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# Image-based Biophysical Simulation of Intracardiac Abnormal Ventricular Electrograms

Rocío Cabrera-Lozoya, Benjamin Berte, Hubert Cochet, Pierre Jaïs, Nicholas Ayache, Maxime Sermesant

**Abstract— Goal:** In this work, we used *in silico* patient-specific models constructed from 3D delayed-enhanced magnetic resonance imaging (DE-MRI) to simulate intracardiac electrograms (EGM). These included electrically abnormal electrograms as these are potential radiofrequency ablation (RFA) targets. **Methods:** We generated signals with distinguishable macroscopic normal and abnormal characteristics by constructing MRI-based patient-specific structural heart models and by solving the simplified biophysical Mitchell-Schaeffer model of cardiac electrophysiology. Then, we simulated intracardiac electrograms by modelling a recording catheter using a dipole approach. **Results:** Qualitative results show that simulated EGM resemble clinical signals. Additionally, the quantitative assessment of signal features extracted from the simulated EGM showed statistically significant differences ( $p < 0.0001$ ) between the distributions of normal and abnormal electrograms, similarly to what is observed on clinical data. **Conclusion:** We demonstrate the feasibility of coupling simplified cardiac EP models with imaging data to generate intracardiac EMG. **Significance:** These results are a step forward in the direction of the pre-operative and non-invasive identification of ablation targets to guide RFA therapy.

**Index Terms—**cardiac electrophysiology modelling, intracardiac electrogram modelling, radiofrequency ablation planning, electroanatomical mapping.

## I. INTRODUCTION

**D**espite its numerous advantages over the implantation of a defibrillator, radiofrequency ablation as a curative therapy for patients at risk of sudden cardiac death is still challenging due to the difficulty in finding the appropriate ablation targets. Invasive measures of cardiac activity obtained during an electrophysiology (EP) study provide insightful information about the electrical characteristics of the analysed myocardium. Under pathological conditions, these extracellular recordings deviate from the normal (healthy) signal shape and present multiple deflections and fractionation episodes. However, this lengthy and invasive procedure is difficult for both the patient and the clinician. This is why we are exploring non-invasive approaches to define the ablation targets pre-operatively.

In this work, we focus on a target that recently emerged as an appropriate therapy. On structurally diseased hearts with fibrotic scar, bundles of surviving tissue promote electrical circuit re-entry and are a cause of arrhythmias. Local abnormal ventricular activities (LAVA) are sharp fractionated bipolar potentials occurring during or after the far-field electrogram. They have been shown to indicate surviving fibers within the

scar and have been successfully used as targets for radiofrequency ablation [1].

Previously, fractionated electrograms (EGM) were thought to be caused mainly by artifacts related to the electronics of the acquisition system. Although artifacts indeed may cause complex EGM, most of them are caused by the peculiar behavior of activation fronts, due to structural and electrical complexity of the underlying tissue [2]. Previous studies have used synthetically generated EGM to explore different pathological phenomena. The authors in [3] describe the generation of EGM fractionation from changes in activation wavefront curvature in experimental canine infarction. In [4], the modelling of intracardiac recordings was used to aid in the reconstruction of cardiac ischemia. The work in [5] studied the influence of different catheter angles, locations and filter settings on the morphology of simulated intracardiac EGM and compared them to clinical signals. The study in [6] derived a way to estimate wave direction and conduction velocity from simulated intracardiac EGM recorded using circular mapping catheters and the one in [7] complemented it by applying such estimations on personalized models.

Delayed-enhanced magnetic resonance imaging (DE-MRI) enables a non-invasive 3D assessment of scar topology and heterogeneity with millimetric spatial resolution. It has been hypothesized that areas of intermediate signal intensity in DE-MRI, also referred to as the border zone (BZ), host scarred and surviving myocardium related to arrhythmia in ischemic populations [1].

In this work, we test the feasibility of using DE-MR image-based simulation to reproduce macroscopic abnormal patterns in intracardiac EGM. Through this work, we aim to shine some light on the understanding of the macroscopic mechanisms underlying EGM fractionation in the border zone. The overall scheme of the approach is described in Fig. 1.

## II. CLINICAL DATA

Five patients referred for cardiac ablation of post-infarction ventricular tachycardia (VT) were included in this study. All patients gave written informed consent. They underwent cardiac MRI prior to high-density EP contact mapping of the endocardium (Fig. 1).

### A. Imaging Data

The scar tissue was imaged on a 1.5 Tesla clinical device (Avanto, Siemens Medical Systems) 15 minutes after the injection of a gadolinium contrast agent. A whole heart image

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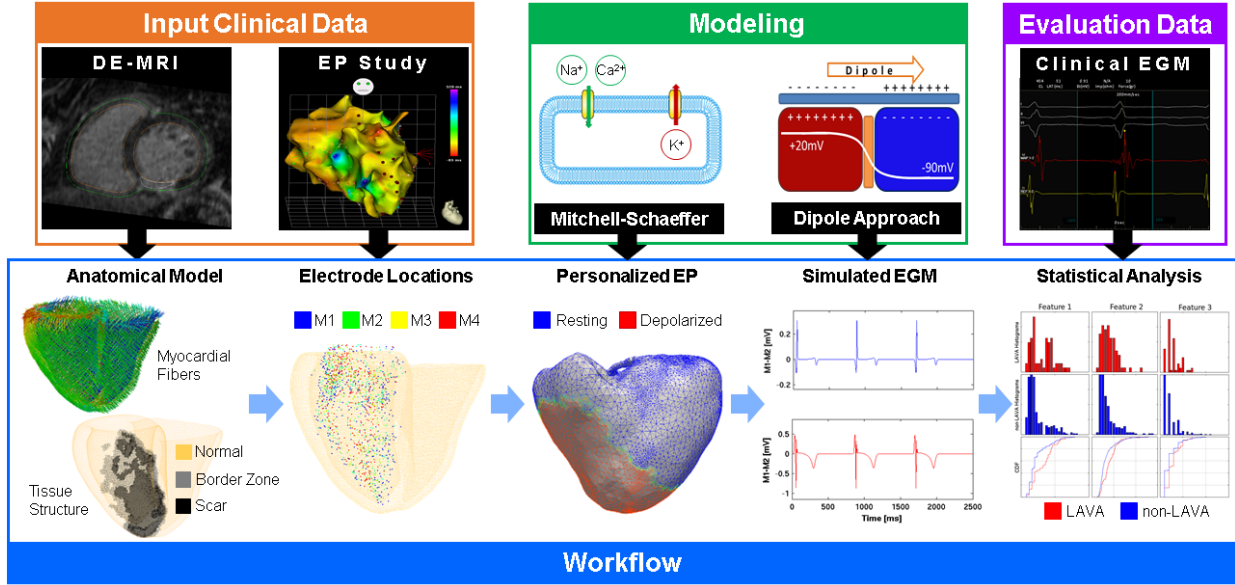


Fig. 1: The processing pipeline includes the generation of a personalized cardiac model from imaging data, the simulation of cardiac electrophysiology and intracardiac electrograms and the statistical analysis of the clinical and simulated signals.

was acquired using an inversion-recovery prepared, ECG-gated, respiratory-navigated, 3D gradient-echo pulse sequence with fat-saturation ( $1.25 \times 1.25 \times 2.5 \text{ mm}^3$ ).

The myocardium was manually segmented on images which were reformatted to have isotropic voxel size ( $0.625 \text{ mm}^3$ ). Abnormal myocardium consisting of dense scar and border zone areas was segmented using adaptive thresholding of the histogram, with a cut-off at 35% of maximal signal intensity. Segmentations were reviewed by an experienced radiologist, with the option of manual correction.

### B. Electrophysiological Data

The CARTO mapping system (Biosense Webster) enables the 3D localization of the catheter tip and provides the distribution of EP signals on cardiac surfaces. Contact mapping was achieved in sinus rhythm on the endocardium (trans-septal approach) with a multi-spline catheter (PentaRay, Biosense Webster). The catheter's five-branched star design (including 20 electrodes located along 5 branches) allows for high density mapping by recording multiple locations at once. Recordings are of 2.5 seconds of duration. Signals were categorized as normal or abnormal by an experienced electrophysiologist. Table I includes details on the EP studies for each patient.

TABLE I: CARTO Electrophysiology Study Details

	P1	P2	P3	P4	P5	$\mu \pm \sigma$
# of Catheter Locations	368	1201	339	214	700	$567 \pm 399$
# of Abnormal EGM	71	44	82	44	33	$55 \pm 21$
# of Normal EGM	297	1157	257	170	667	$510 \pm 409$
% of Abnormal EGM	19	4	24	21	5	$15 \pm 9$

Fig. 2 provides further insight in the distribution of the clinical annotations of the electrode locations for Patient 1. The distance to the border zone is computed by taking the Euclidean distance from the annotated signal location to the

closest edge or point in the mesh generated from the border zone segmentation, as will be detailed in Section III-A. If points are within the border zone mesh, their distance is set to zero. Because the distances are computed using a border zone mesh and not the image intensities themselves, the edges of the BZ are not diffuse. It can be seen that abnormal signals have a higher tendency to remain close to the border zone, nonetheless, healthy signals can also be found in these regions. Similar abnormal EGM distributions were found in the remaining four patients.

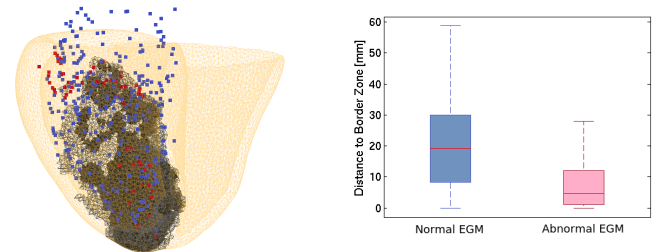


Fig. 2: (Left) Image-driven personalized model and location of abnormal (red) and normal (blue) annotated signals on Patient 1. (Right) Boxplots representing the distance of the annotated signals to the border zone.

### III. CARDIAC MODEL CONSTRUCTION

Fig. 1 shows our processing pipeline. It consists of the generation of a patient-specific cardiac model from imaging data, with inclusion of synthetic myocardial fibers. A cardiac EP model with tissue-specific properties is solved on this geometry and intracardiac EGM are computed at locations obtained from the clinical EP study. Signal feature extraction is performed on both clinical and simulated EGM, which allows us to perform statistical analysis. Each segment of the pipeline will be detailed in the following sections.



Fig. 3: Personalized heart meshes for our five patients depicting healthy myocardium (yellow), scar (black) and border zone (grey) tissues derived from DE-MRI segmentations.

#### A. Anatomical Model Generation

Segmentations of the myocardium, border zone and scar regions on DE-MRI were performed using MUSIC software (multimodality software for specific imaging in cardiology, L’Institut de Rythmologie et de Modélisation Cardiaque, University of Bordeaux, Institut National de Recherche en Informatique et Automatique Sophia Antipolis, Sophia Antipolis, France). The software is a solution that was developed in-house and built on the open-source medInria software architecture (<http://med.inria.fr>). Segmentations masks were the basis to generate personalized 3D tetrahedral meshes of the biventricular myocardium using the CGAL library [8]. Mesh details including mean edge length and number of tetrahedra are shown in Table III.

Myocardial fiber directions were created synthetically as proposed in [9]. The elevation angle (measured w.r.t. the short axis plane) was varied from  $-80^\circ$  on the epicardium to  $0^\circ$  at mid-myocardial wall to  $+80^\circ$  on the endocardium.

#### B. Electrophysiology (EP) Model

Cardiac electrophysiology can be described through a variety of mathematical models [10], [11], [12], [13]. More detailed cell-specific models also exist which aim to describe cell-to-cell variability in the cardiac tissue [14], [15]. These models have evolved in the last decades to better represent physiological phenomena [16], e.g. some have been used to study VT or other arrhythmia episodes [17], [18], [19], [20].

We chose a model able to represent complex cardiac electrical phenomena while keeping the number and variation of the involved variables tractable: the Mitchell-Schaeffer (MS) [13] model. Its main advantages include:

- its simplicity, as it only includes two differential equations
- the relationship between its parameters and physiological behavior makes it easier to interpret
- its ability to simulate arrhythmia macroscopically due to its restitution parameters

It has been used for patient-specific personalization for VT simulation [19] and interactive simulation of patient-specific EP [21].

The Mitchell-Schaeffer (MS) model describes the transmembrane potential as the sum of a passive diffusive current and active reactive currents (inward and outward ionic currents). The system of equations used is the following:

$$\begin{cases} \partial_t u = \text{div}(D\nabla u) + \frac{zu^2(1-u)}{\tau_{in}} - \frac{u}{\tau_{out}} + J_{stim}(t) \\ \partial_t z = \begin{cases} \frac{(1-z)}{\tau_{open}} & \text{if } u < u_{gate} \\ \frac{-z}{\tau_{close}} & \text{if } u > u_{gate} \end{cases} \end{cases} \quad (1)$$

Where:

- $u$ : normalised transmembrane potential variable
- $z$ : gating variable depicting depolarization and repolarization phases by opening and closing the current gates
- $J_{in} = zu^2(1-u)/\tau_{in}$ : inward  $Na^+$  and  $Ca^{2+}$  ionic currents which raise the action potential voltage
- $J_{out} = u/\tau_{out}$ : outward ionic  $K^+$  current which decreases the action potential voltage during repolarization
- $J_{stim}$ : stimulation current at the pacing location
- $\tau_{in}, \tau_{out}, \tau_{open}, \tau_{close}$  have units of seconds
- $D = d \cdot \text{diag}(1, r, r)$ : anisotropic diffusion tensor in fiber coordinates which enables conduction velocity in the fiber direction to be 2.5 larger than in the transverse plane ( $r = 1/2.5^2$ )

This model incorporates both action potential duration (APD) and conduction velocity restitution effects. We used the MS model implementation in the SOFA framework [21].

#### C. Pseudo-personalization: Tissue-specific EP Properties

The EP properties of the infarcted and border zone tissues differ from those of the healthy myocardium. For the construction of our personalized model, DE-MRI was used to assess the viability of the cardiac tissue. We identified tissue as either healthy myocardium, scar or border zone. Fig. 3 shows the derived volumetric ventricular models for each of our patients.

Conductivity in the border zone was decreased by 90% with respect to its value in the healthy myocardium [22]. Studies in infarcted hearts [23] [24] [25] [26] reported variations in ionic currents in the border zone tissue generating action potentials that differ from the healthy myocardium, with:

- 32% lower peak action potential amplitudes
- 31% smaller upstroke velocity
- 25% longer action potential duration (APD)

Consequently, in our model, the term  $J_{in} = zu^2(1-u)/\tau_{in}$  in (1) was modified to  $zu^2(a-u)/\tau_{in}$ , where variable  $a$  controls the peak amplitude for the action potential. To obtain a smaller upstroke velocity,  $\tau_{in}$  in the border zone was increased 31% with respect to the value in the healthy tissue.

The analytical expression for the maximum APD for the Mitchell-Schaeffer model is given by  $APD =$

TABLE II: Simulation Parameter Values

Parameter	Tissue Type		
	Healthy	Border Zone	Scar
$\tau_{in}$ [ms]	0.3	0.42	$10^4$
$\tau_{out}$ [ms]	6	8.4	$10^4$
$\tau_{close}$ [ms]	150	187.5	187.5
$\tau_{open}$ [ms]	120	120	120
$u_{gate}$	0.13	0.13	0.13
Action Potential Amplitude	1.0	0.7	0.7
Conductivity [ $\times 10^{-3} m^2/s$ ]	4.0	0.4	0.4

$\tau_{close} \ln(\tau_{out}/4\tau_{in})$ , implying a linear relationship between  $\tau_{close}$  and the APD. Therefore, the values for this parameter in the border zone were increased by 25%. Finally, because  $\tau_{in}$  had been modified to account for a smaller upstroke velocity,  $\tau_{out}$  was modified in the same proportion to keep the ratio  $\tau_{out}/\tau_{in}$  constant.

Scar tissue was modelled to have a null reaction component on the MS model by setting much higher values of  $\tau_{in}$  and  $\tau_{out}$  with respect to those of the healthy myocardium. The diffusion component was left with the values of the border zone. Healthy myocardial values were set to the default values in [13]. Furthermore, patient-specific sinus rhythm was extracted from surface electrocardiogram (ECG) recordings. Table II summarizes the parameters used for our personalized simulations.

The current tissue-specific model parameters were not modified to account for the heterogeneity in action potential durations between the endocardial and epicardial [27] nor between the apex to base [28] tissue, which affect the repolarization phase. This study focuses on EGM abnormalities found in the depolarization complex. This can explain the difference between simulated and recorded T-wave patterns.

#### IV. INTRACARDIAC EGM SIMULATION

Extracellular electrograms arise due to transmembrane currents occurring from differences in the axial voltage gradient at the interface between activated and inactivated myocardial cells [2]. If an activation wave front travels from left to right throughout the myocardium, at the boundary of the propagation front, the depolarized cell has an intracellular potential of +20mV whereas that of the cell in resting state remains at -90mV. This creates a current flow from the activated to the inactivated myocyte. To comply with the preservation of charge, the back of the activation front presents a current flow from the extracellular space into the intracellular space, whereas the current flow direction is reversed in the regions ahead of the activation front. This flow of current in the extracellular space generates an extracellular voltage difference and therefore a potential dipole.

##### A. The Dipole Approach

The notion of resulting dipoles at the depolarization wave front can be used to compute the resulting potential at a given coordinate in the extracellular space [29]. This approach was used by [30] to compute pseudo pre-cordial and limb-lead ECGs from a Purkinje muscle model. We introduce a similar

approach to simulate unipolar and bipolar intracardiac EGM at a given position representing the catheter location.

The monodomain formulation describes the action potential propagation with the following reaction diffusion equation:

$$C_m \frac{\partial v}{\partial t} + I_{ion} = \nabla \cdot \sigma_m \nabla v \quad (2)$$

where  $v$  represents the transmembrane potential,  $C_m$  is the membrane capacitance,  $\sigma_m$  corresponds to the local conductivity and  $I_{ion}$  is the current through the cell membrane per unit of area. We define the equivalent current density  $\mathbf{j}_{eq}$  as:

$$\mathbf{j}_{eq} = -\sigma_m \nabla v \quad (3)$$

$-\mathbf{j}_{eq}$  behaves like a flow source density, and it can also be seen as a dipole moment per unit of volume:

$$\mathbf{p} = \int \mathbf{j}_{eq} dV \quad (4)$$

According to the volume conductor theory [29], the electric potential registered at a distance  $\mathbf{R}$  in a homogeneous volume conductor of conductivity  $\sigma$  outside the region occupied by the volume source  $V$  is :

$$\Psi(\mathbf{R}) = \frac{1}{4\pi\sigma} \int_V \mathbf{j}_{eq} \cdot \nabla \left( \frac{1}{\mathbf{R}} \right) dV \quad (5)$$

In our case, we model the moving propagation front as a local dipole. As this dipole is proportional to the potential gradient, and we use linear elements to solve the model equations, the dipole momentum  $\mathbf{p}_{\mathbf{X}}$  at a position  $\mathbf{X}$  is spatially invariant over a single tetrahedron  $H$ :  $\forall \mathbf{X} \in H$ ,  $\mathbf{p}_{\mathbf{X}} = \mathbf{p}_H$ .

Then we sum the potential over all the myocardial regions: those being activated, those depolarized and those at rest. Non-activated regions give almost null dipole momenta and it simplifies the overall calculation.

We discretize (4) in space ( $H$  tetrahedra of the mesh) and get the formulation of the dipole moment of the charge in the volume  $V_H$  of tetrahedron  $H$  :

$$\mathbf{p}_H = V_H \mathbf{j}_{eq,H}^t = V_H \sigma_{m,H} \nabla v_H \quad (6)$$

The gradient of the electrical potential in the tetrahedron  $\nabla v_H$  is computed using its node values and its shape vectors as defined in [31], and the dipole is located at the tetrahedron center  $X_H$ .

From (5), the contribution  $\Psi_H(\mathbf{X}_{el})$  of tetrahedron  $H$  to the potential field calculated at the electrode location  $\mathbf{X}_{el}$  is estimated by:

$$\Psi_H(\mathbf{X}_{el}) = \frac{1}{4\pi\sigma} \frac{(V_H \sigma_{m,H} \nabla v_H) \cdot (\mathbf{X}_{el} - \mathbf{X}_H)}{\|\mathbf{X}_{el} - \mathbf{X}_H\|^3} \quad (7)$$

Finally, we sum over the whole mesh to get the potential field at  $\mathbf{X}_{el}$ :  $\Psi(\mathbf{X}_{el}) = \sum_{H=1}^{Nb \text{ tetra}} \Psi_H(\mathbf{X}_{el})$ .

### B. Unipolar and Bipolar Electrogram Computations

Unipolar electrograms are obtained by positioning the exploring electrode in the heart and the indifferent electrode away from the heart so it has little or no cardiac signal [32].

Bipolar EGM correspond to the difference in potential between two unipolar measurements and are useful to study the local activities. The far-field signal is assumed to be similar for both unipolar recordings so it is largely filtered out [32].

Each of the five branches of the recording catheter used in the clinical environment has four electrodes named M1, M2, M3 and M4 from the distal to the proximal. Two bipolar recordings are generated from these unipolar measurements: M1-M2 and M3-M4 (Fig. 4). Electrode positions at recording time are given by the CARTO system and its spatial coordinates are used to generate our computational measurements, as exemplified in Fig. 1.

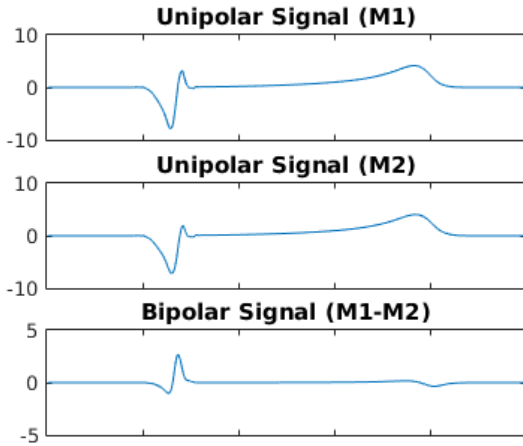


Fig. 4: The subtraction of two unipolar measurements generates a bipolar measurement. Here, the M1 and M2 unipolar signals are used to generate the bipolar M1-M2 signal.

Simulations were performed with a time step of  $1 \times 10^{-5}$  seconds. Further details on EGM simulations per patient are shown in Table III. Computation times correspond to the simulation of a single cardiac cycle and of all the corresponding patient-specific intracardiac EGM, on a computer with Intel Core i5 CPU.

TABLE III: Patient-specific Simulation Details

	P1	P2	P3	P4	P5
# Tetrahedra in Mesh	112,308	96,260	150,480	96,288	29,586
Mean Edge Length [mm]	2.69	2.67	2.69	2.68	3.98
# Nodes in Mesh	23,355	20,306	31,100	21,231	6,742
# Unipolar EGM	1472	4804	1956	856	2800
# Bipolar EGM	736	2402	978	428	1400
Cardiac Cycle [ms]	830	625	1150	1150	830
Computation time [min]	9.7	23.3	17.4	5.8	4.3

### V. SIGNAL ANALYSIS

In addition to a qualitative assessment of signal morphology, a quantitative signal evaluation was performed by extracting the signal characteristics from simulated and clinical EGM.

They are described next and some of them have been illustrated in Fig. 5:

- **Signal Range:** difference between the maximum and minimum amplitude signal values.
- **Number of Inflection Points:** number of points where the signal changes concavity.
- **Signal Energy:** calculated using Teager's operator [33].
- **Dominant Frequency:** from the fast Fourier transform.
- **Mean Slope:** mean absolute value of  $dV/dt$ .
- **Fractionation Index:** number of deflections with an amplitude  $>0.2\text{mV}$  from the signal baseline.
- **Minimum and Maximum Signal Value.**

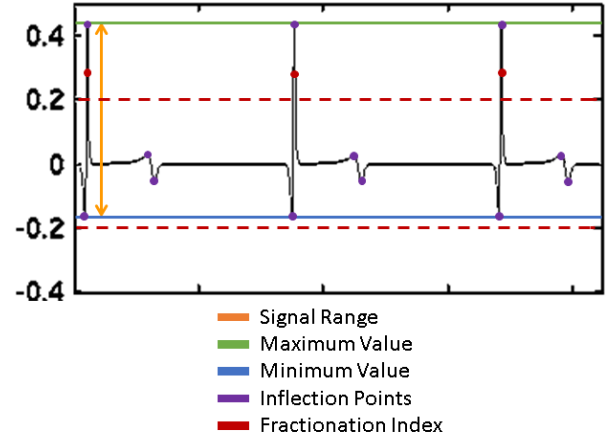


Fig. 5: Illustration of signal features extracted for EGM characterization and quantitative analysis.

Some of these features were inspired by those used clinically [34] to characterize EGM associated with atrial fibrillation. Signal range is expected to be smaller in abnormal electrograms when compared to healthy ones. On the other hand, the number of inflection points, mean slope and fractionation index are interesting characteristics as their values tend to be higher in abnormal electrograms with multiple deflections and fractionation episodes.

The distributions of the values of each of these characteristics in the normal and abnormal signal groups were assessed using the non-parametric Kolmogorov-Smirnov (KS) test, as this test does not assume that data come from a normal distribution [35]. This test compares the cumulative distribution functions of two datasets and computes a  $p$  value dependent on the maximum distance between them. It is useful to detect substantial differences in either shape, spread or median between the distributions. A value of  $p < 0.05$  was considered as statistically significant.

### VI. RESULTS AND DISCUSSION

Catheter measurements were simulated using the image-driven personalized heart model at personalized sinus rhythm with the parameters stated in Table II. Resulting EGM were qualitatively compared to their clinical counterparts and features between normal and abnormal groups were assessed.



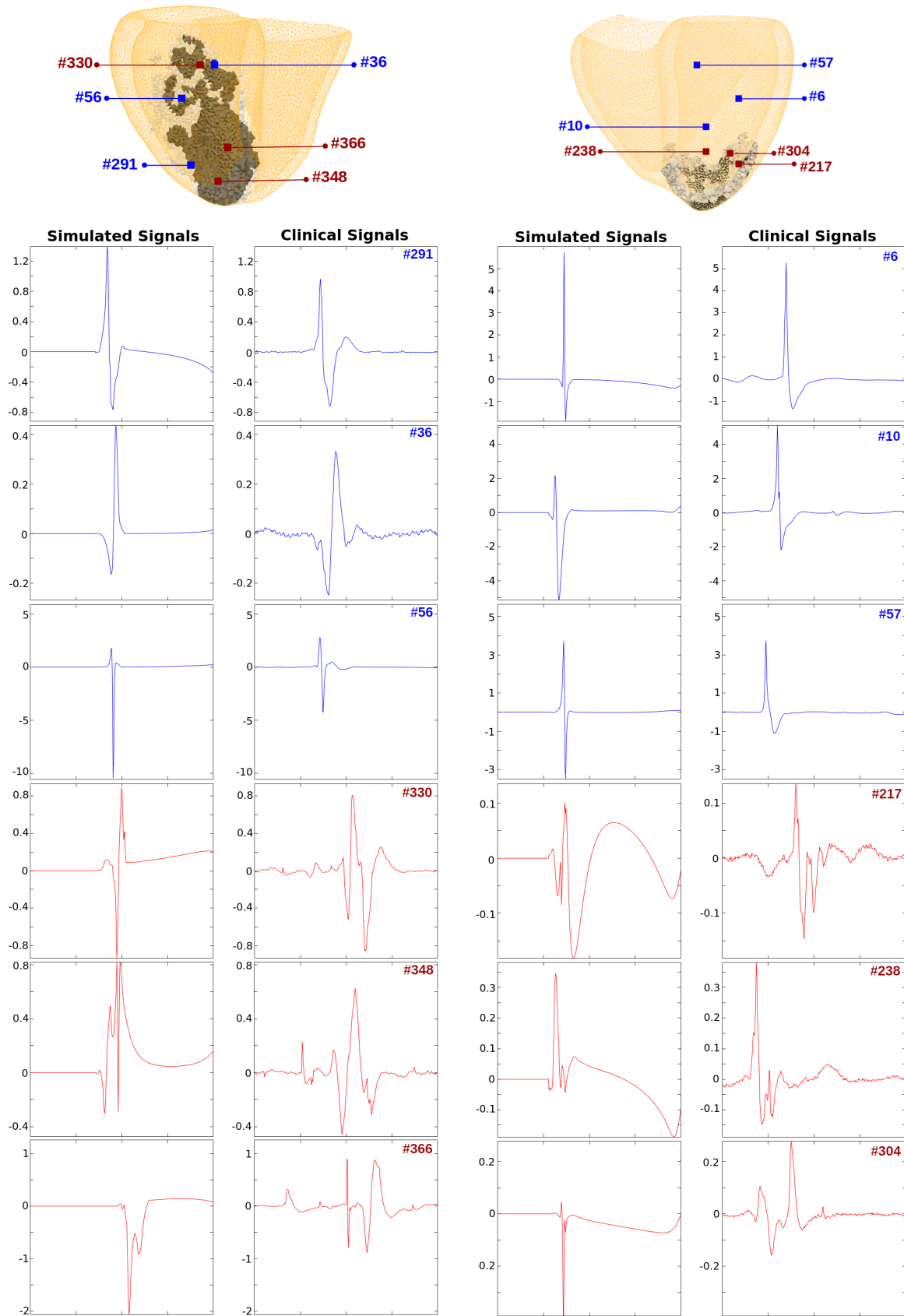


Fig. 6: Simulated and clinical normal (blue) and abnormal (red) bipolar EGM with electrode locations.

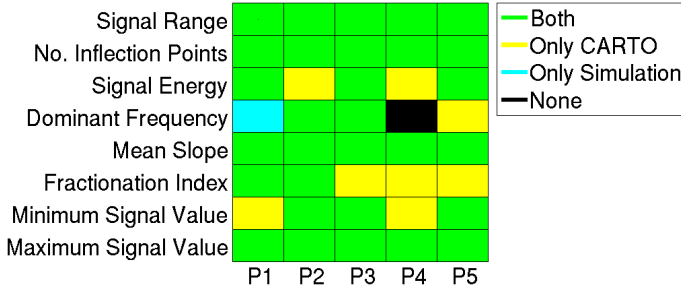


Fig. 7: KS test results. Statistically significant differences ( $p < 0.05$ ) between normal and abnormal feature distributions in (green) both CARTO and simulated signals (yellow) only in CARTO signals (blue) only in simulated signals (black) neither in CARTO nor in simulated signals.

#### A. Qualitative Assessment of Simulated EGM

Fig. 6 shows samples of the locations of the electrodes with respect to the cardiac geometry, and the resulting EGM simulation and clinical bipolar (M1-M2) signals at these locations for Patients 1 and 3. Three normal and three abnormal signals per patient are depicted.

A first qualitative assessment shows that simulated signals are less prone to noise when compared to the CARTO signals. This is expected as the simulated signals are not affected by catheter movement, breathing or cardiac motion, among other factors.

For the normal simulated signals, we observe a normal ventricular depolarization in the form of a steep slope, without any fragmentation inside the EGM signal. For the abnormal signals, fractionation is found anywhere during the QRS complex of the bipolar EGM.

As was mentioned before, the T-wave abnormality is due to the fact that APD heterogeneity across the myocardium was not accounted for in the model.

#### B. Clinical and Simulated Signal Characterization

Results of the KS test on the signal characteristics of the clinical CARTO recordings can be obtained from Fig. 7. With the exception of the dominant frequency in Patients 1 and 4, a statistically significant difference between the signal characteristics of normal and abnormal distributions was found. For reference, most test results yielded as low as  $p$  value of  $< 0.0001$ , denoting a very good separability between the class distributions of normal and abnormal signal features.

For the simulated signals, the results of the KS test can also be obtained from Fig. 7. The distributions of mean slope, number of inflection points and maximum signal value differed significantly among simulated normal and abnormal signal populations, as can be seen by the low  $p$  value obtained after the KS test.

Looking in more detail at these tests, the first two rows in Fig. 8 show the histograms of the distributions for the number of inflection points in the abnormal and normal classes in the simulated signals, respectively. The difference in distributions is perhaps more evident in the bottom row which shows the CDF of both classes. The average KS statistic (maximum

difference between CDF) for this feature across the five patients is 0.34, denoting a statistically significant distance between the distributions.

Figure 7 shows a good agreement between the KS test results in clinical and simulated signals. A statistically significant difference was found in the distributions of features such as signal range, number of inflection points, mean slope and maximum signal value on both clinical and simulated signals.

The distributions of dominant frequency in the signals performed the worst, where a statistically significant difference was only found in simulated signals for Patient 1. For Patient 5, only the CARTO signals had difference in their distribution and no statistically significant difference was found neither in the clinical nor in the simulated signals in Patient 4.

#### C. Discussion

This exploratory research paper seeks to evaluate whether computer simulation combined with medical imaging is able to pinpoint non-invasively the regions of the myocardium that generate abnormal signals in the same way as it is measured in a clinical environment invasively with a catheter, rather than to recreate EGM signals with high fidelity when compared to their clinical counterparts.

Given all the simplifications in modelling both the biophysical phenomenon and the data acquisition procedure, it would be hard to obtain quasi-realistic signals using the approach shown in this paper. Nevertheless, this approach demonstrated statistically significant differences in simulated normal and abnormal intracardiac electrograms which is of high clinical relevance. It opens the door for simulation-based approaches to guide EP interventions in order to guide clinicians to the locations where abnormal electrograms are found. It also restricts the search of abnormalities to subregions in the myocardium and could decrease procedure times. Finally, the clinician would in any case measure the electrical activity in the region before taking the decision of ablating.

## VII. CONCLUSION

We presented the use of a personalized image-based model for the simulation of intracardiac EGM with abnormal electrical characteristics. We showed that the use of a simplified biophysical electrophysiology model with tissue-specific parameters and the use of a dipole approach to simulate intracardiac electrograms were sufficient to generate abnormal signals which are properly localized and distinguishable from their normal counterpart as labeled by an experienced electrophysiologist.

Also, the resemblance of the simulated signals with their clinical CARTO counterpart was qualitatively assessed. Furthermore, feature extraction was performed on both the simulated and clinical signals. Characteristics such as the mean slope and number of inflection points presented significantly different distributions when assessed through the use of a Kolmogorov-Smirnov test in both the simulated ( $p < 0.002$ ) and clinical signals ( $p < 0.0001$ ).

The encouraging results obtained in this work demonstrate that it is feasible to generate intracardiac EGM using a



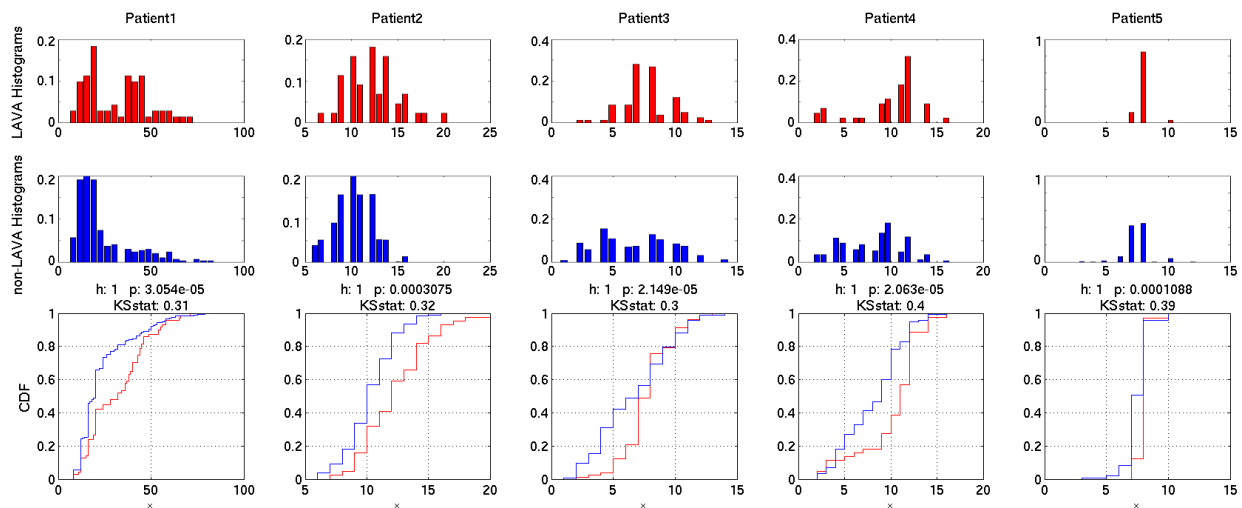


Fig. 8: Histograms of the number of inflection points for abnormal EGM (top) and normal EGM (middle) simulated signals by patient. (bottom) Cumulative distribution functions (CDF) for abnormal EGM (red) and normal EGM (blue) simulated signals.

simplified cardiac EP models personalized with imaging data, with distinct characteristics which could aid in the planning of RFA therapy by pre-operatively and non-invasively identifying ablation targets.

## VIII. ACKNOWLEDGEMENT

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