

# Identification and characterization of tebuconazole transformation products in soil by combining suspect screening and molecular typology

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21	assessment.

## 23 ABSTRACT

24 Once released into the environment, pesticides generate transformation products (TPs) which may be of (eco-)toxicological importance. Past studies have demonstrated the difficulty to 25 predict pesticide TP occurrence and their environmental risk by monitoring-driven approaches 26 mostly used in current regulatory frameworks targeting only known toxicologically relevant 27 TPs. We present a novel combined approach which identifies and categorizes known and 28 unknown pesticide TPs in soil by combining suspect screening time-of-flight mass 29 spectrometry with in silico molecular typology. This approach applies an empirical and 30 theoretical pesticide TP library for compound identification by both non-target and target 31 32 time-of-flight (tandem) mass spectrometry and structural elucidation through a molecular structure correlation program. In silico molecular typology was then used to group the 33 detected TPs according to common molecular descriptors and to indirectly elucidate their 34 35 environmental properties by analogy to known pesticide compounds having similar molecular descriptors. This approach was evaluated via the identification of TPs of the triazole fungicide 36 tebuconazole occurring in a field dissipation study. Overall, 22 empirical and 12 yet unknown 37 TPs were detected and categorized into three groups with defined environmental properties. 38 This approach combining suspect screening time-of-flight mass spectrometry with molecular 39 typology could be extended to other organic pollutants and used to rationalize the choice of 40 TPs to be intensively studied towards a more comprehensive environmental risk assessment 41 scheme. 42

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# 44 CAPSULE ABSTRACT

Combination of time-of-flight mass spectrometry and molecular typology was shown to be of
fundamental interest to identify and categorize tebuconazole transformation products in soil.

# 48 INTRODUCTION

Pesticides, used in agriculture to ensure crop yield and quality, constitute a large group of 49 xenobiotic compounds in the environment.<sup>1,2</sup> They and their transformation products (TPs) 50 have globally been identified as major contaminants of natural water resources. The United 51 States Environmental Protection Agency (U.S. EPA) estimates a global release of 2.4 billion 52 kilograms pesticides per year supporting important economic sectors of agriculture but also 53 causing major environmental pollution.<sup>3</sup> Another aspect which makes pesticides a chemical 54 group of particular interest is their rapidly changing and evolving market due to the 55 continuous introduction of new molecules. In order to reach the market, new pesticide 56 molecules have to undergo an environmental risk assessment (ERA) procedure based on a 57 large set of data submitted by the pesticide companies and evaluated by the official authorities 58 (e.g. the Plant Protection Products and their Residues panel<sup>4</sup> of the European Food Safety 59 60 Authority for Europe (EFSA)). This procedure is periodically repeated in order to re-evaluate the already registered pesticide molecules. Once released into the agro-ecosystem, pesticides 61 and their TPs can diffuse away from their target point, contaminate surrounding water 62 resources<sup>5</sup> and impact non-target organisms supporting key ecosystemic services.<sup>6</sup> The 63 properties of TPs such as bioavailability, mobility and ecotoxicity can be substantially 64 different from the parent compound<sup>7,8</sup> and although their biological efficacy is usually lower,<sup>9</sup> 65 they could be highly (eco-)toxicologically relevant.<sup>10</sup> In Europe, *a posteriori* monitoring of 66 the contamination of water and air resources by pesticides is imposed by corresponding 67 directives.<sup>11,12</sup> In contrast, no monitoring of soil contamination by pesticides is required at EU 68 level. This is the result of the lack of a corresponding directive, although the need for a 69 regulation was proposed to the European Commission.<sup>13</sup> Thus, if an unacceptable risk for the 70 environment becomes evident several years after the introduction of a pesticide to the market, 71 restrictions in its use are imposed and if monitoring studies reveal that no alleviation of the 72

risk was achieved, the pesticide is taken off the market. Past experience has shown that the 73 knowledge of the existence of (eco-)toxicologically relevant TPs typically emerges only about 74 20 to 30 years after the introduction of a given pesticide to the market.<sup>10</sup> Currently used 75 monitoring-driven approaches for pinpointing environmentally and (eco-)toxicologically 76 relevant pesticide TPs often concentrate on water samples and do not prevent the 77 contamination of natural resources and impose mitigation actions only after contamination is 78 79 reported. Novel approaches able to identify potential TPs and *in silico* assessment of their toxicity and environmental contamination could be part of the solution of this issue. 80

The gap of knowledge about the existence or properties of pesticide TPs derived by abiotic 81 and biotic processes in the environment may be the reason why relevant TPs have been 82 overlooked for so long. Conventional analytical methods focus on the monitoring of the so-83 defined '(eco-)toxicologically relevant' pesticide TPs or TPs 'relevant for groundwater 84 contamination'.<sup>4,14</sup> Consequently, most studies focus on a limited number of molecules which 85 have been identified as targets by monitoring-driven approaches and are calibrated against 86 analytical standards. Therefore, they often provide only a partial view of the complex 87 transformation patterns of pesticides in the environment. Thus, there is a need to develop an 88 innovative approach by combining powerful analytical methods detecting also unknown TPs 89 90 with categorization methods based on the chemical features of the detected TPs.

The analytical techniques suitable to detect unknown and suspected molecules in the absence of analytical standards are high resolution mass spectrometry (MS), namely quadrupole timeof-flight MS (QTOF-MS) and Fourier transform MS (FT-MS), as well as nuclear magnetic resonance (NMR) spectroscopy.<sup>15</sup> These techniques are often combined in forensics<sup>16</sup> or medicine,<sup>17</sup> where QTOF-MS is used for screening for new unknown molecules and then is supplemented by NMR spectra for the structural elucidation of a compound. However, NMR spectroscopy is an effective method for the confirmation of molecular structures but is rather

less appropriate for the analysis of TPs in environmental samples due to its rather poor
sensitivity, which generally is three orders of magnitude lower than MS.<sup>15</sup>

Categorization of unknown and suspected molecules can be achieved by *in silico* molecular
 typology that enables the property-related clustering of potential TPs.<sup>18</sup>

In this study, we present a new approach for the *a posteriori* analysis of agricultural soil 102 samples treated with a pesticide by combining high-resolution QTOF-MS with in silico 103 molecular typology. Our approach enables profiling and molecular structure elucidation of 104 105 unknown and suspected pesticide TPs in agricultural soil by the use of a compound library containing empirical (already known) and theoretical (yet unknown) TPs. The proposed 106 approach consists of three principal steps: (i) construction of an in-house pesticide-specific TP 107 library based on literature survey and establishment of suspect TPs, (ii) QTOF-MS analyses 108 of soil extracts by non-target MS to screen TPs against this library, followed by target tandem 109 110 MS using an ion inclusion list to propose their structure via accurate mass daughter ions and a molecular structure correlation (MSC) program and (iii) categorization of detected TPs based 111 on *in silico* analysis using molecular typology. 112

113 To demonstrate the potential of this novel approach, the dissipation of the triazole fungicide tebuconazole (TCZ) was investigated in a field dissipation study. TCZ was chosen as a model 114 compound because, despite being on the market for quite some time (27 years),<sup>19</sup> there are 115 still significant knowledge gaps regarding its environmental fate, its transformation in soil and 116 the ecotoxicological impact of potential TPs.<sup>20</sup> The potential formation of various triazole TPs 117 known to be recalcitrant to biodegradation and to possibly interact with the hormone 118 regulation of non-target organisms by inhibition of the cytochrome P450-dependent 119 conversion of lanosterol to ergosterol is particularly relevant.<sup>21,22</sup> The applied approach is of 120 interest for improving future environmental fate studies to become more comprehensive, thus 121 strengthening ERA. 122

#### **123 MATERIAL AND METHODS**

# 124 Library setup of TCZ TPs

An in-house TCZ TP library was constructed based on a thorough literature review including 125 research papers (cited in the results section entitled 'Empirical TCZ TPs') and data sets of the 126 pesticide company on the transformation pattern of TCZ in various matrices. Search was done 127 by means of 'Web of Science', 'Google' and others using search terms like 'tebuconazole', 128 'metabolite', 'transformation product', 'degradation' and others. The developed library was 129 composed of 47 empirical (already detected in at least one study) and 29 theoretical TPs. The 130 latter were created based on expert knowledge taking into account common reaction processes 131 132 in organic chemistry and biochemistry likely leading to probable molecules. The TCZ TP library contains the name, molecular formula, molecular structure and mass of each molecule 133 (Supporting Information, table S1). Initial pretests of chemical analysis were carried out on a 134 subset of soil samples randomly chosen from the field study to search for TCZ TPs. Retention 135 times of TPs proposed via MSC were then added to the library as an additional parameter for 136 identification purposes in MS screening. 137

#### **TCZ field dissipation study**

The field dissipation study was conducted on a North Italian field (45°04'80''N, 09°45'60''E) 139 in autumn/winter 2013/14. The field soil was characterized as loamy sand (4.2 % clay, 13.5 % 140 silt, 82.3 % sand) with a pH of 7.3 and 4.5 % of organic matter. It was not treated with 141 pesticides for more than five years and cropped with winter wheat. A commercial formulation 142 of TCZ (Folicur SE, Bayer Garden) was applied on field plots of 60 m<sup>2</sup> each at 2.5 kg TCZ/ha 143 (5x the recommended dose).<sup>23</sup> Soil samples were taken at 0, 3, 7, 21, 35, 56, 70, 105 and 125 144 days after pesticide application according to the ISO standard.<sup>24</sup> For each plot and each 145 sampling date, a composite soil sample was prepared from 10 sub-samples collected with an 146 iron cylinder of 5 cm diameter, taking a soil depth of 5 cm. Soil samples were manually 147

homogenized, sieved (5 mm) and kept frozen at -20°C until further processing. The field 148 dissipation study was part of a larger study investigating field dissipation of three model 149 pesticides including TCZ at various application rates (four plots are used in this study: two 150 non-treated control plots and two TCZ 5x dose plots). The field dissipation of TCZ was 151 monitored in soil extracts via high performance liquid chromatography coupled to photodiode 152 array detector (HPLC-PDA) and followed a biphasic pattern with a half-life ( $DT_{50}$ ) of 2.5 153 days, as calculated with a Hockey-stick kinetic model.<sup>25</sup> After 125 days, 14.2 % of applied 154 TCZ were remaining in the investigated soil layer. 155

# 156 TCZ and TP extraction from soil

157 TCZ and its related TPs were extracted with acetone from 40 g of soil as explained in the 158 Supporting Information (SI-2). The final obtained residues were re-dissolved in 1 mL of 159 70 v% methanol and 30 v% ultrapure water containing 0.01 % phosphoric acid and used for 160 chemical analysis. The extraction efficiency for TCZ was 86.36 %  $\pm$  0.45 %.

# 161 UHPLC-ESI-QTOF MS analysis

In order to investigate TCZ TPs, samples were first analyzed by non-target UHPLC-ESI-162 QTOF mass spectrometry. TCZ TPs were primarily identified by screening of the MS-only 163 raw data against the TP library, using accurate mass, isotope spacing and isotope ratio for 164 identification. A difference in accurate mass of 5 ppm, together with an isotopic pattern 165 overall score higher than 85 % were used as cutoff values. Compounds that passed both 166 thresholds, that had a plausible peak chromatogram feature and that were not detected in the 167 controls (non-treated soil samples), were chosen for further analysis by target UHPLC-ESI-168 QTOF tandem MS. For this purpose, the three most abundant precursor ions (generated 169 during MS-only) and the retention time (RT) of each MS-only detected TP were exported to 170 be included into a target list for tandem MS. Method details are explained in the Supporting 171 Information (SI-3). 172

#### 173 Molecular structure correlation

Elucidation of the molecular structure of compounds detected by tandem MS was obtained 174 through a molecular structure correlation (MSC) program (Agilent MassHunter Molecular 175 Structure Correlator B.05.00). The software correlated accurate mass daughter ions for a 176 compound of interest with the molecular structures available in the in-house custom library by 177 using a 'systematic bond-breaking' approach as described by Hill and Mortishire-Smith.<sup>26</sup> 178 Briefly, an overall correlation score was generated by taking into account the individual 179 scores from each daughter ion, the mass accuracy of these fragments, and the overall 180 percentage of ion intensities being plausibly explained with substructures. 181

# 182 Molecular typology for categorization of detected TCZ TPs

TCZ and its detected TPs were categorized using the molecular typology approach 'TyPol' 183 (Typology of Pollutants) proposed by Servien et al. (2014) allowing clustering of chemicals 184 based on the use of molecular descriptors and of environmental parameters.<sup>18</sup> Environmental 185 parameters of TPs of TCZ being unknown, clustering was only done using a set of 40 186 molecular descriptors. After clustering, TyPol was used to estimate the environmental 187 parameters (vapor pressure, dissipation half-life, sorption coefficient and bioconcentration 188 factor) of TCZ TP clusters by analogy to reference compounds for which environmental 189 190 parameters are known. Details about the method are given in the Supporting Information (SI-4). 191

#### 192 **RESULTS AND DISCUSSION**

# 193 Empirical TCZ TPs

Literature review revealed that studies reporting TCZ dissipation were often ignoring potential TP occurrence and were mainly following targeted analytical approaches.<sup>27,28</sup> It is noteworthy that newly discovered TPs detected by non-target approaches were observed during degradation of TCZ by bacterial or fungal cultures,<sup>29</sup> as photodegradation products,<sup>30</sup> in field soil studies<sup>31,3</sup>, in soil microcosm studies<sup>33,34</sup> and in fruits.<sup>35</sup> Additional information
was found in regulatory documents mainly citing unpublished or confidential studies
reporting TPs in soil and crops partially based on MS and NMR spectroscopy.<sup>36,37,38</sup>
Altogether, literature review led to the construction of a library containing 47 empirical and
29 theoretical TCZ TPs (Supporting Information, table S1) which was used for TCZ TP
suspect screening in soil samples from the field dissipation study described before.

#### 204 Detection of TCZ TPs

The QTOF-MS analyses of soil extracts gained from the field dissipation study led to the 205 detection of the parent compound TCZ, as well as of 22 empirical and 12 yet unknown TPs, 206 207 which were contained in the library. Among the 34 detected features, some of them showed the same exact mass and retention times. 12 TPs passed all steps of identification from non-208 target MS-only identification to target tandem MS and structure correlation. Three TPs passed 209 only the first two steps (selected by tandem MS but not further identified through structural 210 correlation) and 19 TPs were only detected by MS identification with plausible chromatogram 211 features, but were not further selected by tandem MS due to either method restrictions or used 212 selection criteria or molecular characteristics (e.g. low mass, high volatility,...). Table 1 213 summarizes the data of all detected TCZ TPs, reporting compound name, molecular structure, 214 mass, reference if empirical, and retention time of each TP. Moreover, the time points are 215 provided (expressed in days after the application of the TCZ treatment) at which the TCZ TPs 216 were detected and information regarding the confidence of identification. TPs are coded 217 according to their level of identification confidence (a and orange hooklet: detected by MS-218 only; a,b and yellow hooklet: detected by MS and selected by tandem MS; a,b,c and green 219 hooklet: detected by MS, selected by tandem MS and identified by MSC). According to 220 Schymanski et al. (2014) level 'a' (MS-only detection) could be assigned to 'level 4 -221 identification confidence by unequivocal molecular formula'; level 'a,b' (selection by tandem 222

MS) to 'level 3 - tentative candidate - identification confidence by structure, substituent and 223 class'; level 'a,b,c' (structure proposition by MSC) to 'level 2,b - probable structure -224 identification confidence by diagnostic evidence'.<sup>39</sup> The molecular formulas of compounds 225 that have only been detected by non-target MS but were not further selected by target tandem 226 MS are rather unlikely to be false positives, as they (i) were not detected in the controls, (ii) 227 have plausible chromatogram features and isotopic patterns and (iii) do not differ in RT 228 229 between replicates and different time points. However, their molecular structure rests tentative as the compound was not selected by tandem MS and as the molecular structure elucidation 230 by MSC is only possible based on the detected product ions produced during tandem MS. 231 232 More likely, reasons for the impossibility of selection by target tandem MS could be either peak saturation at high concentrations (as it has been seen for the parent molecule TCZ when 233 injected at high volumes), or a high sensitivity of compounds present at low concentrations, or 234 235 too low masses of the product ions or a high volatility of the precursor. Indeed, electron spray ionization (ESI) conditions and the chosen collision energies have to be efficient enough to 236 produce precursor and product ions while avoiding fragmenting the precursor into too small 237 product ions, as the minimum m/z value was chosen to be 60 due to noise exclusion. 238 Regarding low m/z values, it must be mentioned that the low molecular weight of putative 239 TPs not detected by the non-target MS-only screening such as the well-known triazole 240 fungicide TP 1-H-1,2,4-triazole (TP 63)<sup>35-38</sup> may be considered as false negatives as they may 241 not be efficiently ionized by ESI due to their volatility at the high temperature of the used ion 242 source. Nevertheless, it should be noted that a compound, which was detected by MS-only 243 and selected by target tandem MS but whose molecular structure could not be proposed by 244 MSC, might be correct in its molecular formula but might have a molecular structure not 245 present in the TP library. Additionally, the poor observed correlation for the low molecular 246 weight TPs might be affected by the formation of product ions, which were below the m/z 247

threshold and thus were missing for MSC. Strictly, all the detected molecules by our approach
still remain inferred until confirmed by retention time and fragmentation pattern of pure
standards, which are not available on the market for most of them.

251 Despite of this weakness, the benefits of suspect screening coupled to MSC for the elucidation of molecular structures represent a powerful alternative when standards are not available. 252 These benefits can be illustrated by the example of TPs 13 and 15, which are constitutional 253 254 isomers. As they have the same molecular formula and mass, they could not be discriminated by MS-only and tandem MS (Supporting Information, figure S1) even using MS instruments 255 with higher resolution power. MSC was able to correlate the product ions to the respective 256 257 precursor for elucidation of their molecular structure (Supporting Information, figure S2) and hence had the capacity to discriminate between these TPs without need of any additional 258 separation step or the use of standards. Furthermore, it must be pointed out that an even 259 260 higher number of TPs can be taken into account through suspect screening than in targeted approaches, thus providing more comprehensive information on the transformation of one or 261 more pesticides to a series of TPs in the environment. 262

The pesticide dissipation study carried out at field scale was found to be suitable for the 263 purposes of our study. The applied TCZ concentration was in the range of concentrations 264 commonly applied in agriculture<sup>23</sup> and nine time points within 125 days after treatment 265 enabled the comprehensive exploration of TP formation and decay patterns although detailed 266 data on the temporal patterns were not established due to the absence of analytical standards. 267 As TCZ TPs were extracted from the top 5 cm layer of soil, it has to be noted that only those 268 TPs present at this soil layer could be studied. TP properties can strongly deviate from those 269 of the parent molecule<sup>40</sup> and might be vulnerable to leaching, runoff or volatilization although 270 the parent molecule is considered as relatively immobile or non-volatile.<sup>36,41</sup> Thus, TPs which 271 potentially dissipated from the top soil layer, which were not extractable from soil via the 272

applied extraction method or which were not listed in the TP library were disregarded in thisstudy.

Moreover, TCZ and several of its TPs have two enantiomers or do even possess more than one chiral center. This may be important concerning stereoselective toxicity and dissipation<sup>42</sup> but it was not considered in this study. For the completion of our analytical approach and for the provision of comprehensive information on actual environmental fate, molecular typology can be applied to estimate potential properties of the detected TPs in a theoretical manner. This technique can be used to estimate properties of TPs of any anthropological substance in nature.

#### 282 Categorization of TCZ TPs

TCZ and its 34 detected TPs were categorized by TyPol into 3 different clusters (table 1, 283 figure 1) according to their molecular descriptors including molecular weight, number of 284 285 atoms, dipole moment, polarizability or total energy (Supporting Information, table S2). TCZ and 23 of its TPs were categorized into cluster 1 exhibiting the highest simple and valence 286 molecular connectivity indices (MCIs), energy of the highest occupied molecular orbital 287  $(E_{HOMO})$ , molecular weight (MW), surface (S), polarizability ( $\alpha$ ), number of atoms and the 288 lowest total energy ( $E_{tot}$ ). Cluster 2 was composed of 7 TPs that were characterized by the 289 highest dipole moment ( $\mu$ ), energy of the lowest unoccupied molecular orbital (E<sub>LUMO</sub>) and 290 the lowest number of chlorine atoms. The final 4 TPs were grouped in cluster 3. Those 291 molecules were characterized by the lowest MCIs,  $E_{HOMO}$ , MW, S and  $\alpha$  but by the highest 292 E<sub>tot</sub>. 293

The different molecular properties of the TPs within each cluster also adumbrate deviating environmental behaviors, once more emphasizing the importance of the comprehensive knowledge of the existence of each occurring TP. This means that TPs which were classified into the same cluster as their parent molecule TCZ (including newly detected and yet

unknown TPs) may have similar behavior in the environment compared to TCZ. TPs which 298 were clustered away from TCZ may have different environmental fate and ecotoxicological 299 impact compared to TCZ. This can be especially problematic if their formation is ignored and 300 thus only TCZ and known TPs are quantified and monitored in the environment. To evaluate 301 these hypotheses, TyPol was applied for a second time in order to categorize 116 various 302 well-known pesticides and 19 TPs available in the TyPol database by analysis of both 303 molecular descriptors and their known environmental parameters (half-life in soil  $(DT_{50})$ , 304 sorption coefficient ( $K_{OC}$ ), vapor pressure ( $P_{vap}$ ) and bioconcentration factor (BCF)). The 305 compounds formed 6 clusters and median values of the molecular descriptors for each cluster 306 307 were obtained (an example is given for K<sub>OC</sub> in the Supporting Information, tables S3 and S4, and figure S3). In order to obtain a preliminary idea of the potential environmental behavior 308 of the detected TCZ TPs, their molecular descriptors were compared to the median values of 309 the 6 clusters formed by clustering the 116 pesticides and 19 TPs. When the molecular 310 descriptors of the well-known compounds of one cluster were highly similar to those of a 311 TCZ TP cluster, it was assumed that environmental parameters could be in a similar range. 312 TPs that were in the same cluster as their parent compound TCZ (cluster 1) were found to 313 overlap with a cluster of pesticides characterized by i) low  $P_{vap}$  (median  $P_{vap} = 0.026$  mPa), 314 indicating a low volatility from soil and plants and thus low risk for short or long range 315 transfer through the atmosphere,<sup>43</sup> ii) low mobility in soil (median  $K_{OC} = 825 \text{ L/kg}$ ) 316 suggesting a low risk for groundwater contamination,<sup>44</sup> iii) low persistence in soil 317 (median  $DT_{50} = 37$  days) and iv) low potential for bioaccumulation (median BCF = 78) 318 (Supporting Information, table S2). In the same manner, TCZ TPs forming cluster 2 had 319 molecular descriptor values similar to pesticides and TPs exhibiting i) a higher soil mobility 320 than TCZ (median  $K_{OC} = 217$  L/kg), thus posing a moderate potential risk for groundwater 321 contamination,<sup>44</sup> ii) a higher potential for volatilization losses (median  $P_{vap} = 0.098$  mPa) than 322

TCZ, iii) lower persistence in soil (median  $DT_{50} = 20$  days) and iv) a lower potential for 323 bioaccumulation (median BCF = 44). Finally, TCZ TPs forming cluster 3 were the most 324 mobile molecules among the identified TPs (median Koc of 123 L/kg) suggesting a 325 potentially higher risk for groundwater contamination. Furthermore, the TPs of cluster 3 326 posed the highest risk for volatilization losses (median  $P_{vap} = 1.600$  mPa) suggesting a 327 significant risk for long or short range atmospheric transfer. Their persistence in soil was 328 329 assumed to be similar to that of cluster 2 ( $DT_{50} = 20$  days) and lower than that of TCZ (cluster 1) and their potential for bioaccumulation was estimated to be low (BCF = 1) in comparison 330 to cluster 1 (BCF = 78) and cluster 2 (BCF = 44). These results are in agreement with 331 332 previous observations showing that when E<sub>HOMO</sub>, MCIs, and MW decrease, sorption of organic compounds tend to decrease, and when  $\alpha$ ,  $\mu$  and MW increase, P<sub>vap</sub> tend to decrease.<sup>45</sup> 333 Overall, the *in silico* approach using TyPol suggests that 23 of the 34 TCZ TPs targeted in this 334 study are expected to have a similar environmental behaviour as the parent compound. On the 335 336 other hand, the remaining 11 TPs may be more mobile and volatile than TCZ suggesting a higher risk for groundwater and air contamination. However, their lower persistence and 337 lower bioaccumulation potential than TCZ may counterbalance this risk. This combined 338 339 method allows a first assessment of the fate of theoretical TPs in the environment identifying those of potential concern that need to be intensively studied in a refined risk assessment 340 approach. If we consider TCZ as a case study, certain TPs from each cluster could be selected 341 based on their molecular features (e.g. selecting between some constitutional isomers, keto-342 enol tautomers or others) and further considered for a refined ERA. 343

To our knowledge, our approach is the first which combines (i) comprehensive screening of environmental samples (particularly soils) for suspected pesticide TPs based on a library of empirical and theoretical TPs, (ii) detection of TPs without the need of analytical standards, (iii) proposition of their molecular structures without the need of NMR spectroscopy and then

(iv) categorization of the detected TPs according to their physicochemical properties aiming 348 349 to assess their environmental behavior. The approach described in this study could be applied for refining pesticide risk assessment and thus enable regulators and agrochemical industries 350 351 to identify potential pesticide TPs showing relevant properties for further environmental and ecotoxicological assessment. Once the selected TPs are synthesized, absolute confirmation 352 and quantification can be enabled and thus TP fate, environmental concentration and toxicity 353 354 to ecosystems and human health can be assessed providing a refined and comprehensive risk assessment of a given pesticide. 355

# 356 ASSOCIATED CONTENT

Supporting information is available regarding the TCZ TP library, TP extraction from soil, the analytical method, chromatogram and MSC examples, the molecular typology method, detailed TyPol results for TCZ and its TPs and for the 116 pesticides and 19 TPs from the TyPol database. This material is available free of charge via the Internet.

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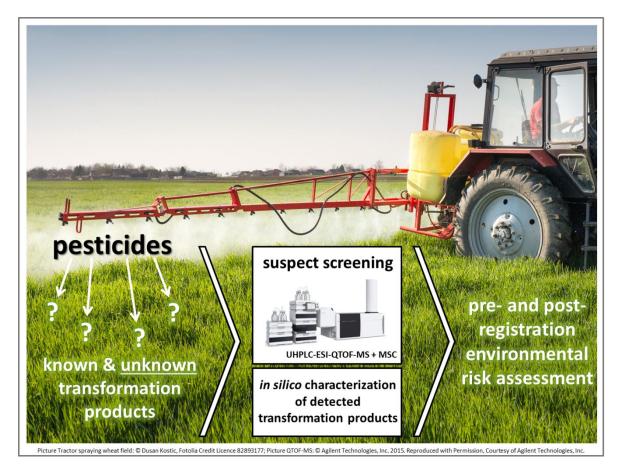
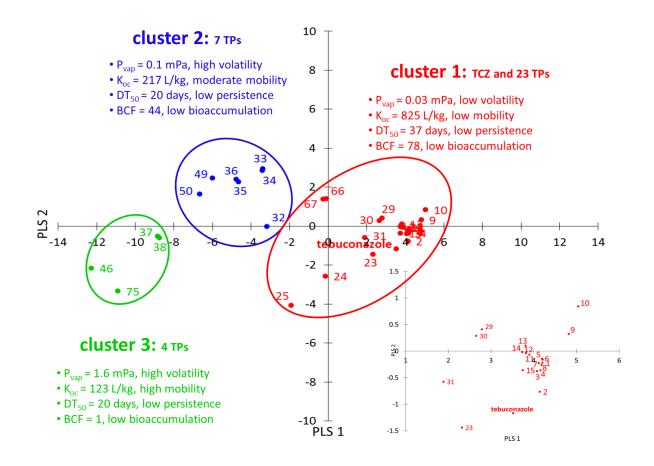


Table 1: TCZ TPs detected in this study by suspect screening analysis. Reference to the degree of confidence on identification (a. detected by MS (orange hooklet); b. selected by target tandem MS (yellow hooklet); c. proposed molecular structure by MSC (green hooklet)), name, structure, mass and reference if empirical of each TP are provided. The time points at which a TP was detected and the cluster into which it was categorized are given. Blue boxes group TPs having the same mass.

compound name	molecular structure	mass [g/mol]	reference	detected in this study	RT [min]	details	TyPol cluster
TCZ, 1-(4-chlorophenyl)-4,4- dimethyl-3-(1,2,4-triazol-1- ylmethyl)pentan-3-ol		307.15	38	a,b,c	9.38	detected at 0,3,7,21,35, 56,70,105 and 125 d	1
TP 1, 5-(4-chlorophenyl)-2,2- dimethyl-3-(1,2,4-triazol-1- ylmethyl)pentane-1,3-diol	он он он он он	323.14	29	✓ a	9.24	detected at 21,35 and 70 d	1
TP 2, 1-(4-chlorophenyl)-4,4- dimethyl-3-(1,2,4-triazol-1- ylmethyl)pentane-2,3-diol		323.14	37	✓ a	9.24	detected at 21,35 and 70 d	1
TP 3, 1-(4-chlorophenyl)-4,4- dimethyl-3-(1,2,4-triazol-1- ylmethyl)pentane-1,3-diol	* OH	323.14	theoretical TP	✓ a	9.24	detected at 21,35 and 70 d	1
TP 4, 2-chloro-5-[3-hydroxy-4,4- dimethyl-3-(1,2,4-triazol-1- ylmethyl)pentyl]phenol	жн сон	323.14	theoretical TP	✓ a	9.24	detected at 21,35 and 70 d	1
TP 5, 5-chloro-2-[3-hydroxy-4,4- dimethyl-3-(1,2,4-triazol-1- ylmethyl)pentyl]phenol	н сн	323.14	37	✓ a	9.24	detected at 21,35 and 70 d	1
TP 6, 2-[2-(4-chlorophenyl)ethyl]- 3,3-dimethyl-1-(1,2,4-triazol- 1-yl)butane-1,2-diol	HO * CH	323.14	theoretical TP	✓ a	9.24	detected at 21,35 and 70 d	1
TP 7, 1-[2-[2-(4- chlorophenyl)ethyl]-2- hydroxy-3,3-dimethyl-butyl]- 1,2,4-triazol-3-ol	HO CI	323.14	theoretical TP	✓ a	9.24	detected at 21,35 and 70 d	1
TP 8, 2-[2-[2-(4- chlorophenyl)ethyl]-2- hydroxy-3,3-dimethyl-butyl]- 1,2,4-triazol-3-ol	* ОН СІ	323.14	theoretical TP	✓ a	9.24	detected at 21,35 and 70 d	1

TP 9, 1-(4-chlorophenyl)-4,4- dimethyl-3-(1,2,4-triazol-1- ylmethyl)pentane-1,3,5-triol		339.14	37	✓ a	9.06	detected at 3,7,21,35, 56,70,105 and 125 d	1
TP 10, 1-[2-[2-(4- chlorophenyl)ethyl]-2- hydroxy-3,3-dimethyl-butyl]- 1,2,4-triazole-3,5-diol	HO H CI	339.14	theoretical TP	✓ a	9.06	detected at 3,7,21,35, 56,70,105 and 125 d	1
TP 11, 1-(4-chlorophenyl)-3- hydroxy-4,4-dimethyl-3- (1,2,4-triazol-1- ylmethyl)pentan-1-one		321.12	30-33	a,b,c	9.41	detected at 3,7,21,35, 56,70,105 and 125 d	1
TP 12, (E)-1-(4-chlorophenyl)-4,4- dimethyl-3-(1,2,4-triazol-1- ylmethyl)pent-1-ene-1,3-diol		321.12	37	a,b,c	9.41	detected at 3,7,21,35, 56,70,105 and 125 d	1
TP 13, 1-(4-chlorophenyl)-3- hydroxy-4,4-dimethyl-3- (1,2,4-triazol-1- ylmethyl)pentan-2-one		321.12	theoretical TP	a,b,c	9.41	detected at 3,7,21,35, 56,70,105 and 125 d	1
TP 14, (Z)-1-(4-chlorophenyl)-4,4- dimethyl-3-(1,2,4-triazol-1- ylmethyl)pent-1-ene-2,3-diol		321.12	theoretical TP	a,b,c	9.41	detected at 3,7,21,35, 56,70,105 and 125 d	1
TP 15, 2-[2-(4-chlorophenyl)ethyl]-2- hydroxy-3,3-dimethyl-1- (1,2,4-triazol-1-yl)butan-1-one		321.12	theoretical TP	a,b,c	9.41	detected at 3,7,21,35, 56,70,105 and 125 d	1
TP 23, 1-(4-chlorophenyl)-4-methyl- 3-(1,2,4-triazol-1- ylmethyl)pentan-3-ol		293.13	37	a,b,c	10.15	detected at 3,7,21,35, 56,70,105 and 125 d	1
TP 24, 4-(4-chlorophenyl)-2-methyl- 1-(1,2,4-triazol-1-yl)butan-2- ol		265.10	37	✓ a	6.54	detected at 105 and 125 d	1
TP 25, 4-(4-chlorophenyl)-1-(1,2,4- triazol-1-yl)butan-2-ol		251.08	37	✓ a	4.24	detected at 3,7,21,35, 56,70,105 and 125 d	1
TP 29, 4-[3-hydroxy-4,4-dimethyl-3- (1,2,4-triazol-1- ylmethyl)pentyl]phenol	н н	289.18	theoretical TP	✓ a,b	8.20	detected at 105 and 125 d	1
TP 30, 2,2-dimethyl-5-phenyl-3- (1,2,4-triazol-1- ylmethyl)pentane-1,3-diol	CH *	289.18	theoretical TP	✓ a,b	8.20	detected at 105 and 125 d	1

TP 31, 4,4-dimethyl-1-phenyl-3- (1,2,4-triazol-1- ylmethyl)pentan-3-ol	CH CH	273.18	theoretical TP	a,b,c	9.65	detected at 3,7,21,35, 56,70,105 and 125 d	1
TP 32, 5-tert-butyl-5-(1,2,4-triazol-1- ylmethyl)tetrahydrofuran-2- one	+*°~~°	223.13	29,34	🗸 a,b,c	5.13	detected at 35,56,70, 105 and 125 d	2
TP 33, 4-hydroxy-5,5-dimethyl-4- (1,2,4-triazol-1- ylmethyl)hexanoic acid	* OH OH	241.14	33	✓ a	8.38	detected at 3,7,21,35 and 56 d	2
TP 34, 5,5-dimethyl-4-(1,2,4-triazol- 1-ylmethyl)hex-1-ene-1,1,4- triol	ж он он он	241.14	33	✓ a	8.38	detected at 3,7,21,35 and 56 d	2
TP 35, 3-hydroxy-4,4-dimethyl-3- (1,2,4-triazol-1- ylmethyl)pentanoic acid		227.13	32	✓ a	4.72	detected at 3,7,21,35 and 56 d	2
TP 36, 4,4-dimethyl-3-(1,2,4-triazol- 1-ylmethyl)pent-1-ene-1,1,3- triol		227.13	32	✓ a	4.72	detected at 3,7,21,35 and 56 d	2
TP 37, 3,3-dimethyl-1-(1,2,4-triazol- 1-yl)butan-2-one		167.11	33	🗸 a,b,c	9.28	detected at 35,56,70, 105 and 125 d	3
TP 38, (E)-3,3-dimethyl-1-(1,2,4- triazol-1-yl)but-1-en-2-ol		167.11	33	a,b,c	9.28	detected at 35,56,70, 105 and 125 d	3
TP 46, 2-(1,2,4-triazol-1-yl)ethane- 1,1-diol	* OH	129.05	35,36	✓ a,b	0.68	detected at 56,70,105 and 125 d	3
TP 49, butyl 2-hydroxy-3-(1,2,4- triazol-1-yl)propanoate		213.11	38	√ a	7.27	detected at 3,7,21,35 and 56 d	2
TP 50, (Z)-1-butoxy-3-(1,2,4-triazol- 1-yl)prop-1-ene-1,2-diol	И ОН	213.11	38	✓ a	7.27	detected at 3,7,21,35 and 56 d	2
TP 66, (NE)-N-[2-[2-(4- chlorophenyl)ethyl]-2- hydroxy-3,3-dimethyl- butylidene]formamidine	* OH	280.13	29	✓ a,b,c	8.14	detected at 7,21,35,56,70 and 105 d	1
TP 67, 1-(4-chlorophenyl)-4,4- dimethyl-3-[(E)- (methylenehydrazono)methyl] pentan-3-ol	DH CH	280.13	29	a,b,c	8.14	detected at 7,21,35,56,70 and 105 d	1
TP 75, 4-chlorobenzoic acid	O OH	156.00	37	✓ a	9.05	detected at 70 and 105 d	3
* Chiral center							



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Figure 1: Categorization of TCZ and its 34 detected TPs into 3 clusters according to their molecular descriptors on the two main components of the PLS regression (PLS 1 and PLS 2) and their estimated environmental parameters. Formed clusters are defined by a different color code. Each TP has its respective number. The inserted graph serves as enlargement of a part of cluster 1.