



**HAL**  
open science

## **Pithovirus sibericum, a new bona fide member of the "Fourth TRUC" club**

Vikas Sharma, Philippe Colson, Olivier Chabrol, Pierre Pontarotti, Didier  
Raoult

► **To cite this version:**

Vikas Sharma, Philippe Colson, Olivier Chabrol, Pierre Pontarotti, Didier Raoult. Pithovirus sibericum, a new bona fide member of the "Fourth TRUC" club. *Frontiers in Microbiology*, 2015, 6, pp.722. 10.3389/fmicb.2015.00722 . hal-01304145

**HAL Id: hal-01304145**

**<https://hal.science/hal-01304145>**

Submitted on 12 Oct 2018

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

# *Pithovirus sibericum*, a new *bona fide* member of the “Fourth TRUC” club

Vikas Sharma<sup>1,2</sup>, Philippe Colson<sup>1,3</sup>, Olivier Chabrol<sup>2</sup>, Pierre Pontarotti<sup>2</sup> and Didier Raoult<sup>1,3\*</sup>

<sup>1</sup> Unité de Recherche sur les Maladies Infectieuses et Tropicales Emergentes, UMG3, Centre National de la Recherche Scientifique 7278, IRD 198, Institut National de la Santé et de la Recherche Médicale U1095, Aix-Marseille University, Marseille, France, <sup>2</sup> I2M UMR 7373, Centre National de la Recherche Scientifique Équipe Evolution Biologique et Modélisation, Aix-Marseille University, Marseille, France, <sup>3</sup> Institut Hospitalo-Universitaire (IHU) Méditerranée Infection, Pôle des Maladies Infectieuses et Tropicales Clinique et Biologique, Fédération de Bactériologie-Hygiène-Virologie, Centre Hospitalo-Universitaire Timone, Assistance Publique-Hôpitaux de Marseille, Marseille, France

## OPEN ACCESS

### Edited by:

William Michael McShan,  
University of Oklahoma Health  
Sciences Center, USA

### Reviewed by:

Iara Apolinario Borges,  
Universidade Federal de Minas Gerais,  
Brazil

Betania Paiva Drumond,  
Universidade Federal de Juiz de Fora,  
Brazil

### \*Correspondence:

Didier Raoult,  
Unité des Rickettsies, Faculté de  
Médecine, Aix-Marseille Université, 27  
Boulevard Jean Moulin,  
13385 Marseille, France  
didier.raoult@gmail.com

### Specialty section:

This article was submitted to  
Virology,  
a section of the journal  
Frontiers in Microbiology

**Received:** 10 May 2015

**Accepted:** 02 July 2015

**Published:** 04 August 2015

### Citation:

Sharma V, Colson P, Chabrol O,  
Pontarotti P and Raoult D (2015)  
*Pithovirus sibericum*, a new *bona fide*  
member of the “Fourth TRUC” club.  
*Front. Microbiol.* 6:722.  
doi: 10.3389/fmicb.2015.00722

Nucleocytoplasmic large DNA viruses, or representatives of the proposed order *Megavirales*, include giant viruses of *Acanthamoeba* that were discovered over the last 12 years and are *bona fide* microbes. Phylogenies based on a few genes conserved amongst these megaviruses and shared by microbes classified as *Eukarya*, *Bacteria*, and *Archaea*, allowed for delineation of a fourth monophyletic group or “TRUC” (Things Resisting Uncompleted Classification) composed of the *Megavirales* representatives. A new *Megavirales* member named *Pithovirus sibericum* was isolated from a >30,000-year-old dated Siberian permafrost sample. This virion is as large as recently described pandoraviruses but has a genome that is approximately three to four times shorter. Our objective was to update the classification of *P. sibericum* as a new member of the “Fourth TRUC” club. Phylogenetic trees were constructed based on four conserved ancient genes and a phyletic analysis was concurrently conducted based on the presence/absence patterns of a set of informational genes from members of *Megavirales*, *Bacteria*, *Archaea*, and *Eukarya*. Phylogenetic analyses based on the four conserved genes revealed that *P. sibericum* is part of the fourth TRUC composed of *Megavirales* members, and is closely related to the families *Marseilleviridae* and *Ascoviridae/Iridoviridae*. Additionally, hierarchical clustering delineated four branches, and showed that *P. sibericum* is part of this fourth TRUC. Overall, phylogenetic and phyletic analyses using informational genes clearly indicate that *P. sibericum* is a new *bona fide* member of the “Fourth TRUC” club composed of representatives of *Megavirales*, alongside *Bacteria*, *Archaea*, and *Eukarya*.

**Keywords:** giant virus, *Pithovirus sibericum*, *Megavirales*, nucleocytoplasmic large DNA viruses, informational genes, TRUC, domains of life, phylogeny

## Introduction

Nucleocytoplasmic large DNA viruses (NCLDV), or members of the proposed order *Megavirales*, are the largest known viruses thus far; their genome ranges in size from ≈100 to 2500 kilobase pairs (kbp) and they are visible on photonic microscopy (Iyer et al., 2006; Raoult and Forterre, 2008; Yutin et al., 2009; Yutin and Koonin, 2012; Colson et al., 2013; Philippe et al., 2013; Legendre et al., 2014; Raoult, 2014). These viruses rely on very diverse eukaryotic hosts for their life cycle, which

include protists, algae, vertebrate animals, and insects. They include giant viruses whose existence was revealed 12 years ago with the discovery of Mimivirus, a *Acanthamoeba polyphaga*-resisting microbe that was initially considered a Gram positive bacterium (La Scola et al., 2003; Raoult et al., 2004) and then discovered to have a gigantic 1200 kbp-large genome encompassing approximately 1000 genes (Raoult et al., 2007; Raoult, 2014). Moreover, this genome was predicted to encode several cellular trademark genes never previously reported in any kind of viruses. Afterwards, several dozen new giant amoebal viruses were discovered that were almost all isolated on *Acanthamoeba* spp., including other mimiviruses (Fischer et al., 2010; Arslan et al., 2011; Yoosuf et al., 2012), but also smaller giants that founded the family *Marseilleviridae* (Aherfi et al., 2014), then pandoraviruses, which are the current record holders for virion and genome size (Philippe et al., 2013; Antwerpen et al., 2015). These *Megavirales* members were shown to share a putative ancient common ancestor that was inferred to harbor about 50 core genes (Koonin and Yutin, 2010; Yutin and Koonin, 2012). Still more recently, other researchers described another giant virus, *Pithovirus sibericum*, which also infects *Acanthamoeba* and was isolated in Siberia from a >30,000-year-old frozen soil sample (Legendre et al., 2014). The *P. sibericum* virion has a very similar morphology and size compared to pandoraviruses but its genome was unexpectedly about 3–4 times shorter than those of pandoraviruses, and comparative genomics showed low similarity between these new clades of giant viruses. In addition, only 11 and 32% of the *P. sibericum* gene complement were found to have counterparts in other viruses and the NCBI GenBank sequence database, respectively. The greatest numbers of best matches with viruses were found to be with representatives from the *Marseilleviridae*, then *Mimiviridae* and *Iridoviridae* families. It is noteworthy that only a very weak homology with a major capsid protein from an unclassified megalocytivirus of the family *Iridoviridae* was found in the *P. sibericum* gene content.

Cellular organisms were divided in the 1970s by C. Woese into three domains (*Bacteria*, *Archaea*, and *Eukarya*) comprising a universal tree of life on the basis of ribosomal genes (Woese et al., 1990). This way, viruses were neglected and excluded from this tree of life as devoid of ribosomes (Raoult, 2013, 2014). After the giant virus discovery, this classification was challenged because a few conserved genes from these viruses appeared to share a common ancestor with homologs from cellular organisms. Thus, starting from the initial description of the Mimivirus genome, several phylogeny reconstructions and phyletic analyses based on such genes showed that *Megavirales* comprised a fourth major monophyletic group alongside *Bacteria*, *Archaea*, and *Eukarya* in a revised universal tree of life (Boyer et al., 2010; Raoult, 2014; Sharma et al., 2014, 2015). The four branches topology was criticized by other groups that claim it was biased by long branch attraction or lateral transfer of genes to giant viruses from their hosts or other sources (Filee et al., 2008; Moreira and Brochier-Armanet, 2008; Williams et al., 2011; Yutin et al., 2014). However, alternative trees failed to show a monophyly for *Eukarya* (Williams et al., 2011). In addition,

although giant viruses have mosaic genomes that harbor genes from multiple origins and with a complex evolutionary history (Filee et al., 2008; Boyer et al., 2009; Yutin and Koonin, 2012), there are a few genes not prone to transfer (Sharma et al., 2014). Using two of them, which are universal and encode DNA-dependent RNA polymerase subunits, recent phylogenetic analyses with a comprehensive sequence set further supported the four branches of life hypothesis (Sharma et al., 2014). In addition, other research groups strengthened this hypothesis using different methodological approaches or datasets (Wu et al., 2011; Nasir et al., 2012). Eventually, the term “TRUC” (an acronym for Things Resisting Uncompleted Classifications) was introduced for a new classification of life that includes a fourth TRUC, consisting of *Megavirales*, standing out from the ribosome-based three domain classification (Raoult, 2013, 2014), and we recently described pandoraviruses as new members of a “Fourth TRUC” club (Sharma et al., 2015). Here, we tested the membership of the new and highly divergent amoebal giant virus *P. sibericum* to this “Fourth TRUC” club.

## Materials and Methods

### Orthologous Sequences from Viruses

Analysis was performed using a strategy similar to the one we have used in previous works (Boyer et al., 2010; Sharma et al., 2014) to update the fourth TRUC hypothesis. The genes used in the present study were identified from clusters of orthologous groups of proteins (COGs) involved in nucleotide transport and metabolism and information storage and processing (i.e., categories F, J, A, K, L, and B). These genes included five genes conserved among previously identified *Megavirales* representatives and that encode family B DNA polymerase (DNAPol), DNA-dependent RNA polymerase subunits 1 (RNAP1) and 2 (RNAP2), transcription factor II B (TFIIB), and ATP-dependent DNA ligase (DNA ligase). All these genes have an important function in replication and transcription processes (Yutin and Koonin, 2009; Boyer et al., 2010). DNAPol plays a critical role in replication, thus transferring genetic information from one generation to another; it is one of the most widely used ancient gene markers for taxonomic mapping across a wide range of organisms from *Archaea*, *Eukarya*, and *Megavirales*, but was found less conserved among *Bacteria* (Filee et al., 2002). Genes encoding RNAP subunits are considered as an appropriate alternative to ribosomal genes because they are conserved in *Bacteria*, *Archaea*, *Eukarya*, and *Megavirales*, and poorly prone to horizontal gene transfer and recombination (Case et al., 2007; Adekambi et al., 2009; Sharma et al., 2014). TFIIB is a transcriptional regulator that plays an important role in transcription and was found to be conserved in a few *Megavirales* representatives, among which *P. sibericum* is found, as well as in a wide range of cellular organisms among *Eukarya* and *Archaea*. Finally, ATP-dependent DNA ligase is involved in the process of replication, repair, and recombination, and is widely conserved in *Bacteria*, *Eukarya*, phages, and some *Megavirales* representatives (including phycodnaviruses, chordopoxviruses, asfarviruses, marseilleviruses, and *P. sibericum*) (Yutin and

Koonin, 2009). Viral orthologs for these four genes were extracted using the OrthoMCL program (Li et al., 2003) from the gene complements of a set of 317 viral genomes harboring more than 100 genes, which were directly downloaded from the NCBI sequence databases (<ftp://ftp.ncbi.nih.gov/genomes/Viruses/>) and were completed with the gene complement previously determined from the *P. sibiricum* (GenBank Accession no. NC\_023423.1).

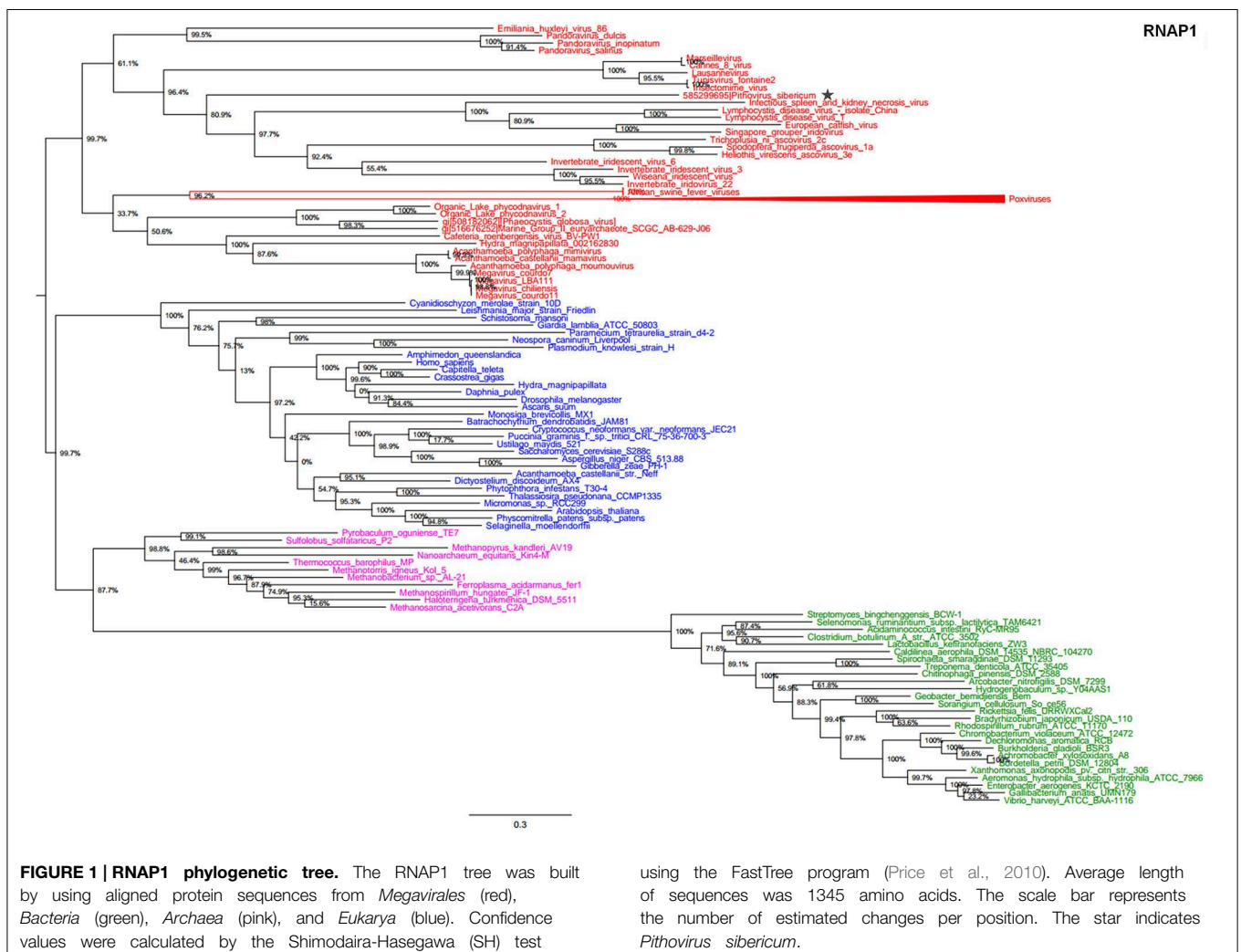
## Orthologous Sequences from the Cellular Life Forms

Stand-alone BLAST 2.2.27 searches were performed to retrieve homologs for informational genes from the members of cellular domains of life, using viral sequences as a query against the NCBI GenBank non-redundant (nr) protein sequence database; the maximum number of target sequences was 20,000 (Altschul et al., 1990). Because informational genes are the most conserved genes among all life forms, they comprise large amounts of homologous sequences in public databases. To obtain an informative equilibrated phylogenetic tree, we selected homologous sequences using TimeTree, a professional

informational data bank where the divergence times of species have been reported based on studies of molecular clocks from peer-reviewed journals (Hedges et al., 2006). Here, we have selected representative species that have been diverging since around 500 million years ago to constitute a set of representatives from *Bacteria*, *Archaea*, and *Eukarya* (Sharma et al., 2014). A Perl script was used to filter out from the BLASTp results, by Taxon identifiers, sequences from these selected representatives. Then, selected protein sequences were directly downloaded from the NCBI nr database using the GenBank identifiers. Identical sequences were removed by clustering using the CD-HIT suite, as previously described (Sharma et al., 2014, 2015).

## Multiple Sequence Alignments and Phylogeny Reconstructions

Sequences were aligned by the Muscle program (Edgar, 2004). Alignment quality was analyzed visually and manually curated, and phylogeny reconstructions were performed with the maximum likelihood method and the WAG model; confidence values were calculated by the Shimodaira-Hasegawa (SH) test using FastTree (Price et al., 2010). The FigTree software was used



for the visualization of phylogenetic trees (<http://tree.bio.ed.ac.uk/software/figtree/>).

### Phyletic Pattern Analysis with COGs

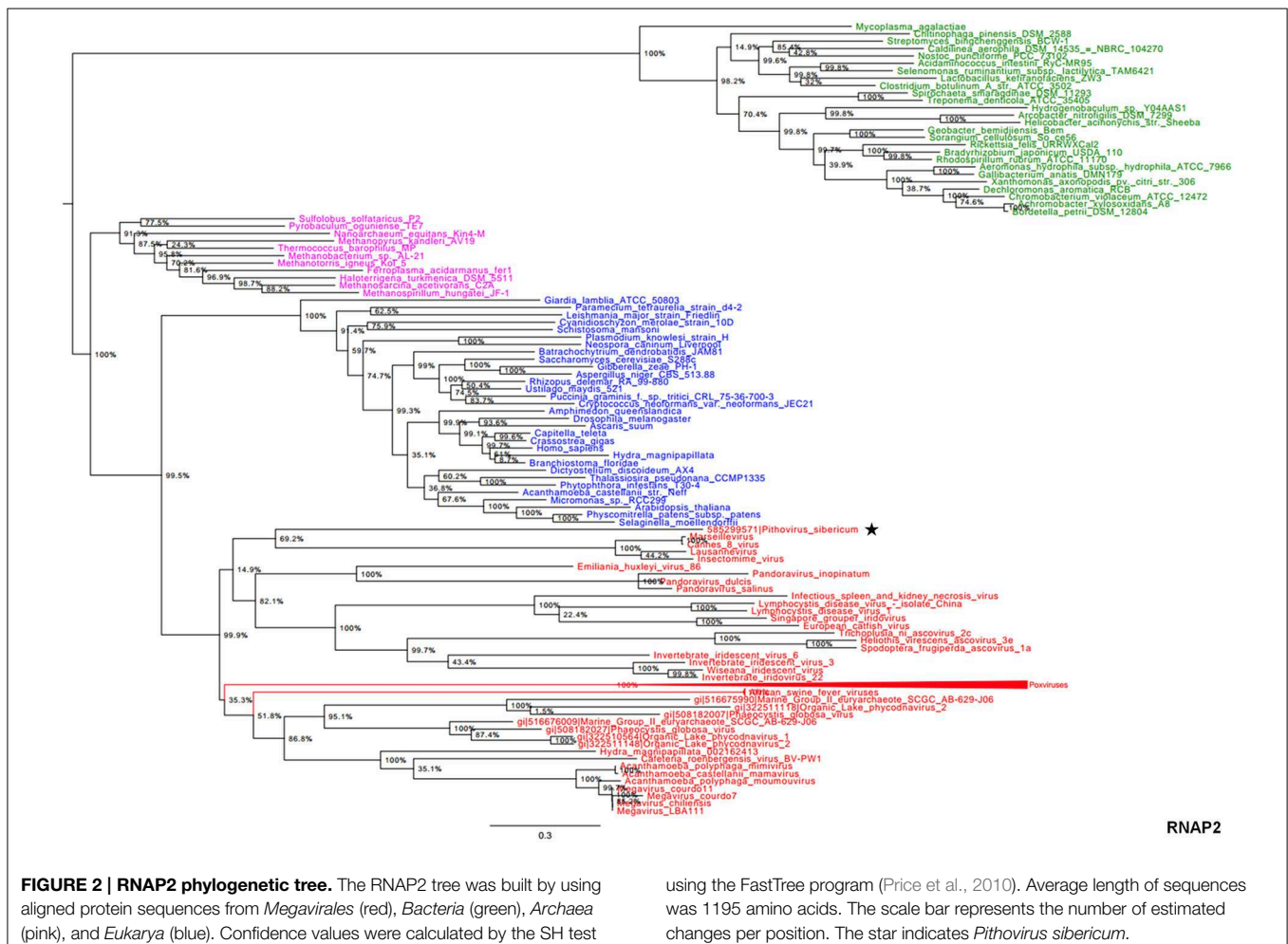
DNA processing and nucleotide metabolism genes are highly conserved in all cellular organisms and in some viruses (Wolf et al., 2002; Boyer et al., 2010). In addition, clusters of orthologous groups of proteins (COGs) have been documented as an ideal set to study functional annotations (Tatusov et al., 2003). Therefore, a set of 727 COGs involved in information storage, processing, nucleotide transport, and metabolism (categories J, A, K, L, B, and F) have been used for the current analysis. Stand-alone BLASTp was performed for the corresponding 727 COGs against selected representatives from *Bacteria*, *Archaea*, *Eukarya*, and *Megavirales*, with stringent parameters ( $e < 1e-3$ , query coverage >70%, and identity >30%) (Boyer et al., 2010). Then, a binary matrix was constructed using BLASTp results based on patterns of presence (1) and absence (0). Finally, a dendrogram was constructed by hierarchical clustering amended with the Pearson distance method using the TM4 multi-package software (Saeed et al., 2003).

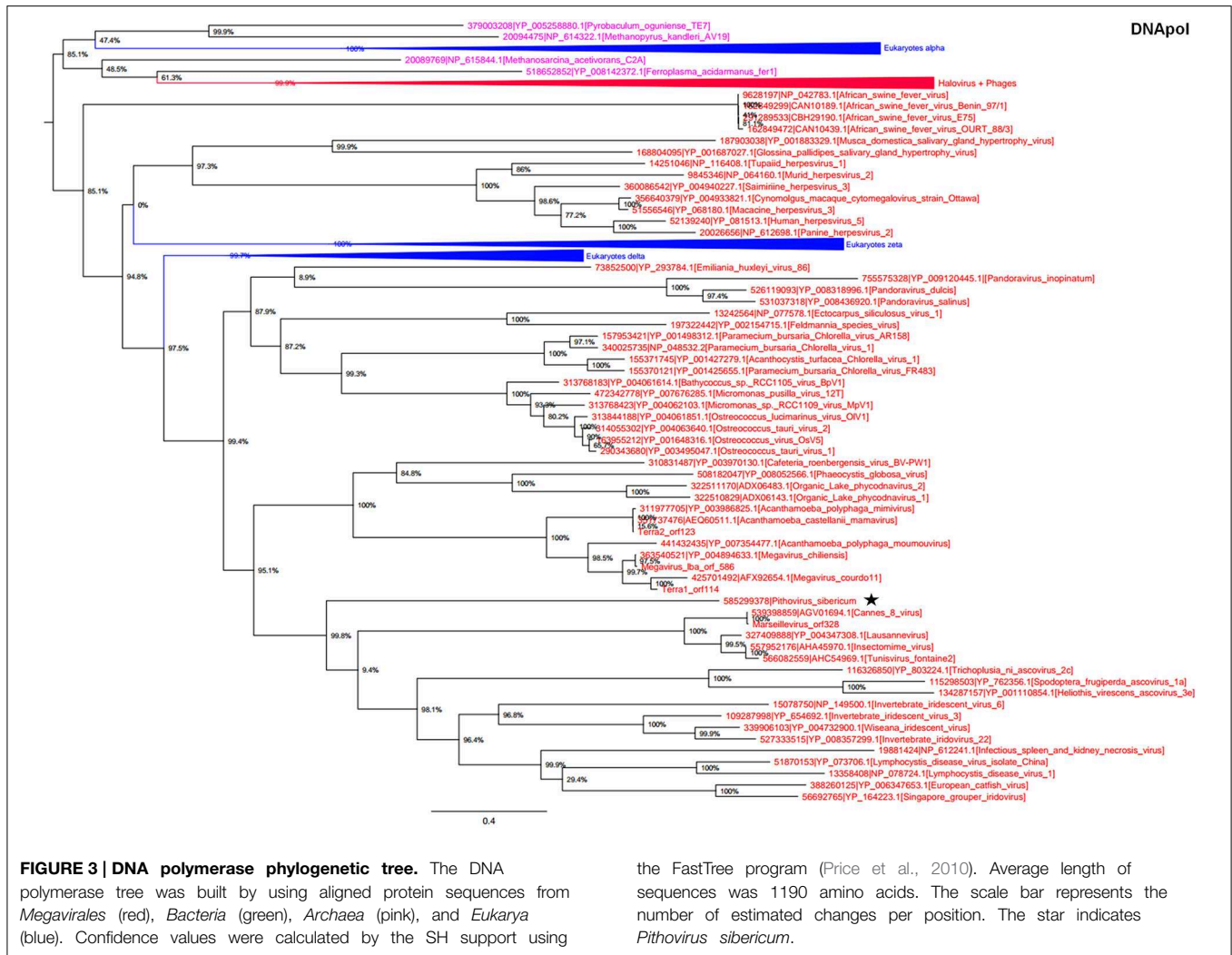
## Results

### Phylogenetic Analyses

In the present study, we reconstructed phylogenies using five informational genes. *P. sibericum* position in both RNAP1 and RNAP2 trees indicated that this species belongs to a new family of *Megavirales*, and was most closely related to families *Iridoviridae*/*Ascoviridae* and *Marseilleviridae* (Figures 1, 2). In the RNAP1 tree, *P. sibericum* was clustered with iridoviruses and ascoviruses, whereas marseilleviruses appeared as an outgroup. In the RNAP2 tree, *P. sibericum* was confidently clustered with marseilleviruses. In addition, these phylogenetic trees constructed using both RNAP1 and 2 sequences from representatives of *Bacteria*, *Archaea*, *Eukarya*, and *Megavirales*, including pandoraviruses and *P. sibericum*, clearly delineated four branches.

In the phylogeny reconstruction based on DNAPol that is present in archaea, eukaryotes, and megaviruses, *P. sibericum* branched deeply in a cluster that also included marseilleviruses, iridoviruses, and ascoviruses (Figure 3). In addition, all these viruses were clustered together with other *Megavirales* members, apart from asfarviruses.





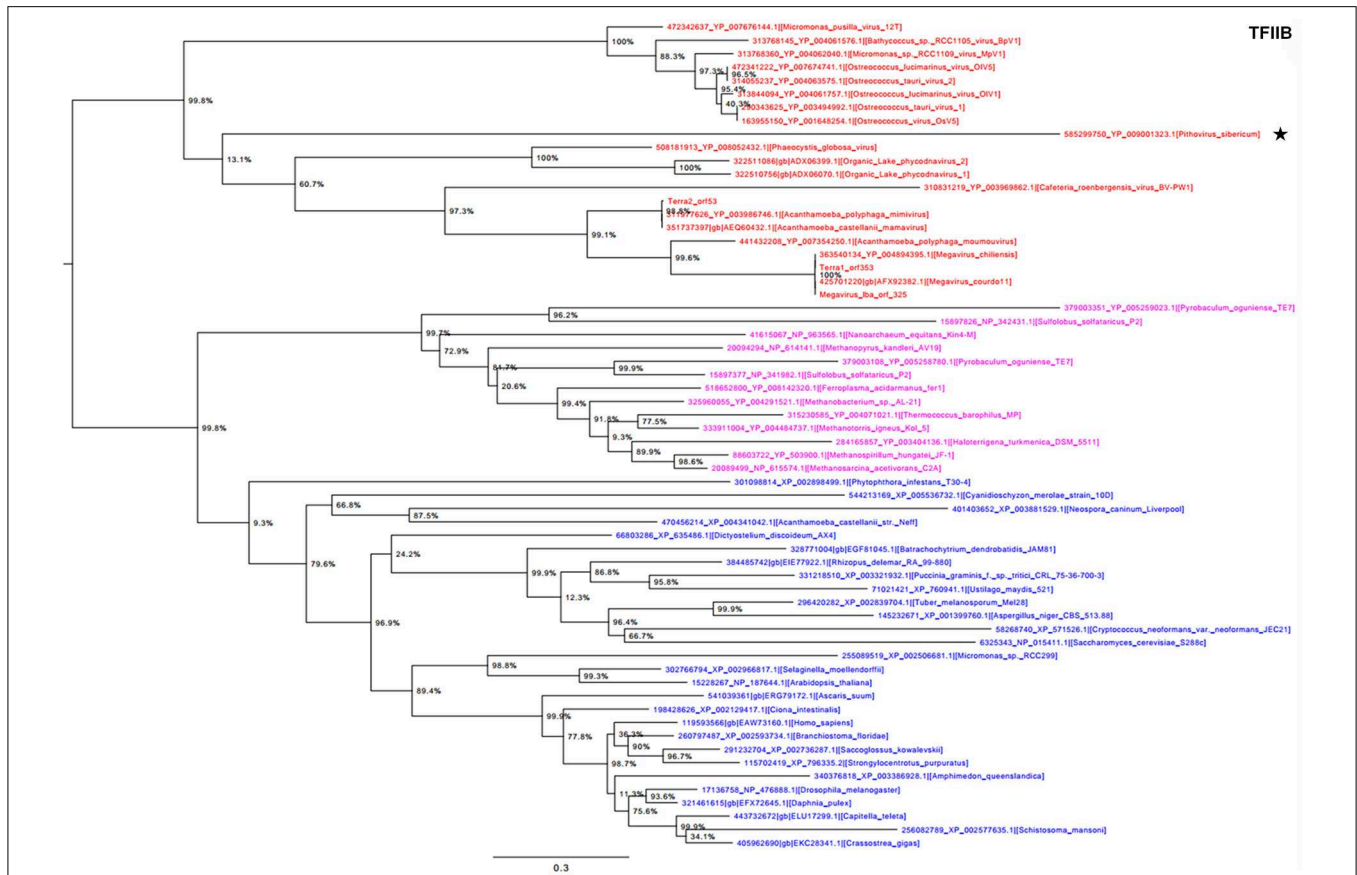
Phylogeny reconstructed using TFIIB showed a strongly supported monophyletic group composed of the *Megavirales* representatives (Figure 4). *P. sibericum* was part of this monophyletic viral clade. Nevertheless, *P. sibericum* was not clustered with any other viral families or putative families. Finally, phylogenetic analysis performed based on the ATP-dependent DNA ligase, which is not conserved in the *Iridoviridae*, *Ascoviridae*, and *Mimiviridae* families apart from some distant representatives, showed a complex evolutionary history. In this phylogenetic reconstruction, megaviruses appeared as a paraphyletic clade scattered in two groups (Supplementary Figure S1). *P. sibericum* was clustered deeply with the *Marseilleviridae* and *Asfarviridae* families, which supports its evolutionary relationship with *Megavirales*; bacteriophages formed a sister group of these viruses.

**Phyletic Analysis**

Hierarchical clustering analysis was performed based on a binary presence/absence matrix constructed using 727 informational

COGs that were known to be conserved in all cellular organisms among *Bacteria*, *Archaea*, and *Eukarya*, and some *Megavirales* members.

The corresponding binary matrix was composed of 142 representatives from *Bacteria*, *Archaea*, *Eukarya*, and *Megavirales*. The *P. sibericum* genome was found to encode homologs to a total of 19 COGs. Among them, the largest numbers of COGs were shared with representatives from the *Marseilleviridae* and *Mimiviridae* families (13 and 10, respectively). In addition, five COGs were shared with another *Megavirales* member, and only one of the 19 COGs present in *P. sibericum* was absent from any other *Megavirales* representatives. A total of 74 COGs were absent from *P. sibericum* while present in other *Megavirales* representatives, eight being present in Marseillevirus, and 23 being present in Mimivirus. Finally, this phyletic analysis based on a set of informational genes also showed a topology of four branches with *Megavirales* being a distinct and separate branch alongside *Bacteria*, *Archaea*, and *Eukarya*, and supports *P. sibericum*'s belonging to the fourth TRUC (Figure 5).



**FIGURE 4 | Transcription factor II B (TFIIB) phylogenetic tree.**  
 The TFIIB tree was built by using aligned protein sequences from *Megavirales* (red), *Bacteria* (green), *Archaea* (pink), and *Eukarya* (blue). Confidence values were calculated by the SH support using

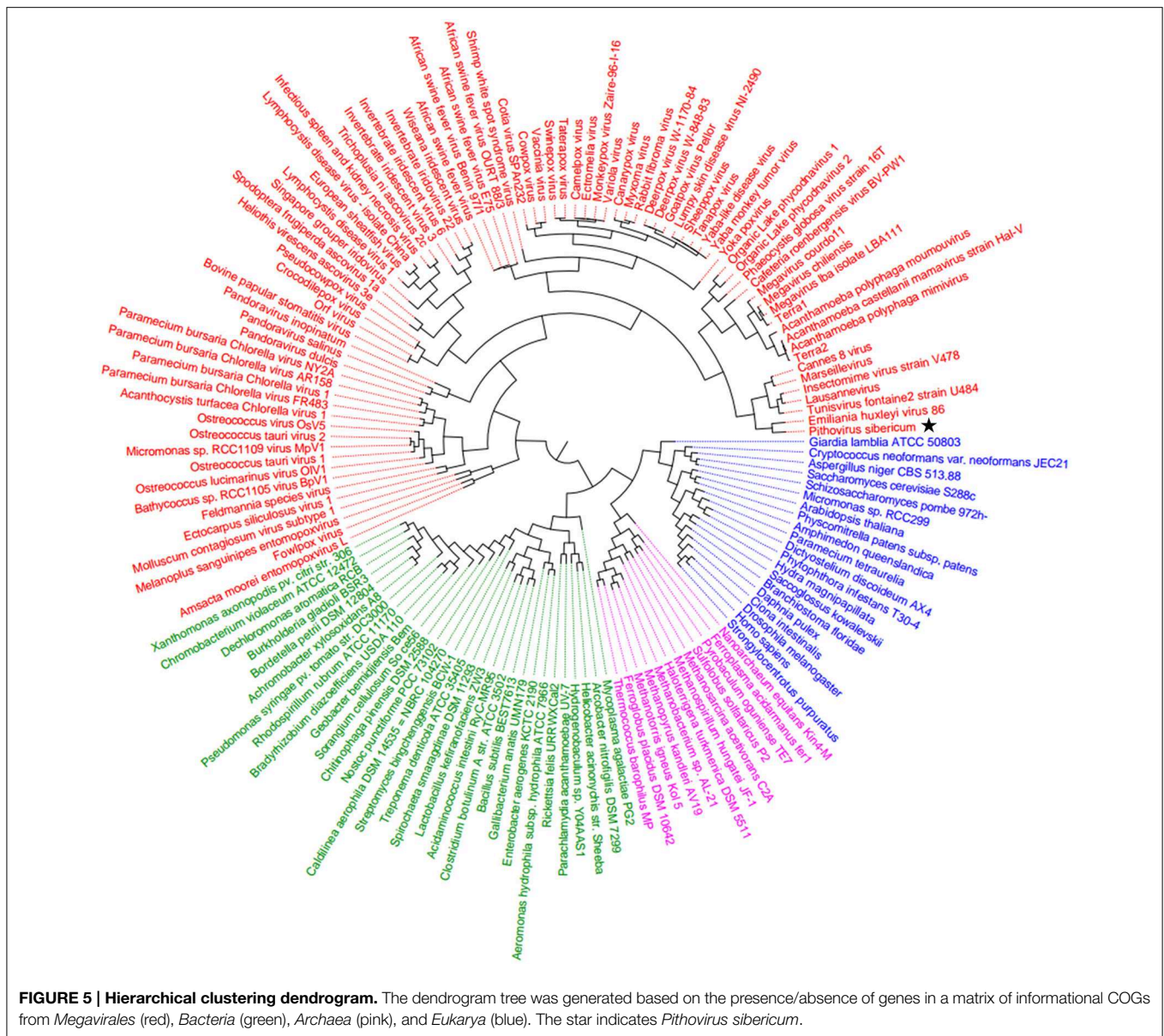
the FastTree program (Price et al., 2010). Average length of sequences was 348 amino acids. The scale bar represents the number of estimated changes per position. The star indicates *Pithovirus sibericum*.

## Discussion

The present phylogenetic and phyletic analyses including *P. sibericum* suggests that this giant virus is linked to *Megavirales*, which comprises a monophyletic group alongside *Eukarya*, *Bacteria*, and *Archaea*. Thus, phylogenies constructed based on three ancient genes (RNAP1/2, DNA polymerase, and TFIIB) strongly support *Megavirales* as an independent monophyletic group, as previously reported (Boyer et al., 2010; Sharma et al., 2014, 2015). Moreover, these analyses suggest that *P. sibericum* is the founder of a new viral family most closely related to the *Marseilleviridae*, *Iridoviridae*, and *Ascoviridae* families. *P. sibericum* position in both RNAP1 and RNAP2 trees indicated that this species belongs to a new family of *Megavirales*, and was most closely related to families *Iridoviridae/Ascoviridae* and *Marseilleviridae*, in congruence with previous analyses (Legendre et al., 2014; Yutin et al., 2014). Phylogeny reconstruction based on DNAPol showed that *P. sibericum* branched deeply in a cluster that also included marseilleviruses, iridoviruses and ascoviruses, in congruence with a previous report (Legendre et al.,

2014). Monophyly observed here for *Megavirales* members based on DNAPol gene was not found in a previous study (Yutin and Koonin, 2012), whereas the one found based on the TFIIB gene was already reported (Yutin and Koonin, 2012). Otherwise, in accordance with the results from our phylogenetic analyses, hierarchical clustering performed here using a set of informational COGs showed that *P. sibericum* is a member of the order *Megavirales*, which comprises, as previously described, an independent branch, a fourth TRUC alongside cellular branches of life (Boyer et al., 2010; Raoult, 2013).

It was described initially based on comparative genomics and cladistic analyses that four viral families comprised a monophyletic group and shared an ancient common ancestor (Iyer et al., 2001; Koonin and Yutin, 2010). Later on, mimiviruses and additional new giant viruses have grown this monophyletic group and founded new families or putative families, which were proposed to be reclassified in the new order *Megavirales* (Colson et al., 2013; Sharma et al., 2015). The order *Megavirales* is a matter of debate because giant virus genomes comprise numerous genes that have orthologs in cellular organisms but a



limited set of viral core genes (Yutin et al., 2014). Phylogenomic analyses using multiple core genes revealed complex evolutionary histories, and it was suggested that *Megavirales* members were gene robbers, and had acquired multiple genes from other megaviruses or from cellular organisms (Moreira and Brochier-Armanet, 2008; Williams et al., 2011; Yutin and Koonin, 2012). Overall, using different components of such chimeric gene repertoires as those of *Megavirales* representatives leads to different scenario based on the genes used. It has been put forward that part of the *Megavirales* genes have a cellular origin, but notwithstanding, a viral origin has been inferred for other genes in the genomes of these viruses, which have no detectable homologs in cellular life forms. Ancient viral genes with cellular counterparts also exist, and these latter show *Megavirales* as gathered biological entities apart from *Bacteria*, *Archaea*, and

*Eukarya* (Boyer et al., 2010; Colson et al., 2011; Sharma et al., 2014, 2015).

Some previous studies have stated that *Megavirales* does not comprise a separate clade and should not be included in the tree of life, and are opposed to the fourth TRUC hypothesis (Moreira and Brochier-Armanet, 2008; Williams et al., 2011; Yutin et al., 2014). Notably, it was assumed that even ancient genes such as RNAP have a polyphyletic origin (Yutin et al., 2014). Thus, a polyphyletic origin of RNAP1/2 from *Megavirales* representatives was previously described, but in these studies the three eukaryotic paralogous genes were included whereas here we used RNA polymerase III, because it was found to be the most conserved of these three eukaryotic paralogs among *Megavirales* members and cellular living forms (Yutin and Koonin, 2012; Sharma et al., 2014, 2015; Yutin et al., 2014). In addition, RNAP1/2 genes from



*Phytophthora parasitica*, a eukaryotic plant pathogen from the *Oomycetes* class, were found to be clustered with *Mimiviridae* (Yutin et al., 2014), but *Phytophthora parasitica* sequences were previously described to have *Megavirales* member sequences as best hits, and might represent overlooked megaviruses (Sharma et al., 2014). Misannotations exist in public databases, and contigs composed of viral genes have been detected within eukaryotic genomes (Filee, 2014; Maumus et al., 2014; Sharma et al., 2014). This might be explained by contamination with viral DNA, the sequencing of giant virus DNA together with that of a eukaryotic host, or the presence of giant viral genes in eukaryotic DNA. In any case, phylogenetic analyses using RNAP genes allowed us to resolve the phylogenetic and taxonomic status of megaviruses (Sharma et al., 2014). Certainly, giant viruses are chimeric organisms that harbor genes with multiple origins and a complex evolutionary history (Filee et al., 2008; Yutin and Koonin, 2012). However, there are still a few genes that are not prone to transfer, including those encoding DNA-dependent RNA polymerase subunits, or other informational genes, which enabled a better understanding of the origin and evolution of the giant viruses (Sharma et al., 2014). We believe that the *Megavirales* ancestor would have harbored these ancient stigma genes that exhibit few or no gene transfers. Then, *Megavirales* representatives might have lost some of these genes and acquired new ones from their respective hosts or from various organisms. Interestingly, previous analyses indicated that *P. sibericum* dramatically diverged from other viruses, similarly to pandoraviruses. Thus, *P. sibericum* shares only a small fraction

of its genes with other known members of the *Megavirales* as only one third of its gene repertoire matches known viral genomes, and notably this repertoire lacks many of the megaviral core genes (Legendre et al., 2014). Nonetheless, the small core gene repertoire of this giant virus contains stigma genes that are sufficient to link it to the *Megavirales*.

Taken together, our phylogenetic and phyletic analyses indicate that *P. sibericum* is a new genuine member of the “Fourth TRUC” club, and appears to be closely related to the *Marseilleviridae* and *Iridoviridae/Ascoviridae* families.

## Acknowledgments

VS was financially supported through a grant from the Infectiopol Sud foundation.

## Supplementary Material

The Supplementary Material for this article can be found online at: <http://journal.frontiersin.org/article/10.3389/fmicb.2015.00722>

### Supplementary Figure S1 | ATP-dependent DNA ligase phylogenetic tree.

The DNA ligase tree was built by using aligned protein sequences from *Megavirales* (red), *Bacteria* (green), *Archaea* (pink), and *Eukarya* (blue). Confidence values were calculated by the SH support using the FastTree program (Price et al., 2010). The scale bar represents the number of estimated changes per position. The star indicates *Pithovirus sibericum*.

## References

- Adekambi, T., Drancourt, M., and Raoult, D. (2009). The *rpoB* gene as a tool for clinical microbiologists. *Trends Microbiol.* 17, 37–45. doi: 10.1016/j.tim.2008.09.008
- Aherfi, S., Boughalmi, M., Pagnier, I., Fournous, G., La Scola, B., Raoult, D., et al. (2014). Complete genome sequence of tunisivirus, a new member of the proposed family marseilleviridae. *Arch. Virol.* 159, 2349–2358. doi: 10.1007/s00705-014-2023-5
- Altschul, S. F., Gish, W., Miller, W., Myers, E. W., and Lipman, D. J. (1990). Basic local alignment search tool. *J. Mol. Biol.* 215, 403–410. doi: 10.1016/S0022-2836(05)80360-2
- Antwerpen, M. H., Georgi, E., Zoeller, L., Woelfel, R., Stoecker, K., and Scheid, P. (2015). Whole-genome sequencing of a pandoravirus isolated from keratitis-inducing acanthamoeba. *Genome Announc.* 3, E00136–E00115. doi: 10.1128/genomeA.00136-15
- Arslan, D., Legendre, M., Seltzer, V., Abergel, C., and Claverie, J. M. (2011). Distant mimivirus relative with a larger genome highlights the fundamental features of megaviridae. *Proc. Natl. Acad. Sci. U.S.A.* 108, 17486–17491. doi: 10.1073/pnas.1110889108
- Boyer, M., Madoui, M. A., Gimenez, G., La Scola, B., and Raoult, D. (2010). Phylogenetic and phyletic studies of informational genes in genomes highlight existence of a 4 domain of life including giant viruses. *PLoS ONE* 5:e15530. doi: 10.1371/journal.pone.0015530
- Boyer, M., Yutin, N., Pagnier, I., Barrassi, L., Fournous, G., Espinosa, L., et al. (2009). Giant marseillevirus highlights the role of amoebae as a melting pot in emergence of chimeric microorganisms. *Proc. Natl. Acad. Sci. U.S.A.* 106, 21848–21853. doi: 10.1073/pnas.0911354106
- Case, R. J., Boucher, Y., Dahlfors, I., Holmstrom, C., Doolittle, W. F., and Kjelleberg, S. (2007). Use of 16S rRNA and *rpoB* genes as molecular markers for microbial ecology studies. *Appl. Environ. Microbiol.* 73, 278–288. doi: 10.1128/AEM.01177-06
- Colson, P., De Lamballerie, X., Yutin, N., Asgari, S., Bigot, Y., Bideshi, D. K. et al. (2013). “Megavirales,” a proposed new order for eukaryotic nucleocytoplasmic large DNA viruses. *Arch. Virol.* 158, 2517–2521. doi: 10.1007/s00705-013-1768-6
- Colson, P., Gimenez, G., Boyer, M., Fournous, G., and Raoult, D. (2011). The giant cafeteria roenbergensis virus that infects a widespread marine phagocytic protist is a new member of the fourth domain of life. *PLoS ONE* 6:e18935. doi: 10.1371/journal.pone.0018935
- Edgar, R. C. (2004). Muscle: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res.* 32, 1792–1797. doi: 10.1093/nar/gkh340
- Filee, J. (2014). Multiple occurrences of giant virus core genes acquired by eukaryotic genomes: the visible part of The Iceberg? *Virology* 466–467, 53–59. doi: 10.1016/j.virol.2014.06.004
- Filee, J., Forterre, P., Sen-Lin, T., and Laurent, J. (2002). Evolution of DNA polymerase families: evidences for multiple gene exchange between cellular and viral proteins. *J. Mol. Evol.* 54, 763–773. doi: 10.1007/s00239-001-0078-x
- Filee, J., Pouget, N., and Chandler, M. (2008). Phylogenetic evidence for extensive lateral acquisition of cellular genes by nucleocytoplasmic large DNA viruses. *BMC Evol. Biol.* 8:320. doi: 10.1186/1471-2148-8-320
- Fischer, M. G., Allen, M. J., Wilson, W. H., and Suttle, C. A. (2010). Giant virus with a remarkable complement of genes infects marine zooplankton. *Proc. Natl. Acad. Sci. U.S.A.* 107, 19508–19513. doi: 10.1073/pnas.1007615107
- Hedges, S. B., Dudley, J., and Kumar, S. (2006). Timetree: a public knowledge-base of divergence times among organisms. *Bioinformatics* 22, 2971–2972. doi: 10.1093/bioinformatics/btl505
- Iyer, L. M., Aravind, L., and Koonin, E. V. (2001). Common origin of four diverse families of large eukaryotic DNA viruses. *J. Virol.* 75, 11720–11734. doi: 10.1128/JVI.75.23.11720-11734.2001

- Iyer, L. M., Balaji, S., Koonin, E. V., and Aravind, L. (2006). Evolutionary genomics of nucleocytoplasmic large DNA viruses. *Virus Res.* 117, 156–184. doi: 10.1016/j.virusres.2006.01.009
- Koonin, E. V., and Yutin, N. (2010). Origin and evolution of eukaryotic large nucleocytoplasmic DNA viruses. *Intervirology* 53, 284–292. doi: 10.1159/000312913
- La Scola, B., Audic, S., Robert, C., Jungang, L., De Lamballerie, X., Drancourt, M., et al. (2003). A giant virus in amoebae. *Science* 299, 2033. doi: 10.1126/science.1081867
- Legendre, M., Bartoli, J., Shmakova, L., Jeudy, S., Labadie, K., Adrait, A., et al. (2014). Thirty-thousand-year-old distant relative of giant icosahedral DNA viruses with a pandoravirus morphology. *Proc. Natl. Acad. Sci. U.S.A.* 111, 4274–4279. doi: 10.1073/pnas.1320670111
- Li, L., Stoeckert, C. J. Jr., and Roos, D. S. (2003). Orthomcl: identification of ortholog groups for eukaryotic genomes. *Genome Res.* 13, 2178–2189. doi: 10.1101/gr.1224503
- Maumus, F., Epert, A., Nogue, F., and Blanc, G. (2014). Plant genomes enclose footprints of past infections by giant virus relatives. *Nat. Commun.* 5:4268. doi: 10.1038/ncomms5268
- Moreira, D., and Brochier-Armanet, C. (2008). Giant viruses, giant chimeras: the multiple evolutionary histories of mimivirus genes. *BMC Evol. Biol.* 8:12. doi: 10.1186/1471-2148-8-12
- Nasir, A., Kim, K. M., and Caetano-Anolles, G. (2012). Giant viruses coexisted with the cellular ancestors and represent a distinct supergroup along with superkingdoms archaea, bacteria and eukarya. *BMC Evol. Biol.* 12:156. doi: 10.1186/1471-2148-12-156
- Philippe, N., Legendre, M., Doutre, G., Coute, Y., Poirot, O., Lescot, M., et al. (2013). Pandoraviruses: amoeba viruses with genomes up to 2.5 Mb reaching that of parasitic eukaryotes. *Science* 341, 281–286. doi: 10.1126/science.1239181
- Price, M. N., Dehal, P. S., and Arkin, A. P. (2010). FastTree 2—approximately maximum-likelihood trees for large alignments. *PLoS ONE* 5:e9490. doi: 10.1371/journal.pone.0009490
- Raoult, D. (2013). Truc or the need for a new microbial classification. *Intervirology* 56, 349–353. doi: 10.1159/000354269
- Raoult, D. (2014). How the virophage compels the need to readdress the classification of microbes. *Virology* 477, 119–124. doi: 10.1016/j.virol.2014.11.014
- Raoult, D., Audic, S., Robert, C., Abergel, C., Renesto, P., Ogata, H., et al. (2004). The 1.2-megabase genome sequence of mimivirus. *Science* 306, 1344–1350. doi: 10.1126/science.1101485
- Raoult, D., and Forterre, P. (2008). Redefining viruses: lessons from mimivirus. *Nat. Rev. Microbiol.* 6, 315–319. doi: 10.1038/nrmicro1858
- Raoult, D., La Scola, B., and Birtles, R. (2007). The discovery and characterization of mimivirus, the largest known virus and putative pneumonia agent. *Clin. Infect. Dis.* 45, 95–102. doi: 10.1086/518608
- Saeed, A. I., Sharov, V., White, J., Li, J., Liang, W., Bhagabati, N., et al. (2003). Tm4: a free, open-source system for microarray data management and analysis. *Biotechniques* 34, 374–378.
- Sharma, V., Colson, P., Chabrol, O., Scheid, P., Pontarotti, P., and Raoult, D. (2015). Welcome to pandoraviruses at the “fourth truc” club. *Front. Microbiol.* 6:423. doi: 10.3389/fmicb.2015.00423
- Sharma, V., Colson, P., Giorgi, R., Pontarotti, P., and Raoult, D. (2014). DNA-dependent RNA polymerase detects hidden giant viruses in published databanks. *Genome Biol. Evol.* 6, 1603–1610. doi: 10.1093/gbe/evu128
- Tatusov, R. L., Fedorova, N. D., Jackson, J. D., Jacobs, A. R., Kiryutin, B., Koonin, E. V., et al. (2003). The cog database: an updated version includes eukaryotes. *BMC Bioinformatics* 4:41. doi: 10.1186/1471-2105-4-41
- Williams, T. A., Embley, T. M., and Heinz, E. (2011). Informational gene phylogenies do not support a fourth domain of life for nucleocytoplasmic large DNA viruses. *PLoS ONE* 6:e21080. doi: 10.1371/journal.pone.0021080
- Woese, C. R., Kandler, O., and Wheelis, M. L. (1990). Towards a natural system of organisms: proposal for the domains archaea, bacteria, and eucarya. *Proc. Natl. Acad. Sci. U.S.A.* 87, 4576–4579. doi: 10.1073/pnas.87.12.4576
- Wolf, Y. I., Rogozin, I. B., Grishin, N. V., and Koonin, E. V. (2002). Genome trees and the tree of life. *Trends Genet.* 18, 472–479. doi: 10.1016/S0168-9525(02)02744-0
- Wu, D., Wu, M., Halpern, A., Rusch, D. B., Yoosuf, S., Frazier, M., et al. (2011). Stalking the fourth domain in metagenomic data: searching for, discovering, and interpreting novel, deep branches in marker gene phylogenetic trees. *PLoS ONE* 6:e18011. doi: 10.1371/journal.pone.0018011
- Yoosuf, N., Yutin, N., Colson, P., Shabalina, S. A., Pagnier, I., Robert, C., et al. (2012). Related giant viruses in distant locations and different habitats: acanthamoeba polyphaga mousmouvirus represents a third lineage of the mimiviridae that is close to the megavirus lineage. *Genome Biol. Evol.* 4, 1324–1330. doi: 10.1093/gbe/evs109
- Yutin, N., and Koonin, E. V. (2009). Evolution of DNA ligases of nucleocytoplasmic large DNA viruses of eukaryotes: a case of hidden complexity. *Biol. Direct* 4:51. doi: 10.1186/1745-6150-4-51
- Yutin, N., and Koonin, E. V. (2012). Hidden evolutionary complexity of nucleocytoplasmic large DNA viruses of eukaryotes. *Viol. J.* 9:161. doi: 10.1186/1743-422X-9-161
- Yutin, N., Wolf, Y. I., and Koonin, E. V. (2014). Origin of giant viruses from smaller DNA viruses not from a fourth domain of cellular life. *Virology* 466–467, 38–52. doi: 10.1016/j.virol.2014.06.032
- Yutin, N., Wolf, Y. I., Raoult, D., and Koonin, E. V. (2009). Eukaryotic large nucleocytoplasmic DNA viruses: clusters of orthologous genes and reconstruction of viral genome evolution. *Viol. J.* 17:223. doi: 10.1186/1743-422X-6-223

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2015 Sharma, Colson, Chabrol, Pontarotti and Raoult. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.