final). This study showed that a 2<sup>nd</sup> order reduced polynomial strain energy density  
20 function should be used over a neo-Hookean model for deep soft tissues of the leg.  
21 Also, the differentiation of materials (*i.e.* skin, fat, *fascia cruris*, and muscle) allowed  
22 separating the contributions of each tissue. It is the first time that a thorough analysis is  
23 conducted to really separate the contribution of deep and superficial tissues in the  
24 response to compression tests and this permitted to estimate the passive material  
25 properties of deep soft tissues for four subjects independently of the response of their  
26 other tissues.

27

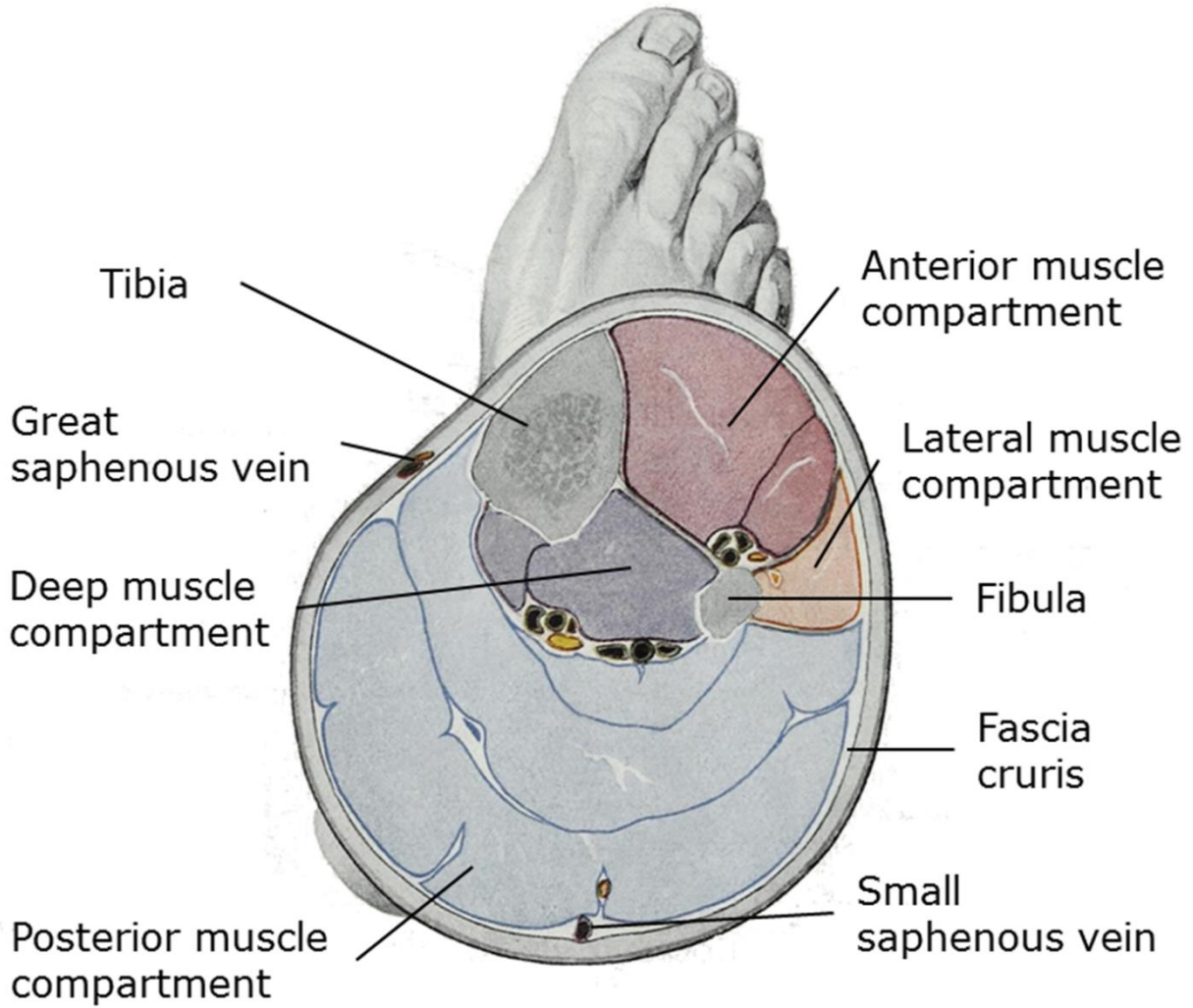
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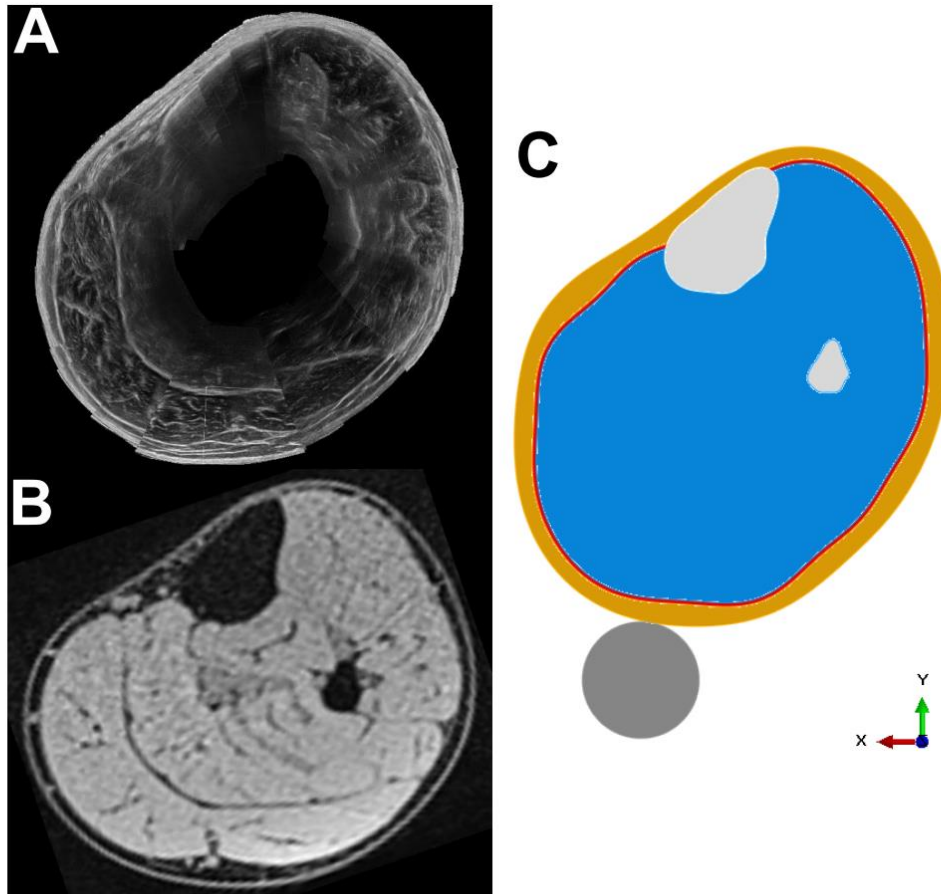
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1  
2 **Figure 1: Anatomy of the human leg – Cross-section of the human right leg. The**  
3 ***fascia cruris*** makes a separation between the superficial tissues, composed by  
4 the adipose tissue, the skin and the superficial veins, and the deep tissues,  
5 composed by the muscular compartments and the deep veins. Adapted from [28].



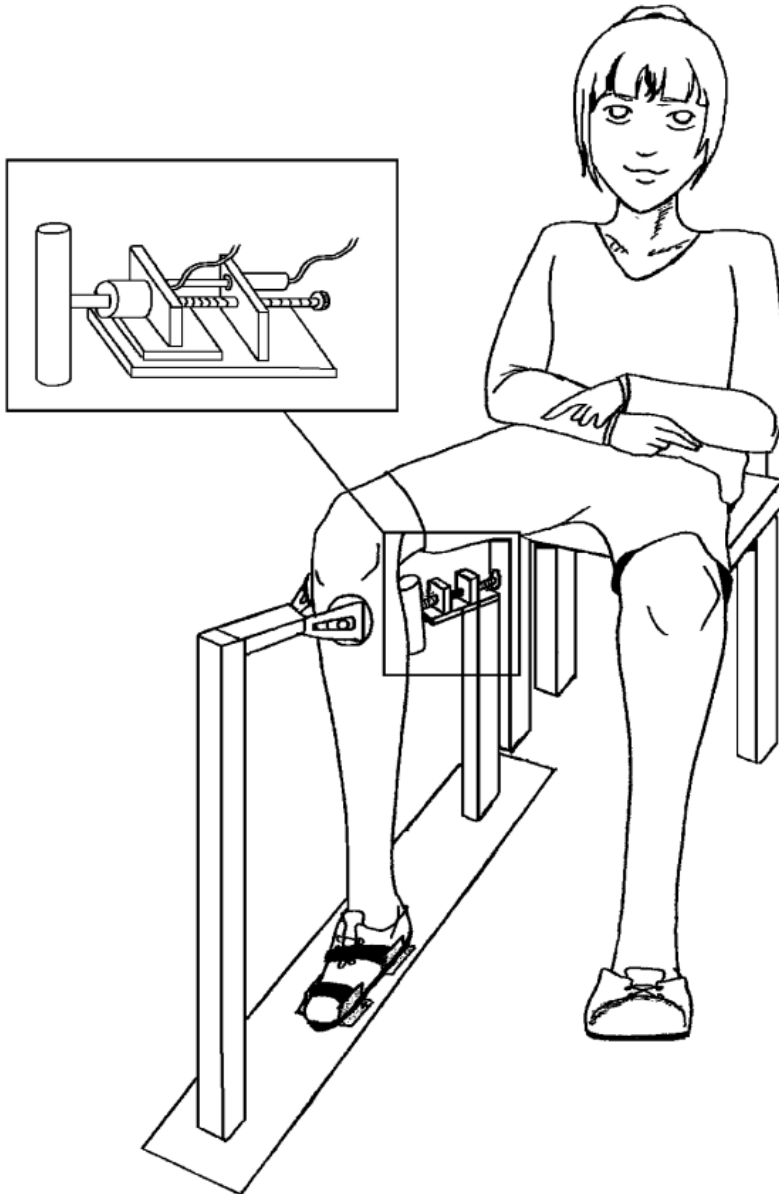
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1 **Figure 2 – Finite element (FE) model implementation – B-mode ultrasound**  
2 **reconstruction of the leg (A), MRI of the leg (B), bi-dimensional FE model (C)**  
3 **composed by the bones (tibia and fibula); the deep soft-tissue compartment, the**  
4 **superficial soft-tissues compartment and the *fascia cruris* separating them.**  
5  
6



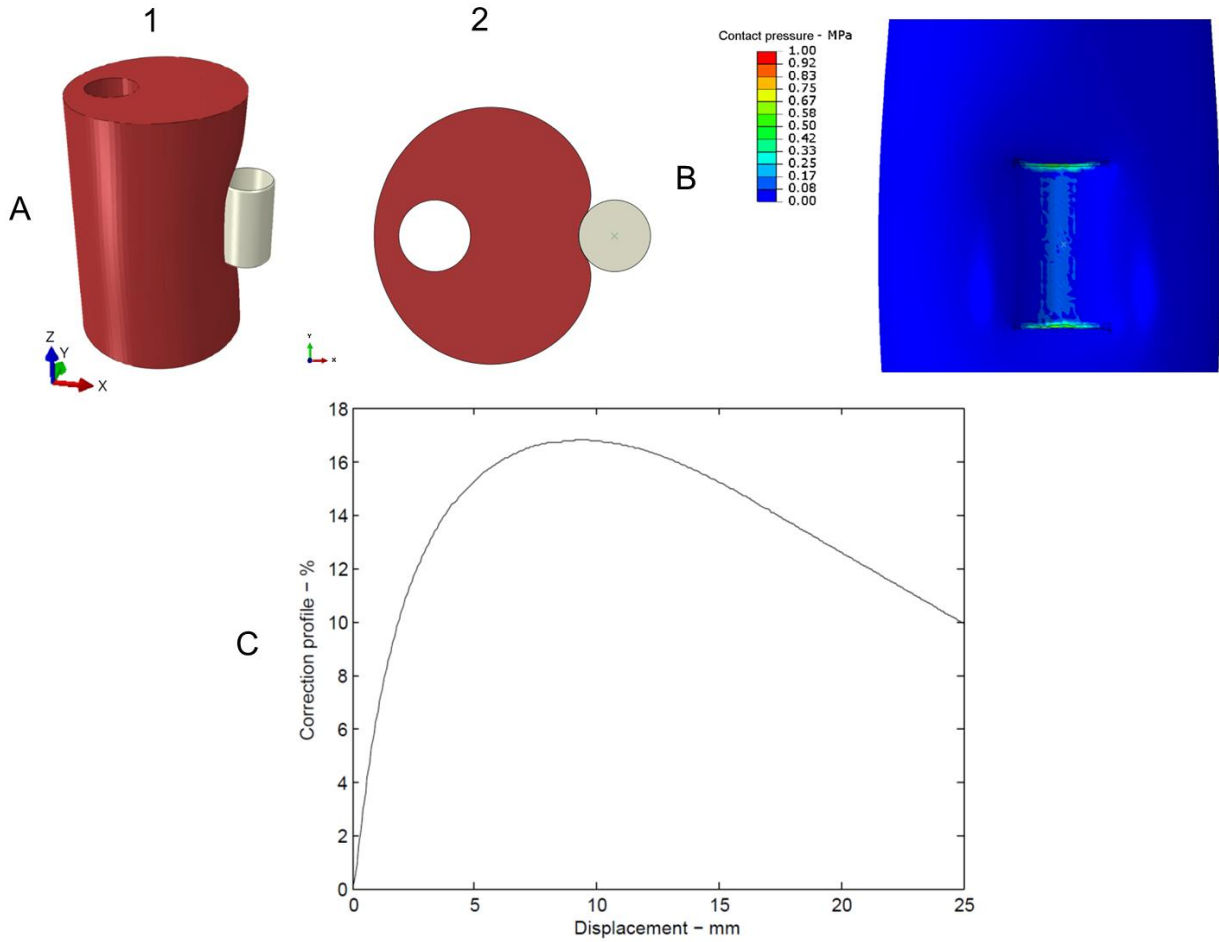
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1 **Figure 3 – Experimental setup – The subject is seated on a chair. The straps**  
2 **around the foot and the rubber buffers behind the knee lead to minimum motion**  
3 **of the leg during data acquisition. A 30 mm diameter cylinder is applied parallelly**  
4 **to the leg axis. A force sensor and a displacement sensor record the desired data.**  
5  
6



7

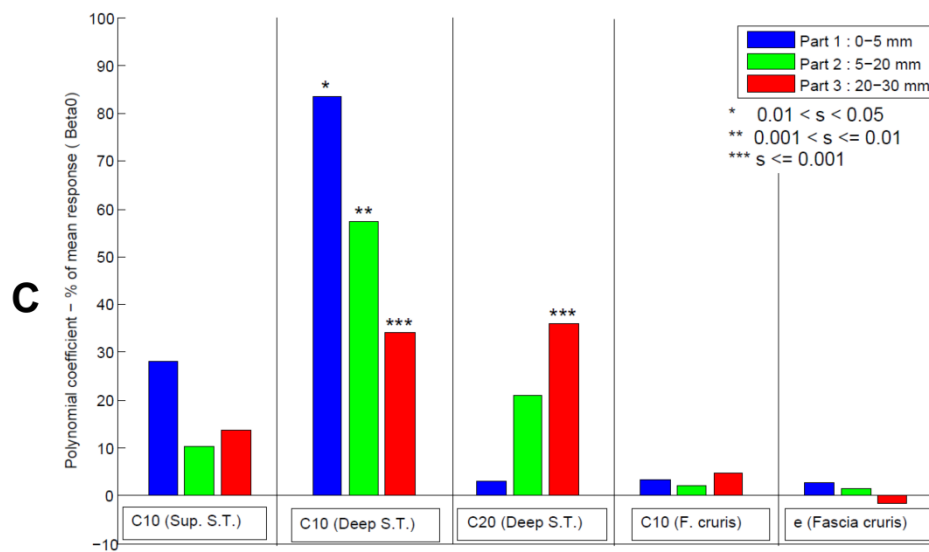
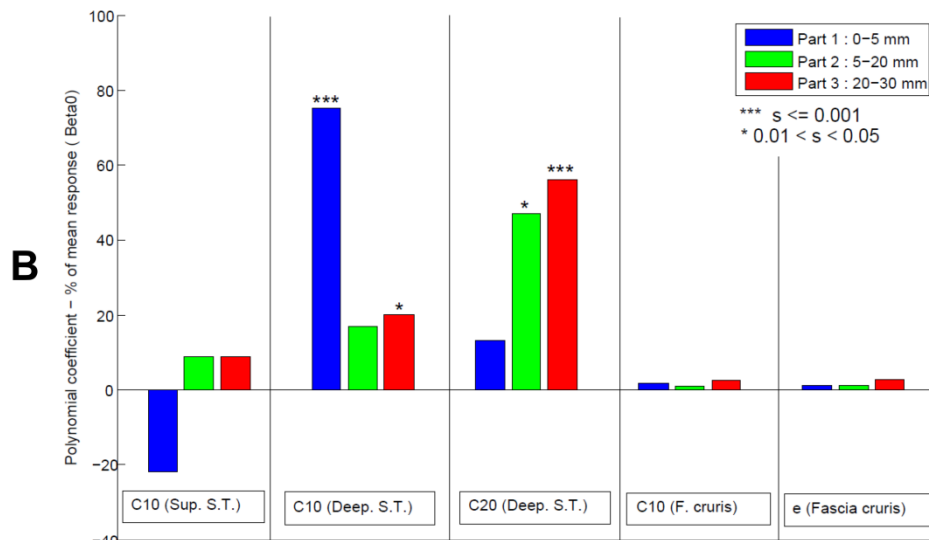
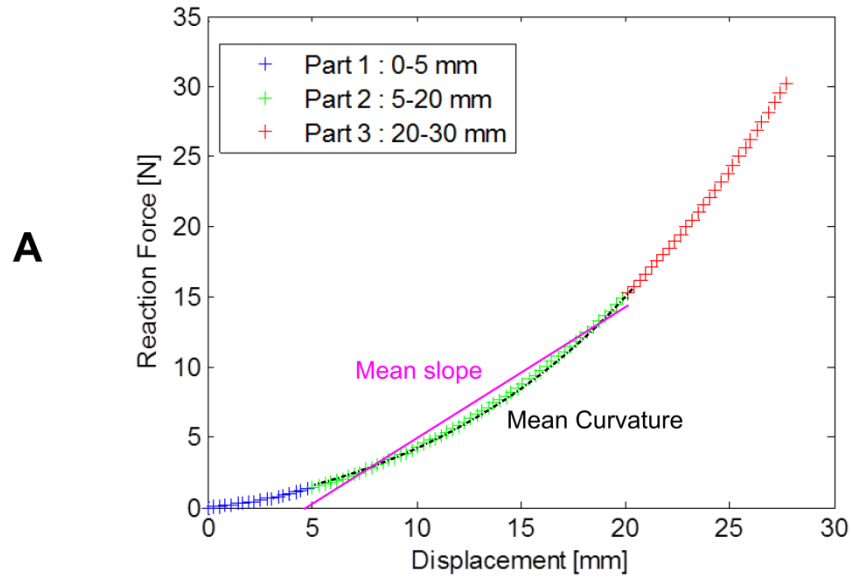
1 **Figure 4 – A –Computations of an idealized leg under a localized compression**  
 2 **using a 30 mm diameter cylinder of different lengths. 1: 3 dimensional simulation;**  
 3 **2: bi-dimensional simulation. B – Pressure distribution applied onto the leg by the**  
 4 **cylinder. C – Function defined by Equation 3 to compensate the underestimation**  
 5 **of the reaction forces made with a 2D plane-strain model.**  
 6



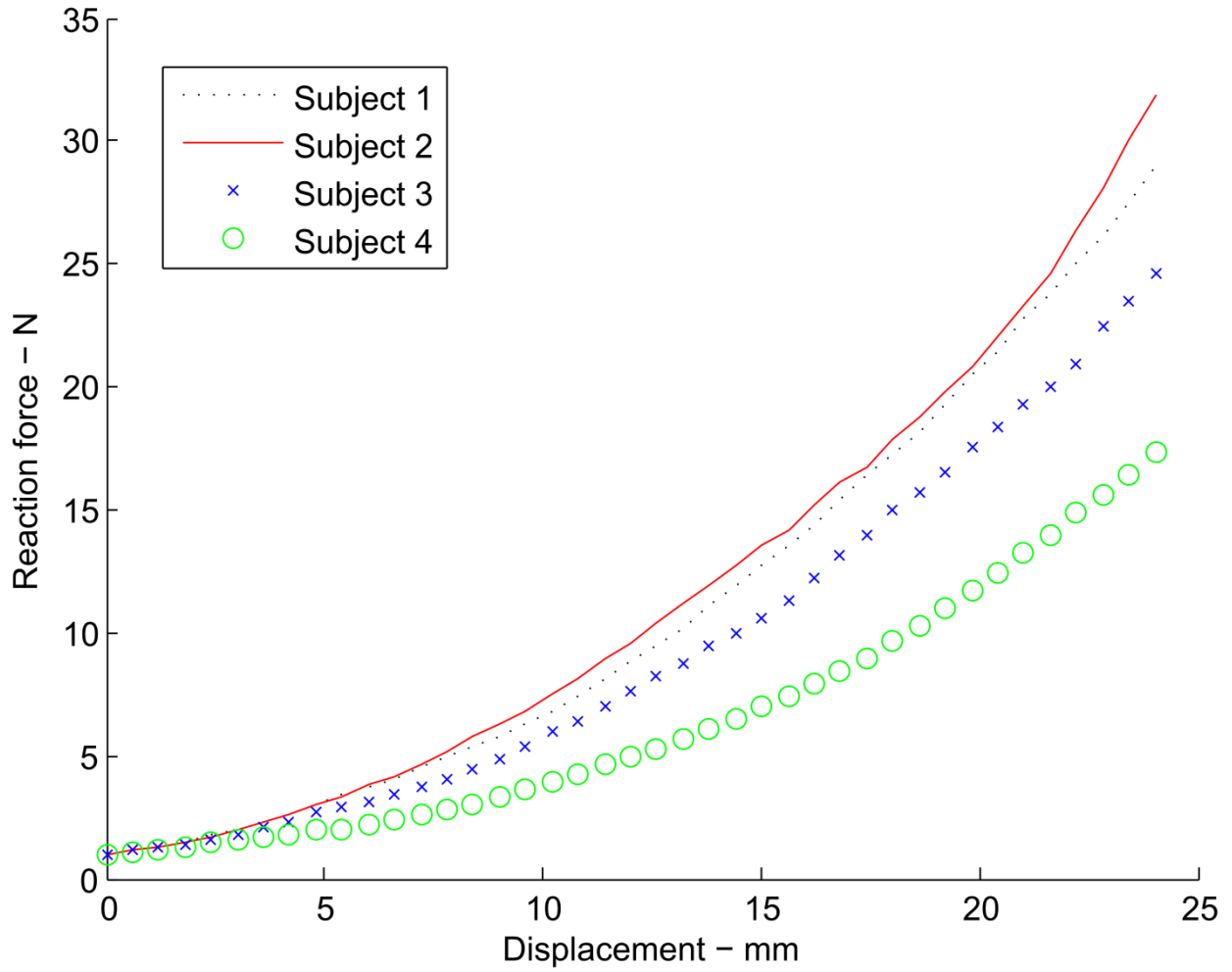
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1 **Figure 5 – Illustration of a force/displacement curve (A) - Sensitivities – 2<sup>nd</sup> order**  
2 **polynomial coefficients adjusted on the model for the mean curvature response**  
3 **(B) and mean slope response (C) on the 3 stages of the force/displacement curve.**  
4 **From left to right:  $C_{10}^{\text{Superficial soft tissues}}$ ,  $C_{10}^{\text{Deep soft tissues}}$ ,  $C_{20}^{\text{Deep soft tissues}}$ ,  $C_{10}^{\text{Fascia cruris}}$ ,**  
5 **Thickness  $^{\text{Fascia cruris}}$ . s stands for significance.**  
6  
7



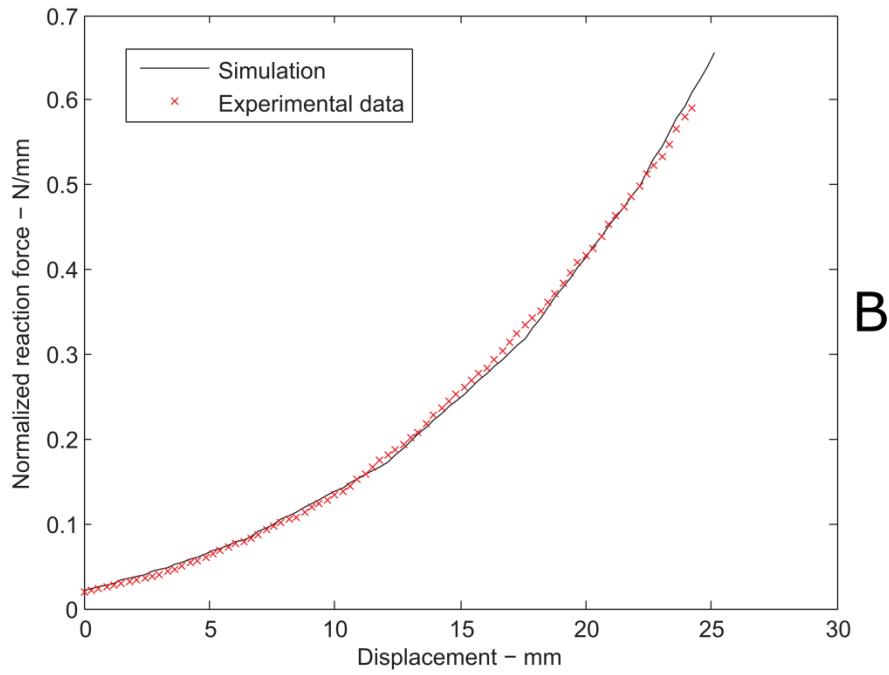
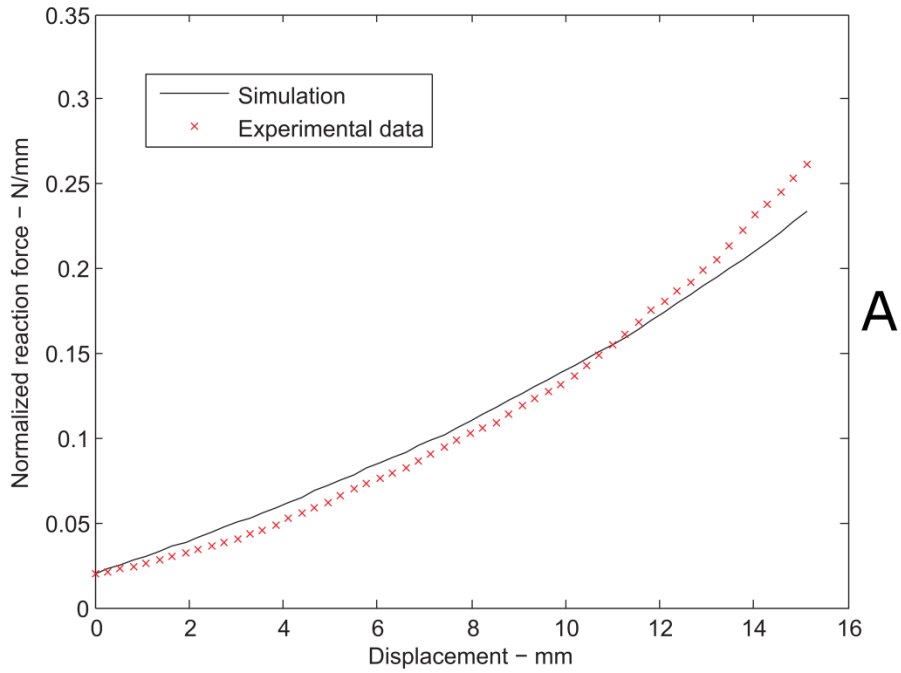


1 **Figure 6 – Experimental data – Response of 4 subjects to a localized compression**  
2 **of 25 mm depth with a 30 mm diameter cylinder.**  
3



4

1 **Figure 7 – Numerical (solid line) and experimental (dash line) force/displacement**  
2 **curves for one subject. A – Result with the neo-Hookean constitutive equation. B**  
3 **– Result of the 2nd order reduced polynomial constitutive equation.**



4

| Material                 | Behavior law                           | Parameters   | Source             |
|--------------------------|--|--|--------------------|
| Deep soft tissues        | Hyper-elastic<br>neo-Hookean           | $C_{10}$<br>$D_1 = 28 \text{ MPa}^{-1}$              | Identified<br>[16] |
|                          | Hyper-elastic<br>reduced<br>polynomial | $C_{10}, C_{20}$<br>$D_1, D_2 = 28 \text{ MPa}^{-1}$ | Identified<br>[16] |
| Superficial soft tissues | Hyper-elastic                          | $C_{10} = 2 \text{ kPa}$                             | [16]               |
|                          | neo-Hookean                            | $D_1 = 22.5 \text{ MPa}^{-1}$                        | [16]               |
| <i>Fascia cruris</i>     | Hyper-elastic                          | $C_{10} = 100 \text{ kPa}$                           | [29]               |
|                          | neo-Hookean                            | $D_1 = 22.5 \text{ MPa}^{-1}$                        | [16]               |

1 **Table 1 – Material properties of the biomechanical model**  
2

| Parameter                                  | Domain               |
|--|----------------------|
| $C_{10}^{\text{Deep soft tissues}}$        | From 0.5 to 30.0 kPa |
| $C_{20}^{\text{Deep soft tissues}}$        | From 0.5 to 30.0 kPa |
| $C_{10}^{\text{Superficial soft tissues}}$ | From 1.0 to 30.0 kPa |
| $C_{10}^{\text{Fascia cruris}}$            | From 0.1 to 10.0 MPa |
| Thickness <sup>Fascia cruris</sup>         | From 0.2 to 1.0 mm   |

1 **Table 2 – Parameters studied in the sensitivity analysis**  
2

| Indicator                  | Regression            |                      | Error                |                      | R <sup>2</sup> |
|----------------------------|-----------------------|----------------------|----------------------|----------------------|----------------|
|                            | Sum of square         | Mean of square       | Sum of square        | Mean of square       |                |
| Curvature from 0 to 5 mm*  | 9.40.10 <sup>-5</sup> | 6.3.10 <sup>-6</sup> | 6.7.10 <sup>-7</sup> | 4.0.10 <sup>-8</sup> | 0.9929         |
| Curvature from 5 to 20 mm* | 3.2.10 <sup>-5</sup>  | 2.1.10 <sup>-6</sup> | 1.2.10 <sup>-7</sup> | 6.9.10 <sup>-9</sup> | 0.9964         |
| Curvature from 20 to 30mm* | 1.8.10 <sup>-4</sup>  | 1.7.10 <sup>-5</sup> | 3.2.10 <sup>-7</sup> | 1.9.10 <sup>-8</sup> | 0.9982         |
| Slope from 0 to 5 mm*      | 2.2.10 <sup>-2</sup>  | 1.5.10 <sup>-3</sup> | 2.1.10 <sup>-4</sup> | 1.2.10 <sup>-5</sup> | 0.9907         |
| Slope from 5 to 20 mm*     | 5.7.10 <sup>-2</sup>  | 3.8.10 <sup>-3</sup> | 1.1.10 <sup>-4</sup> | 6.7.10 <sup>-6</sup> | 0.9980         |
| Slope from 20 to 30 mm*    | 0.19                  | 1.3.10 <sup>-2</sup> | 1.4.10 <sup>-2</sup> | 8.0.10 <sup>-4</sup> | 0.9336         |

Fisher's test probability: \*≤0.001

1 **Table 3 – Statistical analysis (ANOVA) of the response surface**

2

| Subject | $C_{10}$ identified<br>(kPa) | Cost function value<br>(N <sup>2</sup> ) |
|---------|------------------------------|--|
| 1       | 2.11                         | 0.45                                     |
| 2       | 1.72                         | 0.48                                     |
| 3       | 1.21                         | 0.23                                     |
| 4       | 0.51                         | 0.16                                     |

2 **Table 4 – Identification of  $C_{10}$ <sup>Deep soft tissues</sup> for 4 subjects**  
3

| Subject | C <sub>10</sub> identified<br>(kPa) | C <sub>20</sub> identified<br>(kPa) | Cost function value<br>(N <sup>2</sup> ) |
|---------|-------------------------------------|-------------------------------------|--|
| 1       | 0.82                                | 2.61                                | 6.5.10 <sup>-3</sup>                     |
| 2       | 0.80                                | 2.00                                | 2.5.10 <sup>-2</sup>                     |
| 3       | 0.70                                | 0.80                                | 3.5.10 <sup>-1</sup>                     |
| 4       | 0.19                                | 0.24                                | 4.3.10 <sup>-1</sup>                     |

1 **Table 5 – Identification of C<sub>10</sub><sup>Deep soft tissues</sup> and C<sub>20</sub><sup>Deep soft tissues</sup> for 4 subjects**