

Purification of organic acids by chromatography with strong anionic resins: Investigation of uptake mechanisms

Julien Lemaire, Claire-Line Blanc, Florence Lutin, Marc-André Theoleyre, Moncef Stambouli, Dominique Pareau

▶ To cite this version:

Julien Lemaire, Claire-Line Blanc, Florence Lutin, Marc-André Theoleyre, Moncef Stambouli, et al.. Purification of organic acids by chromatography with strong anionic resins: Investigation of uptake mechanisms. Journal of Chromatography A, 2016, 1458, pp.63-69. 10.1016/j.chroma.2016.06.057 . hal-01363370

HAL Id: hal-01363370 https://hal.science/hal-01363370v1

Submitted on 9 Nov 2016

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1	ORGANIC ACIDS CHROMATOGRAPHY WITH STRONG ANIONIC RESINS
2	
3	
4	
5	
6	
7	Purification of organic acids by chromatography with strong anionic resins:
8	investigation of uptake mechanisms
9	
10	Julien Lemaire ^{a*} , Claire-Line Blanc ^{a,b} , Florence Lutin ^b , Marc-André Theoleyre ^a ,
11	MoncefStambouli ^a , Dominique Pareau ^a
12	^a LGPM, CentraleSupélec, Université Paris-Saclay, SFR Condorcet FR CNRS 3417, Centre Européen
13	de Biotechnologie et de Bioéconomie (CEBB), 3 rue des Rouges Terres 51110 Pomacle, France
14	^b Eurodia Industrie, ZAC Saint Martin, Impasse Saint Martin, 84120 Pertuis, France
15	
16	* Corresponding author.
17	Present address: CentraleSupélec, CEBB, 3 rue des Rouges Terres 51110 Pomacle,
18	France.
19	Tel.: +33(0) 352 620502.
20	Email address: julien.lemaire@centralesupelec.fr

- 1 Abstract
- 2

Bio-based organic acids are promising renewable carbon sources for the chemical
industry.However energy-consuming purification processes are used, like distillation or
crystallization, to reach high purities required in some applications. That is why preparative
chromatography was studied as an alternative separation technique. In a previous work
dealing with the purification of lactic, succinic and citric acids, theLangmuir model was
insufficient to explain the elution profiles obtained with a strong anionic resin.

9 Consequentlythe Langmuir model was coupled with a usual ion-exchange model to take 10 into account the retention of their conjugate bases (<2%), which are commonly neglected at 11 low pH (< 1.5). Elution simulationswith both uptake mechanisms fitted very well with 12 experimental pulse tests. Only two parameters were optimized (equilibrium constant of acid 13 uptake and ion-exchange selectivity coefficient of conjugate base)and their value were 14 coherent with experimental and resin suppliers' data.

These results confirmed that the singular tailing and apparent delay observed with succinic and citric acids can be explained by the high affinity of succinate and citrate for resin cationic sites. Themodel was implemented in a preparative chromatography simulation program in order tooptimize operating parameters of our pilot-scale ISMBunit (Improved Simulated Moving Bed). The comparison with experimental ISMB profiles was conclusive. 1 **Keywords:**chromatography; modelling;anionic resin; organic acid;sustainable processing.

2

3 Highlights

- Uptake mechanisms of organic acids with strong anionic resin were investigated.
- Langmuir model coupled with ion-exchange model fitted with experimental data.
- Singular peak tailing and delay were better explained, particularly for citric acid.
- Retention of someorganic acids in the dissociated form cannot be neglected.

1 1. Introduction

2 In the frame of sustainable development, biomass valorization as a renewable carbon 3 source for the chemical industry is growing. Indeed, fossil resources are running out in the 4 medium to long term. Moreover carbon balances have to be improved so as to reduce the 5 CO₂ level in the atmosphere, whose rapid rise is linked to global warming. In the famous 6 report of the Pacific Northwest National Laboratory (PNNL) and the National Renewable 7 Energy Laboratory (NREL)[1], bio-based organic acids are identified as very promising 8 sources of renewable carbon. Many compounds can be produced from these moleculesinthe 9 chemical industry, particularly in the polymer industry. Up to now, the production of bio-based 10 organic acids keeps small because they are stilluncompetitive compared to the 11 corresponding petroleum-derived products[2]. That is why the reduction of their production 12 cost is a major concern of the chemical industry.

13 Bio-based organic acids are usually produced by microbial fermentation for the food 14 industry but further purification techniques are required for others applications which need a 15 much higher purity, particularly polymer synthesis. Currently, distillation [3-4], esterification 16 [5-7]and crystallization [8]are commonly used to reach very high purities (> 99.5%)[9]. 17 However, these techniques are energy-consuming and give quite low recovery 18 rates.Preparative chromatography is a promising solution for downstream processes 19 because its energy, chemical and water consumptions are generally quite lower and 20 consequently effluent volumes too. Initially developed for extraction and purification of 21 complex vegetal mixtures [10], this separation method is spreading in different fields: 22 biotechnology [11-12], pharmaceutic[13-14] and the sugar industry [15], for instance. Indeed, 23 the same separation quality may be reached with better yields, lower costs and impacts on 24 the environment. However, for several years now, the state of the world economy has not 25 been favorable to industrial investments for theimprovement of downstream purification 26 processes. Moreover, chromatography needs to prove itselffor high purity application (> 27 99.5%) at the industrial scale.

1 In the frame of organic acids purification in fermentation broths, chromatography is often 2 performed with ion-exchange resinsto separate mineral salts, sugars and organic acids. The 3 eluent is an aqueous solution whose pH can be adjusted with the inorganic acid or base 4 corresponding to the counter-ions fixed on the resin so as to keep its composition constant. 5 The separation of compounds is based on affinity differences for the resins. Higher is the 6 affinity, slower migrates the compound along the resin bed. In the sugar industry, cationic 7 resins are used to separate sucrose from saltsas well as sugars from each other. The 8 glucose-fructose separation is among the most famous applications of preparative 9 chromatography. Otherwise, anionic resins are performed for the citric acid purification. 10 Uptake and slowdown mechanisms are not well known and complex, coupling size 11 exclusion, ligand exchange, hydrophilic or hydrophobic interactions and ion-exchange[16-12 17].

13 In a previous work dealing with the purification of lactic, succinic and citric acids by 14 chromatography with a strong anionic resin (quaternary ammonium), the Langmuir model 15 fitted well with the experimental equilibrium isotherms obtained by frontal analysis[18]. 16 However, the corresponding affinity coefficients were significantly lower than those obtained 17 by pulse tests for succinic and citric acids. Their peaks had a singular tailing and were 18 delayedcompared to simulated ones with the Langmuir model and experimental coefficients 19 of isotherms. Indeed, the Langmuir model was insufficient to describe the organic acids 20 retention on the strong anionic resin and frontal analysis gave underestimated affinity 21 coefficients.

These results lead us to suspect the impact of conjugate bases (dissociated organic acids) retention by ion-exchange, despite their very low concentrations. Indeed, the citrate anion affinity is high for quaternary ammonium anionic resin according to selectivity coefficients given by resins producers. Usually, this uptake mechanism is not taken into account because the strong anionic resins are performed at such acid pH (1 to 1.5) that only 0.2% to 0.4% of lactic acid, 0.1% to 0.2% of succinic acid and 1.2% to 2.3% of citric acid are

dissociated.Thus, the organic acids dissociation is generally not considered although such
small amount of conjugate bases may have a significant impact if they are strongly retained.

The present work aimed at modelling uptake mechanisms on a strong anionic resin. The Langmuir model, commonly used for the retention of organic acids, was coupled with a usual ion-exchange model for the retention of conjugate bases. Simulation results were compared with experimental pulse tests to check the model. The final goalwas to improve the accuracy of apreparative chromatographysimulation program in order to optimize the operating parameters of our pilot-scale ISMB unit (Improved Simulated Moving Bed).

9 2. Chromatography modelling

10 2.1. Mass transfer model

11 The Martin and Synge plate model [19] was chosen to describe the elution of organic 12 acids through a strong anionic resin bed. In each plate of volume V, the mobile phase takes 13 up a volume ε V and the stationary phase a volume (1- ε)V, with ε the bed voidage. Usually, 14 for preparative chromatography,volumes are expressed in BV (bed volume)[20-22], 15 corresponding to the ratio of volume to the resin bed volume.Both phases arealways 16 supposed in equilibriumin all plates (nonlinear equilibrium in our case). The axial dispersion 17 is characterized by the plate number N.

18 2.2. Uptakemechanisms- reactions in stationary phase

The strong anionic resin is a styrene-divinylbenzene resin functionalized with quaternary ammonium groups RMe_3N^+ . The amount of these cationic sites corresponds to the ionexchange capacity $q_{RMe_3N^+}$ of the resin, expressed in mol. L_{resin}^{-1} . The resin is initially in the sulfate form, namely with the counteranions HSO_4^- and SO_4^{-2-} in equilibrium. The proportion of both forms depends on their concentrationin the mobile phase, so on its pH. The concentrations of hydroxyl anion OH⁻ in both phases were considered negligible at pH < 1.5.

The proposed model was based on two classical uptake mechanisms, with some
 assumptions:

Uptake of the organic acid in the dissociated form A⁻ by ion-exchange with resin
 counteranions HSO₄⁻ and SO₄²⁻.

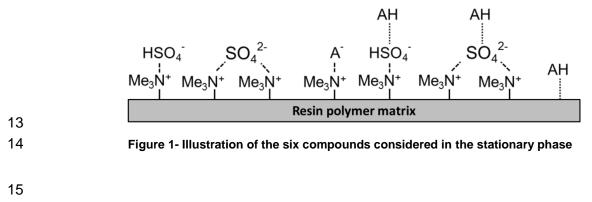
Retention of the organic acid in the molecular form AH according to the Langmuir
 model, by weak bonds (hydrogen or van der Waals), on free sites either on
 counteranions HSO₄⁻ and SO₄²⁻ (weak nucleophiles, *cf.* UOP patents [23-24]) or on the

- 8 polymer matrix whose capacity q_{mat} is also expressed in mol. L_{resin}^{-1} . We supposed that
- 9 the AH retention does not modify the selectivity coefficients of counteranions.

10 Consequently, 6 compounds were considered in the stationary phase: HSO_4^- , SO_4^{-2} , A⁻,

11 AH-site(HSO₄⁻), AH-site(SO₄²⁻) and AH-site(matrix) (figure 1).

12



16 The ion-exchange reactions correspond to the following equilibria:

$$(RMe_{3}N^{+})_{2}SO_{4}^{2-} + 2A^{-} \leftrightarrow 2(RMe_{3}N^{+})A^{-} + SO_{4}^{2-}$$
$$(RMe_{3}N^{+})_{2}SO_{4}^{2-} + 2HSO_{4}^{-} \leftrightarrow 2(RMe_{3}N^{+})HSO_{4}^{-} + SO_{4}^{2-}$$
$$(RMe_{3}N^{+})HSO_{4}^{-} + A^{-} \leftrightarrow (RMe_{3}N^{+})A^{-} + HSO_{4}^{-}$$

Since the affinity of counteranions HSO₄ and SO₄² for resin cationic sites was supposed
not influenced by AH uptake, their equilibrium constants can be written as follows:

$$Ke_{A^{-}/SO_{4}^{2-}} = \frac{[SO_{4}^{2-}]q_{A^{-}}^{2}}{[A^{-}]^{2}q_{SO_{4}^{2-}}} \text{Eq. 1} \qquad Ke_{HSO_{4}^{-}/SO_{4}^{2-}} = \frac{[SO_{4}^{2-}]q_{HSO_{4}^{-}}^{2}}{[HSO_{4}^{-}]^{2}q_{SO_{4}^{2-}}} \text{Eq. 2}$$
$$Ke_{A^{-}/HSO_{4}^{-}} = \frac{[HSO_{4}^{-}]q_{A^{-}}}{[A^{-}]q_{HSO_{4}^{-}}} = \sqrt{\frac{Ke_{A^{-}/SO_{4}^{2-}}}{Ke_{HSO_{4}^{-}/SO_{4}^{2-}}}} \quad \text{Eq. 3}$$

1 where $q_{HSO_{4}}$ and $q_{SO_{4}}$ represent the HSO₄ and SO₄² contents in the stationary phase (free or 2 bound to AH), q_{A} the A⁻ content and [X] the concentration of X in the mobile phase. 3 The ion-exchange equilibrium constants Kecan be estimated from data of resin 4 suppliers by equations 5-7. For example, the Dow Chemical Company gives the selectivity 5 coefficients of anions X⁻ for a strong anionic resin in the hydroxide form, $k_{X/OH}$, defined as

6 follows:

$$k_{X/OH} = \frac{q_X - [OH^-]}{[X^-]q_{OH^-}}$$
 Eq. 4

$$Ke_{HSO_{4}^{-}/SO_{4}^{2-}} = \frac{k_{HSO_{4}/OH}^{2}}{k_{SO_{4}/OH}}$$
 Eq. 5 $Ke_{A^{-}/SO_{4}^{2-}} = \frac{k_{A/OH}^{2}}{k_{SO_{4}/OH}}$ Eq. 6 $Ke_{A^{-}/HSO_{4}^{-}} = \frac{k_{A/OH}}{k_{HSO_{4}/OH}}$ Eq. 7

According to the Langmuir model, the AH uptake reactions on free sites on counteranions
HSO₄⁻ and SO₄²⁻ and on the polymer matrix correspond to the following equilibria:

 $Free \ HSO_4^- \ site + AH \ \leftrightarrow \ Occupied \ HSO_4^- \ site$

Free SO_4^{2-} site + AH \leftrightarrow Occupied SO_4^{2-} site

Free matrix site + $AH \leftrightarrow Occupied$ matrix site

9 The corresponding equilibrium constants Ksare defined as follows:

$$Ks_{AH/HSO_{4}^{-}} = \frac{q_{AH}-HSO_{4}}{[AH]q_{HSO_{4}^{-}}^{free}} Eq. 8 \qquad \qquad Ks_{AH/SO_{4}^{-}} = \frac{q_{AH}-SO_{4}}{[AH]q_{SO_{4}^{-}}^{free}} Eq. 9$$

 $Ks_{AH/mat} = \frac{q_{AH-mat}}{[AH]q_{mat}^{free}}$ Eq. 10

where q_{AH-HSO4}, q_{AH-SO4} and q_{AH-mat} represent the AH concentrations bound touptake sites
 respectively on counteranions HSO₄⁻ and SO₄² and on the polymer matrix, meanwhile q^{free}_{HSO4},
 q^{free}_{SO4} and q^{free}_{mat} represent the concentrations of corresponding free uptake sites.

4 Mass balances on the three kinds of uptake siteslead to the followingequations:

$$q_{HSO_{4}^{-}}^{free} = xq_{HSO_{4}^{-}} - q_{AH-HSO_{4}}$$
Eq. 11 $q_{SO_{4}^{-}}^{free} = yq_{SO_{4}^{-}} - q_{AH-SO_{4}}$ Eq. 12
 $q_{mat}^{free} = q_{mat} - q_{AH-mat}$ Eq. 13

5 where x and y correspond to the uptake capacity of counteranions HSO_4^- and $SO_4^{2^-}$, 6 namely the number ofmolecule that can be retained per counteranion. Indeed, we assumed 7 that several molecules can eventually be retained on each counteranion. These numbers are 8 not necessarily integers and can be smaller than 1 if one molecule AH occupies several 9 counteranions HSO_4^- or $SO_4^{2^-}$ for steric reasons.

For each type of uptake sites, the combination of one mass balance with its corresponding equilibrium constant gives the following usual equations of the Langmuir model. They relate concentration of AH retained respectively on the three kind of sites to AH concentration in mobile phase and total concentration of corresponding sites.

$$q_{AH-HSO_4} = \frac{K_{S_{AH/HSO_{4}}}[AH](xq_{HSO_{4}})}{1+K_{S_{AH/HSO_{4}}}[AH]} \text{Eq. 14} \qquad q_{AH-SO_4} = \frac{K_{S_{AH/SO_{4}}}[AH](yq_{SO_{4}})}{1+K_{S_{AH/SO_{4}}}[AH]} \text{Eq. 15}$$
$$q_{AH-mat} = \frac{K_{S_{AH/mat}}[AH]q_{mat}}{1+K_{S_{AH/mat}}[AH]} \quad \text{Eq. 16}$$

14 2.3. <u>Reactions in mobile phase</u>

At low pH (<1.5), organic acids are mainly in themolecular form AH, howeverthe retention
 of the dissociated form A⁻ was studied(for polyacids like succinic or citric acids, only the
 monodissociated formwas considered). So 5 species were considered in the mobile phase:

H⁺, HSO₄⁻, SO₄²⁻, AH and A⁻. The main reactions are both acid-base equilibria, whose
equilibrium constantsKa are defined as follows:

$$AH \leftrightarrow A^{-} + H^{+} \qquad \qquad HSO_{4}^{-} \leftrightarrow SO_{4}^{2-} + H^{+}$$

$$Ka_{AH} = \frac{[A^-][H^+]}{[AH]}$$
 Eq. 17 $Ka_{HSO_{4}^{-}} = \frac{[SO_{4}^{2-}][H^+]}{[HSO_{4}^{-}]}$ Eq. 18

The pKa of lactic, succinic and citric acids and hydrogen sulfate at 60°C (operating
temperature) are respectively 3.86, 4.13, 3.05 and 2.39.

5

6 2.4. Plate mass balances

7 The plate content is represented by 11 unknown variables (5 in the mobile phase and 6 in

- 8 the stationary phase) that obey 7 independent equilibrium relations (Eq. 1-2 and 14-18).
- 9 Consequently, 4 mass balancesare necessary to solve the system and estimate the content

10 of eachplate at time"t" from the composition of all platesat time"t-1".

11 Mass balance of ion-exchange site RMe_3N^+ whatever the time t,

$$2 q_{SO_4^{2-n}t}^{n} + q_{HSO_4^{n}t}^{n} + q_{A^{-n}t}^{n} = q_{RMe_3N^{+}}$$
 Eq. 19

12 Mass balance of sulfate group SO₄between t-1 and t:

$$\epsilon V \left(\frac{d[HSO_4^-]_t^n}{dt} + \frac{d[SO_4^{2-}]_t^n}{dt} \right) + (1-\epsilon) V \left(\frac{dq_{SO_4^{2-}t}^n}{dt} + \frac{dq_{HSO_4^-}^n}{dt} \right)$$

$$= Q([HSO_4^-]_t^{n-1} - [HSO_4^-]_t^n) + Q([SO_4^{2-}]_t^{n-1} - [SO_4^{2-}]_t^n)$$
Eq. 20

13 Mass balance ofacids (H⁺ element) between t-1 and t (water dissociation negligible):

$$\epsilon V \left(\frac{d[H^+]_t^n}{dt} + \frac{d[HSO_4^-]_t^n}{dt} + \frac{d[AH]_t^n}{dt} \right) + (1 - \epsilon) V \left(\frac{dq_{HSO_4^-}}{dt}^n + \frac{dq_{AH-HSO_4^-}}{dt} + \frac{dq_{AH-SO_4^-}}{dt} + \frac{dq_{AH-mat}^n}{dt} \right)$$

$$= Q([H^+]_t^{n-1} - [H^+]_t^n) + Q([HSO_4^-]_t^{n-1} - [HSO_4^-]_t^n) + Q([AH]_t^{n-1} - [AH]_t^n)$$
Eq. 21

14 Mass balance of organic compoundA (molecular and dissociated forms)between t-1 and t:

$$\varepsilon V\left(\frac{d[A^{-}]_{t}^{n}}{dt} + \frac{d[AH]_{t}^{n}}{dt}\right) + (1-\varepsilon)V\left(\frac{dq_{A^{-}t}^{n}}{dt} + \frac{dq_{AH-HSO_{4}t}^{n}}{dt} + \frac{dq_{AH-SO_{4}t}^{n}}{dt} + \frac{dq_{AH-mat_{t}}^{n}}{dt}\right)$$

$$= Q([A^{-}]_{t}^{n-1} - [A^{-}]_{t}^{n}) + Q([AH]_{t}^{n-1} - [AH]_{t}^{n})$$
Eq. 22

1	In the last three mass balances, the terms on the left-hand side of the equal sign
2	correspond respectivelyto the accumulation terms in the mobile phase and in the stationary
3	phase, meanwhile the expressionson the right-hand side correspond to the difference
4	between inputs (fromthe (n-1) th plate) and outputs (from the n th plate).Q represents the
5	flowrate.
6	The proposed model depends on 10 parameterswhose value is more or less known
7	accuratelyaccording to data from resin suppliers, literature or our own experimental results:
8	Plate number: N,
9	• Resin ion-exchange capacity: $q_{RMe_3N^+}$ expressed in mol. L_{resin}^{-1} ,
10	• Polymer matrix uptake capacity: q_{mat} expressed in mol. L_{resin}^{-1} ,
11	 HSO₄⁻ uptake capacity:x,
12	• SO_4^{2-} uptake capacity: <i>y</i> ,
13	• Equilibrium constant of ion-exchange between A and HSO_4 : $Ke_{A^-/HSO_{\overline{4}}}$,
14	• Equilibrium constant of ion-exchange between A ⁻ and SO ₄ ²⁻ : $Ke_{HSO_{4}^{-}/SO_{4}^{2-}}$,
15	• Equilibrium constant of AH uptake on HSO ₄ ⁻ counteranions: $Ks_{AH/HSO_{4}}$,
16	• Equilibrium constant of AH uptake on SO_4^{2-} counteranions: $Ks_{AH/SO_4^{2-}}$,
17	• Equilibrium constant of AH uptake onresin polymer matrix: $Ks_{AH/mat}$.
18	2.5. Numerical methods
19	The finite-difference method with implicit schemewas used to discretize the 3 differential
20	equations (eq. 20-22). The Newton-Raphson iterative methodwas chosen to solve the
21	nonlinearsystem of 4 remaining equations after simplification. The model and its numerical
22	solving were computed using the software MATLAB [®] . The initial conditions (concentrations in

both phases) need to be given as well as the boundary conditions: composition of the feed
(organic acid solution then eluent) as a function of time.

3 3. Results and discussion

4 3.1. <u>Simulation withonly the Langmuir model and coefficients of equilibrium isotherms</u>

5 First, the simulations of organic acids elution through a strong anionic resin bed were 6 performed considering that AH was only retained on the polymer matrix, according to the 7 Langmuir model.We used the coefficients of equilibrium isotherms obtained previously by 8 frontal analysis (table 1). So, the elution profiles were only influenced by 3 parameters: the 9 plate number N, the polymer matrix uptake capacity q_{mat} equated to q_{max} and the equilibrium 10 constant of AH uptake on resin polymer matrixKs_{AH/mat} equated to*Ks_{AH}*.

11

12Table 1- Resin uptake capacity q_{max} and equilibrium constant Ks_{AH} of 3 organic acids estimated by frontal13analysis at 60°C for a strong anionic resin [18]

-		
	q_{max}	Ks _{AH}
	$mol. L_{resin}^{-1}$	$10^{-1}L_{solution}$. mol^{-1}
Lacticacid	6.8 ± 2.6	1.4 ± 0.5
Succinicacid	2.7 ± 0.9	7.2 ± 2.5
Citricacid	2.5 ± 0.2	9.0 ± 1.7

14

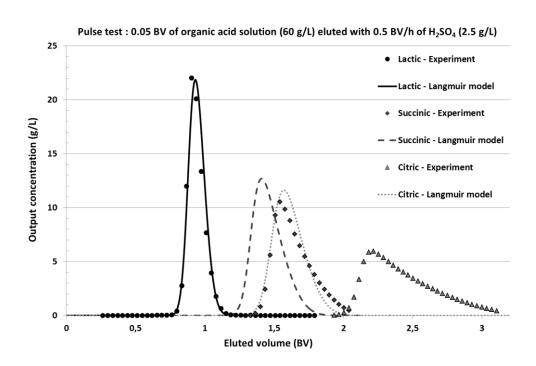
For lactic and succinic acids, the resin uptake capacity q_{max} and the equilibrium constant *Ks*_{AH} have significant error margin because it was not possible to get enough experimental points in the nonlinear region of equilibrium isotherms [18].

According to our previous study[18], the plate number Nwas set to250for the 3 organic acids. It corresponds to the experimental value obtained by pulse testswith lactic acid whose symmetric profile is the most appropriate to characterize axial dispersion. The plate number N was considered equal for all species. Indeed, the study of axial dispersion (Van Deemter
curves) in this work showed that the molecular diffusion is negligible compared to the
mechanical dispersion and the influence of mass transfer limitation, which were quite similar
between compounds in our experimental operating conditions.

These conditions were applied in the model so as to compare with experimental results:
0.05 BV of 60 g.L⁻¹ organic acid solution, whose pH was adjusted to 1.4 with sulfuric acid,
were eluted with a 2.5 g.L⁻¹ sulfuric acid solution. The temperature was 60 °C and the flow
rate was 0.5 BV.h⁻¹.

9 For lactic acid, the simulation results are quite close to the experimental ones (figure 2). 10 So, the lactic acid uptake mechanisms by a strong anionic resin can be well fitted with only 11 the Langmuir model. However, this model is insufficient to simulateelution of succinic and 12 citric acids. Indeed, the simulation gives much earlier and narrower peaks than the 13 experiments, particularly for citric acid.Another uptake mechanism must then be responsible 14 for the delay and peak tailingobserved experimentally and should be taken into account.

15



- Figure 2 Comparison of experimental pulse test profiles with simulated ones considering the Langmuir
 model with experimental coefficients of equilibrium isotherms
- 3

4

3.2. Simulation with taking into account the retention of the dissociated form A⁻

Simulations of organic acids elution were performed with the proposed model which takes
into account the retention of the molecular form AH according to the Langmuir model and the
retention of the dissociated form A⁻ by ion-exchange with resin counteranions.

8 To simplify, as a start, we considered that the molecular form AH was retained only on 9 free uptake sites of counteranions HSO_4^{-1} and SO_4^{-2-} according to UOP patents [23-24]. 10 Moreover, we supposed that the equilibrium constants of AH uptake on both counteranions 11 were equal. They were estimated first from the experimental equilibrium constant Ks_{AH} (table 12 1), then optimized by fitting simulation results to experimental profiles (least squares 13 method). They were expected lower than Ks_{AH} since it includes also the contribution of ion-14 exchange in retention.

According to the supplier, the ion-exchange capacity $q_{RMe_3N^+}$ is about 1.35 mol. L_{bed}^{-1} , 15 equivalent to 2.4 mol. L_{resin}^{-1} . The uptake capacities of counteranions HSO₄⁻ and SO₄²⁻, x and 16 17 y, were supposed equal and defined from the experimental uptake capacity q_{max} (table 1). For 18 instance, q_{max} of lactic acid is twofold to threefold higher than $q_{RMe_3N^+}$, so x and y were supposed equal to 2 (table 3). It could be related to both nucleophile carbonyl groups of 19 counteranions HSO_4^{-1} and SO_4^{-2} while lactic acid gets only one electrophile hydroxyl group. It 20 21 means that 2 molecules of lactic acid can be retained on each counteranion. For succinic and 22 citric acids, x and y were supposed equal to 1 (table 3).

The selectivity coefficients $k_{X/0H}$ were first estimated from data of the Dow Chemical Company (table 2), then optimized by fitting simulation results to experimental profiles (least squares method).

1	Table	e 2- Selectivity coefficients of some anions (compared with OH ⁻) on a strong anionic resin						
2			(Data from Dow Chemical Company)					
			HSO4	SO4 ²⁻	Citrate*	Formate	Acetate	Propionate
		$\mathbf{k}_{\mathrm{X/OH}}$	85	150	220	4.6	3.2	2.6
3		* Valence not mentioned						
4								
5	The pla	plate number N was still set to 250 for the 3 organic acids.Values of simulation						
6	parameters for the 3 organic acids were gathered in the table 3.							
7								

8 Table 3 - Optimized parameters for simulation of lactic, succinic and citric acids elution with the proposed 9 model which also takes into account A⁻ion-exchange

	_		<u> </u>	
		Lacticacid	Succinicacid	Citricacid
N	_	250	250	250
$q_{RMe_3N^+}$	$mol. L_{resin}^{-1}$	2.4	2.4	2.4
k _{A/OH}	-	2.5	330	110
k _{HSO4/OH}	-	85	85	85
k _{S04/0H}	L_{resin} . $L_{solution}^{-1}$	150	150	150
Ks _{AH/HSO4}	$L_{solution}$. mol^{-1}	0.225	0.82	0.70
$Ks_{AH/SO_4^{2-}}$	$L_{solution}$. mol^{-1}	0.225	0.82	0.70
x	_	2	1	1
у	-	2	1	1

10

The figure 3 shows that the proposed model fitted very well the experimental profiles with coherent value for each parameter compared to experimental and resin supplier'sdata.The delay and peak tailing observed mainly withsuccinic and citric acids can be reproduced with this model.

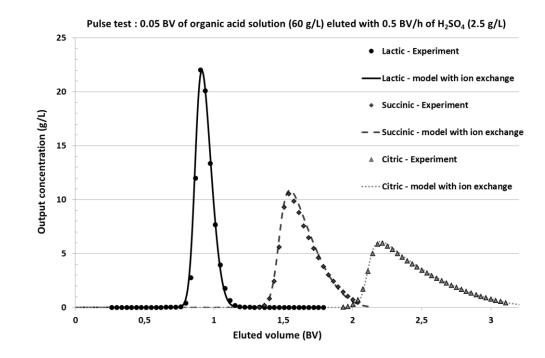


Figure 3 - Comparison of experimental pulse test profiles and simulations ones considering the Langmuir
 model combined with ion-exchange of conjugate bases

5

2

It confirms that the retention of the dissociated organic acids by ion-exchange with resin
counteranions must be considered despite of the very lowdissociation of the acid at acidic pH
(0.1 to 2%). Indeed, the conjugate base of some organic acids, like citrate, can have a very
high affinity for resin cationic sites, so the organic acid can be much more retained than
expected. Moreover, it shows that assumptions added in this simulation to simplify the model
wereacceptable.

12

3.3. Implementation in a ISMB chromatography simulation program

This uptake model was developed in order to improve simulation of preparative chromatography like ISMB (Improved Simulated Moving Bed) that is currently used at pilot scale in our laboratory. We aimed at predicting the ISMB performance more accurately with strong anionic resins and optimizing operating parameters according to thewanted recovery rates and purities. An ISMB unit is composed of 4 columns in series filled with the stationary phase. The ISMB process consists in changing sequentially position of inputs (feed and eluent) and outputs (extract enriched with the more retained component, succinic acid, and raffinate enriched with the less retained, salts and lactic acid). An operating step is composed of four different sub-steps, where each column corresponds to one zone:

6 1) Feed input in the zone 3 and raffinate output from the zone 3.

7 2) Eluent input in the zone 1 and extract output from the zone 1.

8 3) Eluent input in the zone 1 and raffinate output from the zone 3.

9 4) Recirculation loop with all zonesconnected in series.

After one step, each zone is shifted by one column: the zone 1 becomes the zone 2,the
zone 2 becomes the zone 3, etc. So, a cycle is performed after 4 steps. ISMB performance
depends on different volumes eluted during these four sub-steps.

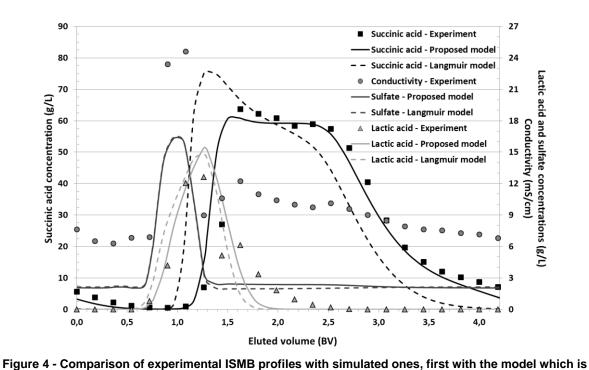
13 ISMB simulations were performed with the Langmuir model whose coefficients were
14 optimized previously from an experimental pulse test at pilot scale, then with the model
15 developed whose coefficients were also optimized in the same way. The feed was mainly
16 composed of sulfatesalts (16 g/L), succinic acid (65 g/L) and lactic acid (14 g/L) while the
17 eluent was a 2.5 g/L sulfuric acid solution.

18 The figure 4 shows that the proposed model fitted better ISMB experimental profiles than

19 the Langmuir model. The accuracy of ISMB simulation with strong anionic resin was

20 improved by taking into account the retention of dissociated organic acids by ion-exchange,

21 despite their very low concentrations.



developed, then with the Langmuir model only

1 2 3

4

5 4. Conclusion

6 Preparative chromatography is an efficient separation technique which is spreading in 7 different industrial fields. For several years now, anionic resins have been performed for the 8 citric acid purification, however uptake mechanisms are still not well known. In a previous 9 work [18], peaks corresponding to succinic and citric acids exhibited singular tailing and 10 delay which could not be explained only by the Langmuir model.

11 The uptake mechanisms supposed in the proposed model are relevant to explain the 12 characteristic elution profiles of some organic acids obtained in pulse tests. To sum up, we 13 assume that the acid molecular form can be retained on resin counteranions or polymer 14 matrix while the dissociated form can be fixed to cationic sites by ion-exchange. Usually, the 15 organic acids dissociation is not considered at such low pH (<1.5), but we proved that even a 16 very small proportion of dissociated form can have a significant impact when it is strongly 17 retained. This is the reason why the Langmuir model is sufficient for lactic acid because of the low affinity of lactate but insufficient for citric acid since citrate affinity is very high for
 quaternary ammonium anionic resins.

The developed model was implemented in a preparative chromatography (ISMB) simulation program in order to optimize the operating parameters of our pilot-scale unit and improve estimation of its performance. Experimental ISMB profiles confirmed that the proposed model was better than the Langmuir model to describe the organic acids retention on strong anionic resins.

8 Next, a plug flow reactor model will be used to take into account different axial dispersions
9 of compounds. Moreover, it would be interesting to further investigate so as to support the
10 assumptions which were made to simplify the model and eventually to refine it.

11 Acknowledgements

12 The authors would like to thank the Fondation Paris-Reims, the RégionChampagne-

13 Ardenne, the Département de la Marne and ReimsMétropole for their financial support.

1 References

- 2 [1] National Renewable Energy Laboratory, Top value added chemicals from biomass, vol. I, 2004.
- 3 [2] Sauer M, Porro D, Mattanovich D and Branduardi P, Microbial production of organic acids:
- 4 expanding the markets, Trends Biotechnol. 26 (2008) 100-108.
- 5 http://dx.doi.org/10.1016/j.tibtech.2007.11.006
- 6 [3] Dierdorp BM, Groot WJ, Van BJ and Veldhuis-Stribos BI, Continuous process for preparing lactic
- 7 acid, WO Patent 0127064 A1, 2001.
- 8 [4] JoglekarHG, Rahman I, Babu S, Kulkarni BD and Joshi A, Comparative assessment of
- 9 downstream processing options for lactic acid, Sep.Purif. Technol. 52 (2006) 1-17.
- 10 <u>http://dx.doi.org/10.1016/j.seppur.2006.03.015</u>
- 11 [5] Sterzel HJ, Vogel H, Exner H and DetlefKratz, Preparation of lactates, US Patent 5453365, 1995.
- 12 [6] Sun X, Wang Q, Zhao W, Ma H and Sakata K, Extraction and purification of lactic acid from
- 13 fermentation broth by esterification and hydrolysis method, Sep.Purif. Technol. 49 (2006) 43-48.
- 14 <u>http://dx.doi.org/10.1016/j.seppur.2005.08.005</u>
- 15 [7] Kumar R, Mahajani SM, Nanavati H and Noronha SB, Recovery of lactic acid by batch reactive
- 16 distillation, J. Chem. Technol.Biotechnol. 81 (2006) 1141-1150.
- 17 <u>http://dx.doi.org/10.1002/jctb.1444</u>
- 18 [8] Coszach P, Bogaert JC, Mariage PA, Chianese A and Parisi MP, Method for purifying lactic acid
- 19 by crystallization, WO Patent 072473 A1,2010.
- 20 [9] Huh YS, Jun YS, Hong YK, Song H, Lee SY and Hong WH, Effective
- 21 purificationofsuccinicacidfromfermentationbrothproducedby Mannheimiasucciniciproducens, Process
- 22 Biochem. 41 (2006)1461-1465.
- 23 <u>http://dx.doi.org/10.1016/j.procbio.2006.01.020</u>
- [10] Guiochon G, Preparative liquid chromatography, J.Chromatogr. A 965 (2002) 129-161.
- 25 <u>http://dx.doi.org/10.1016/S0021-9673(01)01471-6</u>
- 26 [11] MolnárZ, Nagy M, Aranyi A, Hanák L, Argyelán J, Pencz I and Szánya T, Separation of amino
- acids with simulated moving bed chromatography, J. Chromatogr. A 1075 (2005) 77-86.

1 http://dx.doi.org/10.1016/j.chroma.2005.03.083

- 2 [12] Ilisz I, Aranyi A, Pataj Z and Péter A, Enantiomeric separation of nonproteinogenic amino acids
- 3 by high-performance liquid chromatography, J. Chromatogr. A 1269 (2012) 94-121.
- 4 <u>http://dx.doi.org/10.1016/j.chroma.2012.07.011</u>
- 5 [13] Kumar V, Leweke S, von LieresE and Rathore AS, Mechanistic modeling of ion-exchange process
- 6 chromatography of charge variants of monoclonal antibody products, J. Chromatogr. A 1426 (2015)
- 7 140-153.
- 8 <u>http://dx.doi.org/10.1016/j.chroma.2015.11.062</u>
- 9 [14] Guélat B,Khalaf R, Lattuada M, Costioliand Morbidelli M, Protein adsorption on ion exchange
- 10 resins and monoclonal antibody charge variant modulation, J. Chromatogr. A 1447 (2016) 82-91.
- 11 <u>http://dx.doi.org/10.1016/j.chroma.2016.04.018</u>
- 12 [15] Huh YS, Jun YS, Hong YK, Song H, Lee SY and Hong WH, Fructose–glucose separation in a
- 13 SMB pilot unit: Modeling, simulation, design, and operation, AIChE J. 47 (2001) 2042-2051.
- 14 <u>http://dx.doi.org/10.1002/aic.690470915</u>
- 15 [16] Stefansson M andWesterlund D,Ligand-exchangechromatographyof carbohydrates and
- 16 glycoconjugates, J.Chromatogr. A 720 (1996) 127-136.
- 17 <u>http://dx.doi.org/10.1016/0021-9673(95)00276-6</u>
- 18 [17] Nobre C, Santos MJ, Dominguez A, Torres D, Rocha O, Peres AM, Rocha I, Ferreira EC,
- 19 Teixeira JA and Rodrigues LR, Comparison of adsorption equilibrium of fructose, glucose and sucrose
- 20 on potassium gel-type and macroporous sodium ion-exchange resins, Anal.Chim.Acta 654 (2009) 71-
- 21 76.
- 22 http://dx.doi.org/10.1016/j.aca.2009.06.043
- 23 [18] Blanc CL, Theoleyre MA, Lutin F, Pareau D and Stambouli M, Purification of organic acids by
- chromatography: adsorption isotherms and impact of elution flow rate, Sep.Purif. Technol. 141(2015)
- 25 105-112.
- 26 <u>http://dx.doi.org/10.1016/j.seppur.2014.11.032</u>
- 27 [19] Guiochon G, Shirazi DG, Felinger A and Katti AM, Fundamentals of Preparative and Nonlinear
- 28 Chromatography, Academic Press (2006) 283-286. ISBN 0123705371, 9780123705372

- 1 [20] Thang VH and Novalin S, Green biorefinery: separation of lactic acid from grass silage juice by
- 2 chromatography using neutral polymeric resin, Bioresour. Technol. 99(2008) 4368-4379.
- 3 <u>http://dx.doi.org/10.1016/j.biortech.2007.08.045</u>
- 4 [21] Wu J, Hu Y, Zhou J, Qian W, Lin X, Chen Y, Chen X, Xie J, Bai J and Ying H, Separation of d-
- 5 lactic acid from aqueous solutions based on the adsorption technology, Colloids
- 6 Surf.Physicochem.Eng.Asp.407 (2012) 29-37.
- 7 <u>http://dx.doi.org/10.1016/j.colsurfa.2012.04.051</u>
- 8 [22] Morin-Couallier E, Fargues C, Lewandowski R, Decloux M and Lameloise ML, Reducing water
- 9 consumption in beet distilleries by recycling the condensates to the fermentation phase, J. Clean.
- 10 Prod. 16 (2008) 655-663.
- 11 <u>http://dx.doi.org/10.1016/j.jclepro.2007.02.009</u>
- 12 [23] Kulprathipanja S and Oroskar AR, Separation of lactic acid from fermentation broth with an
- 13 anionic polymeric absorbent, US Patent 5068418 (1991).
- 14 [24] Kulprathipanja S and Oroskar AR, Separation of an organic acid from fermentation broth with an
- 15 anionic polymeric adsorbent, US Patent 5068419 (1991).