



Molecular identification of symbiotic dinoflagellates in Pacific corals in the genus Pocillopora (erratum)

Hélène Magalon, Jean-François Flot, Emmanuelle Baudry

► To cite this version:

Hélène Magalon, Jean-François Flot, Emmanuelle Baudry. Molecular identification of symbiotic dinoflagellates in Pacific corals in the genus Pocillopora (erratum). *Coral Reefs*, 2008, 27 (4), pp.997-997. 10.1007/s00338-008-0402-7 . hal-01312396

HAL Id: hal-01312396

<https://hal.science/hal-01312396>

Submitted on 6 May 2016

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Molecular identification of symbiotic dinoflagellates in Pacific corals in the genus *Pocillopora*

H. Magalon · J.-F. Flot · E. Baudry

The authors regret that they made an error in reporting the primers used to amplify and sequence the ITS1 region, mentioned in the Material and methods section. The primer ITSint-rev was incorrectly reported and an internal primer which was incorrectly omitted has been added below.

Materials and methods

PCR amplifications of zooxanthellae nrDNA

[The second paragraph of this section should be replaced with the following text]

To investigate zooxanthella diversity in more detail, the Internal Transcribed Spacer 1 region (ITS1), which is a non-coding region that evolves faster than the 28S nrDNA

(Santos et al. 2002a), was sequenced. Two primers were used to amplify both ITS1 and ITS2: one designed in the 3'-end of the 18S DNA [ITS-for: 5'-CGG TGA ATT ATT CGG ACT GAC-3'; reverse of SYM3, modified from Hunter et al. (1997)] and the other in the 5'-end of the 28S DNA [ITS-rev: 5'-TCC TCC GCT TAT TGA TAT GC-3' from Hunter et al. (1997)]. The PCR protocol was identical to that of 28S nrDNA amplification, except amplifications were carried out in a total volume of 30 µl. Amplified products were purified using QIAquick PCR purification kit (Qiagen) following the manufacturer's instructions and ITS1 region was directly sequenced in both directions using 3.2 pmol of the primer ITS-for and an internal primer in the 5.8S region [ITSint-rev: 5'-CAC GGA GTT CTG CAA TTC-3' (reverse of ITSintfor2 from LaJeunesse and Trench (2000))]. [The region ITS2 can be sequenced using ITS-rev and ITSintfor2 from LaJeunesse and Trench (2000).] Reagents and cycling conditions were as specified in the ABI Prism Big Dye Terminator Cycle Sequencing Ready Reaction kit (PE Applied Biosystems, Foster City, California).

H. Magalon (✉)
UMR CNRS 7625, Laboratoire Fonctionnement et Evolution des Systèmes Ecologiques, Université Pierre et Marie Curie,
7 quai saint Bernard, 75005 Paris, France
e-mail: hmagalon@svn.jussieu.fr

J.-F. Flot
UMR UPMC-CNRS-MNHN-IRD 7138, Département Systématique et Evolution, Muséum National d'Histoire Naturelle, 43 rue Cuvier, 75005 Paris, France

E. Baudry
UMR CNRS 8079, Laboratoire Ecologie, Systématique et Evolution, Université Paris-Sud, Bat 362, 91405 Orsay Cedex, France