

The taxonomic challenge posed by the Antarctic echinoids Abatus bidens and Abatus cavernosus (Schizasteridae, Echinoidea).

Bruno David, Thomas Saucède, Anne Chenuil, Emilie Steimetz, Chantal de

Ridder

► To cite this version:

Bruno David, Thomas Saucède, Anne Chenuil, Emilie Steimetz, Chantal de Ridder. The taxonomic challenge posed by the Antarctic echinoids Abatus bidens and Abatus cavernosus (Schizasteridae, Echinoidea).. Polar Biology, 2016, 39 (5), pp.897-912. 10.1007/s00300-015-1842-5. hal-01310170

HAL Id: hal-01310170 https://hal.science/hal-01310170

Submitted on 23 Feb 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Click here to view linked References

1	The taxonomic challenge posed by the Antarctic echinoids Abatus bidens and Abatus cavernosus
2	(Schizasteridae, Echinoidea)
3	
4	
5	Bruno David ^{1,4} , Thomas Saucède ¹ , Anne Chenuil ² , Emilie Steimetz ¹ & Chantal De Ridder ³
6	
7	1 UMR6282 Biogéosciences, CNRS - Université de Bourgogne Franche-Comté, 6 boulevard Gabriel, F-21000 Dijon,
8	France
9	2 Institut Méditerranéen de Biodiversité et d'Ecologie marine et continentale, UMR CNRS 7263, Aix-Marseille
10	Université, IRD, Station Marine d'Endoume, Chemin de la Batterie des Lions, 13007 Marseille, France
11	3 Laboratoire de Biologie Marine (CP 160/15), Université Libre de Bruxelles, 50 avenue F.D. Roosevelt, B-1050
12	Brussels, Belgium
13	4 Muséum national d'Histoire naturelle, 57 rue Cuvier, 75005 Paris, France
14	
15	Corresponding author: Thomas Saucède
16	UMR6282 Biogéosciences, CNRS - Université de Bourgogne Franche-Comté, 6 boulevard Gabriel, F-21000 Dijon,
17	France
18	E-mail: thomas.saucede@u-bourgogne.fr
19	Phone number: +333 80 39 63 79
20	Fax number: +333 80 39 63 87
21	
าา	A history of

22 Abstract

23 Cryptic species have been repeatedly described for two decades among the Antarctic fauna, challenging the classic 24 model of Antarctic species with circumpolar distributions and leading to revisit the richness of the Antarctic fauna. No 25 cryptic species had been so far recorded among Antarctic echinoids, which are, however, relatively well-diversified in 26 the Southern Ocean. The R/V Polarstern cruise PS81 (ANT XXIX/3) came across populations of Abatus bidens, a 27 schizasterid so far known by few specimens that were found living in sympatry with the species Abatus cavernosus. 28 The species A. cavernosus is reported to have a circum-Antarctic distribution, while A. bidens is only recorded with 29 certainty in South Georgia and at the northern tip of the Antarctic Peninsula. Based on genetic and morphological 30 analyses, our results clearly show that A. bidens and A. cavernosus are two distinct species. The analyzed specimens 31 of A. bidens group together in two haplogroups separated from one another by 2.7% of genetic difference. They are 32 located in the Weddell Sea and in the Bransfield Strait. Specimens of A. cavernosus form one single haplogroup 33 separated from haplogroups of A. bidens by 5% and 3.5% of genetic difference respectively. The species was 34 collected in the Drake Passage and in the Bransfield Strait. Morphological analyses allow to differentiate A. bidens 35 from A. cavernosus. In contrast, the two genetic groups of A. bidens cannot be differentiated from one another based 36 on morphology alone, suggesting that they could represent a case of cryptic species, common in many Antarctic taxa, 37 but not yet reported in Antarctic echinoids. This need to be confirmed by complementary analyses of independent 38 genetic markers.

- 39
- 40 Keywords: Abatus bidens, Abatus cavernosus, cryptic species, Echinoidea, Schizasteridae, Southern Ocean

43 Introduction

44

45 For two decades, cryptic species have been repeatedly described among the Antarctic fauna, challenging the former, 46 classic biogeographic model of Antarctic species with wide, circumpolar distributions (De Broyer et al. 2011; Dettai 47 et al. 2011). The richness and spatial distribution of the Antarctic fauna have been revisited and the underpinning 48 evolutionary processes re-assessed (Kaiser et al. 2013). The occurrence of cryptic species in the Southern Ocean, and 49 on the Antarctic continental shelf in particular was highlighted in many studies of population genetics, in some cases 50 even signing the existence of species flocks (Lecointre et al. 2013). Cryptic species have been described in various 51 Antarctic groups: polychaetes (Schüller 2011), gastropods (Wilson et al. 2013), bivalves (Linse et al. 2007), 52 crustaceans (Held and Wägele 2005; Raupach and Wägele 2006), pycnogonids (Krabbe et al. 2009), cephalopods 53 (Allcock et al. 2010), crinoids (Wilson et al. 2007; Hemery et al. 2012), holothuroids (O'Loughlin et al. 2011), and 54 fish (Smith et al. 2011). Whatever the clade considered, depending on historical and environmental contingencies, the 55 processes leading to cryptic speciation can be related to spatial isolation, disruptive selection, and ecological 56 divergence. Cryptic species being recurrently described in various Antarctic clades, its origin must be soughed for in 57 mechanisms impacting the entire Antarctic fauna and linked to relatively fast landscape dynamics that is, rapid 58 alternation between allopatry and sympatry (Aguilée et al. 2012). A common explanation resides in the alternation 59 between glacial and interglacial periods that forces populations to isolated, glacial refugia or otherwise allows species 60 to expand their spatial ranges (Thatje et al. 2005; Allcock and Strugnell 2012; Aguilée et al. 2012).

61 Unexpectedly, no cryptic species had been so far recorded among Antarctic echinoids, which are, however, 62 relatively well-diversified in the Southern Ocean (Saucède et al. 2014). Spatangoid irregular echinoids in particular 63 form the most speciose group of echinoids in the Southern Ocean. It is exclusively represented there by the family 64 Schizasteridae and counts 30 morphologically recognized species and eight genera (David et al. 2005a, b; Pierrat et al. 65 2012, Saucède et al. 2014). Most species are brooders, the females incubating their young in deepened petals that 66 form kind of pouches or marsupia. Such a life history trait implies limited dispersal capabilities that might have 67 facilitated this outstanding diversification of the group also promoted by the alternation between glacial and 68 interglacial periods and the flow of the Antarctic Circumpolar Current (Poulin et al. 2002; Pearse et al. 2009). All 69 Antarctic species of Schizasteridae constitute a single monophyletic grouping, which presumably represent a species 70 "core flock" (sensu Lecointre et al. 2013) endemic to the Antarctic continental shelf including its deepest parts (David 71 et al. 2005a; Pearse et al. 2009; Lecointre et al. 2013; Saucède et al. 2014). Schizasterids are deposit feeders and soft-72 bottom dwellers, they live more or less buried into the superficial layer of sediments.

73 The systematics of Antarctic Schizasteridae has long been explored based on morphological observations (e.g. 74 Mortensen 1909, 1910; Koehler 1912; Pawson 1969; David et al. 2005a, b for the most recent reviews) but several 75 species are known by few specimens and still remain poorly described. Considering the significant variation of 76 morphological characters such as test outline and plating, development and position of fasciolar bands and of other 77 appendages, the taxonomy of Antarctic Schizasteridae and the phylogenetic relationships among species remain 78 puzzling, pending for new appraisals grounded on molecular markers. The R/V Polarstern cruise PS81 - ANT 79 XXIX/3 (Gutt 2013) came across populations of Abatus bidens, a schizasterid so far known by few specimens that 80 were found living in sympatry with the species Abatus cavernosus. The species were formerly considered as two

81 varieties of a same species (Mortensen 1951) before they were placed in two distinct entities (David et al. 2005a). The 82 species A. cavernosus is so far reported to have a circum-antarctic distribution, while A. bidens is only recorded with 83 certainty in South Georgia and at the northern tip of the Antarctic Peninsula. The later species was also reported in 84 Adelie Land where its occurrence remains uncertain (David et al. 2005a). The present study makes possible to fully 85 describe the species A. bidens for the first time and to assess morphological and genetic variations between the two 86 forms. What are the morphological and genetic differences between A. bidens and A. cavernosus? Are there reliable, 87 genetic differences to support the distinction between the two nominal species ? Are there transitional forms to 88 suggest that they are cryptic species if any?

89

90 Materials and methods

91

92 Material studied

93 Seventy nine specimens of A. bidens and A. cavernosus were collected at 19 stations in the northwest of the Weddell 94 Sea, in the Bransfield Strait, and in the Drake Passage during cruise PS81 of the R/V Polarstern (Fig. 1; Table 1). 95 Specimens were collected between 102 and 782 m depth using an Agassiz trawl (Gutt 2013). The two species were 96 found in sympatry at five of the 19 stations. Specimens were subsequently fixed in 96% ethanol and are now housed 97 in collections of the University of Burgundy (Dijon, France). Taxonomy of Antarctic species of the family 98 Schizasteridae is really challenging as shown for long by previous studies (David et al. 2005a, Saucède et al. 2014). 99 We therefore concentrated our study on recently sampled specimens during cruise PS81, for which a special care was 100 taken to control sample preservation, morphological study, and taxonomic identification. Genetic and morphometric 101 analyses were performed independently.

102

103 Genetic analyses

104 Genetic analyses initially included 62 specimens among which 49 provided informative sequences to be analyzed. 105 Tissue samples were taken on echinoid spines and DNA was extracted using the Chelex method following Chenuil 106 and Féral (2003). A typical barcoding region of the mitochondrial cytochrome oxidase gene was amplified by PCR 107 (5'- GCTCGTGC(A/G)TCTAC(A/G)TCCAT using the primers COIe3 -3') and COIe5 (5'-108 GC(C/T)TGAGC(A/T)GGCATGGTAGG -3') from Stockley et al. (2005). PCR amplicons were sent to the industry 109 (Eurofins Genomics) for Sanger sequencing. DNA sequences were manually trimmed and checked using the program 110 Bioedit (Hall 1999). Haplotype networks were built by the median-joining method using the Network software 111 (Bandelt et al. 1999). A color code was used to identify the morphological species, the distribution area (inside the 112 Weddell Sea, the Bransfield Strait, and the Drake Passage), and the sampling stations of each haplotype owing to the 113 NetworkPublisher software (Bandelt et al. 1999).

114

115 Morphological analyses

Echinoids were observed under a binocular microscope and morphology was analyzed using 10 quantitative parameters measured on echinoid tests with a digital caliper to the nearest 0.1 mm. These measurements were defined based on the morphological characters described in diagnoses of Antarctic echinoid species (David et al. 2005a). They are total test length (LL), total test width (WD), total test height (HT), distance between the apical system and the back of the test (AS), length of posterior petal I (PPT), length of anterior petal II (APT), distance between center of 121 the apical system and beginning of petal II (DP), distance between the periproct and the base of the test (PP), distance 122 between the peristome and the back of the test (PS), and total number of adjacent ambulacral plates abuting the 123 labrum (LB) (Fig. 2). Echinoid size was estimated using the geometric mean of test length, width and height (i.e., the 124 cube root of their product) and test shape (i.e., test proportions) was analyzed using seven indices computed from 125 ratios between some of the 10 measured quantitative parameters: shape at the ambitus (AMB=WD/LL), gibbosity 126 (GIB=HT/[\((LL.WD))), relative position of the peristome (RPS=PS/LL), relative position of the periproct 127 (RPP=PP/HT), relative position of the apical system (RAS=AS/LL), relative size of the petals (RPET=PPT/APT), 128 relative importance of the gap between the apical system and the beginning of petal II (RDP=DP/APT). 129 Morphometric analyses were performed on 75 specimens, 57 identified as A. bidens and 18 as A. cavernosus, using 130 the softwares Past (Hammer et al. 2001) and Statistica 6.1 (Statsoft 2002). Principal component analyses (PCAs) were 131 performed independently on the 10 quantitative parameters and on the seven indices to analyze morphological 132 variations within and between the two species. Correlation matrices were used to compute the PCAs as raw data show 133 contrasting units and range variations. Morphological differentiation between the two species was analyzed using 134 discriminant analyses based on quantitative measurements and on the indices independently. A priori groups were 135 determined based on genetic data (see below). These analyses were complemented by parametric tests as 136 homoscedasticity assumptions are respected (only parameters PP and LB and indices RPS and RPP show significantly 137 different variances between the two species).

138

Appendages were prepared and examined following David and Mooi (1990). Pedicellariae were removed from tests and placed into 96 % ethanol. They were bleached with a 10 % solution of sodium hypochlorite to remove soft tissue and to separate the valves. Then, they were washed in water, dried for 24 h, and mounted on SEM stubs. Details of pedicellariae were examined with a tabletop scanning electron microscope, and digital images were recorded.

143

144 **Results**

145

146 Genetic analyses

147 After trimming, the alignment file contained 633 homologous base pairs for 49 individuals. Seven haplotypes were 148 identified and form three divergent haplogroups, each group including two or three haplotypes (Fig. 3). Haplogroups 149 G1 (26 specimens) and G2 (18 specimens) are composed of specimens of A. bidens and are separated from one 150 another by a minimum of 17 mutations (i.e., 2.7% of genetic difference). The third haplogroup G3 (5 specimens) is 151 composed of specimens of A. cavernosus only, and is separated from haplogroups G1 and G2 by a minimum of 31 152 and 23 mutations respectively (i.e., 5% and 3.5% of genetic difference) (Fig. 3a). Haplogroups were identified at 11 of 153 the 19 sampling stations. Haplogroup G3 was identified at stations of the Drake Passage and in the Bransfield Strait, 154 while the two divergent haplogroups of A. bidens were found at stations of the Weddell Sea and of the Bransfield 155 Strait (Fig 3b). The three haplogroups occur in sympatry at two stations only (Sts. 220-2 and 224-3), the two 156 haplogroups of A. bidens, G1 and G2, are present together in 3 other stations (Sts. 164-4, 204-2, and 227-2), one 157 single haplotype was identified at the six remaining stations (Fig. 3c).

158

159 Morphological analyses

160 Within the 75 specimens measured, 41 were identified as *A. bidens* by genetic analyses, 26 belong to haplogroup G1,

- 161 15 to haplogroup G2, and 5 more specimens were identified as *A. cavernosus* and belong to haplogroup G3. A total of 162 29 specimens could not be analyzed for genetics and were finally attributed to their respective species based on 163 morphological results that is, 16 specimens to *A. bidens* and 13 to *A. cavernosus*. The distribution of specimens is 164 detailed in Table 1.
- 165 Statistical analyses of morphological data allow to clearly differentiate the two morphological species A. 166 bidens and A. cavernosus. Five parameters, in particular, were tested significantly different between the two species 167 (Student bilateral tests are detailed in Table 2): the distance between the peristome and the back of the test (PS), the 168 distance between the apical system and the back of the test (AS), the length of the posterior petal I (PPT), the distance 169 between the center of the apical system and the beginning of petal II (DP), and the total number of adjacent 170 ambulacral plates abuting the labrum (LB). Size was not tested significant between the two species (bilateral t-test 171 under homoscedasticity: p=0.47) so that morphological differences between species are not due to allometry. Sexual 172 dimorphism was not tested significant either for shape differences (Hotelling's T-squared: p=0.19 and p=0.29 for raw 173 parameters and shape indices respectively). Size only differs between females and males, females being larger than 174 males (unilateral t-test: p=0.014, mean size of females and males: 38.5 mm and 35.4 mm respectively).
- 175 The principal component analysis performed on the 10 quantitative parameters (Fig. 4) shows that the two 176 species mainly differentiate from one another along PC2 (14.2% of the total variance), which is mainly due to three 177 parameters: labrum extension (LB), length of posterior petal I (PPT), and position of the apical system (AS). The 178 principal component analysis performed on shape indices gave congruent results with a clear differentiation of the two 179 species along the two first PCs (62.9% of the total variance). Differentiation mainly concerns three indices: the shape 180 of the ambitus (AMB), the relative position of the apical system (RAS) and the posterior/ anterior ratio between petals 181 (RPET). Congruent results of the two PCAs show that specimens of A. bidens have a more extended labrum, a more 182 elongated ambitus, a more anterior apical system and a longer posterior petal than specimens of A. cavernosus. This 183 morphological difference is also supported by the discriminant analyses performed on raw parameters and shape 184 indices, with a high attribution ratio of 98.7% for both parameters and indices (Hotelling's T-squared: $p=9.3 \ 10^{-15}$ and 185 $p=2.3 \ 10^{-19}$ respectively).
- 186 Morphological variations between the two genetic groups of A. bidens, G1 and G2, were also analyzed. No 187 conspicuous difference was noticed between the two groups when specimens were observed at the naked eye and 188 under the binocular microscope, but statistical analyses revealed significant differences for three following 189 parameters: test height (HT), labrum extension (LB), and distance between the apical system and the beginning of 190 petal II (DP) (Table 3). Variation ranges, however, widely overlap and clear, diagnostic morphological characters 191 cannot be identified to differentiate the two groups. These results are also supported by the PCAs performed on raw 192 parameters (Fig. 5) and shape indices independently. The PCA of raw parameters shows no clear differentiation 193 between morphologies of the two groups along the first two PCs (Fig. 5a), the two groups best differentiating along 194 PC3, which only accounts for 9.35% of the total variance, though widely overlapping with one another (Fig. 5b). 195 These results are congruent with those of the discriminant analyses (Hotelling's T-squared: $p=1.6 \ 10^{-5}$ and p=0.005 for 196 raw parameters and shape indices respectively), the two groups best differentiating for raw parameters. This 197 discriminant analysis performed on raw parameters also shows that the two groups of A. bidens partly overlap with 198 one another in morphology but clearly differentiate from A. cavernosus (Fig. 6).

199 Morphological comparison of the three haplotypes demonstrates that they all differ significantly from each 200 other in some of the measured, raw parameters. Pairwise Mahalanobis distances between the three groups were all 201 tested significant regarding raw parameters, while for shape indices only the Mahalanobis distance between G1 and 202 G2 is not significant (Table 4). Haplogroup G1 turns out to be the most different from G3 in morphology, while G2 is 203 intermediate in position. This totally agrees with genetic results, the distance between G1 and G3 being the longest as 204 well (Fig. 3a).

205

206 Besides morphometric analyses, qualitative characters were also examined to complement the morphological 207 study, including observation of appendages (Table 5; Fig. 7). Some characters appear clearly diagnostic of A. 208 cavernosus and allow to differentiate it from A. bidens. In contrast, no single character allows to differentiate the two 209 groups of A. bidens from one another.

210 The test outline is heart-shaped in A. cavernosus, it is angular with a truncated posterior end in A. bidens. The 211 width/length ratio is significantly different between the two species, A. bidens being the most elongated (unilateral t 212 test on AMB: p=2.16 10⁻⁶). All petals are almost of the same size in A. bidens (mean posterior petals size reaches 93% 213 of that of anterior ones), while posterior petals are shorter in A. cavernosus (mean ratio = 64%). The color of test and 214 spines is light brown in A. bidens, darker in A. cavernosus. The fasciolar band appears beige in A. bidens, in which it 215 passes very low anteriorly, and dark purple in A. cavernosus. The apical system is anterior in position in A. bidens, 216 centered in A. cavernosus (it is situated at 39% and 51% of total test length in A. bidens and A. cavernosus 217 respectively).

218 Regarding plate patterns, in A. cavernosus the labrum is short and does not extend farther than the 1st, 219 infrequently the 2nd adjacent ambulacral plate, while in A. bidens the labrum extends as far as the 3rd adjoining 220 ambulacral plate, more rarely the 2nd and very seldom the 1st one. Marsupia abut the apical system in A. cavernosus, 221 what is the most frequent pattern showed by Antarctic brooding schizasterids, while they often do not start before 222 plates 8 or 9 in anterior petals of A. bidens, and plates 5 or 6 in the posterior ones (Fig. 8).

223 Pedicellariae of the two species were observed in details (Fig. 7). Globiferous pedicellariae are drastically 224 distinct between the two species. They are not very numerous in A. cavernosus, appear black in living specimens 225 when not cleaned of their soft tissue, and valves terminate in a series of four (rarely three) sharp, tiny teeth that are 226 rooted on the upper margin of a circular opening (Fig. 7a). In A. bidens, globiferous pedicellariae are extremely 227 numerous on the apical side, they are embedded in white tissue in living specimens, which turns dark in the specimens 228 fixed in ethanol. Valves terminate in two to three very long hooks (Fig. 7d). Rostrate pedicellariae of the two species 229 are relatively similar in shape except for the presence of a narrow, spiky distal tip in A. bidens (Fig. 7f), while it is 230 blunt and serrated in A. cavernosus (Fig. 7b). Tridentate pedicellariae belong to two main types: (1) the classic type 231 with a spoon-shaped distal part that tappers progressively toward the base and (2) a showel-shaped type with the distal 232 part separated from the base by a more or less straight shaft (Fig. 7c, e). Both types are present in the two species, but 233 they are more elongated in A. cavernosus (Fig. 7c) than in A. bidens (Fig. 7e).

234

Sphaeridae are somehow rounded in A. cavernosus (Fig. 7g), more elongated in A. bidens (Fig. 7h).

235

236 **Systematics**

237 The systematics and taxonomy of A. bidens is revised below based on the new genetic and morphological results 238 obtained in the present work. All results are congruent and show that A. bidens should be regarded as a distinct

239	species, following David et al. (2005a), and not a variety (sub-species) of A. cavernosus, contrary to Mortensen
240	(1951) and Kroh (2015). The description includes the two haplogroups of A. bidens identified in this study, G1 and
241	G2, herein treated as two putative cryptic species. All the specimens figured and studied in the present work are
242	housed in collections of the University of Burgundy, Dijon (France).
243	
244	Abatus bidens Mortensen 1910
245	Figures 7 and 8
246	
247	Abatus cavernosus var. bidens Mortensen, 1910; p. 73; Pl. 19: 32, 35, 39, 42.
248	Abatus cavernosus var. bidens Mortensen, 1951; p. 256.
249	Non Abatus bidens Bernasconi, 1953; p. 44; Pl. 24: 1-6; Pl. 25: 1-4.
250	Abatus bidens David et al., 2005a: 190–191.
251	Abatus cavernosus bidens Kroh, 2015:http://www.marinespecies.org/ophiuroidea/aphia.php?p=taxdetails&id=513701
252	Abatus bidens Kroh, 2015: http://www.marinespecies.org/ophiuroidea/aphia.php?p=taxdetails&id=160761
253	
254	Material: see Table 1 for location of material examined.
255	
256	Diagnosis (Table 5): apical system anterior in position. Anterior and posterior petals of equal length. Anterior branch
257	of the fasciole low on the test. Labrum extending posteriorly to the 2 nd or 3 rd adjacent ambulacral plate. Globiferous
258	pedicellariae very numerous on the apical side, particularly between petals and on the two margins of the anterior
259	ambulacrum, with valves terminating in two or three long hooks positioned at the end of a narrow, elongated aperture
260	(Fig. 7d). Living specimens yellow with whitish globiferous pedicellariae.
261	
262	Description
263	General morphology and plate pattern.
264	Size: The mean length of adult specimens is 46.7 mm, the females being slightly larger than males (unilateral t
265	test: $p = 0.008$).
266	Color: Living specimens beige to light brown, sometimes almost yellow (Fig. 8). The color of spines is light
267	beige. The fasciole is of the same color, turning dark in dry specimens. Plate pattern of the apical side is more or less
268	shown by subtle changes in coloration.
269	Outline of the test: Ambitus anteriorly rounded with a faint frontal notch and somehow posteriorly truncated,
270	making an angle covered with a tuft of spines on each side. Test width is 89% to 98% of test length (94% on average).
271	The posterior end of the test is vertically truncated, otherwise dome-shaped. Test height is 60% of test length.
272	Posterior petals are slightly shorter than anterior ones and converge at a more acute angle than anterior ones.
273	Apical system: It is slightly anterior in position, at 39% of the total test length from the anterior. Plating
274	follows the classic Abatus ethnolytic pattern with three gonopores (there is no gonopore in genital plate 2).
275	Gonopores larger in females.
276	Periproct: Located on the vertical posterior side, its adoral margin situated at 42% of test height from the lower
277	surface. It is scarcely visible in either apical or oral views. It is embedded between interambulacral plates 5.a.4/5.b.5

278 or 5.a.5/5.b.6 adorally, and 5.a.7/5.b.7 or 5.a.7/5.b.8 apically.

279 Peristome: It is relatively large and discernible as the anterior part of the labrum does not overhang over it. It is280 anterior in position, located at 31% of test length from the anterior of the test.

Plastron: In interambulacrum 5, the labrum is long, extending backward to the 3rd or 2nd adjacent ambulacral
 plates (sometimes the 4th).

Marsupiae (petals) and sexual dimorphism: Anterior and posterior petals almost of the same size, especially in haplogroup G1 (mean ratio = 0.97) compared to haplogroup G2 (mean ratio = 0.87). In most specimens, the adapical extremity of brood pouches is distant from the apical system, a morphological feature diagnostic of *A. bidens*. It is particularly conspicuous in females, in which anterior pouches initiate as far as the 8th to 9th ambulacral plates, while posterior ones are less distant from the apical system. This character is reminiscent of the plate pattern observed in *Abatus nimrodi*.

Fasciole: The peripetalous orthofasciole (*sensu* Néraudeau et al. 1998) is very conspicuous and form a broad band of 7 to over 15 rows of miliaries, depending on the segment considered and on test size. The fasciole is excentric anteriorly with the posterior segment distant from the posterior end of the test, and the anterior segment passing very close to the ambitus, barely visible in apical view.

294 Appendages

293

Primary spines: They are relatively coarse and of the usual schizasterid shape with finely serrated longitudinal ridges. The largest spines occur on the adoral side, in the plastronal and other interambulacral areas, and are used for locomotion. Their distal extremity is gently curved. The smallest spines are on the apical side and show rounded tips.

Secondary spines: Those observed on specimens collected at station 163-9 (Weddell Sea) are club-shaped and
 densely distributed.

300 Miliaries: They are made of a relatively loose stereom meshwork composed of 7 to 8 longitudinal ridges 301 separated by large openings. They end in a swollen serrated tip.

Clavulae: They are straight, made of fenestrated stereom, and end in a tuft of trabeculae. They are very similar
 in *A. bidens* and *A. cavernosus*.

Pedicellariae: The globiferous pedicellariae are extremely numerous, particularly on the apical side in the vicinity of petals and along each side of ambulacrum III. In living specimens, they appear as vivid yellow or whitish spots; they are brownish in specimens preserved in ethanol. Their valves terminate in two or three long hooks located at the top of an elongated opening (Fig. 7d). This character is shared with several other species of *Abatus* (e.g. *A. curvidens*, *A. agassizi*, and *A. elongatus*) and can putatively be regarded as plesiomorphic as it is also present in several species of *Amphipneustes* and *Tripylus*.

Tridentate pedicellariae (dentate pedicellariae are very commonly tridentate, sometimes bidentate) belong to two main types. As indicated above, valves of type 1 have a spoon-shaped distal part that tappers progressively toward the base (Fig. 7e). Valves of type 2 comprise three parts: the base, the intermediary tubular part, and the spoon-shaped, and finely serrated distal part (Fig. 7e). Rostrate pedicellariae display a very long and curved blade with small lateral teeth along the rim of the blade (Fig. 7f).

315

Sphaeridiae: They are slightly ovoid and ornamented with smal meridian ridges and grooves (Fig. 7h).

316

Distribution: A. bidens was reported with confidence by the Swedish South Polar and Discovery expeditions from
the South Georgia Islands, between 64 and 270 m (Mortensen 1910), and along the Antarctic Peninsula in the
Bransfield Strait and in the northwest of the Weddell Sea, between 102 and 782 m (this study).

320

Remarks: Mortensen (1951) regarded *A. bidens* as a subspecies of *A. cavernosus* but suggested that it could be a distinct species. David et al. (2005a) considered *A. bidens* as a separate species because globiferous pedicellariae of the two species are very different. Following Mortensen (1951) and David et al. (2005a), the specimens described by Bernasconi (1953) as *A. bidens* are considered here as representatives of *Tripylus excavatus* because adult individuals possess a conspicuous latero-anal fasciolar branch, absent in *A. bidens*. The two cryptic species (genetic groups G1 and G2) are here referred to as *A. bidens* pending for new data.

327

328 Discussion

In the present study, morphological and genetic results show that the two nominal species *A. cavernosus* and *A. bidens* are clearly distinct and should be regarded as truly different species as formerly suggested by David et al. (2005a). Several morphological features were recognized as clear, diagnostic characters of *A. bidens*, among which the number and shape of globiferous pedicellariae are the most remarkable (Table 5). Other distinctive, and more accessible characters, albeit not so clear cut, are petal relative size (anterior and posterior petals are of the same size in *A. bidens*) and the position of the apical system (anterior in *A. bidens*). These characters are reliable enough for the field determination of the species.

336 Unexpectedly, two genetic groups of A. bidens were identified. The genetic divergence between the two groups 337 (2.7%) is significant enough to suggest that they could be reproductively isolated from each other. This is congruent 338 with results obtained with molecule COI for other echinoderms. Genetic distance between closely related species of 339 echinoderms usually ranges between 0.62% and 1.3% (Ward et al. 2008; Hoareau and Boissin 2010). Some cryptic 340 species can even display up to 3% of genetic divergence (Egea et al. 2015). For instance, the most closely related 341 lineages within the species complex Ophioderma longicauda display a COI divergence of 2.2% (Boissin et al. 2011), 342 and cryptic species of the sand dollar Mellita longifissa show a genetic divergence comprised between 5.60% and 343 3.97% (Coppard et al. 2013). Hence, in the present study, the 2.7% of divergence between haplogroups G1 and G2 of 344 A. bidens, the absence of intermediate forms between them, and the short genetic distance between them and A. 345 cavernosus (3.5% to 5%) strongly suggest that they constitute two cryptic species, all the more as schizasterid 346 echinoids were shown to present slow rates of mitochondrial DNA evolution (Chenuil et al. 2008, 2010). The lack of 347 clear-cut discrimination within morphological variations of the two forms of A. bidens is in line with this 348 interpretation. Some morphological differences between the two genetic groups were tested significant but most 349 variations significantly overlap. Three evolutionary scenarios, however, can challenge our interpretation because they 350 too can generate divergent lineages in a biological species with no intermediate forms: a past demographic bottleneck, 351 a selective sweep, and a long-lasting vicariance that did not result in an efficient reproductive barrier before 352 "secondary" contact. The only way to rule these scenarios out and prove that haplogroups G1 and G2 form true 353 cryptic species of Abatus bidens, experimental crosses excepted, would consist in characterizing at least one 354 additional genetic marker independent from the mitochondrial COI gene that is, a DNA nuclear marker.

The two groups of *A. bidens* were collected in sympatry at the scale of sampling stations (they were found together at five of the eight stations at which they were identified). *A. cavernosus* was found living in sympatry with 357 A. bidens at five stations of the 12 stations at which it was collected, and the three haplogroups were found together at 358 four of these stations. Despite the limited number of sampling sites, the occurrence of species together was frequent 359 and attests that they can be living in sympatry. The co-existence of closely related species of similar forms is 360 generally considered a rare phenomenon as sympatry presumes the existence of differences in ecological niches and 361 life history traits. In environments of the Southern Ocean, such cases are known in amphipods (Baird et al. 2011), 362 isopods (Held 2001), and crinoids (Hemery et al. 2012). In irregular echinoids, sympatry was observed between 363 closely related species of the genus Echinocardium (between Echinocardium cordatum and Echinocardium flavescens 364 in Norway, pers. obs.), as well as between species of the sand dollar Mellita along the tropical coasts of the Americas 365 (Coppard et al. 2013). Sympatry between attested, cryptic species is much more rare. Examples are documented in the 366 sand dollar Mellita, in which only one of nine possible cases of sympatry between cryptic species was recorded 367 (Coppard et al. 2013), and in the species complex E. cordatum (Egea et al. 2011; Egea et al. 2015). Contrary to these 368 two case studies, all species of Abatus are brooders and consequently, show limited dispersal capability compared to 369 sea urchins with planktonic larvae. The co-existence of A. bidens and A. cavernosus at the same sites (Fig. 3c) is 370 therefore unexpected, all the more that previous studies revealed significant genetic differentiation between 371 populations of Abatus species at small spatial scale such as in Abatus cordatus (Poulin and Féral 1995; Ledoux et al. 372 2012), Abatus agassizi (Díaz et al. 2012), and Abatus nimrodi (Chenuil et al. 2004). In the present case study, the 373 mechanisms involved in the separation between the three forms cannot be uncovered with confidence. Morphological 374 similarities exclude the hypothesis of a strict ecological mechanism at the origin of the divergence for the three 375 groups, and the two forms of A. bidens in particular probably have similar modes of life. The separation is more likely 376 rooted in life history traits (e.g. shifts in spawning seasons) or in prezygotic isolation (i.e. incompatibility between 377 gamete recognition proteins) (Lessios 2011). Finally, it cannot be excluded that haplogroups of A. bidens may 378 interbreed with no fitness cost and belong to the same biological species, nor that post-zygotic incompatibility only is 379 involved in the isolation of the two forms despite the evolutionary cost it would imply.

380 At the scale of the three geographic areas explored, the two forms of A. bidens were not collected in the Drake 381 Passage, while conversely, A. cavernosus was frequently found there (at 83% of the sampling sites). The latter species 382 is also present in the Bransfield Strait but it was less frequently found than A. bidens, and it is absent from the 383 northwest of the Weddell Sea (namely no sequence of A. cavernosus was identified in the Weddell Sea) (Fig. 9). Two 384 distinct distribution patterns seem to appear and to follow reverse, latitudinal gradients: A. cavernosus is the most 385 frequently encountered to the north, and most often outside the areas strongly impacted by sea ice, while the two 386 forms of A. bidens are the most frequent to the south in areas under sea ice influence (Fig. 9). Though still in need for 387 further research, these apparent patterns are supported by records of A. cavernosus at the tip of South America 388 (Bernasconi 1925, 1966) and by the putative presence of A. bidens in Adelie Land (Koehler 1926) described 389 globiferous pedicellariae of "A. cavernosus" with two long hooks, suggesting that its specimens should be regarded as 390 A. bidens). The two species have been also unquestionably recognized to co-occur in South Georgia (Bernasconi 391 1953). This is not contradictory with the proposed pattern as South Georgia has been often regarded as a 392 biogeographic area under the cross influence of Antarctic and sub-Antarctic regions. Although unevenly distributed, 393 the three haplogroups co-occur in the Bransfield Strait where they can be found in sympatry. Such a diversity pattern 394 has already been highlighted in previous studies for other organisms (Wilson et al. 2007; Hemery et al. 2012). Along 395 with the Scotia Arc region and the northern tip of the Antarctic Peninsula, the Bransfield Strait is a favored zone of connectivity among populations due to the strong current systems present near the tip of the Antarctic Peninsula(Thompson et al. 2009).

At the scale of the entire Southern Ocean, the attested occurrence of *A.bidens* in the Peninsula area as well as in Adelie Land suggests that the species has a wide, potentially circumpolar distribution. The respective distribution of the two halpogroups still remain to be clarified, in East Antarctica in particular. In contrast, *A. cavernosus* seems to have a more northern distribution, potentially restricted to the Magellanic and Scotia Arc regions as no report of *A. cavernosus* outside this area is fully certain. This distribution pattern is in complete opposition with previous works (David et al. 2005a).

404

405 Conclusion

406 Pending for further genetic analyses including closely related species of *Abatus* (i.e. *Abatus philippi*, *A. agassizi*, and 407 *A. cordatus*) and additional samplings in other areas of the Southern Ocean, the present study, however, suggests that 408 the *A. cavernosus - A. bidens* complex comprises at least two morphologically close species and two cryptic species. 409 It contributes to clarifying the systematics of *A. cavernosus* and *A. bidens* and shows that *A. bidens* should definitely 410 be regarded as distinct from *A. cavernosus*, based on congruent morphological characters and genetic results. In 411 addition, *A. cavernosus* and the two forms of *A. bidens* were shown to have distinct distribution patterns.

The present work is also the first study to report the occurrence of cryptic species of echinoids in the Southern Ocean, in accordance with many observations of various marine invertebrates of the Southern Ocean. It leads to revisit the richness of the Antarctic echinoid fauna, its systematics, macroecological patterns, and to re-assess the underpinning evolutionary processes and history. It also highlights the crucial importance of associated morphological and genetic studies for improving our knowledge of Antarctic biodiversity. This is a prerequisite for further processbased studies, from ecology to phylogeography, and for uncovering the origin of the rich Antarctic biodiversity.

418

419 Acknowledgements

Samples were collected during the oceanographic campaign PS81 - ANT-XXIX/3 of the R/V *Polarstern*. The authors
would like to thank Prof. Dr. Julian Gutt, chief scientist of this campaign, as well as the crew and the scientific staff.
Chantal De Ridder was supported by F.R.S-FNRS "short stay abroad" travel grants (grant nr. 2013/V3/5/035). This is
contribution nr. XX to the vERSO project (www.versoproject.be), funded by the Belgian Science Policy Office (BELSPO,
contract nr. BR/132/A1/vERSO). This is a contribution to team BioME of the CNRS laboratory Biogéosciences
(UMR 6282), and MARE publication nr. YY.

- 426
- 427

428 **Conflict of interest**

429 The authors declare that they have no conflict of interest.

430

431

432 References

- 433 Aguilée R, Claessen D, Lambert A (2012) Adaptive radiation driven by the interplay of eco-evolutionary and 434 landscape dynamics. Evolution 67(5):1291-1306. doi:10.1111/evo.12008
- 435 Allcock AL, Barratt I, Eléaume M, Linse K, Smith PJ, Steinke G, Stevens DW, Norman MD, Strugnell JM (2010)

- 436 Cryptic speciation and the circumpolarity debate: A case study on endemic Southern Ocean octopuses using
 437 the COI barcode of life. Deep-Sea Res II 58:242-249.
- Allcock AL, Strugnell JM (2012) Southern Ocean diversity: new paradigms from molecular ecology. Trends Ecol
 Evol 27:520-528
- Bandelt HJ, Forster P, Röhl A (1999) Median-joining networks for inferring intraspecific phylogenies. Mol Biol Evol
 16:37-48
- 442 Bernasconi I (1925) Resultados de la primera expedicion a Tierra del Fuego (1921). Echinodermos. I Equinoideos.
 443 Anales Soc Ci Argent 98:3-17
- 444 Bernasconi I (1953) Monografia de los equinoideos argentinos. Anales Mus Nac Hist Nat Buenos Aires 2(6):1-58
- Bernasconi I (1966) Los equinoideos y asteroideos colectados por el buque oceanografico R/V "Vema", frente a las
 costas argentinas, uruguayas y sur de Chile. Revista Mus Argent Ci Nat "Bernardino Rivadavia" 9:147-175
- Coppard S, Zigler KS, Lessios HA (2013) Phylogeography of the sand dollar genus *Mellita*: Cryptic speciation along
 the coasts of the Americas. Mol Phylogenet Evol 69:1033-1042
- Boissin E, Stöhr S, Chenuil A (2011) Did vicariance and adaptation drive cryptic speciation and evolution of brooding
 in *Ophioderma longicauda* (Echinodermata: Ophiuroidea), a common atlanto-mediterranean ophiuroid? Mol
 Ecol 20:4737-4755
- 452 Chenuil A, Gault A, Féral JP (2004) Paternity analysis in the Antarctic brooding sea urchin *Abatus nimrodi*. A pilot
 453 study. Polar Biol 27:117-182
- 454 Chenuil A, Egea E, Rocher C, Touzet H, Féral JP (2008) Does hybridization increase evolutionary rates ? Data from
 455 the 28S-rDNA D8 domain in echinoderms. J Mol Evol 67:539–550
- 456 Chenuil A, Féral JP (2003) Sequences of mitochondrial DNA suggest that *Echinocardium cordatum* is a complex of
 457 several sympatric or hybridizing species. A pilot study. In: Féral JP & David B (eds) Echinoderm Research
 458 2001, Swets & Zeitlinger, Lisse, pp 15-21
- Chenuil A, Egea E, Rocher C, Féral JP (2010) Comparing substitution rates in spatangoid sea urchins with putatively
 different effective sizes, and other echinoderm datasets. In: Harris LG, Böttger SA, Walker CW, Lesser MP
 (eds) Echinoderms Durham, Balkema, Leiden, pp 159-161
- 462 David B, Choné T, Mooi R, De Ridder C (2005a) Antarctic Echinoidea. Synopses of the Antarctic Benthos. Koeltz
 463 Scientific Books, Königstein
- 464 David B, Choné T, Mooi R, De Ridder C (2005b) Biodiversity of Antarctic echinoids: a comprehensive and
 465 interactive database. Sci Mar 69:201–203
- 466 De Broyer C, Danis B et al (2011) How many species in the Southern Ocean? Towards a dynamic inventory of the
 467 antarctic marine species. Deep-Sea Res II 58:5-17
- 468 Dettai A, Adamowizc SJ, Allcock L, Arango CP, Barnes DKA, Barratt I, Chenuil A, Couloux A et al (2011) DNA
 469 Barcoding and molecular systematics of the benthic and demersal organisms of the CEAMARC survey.
 470 Polar Science 5:298–312
- 471 Díaz A, González-Wevar CA, Maturana CA, Palma AT, Poulin E, Gérard K (2012) Restricted geographic distribution
 472 and low genetic diversity of the brooding sea urchin *Abatus agassizii* (Spatangoidea: Schizasteridae) in the
 473 South Shetland Islands: a bridgehead population before the spread to the northern Antarctic Peninsula. Rev
 474 Chil Hist Nat 85:457-468

- 475 Egea E, David B, Choné T, Laurin B, Féral JP, Chenuil A (2015) Morphological and genetic analyses reveal a cryptic
 476 species complex in the echinoid *Echinocardium cordatum* and rule out a stabilizing selection explanation. Mol
 477 Phylogenet Evol. doi:10.1016/j.ympev.2015.07.023
- 478 Egea E, Mérigot B, Mahé-Bézac C, Féral JP, Chenuil A (2011) Differential reproductive timing in *Echinocardium* 479 *spp.*: the first Mediterranean survey allows inter-oceanic and inter-specific comparisons. C R Biol 334:13-23
- 480 Gutt J (2013) The expedition of the research vessel "Polarstern" to the Antarctic in 2013 (ANT-XXIX/3). Berichte zur
 481 Polar- und Meeresforsch 665:1-151
- Hall TA (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows
 95/98/NT. Nucleic Acids Symp Ser 41:95-98
- Hammer Ø, Harper DAT, Ryan PD (2001) PAST: Paleontological Statistics Software Package for Education and Data
 Analysis. Pal Elec 4(1):1-9
- Held C (2001) No evidence for slow-down of molecular substitution rates at subzero temperatures in Antarctic serolid
 isopods(Crustacea,Isopoda, Serolidae). Polar Biol 24:497-501
- Held C, Wägele JW (2005) Cryptic speciation in the giant Antarctic isopod *Glyptonotus antarcticus* (Isopoda:
 Valvifera: Chaetiliidae). Sci Mar 69(2):175-181
- Hemery LG, Eléaume M, Roussel V, Améziane N, Gallut C, Steinke D, Cruaud C, Couloux A, Wilson NG (2012)
 Comprehensive sampling reveals circumpolarity and sympatry in seven mitochondrial lineages of the Southern
 Ocean crinoid species *Promachocrinus kerguelensis* (Echinodermata). Mol Ecol 21:2502-2518
- Hoareau TB, Boissin E (2010) Design of phylum-specific hybrid primers for DNA barcoding: addressing the need for
 efficient COI amplification in the Echinodermata. Mol Ecol Res 10:960–967
- Kaiser S, Brandão SN, Brix S, Barnes DKA, Bowden DA, David B, Gutt J, et al (2013) Pattern, process and
 vulnerability of Antarctic and Southern Ocean benthos a decadal leap in knowledge and understanding. Mar
 Biol 160:2295-2317
- Koehler R (1912) Echinodermes nouveaux recueillis dans les mers antarctiques par le Pourquoi-pas (astéries,
 ophiures et échinides). Zool Anz 39:151-163
- Koehler R (1926) Echinodermata Echinoidea. In: Australasian Antarctic expedition 1911-1914. Sci Rep (C8):1 134
- Krabbe K, Leese F, Mayer C, Tollrian R, Held C (2009) Cryptic mitochondrial lineages in the widespread pycnogonid
 Colossendeis megalonyx Hoek, 1881 from Antarctic and Subantarctic waters. Polar Biol 33:281-292
- Kroh A (2015) *Abatus cavernosus bidens* Mortensen, 1910. In: Kroh A and Mooi R(eds) World Echinoidea Database.
 Accessed through: World Register of Marine Species
- Lecointre G, Améziane N, Boisselier MC, Bonillo C, Busson F, Causse R, Chenuil A, Couloux A et al (2013) Is the
 Species Flock Concept Operational? The Antarctic Shelf Case. PloS ONE 8:e68787
- Ledoux JB, Tarnowska K, Gérard K, Lhuillier E, Jacquemin B, Veydmann A, Féral JP, Chenuil A (2012) Fine-scale
 spatial genetic structure in the brooding sea urchin *Abatus cordatus* suggests vulnerability of the Southern
 Ocean marine invertebrates facing global change. Polar Biol 35:611-623
- 511 Lessios HA (2011) Speciation genes in free-spawning marine invertebrates. Integr Comp Biol 51:456-465
- 512 Linse K, Cope T, Lörz AN, Sands C (2007) Is the Scotia Sea a centre of Antarctic marine diversification? Some
- 513 evidence of cryptic speciation in the circum-Antarctic bivalve *Lissarca notorcadensis* (Arcoidea:
- 514 Philobryidae). Polar Biol 30:1059-1068

515 Mortensen T (1909) Die Echinoiden der Deutschen Südpolar Expedition 1901-1903. Deutsche Südpolar 516 Expedition. G. Reimer, Berlin 517 Mortensen T (1910) The Echinoidea of the Swedish South Polar Expedition. Wissenschaftliche Ergebnisse der 518 Schwedischen Südpolar-Expedition 1901-03. Lithographisches Institut des Generalstabs, Stockholm 519 Mortensen T (1951) A monograph of the Echinoidea. Vol. 5.2 Spatangoida II. Reitzel CA, Copenhagen 520 Néraudeau D, David B, Madon C (1998) Tuberculation in spatangoid fascioles: Delineating plausible homologies. 521 Lethaia 31:323-334 522 O'Loughlin, PM, Paulay G, Davey N, Michonneau F (2011) The Antarctic region as a marine biodiversity hotspot for 523 echinoderms: diversity and diversification of sea cucumbers. Deep-Sea Res II 58, 264-275 524 Pawson DL (1969) Echinoidea. In: Bushnell VC, Hedgpeth JW (eds) Distribution of selected groups of marine 525 invertebrates in water south of 35° S latitude. Antarctic map folio. Amer Geograph Soc, New York, pp 526 38-41 527 Pearse J, Mooi R, Lockhart SJ, Brandt A (2009) Brooding and Species Diversity in the Southern Ocean: Selection for 528 Brooders or Speciation within Brooding Clades? In: Krupnik I, Lang MA, Miller SE (eds) Smithsonian at the 529 Poles: Contributions to International Polar Year science. Smithsonian Institution Scholarly Press, 530 Washington, pp 181-196 531 Pierrat B, Saucède T, Festeau A, David B (2012) Antarctic, sub-Antarctic and cold temperate echinoid database. 532 ZooKeys 204:47-52. doi:10.3897/zookeys.204.3134 533 Poulin E, Féral J-P (1995) Pattern of spatial distribution of a brood-protecting Schizasterid Echinoid, Abatus cordatus, 534 endemic to Kerguelen Islands, Mar Ecol Prog Ser 118:179-186 535 Poulin E, Palma AT, Féral J-P (2002) Evolutionary versus ecological success in Antarctic benthic invertebrates. 536 Trends Ecol Evol 17:218–222 537 Raupach MJ, Wägele JW (2006) Distinguishing cryptic species in Antarctic Asellota (Crustacea: Isopoda) - a 538 preliminary study of mitochondrial DNA in Acanthaspidia drygalskii. Antarctic Sci 18:191-198 539 Saucède T, Pierrat B, David B (2014) Chapter 5.26. Echinoids. In: De Broyer C, Koubbi P, Griffiths HJ, Raymond B, 540 d'Udekem d'Acoz C et al (eds) Biogeographic atlas of the Southern Ocean. Scientific Committee on Antarctic 541 Research, Cambridge, pp 213-220 542 Schüller M (2011) Evidence for a role of bathymetry and emergence in speciation in the genus Glycera (Glyceridae, 543 Polychaeta) from the deep Eastern Weddell Sea. Polar Biol 34:549-564 544 Smith PJ, Steinke D, McMillan PJ, Stewart AL, McVeagh SM, Diaz de Astarloa JM, Welsford D, Ward RD (2011) 545 DNA barcoding highlights a cryptic species of grenadier Macrourus in the Southern Ocean. J Fish Biol 546 78:355-365 547 Statsoft France (2002) STATISTICA (logiciel d'analyse de données). Versio 6, www.Statsoft.com 548 Thatje S, Hillenbrand C, Larter R (2005) On the origin of Antarctic marine benthic community structure. Trends Ecol 549 Evol 20:534-540 550 Thompson AF, Heywood KJ, Thorpe SE, Renner AHH, Trasvina A (2009) Surface circulation at the tip of the 551 Antarctic Peninsula from drifters. J Phys Oceanogr 39:3-26 552 Ward RD, Holmes BH, O'Hara TD (2008) DNA barcoding discriminates echinoderm species. Mol Ecol Res 8:1202-553 1211

- 554
- Wilson NG, Hunter RL, Lockhart SJ, Halanych KM (2007) Multiple lineages and absence of panmixia in the 555 "circumpolar" crinoid Promachocrinus kerguelensis from the Atlantic sector of Antarctica. Mar Biol 152:895-556 904
- 557 Wilson NG, Maschek A, Baker BJ (2013) A Species Flock Driven by Predation? Secondary Metabolites Support 558 Diversification of Slugs in Antarctica. PloS ONE 8:e80277
- 559

560 Figure and table captions

561

562 Fig. 1 The 19 stations in the Weddell Sea (vellow dots), Bransfield Strait (orange dots), and Drake Passage (blue dots) 563 where the studied specimens of Abatus cavernosus and Abatus bidens were collected during cruise PS81 of the R/V 564 Polarstern

565

566 Fig. 2 Measurements performed in the morphometric analysis. a, echinoid test drawn in apical view (fasciolar band in 567 light grey, brood pouches in black, apical system detailed in the center). b, test in posterior view (fasciolar band in 568 light grey, brood pouches and periproct in black). c, test in oral view with details of plate patterns (peristome in 569 black). a, total test length (LL) and width (WD), distance between the apical system and the back of the test (AS), 570 length of posterior petal I (PPT), length of anterior petal II (APT), distance between center of the apical system and 571 beginning of petal (pouch) II (DP). b, total test height (HT), distance between the periproct and the base of the test 572 (PP). c, distance between the peristome and the back of the test (PS); total number of adjacent ambulacral plates 573 abuting the labrum (LB). A maximum of six ambulacral plates (3 per ambulacrum) can abut the labrum in Abatus 574 *bidens*; two ambulacral plates only abut the labrum in the specimen drawn in Figure 2c

575

576 Fig. 3 Median-joining network of haplotypes based on mtDNA COI sequences obtained from 5 specimens of Abatus 577 cavernosus and 44 specimens of Abatus bidens. a, each haplotype is labelled from H1 to H8, and shown in a colored 578 circle indicating the genetic unit to which it belongs: G1 (light green), G2 (dark green), and G3 (red). Size of circles 579 proportional to frequency in the sample analyzed. Mutations shown with unlabelled red dots. Minimum pairwise 580 distances (minimum number of mutations) between specimens of the three groups shown in the table inserted. b, 581 colors indicating the sampling geographic area of each haplotype: the Bransfield Strait (orange), Weddell Sea 582 (yellow), and Drake Passage (blue). c, colors indicating the sampling stations of each haplotype

583

584 Fig. 4 Principal component analysis of the ten raw morphometric parameters measured in specimens of Abatus bidens 585 (green dots) and Abatus cavernosus (red dots). The two species differentiate along the first two principal components, 586 which account for 75.06% of the total variance

587

588 Fig. 5 Principal component analysis of the ten raw morphometric parameters measured in specimens of Abatus bidens 589 (G1, light green dots; G2, dark green dots). a, the first two principal components, which acount for 78.82% of the total 590 variance. b, first and third principal components, which accounts for 76.53% of the total variance

591

592 Fig. 6 Discriminant analysis of the ten raw morphometric parameters measured in specimens of Abatus bidens (G1, 593 light green dots; G2, dark green dots) and *Abatus cavernosus* (G3, red dots)

595 Fig. 7 Pedicellariae and sphaeridiae in Abatus cavernosus (a-c, g) and Abatus bidens (d-f, h). a, isolated valve of 596 globiferous pedicellaria from specimen UBGD 279061 (G3). b, valves of rostrate pedicellariae from specimens 597 UBGD 279063 (G3), UBGD 279061 (G3), and UBGD 279064 (G3) from left to right. c, valves of tridentate 598 pedicellariae from specimens UBGD 279062 (G3), UBGD 279064 (G3), and UBGD 279063 (G3) from left to right. 599 d, valves of globiferous pedicellariae from specimens UBGD 279067 (G1) and UBGD 279070. e, tridentate 600 pedicellariae from specimen UBGD 279067 (G1). f, rostrate pedicellariae from specimens UBGD 279069 (G1), and 601 UBGD 279070. g, sphaeridiae from specimens UBGD 279062 (G3) and UBGD 279064 (G3). h, sphaeridiae from 602 specimens UBGD 279070 and UBGD 279068

603

Fig. 8 *Abatus cavernosus* (a-d) and *Abatus bidens* (e-h). a, apical view of fresh specimen UBGD 279059. b, apical, c,
oral, and d, posterior views of denuded test of specimen UBGD 279060. e, apical view of fresh specimen UBGD
279065. f, apical, g, oral, and h, posterior views of denuded test of specimen UBGD 279066. Scale bars: 10 mm

607

Fig. 9 Distribution map of the genetic groups of *Abatus cavernosus* and *Abatus bidens* (G1, light green; G2, dark

green; G3, red) among the 14 stations where they were identified. Circle wedges indicate presence/absence of geneticgroups, not abundance

611

612 **Table 1** Sites where specimens of the species complex *Abatus cavernosus – Abatus bidens* were collected. Crosses

- 613 indicate where the haplogroups were formally identified and where non-sequenced specimens were identified based
- on morphology alone.
- 615

Sampling sites		Abatus bidens	Abatus cavernosus		
Sampling sites	haplogroup G1	haplogroup G2	non-sequenced	haplogroup G3	non-sequenced
Weddell (162-7)		Х			
Weddell (163-9)	X	Х			Х
Weddell (164-4)	X	Х			Х
Weddell (188-4)			Х		
Weddell (190-2)			Х		
Bransfield (116-9)				Х	
Bransfield (196-8)			Х		
Bransfield (197-5)	X				
Bransfield (198-5)		Х			Х
Bransfield (199-4)			Х		
Bransfield (204-2)	X	Х			
Bransfield (205-1)			Х		
Bransfield (220-2)	X	Х		Х	
Bransfield (224-3)	X	Х		Х	
Bransfield (227-2)	X				
Drake (234-5)				Х	
Drake (237-3)					Х
Drake (240-3)					Х
Drake (245-1)					Х
_	1				
Table 2 Morphologicprobability values for	al differences betwe Student bilateral tes	en haplogroups of a	Abatus bidens and a es only are detailed	A <i>batus cavernosus</i> . I (abbreviations are	The table gives th given in Fig. 2)

619
620

	Parameters	LL, WD, HT, PP, APT	PS	AS	PPT	DP	LB		
	p values	> 0.05	0.004	1.6 10-7	1.2 10-9	0.015	1.3 10-6		
621		I							
622									
623	Table 3 Mo	rphological differences be	tween ha	plogroups	G1 and G	62 of Ab	atus bidens	s. The table gives the probabili	ty
624	values for Student bilateral tests. Significant values only are detailed (abbreviations are given in Fig. 2)								
625									
	Parameters	LL, WD, PS, PP, AS, PF	PT, APT	HT	DP LB				

> 0.05

0.02

0.01

0.01

p values

Table 4 Pairwise Mahalanobis distances between haplogroups G1, G2, and G3 of *Abatus bidens* and *Abatus cavernosus* respectively. The table gives the probability values for comparisons using raw variables (matrix upper
 half) and shape indices (matrix lower half). Asterisks indicate if differences are significant (*) or highly significant
 (**)



Table 5 Diagnostic morphological characters of *Abatus bidens* (including the two genetic groups) that clearly differ
 from those of *Abatus cavernosus*. Characters are listed in order of importance for distinguishing the two species

	A. bidens	A. cavernosus	
Globiferous pedicellariae	They are numerous, particularly on	They are rare. When present, a	
	the apical side. Their valves	series of three to four teeth borders	
	terminate by two long hooks	the upper rim of a rounded	
	located above a narrow, elongated	aperture.	
	aperture.		
Petals	Posterior petals about the same	Posterior petals shorter than the	
	length as the anterior ones.	anterior ones.	
Color	Living specimens yellow.	Living specimens brownish.	
	Globiferous pedicellaria whitish.		
Apical system and fasciole	Shifted anteriorly.	Centered.	
Labrum	It extends posteriorly to the 2 nd or	It extends posteriorly to the 1st (or	
	3 rd adjacent ambulacral plates.	2 nd) adjacent ambulacral plates.	
Brooding pouches (in females)	They often start away from the	Always starting close to the apical	
	apical system.	system.	
Sphaeridiae	Elongated	Rounded or flattened	







а

b

С









root 2 4 3 2 \bigcirc \bigcirc 1 \bigcirc \bigcirc 0 0 \bigcirc • G3 -1 \bigcirc 6 \bigcirc • G2 \bigcirc • G1 -2 root 1 -3 -3 -2 -1 0 2 3 -4 1 4 5





