

Permian-Triassic Osteichthyes (bony fishes): diversity dynamics and body size evolution.

Carlo Romano, Martha B. Koot, Ilja Kogan, Arnaud Brayard, Alla V. Minikh,

Winand Brinkmann, Hugo Bucher, Jürgen Kriwet

► To cite this version:

Carlo Romano, Martha B. Koot, Ilja Kogan, Arnaud Brayard, Alla V. Minikh, et al.. Permian-Triassic Osteichthyes (bony fishes): diversity dynamics and body size evolution.. Biological Reviews, 2016, 91 (1), pp.106-147. 10.1111/brv.12161 . hal-01253154

HAL Id: hal-01253154 https://hal.science/hal-01253154

Submitted on 8 Feb 2024

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1	Permian-Triassic Osteichthyes (bony fishes): Diversity dynamics and body size evolution
2	
3	Carlo Romano ^{1*} , Martha B. Koot ² , Ilja Kogan ³ , Arnaud Brayard ⁴ , Alla V. Minikh ⁵ , Winand
4	Brinkmann ¹ , Hugo Bucher ^{1,6} and Jürgen Kriwet ⁷
5	
6	*corresponding author: carlo.romano@pim.uzh.ch, phone: +41 44 634 23 47
7	
8	¹ Palaeontological Institute and Museum, University of Zurich, Karl Schmid-Strasse 4, 8006
9	Zurich, Switzerland
10	² School of Geography, Earth and Environmental Sciences (Faculty of Science and
11	Technology), Plymouth University, Fitzroy Building, Drake Circus, Plymouth, Devon, PL4
12	8AA, England; Current address: 25 Honicknowle Lane, Plymouth, PL2 3QS, United
13	Kingdom
14	³ Department of Palaeontology, Geological Institute, TU Bergakademie Freiberg, Bernhard-
15	von-Cotta-Strasse 2, 09596 Freiberg, Germany
16	⁴ UMR CNRS 6282 Biogéosciences, Université de Bourgogne, 6 Boulevard Gabriel, F-21000,
17	Dijon, France
18	⁵ Saratov State University, 83 Astrakhanskaya Street, Saratov 410012, Russia
19	⁶ Department of Earth Sciences, ETH Zurich, Sonneggstrasse 5, 8092 Zurich, Switzerland
20	⁷ Department of Palaeontology, University of Vienna, Geozentrum, Althanstrasse 14, 1090
21	Vienna, Austria
22	
23	ABSTRACT
24	
25	The Permian and Triassic were key time intervals in the history of life on Earth. Both

26 periods are marked by a series of biotic crises including the most catastrophic of such events,

1 the end-Permian mass extinction, which eventually led to a major turnover from typical 2 Palaeozoic faunas and floras to those that are emblematic for the Mesozoic and Cenozoic. The 3 aim of this review is to examine trends in Permian and Triassic osteichthyans (Dipnoi, 4 Actinistia, Actinopterygii) in order to elucidate their response to important biotic events 5 during these periods. Our analyses are based on an updated data matrix adapted from primary 6 literature. We investigate different metrics, including taxonomic diversity and body size. 7 Diversity is measured separately for marine and freshwater bony fishes but also on global 8 scale (total diversity), across palaeolatitudinal belts and within different palaeogeographical 9 provinces. In addition, we examine body size to infer changes in trophic position. Our results 10 suggest a general trend from low osteichthyan diversity in the Permian to higher levels in the 11 Triassic. Diversity dynamics in the Permian are marked by a decline in freshwater taxa during 12 the Cisuralian, followed by a slight rise in global osteichthyan diversity towards the end of the 13 Permian. An extinction event during the end-Guadalupian crisis is not evident from our data. 14 However, basal actinopterygians ('palaeopterygians') experienced a significant body size 15 increase across the Guadalupian-Lopingian boundary and these fishes upheld their position as 16 large, top predators from the Late Permian to the Late Triassic. Smaller osteichthyans at lower 17 trophic levels mainly comprised neopterygians and 'subholosteans'. The effects of the end-18 Permian mass extinction on bony fishes remain elusive due to taxonomic difficulties, however 19 two distinct diversification events are noted in the wake of this biotic crisis, a first one during 20 the Early Triassic (dipnoans, actinistians, 'palaeopterygians', 'subholosteans') and a second 21 one during the Middle Triassic ('subholosteans', neopterygians) – the latter event mediating a 22 significant global decrease in osteichthyan body size. Neopterygii, the clade that encompasses 23 the vast majority of extant fishes, experienced another radiation event in the Late Triassic. 24 The Triassic diversification of Osteichthyes, predominantly of Actinopterygii, which only 25 occurred after severe extinctions among Chondrichthyes during the Middle-Late Permian, 26 resulted in a profound change within global fish communities, from the chondrichthyan-rich

1	faunas of the Permo-Carboniferous to the typical Mesozoic and Cenozoic associations	5
2	dominated by actinopterygians. This ichthyofaunal turnover was not sudden but follow	wed a
3	stepwise pattern over tens of millions of years, with leaps during extinction events.	
4		
5	Key words: Dipnoi, Actinistia, Actinopterygii, Osteichthyes, mass extinction, biotic re	ecovery,
6	Permian-Triassic boundary, diversity, body size	
7		
8	CONTENTS	
9	I. Introduction	4
10	II. Material and methods	6
11	(1) Dataset	6
12	(2) Diversity analyses	9
13	(3) Body size analyses	
14	(4) Quality of the osteichthyan fossil record	11
15	(5) Lagerstätten effect	
16	(6) First and last appearance datums	14
17	III. Results	15
18	(1) Estimation of sampling bias and Lagerstätten effect	15
19	(2) General trends: Genus diversity, turnovers and turnover rates	17
20	(3) Diversity dynamics within Osteichthyes	
21	(4) Palaeolatitudinal diversity	
22	(5) Body size	
23	IV. Discussion	
24	(1) The Permian record	
25	<i>(a)</i> "Olson's gap"	
26	(b) The end-Guadalupian extinction event	
27	(c) The end-Permian mass extinction event	
28	(2) The Triassic record	
29	(a) Early Triassic recovery	
30	(b) The Triassic actinopterygian revolution	
31	(c) "Carnian Pluvial Event"	
32	(3) Permian-Triassic turnover of fish communities	
33	V. Conclusions	
34	VI. Acknowledgements	
35	VII. References	
36	VIII. Supporting Information	
37		

39 I. INTRODUCTION

1 Fishes (non-tetrapod vertebrates) comprise more than half of today's global vertebrate 2 diversity and they are important predators in aquatic ecosystems. Marine and freshwater 3 ichthyofaunas have been dominated by teleosteans since the later part of the Mesozoic, with 4 presently >28'000 species, followed by chondrichthyans (sharks, rays, ratfishes) as the second 5 largest clade with >1'100 extant species (Arratia, 2004; Nelson, 2006; Sallan, 2014). Other 6 groups, such as non-teleostean ray-finned fishes (bichirs, reedfishes, sturgeons, bowfin, gars), 7 lobe-finned fishes (lungfishes, 'coelacanths'), and 'Cyclostomata' (hagfishes, lampreys), 8 today constitute only a minor component, whereas other groups are only known as fossils 9 (Janvier, 1996, 2007; Nelson, 2006; Friedman & Sallan, 2012; Sallan, 2014). The diversity of 10 fishes has fluctuated in the past and groups that are rare or extinct today flourished at different 11 times (Thomson, 1977; McCune & Schaeffer, 1986; Patterson, 1994; Schultze, 2004; Klug et 12 al., 2010; Sallan & Coates, 2010; Anderson et al., 2011; Friedman & Sallan, 2012; Tintori et 13 al., 2013). Questions concerning which factors led to the observed diversity patterns in fishes 14 and to what extent mass extinctions played a role have become increasingly important in 15 modern palaeoichthyology (e.g. McCune & Schaeffer, 1986; Patterson & Smith, 1987; 16 Benton, 1998; Cavin, 2002; Kriwet & Benton, 2004; Cavin, Forey & Lécuyer, 2007; Sallan & 17 Coates, 2010; Friedman & Sallan, 2012; Lloyd & Friedman, 2013; Sallan, 2014). 18 A critical time for both marine and continental life on Earth was the Permian-Triassic 19 interval (ca. 298.9–201.3 myr ago; Gradstein et al., 2012), known for a series of extinction 20 events, amongst others the most devastating of all time: the end-Permian mass extinction 21 (~252.17 myr ago; e.g. Raup, 1978; Sepkoski, 1984; Benton, 2003; Erwin, 2006; Shen et al., 22 2011). A few studies have already focused on diversity patterns of bony fishes (Osteichthyes) 23 or fishes in general in relation to the end-Permian mass extinction and most of them arrived at 24 the conclusion that these groups were only minorly impacted by this event (Schaeffer, 1973; 25 Thomson, 1976, 1977; Patterson & Smith, 1987; Benton, 1998; Reguant, 2007; Blieck, 2011). 26 Nevertheless, possible crises have been suggested for certain subsets of fishes, such as

1	durophagous forms (Friedman & Sallan, 2012) or marine taxa (Pitrat, 1973). Little is known
2	about the response of fishes to other extinction events of the Permian and Triassic, such as the
3	end-Guadalupian (Groves & Wang, 2013) and end-Smithian crises (Galfetti et al., 2007a, b).
4	Moreover, previous studies were only based on anecdotal evidence or primarily on data from
5	old compendia, such as Benton (1993) or Sepkoski (2002). Note that the database of Sepkoski
6	(2002) was last updated in 1998 and his analyses were not specifically aimed at fishes. Hence,
7	earlier studies lack the data compiled over the last 15 to 20 years, including discoveries and
8	revisions of taxa and refinements in biochronostratigraphy. Finally, most previous studies
9	have only looked at taxonomic changes but not at other metrics.
10	Here we present a literature review and state-of-the-art study of the evolutionary
11	dynamics of the three dominant groups of Osteichthyes during the Permian and Triassic:
12	Actinopterygii (ray-finned fishes), Actinistia ('coelacanths') and Dipnoi (lungfishes). Trends
13	within Permian-Triassic Chondrichthyes (cartilaginous fishes) are examined elsewhere (Koot,
14	2013). Our analyses rely on an updated data matrix based on primary literature (see
15	Supplementary Tables A1–A3) and focus on changes in taxonomic diversity among various
16	osteichthyan groups, within different environments (marine, freshwater, global), and across
17	palaeolatitudes, but also on fluctuations in body size and accompanied implications for
18	aquatic trophic networks. The results are discussed primarily in the context of different biotic
19	crises and recoveries of the Permian-Triassic and the evolutionary dynamics of fishes are
20	compared to those of other taxa. The goal of this paper is to review the current state of
21	research and to highlight gaps in our knowledge as a guide for future research.
22	
23	II. MATERIAL AND METHODS
24	
25	(1) Dataset
26	

1	This study is based on the more diverse osteichthyan groups of the Permian-Triassic,
2	namely Actinopterygii, Actinistia, and Dipnoi, which have been the dominant clades among
3	bony fishes since the Early Carboniferous and also include extant representatives (e.g.,
4	Nelson, 2006; Sallan & Coates, 2010; Friedman & Sallan, 2012). Chondrichthyes have been
5	studied separately (Koot, 2013). Conodonts, whose vertebrate affinities are still debated (see
6	Turner et al., 2010), were studied by Orchard (2007). Other, rarer groups that lived during the
7	Permian-Triassic interval, such as 'Cyclostomata' (Janvier, 1996; Kuraku & Kuratani, 2006),
8	'Acanthodii' (early gnathostomes; Mutter & Richter, 2007), and megalichthyid and
9	rhizodopsid sarcopterygians (Schultze & Heidtke, 1986; Witzmann & Schoch, 2012), are
10	excluded because of insufficient data for sound analyses. However, all groups of fishes, both
11	cartilaginous and bony clades, are taken into account in the discussion section.
12	Osteichthyes are divided into two major subclades: Sarcopterygii (lobe-finned fishes)
13	and Actinopterygii (ray-finned fishes). Sarcopterygians comprise the Dipnoi and Actinistia,
14	but also the Tetrapodomorpha (e.g. the Late Palaeozoic rhizodopsids and megalichthyids;
15	Janvier, 2007). Permian-Triassic actinopterygians are subdivided into the monophyletic
16	Neopterygii, which include Teleosteomorpha (Arratia, 2004) and Holostei (to this group we
17	herein refer all non-teleosteomorph neopterygians, see also Grande, 2010 and López-
18	Arbarello, 2012), as well as a paraphyletic group of non-neopterygian actinopterygians coined
19	'Palaeopterygii' by Regan (1923). Many predominantly Triassic 'palaeopterygians' exhibit a
20	mosaic pattern of neopterygian characters (e.g. Brough, 1936; Schaeffer 1956). However,
21	these taxa have traditionally been excluded from Neopterygii and the term 'Subholostei' was
22	established for them by Brough (1936). Their similarities with neopterygians are mostly
23	considered the result of convergent evolution (Brough, 1936; Schaeffer, 1956).
24	Although some Permian-Triassic bony fishes have already been included in cladistic
25	analyses (e.g. Lund, Poplin & McCarthy, 1995; Forey, 1998; Coates, 1999; Schultze, 2004;
26	Gardiner, Schaeffer & Masserie, 2005; Poplin & Dutheil, 2005; López-Arbarello, 2012;

1 Benton et al., 2013; Cavin, Furrer & Obrist, 2013), the phylogenetic position of many taxa is 2 still not well known. The taxonomic framework at family and order level is poorly resolved for many osteichthyans, specifically for 'palaeopterygians' and 'subholosteans' (McCune & 3 4 Schaeffer, 1986; Patterson & Smith, 1987; Sallan, 2014). For these reasons, we refrain from 5 analysing osteichthyan diversity at family or order level and instead use the classical higher 6 groupings, i.e. Dipnoi, Actinistia, 'Palaeopterygii' (), 'Subholostei', Holostei, and 7 Teleosteomorpha. We largely adopt Brough's (1936) concept of 'Subholostei' but exclude 8 Saurichthyidae due to a lack of neopterygian-like traits (note that the sole character mentioned 9 by Brough, a vertical suspensorium, is absent in saurichthyids, cf. Stensiö, 1925). 10 For our analyses, we compiled a new database (last update: December 2013) and we 11 mostly follow the latest taxonomic interpretations (Supplementary Tables A1–A3). Our 12 database, resolved at species level (including species in open nomenclature), contains 13 information on geographical and stratigraphical occurrences as well as body size. Only taxa 14 whose stratigraphic occurrence can be dated at least at the epoch level are incorporated. 15 Occurrences of taxa represented by isolated skeletal elements (e.g. skull bones or scales) are 16 only included in the database if the remains can be identified with reasonable certainty at 17 genus level (e.g. jaw fragments of Saurichthys). Nominal taxa based on material we deem 18 non-diagnostic are omitted. Questionable occurrences are indicated by a '?' or genera listed in 19 inverted commas in Supplementary Tables A1-A3. 20 Our database (see Supplementary Tables A1-A3; nomina dubia/nuda omitted) includes 21 a total of 697 named species (59 Dipnoi, 43 Actinistia, 595 Actinopterygii) belonging to 363 22 nominal genera (18 Dipnoi, 32 Actinistia, 313 Actinopterygii). Marine and freshwater 23 occurrences are treated as separate datasets. Assignment of occurrences as either marine or

24 freshwater mostly adheres to the interpretation of previous authors. Taxa from brackish water

25 environments are included in the marine dataset. All lungfishes are incorporated in the non-

26 marine dataset (note, however, that some dipnoans may have also dwelled in marine habitats;

1 e.g. Schultze, 2004). For the Permian-Triassic, we identified 212 marine genera (28

2 Actinistia, 184 Actinopterygii) and 172 freshwater genera (18 Dipnoi, 6 Actinistia, 148

Actinopterygii). Furthermore, 20 bony fish genera (2 Actinistia, 18 Actinopterygii) have been reported from both marine and freshwater deposits and can thus be considered euryhaline. For estimation of diversity on a global scale (i.e. marine plus freshwater combined), euryhaline taxa were only counted once per interval. For comparison, the global average number of genera per each Permian-Triassic epoch corresponds to ca. 0.5–1% of all extant osteichthyan

8 genera (~4'300; Nelson, 2006).

9 The Permian and Triassic are divided into three epochs each (Early, Middle, Late). 10 Permian epochs were originally named after geographically separated outcrop areas in Russia 11 (Cisuralian = Early Permian), the United States (Guadalupian = Middle Permian), and China 12 (Lopingian = Late Permian). The Cisuralian includes four stages (Asselian, Sakmarian, 13 Artinskian, Kungurian), the Guadalupian three (Roadian, Wordian, Capitanian), and the 14 Lopingian two (Wuchiapingian, Changhsingian). The Early Triassic is officially subdivided into two stages (Induan, Olenekian; Kiparisova & Popov, 1956; Gradstein et al., 2012) but 15 16 more commonly into four substages (Tozer, 1965) well defined by global faunal events 17 (Griesbachian, Dienerian, Smithian, Spathian). The Middle Triassic comprises two stages 18 (Anisian, Ladinian), and the Late Triassic three (Carnian, Norian, Rhaetian).. Occurrences of 19 bony fishes are known from all stages of the Permian-Triassic interval (and substages of the 20 Early Triassic) and from all continents (Fig. 1a), with a focus on Europe due to the much longer research history, starting with the dawn of palaeoichthyology in the early 19th century 21 22 (e.g. Agassiz, 1833–43).

23

24 (2). Diversity analyses

1 Diversity analyses herein are based on occurrences of genera (Supplementary Tables 2 A1-A3). Diversity of bony fishes is measured in terms of genus richness (counts of genera per bin), boundary crossers (number of genera surviving an interval boundary), and mean 3 4 standing diversity. Mean standing diversity was calculated by counting first and last 5 occurrences of a given genus as 0.5 (assuming that taxa persisted, on average, for about half 6 of the duration of the origination or extinction interval, respectively) and range-through 7 occurrences as 1, whereas singletons (genera occurring in only one interval) are weighted as 8 0. Lazarus occurrences (virtual occurrences between known occurrences) were incorporated 9 for calculation of genus richness and boundary crossers. However, Lazarus occurrences 10 between intervals of questionable occurrence were omitted from the analyses. Numbers of 11 originations and extinctions were calculated, as well as per taxon origination and extinction 12 rates (counts of originations/extinctions divided by genus richness). Questionable occurrences 13 of taxa were ignored for these calculations. 14 For the assessment of palaeolatitudinal diversity, occurrences of bony fishes were 15 grouped into palaeolatitudinal belts (according to the palaeogeographic position of their find

locality, Fig. 1a) and equivalent belts of the northern and southern palaeohemisphere were
combined as follows: (1) low-palaeolatitudinal zone (30°N to 30°S), (2) mid-palaeolatitudinal
zone (31°N to 60°N and 31°S to 60°S), and (3) high-palaeolatitudinal zone (61°N to 90°N and
61°S to 90°S). Palaeolatitudinal diversity (marine, freshwater, global) is measured at epoch
level. Most Permian-Triassic bony fish localities in the northern palaeohemisphere were
deposited under marine conditions, whereas those from the southern palaeohemisphere
mainly represent continental deposits (Allison & Briggs, 1993; Fig. 1a).

23

24 (3) Body size analyses

1 In addition to diversity, we also examined variations in osteichthyan body size during 2 the Permian-Triassic. For this purpose, we compiled data on maximum standard lengths 3 (MSL) at species level (inclusive of some species in open nomenclature, Supplementary 4 Tables A1–A3). MSL of species only known from partial skeletons was estimated by 5 comparison with complete material of other species of the same genus. Based on the amount 6 of available data on body size for each time bin the data is pooled at the epoch level for the 7 Permian and Early Triassic, and at the stage-level for the Middle and Late Triassic (data for 8 the Norian and Rhaetian stages are combined). MSL of each species was counted only once 9 per interval regardless of number of occurrences. A Log₂ transformation was chosen to make 10 differences at smaller sizes better visible. Statistical analyses were performed using PAST 11 (Hammer, Harper & Ryan, 2001).

12 A recent study by Romanuk, Hayward & Hutchings (2011) demonstrates that body size 13 of extant fishes significantly correlates with trophic level (see also Trebilco et al., 2013). 14 Accordingly, as a working hypothesis, body size of fossil fishes can be used as a proxy for 15 trophic position within ancient aquatic food webs. Note that a few extant taxa (e.g. large 16 planktivores) do show a negative correlation between body size and trophic level (Romanuk, 17 Hayward & Hutchings, 2011). Dietary data for Permian-Triassic bony fishes are limited (e.g. 18 Schaeffer & Mangus, 1976; Bürgin, 1996; Esin, 1997; Lombardo & Tintori, 2005; Boy & 19 Schindler, 2012; Bellwood et al., 2014) and hence are only qualitatively discussed herein, but 20 many large-bodied species possess conical teeth typical for carnivores, suggesting that they 21 conform to the general trend. In fact, large suspension-feeding Osteichthyes did not evolve 22 before the Jurassic (Friedman, 2012). We herein use the following rough body length 23 categories as proxies for the spectrum (range) and dominance (median plus 25–75% quartiles) 24 of trophic position of Permian-Triassic Osteichthyes: small consumers (below ~200 mm 25 MSL), mid-sized consumers (ca. 200-500 mm MSL) and large consumers (above ca. 500 mm 26 MSL).

2 (4). Quality of the osteichthyan fossil record

3

4 Inferring biological signals from the fossil record is often complicated because the 5 patterns that emerge from it can be skewed by other factors, including biases in preservation, 6 differences in sampling, and variations in research effort (see Benton et al., 2011 for 7 discussion). This is especially true for vertebrates because of their more sporadic stratigraphic 8 distribution compared to other groups (e.g. many invertebrates). Nevertheless, fishes are an 9 abundant group within the vertebrate fossil record and potentially useful for the study of 10 evolutionary and macroecological patterns through time (e.g. Lloyd & Friedman, 2013). 11 For the evaluation of biases related to differences in palaeogeographical sampling, we 12 compared the relative contribution of six macrogeographical provinces to the global fossil 13 record of Permian-Triassic osteichthyans. These provinces are: (1) Boreal Sea, (2) east-14 equatorial Panthalassa, (3) south-western Pangaea, (4) Palaeotethys, (5) Neotethys, and (6) 15 western and south-western Panthalassa (Fig. 1a). 16 To circumvent potential biases when comparing unevenly sampled assemblages, 17 rarefaction methods are traditionally used (e.g. Gotelli & Colwell, 2011). However, this 18 approach is not flawless. For instance, a rarefied richness of a Tropical assemblage may be 19 strongly underestimated when compared to the richness of a Boreal assemblage. Chao & Jost 20 (2012) provided an analytical solution for coverage-based rarefaction, "coverage" being a 21 measure of sample completeness. Contrary to classical, sample size-based rarefaction, this 22 rarefaction preserves the real degree of difference between the communities' richnesses, even for small sample size, and provides a sampling 'stopping rule'. Chao & Jost (2012) and 23 24 Colwell et al. (2012) also provided an analytical solution for the extrapolation of sample 25 richness to higher coverage instead of larger sample size, thus unifying coverage-based 26 interpolation and extrapolation into a single curve [see Section III(1)].

2 (5) Lagerstätten effect

3

4 One of the most important biases in the fossil record of bony fishes arises from the 5 occurrence of Lagerstätten (Seilacher et al., 1985). This is related to the fact that well-6 preserved material is essential for the identification of osteichthyan fossils at genus level, and 7 this kind of preservation is usually linked to occurrences of dysoxic to anoxic facies (Tintori, 8 1992; Ware et al., 2011). Because taxonomic richness at a particular site is dependent on the 9 amount of specimens recovered, time slices containing fossiliferous localities with good 10 quality of preservation frequently yield increased diversity compared to intervals without such 11 sites. Moreover, localities yielding large quantities of well-preserved fossils are often also 12 more extensively studied, leading to another bias through increased research effort. 13 In order to estimate the impact of the Lagerstätten effect we produced separate diversity 14 curves for the marine and freshwater realms in which osteichthyan genera that are known 15 solely from Lagerstätten are excluded and compared them with the trend lines derived from 16 the complete datasets [see Section III(1)]. In addition, we calculated the degree of endemism

17 of each Lagerstätte (i.e. percentage of genera that are exclusively known from that site during

18 a given epoch, Fig. 3). In general, the higher the degree of endemism, the larger the

19 Lagerstätten effect will be.

We herein identify osteichthyan Lagerstätten based on two criteria: (1) exceptional
preservation quality, i.e. presence of well-preserved, (largely) complete specimens, and (2)
high abundance of such fossils. Based on these criteria, the following Permian-Triassic
Lagerstätten are determined for marine bony fishes: Marl Slate and Kupferschiefer formations
(England and Germany, respectively; Late Permian; here treated together as a single
Lagerstätte), Wordie Creek Formation (Greenland; Early Triassic), «couches à poissons et à
ammonites» (NW Madagascar; Early Triassic), «fish horizon» (Spitsbergen; Early Triassic),

1	Sulphur Mountain Formation (British Columbia, Canada; Early Triassic), Luoping Biota
2	(Guizhou and Yunnan, China; Middle Triassic), Besano and Meride formations (Monte San
3	Giorgio area, Swiss-Italian boundary; Middle Triassic), Prosanto Formation (Switzerland;
4	Middle Triassic), and Zorzino Limestone (Bergamo Alps, Italy; Late Triassic). Note the
5	absence of marine Lagerstätten for the Early and Middle Permian (Fig. 3a).
6	For the freshwater record, we identify the following Lagerstätten: Autun, Bourbon
7	l'Archambault and Lodève basins (France; Early Permian; all three treated as a single
8	Lagerstätte), Krkonoše Piedmont Basin and Boskovice Graben (Czech Republic; Early
9	Permian), Upper Beaufort Series (Karoo Basin, South Africa; Early Triassic), Hawkesbury
10	Sandstone and Wianamatta Shale (New South Wales, Australia; Middle Triassic), and Chinle,
11	Dockum and Newark Groups (USA; Late Triassic; all three treated as one Lagerstätte). Note
12	that based on the criteria outlined above, no freshwater Lagerstätten could be identified for the
13	Middle and Late Permian (Fig. 3b).
14	Other methods to estimate sampling biases include division of taxonomic richness by (i)
15	number of localities or (ii) interval length (Supplementary Table B1). Note that division by
16	bin duration provides a time-standardised quantification of diversity, whereas division by
17	locality (or formation) number may suffer from redundancy (see e.g. Benton et al., 2011). We
18	did not test for correlations between diversity and rock volume (see e.g. Thomson, 1977 for
19	discussion) but these two variables are likely governed by other factors (Hannisdal & Peters
20	2011).
21	
22	(6) First and last appearance datums
23	
24	The general incompleteness of the fossil record has important implications for the first
25	appearance datum (FAD) and last appearance datum (LAD) of taxa. Although, for practical

26 reasons, FADs are often equated with true originations and LADs with real extinctions, in

many cases this assumption is not correct (e.g. Signor-Lipps effect, Signor & Lipps, 1982).
Moreover, different biological meanings can be attributed to FADs and LADs. Apart from
true origination or extinction, the FAD and LAD of a given taxon could also suggest a
migration, for instance from a palaeoenvironment with a poor fossil record into one that has a
good record, or vice versa (cf. *'refugia* taxa' during environmental crises, Kauffman &
Harries, 1986). Alternatively, a taxon may have stayed in its habitat prior to its FAD and/or
after its LAD but preservational conditions in that palaeoenvironment changed.

8 A famous example for migration as a likely explanation for a LAD is demonstrated for 9 actinistians. They are known from the Devonian up to the present, but lack a Cenozoic fossil 10 record (~70 Ma; Casane & Laurenti, 2013; Cavin, Furrer & Obrist, 2013). Until the discovery 11 of the extant coelacanth Latimeria in the 1930s (Smith, 1939), they were wrongly considered 12 extinct. Latimeria dwells in deep water environments, which have a poor fossil record. It was 13 hypothesized that such habitats could have been typical for post-Cretaceous actinistians, 14 providing a possible explanation for the gap in their fossil record (White, 1953; Forey, 1998). 15 However, it is noteworthy that actinistians are usually rare within fish assemblages and their 16 diversity in the fossil record is generally low, with a few exceptions (e.g. during the Early 17 Triassic; Schultze, 2004; Cavin, Furrer & Obrist, 2013). Although the migration scenario may 18 be a potential biological explanation for FADs and LADs of such less diverse or rare groups, 19 and generally at lower taxonomic ranks, it seems to be a more extraordinary assumption for 20 groups that are more diverse and abundant in the fossil record. At any rate, migration as well 21 as true origination and extinction of many taxa at the same time could still suggest an 22 underlying biotic event. As a working hypothesis, we herein follow previous authors and treat 23 FADs and LADs of taxa as genuine originations or extinctions, respectively, although 24 potential biases in the fossil record as well as the migration scenario may in some cases offer 25 alternative explanations.

26

2 III. RESULTS

3

4 (1) Estimation of sampling bias and Lagerstätten effect

5

6 Most of the record for the Permian and the Middle–Late Triassic is dominated by data 7 from the Palaeotethys, with important contributions from the western and south-western 8 Panthalassa region, south-western Pangaea, and the east-equatorial Panthalassa province (Fig. 9 1b). In the Early Triassic, however, trends within osteichthyans are mainly based on either the 10 Boreal Sea record (Griesbachian and Smithian) or the Neotethys record (Dienerian) because 11 of occurrences of Lagerstätten [see Section II(5)]. The Spathian record of Osteichthyes is poor 12 due to the absence of well-explored Lagerstätten and the data mostly come from China 13 (western and south-western Panthalassa province, Fig. 1b). Other important sampling biases 14 include large gaps in the Neotethys and western and south-western Panthalassa records during 15 the Permian (Fig. 1b).

16 Sample-size rarefaction and extrapolation curves (following Chao & Jost, 2012 and 17 Colwell et al., 2012) indicate that the estimated osteichthyan richness at the species and genus 18 levels is much higher for the Triassic than for the Permian (Fig. 2). Both time intervals show 19 rather similar sample coverage values (~ 0.6 at the genus level and ~ 0.25 at the species level), 20 indicating that data can be directly compared and are not likely to result from major 21 differential sampling artefacts. However, Permian and Triassic sample coverage values are 22 relatively low (especially at species level) suggesting that sampling effort for these two 23 periods can be improved in the future and that endemic taxa largely dominate known bony 24 fish records.

Our tentative evaluation of the Lagerstätten effect yielded different results for the
marine and non-marine records. In the marine realm (Fig. 3a), we find the same main long-

1 term trend from lower diversity in the Permian to higher diversity in the Triassic, with a peak 2 in the Middle Triassic, whether taxa known only from Lagerstätten [see Section II(5)] are 3 omitted or not. The main difference between the two curves is, however, found in the Early 4 Triassic: here, osteichthyan diversity is similar to that found in the Late Permian if 5 Lagerstätten taxa are excluded from the dataset, whereas genus richness during the Early 6 Triassic is intermediate to Late Permian and Middle Triassic values if the complete marine 7 dataset is considered (Fig. 3a). The reason for this discrepancy lies in the similarity among 8 bony fish faunas in Early Triassic Lagerstätten as a result of increased cosmopolitanism 9 during this epoch [Piveteau, 1935; Schaeffer & Mangus, 1976; see Section IV(2a)]. Despite 10 the presence of high-diversity Lagerstätten in the Middle Triassic (Monte San Giorgio, 11 Luoping), removal of taxa restricted to these sites does not obliterate the peak in genus 12 richness during this epoch. In fact, this peak is due to high endemism at Middle Triassic 13 localities (Fig. 3).

14 For the freshwater record, we find high genus richness in the Early Permian and in the 15 Early to Late Triassic, and relatively lower diversity in the Middle to Late Permian if the 16 complete dataset (including Lagerstätten taxa) is considered. However, if genera restricted to 17 Lagerstätten are removed from the analysis, similar levels of genus richness are found 18 throughout the Permian-Triassic interval (Fig. 3b). Hence, it appears that the trends in the 19 freshwater record of Permian-Triassic bony fishes are influenced by the Lagerstätten effect. 20 The reasons for this may be twofold: (1) relatively high endemism at genus level in 21 freshwater Lagerstätten (>48%, Fig. 3b), and (2) mostly low freshwater diversity.

Division of genus richness (with or without inclusion of Lagerstätten taxa) by the number of localities results in higher diversity in the Early and Middle Permian and again in the Middle and Late Triassic (see Supplementary Tables A1–A3, B1). Lower genus richness is found during the Late Permian and Early Triassic. These trends are evident in the marine and the freshwater datasets, although in the marine record genus richness of bony fishes

1	reaches similar levels in the Early-Middle Permian and the Middle-Late Triassic intervals,
2	whereas in the freshwater record Middle-Late Triassic diversity is only about half of that
3	found in the Early-Middle Permian interval. Division of genus richness by interval length
4	yields peaks in the Early Triassic, usually followed by high values in the Middle Triassic and
5	the Late Permian, both with and without inclusion of Lagerstätten genera (Supplementary
6	Table B1). This pattern, which is found in the marine, freshwater and global datasets, mainly
7	results from relatively short interval lengths of the Late Permian and Early Triassic and high
8	diversity and short interval duration for the Middle Triassic.
9	Correlation indices (Spearman's ρ , Kendall's τ) calculated for genus richness versus
10	number of localities and for genus richness versus bin length suggest that global osteichthyan
11	diversity is independent of number of localities and interval length. A correlation ($p < 0.05$)
12	does exist, however, between freshwater genus richness and stage/substage duration, and
13	between marine genus richness and number of localities (Supplementary Table B2).
14	
15	(2) General trends: Genus diversity, turnovers and turnover rates
16	
17	Diversity dynamics of marine osteichthyans (Fig. 4a, Supplementary Table B3) show an
18	overall trend from low levels in the Permian to much higher values in the Triassic. Genus
19	richness of marine bony fishes gently rises from the Early to the Late Permian. However, this
20	trend is not seen in the mean standing diversity and boundary crossers curves. Two distinct
21	diversity peaks can be discerned in the marine record, one during the Early Triassic and one
22	in the Middle Triassic, followed by decreasing diversity towards the Triassic-Jurassic
23	boundary. The Early Triassic diversity peak spans the Griesbachian to Smithian interval
24	(maxima in the Dienerian) and is succeeded by low diversity during the Spathian [the latter
25	being due to incomplete sampling, see Sections II(4) and $IV(2a)$]. Marine diversity is highest
26	in the Ladinian.

1	Turnovers (sum of originations and extinctions) among marine bony fishes are higher in
2	the Triassic than in the Permian (Fig. 4b). Marine per taxon turnover rates (TR, relative to
3	genus richness) are also high throughout the Permian-Triassic interval when singletons are
4	included, with a peak in per taxon origination rates (OR) at the Spathian-Anisian boundary
5	(Fig. 3b, Supplementary Table B4). Per taxon extinction rates (ER) show a peak in the
6	Cisuralian (Sakmarian-Artinskian boundary), increase towards the end of the Permian and
7	remain elevated during the Triassic. If singleton genera are omitted (Fig. 4c, Supplementary
8	Table B4), marine TR mainly peak (≥50%) in the late Cisuralian (Artinskian-Kungurian
9	boundary: 50% OR), at the Permian-Triassic boundary (65% OR, 57% ER), the Spathian-
10	Anisian boundary (94% OR, 64% ER) and the Ladinian-Carnian boundary (51% ER). ER
11	remain high (>30%) from the Middle-Late Triassic boundary onwards (Fig. 4c,

12 Supplementary Table B4).

13 Diversity trends of the freshwater osteichthyan record (Fig. 5a, Supplementary Table 14 B3) differ from those described for the marine dataset. Permian freshwater diversity is largely 15 similar to that of the Triassic when comparing numbers of boundary crossers as well as mean 16 standing diversity. However, freshwater genus richness is distinctly higher in the Middle and 17 Late Triassic compared to the Middle Permian-Early Triassic interval. All three diversity 18 estimates studied herein (genus richness, boundary crossers, mean standing diversity) show 19 relatively high freshwater bony fish diversity during the early Cisuralian. Generally low 20 diversity levels are found in the late Early Permian to early Middle Permian and again in the 21 Early Triassic. Note, however, that low Early Triassic diversity is a sampling artefact and 22 related to the circumstance that many freshwater fishes from this interval (mostly from the 23 Karoo Basin in South Africa and the Tunguska Basin of Siberia, Russia) cannot be accurately 24 dated at substage level. At epoch level, a diversity minimum is not evident in the Early 25 Triassic (Fig. 3b).

1	TR of freshwater osteichthyans (including singletons; Fig. 5b, Supplementary Table B4)
2	peak at the Sakmarian-Artinskian boundary (62% ER), the Artinskian-Kungurian boundary
3	(56% OR), the Wordian-Capitanian boundary (62% OR), the Permian-Triassic boundary
4	(64% ER), and several times during the Triassic (Spathian-Anisian boundary: 73% OR;
5	Anisian-Ladinian boundary: 63% OR, 70% ER; Ladinian-Carnian boundary: 67% OR, 63%
6	ER; Norian-Rhaetian boundary: 56% ER). Increased TR in the Triassic are linked to
7	occurrences of relatively short-lived endemic taxa (mainly Sydney Basin, New South Wales,
8	Australia). In fact, when singleton genera are removed (Fig. 5c, Supplementary Table B4),
9	Triassic TR of freshwater bony fishes tend to be much lower. Omission of singletons restricts
10	peaks in OR to the Wordian-Capitanian boundary (55%) and the Ladinian-Carnian boundary
11	(56%), whereas maxima in ER are only found at the Sakmarian-Artinskian boundary (65%),
12	the Permian-Triassic boundary (60%) and the Norian-Rhaetian boundary (50%).
13	Diversity patterns of the global bony fish record (Fig. 6a, Supplementary Table B3)
14	largely resemble those of the marine dataset. This is due to the fact that marine osteichthyans
15	account for the highest taxonomic diversity throughout the Permian-Triassic interval. Global
16	diversity of Osteichthyes decreases during the Cisuralian (minimum in the Artinskian) and
17	rises again until the Wuchiapingian, followed by peak values in the Early Triassic (Dienerian
18	substage) and Middle Triassic. Global diversity of bony fishes then decreases towards the
19	Triassic-Jurassic boundary. The described patterns are evident from all three diversity
20	estimates studied here.
21	The number of turnovers of bony fishes globally is usually higher in the Triassic than in
22	the Permian, as in the marine record, but global TR are similar throughout most of the
23	Permian-Triassic record if singletons are included (Fig. 6b, Supplementary Table B4). Peak
24	OR are observed at the Wordian-Capitanian boundary (50%), the Permian-Triassic boundary
25	(50%) and the Spathian-Anisian boundary (84%), whereas peak ER are found at the
26	Sakmarian-Artinskian boundary (62%), the Permian-Triassic boundary (50%), the Dienerian-

1	Smithian boundary (57%), the Spathian-Anisian boundary (50%), the Anisian-Ladinian
2	boundary (52%), the Ladinian-Carnian boundary (60%) and at the Norian-Rhaetian boundary
3	(54%). However, if singletons are omitted (Fig. 6c, Supplementary Table B4), peaks in global
4	OR are merely found at the Permian-Triassic boundary (50%) and at the Spathian-Anisian
5	boundary (76%). Excluding singletons, peak ER are only obvious at the Sakmarian-
6	Artinskian boundary (55%), but are otherwise absent in the global record.
7	
8	(3) Diversity dynamics within Osteichthyes
9	
10	Actinistia generally show low diversity during the Permian-Triassic. Elevated levels of
11	marine actinistian genus richness (Fig. 7a) are reached during the Changhsingian-Smithian
12	interval and again in the Middle Triassic. Low actinistian diversity during the Spathian is
13	likely due to sampling biases [see Sections II(4) and $IV(2a)$]. Actinistian diversity in
14	freshwater environments (Fig. 7b) is mostly very low, but a diversity increase is observed in
15	the Late Triassic. Among bony fishes, relative genus diversity of actinistians is highest during
16	the Early Permian and in the Early Triassic but generally lower at other times (Fig. 8a;
17	Supplementary Table B5).
18	Actinopterygians are the most diverse group among marine Permian-Triassic
19	osteichthyans (Fig. 8). Permian ray-finned fishes predominantly belong to 'Palaeopterygii'.
20	First representatives of 'Subholostei' (e.g. Bobasatrania, Stemmerik, Bendix-Almgreen &
21	Piasecki, 2001) and Holostei (Acentrophorus, e.g. Schaumberg, 1977) appear in the marine
22	record in the Lopingian. Whereas genus richness of marine non-subholostean
23	'palaeopterygians' remains at similar levels from the Middle Permian to the Middle Triassic,
24	'subholosteans' pass through a massive diversification event during the Early Triassic and
25	again in the Middle Triassic (Figs 7a, 8a). 'Palaeopterygians' and 'subholosteans' show a
26	decline in diversity from the Ladinian onwards (Figs 7a, 8a). In contrast, marine holosteans

(e.g. Semionotiformes) and Teleosteomorpha diversified during the Middle and Late Triassic.
 Although genus richness of marine neopterygians also decreases towards the Triassic-Jurassic
 boundary after their diversity peaks in the Ladinian (Holostei) and Norian (Teleosteomorpha,
 Fig. 7a), their diversity relative to non-neopterygians continuously increases during the
 Middle and Late Triassic and, by the Norian, exceeds that of any other group of marine bony
 fishes (Fig. 8a).

7 Freshwater diversity of sarcopterygians is dominated by dipnoans (Fig. 7b, 8 Supplementary Table B5). Genus richness of lungfishes is generally higher during the 9 Triassic than in the Permian (Fig. 7b), and their relative diversity within freshwater bony fish 10 communities reaches peak values during the Early Triassic (Fig. 8b). Note, however, that the 11 significance of this peak is equivocal due to the generally low diversity during this interval. 12 Freshwater actinistians show relatively low genus richness during the Permian and the Early 13 to Middle Triassic interval, but slightly increased diversity in the Late Triassic (Chinle, 14 Dockum and Newark Groups; Figs 7b, 8b).

15 Diversity of actinopterygians within Permian freshwater environments is dominated by 16 'palaeopterygians' (Figs 7b, 8b). So far, only one putative holostean has been described from 17 Permian freshwater deposits (Brachydegma; Hurley et al., 2007; Near et al., 2012). As in the 18 marine record, 'subholosteans' seemingly first appear in the Late Permian (Ischnolepis; 19 Murray, 2000) and they too become a larger component of freshwater bony fish faunas during 20 the Middle and Late Triassic. However, other than in the marine realm, holosteans and 21 teleosteomorphs account for only a small proportion of Triassic freshwater assemblages. In 22 fact, non-marine actinopterygian diversity in the Triassic is still dominated by 23 'palaeopterygians' and 'subholosteans' (Fig. 8b).

Globally, changes in faunal composition within osteichthyans largely resemble those of the marine record, with minor differences. Most notably, neopterygian dominance in genus richness relative to other bony fishes in the Middle and Late Triassic is less pronounced in the

global record compared to the marine one (Figs 7c, 8c). Sarcopterygian diversity on a global
 scale is relatively constant during the investigated time interval, with elevated genus richness
 in the Artinskian and during the Changhsingian-Spathian (Figs 7c, 8c, Supplementary Table
 B5).

5

6 (4) Palaeolatitudinal diversity

7

8 In the marine record, epoch-level-averaged palaeolatitudinal diversity of osteichthyans 9 is highest in the low-palaeolatitudinal belt throughout the Permian and again in the Middle 10 and Late Triassic. In the Early Triassic, however, low-palaeolatitudinal diversity is low and 11 maximum genus richness is found in the mid-palaeolatitudinal zone (Fig. 9a). Marine 12 diversity at genus level within the low-palaeolatitudinal zone during the Early Triassic is 13 similar to Permian values but distinctly lower compared to the Middle and Late Triassic. For 14 the whole Permian-Triassic record, genus richness of marine osteichthyans at mid-15 palaeolatitudes is highest in the Early Triassic. The highest low-palaeolatitudinal osteichthyan 16 diversity is measured in the Middle Triassic.

17 Changes in palaeolatitudinal genus richness among freshwater bony fishes (Fig. 9b) 18 differ from those observed in the marine record. Non-marine genus richness at low-19 palaeolatitude is similar to or higher than at mid-palaeolatitudes throughout the Permian and 20 again in the Late Triassic. However, in the Early and Middle Triassic, number of genera in the low-palaeolatitudinal zone is distinctly lower than that in the mid-palaeolatitudinal zones. 21 22 Freshwater diversity at mid-palaeolatitudes is generally higher during the Triassic than during 23 the Permian, and highest in the Middle Triassic. High-palaeolatitudinal genus richness 24 normally constitutes the lowest values except in the Early Triassic, where it exceeds low-25 palaeolatitudinal levels (Fig. 9b).

1	Globally, palaeolatitudinal distribution of osteichthyan genus richness during the
2	Permian and Triassic mostly resembles present-day conditions, with highest genus richness at
3	low-palaeolatitudes, lowest diversity at high-palaeolatitudes, and intermediate genus richness
4	at mid-palaeolatitudes (Fig. 9c). The only exception is found in the Early Triassic, where
5	palaeolatitudinal genus diversity is highest within the mid-palaeolatitudinal zones. High-
6	palaeolatitudinal genus richness also reaches maximum values during the Early Triassic. Peak
7	values in low-palaeolatitudinal global diversity are restricted to the Middle and Late Triassic
8	(Fig. 9c). Relative diversity of the three main groups of bony fishes investigated herein
9	(Dipnoi, Actinistia, Actinopterygii) is similar across palaeolatitudinal belts throughout the
10	Permian-Triassic interval (Supplementary Tables A1–A3).
11	
12	(5) Body size
13	
14	Permian-Triassic bony fishes show distinct fluctuations in the range and dominance of
15	species' maximum standard length (MSL). In the marine record, body size spectrum ranges
16	from ca. 20–900 mm in the Early Permian, to ca. 50–620 mm in the Middle Permian, to ca.
17	50–1500 mm in the Late Permian and Early Triassic, to ca. 20–1650 mm in the Anisian stage
18	to the Norian-Rhaetian interval (Fig. 10a, Supplementary Tables A1-A3). The Kolmogorov-
19	Smirnov test for overall equal distribution yields p -values < 0.05 only between the Early and
20	
	Middle Permian and between the Middle and Late Permian, and $p < 0.01$ exclusively between
21	Middle Permian and between the Middle and Late Permian, and $p < 0.01$ exclusively between the Early Triassic and Anisian (Supplementary Table B6). Differences in body size
21 22	Middle Permian and between the Middle and Late Permian, and $p < 0.01$ exclusively between the Early Triassic and Anisian (Supplementary Table B6). Differences in body size distribution of marine osteichthyans between other consecutive intervals are not statistically
21 22 23	Middle Permian and between the Middle and Late Permian, and $p < 0.01$ exclusively between the Early Triassic and Anisian (Supplementary Table B6). Differences in body size distribution of marine osteichthyans between other consecutive intervals are not statistically different ($p > 0.43$). Medians of marine osteichthyan MSL are also only significantly different
21 22 23 24	Middle Permian and between the Middle and Late Permian, and $p < 0.01$ exclusively between the Early Triassic and Anisian (Supplementary Table B6). Differences in body size distribution of marine osteichthyans between other consecutive intervals are not statistically different ($p > 0.43$). Medians of marine osteichthyan MSL are also only significantly different between the Late Permian and Early Triassic (Mann-Whitney U test, $p < 0.05$) and between
21 22 23 24 25	Middle Permian and between the Middle and Late Permian, and $p < 0.01$ exclusively between the Early Triassic and Anisian (Supplementary Table B6). Differences in body size distribution of marine osteichthyans between other consecutive intervals are not statistically different ($p > 0.43$). Medians of marine osteichthyan MSL are also only significantly different between the Late Permian and Early Triassic (Mann-Whitney U test, $p < 0.05$) and between the Early Triassic and Anisian ($p < 0.01$; Supplementary Table B6). In general, body size

consumers in the Early Permian, the Late Permian, and the Early Triassic. A tendency
 towards small sized consumers is found in the Middle Permian and during the Middle–Late
 Triassic (Fig. 10a).

4 MSL range in freshwater osteichthyans varies between ca. 40–1030 mm in the Early Permian, to ca. 50–500 mm in the Middle Permian, to ca. 50–700 mm in the Late Permian, to 5 6 ca. 55–700 mm in the Early Triassic, to ca. 38–1000 mm in the Middle Triassic, to ca. 30– 7 1500 mm in the Late Triassic (Fig. 10b, see Supplementary Tables A1–A3). MSL 8 distributions are statistically different only between the Late Permian and Early Triassic 9 (Kolmogorov-Smirnov, p < 0.01), between the Early and Middle Triassic (p < 0.01) and 10 between the Middle and Late Triassic (p < 0.05; Supplementary Table B6). Differences in 11 median MSL are statistically different only between the Late Permian and Early Triassic as 12 well as between the Early and Middle Triassic (Mann-Whitney U, p < 0.01; Supplementary 13 Table B6). In the non-marine realm, body size distribution is skewed towards mid-sized 14 consumers predominantly in the Middle and Late Permian. In contrast, the bulk of freshwater 15 osteichthyans of the Early Triassic and the Late Triassic can be classified as small consumers 16 (Fig 10b).

17 Globally (Fig. 10c, see Supplementary Tables A1–A3), differences in the distribution of 18 osteichthyan MSL are only statistically significant between the Early Triassic and Anisian 19 (Kolmogorov-Smirnov, p < 0.01; Supplementary Table B6). Significantly different median 20 MSL is also only evident between the Early Triassic and Anisian (Mann-Whitney U, p <21 0.01), although low *p*-values are also yielded between the Middle and Late Permian (p < 0.05; 22 Supplementary Table B6). On a global scale, body size distribution of Osteichthyes suggests 23 dominance of mid-sized consumers during the Permian and Early Triassic and dominance of 24 small consumers during the Middle–Late Triassic (Fig. 10c).

Body size distribution among the different groups of bony fishes during the Permian and
 the Triassic suggests very similar patterns in marine and freshwater environments. Within

1 Actinopterygii, median MSL of 'palaeopterygians' tends to be larger than that of 2 'subholosteans', holosteans or teleosteomorphs throughout the studied interval (Fig 10a). In 3 the marine realm, median MSL of 'Palaeopterygii' varies between ca. 250 mm and 300 mm. 4 These fishes would therefore mostly fall into the mid-sized consumers' trophic category. 5 However, marine 'palaeopterygian' species tend to be much smaller during the Middle 6 Permian (median of 120 mm MSL) and larger during the Late Triassic (median of ca. 350– 7 1000 mm MSL, but note the smaller sample size). A significant size increase in marine 8 'Palaeopterygii' is noted across the Guadalupian-Lopingian boundary (Fig. 10a; Kolmogorov-9 Smirnov and Mann-Whitney U tests, p < 0.01). Marine 'Subholostei' and Neopterygii have 10 median MSL ranging up to ~150 mm and, thus, these groups mainly comprise small 11 consumers throughout the Permian and Triassic (Fig 10a). Marine actinistians show similar 12 median body sizes as 'palaeopterygians', but sample size is generally low. Small to mid-sized 13 'palaeopterygians' become rarer after the Middle Triassic, whereas holosteans continue to 14 extend the upper end of their size spectrum towards the close of the Triassic (although most 15 species remain in the small consumer's guild; Fig 10a). Body size range of teleosteomorphs, 16 on the other hand, remains small (23–155 mm MSL) throughout the studied interval (Fig. 17 10a).

18 Among Permian-Triassic freshwater fishes, there also is a tendency towards larger body 19 sizes in dipnoans and actinistians (median MSL >330mm, but note small sample size) and 20 'palaeopterygians' (median MSL 113-245 mm) compared to 'subholosteans', holosteans, and 21 teleosteomorphs (median MSL 55-120 mm; Fig 10b). Thus, like in the marine realm, 22 freshwater 'palaeopterygians' (and maybe alsosarcopterygians) predominantly occupied the niche of mid- to large-sized consumers, whereas 'subholosteans' and neopterygians mostly 23 24 covered relatively lower trophic positions. However, other than in the marine realm, small 25 'palaeopterygians' do not become scarcer and holosteans do not seem to comprise larger 26 species towards the end of the Triassic in the non-marine domain (Fig. 10b).

1	Changes in osteichthyan body size across palaeolatitudes indicate that the slight increase
2	in global species' MSL between the Guadalupian and the Early Triassic (cf. Fig. 10) can be
3	observed both in the low- and the mid-palaeolatitudinal zones (Fig. 11), although these shifts
4	are not significant (Kolmogorov-Smirnov and Mann-Whitney U tests). The decrease in body
5	size between the Early and Middle Triassic that is evident in the marine realm and on a global
6	scale (Fig. 10), occur within both the low-palaeolatitudinal zone (statistically significant for
7	the marine and global datasets, Kolmogorov-Smirnov and Mann-Whitney U, $p < 0.01$, Fig.
8	11a, c) and within the mid-palaeolatitudinal belts (global, Mann-Whitney U, $p < 0.05$, Fig.
9	11c).
10	
11	
12	IV. DISCUSSION
13	
14	(1) The Permian record
15	
16	(a) "Olson's gap"
17	
18	The Permian is marked by at least three biotic events, each of which affected marine and
19	continental life to varying extent. The first of these three events has been termed "Olson's
20	gap" (Lucas & Heckert, 2001) or "Olson's extinction" (Sahney & Benton, 2008) and it refers
21	to a global hiatus in the terrestrial tetrapod record during an interval spanning the latest
22	Kungurian and most of the Roadian (Lucas, 2004, 2013). It separates the Early Permian North
23	American tetrapod faunas, dominated by basal synapsids ('pelycosaurs') and temnospondyl
24	'amphibians', from the Middle Permian therapsid-dominated communities of Russia and
25	South Africa (Lucas, 2004; Sahney & Benton, 2008; Blieck, 2011). A diversity drop in
26	freshwater and euryhaline osteichthyan families coincident with "Olson's gap" was noted by

Pitrat (1973) and is also implied by the analysis of Blieck (2011), based on data from Benton
 (1993).

3 No extinction event for bony fishes in the Kungurian or Roadian is evident from our 4 data (Figs 4-6). However, genus richness of freshwater osteichthyans notably decreased 5 earlier in the Cisuralian, which is mainly due to the disappearance of taxa native to the great 6 fluvio-lacustrine systems of Central Europe (Czech Republic, France, Germany) and the 7 southern United States (New Mexico, Oklahoma, Texas). Many of these taxa crossed the 8 Carboniferous-Permian boundary, such as the actinopterygians Aeduella, Bourbonella, 9 Paramblypterus, Platysella and Progyrolepis, and the dipnoans Conchopoma, Ctenodus, Gnathorhiza, Megapleuron, Proceratodus and Sagenodus (e.g. Štamberg, 1994, 2007; Heyler, 10 11 2000; Schultze, 2004; Poplin & Dutheil, 2005; see Supplementary Tables A1-A3). A few 12 chondrichthyan clades also vanished during the Early Permian, although the overall diversity 13 of these fishes remained relatively stable (Koot, 2013). Furthermore, megalichthyid and 14 rhizodopsid sarcopterygians (basal tetrapodomorphs, Janvier, 2007), which were confined to Euramerican freshwater basins in the Early Permian, also seem to disappear from the fossil 15 16 record at that time (Schultze & Heidtke, 1986; Witzmann & Schoch, 2012). Another group 17 that potentially became extinct during the Cisuralian are the 'acanthodians', but there is 18 ambiguity regarding their LAD (either Early Permian or Middle-Late Permian, Mutter & 19 Richter, 2007). Both 'acanthodians' and basal tetrapodomorphs were more diverse during 20 their early evolutionary history but became minor faunal elements after the end-Devonian 21 Hangenberg extinction (Sallan & Coates, 2010; Witzmann & Schoch, 2012). 'Acanthodians' 22 and basal tetrapodomorphs are frequently cited among the victims of the end-Permian mass 23 extinction (e.g. Schaeffer, 1973; Janvier, 1996; Mutter & Richter, 2007) but they might 24 already have become extinct during the general decline in freshwater fish diversity during the 25 Cisuralian. The diversity loss among Permo-Carboniferous bony fish taxa in Euramerican 26 fluvio-lacustrine systems during the Early Permian is probably linked to the reduction or

disappearance of their habitats (Pitrat, 1973; Schneider & Werneburg, 2012) as a result of
 contemporaneous aridification trends and other climatic changes (Ziegler *et al.*, 2003;

3 Roscher & Schneider, 2006; Chen *et al.*, 2013).

Although there is no loss in genus richness among marine bony fishes during the
Cisuralian, we note elevated per taxon origination rates at the Kungurian-Roadian boundary
(Fig. 4c) and a possible reduction in body size across the Cisuralian-Guadalupian boundary
(Fig. 10a). Most Early Permian marine osteichthyans originate from the Eastern European
platform, based on which Esin (1997) observed a decrease in body size in the late Kazanian
(late Roadian; Menning *et al.*, 2006). However, whether this change in body length of marine
bony fishes is related to the continental "Olson's gap" remains unknown.

11

12 *(b) The end-Guadalupian extinction event*

13

14 The second Permian event is the end-Guadalupian (or mid-Capitanian; Wignall et al., 15 2012) extinction. This event is associated with shifts in the carbon and oxygen isotope record 16 (Chen et al., 2013) and a first-order sea-level lowstand (see Groves & Wang, 2013 for a 17 summary). Although there is still disagreement about the timing and main triggers of the end-18 Guadalupian extinction (Clapham, Shen & Bottjer, 2009; Groves & Wang, 2013), it seems 19 that the event is prolonged and possibly linked to a cool phase during the Capitanian 20 ('Kamura event', Isozaki, Kawahata & Ota, 2007) as well as to eruptive activity of the 21 Emeishan Traps (southern China) and the sea-level lowstand during the Capitanian-22 Wuchiapingian transition (e.g. Wignall et al., 2012; Clapham & Payne, 2011; Groves & 23 Wang, 2013). Victims of the Guadalupian crisis were predominantly marine invertebrates, 24 including fusulinoidean foraminifera, corals, bryozoans, brachiopods, and ammonoids 25 (Glenister & Furnish, 1981; Stanley & Yang, 1994; Clapham, Shen & Bottjer, 2009; Groves 26 & Wang, 2013; Wang & Sugiyama, 2000), although some terrestrial tetrapods also suffered

extinctions during the Middle Permian (Lucas, 2009; Fröbisch, 2013). Clapham, Shen &
Bottjer (2009) pointed out that the diversity decrease of various marine invertebrate groups
occurred over an extended interval (Wordian–Changhsingian) and mainly resulted from low
origination rates. Increased extinction rates at the Capitanian-Wuchiapingian boundary are,
nevertheless, found for some fusulinoidean families (Groves & Wang, 2013) and corals
(Wang & Sugiyama, 2000).

7 Chondrichthyan diversity trends imply that the Guadalupian crisis had an important 8 impact on these fishes, as it largely marks the tipping point of their dramatic diversity drop 9 towards the end of the Permian (Fig. 12; Pitrat, 1973; Koot, 2013). Strikingly, patterns in 10 bony fishes are very different. Global diversity of Osteichthyes is slowly rising between the 11 Kungurian and Wuchiapingian (Fig. 6a) and high per taxon turnover rates at the Guadalupian-12 Lopingian boundary are not evident (Fig. 6c). Nevertheless, changes in trophic composition 13 of Middle-Late Permian ichthyofaunas from basins within the European part of Russia have 14 been documented by Esin (1997). Amongst others, he noted a decrease in diversity of 15 actinopterygians with a presumed durophagous diet in the late Urzhumian (late Wordian; 16 Menning et al., 2006), in tandem with lowered benthic taxonomic diversity. This was 17 followed by reduced trophic diversity in the Sverodvinian (Capitanian-early Wuchiapingian; 18 Menning et al., 2006), characterised by a high abundance of algal grazers. Diversity of 19 durophagous 'palaeopterygians' did not recover until the Vyatkian (late Wuchiapingian and 20 Changhsingian; Menning et al., 2006). Esin (1997) interpreted these changes in trophic 21 communities as being related with alterations in local aquatic environments and sea-level 22 fluctuations, but they could be evidence for a potential crisis in relation to the end-23 Guadalupian event.

During the Lopingian aftermath, global genus richness of Osteichthyes reached similar levels to that of Chondrichthyes for the first time (Fig. 12; Koot, 2013). This diversity trend is coupled with a significant body size increase among marine 'palaeopterygians' across the

1	Middle-Late Permian boundary. Following the scarcity of large osteichthyan consumers in the
2	Early and Middle Permian, Lopingian faunas bear several large-sized marine taxa (e.g.
3	species of Acropholis, Mutovinia, Plegmolepis) with similar body lengths as the osteichthyan
4	apex predators of the Triassic (e.g. species of Birgeria, Saurichthys, Lombardo & Tintori,
5	2005; Romano & Brinkmann, 2009; Romano et al., 2012; see Supplementary Tables A1–A3).
6	The Late Permian is further characterized by the first appearance of 'subholosteans'
7	(Bobasatrania, Dorypterus, Ischnolepis) in marine and freshwater environments, and small-
8	sized holosteans (Acentrophorus, Archaeolepidotus) in the marine realm (Figs 7, 8, 10; see
9	Supplementary Tables A1-A3). However, these groups, which greatly diversified later in the
10	Triassic [see Section IV(2)], make up only a small component of Lopingian ichthyofaunas
11	(e.g. Accordi, 1955; Jubb & Gardiner, 1975; Schaumberg, 1977; Stemmerik, Bendix-
12	Almgreen & Piasecki, 2001; Brinkmann et al., 2010).
13	
14	(c) The end-Permian mass extinction event
15	
16	The end-Permian (late Changhsingian; Shen et al., 2011) mass extinction is the most
17	devastating biotic crisis of the Phanerozoic (Erwin, 2006; Benton, 2003), wiping out an
18	estimated 96% of marine species (Raup, 1979) and about 49% of terrestrial tetrapod families
19	(Benton & King, 1989). This event, which was possibly linked to eruptions of the Siberian
20	Large Igneous Province (Ivanov et al., 2013) and associated climatic and environmental
21	changes (e.g. Schneebeli-Hermann et al., 2013; Benton & Newell, 2013), had a profound
22	impact on various marine invertebrate groups, including ammonoids (Brayard et al., 2009),

23 gastropods (e.g. Batten, 1973; Payne, 2005), brachiopods (Shen & Shi, 2002), corals (e.g.

24 Wang & Sugiyama, 2000), and echinoderms (Twitchett & Oji, 2005), but also affected

25 terrestrial tetrapods (Lucas, 2009; Fröbisch, 2013). Land plants experienced a major change

26 from dominance of seed plants to spore plants at the Permian-Triassic boundary (Schneebeli-

Hermann *et al.*, 2013). However, fishes (and conodonts) are considered largely unimpaired by
 the end-Permian event (e.g. Schaeffer, 1973; Thomson, 1976, 1977; Benton, 1998; Orchard,
 2007; Reguant, 2007; Blieck, 2011).

4 Friedman & Sallan (2012) argued that the Late Permian record of fishes is very poor, 5 with many lineages ending before the boundary with the Triassic. However, our updated 6 database shows that the Lopingian record has improved with several lineages ending in the 7 Changhsingian, leading to elevated extinction rates at the Permian-Triassic boundary in both 8 the marine (57–70%, Fig. 4, Supplementary Table B4) and the freshwater record (60–64%, 9 Fig. 5, Supplementary Table B4). It should be noted that these high extinction rates are also a 10 consequence of the relatively low genus richness at the end of the Permian. Moreover, 11 extinction rates in the Changhsingian are lower at global scale (38–50%, Fig. 6, 12 Supplementary Table B4) due to the fact that some genera switch between marine and 13 freshwater environments during the Permian-Triassic transition (Acrolepis, Atherstonia, 14 Palaeoniscum, Pteronisculus?; see Supplementary Tables A1–A3). Others seemingly 15 survived the boundary without invading new habitats (e.g. Bobasatrania, Elonichthys, 16 Pygopterus; see Supplementary Tables A1–A3). Many genera crossing the Permian-Triassic 17 boundary are peculiarly long-lived and some probably represent 'Elvis taxa' (Erwin & 18 Droser, 1993). The impact of the end-Permian event on bony fish diversity remains elusive 19 until these taxa are subjected to critical re-examination and revision. 20 Body size changes across the Permian-Triassic boundary indicate no major shiftsamong 21 marine Osteichthyes suggesting relatively stable trophic communities in the oceans at epoch 22 level, whereas the significant body length decrease of freshwater bony fishes could indicate a 23 potential crisis for continental faunas (Figs 10, 11, Supplementary Table B6). Most Lopingian 24 marine apex predators among bony fishes (e.g. Acropholis, Mutovinia, Plegmolepis) did not 25 cross the Permian-Triassic boundary and by the Early Triassic, new osteichthyan taxa such as 26 Birgeria and Saurichthys (as well as marine tetrapods) became abundant and widespread

1	(Romano et al., 2012; Scheyer et al., 2014; Supplementary Tables A1–A3). Despite the rapid
2	recovery of oceanic trophic networks after the end-Permian event (Scheyer et al., 2014), the
3	taxonomic turnover within marine apex predatory guilds across the Permian-Triassic
4	boundary suggests that higher trophic levels nonetheless suffered during the mass extinction.
5	
6	(2) The Triassic record
7	
8	(a) Early Triassic recovery
9	
10	The Early Triassic has received increased research interest during recent decades due to
11	the study of biotic recovery following the end-Permian mass extinction (e.g. Benton, 2003;
12	Erwin, 2006, Foster & Twitchett, 2014). It is frequently assumed that the Triassic recovery
13	was unusually slow and protracted, with groups like corals (e.g. Deng & Kong, 1984; Qi &
14	Stanley, 1989; Roniewicz & Stanley, 1998; Payne et al., 2006), some echinoderms (Twitchett
15	& Oji, 2005; Hagdorn, 2011), gastropods (Batten, 1973; Erwin, 1996; Fraiser & Bottjer,
16	2004; Nützel & Schulbert, 2005) or terrestrial vertebrates (e.g. Sahney & Benton, 2008)
17	possibly exhibiting delayed re-establishment of taxonomic diversity (i.e. not before the
18	Middle Triassic or later). However, in recent years it has been demonstrated that a number of
19	groups such as conodonts (Orchard, 2007), ammonoids (e.g. Brayard et al., 2009) or the
20	benthos (Brayard et al., 2011; Hautmann et al., 2013; Hofmann et al., 2013; Foster &
21	Twitchett, 2014) already display diversity peaks during the Early Triassic. These faster
22	recovering groups nevertheless experienced relapses in their diversity during the Early
23	Triassic (see below).
24	Carbon and oxygen isotope records, as well as spore-pollen ratios within sporomorph
25	assemblages show large fluctuations indicating iterative climatic changes in the wake of the

26 end-Permian mass extinction (Payne et al., 2004; Galfetti et al., 2007a, b; Hermann et al.,

1 2012; Sun et al., 2012; Romano et al., 2013), which may be linked to protracted eruptive 2 phases of the Siberian Traps (Ivanov et al., 2013 and references therein). It has been hypothesized that biotic recovery in the Early Triassic may have occurred whenever 3 4 favourable conditions prevailed, for instance during amelioration of temperatures, cessation of 5 anoxia or during transgressions (Orchard, 2007; Brayard et al., 2011; Romano et al., 2013). 6 Soon after the end-Permian mass extinction, bony fishes experienced a great increase in 7 global diversity (Fig. 6). This first osteichthyan diversification event at the onset of the Early 8 Triassic took place in both marine and freshwater environments (Figs 4–5) and involved both 9 sarcopterygians and actinopterygians (Figs 7-8; also see Cavin, Furrer & Obrist, 2013; Tintori 10 et al., 2013). This origination peak at the onset of the Triassic cannot be dismissed as a mere 11 taxonomic artefact. Most Early Triassic bony fishes are morphologically very distinct from 12 their Permian relatives and their attribution to different genera is well justified. The Early 13 Triassic diversification pulse may partly be related to the prevalence of Lagerstätten (i.e. 14 Greenland, NW Madagascar, Spitsbergen, western Canada; Fig. 3) as well as to ideal 15 preservational conditions linked to widespread episodic anoxic events, i.e. in the Dienerian 16 and Late Smithian (Tintori, 1992; Hermann et al., 2011; Ware et al., 2011). Strikingly, 17 however, Early Triassic marine bony fish Lagerstätten comprise many shared genera 18 (indicated by low endemism rates, Fig. 3), a fact that has already puzzled previous researchers 19 (e.g. Piveteau, 1935; Schaeffer & Mangus, 1976; Brinkmann et al., 2010). Cosmopolitanism 20 was also recognized for Early Triassic ammonoids, although only during certain intervals 21 (Tozer, 1982; Dagys, 1988; Brayard et al., 2006; Brayard, Escarguel & Bucher, 2007). Some 22 sea-going tetrapods like trematosauroid 'amphibians' (Aphaneramma, Wantzosaurus) or early 23 ichthyopterygian reptiles like Utatsusaurus were also wide-ranging during the Early Triassic 24 (Scheyer et al., 2014 and references therein).

Cosmopolitan taxa are typical for post-extinction faunas and they are commonly
referred to as 'disaster taxa' (e.g. Sahney & Benton, 2008; Benton & Newell, 2013). Apart

from their wide palaeogeographic distribution, 'disaster taxa' (*sensu* Kauffman & Harries, 1996) are also typified as being r-strategists and as showing high fossil abundance within post-extinction assemblages (short-term blooms). 'Disaster taxa' are usually rare among preextinction communities and only become more frequent after competitor taxa declined. They are also interpreted as having evolved to cope with stressful environmental conditions prior to the extinction event, and hence were able to survive it and to thrive rapidly afterwards (Kauffman & Harries, 1996).

8 , The cosmopolitan Early Triassic bony fish genera cannot be classified as 'disaster 9 taxa' for the following reasons. Firstly, none of them has a dominant occurrence in Early 10 Triassic Lagerstätten (e.g. Lehman, 1952; Nielsen, 1942, 1949, 1961; Schaeffer & Mangus, 11 1976; Stensiö, 1921, 1925, 1932). In fact, based on the inventory list published by Nielsen 12 (1961), Early Triassic osteichthyan Lagerstätten are characterized by relatively high evenness 13 (Simpson index 1-D): East Greenland (0.7907), north-west and south-west Madagascar 14 (0.7919), and Spitsbergen (0.8095). This is in sharp contrast to, for example, the Late Permian 15 German Kupferschiefer, where 90% of all fish fossils are identified as Palaeoniscum 16 freieslebeni (see Schaumberg, 1977). Secondly, although some cosmopolitan genera already 17 appear before the Permian-Triassic boundary, and some continue to exist after the Early 18 Triassic, none of them has yet been demonstrated to exhibits traits suggestive of adaptation to 19 stressful environments. Wen et al. (2013) speculated that Early Triassic actinistians may have 20 been 'disaster taxa' because they could have been adapted to low-oxygenation conditions, like 21 the extant coelacanth Latimeria. However, care must be taken not to assign biological traits of 22 one genus to a whole group, especially in the light of the great morphological variety 23 expressed by Early Triassic actinistians (Casane & Laurenti, 2013). 24 Schaeffer & Mangus (1976) assigned possible feeding habits to a number of Early 25 Triassic genera based on gape size, dentition type and body morphology. According to their

26 interpretations, Early Triassic osteichthyans exhibit a mixture of feeding strategies and

1 include small-bodied planktivores, grazers, and detritus feeders (e.g. Australosomus, 2 Boreosomus, parasemionotids), small to large predators (e.g. Birgeria, Pteronisculus, 3 Saurichthys), as well as durophagous forms (e.g. Bobasatrania). Stensiö (1921) also ascribed 4 a durophagous diet to some large Early Triassic actinistians (e.g. Mylacanthus, 5 *Scleracanthus*). These feeding habits are in agreement with the potential food sources of Early 6 Triassic seas, such as small vertebrates (including juvenile predators, conodonts; Orchard, 7 2007; Scheyer et al., 2014), plankton (e.g. larvae of various stenohaline animals, acritarchs; 8 e.g. Schneebeli-Hermann et al., 2013) and benthos (decapods, ostracods, bivalves, gastropods, 9 brachiopods, echinoderms, sponges, foraminifera; e.g. Garassino & Pasini, 2002; Twitchett & 10 Oji, 2005; Crasquin-Soleau et al., 2007; Brayard et al., 2010, 2011; Song et al., 2011; 11 Hautmann et al., 2013; Hofmann et al., 2013). These potential prey organisms must have 12 been readily available during the Early Triassic to allow for the observed body size ranges of 13 marine osteichthyans (Fig. 10), including top piscine predators of 1 m length or more 14 (Romano et al., 2012; Stensiö, 1921; Wendruff & Wilson, 2012), as well as the presence of 15 large marine tetrapods (Scheyer et al., 2014). Marine predator diversity, distribution, and size 16 spectrum suggest that oceanic productivity during the Early Triassic was sufficient to sustain 17 such large predators (Romanuk, Hayward & Hutchings, 2011; Trebilco et al., 2013; Scheyer 18 et al., 2014).

19 The Early Triassic osteichthyan diversification event was seemingly restricted to mainly 20 mid-palaeolatitudes in the marine realm and largely to mid- to high-palaeolatitudes in the 21 non-marine domain, whereas in both realms the diversity at low-palaeolatitudes decreased 22 across the Permian-Triassic boundary (Fig. 9). Sun et al. (2012) mapped the palaeogeographic 23 distribution of fish and tetrapod localities between the Late Permian and Anisian and argued 24 for an 'equatorial marine vertebrate eclipse' in the Late Smithian – an interval that likely lasted less than 150 kyr (Brühwiler et al., 2010). Besides the fact that the quality of the fossil 25 26 records of Triassic fishes and tetrapods does not allow for recognition of palaeobiogeographic
1 patterns with this stratigraphical precision, the alleged gap in the distribution of vertebrate 2 sites is largely one between the equator and ca. 40°S. This area has an incomplete fossil 3 record throughout the Late Permian to Anisian interval (Sun et al., 2012, fig. S1). Indeed, the 4 record of marine fishes from Gondwana is notoriously poor during the whole Triassic (Fig. 1; 5 e.g. López-Arbarello, 2004), partially related to the fact that sampling in the southern 6 hemisphere is generally poorer than in the northern one (Allison & Briggs, 1993). Hence, our 7 knowledge about low-palaeolatitudinal marine osteichthyan fish faunas of the Early Triassic 8 is entirely based on the northern equatorial realm (ca. 0°–30°N), i.e. on records from South 9 China and western North America (Fig. 1; see Supplementary Tables A1-A3). Note that low-10 palaeolatitudinal vertebrate remains from the Early Triassic of Europe (Buntsandstein) are all 11 from continental deposits (e.g. Brinkmann et al., 2010).

12 Early Triassic osteichthyans from South China are reported to be scarce (Tong *et al.*, 13 2006; Tintori et al., 2013) and they have been much less extensively studied than those from 14 the Middle Triassic of this region. In western North America, Early Triassic fishes have been 15 described from British Columbia and Alberta, Canada, as well as from Idaho and Nevada, 16 USA (Brinkmann et al., 2010). A relatively diverse fish fauna is known from British 17 Columbia, where new discoveries are still being made (e.g. Wendruff & Wilson, 2012). Early 18 Triassic fishes from other North American localities have either received very little research 19 interest during recent decades (Alberta and Idaho; e.g. Schaeffer & Mangus, 1976; Romano et 20 al., 2012) or were only recently discovered (Nevada; Brinkmann et al., 2010; Ware et al., 2011). Peak diversity at mid-palaeolatitudes in the Early Triassic could be affected by the 21 22 presence of Lagerstätten (Greenland, NW Madagascar, Spitsbergen, Fig. 1), although their 23 impact on total diversity is minor due to many shared taxa (see above). In conclusion, bony 24 fish diversity may have been lower within equatorial latitudes than at mid-latitudes during the 25 Early Triassic (Fig. 9), but this discrepancy could at least partly be the result of differences in

sampling and research effort. An 'equatorial eclipse' during the Early Triassic is not evident
 from our data.

3 A number of additional biotic crises occurred in the aftermath of the end-Permian event, 4 starting with the end-Griesbachian extinction. This event severely affected conodonts 5 (Orchard, 2007) but had no discernible effect on osteichthyan (Figs 4-6) and chondrichthyan 6 fishes (Fig. 12; Koot, 2013). Probably the most important crisis during the Early Triassic is 7 the end-Smithian event (Galfetti et al., 2007a, b) less than two million years after the end-8 Permian mass extinction. This event strongly decimated nekto-pelagic clades such as 9 conodonts (Orchard, 2007) and ammonoids (e.g. Brayard et al., 2006, 2009), but seemingly 10 less so benthic communities (Song et al., 2011; Hofmann et al., 2013). The end-Smithian 11 extinction is associated with pronounced shifts in the carbon and oxygen isotope records as 12 well as changes in the net water balance between precipitation and evaporation, all of which 13 point to severe climatic changes possibly related to a late eruptive phase of the Siberian Traps 14 (e.g. Payne et al., 2004; Galfetti et al., 2007a, b; Ivanov et al., 2013; Romano et al., 2013). 15 After the diversification of osteichthyans at the onset of the Triassic and relatively stable 16 fish associations between the Griesbachian and Smithian, a first diversity minimum is noted 17 in the Spathian (Fig. 4). This low, however, is a consequence of the rarity of well-dated 18 Spathian occurrences (also see Tintori et al., 2013) and does not necessarily reflect a crisis 19 across the Smithian-Spathian boundary. Strikingly, the taxonomic compositions of 20 Griesbachian to Smithian bony fish communities differs from that of Middle Triassic 21 associations suggesting that an important turnover occurred in between, analogous to other 22 groups (e.g. Crasquin-Soleau et al., 2007; Hautmann et al., 2013; Scheyer et al., 2014). 23 Whether the Early-Middle Triassic taxonomic turnover within Osteichthyes was gradual or 24 whether extinction events such as the end-Smithian or end-Spathian crises (e.g. Orchard, 25 2007; Brayard et al., 2009) were pivotal triggers remains unknown. If we want to improve our 26 understanding of the evolution of Triassic osteichthyans, filling the gap in their fossil record

during the Spathian, an interval that lasted ~3 million years and thus more than half of the
 Early Triassic, becomes an important task.

3

4 (b) The Triassic actinopterygian revolution

5

6 The Middle Triassic is characterized by the appearance of several new osteichthyan taxa 7 (Figs 4–8). However, this major origination event may potentially have initiated already 8 during the Spathian for reasons outlined above. A diversity increase is coincidentally seen in 9 marine reptiles, for which a link to sea-level rise was postulated (Kelley et al., 2012). This 10 explanation may also apply to fishes (Thomson, 1977). The second Triassic diversification of 11 bony fishes occurred in the marine environment mainly at low-palaeolatitudes, with many 12 taxa possibly originating in the South China region (Lombardo *et al.*, 2011), and in the 13 freshwater realm mostly at mid-palaeolatitudes (Fig. 9). Freshwater genus richness at mid-14 palaeolatitudes remained high throughout the Triassic. This pattern may be explained by the 15 Pangaean palaeogeographical configuration, because continental surface area was largest at 16 mid-palaeolatitudes in both hemispheres during the Triassic (cf. Fig. 1) and freshwater fish 17 diversity generally increases with area (e.g. Lévêque et al., 2008). Scarcity of fishes in low-18 palaeolatitudinal Pangaea in the Early and Middle Triassic could be related to prevalence of 19 generally more arid conditions in this zone (Ziegler et al., 2003) and perhaps sampling bias 20 [see Section II(4)].

In the marine realm, the second Triassic bloom within Osteichthyes spawned a plethora of small-sized consumers, most of which belong to 'Subholostei' and Neopterygii. In fact, the statistically significant decrease of osteichthyan body size (marine, global) across the Early Triassic-Anisian boundary is a direct consequence of the diversifications among these groups (Figs 7, 10, Supplementary Table B6). Although there are differences in palaeogeographic sampling [Early Triassic data mostly come from mid-palaeolatitudes and Anisian data largely

from low-palaeolatitudes, see Section III(4)] and body size tends to increase with latitude as a
 function of temperature and productivity (e.g. Fisher, Frank & Leggett, 2010), the drop in
 body size between the Early and Middle Triassic is unlikely to be a sampling artefact because
 it is evident across palaeolatitudes (Fig. 11).

5 In the freshwater domain, body length increases across the Early Triassic-Anisian 6 boundary due to a rise in size and diversity of 'palaeopterygians' and 'subholosteans' (Figs 7, 7 10, Supplementary Table B6). Interestingly, Triassic freshwater assemblages contain 8 relatively fewer Neopterygii than marine communities (Figs 7–8). Moreover, Middle and Late 9 Triassic osteichthyan freshwater faunas comprise forms exhibiting more archaic bauplans that 10 are either rarer or absent in contemporaneous marine associations (e.g. saurichthyids with 11 more extensive scale coverage, Romano et al., 2012), suggesting that Triassic fluvio-12 lacustrine environments may have served as *refugia* for such groups. The Early Mesozoic 13 radiation of bony fishes led to a pronounced diversity peak in the Middle Triassic (Figs 4-6, 14 12), as the taxonomic richness of these fishes exceeded that of chondrichthyans for the first 15 time (Fig. 12; Sallan & Coates, 2010; Anderson et al., 2011; Koot, 2013) The Triassic 16 osteichthyan revolution was accompanied by the emergence of several evolutionary novelties 17 (Tintori et al., 2013), some of which convergently emerged in neopterygians and 18 'subholosteans' (Brough, 1936; Schaeffer, 1956). These include optimizations of the fins (e.g. 19 reduction of fin ray number and modification of the caudal fin) as well as of the dentition and 20 jaw apparatus (e.g. Brough, 1936; Schaeffer, 1956). These anatomical novelties are associated 21 with new modes of life such as gliding (Thoracopterus; Tintori et al., 2012), refined feeding 22 strategies (e.g. Perleididae; Bürgin, 1996) and new reproductive strategies including internal 23 fertilisation and (ovo-)viviparity. While changes in the feeding and locomotory apparatus 24 mainly pertain to ray-fins, new reproductive strategies are observed within both 25 actinopterygians (species of Saurichthys and Peltopleurus; e.g. Bürgin, 1990; Lombardo, 26 1999; Renesto & Stockar, 2009) and actinistians (Luopingichthys eurylacrimalis; Wen et al.,

1 2013). Note that although saurichthyids are known since the latest Permian, evidence for 2 viviparity has yet only been documented for some Middle and Late Triassic species (Romano 3 et al., 2012). Viviparity evolved numerous times independently within vertebrates 4 (Blackburn, 2014), amongst others also in some Palaeozoic fishes (placoderms, 5 chondrichthyans). However, within osteichthyans evidence for viviparity older than the 6 Middle Triassic is still lacking. The case of viviparity in the Early Triassic Birgeria nielseni is 7 erroneous (Bürgin, 1990; Lombardo & Tintori, 2005). 8

9 (c) "Carnian Pluvial Event"

10

11 The transition between the Middle and Late Triassic is characterized by a drop in genus 12 richness of marine bony fishes, followed by a decreasing diversity trend towards the Triassic-13 Jurassic boundary (Fig. 4). The onset and maximum of the Late Triassic diversity decline of 14 marine Osteichthyes roughly coincides with another biotic crisis known as the "Carnian 15 Pluvial Event" (Simms & Ruffell, 1989). This early Carnian (late Julian) event is associated 16 with sedimentological evidence for increased continental runoff in the western Tethys, which 17 suggests an episode of intensified rainfall during the generally arid Late Triassic (e.g. Simms 18 & Ruffell, 1989; Ziegler et al., 2003; Rigo et al., 2007). Although the existence of a "pluvial 19 event" has been questioned (Visscher et al., 1994), different lines of evidence indicate 20 climatic and biotic changes in the Carnian. These include a negative shift in the carbon 21 isotope record, possibly linked to eruptions of the Wrangellia Large Igneous Province in 22 western North America (Dal Corso et al., 2012), as well as extinctions among, for example, 23 crinoids (Hagdorn, 2011), conodonts (Rigo et al., 2007) and ammonoids (Brayard et al., 24 2009), and a decline of carbonate platforms (Rigo et al., 2007 and references therein). 25 Previously dominant groups, such as anomodont therapsids (Fröbisch, 2008), became rare 26 shortly before the onset of the Late Triassic and several important Mesozoic and Cenozoic

clades diversified in the wake of the Carnian event, such as dinosaurs, scleractinian corals,
 and calcareous nannoplankton (Simms & Ruffell, 1989; Rigo *et al.*, 2007; Dal Corso *et al.*,
 2012).

4 Among marine bony fishes, declining diversity across the Ladinian-Carnian boundary is 5 seen in all groups, yielding total extinction rates of 51-62% (Fig. 4, Supplementary Table 6 B4), followed by a rise in genus richness of Teleosteomorpha during the Carnian and Norian 7 (Fig. 7). Osteichthyan communities of the Late Triassic are characterized by predominantly 8 'palaeopterygians' within the large consumers' guild and 'subholosteans' and neopterygians 9 among small-sized consumers (Fig. 10). Neopterygians underwent a radiation during the Late 10 Triassic and many taxa developed specialisations for durophagy (Tintori, 1998; Lombardo & 11 Tintori, 2005; Tintori et al., 2013). Late Triassic teleosteomorphs occupied both marine and 12 freshwater environments but their diversity remained relatively low until the Late Jurassic, 13 when also several of the extant clades emerged (Arratia, 2004, 2013).

14 The generally decreasing trend of marine osteichthyan diversity towards the end of the 15 Triassic may be linked to sea-level fall (cf. Kelley et al., 2012). However, reduced Late 16 Triassic bony fish diversity could also partially be the result of sampling bias, because these 17 fishes have been less extensively studied than Middle Triassic ones. According to Lombardo 18 & Tintori (2005), many specimens from the Norian Calcare di Zorzino (northern Italy) are yet 19 undescribed, and the diversity may have been much higher than currently known. To what 20 extent a long-term sea-level fall or the Carnian extinction modulated the concurrent 21 ichthyofaunal changes, including the Late Triassic neopterygian radiation, requires further 22 research.

23

24 (3) Permian-Triassic turnover of fish communities

25

1 The series of extinction and diversifications during the Permian-Triassic interval had a 2 profound impact on marine and continental life. Typical Late Palaeozoic faunas and floras 3 were replaced by those that characterise the Mesozoic and Cenozoic (Sepkoski, 1984; Benton, 4 2003; Erwin, 2006). Marine invertebrate communities of the Permian consisting mainly of 5 goniatitic ammonoids, brachiopods, rugose and tabulate corals as well as crinoid and blastoid 6 echinoderms were subsequently transformed into more modern assemblages dominated by 7 ceratitic ammonoids, bivalves, sea urchins, starfishes and scleractinian corals. Similarly, 8 turnover on land meant that tetrapod groups of the Late Palaeozoic gave way to 9 diversification of clades that ultimately evolved into dinosaurs and mammals. Fishes were no 10 exception to this trend.

11

12 The long-term change from low global genus richness of osteichthyans in the Permian to 13 higher levels in the Triassic is in direct contrast to the inverted diversity patterns documented 14 for chondrichthyans (e.g. Koot, 2013). The latter exhibit elevated diversity (at genus level and 15 higher taxonomic ranks) during the Lower and Middle Permian, but show severe extinctions 16 during the end-Guadalupian crisis followed by lower diversity in the Triassic (Fig. 12; Koot, 17 2013). The nature of the fossil record of these two fish clades is different, consisting mostly of 18 ichthyoliths (teeth, denticles or fin/cephalic spines) in Chondrichthyes and articulated material 19 in Osteichthyes (bony fish taxa based on isolated remains are normally considered invalid). 20 Regardless of these differences, however, comparison of the opposing diversity trends of 21 cartilaginous and bony fishes suggests a biological reason behind the patterns observed in 22 their fossil record. Since the radiation of Osteichthyes in the Devonian, global genus-level 23 diversity of actinopterygians remained below that of chondrichthyans and other fishes. 24 Sarcopterygians were diverse in the Devonian but did not recover taxonomically after the end-Devonian Hangenberg extinction (Klug et al., 2010; Sallan & Coates, 2010; Anderson et 25 26 al., 2011). In contrast to the global trends at genus level, however, species richness of

1 actinopterygians was locally higher than that of chondrichthyans during the Carboniferous 2 (Sallan & Coates 2010). Despite their low global genus diversity, Carboniferous and Permian 3 bony fishes already display different body shapes and sizes as well as several trophic 4 adaptations (e.g. Schaumberg, 1977; Esin, 1997; Minikh & Minikh, 2009; Boy & Schindler, 5 2012). Permian bony fishes include general forms, as well as deep-bodied taxa (e.g. 6 *Platysomus*, *Dorypterus*, *Kargalichthys*, *Paranaichthys*) and the enigmatic 7 Discordichthyiformes (Schaumberg, 1977; Minikh & Minikh, 2009). However, advanced 8 anatomical features like those seen in Triassic and later taxa (e.g. viviparity, refined feeding 9 and locomotory apparatus; Brough, 1936; Schaeffer, 1956; Romano et al., 2012; Wen et al., 10 2013; Xu et al., 2013; Blackburn, 2014) are either rare or absent among Palaeozoic 11 osteichthyans.

12 The Triassic saw the rise of the Neopterygii, a group that includes nearly all present-day 13 osteichthyans and more than half of all vertebrates (Nelson, 2006). Although neopterygians 14 experienced their first radiation in the Middle and Late Triassic (Figs 7-8; McCune & 15 Schaeffer, 1986; Tintori, 1998; Arratia, 2004, 2013; Tintori et al., 2013), fossil data and 16 molecular clock analyses suggest that they evolved in the Late Palaeozoic (Hurley et al., 17 2007; Near et al., 2012; Betancur-R. et al., 2013; Sallan, 2014). Palaeozoic neopterygians 18 were either rare and/or lived in environments with a poor fossil record. Neopterygian key 19 innovations in the feeding and locomotory apparatus (e.g. mobile maxilla, reduction of fin 20 rays), some of which convergently evolved in 'Subholostei' (Brough, 1936; Schaeffer, 1956), 21 probably led to the great success of ray-finned fishes during post-Palaeozoic times. The fact 22 that 'subholosteans' and neopterygians (and likely also actinistians and marine tetrapods; 23 Scheyer et al., 2014) did not experience taxonomic blooms before the demise of several 24 chondrichthyan clades (e.g. Petalodontiformes, Eugeneodontiformes) between the late 25 Guadalupian and Early Triassic (Fig. 12, Koot, 2013) highlights the importance of Permian-26 Triassic extinctions for this transformation of fish faunas, suggesting that macroevolutionary

trajectories in fishes mostly follow the Court Jester model (e.g. Benton, 2009). The associated
climatic and environmental changes and profound reorganisation of ecosystems allowed many
bony fish taxa to exploit new niches ultimately leading to the great Mesozoic radiation of
Osteichthyes. The processes and mechanisms involved in this ichthyofaunal turnover deserve
further research.

7 V. CONCLUSIONS

8

9 (1) Diversity of bony fishes changed from low during the Permian to relatively higher levels
10 in the Triassic as a result of origination events in the Early and Middle Triassic. In the marine
11 realm, these diversification pulses probably represent true biological signals and are not
12 merely the results of the Lagerstätten effect.

13

(2) During the Triassic revolution of Osteichthyes, many new taxa appeared exhibiting
evolutionary novelties in their locomotory and feeding apparatus, and for the first time in the
evolutionary history of the group, some species evolved viviparity. In the Early Triassic,
diversifications are noted for both sarcopterygians and actinopterygians, whereas in the
Middle Triassic the increase in taxonomic richness mainly pertains to Actinopterygii
('Subholostei', Neopterygii).

20

(3) For most of the Permian-Triassic, the marine and global palaeolatitudinal diversity
gradients resemble present-day ones, with highest diversity found around the equator and
lowest towards the poles. However, during the Early Triassic, osteichthyan genus richness
was highest at mid-palaeolatitudes, whereas equatorial diversity was reduced compared to
Late Permian and Middle Triassic values. Although there could be an underlying biological

signal, this pattern is largely related to incomplete knowledge of low-palaeolatitudinal faunas
 of Early Triassic age.

3

4 (4) Body size changes in Osteichthyes during the Permian-Triassic include a significant size 5 increase of marine 'Palaeopterygii' across the Middle-Late Permian boundary and these fishes 6 remain among the large apex predators during the Triassic. A significant body size reduction 7 of marine bony fishes is documented across the Early Triassic-Anisian boundary as a result of 8 diversifications of small taxa (mainly 'Subholostei' and Neopterygii). Dietary studies on 9 Permian-Triassic fishes are wanting and could further contribute to a better understanding of 10 macroecological changes during this critical interval. 11 12 (5) Composition of fish faunas changed profoundly during the Permian-Triassic interval: 13 typical chondrichthyan-rich communities of the Permo-Carboniferous diminished during the 14 end-Guadalupian crisis and, during the aftermath of the end-Permian mass extinction, piscine 15 diversity became osteichthyan-dominated – a condition that has prevailed up until today. This 16 ichthyofaunal turnover is a consequence of the series of extinction events during the Permian 17 and Triassic, each associated with severe climatic and environmental changes. 18 19 20 21 **VI. ACKNOWLEDGEMENTS** 22 23 We deeply acknowledge past and present researchers, whose valuable contributions to the 24 taxonomy and stratigraphy of fishes made this study possible. We thank Peter A. Hochuli, 25 Borhan Bagherpour, Christian Klug, David Ware, Heinz Furrer, Richard Hofmann, Morgane 26 Brosse, Maximiliano and Carole Meier, and Torsten M. Scheyer for discussions and Beat

1	Scheffold for drawings (all Palaeontological Institute and Museum, University of Zurich). We
2	are thankful to Adriana López-Arbarello (Bayerische Staatssammlung für Paläontologie,
3	Munich, Germany), John Clarke and Matt Friedman (University of Oxford, UK), Andrew R.
4	C. Milner (St. George Dinosaur Discovery Site at Johnson Farm, St. George, Utah, USA),
5	Kenneth De Baets (University of Erlangen-Nuremberg, Germany), Daisuke B. Koyabu
6	(University of Tokyo, Japan), Jim Jenks (West Jordan, Utah, USA), Jonathan L. Payne
7	(Stanford University, California, USA), Spencer G. Lucas (New Mexico Museum of Natural
8	History and Science, Albuquerque, New Mexico, USA) and Stanislav Štamberg (Regional
9	Museum of Eastern Bohemia, Hradec Králové, Czech Republic) for material and discussions.
10	We also thank two anonymous reviewers for constructive comments. Support by the Swiss
11	National Science Foundation (project numbers 120311/135075 and 144462 to W. B. and H.
12	B.) and the ANR project AFTER (ANR-13-JS06-0001-01 to A. B.) are deeply acknowledged.
13	
14	
15	VII. REFERENCES
16	
17	ACCORDI, B. (1955). Archaeolepidotus leonardii n. gen. n. sp. e altri pesci permo-werfeniani
18	delle Dolomiti. Memorie degli Istituti di Geologia e Mineralogia dell'Università di
19	<i>Padova</i> 19 , 3–28.
20	AGASSIZ, L. (1833–1843). Recherches sur les poissons fossiles II, 2. Petitpierre, Neuchâtel
21	(with supplements).
22	ALLISON, P. A. & BRIGGS, D. E. G. (1993). Paleolatitudinal sampling bias, Phanerozoic
23	species diversity, and the end-Permian extinction. Geology 21, 65-68.
24	ANDERSON, P. S. L., FRIEDMAN, M., BRAZEAU, M. D. & RAYFIELD, E. J. (2011). Initial
25	radiation of jaws demonstrated stability despite faunal and environmental change.
26	<i>Nature</i> 476 , 206–209.

1	ARRATIA, G. (2004). Mesozoic halecostomes and the early radiation of teleosts. In Mesozoic
2	Fishes 3, Systematics, Paleoenvironments and Biodiversity (eds G. ARRATIA and A.
3	TINTORI), p. 279. Dr. Friedrich Pfeil, München.
4	ARRATIA, G. (2013). Morphology, taxonomy, and phylogeny of Triassic pholidophorid fishes
5	(Actinopterygii, Teleostei). Journal of Vertebrate Paleontology 33, 1-138.
6	BATTEN, R. L. (1973). The vicissitudes of the gastropods during the interval of Guadalupian-
7	Ladinian time. In The Permian and Triassic systems and their mutual boundary,
8	Canadian Society of Petroleum Geologists, Memoir 2 (eds A. LOGAN and L. V. HILLS),
9	596–607.
10	BELLWOOD, D. R., HOEY, A. S., BELLWOOD, O. & GOATLEY, C. H. R. (2014). Evolution of
11	long-toothed fishes and the changing nature of fish-benthos interactions on coral reefs.
12	Nature Communications 5, 3144 (doi: 10.1038/ncomms4144).
13	BENTON, M. J. (1993). The Fossil Record 2. Chapman & Hall, London.
14	BENTON, M. J. (1998). The quality of the fossil record of the vertebrates. In The adequacy of
15	the fossil record (eds S. K. DONOVAN and C.R.C. PAUL), p. 269. Wiley, New York.
16	BENTON, M. J. (2003). When Life Nearly Died. Thames & Hudson, London.
17	BENTON, M. J. (2009). The Red Queen and the Court Jester: Species diversity and the role of
18	biotic and abiotic factors through time. Science 323, 728–732.
19	BENTON, M. J. & KING, P. W. (1989). Mass extinctions among tetrapods and the quality of the
20	fossil record. Philosophical Transactions of the Royal Society of London B 325, 369–386.
21	BENTON, M. J. & NEWELL, A. J. (2013). Impacts of global warming on Permo-Triassic
22	terrestrial ecosystems. Gondwana Research
23	(http://dx.doi.org/10.1016/j.gr.2012.12.010).
24	BENTON, M. J., DUNHILL, A. M., LLOYD, G. T. & MARX, F. G. (2011). Assessing the quality of
25	the fossil record: Insights from vertebrates. In Comparing the Geological and Fossil

1	Records: Implications for Biodiversity Studies. Geological Society, London, Special
2	Publications 358 (eds A. J. MCGOWAN and A. B. SMITH), 63–94.
3	Benton, M. J., Zhang, Q., Hu, S., Chen, ZQ., Wen, W., Liu, J., Huang, J., Zhou, C., Xie,
4	T., TONG, J., CHOO, B. (2013). Exceptional vertebrate biotas from the Triassic of China,
5	and the expansion of marine ecosystems after the Permo-Triassic mass extinction.
6	Earth-Science Reviews 125, 199–243.
7	BETANCUR-R., R., BROUGHTON, R. E., WILEY, E. O., CARPENTER, K., LÓPEZ, J. A., LI, C.,
8	HOLCROFT, N. I., ARCILA, D., SANCIANGCO, M., CURETON II, J. C., ZHANG, F., BUSER, T.,
9	CAMPBELL, M. A., BALLESTEROS, J. A., ROA-VARON, A., WILLIS, S., BORDEN, W. C.,
10	Rowley, T., Reneau, P. C., Hough, D. J., Lu, G., Grande, T., Arratia, G. & Ortí, G.
11	(2013). The tree of life and a new classification of bony fishes. PLoS Currents Tree of
12	Life [last modified: 2013 Jun 3]. Edition 1 (doi:
13	10.1371/currents.tol.53ba26640df0ccaee75bb165c8c26288).
14	BLACKBURN, D. G. (2014). Evolution of vertebrate viviparity and specializations for fetal
15	nutrition: a quantitative and qualitative analysis. Journal of Morphology (doi:
16	10.1002/jmor.20272).
17	BLIECK, A. (2011). From adaptive radiations to biotic crises in Palaeozoic vertebrates: A
18	geobiological approach. Geologica Belgica 14, 203–227.
19	BOY, J. A. & SCHINDLER, T. (2012). Ökostratigraphie des Rotliegend. In Stratigraphie von
20	Deutschland, X. Rotliegend. Teil I: Innervariscische Becken. Schriftenreihe der
21	Deutschen Gesellschaft für Geowissenschaften 61 (ed. DEUTSCHE STRATIGRAPHISCHE
22	Kommission), 143–160.
23	BRAYARD, A., BUCHER, H., ESCARGUEL, G., FLUTEAU, F., BOURQUIN, S. & GALFETTI, T.
24	(2006). The Early Triassic ammonoid recovery: Paleoclimatic significance of diversity
25	gradients. Palaeogeography, Palaeoclimatology, Palaeoecology 239, 374–395.

1	BRAYARD, A., ESCARGUEL. G. & BUCHER, H. (2007). The biogeography of Early Triassic
2	ammonoid faunas: Clusters, gradients and networks. Geobios 40, 749-765.
3	BRAYARD, A., ESCARGUEL, G., BUCHER, H., MONNET, C., BRÜHWILER, T., GOUDEMAND, N.,
4	GALFETTI, T., & GUEX, J. (2009). Good genes and good luck: Ammonoid diversity and
5	the end-Permian mass extinction. Science 325, 1118–1121.
6	BRAYARD, A., NÜTZEL, A., STEPHEN, D. A., BYLUND, K. G., JENKS, J. & BUCHER, H. (2010).
7	Gastropod evidence against the Early Triassic Lilliput effect. Geology 38, 147–150.
8	BRAYARD, A., VENNIN, E., OLIVIER, N., BYLUND, K. G., JENKS, J., STEPHEN, D. A., BUCHER,
9	H., HOFMANN, R., GOUDEMAND, N. & ESCARGUEL, G. (2011). Transient metazoan reefs
10	in the aftermath of the end-Permian mass extinction. Nature Geoscience 4,693-697.
11	BRINKMANN, W., ROMANO, C., BUCHER, H., WARE, D. & JENKS, J. (2010).
12	Palaeobiogeography and stratigraphy of advanced gnathostomian fishes
13	(Chondrichthyes and Osteichthyes) in the Early Triassic and from selected Anisian
14	localities (Report 1863–2009). Zentralblatt für Geologie und Paläontologie, Teil 2,
15	2009 , 765–812.
16	BROUGH, J. (1936). On the evolution of bony fishes during the Triassic period. Biological
17	<i>Reviews</i> 11 , 385–405.
18	BRÜHWILER, T., BUCHER, H., BRAYARD, A. & GOUDEMAND, N. (2010). High-resolution
19	biochronology and diversity dynamics of the Early Triassic ammonoid recovery: The
20	Smithian faunas of the Northern Indian margin. Palaeogeography, Palaeoclimatology,
21	<i>Palaeoecology</i> 297 , 491–501.
22	BÜRGIN, T. (1990). Reproduction in Middle Triassic actinopterygians; complex fin structures
23	and evidences of viviparity in fossil fishes. Zoological Journal of the Linnean Society
24	100, 379–391.
25	BÜRGIN, T. (1996). Diversity in the feeding apparatus of perleidid fishes (Actinopterygii)
26	from the Middle Triassic of Monte San Giorgio (Switzerland). In Mesozoic Fishes,

- Systematics and Paleoecology (eds G. ARRATIA and G. VIOHL), p. 555. Dr. Friedrich
 Pfeil, München.
- CASANE, D. & LAURENTI, P. (2013). Why coelacanths are not 'living fossils'. *Bioessays* 35,
 332–338 (doi:10.1002/bies.201200145).
- 5 CAVIN, L. (2002). Effects of the Cretaceous-Tertiary boundary event on bony fishes. In
- 6 *Geological and Biological Effects of Impact Events* (eds E. BUFFETAUT and C.
- 7 KOEBERL), p. 141. Springer, Heidelberg.
- 8 CAVIN, L., FOREY, P. L. & LÉCUYER, C. (2007). Correlation between environment and Late
- 9 Mesozoic ray-finned fish evolution. *Palaeogeography, Palaeoclimatology,*
- 10 *Palaeoecology* **245**, 353–367.
- 11 CAVIN, L., FURRER, H. & OBRIST, C. (2013). New coelacanth material from the Middle
- Triassic of eastern Switzerland, and comments on the taxic diversity of actinistans.
 Swiss Journal of Geoscience 106, 161–177.
- CHAO, A. & JOST, L. (2012). Coverage-based rarefaction and extrapolation: standardizing
 samples by completeness rather than size. *Ecology* 93, 2533–2547.
- 16 CHEN, B., JOACHIMSKI, M. M., SHEN, S.-Z., LAMBERT, L. L., LAI, X.-L., WANG, X.-D., CHEN,
- 17 J. & YUAN, D. X. (2013). Permian ice volume and palaeoclimate history: Oxygen
- 18 isotope proxies revisited. *Gondwana Research* **24**, 77–89.
- 19 CLAPHAM, M. E. & PAYNE, J. L. (2011). Acidification, anoxia, and extinction: A multiple
- 20 logistic regression analysis of extinction selectivity during the Middle and Late Permian.
- 21 *Geology* **39**, 1059–1062.
- 22 CLAPHAM, M. E., SHEN, S.-Z. & BOTTJER, D. J. (2009). The double mass extinction revisited:
- 23 reassessing the severity, selectivity, and causes of the end-Guadalupian biotic crisis
- 24 (Late Permian). *Paleobiology* **35**, 32–50.

1	COATES, M. I. (1999). Endocranial preservation of a Carboniferous actinopterygian from
2	Lancashire, UK, and the interrelation-ships of primitive actinopterygians. Philosophical
3	Transactions of the Royal Society of London B 354, 435–462.
4	COLWELL, R. K., CHAO, A., GOTELLI, N. J., LIN, SY., MAO, C. X., CHAZDON, R. L. &
5	LONGINO, J. T. (2012). Models and estimators linking individual-based and sample-
6	based rarefaction, extrapolation and comparison of assemblages. Journal of Plant
7	<i>Ecology</i> 5 , 3–21.
8	CRASQUIN-SOLEAU, S., GALFETTI, T., BUCHER, H., KERSHAW, S. & FENG, Q. (2007). Ostracod
9	recovery in the aftermath of the Permian-Triassic crisis: Palaeozoic-Mesozoic turnover.
10	Hydrobiologia 585 , 13–27.
11	DAGYS, A. S. (1988). Major features of the geographic differentiation of Triassic ammonoids.
12	In Cephalopods – Present and Past (eds J. WIEDMANN and J. KULLMANN), p. 341.
13	Schweizerbart'sche Verlagsbuchhandlung, Stuttgart.
14	DAL CORSO, J., MIETTO, P., NEWTON, R. J., PANCOST, R. D., PRETO, N., ROGHI, G. &
15	WIGNALL, P. B. (2012). Discovery of a major negative δ^{13} C spike in the Carnian (Late
16	Triassic) linked to the eruption of Wrangellia flood basalts. <i>Geology</i> 40 , 79–82.
17	DENG, Z. & KONG, L. (1984). Middle Triassic corals and sponges from southern Guizhou and
18	eastern Yunnan. Acta Palaeontologica Sinica 23, 489–503.
19	ERWIN, D. H. (1996). Understanding biotic recoveries: Extinction, survival and preservation
20	during the end-Permian mass extinction. In Evolutionary Paleobiology (eds D.
21	JABLONSKI, D. H. ERWIN and J. H. LIPPS), p. 398. University of Chicago Press, Chicago.
22	ERWIN, D. H. (2006). Extinction. How Life on Earth Nearly Ended 250 Million Years Ago.
23	Princeton University Press, Princeton and Oxford.
24	ERWIN, D. H., & DROSER, M. L. (1993). Elvis taxa. Palaios 8, 623–624.
25	ESIN, D. N. (1997). Peculiarities of trophic orientation changes in palaeoniscoid assemblages
26	from the Upper Permian of the European part of Russia. Modern Geology 21, 185–195.

1	FISHER, J. A. D., FRANK, K. T. & LEGGETT, W. C. (2010). Global variation in marine fish body				
2	size and ist role in biodiversity-ecosystem functioning. Marine Ecology Progress Series				
3	405 , 1–13.				
4	FOREY, P. L. (1998). History of the coelacanth fishes. Chapman & Hall, London.				
5	FOSTER, W. J. & TWITCHETT, R. J. (2014). Functional diversity of marine ecosystems after the				
6	Late Permian mass extinction event. Nature Geoscience 7, 233–238.				
7	FRAISER, M. L. & BOTTJER, D. J. (2004). The non-actualistic Early Triassic gastropod fauna: a				
8	case study of the Lower Triassic Sinbad Limestone Member. Palaios 19, 259–275.				
9	FRIEDMAN, M. (2012). Parallel evolutionary trajectories underlie the origin of giant				
10	suspension-feeding whales and bony fishes. Proceedings of the Royal Society B 279,				
11	944–951.				
12	FRIEDMAN, M. & SALLAN, L. C. (2012). Five hundred million years of extinction and				
13	recovery: A Phanerozoic survey of large-scale diversity patterns in fishes.				
14	Palaeontology 55, 707–742.				
15	FRÖBISCH, J. (2008). Global taxonomic diversity of anomodonts (Tetrapoda, Therapsida) and				
16	the terrestrial rock record across the Permian-Triassic boundary. PLoS ONE 3, e3733.				
17	FRÖBISCH, J. (2013). Vertebrate diversity across the end-Permian mass extinction —				
18	Separating biological and geological signals. Palaeogeography, Palaeoclimatology,				
19	<i>Palaeoecology</i> 372 , 50–61.				
20	GALFETTI, T., BUCHER, H., BRAYARD, A., HOCHULI, P. A., WEISSERT, H., GUODUN, K.,				
21	ATUDOREI, V. & GUEX, J. (2007a). Late Early Triassic climate change: Insights from				
22	carbonate carbon isotopes, sedimentary evolution and ammonoid paleobiogeography.				
23	Palaeogeography, Palaeoclimatology, Palaeoecology 243, 394–411.				
24	GALFETTI, T., HOCHULI, P. A., BRAYARD, A., BUCHER, H., WEISSERT, H. & VIGRAN, J. O.				
25	(2007b). Smithian-Spathian boundary event: Evidence for global climatic change in the				
26	wake of the end-Permian biotic crisis. Geology 35, 291–294.				

1	GARASSINO, A. & PASINI, G. (2002). Studies on Permo-Trias of Madagascar. 5. Ambilobeia
2	karojoi n. gen., n. sp. (Crustacea, Decapoda) from the Lower Triassic (Olenekian) of
3	Ambilobe region (NW Madagascar). Atti della Società italiana di Scienze naturali e del
4	Museo Civico di Storia naturale di Milano 143, 95–104.
5	GARDINER, B. G., SCHAEFFER, B. & MASSERIE, J. A. (2005). A review of lower
6	actinopterygian phylogeny. Zoological Journal of the Linnean Society 144, 511-525.
7	GLENISTER, B. F. & FURNISH, W. M. (1981). Permian ammonoids. In The Ammonoidea: The
8	evolution, classification, mode of life and geological usefulness of a major fossil group.
9	The Systematics Association Special Volume 18 (eds M. R. HOUSE and J. R. SENIOR),
10	49–64.
11	GOTELLI, N. & COLWELL, R. K. (2011). Estimating species richness. In Frontiers in
12	Measuring Biodiversity (eds A. E. Magurran and B. J. MCGILL). Oxford University
13	Press, New York.
14	GRADSTEIN, F., OGG, J., SCHMITZ, M. & OGG, G. (2012). The Geologic Time Scale 2012, 1–2.
15	Elsevier, Amsterdam.
16	GRANDE, L. (2010). An empirical synthetic pattern study of gars (Lepisosteiformes) and
17	closely related species, based mostly on skeletal anatomy: the resurrection of Holostei.
18	American Society of Ichthyology and Herpetology, Special Publication 6, 1–871.
19	GROVES, J. R. & WANG, Y. (2013). Timing and size selectivity of the Guadalupian (Middle
20	Permian) fusulinoidean extinction. Journal of Paleontology 87, 183–196.
21	HAGDORN, H. (2011). Triassic: The crucial period of post-Palaeozoic crinoid diversification.
22	Swiss Journal of Palaeontology 130, 91–112.
23	HAMMER, Ø., HARPER, D. A. T. & RYAN, P. D. (2001). Paleontological statistics software
24	package for education and data analysis. <i>Palaeontologia Electronica</i> 4 , 1–9.

1	HAUTMANN, M., SMITH, A. B., MCGOWAN, A. J. & BUCHER, H. (2013). Bivalves from the
2	Olenekian (Early Triassic) of south-western Utah: Systematics and evolutionary
3	significance. Journal of Systematic Palaeontology 11, 263–293.
4	HERMANN, E., HOCHULI, P. A., BUCHER, H., BRÜHWILER, T., HAUTMANN, M., WARE, D.,
5	WEISSERT, H., ROOHI, G., YASEEN, A. & UR-REHMAN, K. (2012). Climatic oscillations at
6	the onset of the Mesozoic inferred from palynological records from the North Indian
7	Margin. Journal of the Geological Society 169, 227–237.
8	HERMANN, E., HOCHULI, P. A., MÉHAY, S., BUCHER, H., BRÜHWILER, T., WARE, D.,
9	HAUTMANN, M., ROOHI, G., UR-REHMAN, K. & YASEEN, A. (2011). Organic matter and
10	palaeoenvironmental signals during the Early Triassic biotic recovery: The Salt Range
11	and Surghar Range records. Sedimentary Geology 234, 19-41.
12	HEYLER, D. (2000). Les actinoptérygiens stéphaniens et autuniens du Massif Central (France)
13	dans les collections du M.N.H.N. (Paris) et du Muséum d'Autun: compléments, mises
14	au point, bilan. Bulletin de la Société d'Histoire Naturelle d'Autun 169, 7–44.
15	HANNISDAL, B. & PETERS, S. E. (2011). Phanerozoic Earth system evolution and marine
16	biodiversity. Science 334, 1121–1124.Hofmann, R., HAUTMANN, M., BRAYARD, A.,
17	NÜTZEL, A., BYLUND, K. G., JENKS, J. F., VENNIN, E., OLIVIER, N. & BUCHER, H. (2013).
18	Recovery of benthic marine communities from the end-Permian mass extinction at the
19	low latitudes of eastern Panthalassa. Palaeontology (doi: 10.1111/pala.12076).
20	HURLEY, I. A., MUELLER, R. L., DUNN, K. A., SCHMIDT, E. J., FRIEDMAN, M., HO, R. K.,
21	PRINCE, V. E., YANG, Z., THOMAS, M. G. & COATES, M. I. (2007). A new time-scale for
22	ray-finned fish evolution. Proceedings of the Royal Society B 274, 489–498.
23	ISOZAKI, Y., KAWAHATA, H. & OTA, A. (2007). A unique carbon isotope record across the
24	Guadalupian–Lopingian (Middle–Upper Permian) boundary in mid-oceanic paleo-atoll
25	carbonates: The high-productivity "Kamura event" and its collapse in Panthalassa.
26	Global and Planetary Change 55, 21–38.

1	IVANOV, A. V., HE, H., YAN, L., RYABOV, V. V., SHEVKO, A. Y., PALESSKII, S. V.,
2	NIKOLAEVA, I. V. (2013). Siberian Traps large igneous province: Evidence for two flood
3	basalt pulses around the Permo-Triassic boundary and in the Middle Triassic, and
4	contemporaneous granitic magmatism. Earth Science Reviews 122, 58-76.
5	JANVIER, P. (1996). Early Vertebrates. Clarendon Press, Oxford.
6	JANVIER, P. (2007). Living primitive fishes and fishes from deep time. In Primitive Fishes.
7	Fish Physiology 26 (eds D. J. MCKENZIE, A. P. FARRELL and C. J. BRAUNER), 1–51.
8	JUBB, R. A. & GARDINER, B. G. (1975). A preliminary catalogue of identifiable fossil fish
9	material from southern Africa. Annals of the South African Museum 67, 381-440.
10	KAUFFMAN, E. G. & HARRIES, P. J. (1996). The importance of crisis progenitors in recovery
11	from mass extinction. In Biotic Recovery from Mass Extinction Events. Geological
12	Society Special Publication 102 (ed. M. B. HART), 15–39.
13	KELLEY, N. P., MOTANI, R., JIANG, DY., RIEPPEL, O., SCHMITZ, L. (2012). Selective
14	extinction of Triassic marine reptiles during long-term sea-level changes illuminated by
15	seawater strontium isotopes. Palaeogeography, Palaeoclimatology, Palaeoecology
16	(http://dx.doi.org/10.1016/j.palaeo.2012.07.026).
17	KIPARISOVA, L. D. & POPOV, J. N. (1956). Subdivision of the lower series of the Triassic
18	System into stages [Raschleneniye nizhnego otdela triasovoj sistemy na yarusy].
19	Doklady Akademii Nauk SSSR 109, 842–845.
20	Klug, C., Kröger, B., Kiessling, W., Mullins, G. L., Servais, T., Frýda, J., Korn, D. &
21	TURNER, S. (2010). The Devonian nekton revolution. Lethaia 43, 465–477.
22	KOOT, M. B. (2013). Effects of the Late Permian mass extinction on chondrichthyan
23	palaeobiodiversity and distribution patterns. PhD thesis: Plymouth University
24	(http://pearl.plymouth.ac.uk/handle/10026.1/1584).

1	KRIWET, J. & BENTON, M. J. (2008). Neoselachian (Chondrichthyes, Elasmobranchii)
2	diversity across the Cretaceous–Tertiary boundary. Palaeogeography,
3	Palaeoclimatology, Palaeoecology 214, 181–194.
4	KURAKU, S. & KURATANI, S. (2006). Time scale for cyclostome evolution inferred with a
5	phylogenetic diagnosis of hagfish and lamprey cDNA sequences. Zoological Science 23,
6	1053–1064.
7	LEHMAN, JP. (1952). Etude complémentaire des poissons de l'Eotrias de Madagascar.
8	Kungliga Svenska Vetenskapsakademiens Handlingar 4 , 1–192.
9	LEVEQUE, C., OBERDORFF, T., PAUGY, D., STIASSNY, M. L. J. & TEDESCO, P. A. (2008). Global
10	diversity of fish (Pisces) in freshwater. Hydrobiologia 595, 545-567.
11	LLOYD, G. T. & FRIEDMAN, M. (2013). A survey of palaeontological sampling biases in fishes
12	based on the Phanerozoic record of Great Britain. Palaeogeography, Palaeoclimatology,
13	Palaeoecology 372 , 5–17.
14	LOMBARDO, C. (1999). Sexual dimorphism in a new species of the actinopterygian
15	Peltopleurus from the Triassic of northern Italy. Palaeontology 42, 741–760.
16	LOMBARDO, C. & TINTORI, A. (2005). Feeding specializations in Late Triassic fishes. Annali
17	dell'Università degli Studi di Ferrara. Museologia Scientifica e Naturalistica, Volume
18	<i>Speciale</i> 2005 , 25–32.
19	LOMBARDO, C., SUN, Z. Y., TINTORI, A., JIANG, D. Y. & HAO, W. C. (2011). A new species of
20	Perleidus (Actinopterygii) from the Middle Triassic of Southern China. Bollettino della
21	Società Paleontologica Italiana 50, 75–83.
22	LÓPEZ-ARBARELLO, A. (2004). The record of Mesozoic fishes from Gondwana (excluding
23	India and Madagascar). In Mesozoic fishes 3. Systematics, Paleoenvironments and
24	Biodiversity (eds G. ARRATIA and A. TINTORI), p. 597. Dr. Friedrich Pfeil, München.
25	LÓPEZ-ARBARELLO, A. (2012). Phylogenetic interrelationships of ginglymodian fishes
26	(Actinopterygii: Neopterygii). PLoS ONE 7, e39370.

1	LUCAS, S. G.	(2004). A	global hiatus	in the Middle	Permian tetra	pod fossil record.
-	100000, 20 00	(D			p • • • • • • • • • • • • • • • •

2 *Stratigraphy* **1**, 47–64.

3	LUCAS, S. G. (2009). Timing and magnitude of tetrapod extinctions across the Permo-Triassic
4	boundary. Journal of Asian Earth Sciences 36, 491–502.

- 5 LUCAS, S. G. (2013). No gap in the Middle Permian record of terrestrial vertebrates:
- 6 Comment. *Geology* **41**, e293 (doi: 10.1130/G33734C.1).
- LUCAS, S. G. & HECKERT, A. B. (2001). Olson's gap: A global hiatus in the record of Middle
 Permian tetrapods. *Journal of Vertebrate Paleontology* 21, 75A.
- 9 LUND, R., POPLIN, C. & MCCARTHY, K. (1995). Preliminary analysis of the interrelationships
- 10 of some Paleozoic Actinopterygii. *Geobios* **19**, 215–220.
- 11 MCCUNE, A. R. & SCHAEFFER, B. (1986). Triassic and Jurassic fishes: Patterns of diversity. In

12 The beginning of the age of dinosaurs. Faunal change across the Triassic-Jurassic

13 *boundary* (ed. K. PADIAN), p. 171. Cambridge University Press, London.

14 MENNING, M., ALEKSEEV, A. S., CHUVASHOV, B. I., DAVYDOV, V. I., DEVUYST, F.-X., FORKE,

- 15 H. C., Grunt, T. A.; Hance, L., Heckel, P. H., Izokh, N. G., Jin, Y.-G., Jones, P. J.,
- 16 Kotlyar, G. V., Kozur, H. W., Nemyrovska, T. I., Schneider, J. W.; Wang, X.-D.,
- 17 WEDDIGE, K., WEYER, D. & WORK, D. M. (2006). Global time scale and regional
- 18 stratigraphic reference scales of Central and West Europe, East Europe, Tethys, South
- 19 China, and North America as used in the Devonian Carboniferous Permian
- 20 Correlation Chart 2003 (DCP 2003). Palaeogeography, Palaeoclimatology,
- 21 *Palaeoecology* **240**, 318–372.
- MINIKH, A. V. & MINIKH, M. G. (2009). *Ichthyofauna of the Permian of European Russia [Ikhtiofauna Permi Evropevskov Rossii]*. Nauka Publishing Centre, Saratov.
- 24 MURRAY, A. M. (2000). The Palaeozoic, Mesozoic and early Cenozoic fishes of Africa.
- 25 Fishes and Fisheries 1, 111–145.

1	MUTTER, R. J. & RICHTER, M. (2007). Acanthodian remains from the Middle-Late Permian of
2	Brazil. <i>Geological Journal</i> 42 , 213–224.

3 NEAR, T.J., EYTAN, R. I., DORNBURG, A., KUHN, K. L., MOORE, J

- 4 WAINWRIGHT, P. C., FRIEDMAN, M. & SMITH, W. L. (2012). Resolution of ray-finned
- 5 fish phylogeny and timing of diversification. *Proceedings of the National Academy of*
- 6 Sciences of the United States of America **109**, 13698–13703.
- 7 NELSON, J. S. (2006). *Fishes of the world*. John Wiley, New York.
- 8 NIELSEN, E. (1942). Studies on Triassic Fishes from East Greenland 1. *Glaucolepis* and

9 *Boreosomus. Palaeozoologica Groenlandica* 1, 1–394.

- 10 NIELSEN, E. (1949). Studies on Triassic fishes from East Greenland 2. Australosomus and
- 11 Birgeria. Palaeozoologica Groenlandica **3**, 1–309.
- NIELSEN, E. (1961). On the Eotriassic fish faunas of central east Greenland. In *Geology of the Arctic 1* (ed. G. O. RAASCH), p. 255. University of Toronto Press, Toronto.

14 NÜTZEL, A. & SCHULBERT, C. (2005). Facies of two important Early Triassic gastropod

- 15 lagerstätten: Implications for diversity patterns in the aftermath of the end-Permian mass
- 16 extinction. *Facies* **51**, 495–515.
- 17 ORCHARD, M. J. (2007). Conodont diversity and evolution through the latest Permian and
- 18 Early Triassic upheavals. *Palaeogeography, Palaeoclimatology, Palaeoecology* 252,
- 19 93–117.
- 20 PATTERSON, C. (1994). Bony fishes. Short Courses in Paleontology 7, 57–84.
- PATTERSON, C. & SMITH, A. B. (1987). Is the periodicity of extinctions a taxonomic artefact?
 Nature 330, 248–252.
- 23 PAYNE, J. L. (2005). Evolutionary dynamics of gastropod size across the end-Permian
- extinction and through the Triassic recovery interval. *Paleobiology* **31**, 269–290.

1	PAYNE, J. L., LEHRMANN, D. J., WEI, J., ORCHARD, M. J, SCHRAG, D. P. & KNOLL, A. H.
2	(2004). Large perturbations of the carbon cycle during recovery from the end-Permian
3	extinction. Science 305 , 506–509.
4	PAYNE, J. L., LEHRMANN, D. J., CHRISTENSEN, S., WEI, J. & KNOLL, A. H. (2006).
5	Environmental and biological controls on the initiation ans growth of a Middle Triassic
6	(Anisian) reef complex on the Great Bank of Guizhou, Guizhou Province, China.
7	<i>Palaios</i> 21 , 325–343.
8	PITRAT, C. W. (1973). Vertebrates and the Permo-Triassic extinction. Palaeogeography,
9	Palaeoclimatology, Palaeoecology 14, 249–264.
10	PIVETEAU, J. (1935). Ressemblances des faunes ichthyologiques du Groenland et du Spitzberg
11	avec celle de Madagascar, au Trias inférieur. Compte Rendu Sommaire des Séances de
12	la Société Géologique de France 1935 , 113–114.
13	POPLIN, C. & DUTHEIL, D. B. 2005. Les Aeduellidae (Pisces, Actinopterygii) carbonifères et
14	permiens: systématique et étude phylogénétique préliminaire. Geodiversitas 27, 17-33.
15	QI, W. & STANLEY, G. D. JR. (1989). New Anisian corals from Qingyan, Guiyang, South
16	China. In Lithospheric Geoscience (ed. THE GEOLOGICAL DEPARTMENT OF PEKING
17	UNIVERSITY), p. 11. Peking University Press, Beijing.
18	RAUP, D. M. (1979). Size of the Permo-Triassic bottleneck and its evolutionary implications.
19	Science, New Series 206, 217–218.
20	REGAN, C. T. (1923). The skeleton of Lepidosteus, with remarks on the origin and evolution
21	of the lower neopterygian fishes. Proceedings of the Zoological Society of London 93,
22	445–461.
23	REGUANT, S. (2007). A contribution to knowledge of the development of marine life during
24	the Permian and Triassic through the analysis of life histories of genera. Journal of
25	<i>Iberian Geology</i> 33 , 173–190.

1	RENESTO, S. & STOCKAR, R. (2009). Exceptional preservation of embryos in the
2	actinopterygian Saurichthys from the Middle Triassic of Monte San Giorgio,
3	Switzerland. Swiss Journal of Geoscience 102, 323-330.
4	RIGO, M., PRETO, N., ROGHI, G., TATEO, F., & MIETTO, P. (2007). A rise in the carbonate
5	compensation depth of western Tethys in the Carnian (Late Triassic): Deep-water
6	evidence for the Carnian Pluvial Event. Palaeogeography, Palaeoclimatology,
7	<i>Palaeoecology</i> 246 , 188–205.
8	ROMANO, C. & BRINKMANN, W. (2009). Reappraisal of the lower actinopterygian Birgeria
9	stensioei Aldinger, 1931 (Osteichthyes; Birgeriidae) from the Middle Triassic of Monte
10	San Giorgio (Switzerland) and Besano (Italy). Neues Jahrbuch für Geologie und
11	Paläontologie, Abhandlungen 252, 17–31.
12	ROMANO, C., KOGAN, I., JENKS, J., JERJEN, I. & BRINKMANN, W. (2012). Saurichthys and other
13	fossil fishes from the late Smithian (Early Triassic) of Bear Lake County (Idaho, USA),
14	with a discussion of saurichthyid palaeogeography and evolution. Bulletin of
15	<i>Geosciences</i> 87 , 543–570.
16	Romano, C., Goudemand, N., Vennemann, T. W., Ware, D., Schneebeli-Hermann, E.,
17	HOCHULI, P. A., BRÜHWILER, T., BRINKMANN, W. & BUCHER, H. (2013). Climatic and
18	biotic upheavals following the end-Permian mass extinction. Nature Geoscience 6, 57–
19	60 (doi: 10.1038/ngeo1667).
20	ROMANUK, T. N., HAYWARD, A. & HUTCHINGS, J. A. (2011). Trophic level scales positively
21	with body size in fishes. Global Ecology and Biogeography 20, 231–240.
22	RONIEWICZ, E. & STANLEY, G. D. (1998). Middle Triassic cnidarians from the New Pass
23	Range, Central Nevada. Journal of Paleontology 72, 246–256.
24	ROSCHER, M. & SCHNEIDER, J. W. (2006). Permo-Carboniferous climate: Early Pennsylvanian
25	to Late Permian climate development of central Europe in a regional and global context.
26	In Non-marine Permian Biostratigraphy and Biochronology. Geological Society,

1 London, Special Publications 265 (eds S. G. LUCAS, G. CASSINIS and J. W. SCHNEIDER),

2 95–136.

- SAHNEY, S. & BENTON, M. J. (2007). Recovery from the most profound mass extinction of all
 time. *Proceedings of the Royal Society B* 275, 759–765.
- 5 SALLAN, L. C. (2014). Major issues in the origins of ray-finned fish (Actinopterygii)
- 6 biodiversity. *Biological Reviews* (doi: 10.1111/brv.12086).
- 7 SALLAN, L. C. & COATES, M. I. (2010). End-Devonian extinction and a bottleneck in the early
- 8 evolution of modern jawed vertebrates. *Proceedings of the National Academy of*
- 9 Sciences of the United States of America **107**, 10131–10135.
- 10 SCHAEFFER, B. (1956). Evolution in the subholostean fishes. *Evolution* 10, 201–212.
- 11 SCHAEFFER, B. (1973). Fishes and the Permian-Triassic boundary. In The Permian and
- 12 Triassic systems and their mutual boundary. Canadian Society of Petroleum Geologists,
- 13 *Memoir 2* (eds A. Logan and L. V. Hills), 493–497.
- 14 SCHAEFFER, B. & MANGUS, M. (1976). An Early Triassic fish assemblage from British
- 15 Columbia. *Bulletin of the American Museum of Natural History* **156**, 127–216.
- 16 SCHAUMBERG, G. (1977). Der Richelsdorfer Kupferschiefer und seine Fossilien, III. Die
- 17 tierischen Fossilien des Kupferschiefers. 2. Vertebraten. *Der Aufschluss* 28, 297–352.
- 18 SCHEYER, T. M., ROMANO, C., JENKS, J. & BUCHER, H. (2014). Early Triassic marine biotic
- 19 recovery: The predators' perspective. *PLoS ONE* **9**, e88987.
- 20 SCHNEEBELI-HERMANN, E., KÜRSCHNER, W. M., HOCHULI, P. A., WARE, D., WEISSERT, H.,
- 21 BERNASCONI, S. M., ROOHI, G., UR-REHMAN, K., GOUDEMAND, N. & BUCHER, H. (2013).
- 22 Evidence for atmospheric carbon injection during the end-Permian extinction. *Geology*
- **41**, 579–582.
- 24 SCHNEIDER, J. W. & WERNEBURG, R. (2012). Biostratigraphie des Rotliegend mit Insekten
- 25 und Amphibien. In Stratigraphie von Deutschland X. Rotliegend. Teil I: Innervariscische

1	Becken. Schriftenreihe der Deutschen Gesellschaft für Geowissenschaften 61 (ed.
2	DEUTSCHE STRATIGRAPHISCHE KOMMISSION), 110–142.
3	SCHULTZE, HP. (2004). Mesozoic sarcopterygians. In Mesozoic fishes 3. Systematics,
4	Paleoenvironments and Biodiversity (eds G. ARRATIA and A. TINTORI), p. 463. Dr.
5	Friedrich Pfeil, München.
6	SCHULTZE, HP., HEIDTKE, U. H. J. (1986). Rhizodopside Rhipidistia (Pisces) aus dem Perm
7	der Pfalz (W-Deutschland). Neues Jahrbuch für Geologie und Paläontologie, Monatshefte
8	1986 , 165–170.
9	SEILACHER, A., REIF, WE., WESTPHAL, F., RIDING, R., CLARKSON, E. N. K. & WHITTINGTON,
10	H. B. (1985). Sedimentological, ecological and temporal patterns of fossil Lagerstätten.
11	Philosophical Transactions of the Royal Society of London, Series B, BiologicalSciences
12	311 , 5–24.
13	SEPKOSKI, J. J. JR. (1984). A kinetic-model of Phanerozoic taxonomic diversity, 3. Post-
14	Paleozoic families and mass extinctions. <i>Paleobiology</i> 10 , 246–267.
15	SEPKOSKI, J. J. JR. (2002). A compendium of fossil marine animal genera. Bulletins of
16	American Paleontology 363 , 1–560.
17	SHEN, SZ., CROWLEY, J. L., WANG, Y., BOWRING, S. A., ERWIN, D. H., SADLER, P. M., CAO,
18	CQ., Rothman, D. H., Henderson, C. M., Ramezani, J., Zhang, H., Shen, Y., Wang,
19	XD., Wang, W., Mu, L., Li, WZ., Tang, Y. G., Liu, XL., Liu, LJ., Zeng, Y., Jiang,
20	Y. F. & JIN, Y. G. (2011). Calibrating the end-Permian mass extinction. Science 334,
21	1367–1372.
22	SHEN, SZ. & SHI, G. R. (2002). Paleobiogeographical extinction patterns of Permian
23	brachiopods in the Asian-western Pacific region. Paleobiology 28, 449-463.
24	SIGNOR, P. W. III & LIPPS, J. H. (1982). Sampling bias, gradual extinction patterns and
25	catastrophes in the fossil record. Geological Society of America Special Paper 190, 291-
26	296.

1	SIMMS, M. J. & RUFFELL, A. H. (1989). Synchroneity of climatic change and extinctions in the
2	Late Triassic. Geology 17, 265–268.
3	SMITH, J. L. B. (1939). A living fish of Mesozoic type. Nature 143, 455–456.

- 4 SONG, H., WIGNALL, P. B., CHEN, Z.-Q., TONG, J., BOND, D. P. G., LAI, X., ZHAO, X., JIANG,
- 5 H., YAN, C., NIU, Z, CHEN, J., YANG, H., & WANG, Y. (2011). Recovery tempo and pattern
- 6 of marine ecosystems after the end-Permian mass extinction. *Geology* **39**, 739–742.
- 7 ŠTAMBERG, S. (1994). Comparison of actinopterygian fishes from the Autun Basin (France)
- 8 and the Krkonoše Piedmont Basin (Eastern Bohemia). Bulletin of the Czech Geological
- 9 Survey [Věstník Českého Geologického Ústavu] 69, 19–24.
- 10 ŠTAMBERG, S. (2007). Permo-Carboniferous Actinopterygians of the Boskovice Graben. Part
- 11 *1. Neslovicella, Bourbonnella, Letovichthys, Elonichthys.* Museum of Eastern Bohemia,
- 12 Hradec Králové.
- STANLEY, S. M. & YANG, X. (1994). A double mass extinction at the end of the Paleozoic era.
 Science 266, 1340–1344.
- 15 STEMMERIK, L., BENDIX-ALMGREEN, S. E. & PIASECKI, S. (2001). The Permian-Triassic
- 16 boundary in central East Greenland: Past and present views. *Bulletin of the Geological*
- 17 Society of Denmark **48**, 159–167.
- 18 STENSIÖ, E. (1921). Triassic fishes from Spitzbergen 1. Adolf Holzhausen, Wien.
- 19 STENSIÖ, E. (1925). Triassic fishes from Spitzbergen 2. Kungliga Svenska
- 20 *Vetenskapsakademiens Handlingar* **3**, 1–261.
- STENSIÖ, E. (1932). Triassic fishes from East Greenland 1–2. *Meddelelser om Grønland* 83,
 1–298.
- 23 SUN, Y. JOACHIMSKI, M. M., WIGNALL, P. B., YAN, C., CHEN, Y., JIANG, H., WANG, L. & LAI.,
- 24 X. (2012). Lethally hot temperatures during the Early Triassic greenhouse. *Science* **338**,
- 25 366–370.

THOMSON, K. S. (1976). Explanation of large scale extinctions of lower vertebrates. *Nature* 261, 578–580.

3	THOMSON, K. S. (1977). The pattern of diversification among fishes. In Patterns of evolution
4	as illustrated by the fossil record (ed. A. HALLAM), p. 377. Elsevier, Amsterdam.
5	TINTORI, A. (1992). Fish taphonomy and Triassic anoxic basins from the Alps: A case history.
6	Rivista Italiana di Paleontologia e Stratigrafia 97, 393–408.
7	TINTORI, A. (1998). Fish biodiversity in the marine Norian (Late Triassic) of northern Italy:
8	The first neopterygian radiation. Italian Journal of Zoology 65, 193–198.
9	TINTORI, A., HITIJ, T., JIANG, DY., LOMBARDO, C. & SUN, ZY. (2013). Triassic
10	actinopterygian fishes: the recovery after the end-Permian crisis. Integrative Zoology
11	(doi: 10.1111/1749-4877.12077).
12	TINTORI, A., SUN, ZY., LOMBARDO, C., JIANG, DY., JI, C. & MOTANI, R. (2012). A new
13	"flying" fish from the Late Ladinian (Middle Triassic) of Wusha (Guizhou Province,
14	southern China). Gortania 33, 39–50.
15	TONG, J., ZHOU, X., ERWIN, D. H., ZUO, J. & ZHAO, L. (2006). Fossil fishes from the Lower
16	Triassic of Majiashan, Chaohu, Anhui Province, China. Journal of Paleontology 80,
17	146–161.
18	TOZER, E. T. (1965). Lower Triassic stages and ammonoid zones of Arctic Canada. Paper of
19	the Geological Survey of Canada 65, 1–14.
20	TOZER, E. T. (1982). Marine Triassic faunas of North America: Their significance for
21	assessing plate and terrane movements. Geologische Rundschau 71, 1077–1104.
22	TREBILCO, R., BAUM, J. K., SALOMON, A. K. & DULVY, N. K. (2013). Ecosystem ecology:
23	Size-based constraints on the pyramids of life. Trends in Ecology & Evolution 28, 423-
24	431.

1	TURNER, S., BURROW, C. J., SCHULTZE, HP., BLIECK, A., REIF, WE., REXROAD, C. B.,
2	BULTYNCK, P. & NOWLAN, G. S. (2010). False teeth: Conodont-vertebrate phylogenetic
3	relationships revisited. Geodiversitas 32, 545–594.
4	ТWITCHETT, R. J. & ОЛ, Т. (2005). Early Triassic recovery of echinoderms. Comptes Rendus
5	<i>Palevol</i> 4 , 531–542.
6	VISSCHER, H., VAN HOUTE, M., BRUGMAN, W. A. & POORT, R. J. (1994). Rejection of a
7	Carnian (Late Triassic) "pluvial event" in Europe. Review of Palaeobotany and
8	Palynology 83 , 217–226.
9	WANG, XD. & SUGIYAMA, T. (2000). Diversity and extinction patterns of Permian coral
10	faunas of China. <i>Lethaia</i> 33 , 285–294.
11	WARE, D., JENKS, J. F., HAUTMANN, M. & BUCHER, H. (2011). Dienerian (Early Triassic)
12	ammonoids from the Candelaria Hills (Nevada, USA) and their significance for
13	palaeobiogeography and palaeoceanography. Swiss Journal of Geosciences 104, 161-
14	181.
15	WEN, W., ZHANG, QY., HU, SX., BENTON, M. J., ZHOU, CY., TAO, X., HUANG, JY., &
16	CHEN, ZQ. (2013). Coelacanths from the Middle Triassic Luoping Biota, Yunnan,
17	South China, with the earliest evidence of ovoviviparity. Acta Palaeontologica Polonica
18	58 , 175–193.
19	WENDRUFF, A. J. & WILSON, M. V. H. (2012). A fork-tailed coelacanth, Rebellatrix
20	divaricerca, gen. et sp. nov. (Actinistia, Rebellatricidae, fam. nov.), from the Lower
21	Triassic of Western Canada. Journal of Vertebrate Paleontology 32, 499-511.
22	WHITE, E. (1953). The coelacanth fishes. <i>Discovery</i> 14, 113–117.
23	WIGNALL, P. B., BOND, D. P. G., HAAS, J., WANG, W., JIANG, H., LAI, X., ALTINER, D.,
24	VÉDRINE, S., HIPS, K., ZAJZON, N., SUN, Y. & NEWTON, R. J. (2012). Capitanian (middle
25	Permian) mass extinction and recovery in western Tethys: A fossil, facies, and $\delta^{13}C$
26	study from Hungary and Hydra Island (Greece). Palaios 27, 78–89.

1	WITZMANN, F. & SCHOCH, R. R. (2012). A megalichthyid sarcopterygian fish from the Lower
2	Permian (Autunian) of the Saar-Nahe Basin, Germany. Geobios 45, 241–248.
3	XU, GH., ZHAO, LJ., GAO, KQ. & WU, FX. (2013). A new stem-neopterygian fish from
4	the Middle Triassic of China shows the earliest over-water gliding strategy of the
5	vertebrates. Proceedings of the Royal Society B 280, 20122261
6	(http://dx.doi.org/10.1098/rspb.2012.2261).
7	ZIEGLER, A. M., ESHEL, G., MCALLISTER REES, P., ROTHFUS, T. A., ROWLEY, D. B. &
8	SUNDERLIN, D. (2003). Tracing the tropics across land and sea: Permian to present.
9	<i>Lethaia</i> 36 , 227–254.
10	
11	VIII. SUPPORTING INFORMATION
12	
13	Supplementary Tables A1-A3. Excel spreadsheets listing occurrences and body size data of
14	Permian and Triassic Osteichthyes: Dipnoi (A1), Actinistia (A2), and Actinopterygii (A3).
15	For references cited in tables A1–A3 see Supplementary References.
16	
17	Supplementary Tables B1-B6. Excel spreadsheets listing diversity data and results of
18	statistical tests.
19	
20	Supplementary References. References for Supplementary Tables A1-A3.
21	
22	
23	Figures captions:
24	
25	Figure 1. Palaeogeography of Permian and Triassic Osteichthyes. A. – Palaeogeographic
26	map of localities (partially summarized, cf. Supplementary Tables A1-A3) yielding bony fish

1	fossils of Permian (above) and Triassic (below) age and their allocation to palaeogeographic
2	provinces (1, Boreal sea; 2, east-equatorial Panthalassa; 3, south-western Pangaea and south-
3	eastern Panthalassa; 4, Palaeotethys; 5, Neotethys; 6, western and south-western Panthalassa).
4	Localities: ACU – Arctic USA (AK), AN – Angola; ANT – Antarctica (Transantarctic
5	Mountains); ARG – Argentina; ASA – Australia, South Australia, AU – Austria, BV –
6	Bolivia; CAA – Canadian Arctic Archipelago, CAN – China, Anhui, CG – China, Gansu,
7	CGX – China, Guangxi, CHB – China, Hubei, CHL – Chile; CHN – China, Hunan, CJI –
8	China, Jiangsu, CR – Czech Republic, CSC – China, Sichuan, CX – China, Xinjiang, CYG –
9	China, Yunnan and Guizhou, CZJ – China, Zhejiang; EE – Eastern Europe, EG – East
10	Greenland, EN – England/Ireland; EUS – eastern USA (MA, CT, NJ, PA, VA); FR – France;
11	GR – Germany; IT – Italy; IS – Israel; KG – Kyrgyzstan; KY – Kenya; KZ – Kazakhstan
12	(eastern, western, Mangyshlak peninsula); LS – Lesotho; LY – Libya; MOR – Morocco; MG
13	– Madagascar; NB – Namibia; NBZ – Northern Brazil (Maranhão, Paraná); NE – The
14	Netherlands; NER – Northern European Russia (Jaroslawl, Kostroma, Moscow Syncline,
15	Vologda, Vyatka-Kama depression, Wladimir); NP – Nepal; NSW – Australia, New South
16	Wales; PL – Poland; QL – Australia, Queensland; RFE – Russia Far East (Magadan,
17	Tunguska); RKJ – Russia, Krasnojarsk; RKP – Russia, Komi/Pechora; RNS – Russia, North
18	Siberia (Lena River Delta, Verkhojansk); SA – Saudi Arabia; SAF – South Africa; SB –
19	Svalbard (Spitsbergen and Bjørnøya); SBZ – Southern Brazil (Rio Grande do Sul, Santa
20	Catarina, São Paulo); SCU – south-central USA (OK, TX); SER – Southern European Russia
21	(Astrachan, Cis-Urals, Kargala, Kazan, Kuznetsk, Orenburg, Perm, Samara, Saratov); SL –
22	Slovenia; SP – Spain; SW – Sweden; SZ – Switzerland; TAS – Australia, Tasmania; TH –
23	Thailand; TK – Turkey; TZ – Tanzania; UR – Uruguay; WA – Western Australia; WC –
24	western Canada (Alberta and British Columbia); WUS – western USA (AZ, CA, CO, ID,
25	NM, NV, UT); ZA – Zambia; ZB – Zimbabwe. B. – Relative contribution of the different

palaeogeographic provinces indicated in A to the total diversity of Permian and Triassic
 Osteichthyes (only known occurrences).

3

Figure 2. Rarefaction and extrapolation curves at species and genus level. A. – Permian
dataset. B. – Triassic dataset. Solid lines: interpolation curves; dashed lines: extrapolation
curves; white circles: sample coverages (see text for details).

7

Figure 3. Tentative evaluation of the Lagerstätten effect in the fossil record of Permian
and Triassic Osteichthyes. A. – Marine and brackisch realm. B. – Freshwater environment.
Upper graphs in A and B show a comparison of the diversity trends derived from the whole
dataset (black line) and with omission of occurrences at Lagerstätten (grey line, see text for
details). Lower graphs in A and B show genus richness at Lagerstätten (black bars).
Percentage values indicate proportions of endemic genera (partially summarized). Also see
Supplementary Table B1.

15

16 Figure 4. Marine (and brackish) diversity dynamics of Permian and Triassic

Osteichthyes (including euryhaline taxa). A. – Diversity (genus richness, mean standing diversity, boundary crossers). B. – Originations/extinctions (bars) and per taxon origination and extinction rates (shades), including singletons. C. – Originations/extinctions (bars) and per taxon origination and extinction rates (shades) without singletons. First three unlabelled intervals of the Early Triassic (in stratigraphic order): Griesbachian, Dienerian, Smithian.

23 Figure 5. Freshwater diversity dynamics of Permian and Triassic Osteichthyes

24 (including euryhaline taxa). A. – Diversity (genus richness, mean standing diversity,

- 25 boundary crossers). **B.** Originations/extinctions (bars) and per taxon origination and
- 26 extinction rates (shades), including singletons. C. Originations/extinctions (bars) and per

taxon origination and extinction rates (shades) without singletons. First three unlabelled
 intervals of the Early Triassic (in stratigraphic order): Griesbachian, Dienerian, Smithian.
 3

4	Figure 6. Global diversity dynamics of Permian and Triassic Osteichthyes (euryhaline
5	taxa only counted once per interval). A. – Diversity (genus richness, mean standing
6	diversity, boundary crossers). B Originations/extinctions (bars) and per taxon origination
7	and extinction rates (shades), including singletons. C Originations/extinctions (bars) and
8	per taxon origination and extinction rates (shades) without singletons. First three unlabelled
9	intervals of the Early Triassic (in stratigraphic order): Griesbachian, Dienerian, Smithian.
10	
11	Figure 7. Diversity dynamics of different groups of Osteichthyes during the Permian
12	and the Triassic. A. – Marine and brackish environment, B. – Freshwater realm, C. – Global.
13	First three unlabelled intervals of the Early Triassic (in stratigraphic order): Griesbachian,
14	Dienerian, Smithian.
15	
16	Figure 8. Relative diversity of different groups of Osteichthyes during the Permian and
17	the Triassic. A. – Marine and brackish realm, B. – Freshwater environment, C. – Global.
18	First three unlabelled intervals of the Early Triassic (in stratigraphic order): Griesbachian,
19	Dienerian, Smithian.
20	
21	Figure 9. Palaeolatitudinal diversity dynamics of Osteichthyes during the Permian and
22	the Triassic. A. – Marine and brackish environment, B. – Freshwater realm, C. – Global. Left
23	column: Palaeolatitudinal gradient of epoch-level pooled genus richness (values of same
24	palaeolatitudinal belts of the northern and southern hemisphere combined). Right column:
25	Palaeolatitudinal diversity through time. Genera occurring within more than one
26	palaeolatitudinal belt are counted once per belt.

2	Figure 10. Body size evolution (Log ₂ of species' maximum standard lengths) of
3	Osteichthyes during the Permian and the Triassic. A. – Marine and brackish realm, B. –
4	Freshwater environment, C. – Global. Left column: Box plots for all Osteichthyes. Solid lines
5	mark the medians, boxes the 25–75% quartiles, whiskers the whole range of data. Right
6	column: Distribution of body sizes of species belonging to different groups of bony fishes.
7	Trend lines connect medians. SL = Standard length. Time intervals: Anis. = Anisian, Carn. =
8	Carnian, EP = Early Permian, ET = Early Triassic, Lad. = Ladinian, LP = Late Permian, LT =
9	Late Triassic, MP = Late Permian, MT = Middle Triassic, NR. = Norian-Rhaetian interval.
10	
11	Figure 11. Body size evolution (Log ₂ of species' maximum standard lengths) of Permian
12	and Triassic Osteichthyes across palaeolatitudinal zones. A. – Marine and brackish realm,
13	B. – Freshwater environment, C. – Global. Solid lines mark the medians, boxes the 25–75%
14	quartiles, whiskers the whole range of data. Time intervals: EP = Early Permian, ET = Early
15	Triassic, LP = Late Permian, LT = Late Triassic, MP = Late Permian, MT = Middle Triassic.
16	
17	Figure 12. Diversity dynamics of Permian and Triassic fishes (Chondrichthyes,
18	Osteichthyes). A. – Genus richness. B. – Relative diversity. Data for Chondrichthyes after
19	Koot (2013), data for Osteichthyes from this study (Supplementary Tables A1-A3). First
20	three unlabelled intervals of the Early Triassic (in stratigraphic order): Griesbachian,
21	Dienerian, Smithian.






















