

Impact of alley cropping agroforestry on stocks, forms and spatial distribution of soil organic carbon: a case study in a Mediterranean context

Rémi Cardinael, Tiphaine Chevallier, Bernard Barthès, Nicolas Saby, Théophile Parent, Christian Dupraz, Martial Bernoux, Claire Chenu

▶ To cite this version:

Rémi Cardinael, Tiphaine Chevallier, Bernard Barthès, Nicolas Saby, Théophile Parent, et al.. Impact of alley cropping agroforestry on stocks, forms and spatial distribution of soil organic carbon: a case study in a Mediterranean context. Geoderma, 2015, 259–260, pp.288-299. 10.1016/j.geoderma.2015.06.015 . hal-01269102

HAL Id: hal-01269102 https://hal.science/hal-01269102

Submitted on 24 Jan 2020 $\,$

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1	Impact of alley cropping agroforestry on stocks, forms and spatial distribution of soil
2	organic carbon - a case study in a Mediterranean context
3	Rémi Cardinael ^{a,d} , Tiphaine Chevallier ^{a*} , Bernard G. Barthès ^a , Nicolas P.A. Saby ^b , Théophile
4	Parent ^a , Christian Dupraz ^c , Martial Bernoux ^a , Claire Chenu ^d
5	
6	^a IRD, UMR 210 Eco&Sols, Montpellier SupAgro, 34060 Montpellier, France
7	^b INRA, US 1106 Infosol, F 45075, Orléans, France
8	^c INRA, UMR 1230 System, Montpellier SupAgro, 34060 Montpellier, France
9	^d AgroParisTech, UMR 1402 Ecosys, Avenue Lucien Brétignières, 78850 Thiverval-Grignon,
10	France
11	* Corresponding author. Tel.: +33 (0)4.99.61.21.30. E-mail address:
12	tiphaine.chevallier@ird.fr
13	
14	ABSTRACT
15	Agroforestry systems, i.e., agroecosystems combining trees with farming practices, are of
16	particular interest as they combine the potential to increase biomass and soil carbon (C)

storage whilst maintaining an agricultural production. However, most present knowledge on the impact of agroforestry systems on soil organic carbon (SOC) storage comes from tropical systems. This study was conducted in southern France, in an 18-year-old agroforestry plot, where hybrid walnuts (Juglans regia × nigra L.) are intercropped with durum wheat (Triticum turgidum L. subsp. durum), and in an adjacent agricultural control plot, where durum wheat is the sole crop. We quantified SOC stocks to 2.0 m depth and their spatial variability in relation to the distance to the trees and to the tree rows. The distribution of additional SOC storage in different soil particle-size fractions was also characterised. SOC accumulation rates between

the agroforestry and the agricultural plots were 248 ± 31 kg C ha⁻¹ yr⁻¹ for an equivalent soil 25 mass (ESM) of 4000 Mg ha⁻¹ (to 26-29 cm depth) and 350 ± 41 kg C ha⁻¹ yr⁻¹ for an ESM of 26 15700 Mg ha⁻¹ (to 93-98 cm depth). SOC stocks were higher in the tree rows where 27 herbaceous vegetation grew and where the soil was not tilled, but no effect of the distance to 28 the trees (0 to 10 m) on SOC stocks was observed. Most of additional SOC storage was found 29 in coarse organic fractions (50-200 and 200-2000 μ m), which may be rather labile fractions. 30 31 All together our study demonstrated the potential of alley cropping agroforestry systems under Mediterranean conditions to store SOC, and questioned the stability of this storage. 32

33

Keywords: Tree-based intercropping system, Soil mapping, Soil organic carbon storage, Soil
 organic carbon saturation, Deep soil organic carbon stocks, Visible and near infrared
 spectroscopy, Particle-size fractionation

37

38 **1. Introduction**

39 Agroforestry systems are defined as agroecosystems associating trees with farming practices (Somarriba, 1992; Torquebiau, 2000). Several types of agroforestry systems can be 40 distinguished depending on the different associations of trees, crops and animals (Torquebiau, 41 42 2000). In temperate regions, an important part of recently established agroforestry systems are alley cropping systems, where parallel tree rows are planted in crop lands, and designed to 43 44 allow mechanization of annual crops. Agroforestry systems are of particular interest as they combine the potential to provide a variety of non-marketed ecosystem services, defined as the 45 benefits people obtain from ecosystems (Millennium Ecosystem Assessment, 2005; Power, 46 47 2010) whilst maintaining a high agricultural production (Clough et al., 2011). For instance, agroforestry systems can contribute to water quality improvement (Bergeron et al., 2011; 48

Tully et al., 2012), biodiversity enhancement (Schroth et al., 2004; Varah et al., 2013), and 49 50 erosion control (Young, 1997). But agroforestry systems are also increasingly recognized as a useful tool to help mitigate global warming (Pandey, 2002; Stavi and Lal, 2013; Verchot et 51 al., 2007). Trees associated to annual crops store the carbon (C) assimilated through 52 photosynthesis into their aboveground and belowground biomass. The residence time of C in 53 the harvested biomass will depend on the fate of woody products, and can reach many 54 decades especially for timber wood (Bauhus et al., 2010; Profft et al., 2009). Agroforestry 55 trees also produce organic matter (OM) inputs to the soil (Jordan, 2004; Peichl et al., 2006), 56 and could thus enhance soil organic carbon (SOC) stocks. Leaf litter and pruning residues are 57 58 left on the soil, whereas OM originating from root mortality and root exudates can be incorporated much deeper into the soil as agroforestry trees may have a very deep rooting to 59 minimize the competition with the annual crop (Cardinael et al., 2015; Mulia and Dupraz, 60 61 2006). Moreover, several studies showed that root-derived C was preferentially stabilized in soil compared to above ground derived C (Balesdent and Balabane, 1996; Rasse et al., 2005), 62 mainly due to physical protection of root hairs within soil aggregates (Gale et al., 2000), to 63 chemical recalcitrance of root components (Bird and Torn, 2006), or to adsorption of root 64 exudates or decomposition products on clay particles (Chenu and Plante, 2006; Oades, 1995). 65 66 Compared to an agricultural field, additional inputs of C from tree roots could therefore be stored deep into the soil, but could also enhance decomposition of SOM, i.e., due to the 67 priming effect (Fontaine et al., 2007). 68

Although it is generally assumed that agroforestry system have the potential to increase SOC
stocks (Lorenz and Lal, 2014), quantitative estimates are scarce, especially for temperate
(Nair et al., 2010; Peichl et al., 2006; Pellerin et al., 2013; Upson and Burgess, 2013) or
Mediterranean (Howlett et al., 2011) agroforestry systems combining crops and tree rows.

Most studies concern tropical regions where agroforestry is a more widespread agricultural
practice (Albrecht and Kandji, 2003; Somarriba et al., 2013).

Moreover, as pointed out by Nair (2012), very few studies assessed the impact of agroforestry 75 trees deep in the soil (Haile et al., 2010; Howlett et al., 2011; Upson and Burgess, 2013). Most 76 of them considered SOC at depths of less than 0.5 m (Bambrick et al., 2010; Oelbermann and 77 Voroney, 2007; Oelbermann et al., 2004; Peichl et al., 2006; Sharrow and Ismail, 2004). This 78 lack of knowledge concerning deep soil is mainly due to difficulties to attain profound soil 79 depths, and to the cost of analyzing soil samples from several soil layers. Recently, new 80 methods such as visible and near infrared reflectance (VNIR) spectroscopy have been 81 developed (Brown et al., 2006; Stevens et al., 2013). They allow time- and cost-effective 82 determination of SOC concentration, in the laboratory but also in the field (Gras et al., 2014). 83 Additionally to the lack of data for deep soil, reference plots were not always available, 84 preventing from estimating the additional storage of SOC due specifically to agroforestry 85 (Howlett et al., 2011). 86

In alley cropping systems, spaces between trees in tree rows are usually covered by natural or 87 sowed herbaceous vegetation, and the soil under tree rows is usually not tilled, which may 88 89 favor SOC storage in soil (Virto et al., 2011). Moreover, while trees strongly affects the depth and spatial distribution of OM inputs to soils (Rhoades, 1997), distribution of SOC stocks 90 91 close and away from trees was seldom considered. Some authors reported higher SOC stocks 92 under the tree canopy than 5 m from the tree to 1 m soil depth (Howlett et al., 2011), others 93 found that spatial distribution of SOC stocks could vary with the age of the trees (Bambrick et al., 2010). Some authors reported that spatial distribution of SOC stocks to 20 cm depth was 94 95 not explained by the distance to the trees but by the design of the agroforestry system, tree rows having higher SOC stocks than inter-rows whatever the distance to the trees (Peichl et 96 al., 2006; Upson and Burgess, 2013). To our knowledge, geostatistical methods (Webster and 97

98 Oliver, 2007) have never been used to describe the spatial distribution of SOC stocks in alley 99 cropping agroforestry system although they have been recognized to be very powerful to map 100 and understand spatial heterogeneity at the plot scale (Philippot et al., 2009) especially when 101 dealing with more diverse and heterogeneous systems.

In addition, it is not known whether additional SOC (compared to an agricultural field) due to 102 103 the presence of trees and tree rows, corresponds to soil fractions with a rapid turnover, such as particulate organic matter (POM), or to clay and silt associated OM, likely to be stabilized in 104 soil for a longer period of time (Balesdent et al., 1998). Takimoto et al. (2008) and Howlett et 105 al. (2011) found that carbon content of coarse organic fractions was increased at different 106 depths under agroforestry systems. But, Haile et al. (2010) found that trees grown in a 107 108 silvopastoral system contributed to most of the SOC associated to the fine silt + clay fractions 109 to 1 m depth. The potential of a soil for SOC storage in a stable form is limited by the amount of fine particles (clay + fine silt) and can be estimated by the difference between the 110 111 theoretical SOC saturation (Hassink, 1997) and the measured SOC saturation value for the fine fraction (Angers et al., 2011; Wiesmeier et al., 2014). 112

In this study, we aimed to assess the effect of introducing rows of timber trees into arable land on SOC storage. For this i) we quantified SOC stocks to a depth of 2.0 m in an agroforestry plot and in an adjacent agricultural control plot, ii) we assessed the spatial distribution of SOC stocks in a geostatistical framework taking into account the distance to the trees and to the tree rows, iii) we studied the distribution of SOC in different soil particle-size fractions.

We hypothesized that SOC stocks would be higher in the agroforestry plot compared to the control plot, also at depth, and that SOC stocks would decrease with increasing distance to the trees at all depths. Moreover, our hypothesis was that additional SOC in the agroforestry plot compared to the control plot would enrich all particle-size fractions. 122

2. Materials and methods

123 2.1. Site description

The experimental site was located in Prades-le-Lez, 15 km North of Montpellier, 124 France (Longitude 04°01' E, Latitude 43°43' N, elevation 54 m a.s.l.). The climate is sub-125 humid Mediterranean with an average temperature of 14.5°C and an average annual rainfall of 126 127 951 mm (years 1996–2003). The soil is a silty and carbonated deep alluvial Fluvisol (IUSS Working Group WRB, 2007). From 1950 to 1960, the site was a vineyard (Vitis vinifera L.), 128 and from 1960 to 1985 the field was occupied by an apple (Malus Mill.) orchard. The apple 129 tree stumps were removed in 1985. Then, durum wheat (Triticum turgidum L. subsp. durum 130 (Desf.) Husn.) was cultivated. In February 1995, a 4.6 hectare agroforestry alley-cropping 131 plot was established after the soil was ploughed to 20 cm depth, with the planting of hybrid 132 walnuts (Juglans regia \times nigra cv. NG23) at 13 \times 4 m spacing, with East–West tree rows 133 (Fig. 1). The remaining part of the plot (1.4 ha) was kept as a control agricultural plot. The 134 walnut trees were planted at an initial density of 200 trees ha⁻¹. They were thinned in 2004 135 down to 110 trees ha⁻¹. In the tree rows, the soil was not tilled and spontaneous herbaceous 136 vegetation grew. The cultivated inter-row was 11 m wide. Since the tree planting, the 137 agroforestry inter-row and the control plot were managed in the same way. The annual crop 138 was most of the time durum wheat, except in 1998, 2001 and 2006, when rapeseed (Brassica 139 napus L.) was cultivated, and in 2010 and 2013, when pea (Pisum sativum L.) was cultivated. 140 The durum wheat crop was fertilized as a conventional crop (120 kg N ha⁻¹ yr⁻¹), and the soil 141 was ploughed annually to 20 cm depth, before durum wheat was sown. 142

143

144



Figure 1. Hybrid walnut-durum wheat agroforestry system. Left panel: in November 2013;
Right panel: in June 2014.

149

150 2.2. Soil core sampling

The experimental site was not designed as traditional agronomical experiments with blocks 151 and replicates, but with two large adjacent plots. First, soil texture was analyzed for 24 152 profiles down to 2 m soil depth, following a random sampling design within the two plots. In 153 May 2013, a sub-plot of 625 m² was sampled in both plots, following an intensive sampling 154 155 scheme (Fig. 2). In the agroforestry plot, this sub-plot included two tree rows, two inter-rows and nine walnut trees. Walnut trees had a mean height of 11.21 ± 0.65 m, a mean height of 156 merchantable timber of 4.49 ± 0.39 m and a mean diameter at breast height of 25.54 ± 1.36 157 158 cm. Soil cores (n=36) were sampled on a regular grid, every 5 m (Fig. 2). Around each tree, a soil core was collected at 1 m, 2 m and 3 m distance from the tree (n=57), in the tree row and 159 160 perpendicular to the tree row. Seven soil cores were sampled additionally in the middle of the inter-row to study short scale (1 m distance) spatial heterogeneity of SOC stocks far from the 161 trees (Fig. 2). The same sampling scheme was followed in the control plot without these seven 162 163 additional soil cores. Thus, 100 soil cores were sampled in the agroforestry sub-plot (40 in tree rows, 60 in inter-rows) and 93 in the agricultural sub-plot (Fig. 2). All cores were sampled down to 2 m depth using a motor-driven micro caterpillar driller (8.5-cm diameter and 1-m long soil probe). The soil probe was successively pushed two times into the soil, to get 0-1 m and 1-2 m cores at each sampling point. Each soil core was then cut into ten segments, corresponding to the following depth increments: 0-10, 10-30, 30-50, 50-70, 70-100, 100-120, 120-140, 140-160, 160-180, and 180-200 cm.





Figure 2. Description of the intensive sampling scheme in the agroforestry and in the control
sub-plots. Circles represent hybrid walnuts, the grey strips represents the tree rows,
triangles are for soil cores on the regular grid (every 5 m), squares are for soil cores
on transects (every 1 m).

175

176 2.3. Use of field visible and near infrared spectroscopy to predict SOC

As core surface had been smoothed by the soil probe, each segment was refreshed with a knife before being scanned, in order to provide a plane but un-smoothed surface. Then, four VNIR spectra (from 350 to 2500 nm at 1 nm increment) were acquired in the field at different

places of each segment, using a portable spectrophotometer ASD LabSpec 2500 (Analytical 180 Spectral Devices, Boulder, CO, USA), and were then averaged. Reflectance spectra were 181 recorded as absorbance, which is the logarithm of the inverse of reflectance. The whole 182 183 spectrum population was composed of 1908 mean spectra (i.e. 193 cores with 10 sub-cores per core but a few samples were lost due to mechanical problems). In topsoil (0-30 cm), the 184 soil was dry and crumbled whereas in deeper soil horizons, it was moister and had higher 185 cohesion. Thus, two different predictive models were built: one for topsoil samples, the other 186 for subsoil (30-200 cm) samples. The "topsoil model" for predicting SOC was built using the 187 116 most representative topsoil samples, out of 380 samples, and the "subsoil model", using 188 189 the 142 most representative subsoil samples, out of 1488 samples. The procedure to select the most representative samples is presented below. The 0-10 cm soil layer from the tree rows (40 190 191 samples) was not used for the topsoil model as it contained abundant plant debris < 2 mm192 (roots, leaves, etc.) and a PCA revealed that these VNIR spectra were different from the whole spectra population. SOC concentration of these samples was therefore determined with 193 194 a CHN elemental analyzer, and, thus, not predicted by VNIR. The SOC concentration of the 258 samples selected for building the VNIR prediction models was also analyzed using a 195 CHN elemental analyzer. 196

197

198

2.4. VNIR spectra analysis and construction of predictive models

199 VNIR spectra analysis was conducted on topsoil and subsoil samples separately, using
200 the WinISI 4 software (Foss NIRSystems/ Tecator Infrasoft International, LLC, Silver Spring,
201 MD, USA) and R software version 3.1.1 (R Development Core Team, 2013). The most
202 representative samples, from a spectral viewpoint, were selected using the Kennard-Stone
203 algorithm, which is based on distance calculation between sample spectra in the principal

component space (Kennard and Stone, 1969). For the topsoil model, the calibration subset 204 included 104 samples (90%) selected as the most representative spectrally, and the validation 205 subset 12 samples (10%). For the subsoil model, the calibration subset included 128 samples 206 (90%), and the validation subset 14 samples. Fitting the spectra to the SOC concentrations 207 determined with a CHN elemental analyzer was performed using partial least squares 208 regression (PLSR; Martens and Naes, 1989). We tested common spectrum preprocessing 209 techniques including first and second derivatives, de-trending, standard normal variate 210 transformation and multiplicative scatter correction, but the best models were obtained when 211 no pre-treatment was applied on the spectra (data not shown). Then cross-validation was 212 performed within the calibration subset, using groups that were randomly selected (10 213 groups), in order to build the model used for making predictions on the samples not analyzed 214 in the laboratory. No outlier was removed. The number of components (latent variables) that 215 216 minimized the standard error of cross-validation (SECV) was retained for the PLSR. The performance of the models was assessed on the validation subsets using the coefficient of 217 218 determination (R²) and the standard error of prediction (SEP) between predicted and measured values, and also the ratio of standard deviation to SEP, denoted RPD, and the RPIQ, which is 219 the ratio of performance to IQ (interquartile distance), i.e. IQ/SEP = (Q3 - Q1)/SEP, where 220 Q1 is the 25th percentile and Q3 is the 75th percentile (Bellon-Maurel et al., 2010). Then all 221 sub-set samples (i.e., calibration and validation samples) were used to build models that were 222 applied on the samples not analyzed in the laboratory. The performance of these models was 223 also assessed according to R², SECV, RPD and RPIQ. 224

225

226

Table 1. External validation and prediction model results for soil organic carbon. N: numbers of samples; SD: standard deviation (mean and

standard deviation of the conventional determinations); R^2 : coefficient of determination; RPD is the ratio of performance to deviation,

i.e. the ratio of SD to SEP or SECV. RPIQ is the ratio of performance to IQ (interquartile distance), i.e. IQ/SEP (or SECV) = (Q3 -

231 Q1)/SEP (or SECV).

							То	psoil							
	External validation on 10% samples after								Prediction model using 100% samples						
calibration using 90% samples									(10-group cross-validation)						
Ν	Mean	SD	SEP	Bias	\mathbb{R}^2	RPD	RPIQ		Ν	Mean	SD	SECV	\mathbb{R}^2	RPD	RPIQ
	mg g ⁻¹	mg g ⁻¹	mg g ⁻¹	mg g ⁻¹						mg g ⁻¹	mg g ⁻¹	mg g ⁻¹			
12	9.71	2.09	1.04	-0.59	0.78	1.75	2.60		116	9.18	1.99	1.20	0.63	1.66	4.35

	Subsoil														
	External validation on 10% samples after									Prediction model using 100% samples					
	calibration using 90% samples								(10-group cross-validation)						
Ν	Mean	SD	SEP	Bias	\mathbb{R}^2	RPD	RPIQ		Ν	Mean	SD	SECV	\mathbb{R}^2	RPD	RPIQ
	mg g ⁻¹	mg g ⁻¹	mg g ⁻¹	mg g ⁻¹						mg g ⁻¹	mg g⁻¹	mg g ⁻¹			
14	6.19	1.80	0.83	0.01	0.74	2.03	3.03		142	6.06	1.86	0.77	0.83	2.40	4.85

232

Subsoil models performed better than topsoil models (Table 1, Fig. S1). In external 234 235 validation, RPD was higher than 2 for the subsoil, which has been considered a threshold for accurate NIRS prediction of soil properties in the laboratory (Chang et al., 2001). This RPD 236 237 threshold was not achieved for the topsoil model, but SOC concentrations were predicted for less than 60% of topsoil samples, the rest was directly analyzed with a CHN elemental 238 239 analyzer. It is worth noting that cross-validation on the whole set (for making prediction on 240 the samples not analyzed in the lab) yielded better results than external validation (on 10% of analyzed samples) in the subsoil, but the opposite was observed in the topsoil. 241

242

243

2.5. Bulk densities determination

Each segment was weighed in the field to determine its humid mass. Following this 244 step, each segment was crumbled and homogenized, and a representative sub-sample of about 245 246 300 g was sampled. Sub-samples were sieved at 2 mm to separate coarse fragments such as 247 stones and living roots. Coarse fragments represented less than 1% of each soil mass and were considered as negligible. Moisture contents were determined for 23 soil cores (i.e. 230 248 samples) after 48 h drying at 105°C, and were used to calculate the dry mass of all samples. 249 Bulk density (BD) was determined for each sample by dividing the dry mass of soil by its 250 volume in the soil corer tube. 251

252

253

2.6. Reference analysis measurements

After air drying, soil samples were oven dried at 40°C for 48 hours, sieved at 2 mm, and ball milled until they passed a 200 µm mesh sieve. Carbonates were removed by acid fumigation, following Harris et al., (2001). For this, 30 mg of soil was placed in open Ag-foil

capsules. The capsules were then placed in the wells of a microtiter plate and 50 µL of 257 demineralized water was added in each capsule. The microtiter plate was then placed in a 258 vacuum desiccator with a beaker filled with 100 mL of concentrated HCl (37%). The samples 259 260 were exposed to HCl vapors for 8 hours, and were then dried at 60°C for 48 hours. Capsules were then closed in a bigger tin capsule. Decarbonated samples were analyzed for organic 261 carbon concentration with a CHN elemental analyzer (Carlo Erba NA 2000, Milan, Italy). 262 263 Isotopic measurements were performed on a few samples to check that decarbonation was well performed (δ^{13} C OM = -25 ‰). 264

265

266

2.7. Soil organic carbon stock calculation

In most studies comparing SOC stocks between treatments or over time periods, SOC 267 stocks have been quantified to a fixed depth as the product of soil bulk density, depth and 268 269 SOC concentration. However, if soil bulk density differs between the treatments being 270 compared, the fixed-depth method has been shown to introduce errors (Ellert et al., 2002). A more accurate method is to use an equivalent soil mass (ESM) (Ellert and Bettany, 1995). We 271 defined a reference soil mass profile that was used as the basis for comparison, based on the 272 lowest soil mass observed at each sampling depth and location. For this reference, soil mass 273 layers (0-1000, 1000-4000, 4000-7300, 7300-10700, 10700-15700, 15700-18700, 18700-274 21900, 21900-25100, 25100-28300, 28300-31500 Mg ha⁻¹) corresponded roughly to soil 275 depth layers (0-10, 10-30, 30-50, 50-70, 70-100, 100-120, 120-140, 140-160, 160-180, 180-276 277 200 cm, respectively). For the different treatments (control, tree row, inter-row), SOC stocks were calculated on this basis, soil mass was the same, whereas depth layer varied (Table 2). 278 279 The effect of the ESM correction can be seen in Table S1. SOC stocks in the agroforestry plot 280 were calculated with tree rows representing 16% of the plot surface area and inter-rows 84%:

281 SOC stock_{Agroforestry} =
$$0.16 \times \text{SOC stock}_{Tree \, row} + 0.84 \times \text{SOC stock}_{Inter \, row}$$
 (1)

We defined delta SOC stock as the difference between SOC stock in the agroforestry plot and in the control plot:

$$\Delta \text{ SOC stock} = \text{SOC stock}_{Agroforestry} - \text{ SOC stock}_{Control}$$
(2)

All SOC stocks were expressed in Mg C ha⁻¹. SOC accumulation rates (kg C ha⁻¹ yr⁻¹) were calculated by dividing delta stocks by the number of years since the tree planting (18 years):

287 SOC accumulation rate =
$$\frac{\Delta \text{ SOC stock}}{18} \times 1000$$
 (3)

288 2.8. Particle-size fractionation

Particle-size fractionation was performed for five soil cores from the inter-rows, five from the 289 tree rows and six from the control plot, and at four depths: 0-10, 10-30, 70-100 and 160-290 291 180 cm. Thus, 64 soil samples were fractionated, as described in Balesdent et al. (1998) and Gavinelli et al. (1995). Briefly, 20 g of 2-mm sieved samples were soaked overnight at 4°C in 292 300 mL of deionized water, with 10 mL of sodium metaphosphate (HMP, 50 g L⁻¹). Samples 293 were then shaken 2 h with 10 glass balls in a rotary shaker, at 43 rpm. The soil suspension 294 was wet-sieved through 200-µm and 50-µm sieves, successively. The fractions remaining on 295 296 the sieves were density-separated into organic fractions, floating in water, and remaining mineral fractions. The 0-50 µm suspension was ultrasonicated during 10 min with a probe-297 type ultrasound generating unit (Fisher Bioblock Scientific, Illkirch, France) having a power 298 299 output of 600 watts and working in 0.7:0.3 operating/interruption intervals. This 0-50 µm suspension was then sieved through a 20-µm sieve. The resulting 0-20 µm suspension was 300 transferred to 1-L glass cylinders, which were then shaken by hand and 50 mL of the 301 suspension were withdrawn immediately after. They constituted an aliquot of the entire 0-20 302 µm fraction. After a settling time of 8 h approximately, a second aliquot of 50 mL was 303

removed by siphoning the upper 10 cm of the suspension left after the first sampling. This 304 represented an aliquot of the 0-2 µm fraction. A third aliquot was also collected in the upper 305 10 cm, and centrifuged two times 35 min, at 4000 rpm. This aliquot was then filtered at 2 µm 306 to get the hydrosoluble fraction. Fractions were then dried at 40°C, finely ground, 307 decarbonated and analyzed with a CHN elemental analyzer. A binocular microscope was used 308 to check if separation of coarse mineral fractions and of light organic coarse fractions (200-309 2000 and 50-200 µm) was well done. No pyrogenic particles were observed. Organic carbon 310 contents of coarse mineral fractions were then assumed to be 0 mg C g⁻¹. A sub-sample of 311 each of the 64 selected samples was used to perform a classical textural analysis after 312 destruction of organic matter. These texture analyses were used to evaluate the quality of the 313 dispersion for soil particle size fractionation. 314

315

316 2.9. Calculation of SOC saturation

The theoretical value of SOC saturation was calculated according to the equation proposed by(Hassink, 1997):

319
$$SOC_{sat-pot} = 4.09 + 0.37 \times \text{particles} < 20 \,\mu\text{m}$$
 (4)

where SOC_{*sat-pot*} is the potential SOC saturation (mg C g⁻¹) and where particles < 20 μ m represents the proportion of fine soil particles <20 μ m (%).

To calculate the SOC saturation deficit (Angers et al., 2011; Wiesmeier et al., 2014), the estimated current SOC concentrations of the fine fraction were subtracted from the potential SOC saturation:

$$SOC_{sat-def} = SOC_{sat-pot} - SOC_{cur}$$
(5)

where SOC_{*sat-def*} is the SOC saturation deficit (mg C g⁻¹) and SOC_{*cur*} is the current mean SOC concentration of the fine fraction <20 μ m (mg C g⁻¹). The total amount of the SOC storage potential (SOC_{*stor-pot*}, Mg C ha⁻¹) was calculated multiplying SOC_{*sat-def*} by soil bulk density and soil layer thickness.

These calculations were performed for the four depths where particle-size fractionation was done (0-10, 10-30, 70-100 and 160-180 cm). But as the equation proposed by (Hassink, 1997) was calibrated for topsoil layers, calculations for deep soil layers are only indicative.

333

The observed variability in a soil property *Z* such as SOC concentration results from complex processes operating over various spatial scales. A simple but useful statistical model for *Z* at a set of observations that could be spatially located, $s_i = \{s_1, s_2, \dots, s_q\}$ is

338
$$Z(\mathbf{s}_i) = \mu(\mathbf{s}_i) + \varepsilon(\mathbf{s}_i)$$
(6)

where $\mu(s_i)$ is a deterministic component and $\varepsilon(s_i)$ is a correlated random component that can include a pure noise random one. A soil property can be correlated with other environmental variables such as, in this work, the distance to the closest tree. This can be represented in Equation 6 by assuming that $\mu(s_i)$ comprises an additive combination of one or more fixed effect:

344
$$\mu(\boldsymbol{s}_i) = \beta_0 + \sum_{j=1}^q \beta_j x_j(\boldsymbol{s}_i)$$
(7)

where x_j ($j = 1, 2, \dots, q$) are q auxiliary variables and β_0, \dots, β_q are the associated fixed effects. This model is referred as a Mixed Effects Model which offers a flexible framework by which to model the sources of variation and correlation that arise from grouped data (Lark et al., 2006; Pinheiro and Bates, 2000). In this work, we fitted two different linear mixed models(LMM).

We first fitted a LMM using the whole set of the bulk densities, SOC concentrations, and 350 SOC stocks observations at the different depths. We used the *nlme* package (Pinheiro et al., 351 2013). Soil core ID was considered as a random effect to take into account a sample effect. 352 These soil properties were then compared by depth and per location (control, tree row, inter-353 row). An ANOVA was performed on these models. We then used the multcomp package 354 355 (Hothorn et al., 2008) to perform a post hoc analysis and determine which means differed significantly between the control, tree rows and inter-rows, using the Tukey-Kramer test, 356 designed for unbalanced data. To study spatial influence on SOC stocks, "distance to the 357 closest tree" was added to the LMM model, and an ANOVA was performed. 358

Secondly, we fitted a LMM in a geostatistical framework using the cumulated SOC stock 359 observations for 3 depths (0-30 cm, 0-100 cm and 0-200 cm). In a spatial context, the random 360 effects of the LMM describe spatially-correlated random variation. The LMM model is then 361 parameterized by a global vector, called $\boldsymbol{\Theta}$, of model parameters which include the parameters 362 of the covariance function and the fixed effects coefficients. These can be fitted to the data by 363 364 a likelihood method. Lark et al. (2006) described how the maximum likelihood estimator is 365 biased in the presence of fixed effects and suggested that the restricted maximum likelihood 366 estimator (REML) should be applied. Following Villanneau et al., (2011) we have tested the assumption that the random effects are spatially correlated by comparing the quality of the 367 model-fit for spatially correlated and spatially independent models (usually called pure nugget 368 model). Webster and McBratney, (1989) suggested that the Akaike information criterion 369 (AIC, Akaike, 1974) should be used to compare different spatially correlated models. Once 370 the parameters of the LMM have been fitted, they may be plugged into the best linear 371 unbiased predictor to form the empirical best linear unbiased predictor (E-BLUP) of the 372

property at unsampled sites (Lark et al., 2006). The error variance of the E-BLUP can also be 373 computed at any unsampled site. For this, the value of fixed effects covariates must be known 374 at each prediction site. We therefore calculated several grids of the fixed effects with a 25 cm 375 376 cell size. The use of any model of spatial variation implies that assumptions have been made about the type of variation the data exhibit. Once the model has been fitted, cross-validation 377 378 can be used to confirm that these assumptions are reasonable and that the spatial model 379 appropriately describes the variation. We therefore computed a 'leave-one-out crossvalidation'. For each sampling location, s_i ($i = 1, 2, \dots, q$), the value of the property at s_i was 380 predicted by the E-BLUP upon the vector of observations excluding $Z(s_i)$, in order to 381 compute the standardized squared prediction error (SSPE: the squared difference between the 382 E-BLUP and the observed value divided by the computed prediction error variance (PEV)). 383 384 Under an assumption of normal prediction errors, the expected mean SSPE is 1.0 if the PEVs are reliable (which requires an appropriate variogram model), and the expected median SSPE 385 is 0.455. The spatial analysis package *GeoR* (Ribeiro and Diggle, 2001) was used for REML 386 387 fitting and kriging.

Finally, a Kruskal-Wallis test (Kruskal and Wallis, 1952) was performed to analyze SOC concentration in soil fractions per depth and per location (5 or 6 replicates). This test was followed by a post hoc analysis using Dunn's test (Dunn, 1964) with a Bonferroni correction (p-value=0.017).

All the statistical analyses were performed using R software version 3.1.1 (R Development
Core Team, 2013), at a significance level of <0.05.

394

395

Table 2. Soil organic carbon stocks (Mg C ha⁻¹) and SOC accumulation rates (kg C ha⁻¹ yr⁻¹). Associated errors are standard errors (40 replicates for the tree-row, 60 replicates for the inter-row, and 93 replicates for the control plot). ESM = Equivalent Soil Mass. Significantly (pvalue<0.05) different SOC stocks are followed by different letters.

Cumulated ESM (Mg ha ⁻¹)	Cumulate	ed calculated ESM (cm)	depth to		Cumulated SOC s	stocks (Mg C ha ⁻¹)		Δ SOC stocks (Mg C ha ⁻¹)	lation rates 1 ⁻¹ yr ⁻¹)	
	Tree-row	Inter-row	Control	Tree-row	Inter-row	Agroforestry	Control	Δ (Agroforestry – Control)	Agroforestry vs Control	Inter-row vs Control
1000	0-9	0-8	0-7	21.6 ± 1.0 a	9.8 ± 0.4 c	11.7 ± 0.3 b	9.3 ± 0.1 c	2.3 ± 0.4	129 ± 20	24 ± 21
4000	0-29	0-27	0-26	52.8 ± 1.4 a	$\begin{array}{c} 37.9\pm0.6\\ c\end{array}$	$\begin{array}{c} 40.3\pm0.5\\ b\end{array}$	$\begin{array}{c} 35.8\pm0.2\\ d\end{array}$	4.5 ± 0.6	248 ± 31	115 ± 33
7300	0-49	0-47	0-45	77.1 ± 1.5 a	$\begin{array}{c} 62.0\pm0.7\\ \text{c}\end{array}$	$\begin{array}{c} 64.4\pm0.6\\ b\end{array}$	$59.4 \pm 0.2 \\ d$	5.0 ± 0.6	276 ± 36	141 ± 39
10700	0-69	0-66	0-64	98.1 ± 1.5 a	$\begin{array}{c} 82.4\pm0.7\\ c\end{array}$	$\begin{array}{c} 84.9\pm0.6\\ b\end{array}$	$79.7\pm0.3\\d$	5.1 ± 0.7	286 ± 39	147 ± 43
15700	0-98	0-95	0-93	130.4 ± 1.5 a	113.7 ± 0.7 c	116.4 ± 0.7 b	110.1 ± 0.3 d	6.3 ± 0.7	350 ± 41	202 ± 45
18700	0-118	0-115	0-112	150.3 ± 1.5 a	133.1 ± 0.8 c	$\begin{array}{c} 135.9\pm0.7\\ b\end{array}$	$\begin{array}{c} 129.3 \pm 0.4 \\ d \end{array}$	6.5 ± 0.8	363 ± 43	210 ± 46
21900	0-137	0-134	0-131	170.9 ± 1.5 a	152.8 ± 0.8 c	$\begin{array}{c} 155.7\pm0.7\\ b\end{array}$	$\begin{array}{c} 149.5\pm0.4\\ c\end{array}$	6.2 ± 0.8	345 ± 44	185 ± 48
25100	0-157	0-154	0-150	191.0 ± 1.6 a	172.4 ± 0.8 c	$\begin{array}{c} 175.4\pm0.7\\ b\end{array}$	169.9 ± 0.4 c	5.5 ± 0.8	306 ± 45	140 ± 49
28300	0-176	0-173	0-170	209.5 ± 1.6 a	190.5 ± 0.8 c	$\begin{array}{c} 193.5\pm0.7\\ b\end{array}$	189.3 ± 0.4 c	4.3 ± 0.8	238 ± 47	69 ± 51
31500	0-196	0-193	0-189	226.1 ± 1.6 a	$\begin{array}{c} 206.0 \pm 0.84 \\ c \end{array}$	$\begin{array}{c} 209.2\pm0.7\\ b\end{array}$	205.9 ± 0.4 c	3.3 ± 0.9	183 ± 48	5 ± 53

400 **3. Results**

401

3.1. Changes in soil texture with depth

Clay, silt and sand profiles were very similar at both plots (Fig. 3). Soil texture was homogeneous in the first 50 cm. Clay and silt contents linearly increased till 100 cm soil depth to reach about 325 g kg⁻¹ and 575 g kg⁻¹ respectively, while sand content decreased. Soil texture did not change between 100 and 200 cm soil depth. Below 140 cm depth, clay and sand content were significantly different (F=71.31, P<0.001) in both plots, but the maximum difference was less than 20 g kg⁻¹.



408

Figure 3. Changes in soil texture with depth in the control plot and in the agroforestry plot.
Error bars represent standard errors (n=100 in the agroforestry, n=93 in the control).

411

412

414 *3.2. Soil bulk densities*

419

Soil bulk densities were significantly higher in the control plot than in the tree row at all depths except for 30-50 and 140-160 cm, and higher than in the inter-row, except for 10-30 and below 140 cm depth (Table 3). In the agroforestry system, soil bulk densities were higher in the inter-row than in the tree row for 0-10 and 10-30 cm.



Figure 4. Soil organic carbon concentration (mg C g⁻¹ soil) of soil layers to 2-m depth in the
control plot and in the agroforestry plot. Error bars represent standard errors (n=100
in the agroforestry, n=93 in the control). Significantly (p-value<0.05) different SOC
concentrations per depth are followed by different letters.

426	Depth (cm)	Agroforestry – tree row	Agroforestry – inter-row	Control plot
	0-10	$1.10 \pm 0.02 \text{ c}$	$1.23\pm0.03~b$	1.41 ± 0.01 a
427	10-30	$1.49\pm0.01~b$	1.60 ± 0.02 a	$1.61 \pm 0.00 \ a$
427	30-50	$1.71 \pm 0.01 \text{ ab}$	$1.67\pm0.02~b$	$1.73 \pm 0.00 \text{ a}$
	50-70	$1.73 \pm 0.01 \text{ c}$	$1.77 \pm 0.01 \text{ b}$	$1.80 \pm 0.00 \ a$
428	70-100	$1.68\pm0.00~\mathrm{c}$	$1.71\pm0.00~b$	$1.74 \pm 0.00 \ a$
120	100-120	$1.55 \pm 0.01 \text{ b}$	$1.55\pm0.01~b$	$1.61 \pm 0.00 \ a$
	120-140	$1.63\pm0.00~b$	$1.64 \pm 0.01 \text{ b}$	$1.65 \pm 0.00 \text{ a}$
429	140-160	$1.64 \pm 0.00 \text{ a}$	1.64 ± 0.01 a	$1.65 \pm 0.00 \text{ a}$
	160-180	$1.62 \pm 0.01 \text{ b}$	1.65 ± 0.01 a	$1.65 \pm 0.00 \text{ a}$
	180-200	$1.64\pm0.00~b$	$1.65 \pm 0.00 \ a$	$1.65 \pm 0.00 \text{ a}$
430				

425 Associated errors are standard errors (40 replicates for the tree-row, 60 replicates for the inter-row, and 93 replicates for the control plot).

Table 3. Mean soil bulk densities (g cm⁻³). For a given depth, means followed by the same letters do not differ significantly at p = 0.05.

An ANOVA performed on the LMM model revealed that soil depth (F-value=270, P<0.0001) 436 and location, i.e., tree row vs. inter-row (F-value=171, P<0.0001), were the only variables 437 affecting significantly SOC concentrations. Distance to the closest tree had no significant 438 effect (F-value=1.3, P=0.28). As shown in Fig. 4, for 0-10 cm, SOC concentration doubled in 439 the tree row (21.6 \pm 0.8 mg C g⁻¹) compared to the inter-row (9.8 \pm 0.1 mg C g⁻¹) and to the 440 control (9.3 \pm 0.1 mg C g⁻¹), whereas the latter two were not significantly different. SOC 441 442 concentration was significantly higher in the tree row than in the control plot to 120 cm soil depth, except in the 50-70 cm soil layer where no difference was observed. SOC 443 concentration was significantly higher in the tree row than in the inter-row to 30 cm soil 444 depth. 445

446

447

3.4. Soil organic carbon stocks

Fig. 5 represents SOC stocks in the agroforestry plot as a function of soil depth, location and distance to the closest tree. For a given depth and distance to the closest tree, variability of SOC stocks was high, and there was no effect of the distance to the closest tree on SOC stocks (Fig. 5). An ANOVA performed on the LMM model confirmed that SOC stocks were significantly influenced by soil depth (F-value=483, P<0.0001) and location, i.e., tree row vs. inter-row (F-value=66, P<0.0001), but not by the distance to the closest tree (F-value=1.5, P=0.22).

For an equivalent soil mass (ESM) of 4000 Mg ha⁻¹ (to 26-29 cm depth), SOC stocks were significantly higher in the tree row than in the inter row and in the control (Table 2). For an ESM of 31500 Mg ha⁻¹ (to 189-196 cm depth), SOC stocks were about 20 Mg C ha⁻¹ higher in the tree rows compared to the inter-rows or to the control. Cumulated SOC stocks were 459 significantly higher in the inter-row than in the control plot to an ESM of 18700 Mg ha⁻¹ (to
460 112-115 cm depth), except for an ESM of 1000 Mg ha⁻¹ where not difference was found
461 (Table 2).

At the plot scale, cumulated SOC stocks in the agroforestry plot were significantly higher than in the control plot at all depths (Table 2). For an ESM of 4000 Mg ha⁻¹ (to 26-29 cm depth), SOC stocks were 40.3 ± 0.5 Mg C ha⁻¹ and 35.8 ± 0.2 Mg C ha⁻¹ in the agroforestry and in the control, respectively. For a soil mass of 15700 Mg ha⁻¹ (to 93-98 cm depth), Δ SOC stock between the agroforestry and the control was 6.3 ± 0.7 Mg C ha⁻¹. This difference was much lower without the ESM correction (Table S1).

468

469

3.5. Soil organic carbon accumulation rates

470 Compared to the control, inter-rows accumulated $115 \pm 33 \text{ kg C} \text{ ha}^{-1} \text{ yr}^{-1}$ for an ESM of 4000 471 Mg ha⁻¹ (26-29 cm) (Table 2), and $202 \pm 45 \text{ kg C} \text{ ha}^{-1} \text{ yr}^{-1}$ for an ESM of 15700 Mg ha⁻¹ (93-472 98 cm). SOC accumulation rates in the agroforestry plot compared to the control were 248 ± 473 31 kg C ha⁻¹ yr⁻¹ for an ESM of 4000 Mg ha⁻¹, 350 ± 41 kg C ha⁻¹ yr⁻¹ an ESM of 15700 Mg 474 ha⁻¹, and 183 ± 48 kg C ha⁻¹ yr⁻¹ an ESM of 31500 Mg ha⁻¹ (Table 2). The additional SOC 475 storage rates for 0-10 cm and 10-30 cm were respectively explained at 80% and 60% by the 476 tree rows.

477

478

479



Figure 5. Soil organic carbon stocks (Mg C ha⁻¹) in the agroforestry plot as a function of depth, location (tree row vs. inter-row) and distance to
the closest tree. The lines represent the regression lines fitted using soil samples per investigated depth. The gray shades display the
prediction confidence interval at the 0.95 level.

Table 4. Summary of selected models fitted to the data on cumulated soil organic carbon stocks at 3 depths (0-30 cm, 0-100 cm and 0-200 cm) for the 2 plots, and cross validation. SSPE, standardized squared prediction errors; ME, mean error (Mg C ha⁻¹); RMSQE, root mean squared error (Mg C ha⁻¹); AIC, AIC of the spatially correlated model; AIC.ns, AIC of the non-spatially correlated model; β_0 and β_1 the fixed effects (Mg C ha⁻¹). Bold characters represent the smallest AIC for each depth. The medians and the mean of the cross validation statistics are within the 95% confidence interval.

	Depth (cm)	Mean SSPE	Median SSPE	ME	RMQSE	AIC	AIC.ns	eta_0	eta_1	Nugget	Sill	Range	Nugget to Sill ratio
A groforestry	0-30	0.99	0.36	-0.004	20.7 43 3	585 662	583	38.1 114 1	14.8 16.4	19.7 36.0	1.3 16.3	15.2 12.8	0.94
Agiololesuy	0-200	0.99	0.45	0.055	123.1	002 769	780	207.1	19.4	97.8	79.2	12.8	0.55
Control	0-30	1.01	0.33	0.000	2.6	361	357	35.9	-	2.4	0.2	19.4 12.6	0.93
Control	0-100	0.98	0.30	0.061	23.7 57.5	578 665	579 681	208.9	-	20.2 16.4	85.8	6.3	0.65

3.6. Spatial distribution of SOC stocks

The AIC (Table 4) of the spatially correlated model were less than that of the spatially 491 uncorrelated model for 2 depths (0-100 cm and 0-200 cm for the agroforestry and the control 492 plots), indicating that spatial correlation should be included in the model of variation. We 493 tested several models of spatial variation and retained the spherical model (Webster and 494 495 Oliver, 2007). For top soil depth of the two plots (0-30 cm), the AIC of the spatially uncorrelated model was slightly the smallest indicating that the residual variation could be 496 independent once fixed effects had been included in the model. But the difference was very 497 small so we considered the spatially correlated model for the rest of the study. The cross-498 validation results confirmed the validity of the fitted LMM. The nugget to sill ratio measures 499 500 the unexplained part of the observed variability. The smallest value was observed for the 0-200 cm depth in the control plot and the higher was observed for the 0-30 cm depth in both 501 plots. When mapping the SOC stocks for three fixed depths with the BLUP in the two plots, a 502 503 clear pattern can be observed in the agroforestry plot, with high SOC stocks in the tree rows (Fig. 6). The fitted fixed effects indicate that, in average, the SOC stocks were 15 to 20 Mg C 504 ha⁻¹ higher in the tree rows to 30 to 200 cm depth (Table 4). At the opposite, the control plot 505 did not exhibit any spatial pattern. 506

507

508

- 509
- 510

511



Figure 6. Krieged maps of cumulated soil organic carbon stocks (Mg C ha⁻¹) in the
agroforestry and in the control plot.

517 *3.7. Organic carbon distribution in soil fractions*

An average mass yield of 98% and an average carbon yield of 96% were obtained, showing the quality of the particle size fractionation. Furthermore, the variation between soil texture and soil fractionation was only 5-6% (data not shown). Soil segments used for soil fractionation had similar total SOC concentrations compared to mean SOC concentrations at the same depth (Fig. S2). However, the small differences found between SOC concentrations in the inter row and in the control was not visible with the soil segments used for fractionation.

For 0-10 cm depth, the distribution of OC in particle size fractions was strongly modified in the tree rows, with an important increase of C in particulate organic matter (POM) fractions (50-200 μ m and 200-2000 μ m) compared to the inter-row and to the control (Fig. 7). An increase of C in silt size fractions (2-20 μ m and 20-50 μ m) of the tree rows compared to the inter row and to the control was also observed. Significantly higher C concentrations in the clay fraction (0-2 μ m) were observed in the tree row than in the inter-row (Fig. S3), but it was not the case for the amount of C in the clay fraction per gram of soil (Fig. 7).

Similar trends in C distribution in fractions were observed at 10-30 cm depth compared to 0-10 cm, although with much smaller differences (Figs. 7, S3). At deeper depths (70-100 and 160-180 cm) there were no differences between the three locations (tree row, inter-row and control) except a lower amount of C in the soluble fraction in the tree row. The potential SOC saturation of particles <20 μ m was not reached at any depths (Table 5), and the SOC deficit was high. The saturation capacity was far from being reached, as it amounted 17 to 40% of saturation capacity in the tree rows.

539





Figure 7. Organic carbon contents in each soil fraction (mg C g^{-1} soil). Error bars represent standard errors (n=6 in the control, n=5 in the interrow and in the tree row). OF = Organic fraction, F = organo-mineral fraction. 0-2, 2-20, 20-50, 50-200 and 200-2000 represent particle size (μ m). Means followed by the same letters do not differ significantly at p=0.017 (Dunn's test with Bonferroni correction).

Table 5. Soil organic carbon saturation of the fractionated soil samples in the agroforestry plot. $SOC_{sat-pot}$, potential SOC saturation (mg C g⁻¹); SOC_{cur}, current mean SOC concentration of the fine fraction <20 µm (mg C g⁻¹); SOC_{sat-def}, SOC saturation deficit (mg C g⁻¹); SOC_{stor-pot},

547 total amount of the SOC storage potential (Mg C ha⁻¹). Associated errors are standard errors (n=5). Values of SOC saturation for deep soil

548 layers are only indicative.

	SOC _{sat-pot}	SOC _{cur} (mg C g^{-1})	SOC _{sat-def}	(mg C g ⁻¹)	SO	Cur	SOC _{stor-pot}
_	$(mg C g^{-1})$					SOC _{so}	at-pot	$(Mg C ha^{-1})$
Depth (cm)	Agroforestry	Tree row	Inter-row	Tree row	Inter-row	Tree row	Inter-row	Agroforestry
0-10	18.0 ± 0.4	7.2 ± 0.3	5.4 ± 0.3	10.3 ± 0.4	13.1 ± 0.4	40%	30%	15.3 ± 0.4
10-30	18.7 ± 0.4	6.1 ± 0.1	5.4 ± 0.1	12.6 ± 0.3	13.3 ± 0.3	33%	29%	41.8 ± 0.9
70-100	32.9 ± 0.8	5.6 ± 0.1	5.6 ± 0.1	26.9 ± 0.7	27.6 ± 0.4	17%	17%	140.7 ± 1.9
160-180	32.0 ± 1.1	4.6 ± 0.2	4.6 ± 0.3	26.8 ± 0.7	28.1 ± 0.9	14%	14%	91.9 ± 2.4

549 *3.8. Distribution of additional OC in soil fractions*

For 0-10 cm depth, the additional OC stored between the tree row and the inter-row was explained at 80% by POM fractions, at 15% by silt size fractions, and at 5% by clay fraction, whereas the additional OC stored between the tree row and the control was explained at 80% by POM and at 20% by silt size fractions (Fig. 7). For 10-30 cm, the additional SOC storage between the tree row and the inter-row was explained at 50% by POM fractions, at 25% by coarse and fine silt fractions, and at 25% by clay fraction (Fig. 7), whereas when comparing the tree row and the control these numbers were of 50% (POM) and 50% (silt).

557

558 **4. Discussion**

559

4.1. A shallow additional SOC storage

560 Sampling to 2-m soil depth indicated that the 0-30 cm soil layer contained less than 20% of total SOC stocks to 2-m depth, demonstrating the importance of deeper soil layers for storing 561 562 SOC (Harper and Tibbett, 2013; Jobbagy and Jackson, 2000). SOC stocks observed in 0-30 cm, from 36 to 41 Mg C ha⁻¹, were comparable to reported values for the Mediterranean 563 region, i.e., 25 to 50 Mg C ha⁻¹ (Martin et al., 2011; Muñoz-Rojas et al., 2012). Additional 564 SOC storage in the agroforestry system compared to the agricultural system was mainly 565 566 observed up to 30 cm soil depth in the inter-row and up to 50 cm in the tree row. A 567 companion study at the same site indicated that 60% of additional OM inputs (leaf litter, aboveground and belowground biomass of the natural vegetation in the tree row, tree fine 568 roots) to 2 m depth in the agroforestry plot compared to the control plot were located in the 569 first 50 cm (unpublished data). Even if 50% of tree fine root density was found between 1 and 570 4 m soil depth (Cardinael et al., 2015), it was also proven at this site (Germon et al., 571 submitted) and at other sites (Hendrick and Pregitzer, 1996) that the turnover rate of fine roots 572

decreased with increasing depth, resulting in low OM inputs in deep soil layers. Time since 573 the tree planting (18 years) is probably not long enough to detect changes in SOC stocks at 574 deeper soil depths considering low organic inputs below 1 m depth. For 2012, organic C input 575 due to tree fine root mortality was estimated to be less than 150 kg C ha⁻¹ for 100-200 cm soil 576 depth. Below 1.2 m soil depth, delta of cumulated SOC stocks between the agroforestry and 577 the control plot decreased, due to higher SOC concentrations and stocks in the control at these 578 depths. These higher SOC concentrations were linked to higher SOC concentrations in the 579 clay fraction. This difference may be due to pre-experimental soil heterogeneity, the soil in 580 the agroforestry plot may have had a lower level of SOC below 1.2 m depth before tree 581 planting. An initial heterogeneity was also proposed by Upson and Burgess (2013) who found 582 higher SOC stocks at depth in a control plot compared to an agroforestry plot in an 583 experimental site in England. This shows the limit of paired comparisons - or synchronic 584 585 studies - to evaluate SOC changes after land use change (Junior et al., 2013; Olson et al., 2014), and pleads for long-term diachronic studies in agroforestry systems. An alternative 586 587 explanation could be a positive priming effect, i.e., the acceleration of native SOC decomposition by the supply of fresh organic carbon (Fontaine et al., 2007, 2004) from the 588 trees. However, this seems highly unlikely since positive priming effect could not explain 589 such a high C loss of about 3.2 Mg C ha⁻¹ between 1.2 and 2.0 m soil depth in 18 years, i.e., 590 591 about 180 kg C ha⁻¹ yr⁻¹. Another hypothesis to explain higher SOC stocks below 1.2 m depth in the control plot is a different belowground water regime between the two plots. Water table 592 depth at this site is known to be very variable (between 5 to 7 m). A shallower water table in 593 the agroforestry plot compared to the control plot may promote capillary action, and therefore 594 cause wetting-drying cycles that could enhance SOM decomposition in deep soil layers 595 (Borken and Matzner, 2009). 596

The high SOC stocks observed in tree rows accounted for an important part of SOC stocks of 599 600 the agroforestry plot even though tree rows only represented 16% of the surface area. In a poplar (Populus L.) silvoarable agroforestry experiment in England, Upson and Burgess, 601 (2013) also found that the SOC concentration was greater in the top 40 cm under the tree row 602 (19.6 mg C g^{-1}) in the agroforestry treatment than in the cropped alleys (17 mg C g^{-1}), or the 603 arable control (17.1 mg C g⁻¹). Tree rows are comparable to a natural permanent pasture with 604 605 trees, given that spontaneous herbaceous vegetation grows and that the soil is not tilled. 606 Conversion of arable lands to permanent grasslands is recognized as an efficient land use for climate change mitigation (Soussana et al., 2004). Grasslands can accumulate SOC at a very 607 608 high rate. For instance, it was estimated on about 20 years old field experiments that conversion from crop cultivation to pasture stored SOC at a rate of 1.01 Mg C ha⁻¹ yr⁻¹ in 0-30 609 cm (Conant et al., 2001). In our case, SOC accumulation rate in the tree rows was 0.94 ± 0.09 610 Mg C ha⁻¹ yr⁻¹ in 0-30 cm. Management of tree rows could therefore have an important role in 611 improving agroforestry systems in terms of SOC storage. Improved grass species could be 612 sown in the tree rows, as well as shrubs between trees. Further research should focus on this 613 aspect to evaluate benefits in terms of SOC storage and biodiversity for instance. 614

615

598

616

4.3. Homogeneous distribution of SOC stocks in the cropped alley

There was no significant effect of the distance to the trees on SOC stocks at all depths, either in the tree row or in the inter-row. This was also indicated by the maps of the SOC stocks. Tree density was high at this site, and walnuts were about 13 m in height, which is also the distance between two tree rows. This could explain the homogeneous distribution of leaf litterfall observed in the plot (personal observation). In a similar agroforestry system in terms

of tree density in Canada, Bambrick et al., (2010) and Peichl et al., (2006) also found no 622 effect of the distance to the trees on SOC stocks to 20 cm depth. They also suggested that the 623 18 m high poplar trees distributed litterfall equally in the crop alleys. Close to the tree rows (1 624 to 2 m distance), the intercrop had a lower yield (15% less in 2012) compared to the middle of 625 the inter-row at the study site (Dufour et al., 2013). On the contrary, tree fine root density was 626 higher close to the tree rows (2.79 t DM ha⁻¹ between 0 and 1.5 m from the tree row in the 627 inter row, and to 4-m soil depth) than in the middle of the inter-rows (1.32 t DM ha⁻¹ between 628 3 and 4.5 m from the tree row in the inter row, and to 4-m soil depth) (Cardinael et al., 2015). 629 Thus, lower carbon inputs from crop residues close to the tree rows may be counterbalanced 630 with higher inputs from tree fine root mortality, explaining homogeneous distribution of SOC 631 stocks within the inter-row (Bambrick et al., 2010; Peichl et al., 2006). In the tree row, 632 homogeneous distribution of SOC stocks may be explained by the short distance between 633 634 trees and by the presence of abundant herbaceous vegetation.

635

636

4.4. Agroforestry systems: an efficient land use to improve SOC stocks

Compared to other agroforestry systems having about the same tree density, a lower SOC 637 accumulation rate in 0-30 cm (0.25 Mg C ha⁻¹ yr⁻¹) was observed at our site. Peichl et al. 638 (2006) reported a SOC accumulation rate of 1.04 Mg C ha⁻¹ yr⁻¹ (0-20 cm) in a 13-year old 639 temperate barley (Hordeum vulgare L.)-poplar intercropping system (111 trees ha⁻¹). In a 21-640 year old agroforestry system in Canada where poplars were intercropped with a rotation of 641 642 wheat (Triticum aestivum L.), soybean (Glycine max (L.) Merr.) and corn (Zea mays L.), Bambrick et al. (2010) estimated a SOC accumulation rate of 0.30 Mg C ha⁻¹ yr⁻¹ (0-20 cm). 643 Our lower accumulation rate may be explained by warmer climate, higher temperatures 644 645 enhancing OM decomposition (Hamdi et al., 2013). Moreover, valuable hardwood species like walnut trees have a slower growing rate than fast growing species like poplar (Teck and
Hilt, 1991), and therefore for a same tree age, the amount of OC inputs (leaflitter, fine roots)
to the soil is lower for slow growing species.

Together with other climate-smart farming practices (Lipper et al., 2014), alley-cropping 649 agroforestry systems have the potential to enhance SOC stocks and to contribute to climate 650 651 change mitigation (Nair et al., 2010; Pellerin et al., 2013). No-till farming is a commonly cited agricultural practice supposed to have a positive impact on SOC stocks. But recent meta-652 analyses showed this practice had no effect on SOC stocks to 40 cm depth (Luo et al., 2010) 653 or a smaller one (0.23 Mg C ha⁻¹ yr⁻¹ to 30 cm depth) than previously estimated (Virto et al., 654 2011). A meta-analysis also revealed that the inclusion of cover crops in cropping systems 655 could accumulate SOC at a rate of 0.32 ± 0.08 Mg C ha⁻¹ yr⁻¹ to a depth of 22 cm (Poeplau 656 and Don, 2015). At our site, we found a mean SOC accumulation rate of 0.13 in 0-30 cm in 657 the inter-rows compared to the control. This rate reached 0.25 Mg C ha⁻¹ yr⁻¹ for the whole 658 agroforestry system. A companion study at this site estimated that the tree aboveground C 659 stock was 117 ± 21 kg C tree⁻¹ (unpublished data). With 110 trees ha⁻¹, total organic carbon 660 (SOC to 1 m soil depth + aboveground tree C) accumulation rate was 1.11 ± 0.13 Mg C ha⁻¹ 661 yr⁻¹, making agroforestry systems a possible land use to help mitigating climate change (Lal, 662 2004; Lorenz and Lal, 2014). 663

664

665

4.5. A long-term SOC storage?

Most of additional SOC in the agroforestry plot compared to the control plot was located in coarse soil fractions (50-200 μ m and 200-2000 μ m). These soil fractions are assumed to contain labile fractions (Balesdent et al., 1998), that are not stabilized by interaction with clays and thus prone to be decomposed by soil microorganisms. Our site might not be old

enough to observe a difference in the fine soil fractions as changes in the clay fractions are 670 671 often long-term processes (Balesdent, 1996; Balesdent et al., 1988). For example, Takimoto et al., (2008) found in a 35-year-old Faidherbia albida parkland in Mali, that the silt + clay soil 672 673 fraction ($< 53 \mu m$) was enriched in C at depth compared with treeless systems. But on the other hand, Howlett et al., (2011) did not observe any difference for the same soil fraction in a 674 675 80 year-old Dehesa cork oak (*Ouercus suber* L.) silvopasture, but they found that C storage in 676 the macroaggregate fraction (250-2000 mm) was 68% greater underneath versus away from the tree canopy (in 0-25 cm). Several studies have demonstrated that protection of C within 677 the macroaggregate size class was affected by afforestation (Del Galdo et al., 2003; Denef et 678 679 al., 2013) and cessation of tillage (Tan et al., 2007). The fractionation method that was used in this study disrupted macroaggregates, and part of this labile fractions could be located within 680 681 them and therefore be physically protected from decomposition by soil microorganisms (Six 682 et al., 2000). Further work will focus on this aspect in order to estimate the amount of particulate organic matter located in soil aggregates. Calculation of SOC saturation revealed a 683 high deficit of SOC of this soil compared to the theoretical value, suggesting that 684 accumulation of SOC due to the agroforestry system could continue for decades before 685 686 reaching saturation.

687

688 **5.** Conclusion

This study showed the potential of agroforestry systems to increase SOC stocks. However, despite a deep tree rooting system, additional SOC was mainly located in topsoil layers, and in labile organic fractions, making this C storage vulnerable. Tree rows were shown to be a key factor for SOC storage in alley cropping systems. Combining agroforestry systems with no-till or permanent cover systems could be a very efficient way to increase SOC stocks, but more research is needed on this aspect. To fully estimate the impact of agroforestry systems on SOC sequestration, other aspects should be taken into account. For instance, higher SOC
stocks in the inter-rows could increase soil fertility and reduce the need for chemical fertilizer,
contributing indirectly to a reduction of greenhouse gases emissions; further work should
therefore focus on nutrient cycling in these systems.

699

700 Acknowledgments

This study was financed by the French Environment and Energy Management Agency 701 702 (ADEME), following a call for proposals as part of the REACCTIF program (Research on Climate Change Mitigation in Agriculture and Forestry). This work was part of the funded 703 project AGRIPSOL (Agroforestry for Soil Protection), coordinated by Agroof. R. Cardinael 704 was also supported by La Fondation de France. We are very grateful to our colleagues for 705 706 their help with field and laboratory work and logistics, including Daniel Billiou (UPMC), 707 Emmanuel Bourdon (IRD), Jean-François Bourdoncle (INRA), Lydie Dufour (INRA), Claude 708 Hammecker (IRD), Alain Sellier (INRA) and Manon Villeneuve (IRD). We are also grateful to Valérie Viaud (INRA) for her valuable comments concerning the sampling design and 709 710 geostatistics, and to Michael Clairotte (INRA) for his help concerning analyses of VNIR spectra. We also thank all students without whom this work would not have been possible, 711 especially Catalina Gomà Pumarino, Guillermo Lobos Norambuena, and Eric Zassi. 712

713

714

715

717 **References**

- Akaike, H., 1974. A new look at the statistical model identification. IEEE Trans. Automat.
 Contr. 19, 716–723.
- Albrecht, A., Kandji, S.T., 2003. Carbon sequestration in tropical agroforestry systems. Agric.
 Ecosyst. Environ. 99, 15–27.
- Angers, D.A., Arrouays, D., Saby, N.P.A., Walter, C., 2011. Estimating and mapping the
 carbon saturation deficit of French agricultural topsoils. Soil Use Manag. 27, 448–452.
- Balesdent, J., 1996. The significance of organic separates to carbon dynamics and its
 modelling in some cultivated soils. Eur. J. Soil Sci. 47, 485–493.
- Balesdent, J., Balabane, M., 1996. Major contribution of roots to soil carbon storage inferred
 from maize cultivated soils. Soil Biol. Biochem. 28, 1261–1263.
- Balesdent, J., Besnard, E., Arrouays, D., Chenu, C., 1998. The dynamics of carbon in particlesize fractions of soil in a forest-cultivation sequence. Plant Soil 201, 49–57.
- Balesdent, J., Wagner, G.H., Mariotti, A., 1988. Soil organic matter turnover in long-term
 field experiments as revealed by carbon-13 natural abundance. Soil Sci. Soc. Am. J. 52,
 118–124.
- Bambrick, A.D., Whalen, J.K., Bradley, R.L., Cogliastro, A., Gordon, A.M., Olivier, A.,
 Thevathasan, N. V, 2010. Spatial heterogeneity of soil organic carbon in tree-based
 intercropping systems in Quebec and Ontario, Canada. Agrofor. Syst. 79, 343–353.
- Bauhus, J., van der Meer, P., Kanninen, M., 2010. Ecosystem Goods and Services from
 Plantation Forests. Earthscan, London, UK.

738	Bellon-Maurel, V., Fernandez-Ahumada, E., Palagos, B., Roger, JM., McBratney, A., 2010.
739	Critical review of chemometric indicators commonly used for assessing the quality of the
740	prediction of soil attributes by NIR spectroscopy. Trends Anal. Chem. 29, 1073–1081.

741 Bergeron, M., Lacombe, S., Bradley, R.L., Whalen, J., Cogliastro, A., Jutras, M.-F., Arp, P.,

742 2011. Reduced soil nutrient leaching following the establishment of tree-based
743 intercropping systems in eastern Canada. Agrofor. Syst. 83, 321–330.

- Bird, J.A., Torn, M.S., 2006. Fine roots vs. needles: A comparison of 13C and 15N dynamics
 in a ponderosa pine forest soil. Biogeochemistry 79, 361–382.
- Borken, W., Matzner, E., 2009. Reappraisal of drying and wetting effects on C and N
 mineralization and fluxes in soils. Glob. Chang. Biol. 15, 808–824.
- Brown, D.J., Shepherd, K.D., Walsh, M.G., Dewayne Mays, M., Reinsch, T.G., 2006. Global
 soil characterization with VNIR diffuse reflectance spectroscopy. Geoderma 132, 273–
 290.
- Cardinael, R., Mao, Z., Prieto, I., Stokes, A., Dupraz, C., Jourdan, C., 2015. Competition with
 winter crops induces deeper rooting of walnut trees in a Mediterranean alley cropping
 agroforestry system. Plant Soil 391, 219–235.

Chang, C., Laird, D.A., Mausbach, M.J., Hurburgh, C.R., 2001. Near-Infrared Reflectance
Spectroscopy–Principal Components Regression Analyses of Soil Properties. Soil Sci.
Soc. Am. J. 65, 480–490.

Chenu, C., Plante, A.F., 2006. Clay-sized organo-mineral complexes in a cultivation
chronosequence: revisiting the concept of the "primary organo-mineral complex." Eur. J.
Soil Sci. 57, 596–607.

- Clough, Y., Barkmann, J., Juhrbandt, J., Kessler, M., Wanger, T.C., Anshary, A., Buchori, D.,
 Cicuzza, D., Darras, K., Putra, D.D., Erasmi, S., Pitopang, R., Schmidt, C., Schulze,
 C.H., Seidel, D., Steffan-Dewenter, I., Stenchly, K., Vidal, S., Weist, M., Wielgoss,
 A.C., Tscharntke, T., 2011. Combining high biodiversity with high yields in tropical
 agroforests. PNAS 108, 8311–6.
- Conant, R.T., Paustian, K., Elliott, E.T., 2001. Grassland management and conversion into
 grassland: effects on soil carbon. Ecol. Appl. 11, 343–355.
- 767 Conant, R.T., Ryan, M.G., Ågren, G.I., Birge, H.E., Davidson, E.A., Eliasson, P.E., Evans,
- 768 S.E., Frey, S.D., Giardina, C.P., Hopkins, F.M., Hyvönen, R., Kirschbaum, M.U.F.,
- Lavallee, J.M., Leifeld, J., Parton, W.J., Megan Steinweg, J., Wallenstein, M.D., Martin
 Wetterstedt, J.Å., Bradford, M.A., 2011. Temperature and soil organic matter
 decomposition rates synthesis of current knowledge and a way forward. Glob. Chang.
 Biol. 17, 3392–3404.
- Del Galdo, I., Six, J., Peressotti, A., Cotrufo, M.F., 2003. Assessing the impact of land-use
 change on soil C sequestration in agricultural soils by means of organic matter
 fractionation and stable C isotopes. Glob. Chang. Biol. 9, 1204–1213.
- Denef, K., Galdo, I. Del, Venturi, A., Cotrufo, M.F., 2013. Assessment of Soil C and N
 Stocks and Fractions across 11 European Soils under Varying Land Uses. Open J. Soil
 Sci. 3, 297–313.
- Dufour, L., Metay, A., Talbot, G., Dupraz, C., 2013. Assessing Light Competition for Cereal
 Production in Temperate Agroforestry Systems using Experimentation and Crop
 Modelling. J. Agron. Crop Sci. 199, 217–227.

782 J	Dunn, O.J.,	1964. Mult	ple com	parisons	using rank	sums.	Technometrics	6,	241-	-252
-------	-------------	------------	---------	----------	------------	-------	---------------	----	------	------

- Ellert, B.H., Bettany, J.R., 1995. Calculation of organic matter and nutrients stored in soils
 under contrasting management regimes. Can. J. Soil Sci. 75, 529–538.
- Ellert, B.H., Janzen, H.H., Entz, T., 2002. Assessment of a Method to Measure Temporal
 Change in Soil Carbon Storage. Soil Sci. Soc. Am. J. 66, 1687–1695.
- Fontaine, S., Bardoux, G., Abbadie, L., Mariotti, A., 2004. Carbon input to soil may decrease
 soil carbon content. Ecol. Lett. 7, 314–320.
- Fontaine, S., Barot, S., Barré, P., Bdioui, N., Mary, B., Rumpel, C., 2007. Stability of organic
- carbon in deep soil layers controlled by fresh carbon supply. Nature 450, 277–281.
- Gale, W.J., Cambardella, C.A., Bailey, T.B., 2000. Root-Derived Carbon and the Formation
 and Stabilization of Aggregates. Soil Sci. Soc. Am. J. 64, 201.
- 793 Gavinelli, E., Feller, C., Larré-Larrouy, M., Bacye, B., Djegui, N., Nzila, J. de D., 1995. A
- routine method to study soil organic matter by particle-size fractionation: examples for
 tropical soils. Commun. Soil Sci. Plant Anal. 26, 1749–1760.
- Germon, A., Cardinael, R., Prieto, I., Mao, Z., Kim, J.H., Stokes, A., Dupraz, C., Laclau, J.-
- P., Jourdan, C. Unexpected phenology and lifespan of shallow and deep fine roots ofwalnut trees grown in a Mediterranean agroforestry system (submitted).
- Gras, J.-P., Barthès, B.G., Mahaut, B., Trupin, S., 2014. Best practices for obtaining and
 processing field visible and near infrared (VNIR) spectra of topsoils. Geoderma 214-215,
 126–134.

- Haile, S.G., Nair, V.D., Nair, P.K.R., 2010. Contribution of trees to carbon storage in soils of
 silvopastoral systems in Florida, USA. Glob. Chang. Biol. 16, 427–438.
- Hamdi, S., Moyano, F., Sall, S., Bernoux, M., Chevallier, T., 2013. Synthesis analysis of the
 temperature sensitivity of soil respiration from laboratory studies in relation to
 incubation methods and soil conditions. Soil Biol. Biochem. 58, 115–126.
- Harper, R.J., Tibbett, M., 2013. The hidden organic carbon in deep mineral soils. Plant Soil
 368, 641–648.
- Harris, D., Horwath, W.R., Van Kessel, C., 2001. Acid fumigation of soils to remove
 carbonates prior to total organic carbon or carbon-13 isotopic analysis. Soil Sci. Soc.
 Am. J. 65, 1853–1856.
- Hassink, J., 1997. The capacity of soils to preserve organic C and N by their association with
 clay and silt particles. Plant Soil 191, 77–87.
- Hothorn, T., Bretz, F., Westfall, P., 2008. Simultaneous Inference in General Parametric
 Models. Biometrical J. 50, 346–363.
- Howlett, D.S., Moreno, G., Mosquera Losada, M.R., Nair, P.K.R., Nair, V.D., 2011. Soil
 carbon storage as influenced by tree cover in the Dehesa cork oak silvopasture of centralwestern Spain. J. Environ. Monit. 13, 1897–904.
- IUSS Working Group WRB, 2007. World Reference Base for Soil Resources 2006, first
 update 2007. World Soil Resources Reports No. 103. FAO, Rome.
- Jobbagy, E.G., Jackson, R.B., 2000. The vertical distribution of soil organic carbon and its
 relation to climate and vegetation. Ecol. Appl. 10, 423–436.

Jordan, C.F., 2004. Organic farming and agroforestry: Alleycropping for mulch production
for organic farms of southeastern United States. Agrofor. Syst. 61-62, 79–90.

Junior, C.C., Corbeels, M., Bernoux, M., Piccolo, M.C., Neto, M.S., Feigl, B.J., Cerri, C.E.P.,
Cerri, C.C., Scopel, E., Lal, R., 2013. Assessing soil carbon storage rates under notillage: Comparing the synchronic and diachronic approaches. Soil Tillage Res. 134,
207–212.

- Kennard, R.W., Stone, L.A., 1969. Computer aided design of experiments. Technometrics 11,
 137–148.
- Kruskal, W.H., Wallis, W.A., 1952. Use of Ranks in One-Criterion Variance Analysis. J. Am.
 Stat. Assoc. 47, 583–621.
- Lal, R., 2004. Soil carbon sequestration impacts on global climate change and food security.
 Science (80-.). 304, 1623–7.
- Lal, R., 2004. Soil carbon sequestration to mitigate climate change. Geoderma 123, 1–22.
- Lark, R.M., Cullis, B.R., Welham, S.J., 2006. On spatial prediction of soil properties in the
 presence of a spatial trend: the empirical best linear unbiased predictor (E-BLUP) with
 REML. Eur. J. Soil Sci. 57, 787–799.
- Lipper, L., Thornton, P., Campbell, B.M., Baedeker, T., Braimoh, A., Bwalya, M., Caron, P.,
- 840 Cattaneo, A., Garrity, D., Henry, K., Hottle, R., Jackson, L., Jarvis, A., Kossam, F.,
- 841 Mann, W., McCarthy, N., Meybeck, A., Neufeldt, H., Remington, T., Sen, P.T., Sessa,
- 842 R., Shula, R., Tibu, A., Torquebiau, E.F., 2014. Climate-smart agriculture for food
- security. Nat. Clim. Chang. 4, 1068–1071.

- Lorenz, K., Lal, R., 2014. Soil organic carbon sequestration in agroforestry systems. A
 review. Agron. Sustain. Dev. 34, 443–454.
- Luo, Z., Wang, E., Sun, O.J., 2010. Can no-tillage stimulate carbon sequestration in
 agricultural soils? A meta-analysis of paired experiments. Agric. Ecosyst. Environ. 139,
 224–231.
- 849 Martens, H., Naes, T., 1989. Multivariate calibration. John Wiley & Sons, Ltd, Chichester.
- 850 Martin, M.P., Wattenbach, M., Smith, P., Meersmans, J., Jolivet, C., Boulonne, L., Arrouays,
- D., 2011. Spatial distribution of soil organic carbon stocks in France. Biogeosciences 8,
 1053–1065.
- Millennium Ecosystem Assessment, 2005. Ecosystems and Human Well-being: Synthesis.
 Island Press, Washington, DC.
- Mulia, R., Dupraz, C., 2006. Unusual fine root distributions of two deciduous tree species in
 southern France: What consequences for modelling of tree root dynamics? Plant Soil
 281, 71–85.
- Muñoz-Rojas, M., Jordán, A., Zavala, L.M., De la Rosa, D., Abd-Elmabod, S.K., AnayaRomero, M., 2012. Organic carbon stocks in Mediterranean soil types under different
 land uses (Southern Spain). Solid Earth 3, 375–386.
- Nair, P.K.R., 2012. Carbon sequestration studies in agroforestry systems: a reality-check.
 Agrofor. Syst. 86, 243–253.
- Nair, P.K.R., Nair, V.D., Kumar, B.M., Showalter, J.M., 2010. Carbon sequestration in
 agroforestry systems, in: Advances in Agronomy. pp. 237–307.

- Oades, J., 1995. An overview of processes affecting the cycling of organic carbon in soils, in:
 Zepp, R.G., Sonntag, C. (Eds.), Role of Non-Living Organic Matter in the Earth's
 Carbon Cycle. John Wiley, pp. 293–303.
- Oelbermann, M., Voroney, R.P., 2007. Carbon and nitrogen in a temperate agroforestry
 system: Using stable isotopes as a tool to understand soil dynamics. Ecol. Eng. 29, 342–
 349.
- Oelbermann, M., Voroney, R.P., Gordon, A.M., 2004. Carbon sequestration in tropical and
 temperate agroforestry systems: a review with examples from Costa Rica and southern
 Canada. Agric. Ecosyst. Environ. 104, 359–377.
- Olson, K.R., Al-Kaisi, M., Lal, R., Lowery, B., 2014. Examining the paired comparison
 method approach for determining soil organic carbon sequestration rates. J. Soil Water
 Conserv. 69, 193A–197A.
- Pandey, D.N., 2002. Carbon sequestration in agroforestry systems. Clim. Policy 2, 367–377.
- Peichl, M., Thevathasan, N. V, Gordon, A.M., Huss, J., Abohassan, R.A., 2006. Carbon
 sequestration potentials in temperate tree-based intercropping systems, southern Ontario,
 Canada. Agrofor. Syst. 66, 243–257.
- 881 Pellerin, S., Bamière, L., Angers, D., Béline, F., Benoît, M., Butault, J.P., Chenu, C.,
- Colnenne-David, C., De Cara, S., Delame, N., Doreau, M., Dupraz, P., Faverdin, P.,
 Garcia-Launay, F., Hassouna, M., Hénault, C., Jeuffroy, M., Klumpp, K., Metay, A.,
 Moran, D., Recous, S., Samson, E., Savini, I., Pardon, L., 2013. How can French
 agriculture contribute to reducing greenhouse gas emissions? Abatement potential and
 cost of ten technical measures. Synopsis of the study report, INRA (France).

887	Philippot, L., Čuhel, J., Saby, N.P.A., Chèneby, D., Chroňáková, A., Bru, D., Arrouays, D.,
888	Martin-Laurent, F., Šimek, M., 2009. Mapping field-scale spatial patterns of size and
889	activity of the denitrifier community. Environ. Microbiol. 11, 1518–1526.

- Pinheiro, J., Bates, D., DebRoy, S., Sarkar, D., R Development Core Team, 2013. nlme:
 Linear and Nonlinear Mixed Effects Models. R package version 3.1-111.
- Pinheiro, J.C., Bates, D.M., 2000. Mixed-Effects Models in S and S-PLUS. Springer Science

893 & Business Media.

Poeplau, C., Don, A., 2015. Carbon sequestration in agricultural soils via cultivation of cover

895 crops – A meta-analysis. Agric. Ecosyst. Environ. 200, 33–41.

- Power, A.G., 2010. Ecosystem services and agriculture: tradeoffs and synergies. Philos.
 Trans. R. Soc. Lond. B. Biol. Sci. 365, 2959–2971.
- 898 Profft, I., Mund, M., Weber, G.-E., Weller, E., Schulze, E.-D., 2009. Forest management and

carbon sequestration in wood products. Eur. J. For. Res. 128, 399–413.

- Puget, P., Chenu, C., Balesdent, J., 2000. Dynamics of soil organic matter associated with
 particle-size fractions of water-stable aggregates. Eur. J. Soil Sci. 51, 595–605.
- 902 R Development Core Team, 2013. R: A language and environment for statistical computing.
- 903 Rasse, D.P., Rumpel, C., Dignac, M.F., 2005. Is soil carbon mostly root carbon? Mechanisms
 904 for a specific stabilisation. Plant Soil 269, 341–356.
- Rhoades, C.C., 1997. Single-tree influences on soil properties in agroforestry: lessons from
 natural forest and savanna ecosystems. Agrofor. Syst. 35, 71–94.

- 907 Ribeiro, P.J., Diggle, P.J., 2001. geoR: A package for geostatistical analysis. R-News 1, 15–
 908 18.
- Schroth, G., da Fonseca, G.A.B., Harvey, C.A., Gascon, C., Vasconcelos, H.L., Izac, A.M.N., 2004. Agroforestry and biodiversity conservation in tropical landscapes. Island
 Press, Washington, DC.
- Sharrow, S.H., Ismail, S., 2004. Carbon and nitrogen storage in agroforests, tree plantations,
 and pastures in western Oregon, USA. Agrofor. Syst. 60, 123–130.
- 914 Six, J., Elliott, E.T., Paustian, K., 2000. Soil macroaggregate turnover and microaggregate
 915 formation: a mechanism for C sequestration under no-tillage agriculture. Soil Biol.
 916 Biochem. 32, 2099–2103.
- Six, J., Elliott, E.T., Paustian, K., Doran, J.W., 1998. Aggregation and soil organic matter
 accumulation in cultivated and native grassland soils. Soil Sci. Soc. Am. J. 62, 1367–
 1377.
- 920 Somarriba, E., 1992. Revisiting the past: an essay on agroforestry definition. Agrofor. Syst.
 921 19, 233–240.
- Somarriba, E., Cerda, R., Orozco, L., Cifuentes, M., Dávila, H., Espin, T., Mavisoy, H.,
 Ávila, G., Alvarado, E., Poveda, V., Astorga, C., Say, E., Deheuvels, O., 2013. Carbon
 stocks and cocoa yields in agroforestry systems of Central America. Agric. Ecosyst.
 Environ. 173, 46–57.
- Soussana, J.-F., Loiseau, P., Vuichard, N., Ceschia, E., Balesdent, J., Chevallier, T., Arrouays,
 D., 2004. Carbon cycling and sequestration opportunities in temperate grasslands. Soil
- 928 Use Manag. 20, 219–230.

- Stavi, I., Lal, R., 2013. Agroforestry and biochar to offset climate change: a review. Agron.
 Sustain. Dev. 33, 81–96.
- Stevens, A., Nocita, M., Tóth, G., Montanarella, L., van Wesemael, B., 2013. Prediction of
 Soil Organic Carbon at the European Scale by Visible and Near InfraRed Reflectance
 Spectroscopy. PLoS One 8, 1–13.
- Takimoto, A., Nair, V.D., Nair, P.K.R., 2008. Contribution of trees to soil carbon
 sequestration under agroforestry systems in the West African Sahel. Agrofor. Syst. 76,
 11–25.
- Tan, Z., Lal, R., Owens, L., Izaurralde, R., 2007. Distribution of light and heavy fractions of
 soil organic carbon as related to land use and tillage practice. Soil Tillage Res. 92, 53–
 59.
- Teck, R.M., Hilt, D.E., 1991. Individual-Tree Diameter Growth Model for the Northeastern
 United States. Res. Pap. NE-649. Radnor, PA: US. Department of Agriculture, Forest
 Service, Northeastern Forest Experiment Station. 11 p.
- 943 Torquebiau, E.F., 2000. A renewed perspective on agroforestry concepts and classification.
 944 Life Sci. 323, 1009–1017.
- Tully, K.L., Lawrence, D., Scanlon, T.M., 2012. More trees less loss: Nitrogen leaching
 losses decrease with increasing biomass in coffee agroforests. Agric. Ecosyst. Environ.
 161, 137–144.
- 948 Upson, M.A., Burgess, P.J., 2013. Soil organic carbon and root distribution in a temperate
 949 arable agroforestry system. Plant Soil 373, 43–58.

- Varah, A., Jones, H., Smith, J., Potts, S.G., 2013. Enhanced biodiversity and pollination in
 UK agroforestry systems. J. Sci. Food Agric. 93, 2073–5.
- Verchot, L. V., Noordwijk, M., Kandji, S., Tomich, T., Ong, C., Albrecht, A., Mackensen, J.,
 Bantilan, C., Anupama, K. V., Palm, C., 2007. Climate change: linking adaptation and
 mitigation through agroforestry. Mitig. Adapt. Strateg. Glob. Chang. 12, 901–918.
- Villanneau, E.J., Saby, N.P.A., Marchant, B.P., Jolivet, C.C., Boulonne, L., Caria, G.,
 Barriuso, E., Bispo, A., Briand, O., Arrouays, D., 2011. Which persistent organic
 pollutants can we map in soil using a large spacing systematic soil monitoring design? A
 case study in Northern France. Sci. Total Environ. 409, 3719–3731.
- Virto, I., Barré, P., Burlot, A., Chenu, C., 2011. Carbon input differences as the main factor
 explaining the variability in soil organic C storage in no-tilled compared to inversion
 tilled agrosystems. Biogeochemistry 108, 17–26.
- 962 Von Lützow, M., Kögel-Knabner, I., Ekschmitt, K., Flessa, H., Guggenberger, G., Matzner,
- E., Marschner, B., 2007. SOM fractionation methods: Relevance to functional pools and
 to stabilization mechanisms. Soil Biol. Biochem. 39, 2183–2207.
- Webster, R., McBratney, A.B., 1989. On the Akaike Information Criterion for choosing
 models for variograms of soil properties. J. Soil Sci. 40, 493–496.
- 967 Webster, R., Oliver, M.A., 2007. Geostatistics for Environmental Scientists.
- Wiesmeier, M., Hübner, R., Spörlein, P., Geuß, U., Hangen, E., Reischl, A., Schilling, B., von
 Lützow, M., Kögel-Knabner, I., 2014. Carbon sequestration potential of soils in
 southeast Germany derived from stable soil organic carbon saturation. Glob. Chang.
 Biol. 20, 653–665.

972	Young,	А.,	1997.	Agroforestry	for	Soil	Management,	Second.	ed.	CAB	International,
973	Wa	alling	ford, U	JK.							
974											
975											
976											
977											
978											
979											
980											
981											
982											
983											
984											
985											
986											
987											
988											
989											
990											

991 Supplementary materials

Table S1. Soil organic carbon stocks (Mg C ha⁻¹) and SOC accumulation rates (kg C ha⁻¹ yr⁻¹)
without the equivalent soil mass (ESM) correction. Associated errors are standard errors (100
replicates for the agroforestry plot, 93 for the control plot).

		Cumulated SOC stor	cks (Mg C ha ⁻¹)	Δ SOC stocks (Mg C ha ⁻¹)	SOC accumulation rates (kg C ha ⁻¹ yr ⁻¹)
	Soil depth	Agroforestry	Control	Δ (Agroforestry –	Agroforestry vs
	(cm)			Control)	Control
	0-10	13.9 ± 0.3	13.2 ± 0.1	0.7 ± 0.4	41 ± 20
	0-30	44.5 ± 0.5	41.9 ± 0.2	2.5 ± 0.6	139 ± 32
	0-50	67.9 ± 0.7	65.5 ± 0.4	2.5 ± 0.8	137 ± 42
	0-70	88.8 ± 0.7	86.3 ± 0.4	2.6 ± 0.8	143 ± 46
	0-100	121.2 ± 0.7	118.0 ± 0.5	3.1 ± 0.9	173 ± 54
	0-120	140.9 ± 0.8	138.4 ± 0.6	2.5 ± 1.0	138 ± 56
	0-140	161.4 ± 0.8	159.4 ± 0.6	1.9 ± 1.0	106 ± 58
	0-160	181.5 ± 0.8	180.5 ± 0.6	0.9 ± 1.0	53 ± 60
	0-180	199.5 ± 0.9	199.3 ± 0.6	0.2 ± 1.1	11 ± 60
	0-200	214.6 ± 0.9	214.5 ± 0.6	0.1 ± 1.1	7 ± 62
995					
996					
997					
998					
999					
1000					
1001					
1001					
1002					
1003					
1005					





1005 Figure S1. Measured and cross-validation predicted values of soil organic carbon1006 concentrations for the topsoil and subsoil models.

Fractionnated soil samples



¹⁰¹⁷

Figure S2. Carbon concentration of bulk fractionated samples. Error bars represent standard
errors (n=6 in the control, n=5 in the inter-row and in the tree row).



Figure S3. Carbon concentration of each soil fraction. Error bars represent standard errors (n=6 in the control, n=5 in the inter-row and in the tree row). OF = Organic fraction, F = organo-mineral fraction. Means followed by the same letters do not differ significantly at p=0.017 (Dunn's test with Bonferroni correction).