



HAL
open science

F4+ ETEC infection and oral immunization with F4 fimbriae elicits an IL-17-dominated immune response

Yu Luo, Ut van Nguyen, Pedro Y. de La Fe Rodriguez, Bert Devriendt, Eric Cox

► **To cite this version:**

Yu Luo, Ut van Nguyen, Pedro Y. de La Fe Rodriguez, Bert Devriendt, Eric Cox. F4+ ETEC infection and oral immunization with F4 fimbriae elicits an IL-17-dominated immune response. *Veterinary Research*, 2015, 46 (1), pp.121. 10.1186/s13567-015-0264-2. hal-01341430

HAL Id: hal-01341430

<https://hal.science/hal-01341430v1>

Submitted on 4 Jul 2016

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

RESEARCH ARTICLE

Open Access



F4⁺ ETEC infection and oral immunization with F4 fimbriae elicits an IL-17-dominated immune response

Yu Luo^{1*}, Ut Van Nguyen¹, Pedro Y. de la Fe Rodriguez², Bert Devriendt^{1†} and Eric Cox^{1†}

Abstract

Enterotoxigenic *Escherichia coli* (ETEC) are an important cause of post-weaning diarrhea (PWD) in piglets. Porcine-specific ETEC strains possess different fimbrial subtypes of which F4 fimbriae are the most frequently associated with ETEC-induced diarrhea in piglets. These F4 fimbriae are potent oral immunogens that induce protective F4-specific IgA antibody secreting cells at intestinal tissues. Recently, T-helper 17 (Th17) cells have been implicated in the protection of the host against extracellular pathogens. However, it remains unknown if Th17 effector responses are needed to clear ETEC infections. In the present study, we aimed to elucidate if ETEC elicits a Th17 response in piglets and if F4 fimbriae trigger a similar response. F4⁺ ETEC infection upregulated IL-17A, IL-17F, IL-21 and IL-23p19, but not IL-12 and IFN- γ mRNA expression in the systemic and mucosal immune system. Similarly, oral immunization with F4 fimbriae triggered a Th17 signature evidenced by an upregulated mRNA expression of IL-17F, ROR γ t, IL-23p19 and IL-21 in the peripheral blood mononuclear cells (PBMCs). Intriguingly, IL-17A mRNA levels were unaltered. To further evaluate this difference between systemic and mucosal immune responses, we assayed the cytokine mRNA profile of F4 fimbriae stimulated PBMCs. F4 fimbriae induced IL-17A, IL-17F, IL-22 and IL-23p19, but downregulated IL-17B mRNA expression. Altogether, these data indicate a Th17 dominated response upon oral immunization with F4 fimbriae and F4⁺ ETEC infection. Our work also highlights that IL-17B and IL-17F participate in the immune response to protect the host against F4⁺ ETEC infection and could aid in the design of future ETEC vaccines.

Introduction

In neonatal and recently weaned pigs, ETEC-associated diarrhea is a major cause of illness and mortality and leads to great economic losses in the swine production industry worldwide [1, 2]. ETEC express fimbriae, which are long proteinaceous appendages radiating from the surface of the bacterium. These fimbriae mediate adhesion to host intestinal epithelia through an interaction with specific receptors present on the brush borders of the small intestinal enterocytes, enabling bacterial colonization [3]. Porcine-specific ETEC strains possess five different fimbrial subtypes, of which F4 fimbriae are the

most frequently associated with ETEC-induced diarrhea in piglets [4–6]. Recent data indicate F4 fimbriae are not merely involved in adherence, but also play a role in the modulation of the immune system [7, 8]. In addition, these F4 fimbriae are potent mucosal immunogens, since they elicit a fast secretion of F4-specific secretory IgA (SIgA) at the intestinal tissues upon oral administration, protecting piglets against a challenge infection [3, 9–11].

SIgA responses can be generated by both T cell-dependent and T cell-independent pathways [12]. Recently, Th17 cells and their production of IL-17A and IL-21 have been implicated in the induction of SIgA directed against gut-dwelling pathogens [13–15]. This ability to trigger SIgA responses explains their critical function in the host defense against extracellular pathogens such as *Candida albicans*, *Citrobacter rodentium*, *Salmonella typhimurium*, *Klebsiella pneumoniae*, and *Giardia muris* [16–20]. Effective immunity to pathogens

*Correspondence: yu.luo@ugent.be

†Bert Devriendt and Eric Cox shared senior authorship

¹ Laboratory of Veterinary Immunology, Department of Virology, Parasitology and Immunology, Faculty of Veterinary Medicine, Ghent University, Ghent, Belgium

Full list of author information is available at the end of the article

requires T lymphocytes to be endowed with appropriate effector properties. In this context, naive CD4⁺ T cells differentiate into different effector cells and tailor their functions to the nature of the microbial threat. Besides the classical Th1 and Th2 cells, at least two other CD4⁺ T effector lineages have been identified and described, including Th17 and regulatory T cells (Tregs) [21, 22]. In humans and mice, Th17 cells can be induced from naive CD4⁺ T cells with IL-6 and/or IL-21 in the combination of TGF- β , and mainly secrete IL-17A, IL-17F and IL-21 [23]. IL-17 (also known as IL-17A) is the hallmark cytokine of Th17 cells and is the founding member of the IL-17 cytokine family, which consists of six members: IL-17A, IL-17B, IL-17C, IL-17D, IL-17E (also known as IL-25) and IL-17F [24], [25]. Among the IL-17 family members, IL-17F shares the highest sequence homology with IL-17A [26]. Although both cytokines can bind to the same receptors, regulate inflammatory responses and are involved in mucosal defense, they show a distinct binding affinity for these receptors and as such different roles in triggering immunity [25]. IL-17E on the other hand triggers Th2 immunity and is involved in the clearance of helminths and allergy [27, 28]. Recently, IL-17C produced by goblet cells and enteroendocrine cells has been proposed to mediate the intestinal inflammation in IBD patients [29]. The function of the other IL-17 cytokines in immunity is still poorly understood.

As in humans, porcine Th17 cells arise from naive CD4⁺ lymphocytes via IL-6 in the context of TGF- β and secrete IL-17A and IL-21 [30]. However, whether Th17 cells or IL-17 cytokine family members participate in the immune response against an ETEC infection in pigs or other species is still unclear. The heat labile toxin (LT) from a human ETEC strain was found to enhance IL-17A production by human PBMCs in response to antigen or mitogen stimulation [31]. Similarly, the IL-17A promoting effect of LT was also reported in mice upon *Helicobacter pylori* infection [32]. In pigs, IL-17A mRNA production in the small intestine was upregulated early during F4⁺ ETEC infection [33]. In contrast, serum IL-17A levels were unaltered early during ETEC infection, while in the intestinal tissues a downregulated IL-17A mRNA production 7 days post infection was observed [34]. However, as F4⁺ ETEC infections are usually cleared within 7–8 days, we hypothesized that a potential Th17 response due to ETEC infection should occur earlier. Thus, in the present study, we addressed if an F4⁺ ETEC infection and oral administration with F4 fimbriae could skew the T helper cell differentiation to a Th17 profile by assessing the mRNA expression profile of key transcription factors and cytokines involved in T cell polarization at systemic and intestinal tissues.

Materials and methods

The methodology of the animal experiment was approved by the Ethical Committee of the Faculty of Veterinary Medicine, Ghent University (EC2014/01).

F4⁺ ETEC challenge

Six F4 receptor-positive (F4R⁺) piglets (7–8 week-old, Belgian Landrace) were selected based on the MUC4 TaqMan assay as previously described [35]. Upon arrival in the animal care facilities, all animals were treated orally with colistin (150 000 U/kg of body weight/day; Colivet; Prodivet Pharmaceuticals, Eynatten, Belgium) until 3 days before inoculation to prevent potential ETEC infections due to stress caused by transport and handling of the animals. During the whole trial all pigs had access to water and feed ad libitum. To reduce the bacterial gut flora, the piglets were given orally a broad-spectrum antibiotic (2 mL florfenicol (Nuflor; Schering-Plough, Brussels, Belgium) for each pig for two consecutive days. One day after Nuflor administration, piglets were inoculated with the ETEC reference strain GIS26 (O149:K91:F4ac⁺, LT⁺STa⁺STb⁺) or phosphate buffered saline (PBS) on day 0 (D0) and day 1 (D1). In brief, piglets were sedated with Stressnil (40 mg/mL; Janssen-Cilag, Berchem, Belgium) and the gastric pH was neutralized by intragastric administration of 60 mL NaHCO₃ (1.4% in distilled water) followed by intragastric administration of 10¹⁰ GIS26 in 10 mL sterile PBS. Faeces were collected at D0 and D1 and at D2, D3 and D4 post infection to determine F4⁺ ETEC shedding as previously described [9]. The severity of diarrhea was scored daily as previously described [36]. On D4, piglets were euthanized using pentobarbital (Kela NV, Belgium) and intestinal tissues were excised. Jejunal segments with and without Peyer's patches, ileal segments and mesenteric lymph nodes (MLNs) were collected and washed twice with Krebs-Henseleit buffer (0.12 M NaCl, 0.014 M KCl, 1 mM KH₂PO₄, 0.025 M NaHCO₃, pH 7.4) and once with Krebs-Henseleit buffer containing 1% (v/v) formaldehyde. Next, the MLNs and intestinal samples were frozen in liquid nitrogen and stored at –80 °C until RNA extraction.

Immunohistochemistry

The intestinal tissues were sampled as described above, washed with Krebs-Henseleit buffer, embedded in methocel (Fluka, Bornem, Belgium), snap-frozen in liquid nitrogen and stored at –80 °C until sectioning. Cryosections (14 μ m) were cut and mounted on 3-aminopropyl-triethoxysilane (Sigma-Aldrich, Bornem, Belgium)-coated glass slides. After drying for 30 min at room temperature (RT), the slides were fixed in 4% paraformaldehyde for 20 min at 4 °C and then embedded in 0.1% Triton (TritonTM X-100, Sigma-Aldrich) for

10 min at RT. Slides were washed three times with PBS with gentle agitation and then incubated overnight with anti-CD3 mAb (0.5 µg/mL, mouse IgG₁, clone PPT3) and biotinylated anti-swine IL-17A polyclonal rabbit antibody (2.5 µg/mL, Kingfisher biotech, St. Paul, MN, USA) in PBS at 4 °C in a humidified chamber. Purified Mouse IgG₁ (0.5 µg/mL, Life Technologies, Carlsbad, CA, USA) and irrelevant rabbit polyclonal IgG (2.5 µg/mL, ab27472, Abcam, Cambridge, UK) were used as negative control. The next day, the sections were washed and incubated with streptavidin-FITC (2.5 µg/mL, Biolegend, London, UK) and Texas Red-X conjugated goat anti-mouse IgG (H + L) secondary antibody (5 µg/mL, Life Technologies) at RT for 1.5 h. Subsequently, the sections were washed in PBS and then the nuclei were counterstained with Hoechst (10 µg/mL, Sigma-Aldrich) for 15 min at RT. Finally, the slides were mounted in glycerol containing 0.223 M 1,4-diazobicyclo-(2,2,2)-octane (Sigma-Aldrich) and imaged on a Leica TCS SP2 confocal microscope (Leica Microsystems GmbH).

Isolation of peripheral blood mononuclear cells

Blood was taken on heparin from the jugular vein of piglets on D0 and D1 prior to inoculation and on D2, D3 and D4 post infection. PBMCs were isolated by density gradient centrifugation using Lymphoprep (Axis-shield, Oslo, Norway). Erythrocytes were lysed in ammonium chloride solution. The resulting PBMC fraction was washed twice in ice cold PBS + 1 mM EDTA and counted using a hemocytometer. The viability was confirmed by exclusion of the vital dye Trypan blue. Then, the cells were either put in TRIzol Reagent (1 × 10⁷ cells in 1 mL TRIzol Reagent; Life Technologies) for RNA extraction or cultured at a concentration of 5 × 10⁶ cells/mL in leukocyte medium (RPMI-1640 (Gibco), fetal bovine serum (FBS) (10%) (Gibco), sodium pyruvate (1 mM) (Gibco), L-glutamine (2 mM) (Gibco), penicillin (100 IU/mL), streptomycin (100 µg/mL) (Gibco), and non-essential amino acids (1%)) in the absence or the presence of F4 fimbriae (5 µg/mL) for up to 72 h at 37 °C, 5% CO₂ in a humidified atmosphere. Next, the cells were collected, lysed in 1 mL TRIzol and stored at -80 °C until RNA extraction. Cell supernatants were collected and stored at -80 °C until further processing.

RNA extraction and cDNA synthesis

Prior to RNA extraction, frozen tissue sample were homogenized in liquid nitrogen with mortar and pestle. RNases were removed by baking mortar and pestle at least 3 h at 200 °C. Briefly, mortar and pestle were chilled in liquid nitrogen followed by grinding of the tissues until a fine powder was formed. This powder (600–1200 mg) was added to 1.0 mL prewarmed (37 °C) TRIzol Reagent

and immediately mixed well. Then RNA was extracted following the manufacturer's instructions. RNA samples were treated with DNase I (Promega, Madison, WI, USA) and purified with the RNeasy Mini Kit (Qiagen Benelux, Venlo, The Netherlands) according to the manufacturer's guidelines. The RNA concentration and purity were determined by measuring the optical density at OD₂₆₀/OD₂₈₀ and OD₂₆₀/OD₂₃₀ with a NanoDrop 2000/2000c spectrophotometer (NanoDrop Technologies, Wilmington, DE, USA). All samples had OD₂₆₀/OD₂₈₀ ratios between 1.9 and 2.0 and OD₂₆₀/OD₂₃₀ ratios between 1.9 and 2.1. Total RNA (1 µg) was reverse transcribed using Superscript™ III Reverse Transcriptase (200 U; Life Technologies) and random primers (7.5 ng/µL; Life Technologies). To check the synthesis of amplifiable cDNA in the reverse transcription, a conventional PCR step was performed using GAPDH and β-actin specific primers (Table 1).

Real-time qPCR

Primers (Table 1) were designed using Primer 5 to span an exon–exon junction thereby avoiding amplification of genomic DNA. The primers were purchased from Eurogentec (Liege, Belgium). The amplification efficiency of all the reactions ranged from 94 to 103%. The PCR products were sequenced and subjected to agarose gel electrophoresis to verify their specificity. cDNA was diluted 8x in DEPC-treated ddH₂O and combined with primer pairs and SYBR Green PCR Master Mix (Applied Biosystems, Warrington, UK) according to the manufacturer's recommendations. Quantitative PCR (qPCR) assays were run on the StepOnePlus real-time PCR system (Applied Biosystems) with the following cycling conditions: 95 °C for 3 min, followed by 40 cycles of denaturation at 95 °C for 15 s, annealing for 30 s and elongation at 72 °C for 30 s. Fluorescence acquisition was measured at 72 °C and melting curve analysis was done at 65–95 °C with continuous fluorescence acquisition. The stability of the GAPDH, β-actin, 60S ribosomal protein L19 (RPL-19) and Cyclophilin A (CyPA) mRNA expression levels was evaluated by geNorm [37]. We finally selected GAPDH, β-actin and RPL-19 as reference genes. All reactions were performed in triplicate and relative gene transcription values were calculated using the 2^{-ΔΔCt} method and normalized against these three selected reference genes [38].

Purification of F4 fimbriae

F4 fimbriae were purified from the ETEC reference strain GIS 26 or the IMM01 strain (O147:F4ac⁺LT⁺STb⁺, which lacks flagellin expression) as previously described [9]. Briefly, the bacteria were cultured in tryptone soy broth (Difco Laboratories, Biotrading, Bierbeek, Belgium) at 37 °C for 18 h, collected by centrifugation and

Table 1 List of the primers used in the qPCR assay

Gene	Sequence (5'→3')	Size (bp)	Ta (°C)	Reference
IL-17A	F:ACTCAAACGCTTCACCTCAC R:AGCCCACTGTCAACCACTT	234	58	NM_001005729.1
IL-17B-like	F:CTGGCCAAGAGGAAGTGTGAG R:GGGTCTGGTTGATGCTGTAG	92	60	XM_003124086.1
IL-17F	F:GAGGCAGCAGCTCGAAAAT R:TCCCGGGTGATGTTGTAATCC	173	60	NC_010449.4
IL-21	F:GGCACAGTGGCCATAAATC R:GCAGCAATTCAGGGTCCAAG	124	60	[30]
IL-22	F:TTGACCAGTCCAATTCCAGCAGC R:GCAGCGCTCTCATATTGACTCC	143	60	XM_001926156.1
IL-23p19	F:CCAAGAGAAGAGGGAGATGATGA R:TGCAAGCAGGACTGACTGTTGT	107	57	NM_001130236.1
RORyt	F:TTCAGTACGTGGTGGAGTTC R:TGTGGTTGTCAGCGTTGTAG	141	60	[30]
IL-10	F:CCATGCCAGCTCAGCACTG R:CCCATCACTCTCGCTTCGG	295	60	[65]
IL-13	F:GTCATTGCTCTCACCTGCTT R:TTGGTGCTCGGATGTGCTT	308	58	[66]
GATA-3	F:GCTCTACCACAAAATGAACGGAC R:TCGTTGTGGTTGACAGTTTGC	110	58	NM_001044567.1
IL-12p40	F:GGTTTCAGACCCGACGAACTCT R:CATATGCCACAATGGGAGATG	112	60	[67]
IFN-γ	F:GAGCCAAATTGTCTCCTTCTACT R:CTGACTTCTTCCGCTTCT	262	60	[68]
T-bet	F:TCAATCCTACTGCCCACTAC R:TTAGGAGACTCTGGGTGAAC	151	60	[69]
Foxp3	F:TGCCATTCGCCACAACCTT R:CCTGTCCATCCTTCTTTCCTT	179	60	NM_001128438.1
AID	F:AGAAGTTTCAAAGCCTGGGAG R:TCAACCTCATAAGGGGCAAA	92	57	XM_003126511.1
β-Actin	F:TCATCACCATCGGCAACG R:TTCCTGATGTCACGTCGC	133	60	[70]
GAPDH	F:GGGCATGAACCATGAGAAGT R:AAGCAGGGATGATGTTCTGG	230	60	[65]
RPL19	F:AACTCCCGTCAGCAGATCC R:AGTACCCCTCCGCTTACCG	147	60	[69]

RORyt: RAR-related orphan receptor gamma t, AID: activation-induced (cytidine) deaminase, RPL-19: 60S ribosomal protein L19

washed in sterile PBS. Subsequently, F4 fimbriae were isolated by mechanical shearing of the bacterial suspension followed by centrifugation to remove the remaining bacteria. The fimbriae were precipitated with ammonium sulfate (40% saturation), the pellet was dissolved in PBS and dialysed overnight against PBS at 4 °C. Next, the fimbrial proteins were filtrated (0.22 μm) and the endotoxins were removed by using EndoTrap columns (Hyglos GmbH, Regensburg, Germany) following the manufacturer's guidelines. After endotoxin removal, the fimbrial solution contained almost no endotoxin (0.24 EU/mL) as

determined by the *Limulus* amoebocyte lysate test (Lonza, Walkersville, MD, USA). The protein concentration was determined by the bicinchoninic acid reaction (Sigma-Aldrich) with bovine serum albumin as a standard and the purity was assessed by sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE, 12%).

Oral immunization with purified F4 fimbriae

Eleven F4 receptor-positive (F4R⁺) piglets (4–5-week-old) were selected, housed and treated with antibiotics as described above. The experimental group consisted

of six F4 fimbriae seronegative pigs (three Hypor-west and three Large White × Belgian Landrace), while the control group contained five F4 fimbriae seropositive pigs (two Hypor × Pietrain and three Yorkshire × Large White × Landrace). Prior to the oral immunization piglets were deprived of feed and water for 3 h. Purified F4 fimbriae from the GIS26 strain (1 mg in 10 mL PBS) or PBS were administered orally for three subsequent days to the piglets of the experimental or the control group, respectively. Blood was taken from the jugular vein on the day prior to the initial immunization and at D4 and D9 post immunization to isolate PBMCs for RNA extraction as described above. Blood samples of D0 and D9 were also used to measure F4-specific serum IgG and IgA antibodies by ELISA to monitor immunization success [9].

In vitro culture of PBMCs

Blood was taken from 8–12-week-old healthy conventionally reared, F4 seronegative pigs (Belgian Landrace) and PBMCs were isolated and suspended at a concentration of 5×10^6 cells/mL in leukocyte medium as described above. Subsequently, PBMCs were transferred to a 24-well tissue culture plate and stimulated with 5 µg/mL F4 fimbriae in the presence or absence of polymyxin B (Sigma-Aldrich), endotoxin-free F4 fimbriae (5 µg/mL) or medium at 37 °C, 5% CO₂ in a humidified atmosphere. The cells and supernatants were harvested at 24, 48 and 72 h after stimulation and stored properly as mentioned above. The mRNA expression profile of the PBMCs was analyzed using qPCR as described above.

Cytokine ELISA

The secretion of IL-17A, IL-10, IFN-γ and IL-22 in cell-free supernatants was measured using commercial ELISA kits according to the manufacturer's guidelines (IL-17A and IFN-γ, Kingfisher biotech; IL-10, Life Technologies; IL-22, Sigma-Aldrich).

Statistical analysis

Statistical analysis was performed with the Mann–Whitney U test or Kruskal–Wallis Test for the independent samples and Friedman's two-way analysis for the related samples in the SPSS 22 software package. The significance level was set at $p < 0.05$.

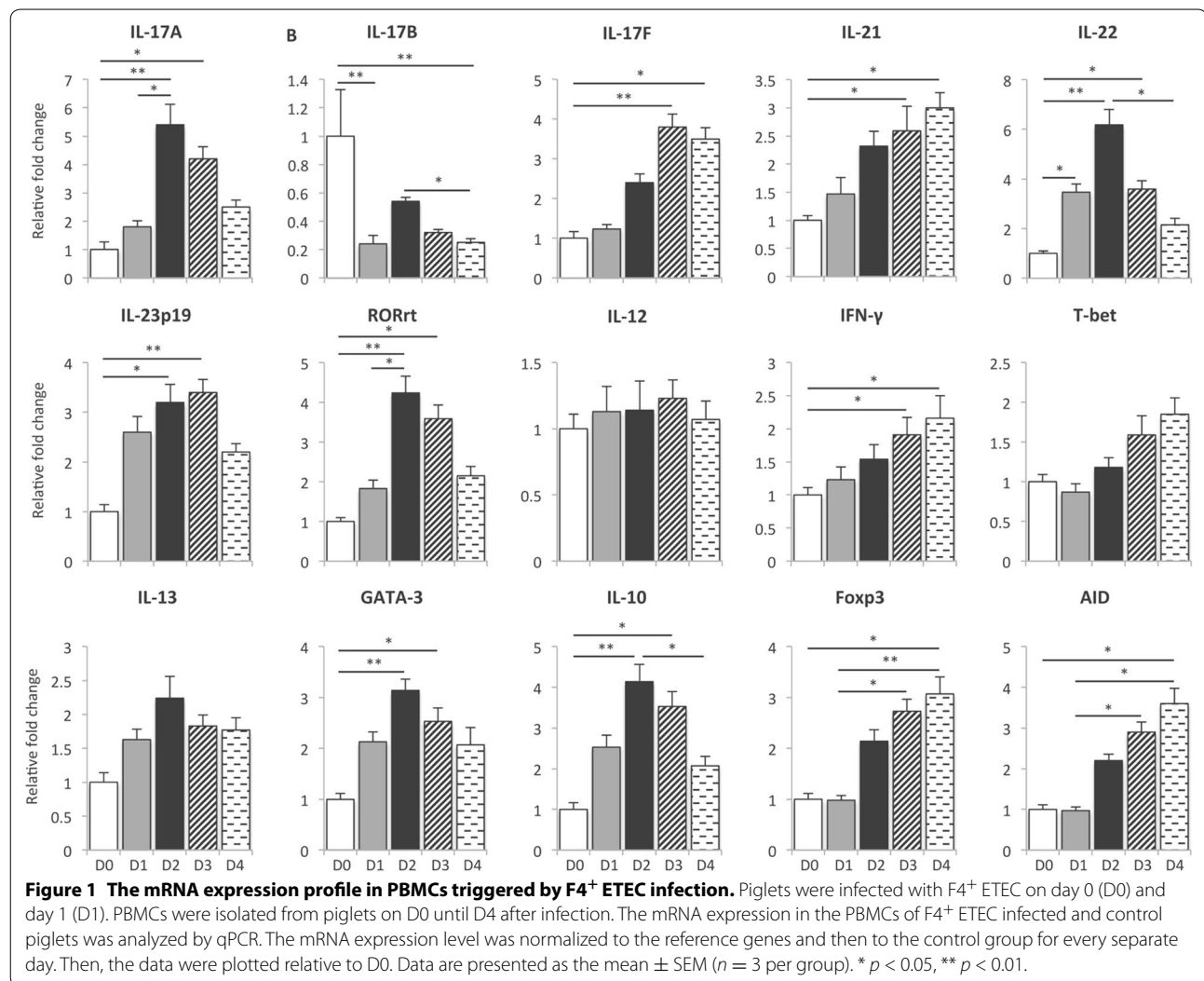
Results

F4⁺ ETEC infection triggers IL-17 signature responses in PBMCs and small intestinal tissues

To analyze the type of immune response elicited by F4⁺ ETEC, we assessed the mRNA expression profile of key cytokines and transcription factors involved in either T cell polarization or their effector functions. Indeed, the

mRNA expression levels of Th1 (IFN-γ, IL-12, T-bet), Th2 (IL-13, GATA-3), Th17 (IL-17A, IL-21, IL-22 IL-23p19, orphan nuclear receptor (RORγt)) and regulatory T cells (Foxp3, IL-10) were evaluated in PBMCs and several intestinal tissues. In addition, the mRNA expression of the IL-17 family cytokines IL-17B and IL-17F as well as activation-induced (cytidine) deaminase (AID), a B-cell specific enzyme required for somatic hypermutation and class switch recombination, was assessed. Following F4⁺ ETEC challenge, only one of the three pigs exhibited severe diarrhea, while F4⁺ ETEC shedding was detected in all infected pigs (Additional file 1). Notably, the expression of IL-17A mRNA was significantly increased in PBMCs at D2 and D3 after F4⁺ ETEC infection (Figure 1). In addition, an increased mRNA expression of IL-23p19 and RORγt was also detected these days (Figure 1). Moreover, F4⁺ ETEC infection also increased the mRNA expression of the Th17 cytokines IL-21, IL-22 and IL-17F. In contrast, IL-17B mRNA expression was downregulated in F4⁺ ETEC infected pigs (Figure 1). With regard to the Th1-related genes, only the mRNA expression of IFN-γ was upregulated at D3 and D4, while the mRNA expression of the Th1-related transcription factor T-bet and the Th1-inducing cytokine IL-12 was not influenced by F4⁺ ETEC infection. ETEC infection also significantly enhanced the mRNA expression of the Th2-related transcription factor GATA-3 and the Tregs-related genes Foxp3 and IL-10. Also the AID mRNA levels increased by F4⁺ ETEC in a time-dependent manner in the PBMCs compared to control pigs, which could indicate the presence of pathogen-specific circulating B cells undergoing class switching.

We further compared the expression profiles of these transcription factors and cytokines in several intestinal tissues on D4 after F4⁺ ETEC infection. As shown in Figure 2, the fold changes of almost all the examined transcripts were higher in the Peyer's patches (PP) and MLNs than those in the jejunal and ileal lamina propria after F4⁺ ETEC infection. Interestingly, these transcripts also displayed higher expression in the PP and MLN of control pigs (Additional file 2). Similar to PBMCs, F4⁺ ETEC infection significantly increased IL-17A, IL-17F, IL-21 and IL-23p19 mRNA levels in all examined tissues as compared to control pigs (Figure 2). In addition, IL-17A, IL-17F and IL-23p19 transcripts were strongly induced in PP and MLN. In contrast to PBMCs, IL-17B mRNA expression was upregulated upon F4⁺ ETEC infection in all intestinal tissues except jejunum (JJ). No significant difference in IL-22 mRNA expression was observed in jejunal and ileal lamina propria, whereas a small upregulation was found in the ileal PP and MLN in F4⁺ ETEC infected pigs as compared to control pigs. Regarding the Th2-related genes, we observed upregulated IL-13 and



GATA-3 mRNA levels with the highest change in the ileal PP and MLN, respectively. Similar to the systemic immune system, F4⁺ ETEC infection did not alter the intestinal mRNA expression of IL-12, IFN- γ and T-bet (Figure 2), while both Foxp3 and IL-10 mRNA levels were significantly upregulated, especially in the PP and MLN. Interestingly, mRNA expression of AID was highly upregulated upon F4⁺ ETEC infection in the PP and MLN, indicating active class switching in those tissues.

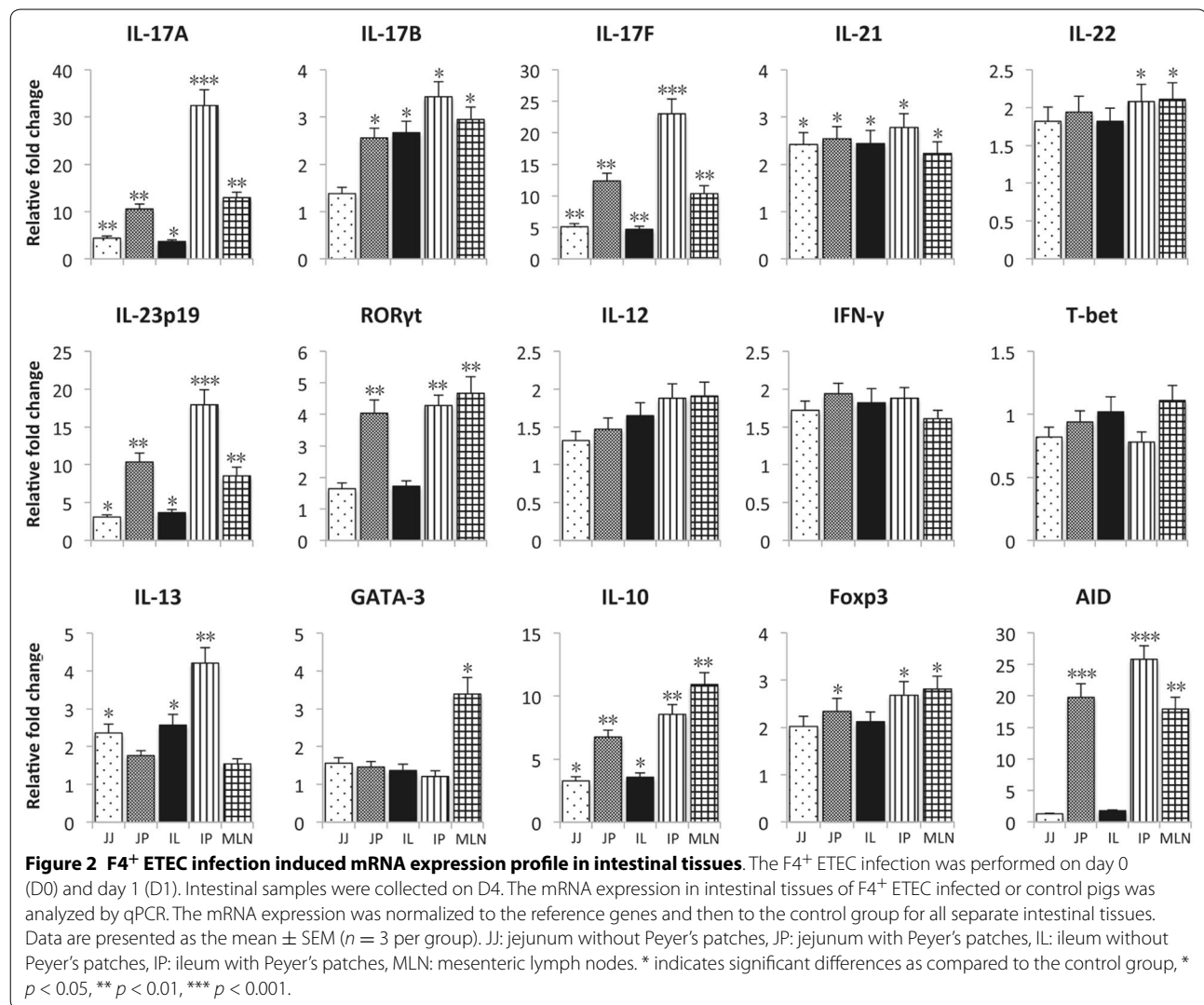
F4⁺ ETEC infection increased CD3⁺IL-17A⁺ T cells in the intestinal tissue

Given the high expression of IL-17A mRNA in the ileal PP of F4⁺ ETEC infected piglets, we assessed if this correlated with an influx of IL-17A⁺ T cells in that tissue. Immunofluorescence analysis of ileal tissue clearly showed an increase in CD3⁺IL-17A⁺ cells in the crypts and villi in F4⁺ ETEC infected piglets as compared to

controls. In contrast, colocalization of CD3 and IL-17A was rarely observed in the tissue of control pigs (Figure 3).

Oral immunization of piglets with F4 fimbriae triggered systemic IL-17 responses

We detected robust Th17 and IL-17-related cytokine responses upon F4⁺ ETEC infection both in intestinal tissues and PBMCs. In a next effort we wanted to determine if these observations could be reproduced by oral immunization of piglets with F4 fimbriae, as these are potent oral immunogens. We only focused on the PBMCs since similar systemic and mucosal responses were found after F4⁺ ETEC infection. Upon immunization, the F4-specific IgG and IgA serum antibody titers were significantly increased as compared to the control group (Additional file 3), indicating a successful immunization. Similar to F4⁺ ETEC infection, oral immunization with F4 fimbriae significantly increased the mRNA

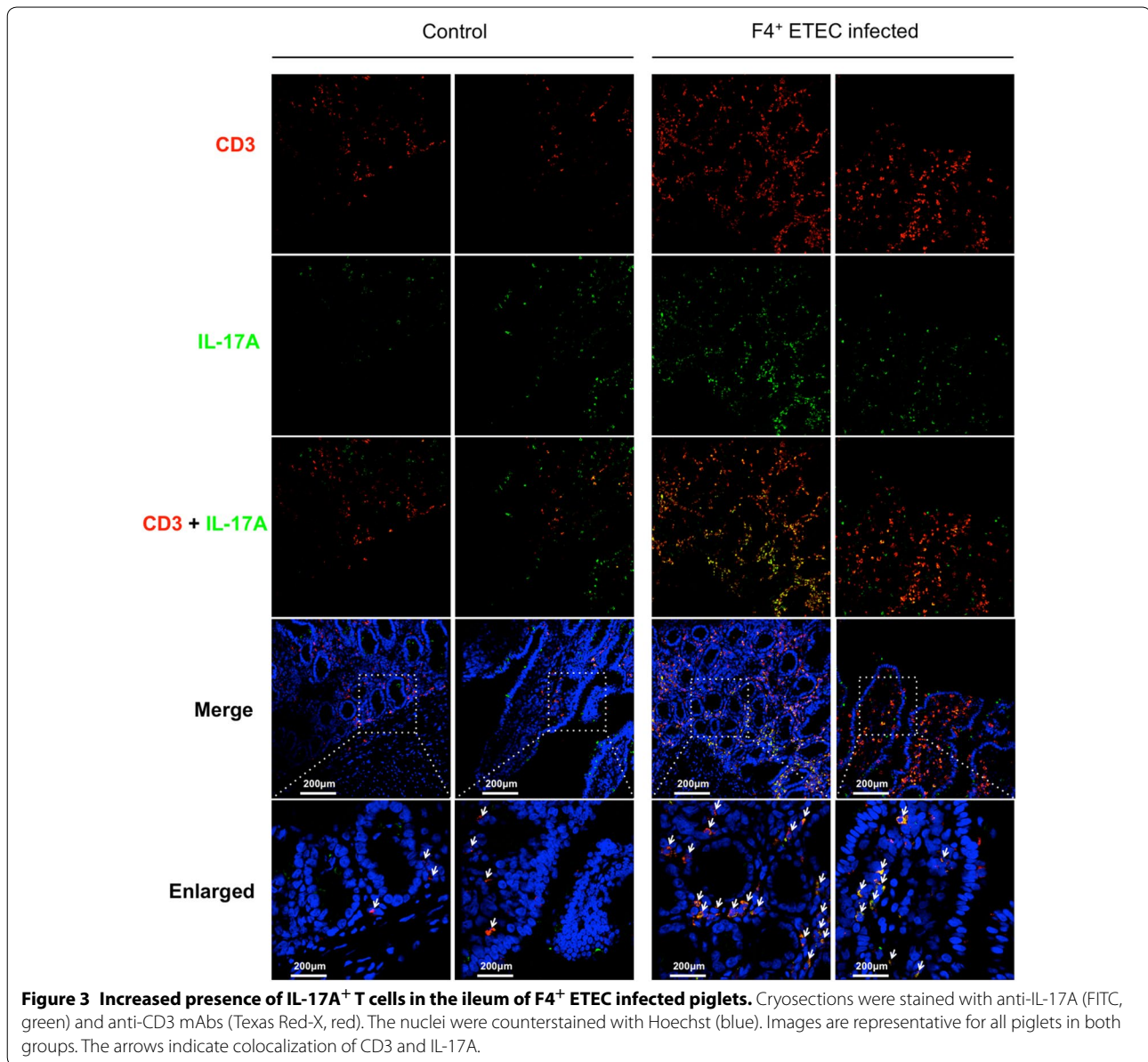


expression of Th17-related genes, including IL-17E, IL-21, IL-22, IL-23p19 and ROR γ t, although with different kinetics (Figure 4). Compared to control pigs, significant changes were only observed at D4 for IL-17F and IL-23p19 and at D9 for IL-22, whereas IL-21 and ROR γ t mRNA expression levels peaked at D4. Unexpectedly, IL-17A mRNA expression was undetectable in porcine PBMCs upon oral immunization with F4 fimbriae. In contrast to the PBMCs of $F4^+$ ETEC infected pigs, oral immunization with F4 fimbriae upregulated IL-17B mRNA expression at D4. In agreement with $F4^+$ ETEC infection, only IFN- γ , but not IL-12 and T-bet, mRNA expression was elevated in immunized pigs. Likewise, GATA-3 levels increased in a time-dependent manner to reach significance at D9. Unexpectedly, we also failed to detect IL-13 mRNA expression in PBMCs upon F4 fimbriae immunization. Regarding the Tregs-related genes, only Foxp3 mRNA expression was enhanced. Similar to

the infection trial, we also found a significant increase of AID mRNA expression, presumably indicating the presence of $F4$ -specific circulating B-cells undergoing class switching.

F4 fimbriae induced a Th17-signature cytokine expression in naive PBMCs

The above data indicate the potential of F4 fimbriae to induce Th17 responses. To further address this potential, PBMCs were isolated from naive animals and stimulated with F4 fimbriae. Upon stimulation for 24 h the mRNA expression levels of IL-17A, IL-17F and IL-22 were significantly increased as compared to unstimulated cells (Figure 5). This upregulated expression level lasted at least till 72 h post stimulation. In addition, F4 fimbriae significantly enhanced IL-23p19 mRNA expression, which peaked at 24 h post stimulation, while IL-21 and ROR γ t mRNA expression only displayed a significant



change at 72 h (Figure 5). Intriguingly, F4 fimbriae stimulated PBMCs downregulated IL-17B mRNA expression. Regarding the Th1-related genes, we only observed a significant upregulation of IFN- γ transcripts, while IL-12 and T-bet mRNA levels were not affected. Furthermore, no significant difference in the mRNA expression of Foxp3, IL-10, GATA-3 and IL-13 was observed. Similar to the infection and immunization experiment, AID mRNA levels were significantly increased in F4 fimbriae stimulated PBMCs (Figure 5).

To confirm that the increased transcript levels upon stimulation of PBMCs with F4 fimbriae resulted in higher protein levels, we measured the secretion of

the corresponding cytokines using ELISA. As shown in Figure 6, stimulation with F4 fimbriae significantly increased IL-17A secretion by PBMCs as fast as 24 h post stimulation. Moreover, endotoxins in the fimbrial solution did not exert a significant effect on the IL-17A secretion (Additional file 4). F4 fimbriae also triggered the secretion of IL-22 by PBMCs, while no significant changes were observed for IL-10 and IFN- γ .

F4 fimbriae boosted Th17 responses in an antigen recall assay

To confirm the presence of circulating F4-specific lymphocytes, PBMCs from F4⁺ ETEC infected pigs were

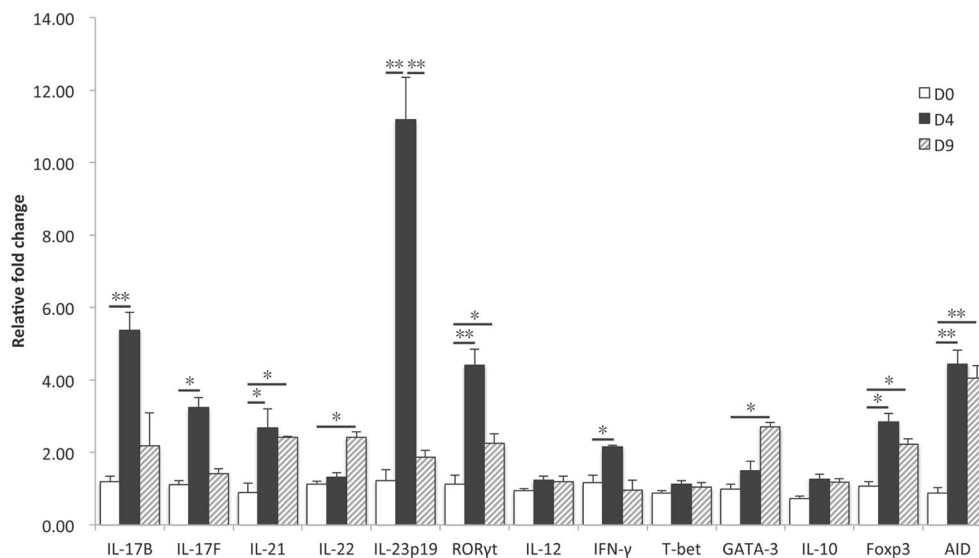


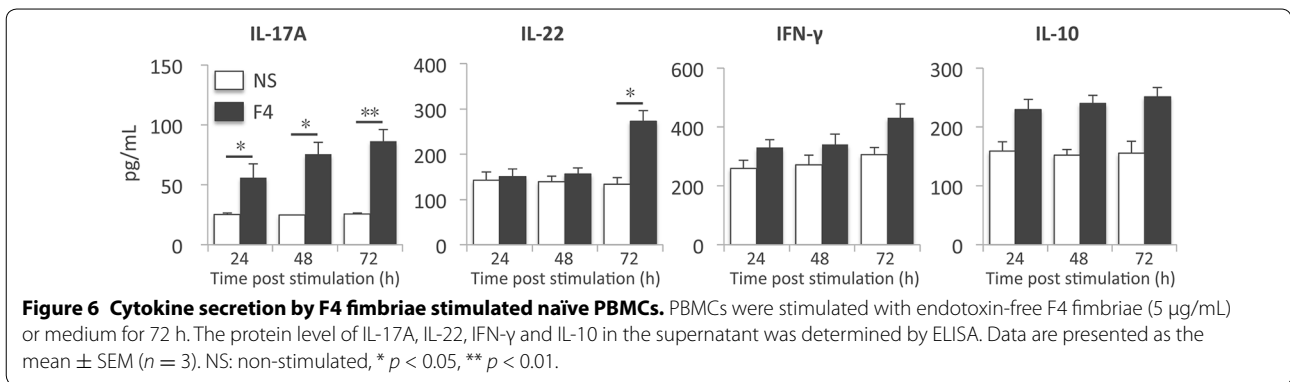
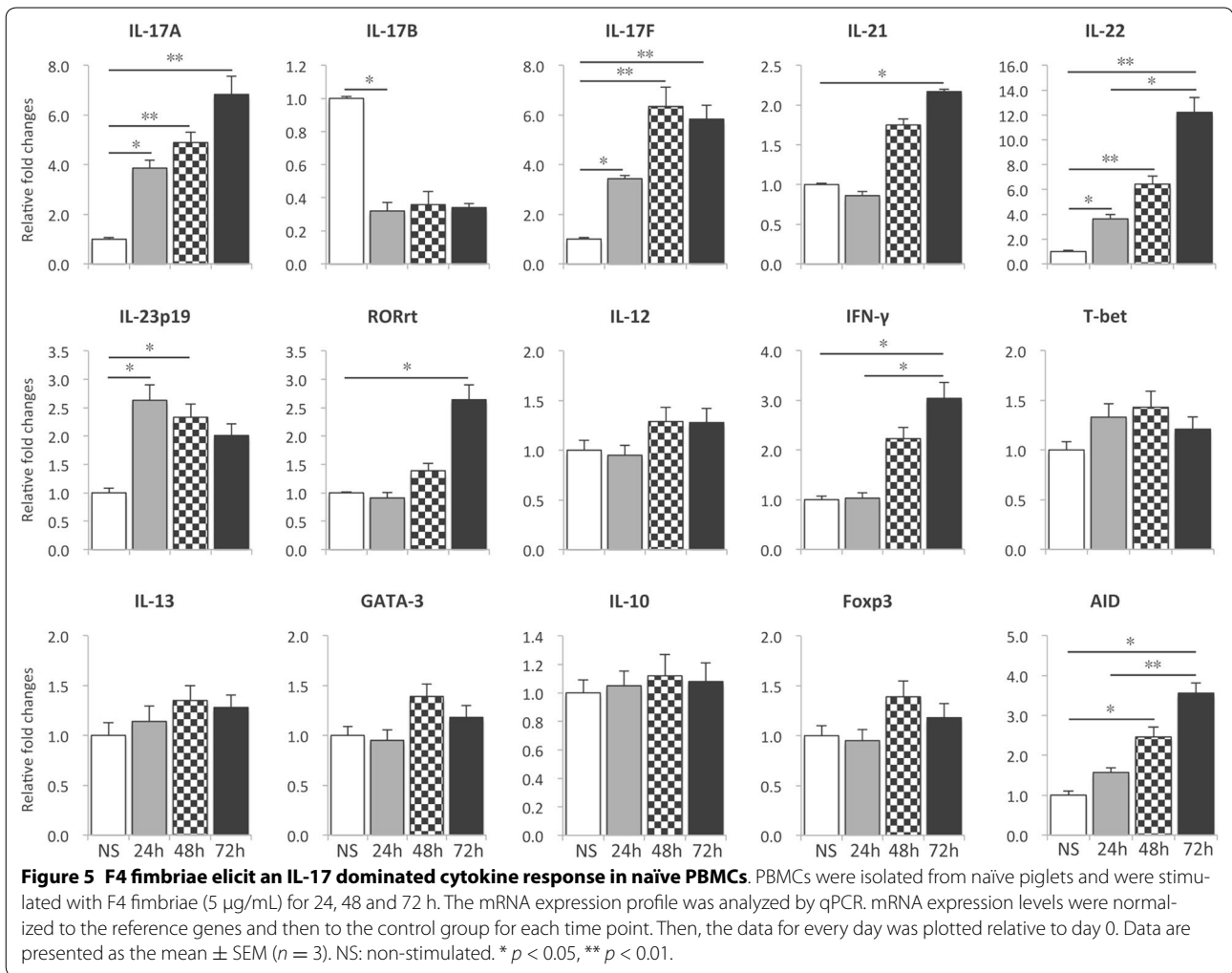
Figure 4 F4 fimbriae trigger mRNA expression of IL-17 cytokines in PBMCs upon oral immunization. The piglets were immunized with 1 mg F4 fimbriae on day 0 (D0), D1 and D2. PBMCs were isolated from piglets on D0 before immunization and on D4 and D9 after immunization. The mRNA expression in PBMCs isolated from F4-immunized or control piglets was analyzed by qPCR. The mRNA expression level was normalized to the reference genes and then to the control group for every separate day. Data are presented as the mean \pm SEM ($n = 5$ per group). * $p < 0.05$, ** $p < 0.01$.

restimulated with F4 fimbriae for 48 h and the mRNA expression levels of the above mentioned cytokines and transcription factors were assessed. As shown in Figure 7A, restimulation with the fimbriae resulted in the upregulation of all examined transcripts, except IL-13 and IL-12. The Th17-related genes IL-17A, IL-17B, IL-17E, RORyt, IL-21, IL-22 and IL-23p19 as well as AID were highly upregulated. In addition, Foxp3, IL-10, GATA-3, IFN- γ and T-bet mRNA expression levels were increased after F4 restimulation, although not to the same extent as the Th17-related cytokines. Furthermore, the upregulated IL-17A and IL-22 transcripts elicited by F4 fimbriae restimulation correlated with an augmented IL-17A and IL-22 cytokine secretion by the PBMCs, while the increase in IFN- γ and IL-10 production was not statistically significant (Figure 7B).

Discussion

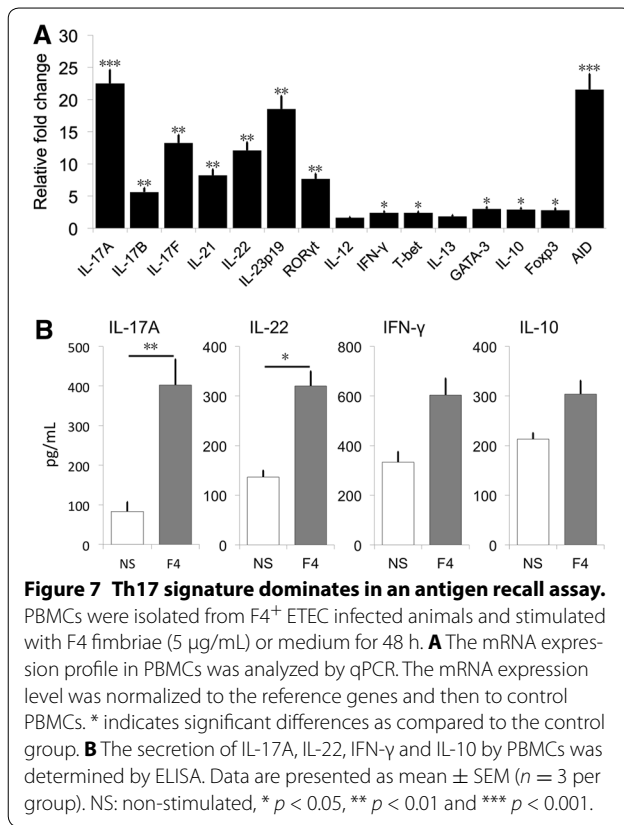
Th17 effector cells are important to eradicate mucosal pathogens including extracellular bacteria, fungi and even helminths [17, 39, 40]. These Th17 cells are characterized by the secretion of IL-17A, IL-17E, IL-21 and IL-22. The latter contribute to the protective function of Th17 cells by inducing the expression of defensins, mucins, tight junction proteins and lipopolysaccharide-binding proteins, which all reinforce the epithelial barrier [41]. Although some progress has been made to elucidate the cytokine response of intestinal epithelial cells to ETEC or its

virulence factors [42–46], only few studies have evaluated the transcriptomic profile in PBMCs and intestinal tissues upon F4⁺ ETEC infection or oral immunization with F4 fimbriae prior to this report [47]. Recent work at our laboratory first hinted at the importance of IL-17A in innate immunity targeted to fend off F4⁺ ETEC infection as IL-17A mRNA levels were upregulated in the small intestine of piglets 4 h after F4⁺ ETEC colonization [33]. Zhu and colleagues on the other hand observed a downregulated expression of IL-17A mRNA in both jejunum and ileum 1 week after F4⁺ ETEC challenge, although this downregulation did not reach the significant level. In addition, no significant changes were observed in the mRNA expression levels of IFN- γ , IL-12p40, IL-4, IL-2, IL-10, Foxp3 and TGF- β , except for an upregulated IL-6 mRNA expression [34]. As F4⁺ ETEC colonize the gut very soon upon ingestion and clearance of this pathogen usually occurs at 7 days post infection [10], any changes in cytokine expression levels should have occurred earlier. Here, we evaluated the mRNA expression profile in PBMCs from day 0 to 4 upon F4⁺ ETEC infection and at day 4 in the intestinal tissues. F4⁺ ETEC infection triggered an increased expression of IL-17A, IL-17E, IL-21, IL-22, IL-23p19 and RORyt in the intestine and PBMC fraction, hinting at a potential role of Th17 cells to clear F4⁺ ETEC infections in piglets. Indeed, IL-17A, IL-17E, IL-21 and IL-22 are preferentially produced by Th17 cells, while the transcription factor RORyt and IL-23 play essential roles in the



differentiation and expansion of Th17 cells, respectively [48, 49]. The involvement of Th17 cells is also evidenced by the occurrence of relatively large amounts of IL-17A producing T cells in the intestinal tissue of F4⁺ ETEC infected piglets. Interestingly, we also observed a strong

upregulation of IL-17F mRNA expression. IL-17A and IL17F have similar biological actions as both cytokines mediate pro-inflammatory responses and play a role in the host defense against certain mucosal pathogens, such as *C. rodentium* [50–52]. Thus, we speculate that both



IL-17A and IL-17F are required for the protection against F4⁺ ETEC infection in piglets. Further research should elucidate the contribution of each cytokine to protection against ETEC, especially as divergent roles for IL-17A and -F in immunity have been reported [53, 54]. Intriguingly, we observed a differential regulation of IL-17B mRNA expression in intestinal tissues and PBMCs upon F4⁺ ETEC infection, which may suggest IL-17B has a different function in mucosal and systemic immunity. Not much is known about IL-17B. This cytokine is expressed by monocytes and neutrophils and induces the secretion of pro-inflammatory cytokines [55–58]. Further research should identify the IL-17B-producing cells in pigs and elucidate their role in the host defense against mucosal pathogens.

In contrast to the increased level of Th17-related cytokines, F4⁺ ETEC infection did not affect the expression level of the Th1-related genes IL-12, IFN-γ and T-bet in both PBMCs and intestinal tissues, although a small increased IFN-γ expression was observed in the PBMC fraction on D3 and 4 upon infection. F4⁺ ETEC infection also resulted in an increased mRNA expression of IL-13, GATA-3, IL-10, and Foxp3, especially in the gastrointestinal tract. Upregulated Foxp3 and IL-10 mRNA expression probably indicates the induction of Tregs

during the later stages of F4⁺ ETEC infection [59]. Since ETEC infection causes inflammation and intestinal damage in piglets, the induction of Tregs is probably required to limit these responses and to avoid immunopathology due to an overwhelming Th17 immunity [60, 61]. It is worth noting that F4⁺ ETEC infection triggered a significant increase in AID mRNA expression in PBMCs, Peyer patches and MLN. AID is a B cell specific enzyme required for the class switch recombination (CSR) in activated B cells [62]. In the F4⁺ ETEC infection model, F4-specific IgG and IgA antibody-secreting cells were observed in most tissues 4 days post infection [3, 10]. Hence, the increased AID mRNA level probably reflects ongoing class switching in B cells. Moreover, the strong induction of AID mRNA expression further supports the involvement of Th17 cells, since these cells also participate in B cell differentiation and subsequent SIgA production [13, 14, 63].

Previous studies in our lab indicated the strong oral immunogenicity of F4 fimbriae [3, 7, 9, 10]. To establish whether oral immunization with F4 fimbriae could elicit similar responses as an F4⁺ ETEC infection, we analyzed the cytokine mRNA expressions in PBMCs. Similar to infection, we observed a robust expression of the Th17-related genes RORyt, IL-23p19, IL-17F, IL-21 and IL-22. Likewise, Foxp3 and AID mRNA levels were also increased from day 4 onwards. These results indicate the capacity of F4 fimbriae to elicit Th17 responses and induce class switching in B cells upon oral administration. Unexpectedly, we failed to detect IL-17A mRNA expression. Upon ETEC infection, we observed a peak expression of IL-17A mRNA at 2 days post infection and the inability to detect IL-17A could be attributed to the later sampling point in the immunization experiment or indicate an important role for other molecules such as the enterotoxins in the IL-17A mRNA expression during infection [33]. The mRNA profile in the PBMCs upon oral immunization were F4 fimbriae specific, since PBMCs from naive piglets showed an upregulated mRNA expression of Th17-related genes and an enhanced secretion of IL-17A and IL-22 upon stimulation with these fimbriae. On top of that, these responses were further amplified in the PBMCs isolated from F4⁺ ETEC infected animals upon F4 fimbriae stimulation. In contrast, all the Th1- (except IFN-γ) and Th2-related genes did not show any significant change in naive PBMCs in the first 72 h, which corroborates a previous study [64]. In addition, Th1 effector cells and Tregs appear to be less important during F4⁺ ETEC infection, since no significant antigen specific IFN-γ and IL-10 recall responses were obtained by restimulation with F4 fimbriae.

In conclusion, F4⁺ ETEC infection and oral immunization with F4 fimbriae elicited robust expression of Th17-related genes and IL-17 producing T cells, indicating that Th17 effector cells participate in the protective immunity to ETEC infection in piglets and that these Th17 responses are in part induced by F4 fimbriae. Moreover, we also hinted at the potential participation of IL-17B and IL-17F in the clearance of F4⁺ ETEC infection. Altogether, our results could facilitate the design of ETEC vaccines.

Additional files

Additional file 1. F4⁺ ETEC number detected in the faeces after F4⁺ ETEC infection. Piglets were orally inoculated with either 10¹⁰ CFU F4⁺ ETEC (Pig 4 to 6) in 10 mL PBS or only PBS (Pig 1 to 3) as control at day 0 and 1. Faecal samples were collected from day 0 to 4 and inoculated onto blood agar plates and then the number of F4-specific hemolytic *E. coli* colonies was determined by dot blotting the colonies on nitrocellulose membranes and detecting F4 fimbriae with an F4ac-specific MAb.

Additional file 2. The distribution of mRNA in intestinal tissues of control pigs. The mRNA expression in intestinal tissues of control pigs was analyzed by qPCR. The mRNA expression was normalized to the reference genes and then to jejunum without Peyer's patches. Data are presented as the mean ± SEM (*n* = 3 per group). JJ = jejunum without Peyer's patches, JP = jejunum with Peyer's patches, IL = ileum without Peyer's patches, IP = ileum with Peyer's patches, MLN = mesenteric lymph nodes. * *p* < 0.05, ** *p* < 0.01.

Additional file 3. Oral immunization with F4 fimbriae induced F4-specific serum antibodies. The piglets were immunized with 1 mg F4 fimbriae on day 0 (D0), D1 and D2. Blood was drawn on D0 before immunization and D9 after immunization. The F4-specific antibody (Ab) titer was tested in serum using ELISA. Data are presented as the mean ± SEM (*n* = 5 per group).

Additional file 4. Effect of endotoxins on IL-17A secretion. PBMCs were stimulated with endotoxin-free F4 fimbriae (5 µg/mL), F4 fimbriae (5 µg/mL) in the presence or absence of polymyxin B (PMB, 25 µg/mL) or medium for 72 h. The IL-17A concentration in the cell-free supernatant was determined by ELISA. Data are presented as the mean ± SEM (*n* = 3).

Authors' contributions

YL, BD and EC conceived and designed the experiments. YL carried out the majority of the experiments, was responsible for data collection and drafted the manuscript. YL, UVN and PYFR performed the infection experiment and YL and UVN the immunization experiment. YL, BD and EC were responsible for data analysis and interpretation. BD and EC critically revised the manuscript. All authors read and approved the final manuscript.

Author details

¹ Laboratory of Veterinary Immunology, Department of Virology, Parasitology and Immunology, Faculty of Veterinary Medicine, Ghent University, Ghent, Belgium. ² Department of Veterinary Medicine and Zootechnics, Universidad Central "Marta Abreu" de Las Villas, Carretera a Camajuani km 5½, 54830 Santa Clara, Villa Clara, Cuba.

Acknowledgements

Y Luo has a Ph.D. fellowship from the China Scholarship Council (CSC Grant; [2011] 3005) and from the Special Research Fund of Ghent University (co-funding of the CSC Grant). BD is supported by a postdoctoral grant from the FWO-Vlaanderen. We gratefully thank S. Brabant, R. Cooman, J. Lamote and K. Grauwet for their technical assistance.

Competing interests

The authors declare that they have no competing interests.

Received: 27 April 2015 Accepted: 2 October 2015

Published online: 21 October 2015

References

- Francis DH (2002) Enterotoxigenic *Escherichia coli* infection in pigs and its diagnosis. *J Swine Health Prod* 10:171–175
- Chen X, Gao S, Jiao X, Liu XF (2004) Prevalence of serogroups and virulence factors of *Escherichia coli* strains isolated from pigs with postweaning diarrhoea in eastern China. *Vet Microbiol* 103:13–20
- Van den Broeck W, Cox E, Goddeeris B (1999) Induction of immune responses in pigs following oral administration of purified F4 fimbriae. *Vaccine* 17:2020–2029
- Wang J, Jiang SW, Chen XH, Liu ZL, Peng J (2006) Prevalence of fimbrial antigen (K88 variants, K99 and 987P) of enterotoxigenic *Escherichia coli* from neonatal and post-weaning piglets with diarrhea in central China. *Asian Aust J Anim Sci* 19:1342–1346
- Fairbrother JM, Nadeau E, Gyles CL (2005) *Escherichia coli* in postweaning diarrhea in pigs: an update on bacterial types, pathogenesis, and prevention strategies. *Anim Health Res Rev* 6:17–39
- Frydendahl K (2002) Prevalence of serogroups and virulence genes in *Escherichia coli* associated with postweaning diarrhoea and edema disease in pigs and a comparison of diagnostic approaches. *Vet Microbiol* 85:169–182
- Verdonck F, Joensuu JJ, Stuyven E, De Meyer J, Muilu M, Pirhonen M, Goddeeris BM, Mast J, Niklander-Teeri V, Cox E (2008) The polymeric stability of the *Escherichia coli* F4 (K88) fimbriae enhances its mucosal immunogenicity following oral immunization. *Vaccine* 26:5728–5735
- Devriendt B, Verdonck F, Summerfield A, Goddeeris BM, Cox E (2010) Targeting of *Escherichia coli* F4 fimbriae to Fcγ receptors enhances the maturation of porcine dendritic cells. *Vet Immunol Immunopathol* 135:188–198
- Van den Broeck W, Cox E, Goddeeris B (1999) Receptor-dependent immune responses in pigs after oral immunization with F4 fimbriae. *Infect Immun* 67:520–526
- Verdonck F, Cox E, Gog K, Stede Y, Duchateau L, Deprez P, Goddeeris B (2002) Different kinetic of antibody responses following infection of newly weaned pigs with an F4 enterotoxigenic *Escherichia coli* strain or an F18 verotoxigenic *Escherichia coli* strain. *Vaccine* 20:2995–3004
- Verdonck F, Cox E, Vancaeneghem S, Goddeeris BM (2004) The interaction of F4 fimbriae with porcine enterocytes as analysed by surface plasmon resonance. *FEMS Immunol Med Microbiol* 41:243–248
- Cerutti A (2008) The regulation of IgA class switching. *Nat Rev Immunol* 8:421–434
- Cao AT, Yao S, Gong B, Elson CO, Cong Y (2012) Th17 cells upregulate polymeric Ig receptor and intestinal IgA and contribute to intestinal homeostasis. *J Immunol* 189:4666–4673
- Hirota K, Turner JE, Villa M, Duarte JH, Demengeot J, Steinmetz OM, Stockinger B (2013) Plasticity of TH17 cells in Peyer's patches is responsible for the induction of T cell-dependent IgA responses. *Nat Immunol* 14:372–379
- Cao AT, Cong Y Control of intestinal IgA by Th17 cells. http://www.utmb.edu/mclaughlin/coll_12/cao-predoc.pdf
- Conti HR, Shen F, Nayyar N, Stocum E, Sun JN, Lindemann MJ, Ho AW, Hai JH, Yu JJ, Jung JW, Filler SG, Masso-Welch P, Edgerton M, Gaffen SL (2009) Th17 cells and IL-17 receptor signaling are essential for mucosal host defense against oral candidiasis. *J Exp Med* 206:299–311
- Khader SA, Gaffen SL, Kolls JK (2009) Th17 cells at the crossroads of innate and adaptive immunity against infectious diseases at the mucosa. *Mucosal Immunol* 2:403–411
- Mayuzumi H, Inagaki-Ohara K, Uyttenhove C, Okamoto Y, Matsuzaki G (2010) Interleukin-17A is required to suppress invasion of *Salmonella enterica* serovar Typhimurium to enteric mucosa. *Immunology* 131:377–385
- Chen K, Fisher C, Zheng MQ, Weaver C, Kolls JK (2011) Immunization of heat-killed *Klebsiella pneumoniae* promotes strong mucosal Th17 responses and protects mice from live infection (Abstract). *J Immunol* 186:59.4

20. Dreesen L, De Bosscher K, Grit G, Staels B, Lubberts E, Bauge E, Geldhof P (2014) *Giardia muris* infection in mice is associated with a protective interleukin 17A response and induction of peroxisome proliferator-activated receptor alpha. *Infect Immun* 82:3333–3340
21. Luckheeram RV, Zhou R, Verma AD, Xia B (2012) CD4⁺T cells differentiation and functions. *Clin Dev Immunol* 2012:925135
22. Zhu J, Yamane H, Paul WE (2010) Differentiation of effector CD4 T cell populations. *Annu Rev Immunol* 28:445–489
23. Korn T, Bettelli E, Oukka M, Kuchroo VK (2009) IL-17 and Th17 Cells. *Annu Rev Immunol* 27:485–517
24. Jin W, Dong C (2013) IL-17 cytokines in immunity and inflammation. *Emerg Microb Infect* 2:e60
25. Iwakura Y, Ishigame H, Saijo S, Nakae S (2011) Functional specialization of interleukin-17 family members. *Immunity* 34:149–162
26. Angkasekwinai P, Dong C (2011) TH17 cytokines: characteristics, regulation, and biological function. In: Jiang SP (ed) TH17 cells in health and disease. Springer, London, pp 27–40
27. Fallon PG, Ballantyne SJ, Mangan NE, Barlow JL, Dasvarma A, Hewett DR, McIlgorm A, Jolin HE, McKenzie AN (2006) Identification of an interleukin (IL)-25-dependent cell population that provides IL-4, IL-5, and IL-13 at the onset of helminth expulsion. *J Exp Med* 203:1105–1116
28. Herberth G, Daegelmann C, Röder S, Behrendt H, Krämer U, Borte M, Heinrich J, Herbarth O, Lehmann I, LISAplus study group (2010) IL-17E but not IL-17A is associated with allergic sensitization: results from the LISA study. *Pediatr Allergy Immunol* 21:1086–1190
29. Friedrich M, Diegelmann J, Schaubert J, Auernhammer CJ, Brand S (2015) Intestinal neuroendocrine cells and goblet cells are mediators of IL-17A-amplified epithelial IL-17C production in human inflammatory bowel disease. *Mucosal Immunol* 8:943–958
30. Kiros TG, van Kessel J, Babiuk LA, Gerds V (2011) Induction, regulation and physiological role of IL-17 secreting helper T-cells isolated from PBMC, thymus, and lung lymphocytes of young pigs. *Vet Immunol Immunopathol* 144:448–454
31. Leach S, Clements JD, Kaim J, Lundgren A (2012) The adjuvant double mutant *Escherichia coli* heat labile toxin enhances IL-17A production in human T cells specific for bacterial vaccine antigens. *PLoS One* 7:e51718
32. Sjökvist Ottsjö L, Flach CF, Clements J, Holmgren J, Raghavan S (2013) A double mutant heat-labile toxin from *Escherichia coli*, LT(R192G/L211A), is an effective mucosal adjuvant for vaccination against *Helicobacter pylori* infection. *Infect Immun* 81:1532–1540
33. Loos M, Geens M, Schauvliege S, Gasthuys F, van der Meulen J, Dubreuil JD, Goddeeris BM, Niewold T, Cox E (2012) Role of heat-stable enterotoxins in the induction of early immune responses in piglets after infection with enterotoxigenic *Escherichia coli*. *PLoS One* 7:e41041
34. Zhu Y-HH, Li X-QQ, Zhang W, Zhou D, Liu HY, Wang JF (2014) Dose-dependent effects of *Lactobacillus rhamnosus* on serum interleukin-17 production and intestinal T-cell responses in pigs challenged with *Escherichia coli*. *Appl Environ Microbiol* 80:1787–1798
35. Nguyen VU, Goetstouwers T, Coddens A, Van Poucke M, Peelman L, Deforce D, Melkebeek V, Cox E (2013) Differentiation of F4 receptor profiles in pigs based on their mucin 4 polymorphism, responsiveness to oral F4 immunization and in vitro binding of F4 to villi. *Vet Immunol Immunopathol* 152:93–100
36. Jamalludeen N, Johnson RP, Shewen PE, Gyles CL (2009) Evaluation of bacteriophages for prevention and treatment of diarrhea due to experimental enterotoxigenic *Escherichia coli* O149 infection of pigs. *Vet Microbiol* 136:135–141
37. Vandesompele J, De Preter K, Pattyn F, Poppe B, Van Roy N, De Paeppe A, Speleman F (2002) Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. *Genome Biol* 3:RESEARCH0034
38. Livak KJ, Schmittgen TD (2001) Analysis of relative gene expression data using real-time quantitative PCR and the 2^{-(Delta Delta C(T))} Method. *Methods* 25:402–408
39. Blaschitz C, Raffatellu M (2010) Th17 cytokines and the gut mucosal barrier. *J Clin Immunol* 30:196–203
40. Bouchery T, Kyle R, Ronchese F, Gros G (2014) The differentiation of CD4⁺ T-helper cell subsets in the context of helminth parasite infection. *Front Immunol* 5:487
41. Bogaert S, Laukens D, Peeters H, Melis L, Olievier K, Boon N, Verbruggen G, Vandesompele J, Elewaut D, De Vos M (2010) Differential mucosal expression of Th17-related genes between the inflamed colon and ileum of patients with inflammatory bowel disease. *BMC Immunol* 11:61
42. Pavlova B, Volf J, Alexa P, Rychlik I, Matiasovic J, Faldyna M (2008) Cytokine mRNA expression in porcine cell lines stimulated by enterotoxigenic *Escherichia coli*. *Vet Microbiol* 132:105–110
43. Devriendt B, Stuyven E, Verdonck F, Goddeeris BM, Cox E (2010) Enterotoxigenic *Escherichia coli* (K88) induce proinflammatory responses in porcine intestinal epithelial cells. *Dev Comp Immunol* 34:1175–1182
44. Geens MM, Niewold TA (2010) Preliminary Characterization of the Transcriptional Response of the Porcine Intestinal Cell Line IPEC-J2 to Enterotoxigenic *Escherichia coli*, *Escherichia coli*, and *E. coli* Lipopolysaccharide. *Comp Funct Genomics* 2010:469583
45. Zanello G, Berri M, Dupont J, Sizart P-YY, D'Inca R, Salmon H, Meurens F (2011) *Saccharomyces cerevisiae* modulates immune gene expressions and inhibits ETEC-mediated ERK1/2 and p38 signaling pathways in intestinal epithelial cells. *PLoS One* 6:e18573
46. Zhou C, Liu Z, Jiang J, Yu Y, Zhang Q (2012) Differential gene expression profiling of porcine epithelial cells infected with three enterotoxigenic *Escherichia coli* strains. *BMC Genomics* 13:330
47. Delisle B, Calinescu C, Mateescu MA, Fairbrother JM, Nadeau É (2012) Oral immunization with f4 fimbriae and CpG formulated with carboxymethyl starch enhances f4-specific mucosal immune response and modulates Th1 and Th2 cytokines in weaned pigs. *J Pharm Pharm Sci* 15:642–656
48. Ouyang W, Kolls JK, Zheng Y (2008) The biological functions of T helper 17 cell effector cytokines in inflammation. *Immunity* 28:454–467
49. McGeachy MJ, Cua DJ (2008) Th17 cell differentiation: the long and winding road. *Immunity* 28:445–453
50. Chang SH, Dong C (2009) IL-17F: regulation, signaling and function in inflammation. *Cytokine* 46:7–11
51. Sabat R, Witte E, Witte K, Wolk K (2013) IL-22 and IL-17: An Overview. In: Quesniaux V, Ryffel V, Di Padova F (eds) IL-17, IL-22 and their producing cells: role in inflammation and autoimmunity progress in inflammation research. Springer, London, pp 11–35
52. Kolls JK, Lindén A (2004) Interleukin-17 family members and inflammation. *Immunity* 21:467–476
53. Yang XO, Chang SH, Park H, Nurieva R, Shah B, Acero L, Wang YH, Schluns KS, Broadbent RR, Zhu Z, Dong C (2008) Regulation of inflammatory responses by IL-17F. *J Exp Med* 205:1063–1075
54. Ishigame H, Kakuta S, Nagai T, Kadoki M, Nambu A, Komiyama Y, Fujikado N, Tanahashi Y, Akitsu A, Kotaki H, Sudo K, Nakae S, Sasakawa C, Iwakura Y (2009) Differential roles of interleukin-17A and -17F in host defense against mucocutaneous bacterial infection and allergic responses. *Immunity* 30:108–119
55. Li H, Chen J, Huang A, Stinson J, Heldens S, Foster J, Dowd P, Gurney AL, Wood WI (2000) Cloning and characterization of IL-17B and IL-17C, two new members of the IL-17 cytokine family. *Proc Natl Acad Sci U S A* 97:773–778
56. Moore EE, Presnell S, Garrigues U, Guilbot A, LeGuern E, Smith D, Yao L, Whitmore TE, Gilbert T, Palmer TD, Horner PJ, Kuestner RE (2002) Expression of IL-17B in neurons and evaluation of its possible role in the chromosome 5q-linked form of Charcot-Marie-Tooth disease. *Neuromuscul Disord* 12:141–150
57. Kokubu T, Haudenschild DR, Moseley TA, Rose L, Reddi AH (2008) Immunolocalization of IL-17A, IL-17B, and their receptors in chondrocytes during fracture healing. *J Histochem Cytochem* 56:89–95
58. Kouri VP, Olkkonen J, Ainola M, Li TF, Björkman L, Kontinen YT, Mandelin J (2014) Neutrophils produce interleukin-17B in rheumatoid synovial tissue. *Rheumatology (Oxford)* 53:39–47
59. Käser T, Gerner W, Hammer SE, Patzl M, Saalmüller A (2008) Phenotypic and functional characterisation of porcine CD4⁺CD25^{high} regulatory T cells. *Vet Immunol Immunopathol* 122:153–158
60. Couper KN, Blount DG, Riley EM (2008) IL-10: the master regulator of immunity to infection. *J Immunol* 180:5771–5777
61. Nyachoti CM, Kiarie E, Bhandari SK, Zhang G, Krause DO (2012) Weaned pig responses to *Escherichia coli* K88 oral challenge when receiving a lysozyme supplement. *J Anim Sci* 90:252–260
62. Delker RK, Fugmann SD, Papavasiliou NF (2009) A coming-of-age story: activation-induced cytidine deaminase turns 10. *Nat Immunol* 10:1147–1153

63. Barbosa RR, Silva SP, Silva SL, Melo AC, Pedro E, Barbosa MP, Pereira-Santos MC, Victorino RMM, Sousa AE (2011) Primary B-cell deficiencies reveal a link between human IL-17-producing CD4 T-cell homeostasis and B-cell differentiation. *PLoS One* 6:e22848
64. Verfaillie T, Cox E, To L, Vanrompay D, Bouchaut H, Buys N, Goddeeris B (2001) Comparative analysis of porcine cytokine production by mRNA and protein detection. *Vet Immunol Immunopathol* 81:97–112
65. Melkebeek V, Verdonck F, Goddeeris BM, Cox E (2007) Comparison of immune responses in parenteral FaeG DNA primed pigs boosted orally with F4 protein or reimmunized with the DNA vaccine. *Vet Immunol Immunopathol* 116:199–214
66. Raymond CR, Wilkie BN (2004) Th-1/Th-2 type cytokine profiles of pig T-cells cultured with antigen-treated monocyte-derived dendritic cells. *Vaccine* 22:1016–1023
67. Devriendt B, Gallois M, Verdonck F, Wache Y, Bimczok D, Oswald IP, Goddeeris BM, Cox E (2009) The food contaminant fumonisin B(1) reduces the maturation of porcine CD11R1(+) intestinal antigen presenting cells and antigen-specific immune responses, leading to a prolonged intestinal ETEC infection. *Vet Res* 40:40
68. Verhelst D, De Craeye S, Dorny P, Melkebeek V, Goddeeris B, Cox E, Jongert E (2011) IFN- γ expression and infectivity of *Toxoplasma* infected tissues are associated with an antibody response against GRA7 in experimentally infected pigs. *Vet Parasitol* 179:14–21
69. Pilon C, Levast B, Meurens F, Le Vern Y, Kerboeuf D, Salmon H, Velge-Roussel F, Lebranchu Y, Baron C (2009) CD40 engagement strongly induces CD25 expression on porcine dendritic cells and polarizes the T cell immune response toward Th1. *Mol Immunol* 46:437–447
70. von der Hardt K, Kandler MA, Fink L, Schoof E, Dötsch J, Brandenstein O, Bohle RM, Rascher W (2004) High frequency oscillatory ventilation suppresses inflammatory response in lung tissue and microdissected alveolar macrophages in surfactant depleted piglets. *Pediatr Res* 55:339–346

**Submit your next manuscript to BioMed Central
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

