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1 **Title**

2 Genetic determinism to primary early-onset osteoarthritis

3

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19

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21 Early-onset osteoarthritis, genetics, mutation

22

23 **Abstract**

24 Osteoarthritis (OA) is the most common joint disease observed worldwide. A minority of cases
25 correspond to familial presentation characterized by early-onset forms and are genetically
26 heterogeneous.

27 This review brings a new point of view of the molecular basis of OA by focusing on gene
28 mutations responsible for early-onset OA determinism. Recently, thanks to whole-exome
29 sequencing, a gain of function mutation in *TNFRSF11B* gene was identified in two distant
30 family members with EO-OA, opening new therapeutic perspectives for OA. Indeed,
31 unravelling the molecular basis of rare Mendelian OA forms will help improve the
32 understanding of molecular processes involved in OA pathogenesis and to ameliorate patient
33 diagnosis, management and therapy.

34

35 **Primary osteoarthritis, a genetic disease?**

36 **Osteoarthritis** (OA, see glossary) is the most common joint disease observed worldwide and
37 affects nearly 27 million US adults [1]. It is the main source of morbidity in developed
38 countries. Symptoms, when present, include pain and reduced mobility causing partial or
39 complete inability to work, difficulties with activities of daily living and impaired quality of
40 life, leading to a very important social-economic cost. OA is mainly characterized by the
41 gradual loss of articular cartilage, particularly affecting the knee, hip, spine, hand and foot
42 (Figure 1). However, OA is not only a disease of cartilage. It affects all the tissues of the joint,
43 including synovium, subchondral bone, capsule, ligaments, periarticular muscles and the
44 sensory nerves whose termini lie within these tissues [2]. Currently, the majority of OA cases
45 are considered as primary, defined as OA lacking underlying factors and structural
46 abnormalities. By contrast, secondary OA results from various factors such as traumatic injury,
47 specific anatomical deformities in the joint or specific abnormalities of the cartilage
48 extracellular matrix (ECM) [3]. Despite the high prevalence of OA and its substantial public
49 health impact, genetic basis of primary OA pathogenesis are not fully elucidated. No efficient
50 treatments are currently available, since they only slow the progression of the disease and
51 reduce pain. A better understanding of the molecular mechanisms and pathways involved in
52 primary OA genesis will help provide improved patient diagnosis, early management, and the
53 development of innovative therapies.

54

55 Premature OA is rarely present without other clinical symptoms and is mainly observed as a
56 part of **syndromes**. In these cases, **early-onset OA** (EO-OA) mostly develops secondary to
57 inflammatory disease or to biomechanical defects, such as **osteochondrodysplasia**. Non-
58 syndromic EO-OA families without dysplasia or other causative pathology are extremely rare.
59 The phenotype of these rare OA families resembles common OA at later ages in the population

60 except for the early age of onset (20–50 years) [4]. However, identification and screening of
61 such families is highly important and should rely on strict inclusion criteria (see Box 2 for
62 criteria that we propose for future genetic studies). In fact, they may reveal causal genes
63 responsible for the genetic determinism to EO-OA and lead to a better understanding of the
64 physiopathological mechanisms involved in OA development.

65

66 First, this review describes the evidence for a genetic component in OA. Second, it focuses only
67 on causal genetic alterations for the determinism of EO-OA, and aims to clearly distinguish
68 causal mutations for EO-OA and those responsible for multiple phenotypes. In fact, most of the
69 pathogenic mutations detected in EO-OA have been identified in patients harboring a
70 syndromic presentation, including EO-OA, but also presenting other prominent secondary
71 features such as ocular, auditory or aortic abnormalities. The presence of multiple clinical
72 features simultaneously explains the difficulty to clearly conclude on the causal role of
73 mutations for the genetic determinism of EO-OA. However, thanks to the use of strict inclusion
74 criteria for cases and controls, and to whole-exome sequencing strategy, a gain of function
75 mutation in *TNFRSF11B* gene was recently identified, leading to innovative therapy
76 perspectives for OA [5].

77

78 **Genetic basis for primary osteoarthritis**

79 Primary OA is a complex disease, caused by a combination of three main risk factors: genetics,
80 environmental factors and age [3]. Evidence for a genetic component in OA comes from family
81 studies, twin studies and animal models. The first evidence for a strong genetic contribution to
82 OA was first reported in 1941 by Stecher *et al.* who demonstrated that **Heberden's nodes** were
83 three times more common in sisters of OA patients than in the general population [6].
84 Subsequently this was confirmed in families with generalized OA with the observation that first

85 degree relatives were twice as likely to present radiographically generalized OA as well [7]. In
86 fact, susceptibility to common OA, which occurs at **late-onset**, is modulated by genetic risk
87 factors, with single nucleotide polymorphisms more frequently observed in patients than in the
88 general population. In 1996, Spector *et al.* estimated the influence of genetic factors between
89 39 and 65% in twins with hand and knee OA [8]. Genetic risk factors associated with primary
90 OA susceptibility (common OA) have been extensively reviewed [9–14]. To date, 21
91 independent OA susceptibility loci were established based on genome-wide significance in
92 GWAS ($p \leq 5 \times 10^{-8}$) and/or proven functional involvement in OA by follow-up studies [15]. By
93 contrast, rare EO-OA forms represent Mendelian diseases with an autosomal dominant
94 inheritance pattern, as reported in families with EO-OA [16–19]. Genetic determinism to EO-
95 OA forms are caused by single gene mutations, which are only carried by patients. In 1993,
96 Nakata *et al.* reported that transgenic mice with a heterozygous central deletion of *COL9A1*,
97 developed OA without **chondrodysplasia** [20]. Since then, several **animal models** have been
98 developed to explore and confirm the causal role of mutations identified in patients, at the
99 genetic level [21]. Indeed, the role of genetics in idiopathic EO-OA pathogenesis has now been
100 clearly established. By contrast, findings from mouse models give new insight into OA
101 pathogenesis and could be further explored in human. As example, a deficiency of mitogen-
102 inducible gene-6 (Mig-6) in mice leads to the development of an EO-OA-like disorder in
103 multiple synovial joints. Recently, this model provides insight into the critical role of
104 chondrocytes in developing the OA-like disorder in the knees, and into the requirement of other
105 cell types for full development of the Mig-6-deficient joint phenotype [22].

106

107 Several approaches have been used to unravel the molecular basis of EO-OA. Thanks to
108 technological improvements, the scale and resolution of screened genomic regions have been
109 enhanced. Target studies of a single candidate genes (e.g. using CSGE or *conformation sensitive*

110 *gel electrophoresis*, RFLP or *restriction fragment length polymorphisms* or Sanger sequencing)
111 have been replaced by high-throughput screening (e.g. genome-wide linkage analysis or whole
112 exome sequencing). Mutations are generally identified in families harboring a severe phenotype
113 and subsequently validated in replication cohorts of cases with the same phenotype. In patients
114 with EO-OA, these strategies have led to the identification of mutations in genes encoding
115 components of the ECM, the TGF- β pathway, vesicular transport or bone remodeling (Figure
116 2, Key Figure). Distinction between mutated genes identified in idiopathic EO-OA patients
117 (Table 2) and those identified in patients having syndromic forms of EO-OA (Table 3) is clearly
118 established.

119

120 **Mutations in genes encoding collagen proteins**

121 The collagen type II, alpha 1 gene (*COL2A1*), encoding one of the three chains of type II
122 collagen (Box 1), has been targeted for extensive analysis in degenerative diseases of the joint
123 due to the abundance of type II collagen in **epiphyseal** and articular cartilage tissue. Mutations
124 in *COL2A1* gene have been identified in patients with various clinical features and are known
125 as type II collagenopathies [23] (Table 1), for a phenotypic spectrum description see [24]. All
126 of these syndromes are characterized by premature OA. As non-syndromic EO-OA (sporadic
127 or familial) are infrequent, *COL2A1* mutations have been rarely screened or identified in these
128 patients. In 1989, a genetic linkage between *COL2A1* gene and familial EO-OA was described
129 in two unrelated Finnish families presenting autosomal dominant inheritance of OA with a
130 mean age of symptoms of 38 years [16]. Patients had radiographically verified polyarticular
131 OA, without evidence of chondrodysplasia. Furthermore, **spondyloepiphyseal dysplasia**,
132 **Stickler syndrome** (STL), or any other congenital cartilage abnormalities were excluded. This
133 linkage was also confirmed in an unrelated family presenting OA with mild chondrodysplasia
134 [25]. Despite exhibiting chondrodysplasia of the metatarsal head and spine, osteoarthritic

135 changes developed progressively in many joints without any epiphyseal deformities or other
136 evidence of chondrodysplasia. In addition, common causes of secondary OA were excluded.
137 This suggested that generalized OA was the main trait in this family and that OA was probably
138 non-syndromic. A missense mutation (c.2155C>T, p.Arg719Cys) was subsequently found in
139 this family, cosegregating with phenotype and absent in 57 unrelated individuals [26]. In 1995,
140 *COL2A1* mutations were detected in one of 45 patients with familial EO-OA, and in more than
141 20% of 36 patients with moderate to severe chondrodysplasias [27]. Therefore, *COL2A1*
142 mutations are not exclusive to non-syndromic EO-OA. However, several patients with *COL2A1*
143 mutations, primarily manifested as generalized OA in late childhood or early adulthood have
144 been reported [19,28], confirming that *COL2A1* mutations are responsible for EO-OA (for
145 review of *COL2A1* mutations -- as a predominant feature -- in OA patients, see [23]).

146 The majority of mutations found in *COL2A1* are missense mutations where glycine is usually
147 replaced by a bulkier amino acid. Among the non-glycine missense mutations, the arginine-to-
148 cysteine substitutions predominate. The α -chain of type II collagen consists of repeat tripeptide
149 sequences, with glycine strictly in place every third amino acid. These tripeptides are essential
150 for collagen triple helix assembly. Thus, mutations resulting in amino acid changes can impair
151 protein stability and thus, helical structure and proper type II collagen function [29]. However,
152 no clear genotype-to-phenotype profiles have been established in type II collagenopathies
153 [24,30].

154 Experimental mouse models do favor a role of *COL2A1* alterations in OA pathogenesis.
155 *Dell*(+/-) mice harboring a small deletion in *Col2a1* have been found to develop early OA-like
156 lesions in their knee joints [31]. Heterozygous *Sedc*/+ mouse, mutant for *Col2a1*, appears
157 phenotypically normal, yet develop premature OA [32]. Homozygous (*sedc/sedc*) mouse
158 expresses dwarfism in addition to the OA-like effect on articular cartilage [33]. Based on
159 immunohistochemistry and electron microscopy analyses, the authors conclude that collagen in

160 the mutant's articular cartilage (both heterozygote and homozygote) fails to provide the normal
161 meshwork required for matrix integrity and overall cartilage stability.

162

163 Mutations in minor collagen types have also been identified in patients presenting EO-OA. The
164 three alpha chains of type IX collagen are encoded by the collagen type IX, alpha 1, 2 and 3
165 genes (*COL9A1*, *COL9A2* and *COL9A3*), respectively. Type IX collagen mutations are
166 responsible for **multiple epiphyseal dysplasia** (MED) [34–36]. Further support for the possible
167 role of type IX collagen in OA has been obtained from animal studies. Mutant mice for *Col9a1*
168 develop a severe degenerative joint disease resembling human OA [20,37]. To date, only one
169 study has screened mutations in the three type IX collagen genes within 72 Finnish non-
170 syndromic EO-OA patients. One unique exonic and five unique intronic variations were found
171 in *COL9A1*, *COL9A2* and *COL9A3*, but none of them altered known splicing consensus
172 sequences or resulted in amino acid substitutions [19]. Recently, a novel missense *COL9A3*
173 mutation were identified in 2 Korean related patients, the proband presented with MED and his
174 father with EO-OA [38]. This supports the hypothesis that *COL9A3* mutations could also be
175 causal for EO-OA.

176

177 The type XI collagen fibrils are encoded by the collagen type XI, alpha 1 and 2 genes (*COL11A1*
178 and *COL11A2*), respectively. *COL11A1* has been implicated in OA development in one mouse
179 model. Heterozygosity for a loss-of-function mutation in *Col11a1* resulted in OA development
180 in the knee and temporomandibular joints of *cho/+* mice [39]. This suggests that *COL11A1*
181 mutations in humans could be causal for OA. Nevertheless, no causal *COL11A1* mutations have
182 been identified to date (one Dutch family, 72 Finnish probands and one US family with primary
183 EO-OA) [17,19]. This may be due to a limited number of OA cases studied.

184 Presently, only one study has identified mutations in the *COL11A2* gene in non-syndromic EO-
185 OA [19]. All other studies have been conducted in patients presenting syndromic OA,
186 **otospondylomegaepiphyseal dysplasia** (OSMED) and other overlapping phenotypes such as
187 **cleft palate**, **Robin sequence**, and **non-ophthalmic STL**. In the former study, *COL11A2*
188 mutations have been identified in two families presenting EO-OA. Despite great efforts to
189 define strict selection criteria for probands and to establish OA diagnosis on radiological data,
190 related patients have unclear clinical definition. In fact, the first mutation is a single nucleotide
191 deletion affecting a consensus splice site and leading to an in-frame deletion of 36 amino-acids
192 of the exon 43 (c.3151-2delA, p.Gly1051_Lys1086del) This splice variant was identified in a
193 small US family with EO-OA, segregating with the OA phenotype in the two affected cases.
194 One of them has experienced knee pain since age 17 after participating in athletic activities,
195 suggesting that OA development is secondary to excessive biomechanical constraints. This
196 splice mutation leads to exon 43 skipping and thus to an in-frame deletion of 36 amino-acids.
197 The authors suggest that this deletion leads to only partially folded molecules of type XI
198 collagen. The second mutation is a missense variant (c.1615C>T, p.Arg539Trp) identified in a
199 Finnish proband with primary bilateral hip EO-OA. This mutation segregates with the OA
200 phenotype in the four EO-OA cases of the family. One member presenting late-onset OA does
201 not share the mutation, suggesting that the mutation is specific to EO-OA, and responsible for
202 the early-onset OA in these patients. However, one mutation carrier has a loose cartilage
203 fragment caused by **osteochondritis dissecans** (OD) (which could explain secondary OA
204 development), and one other mutation carrier is asymptomatic at 45 years of age. This missense
205 mutation leads to the substitution of an arginine to a tryptophan in the major triple-helix and
206 has been predicted to be probably damaging by **Polyphen *in silico* analysis**. The authors
207 assume that this tryptophan substitution (the most hydrophobic amino acid), has an effect on
208 the collagen triple-helix conformation or affects the interactions with other matrix molecules.

209

210 **Mutations in genes encoding other components of the cartilage extracellular matrix**

211 Proteoglycans represent another major protein class in the ECM. The aggrecan gene (*ACAN*)
212 encodes aggrecan, the most abundant proteoglycan in cartilage. Its main function is to resist
213 compression, protecting bones and joints. Heterozygous mutations in *ACAN* have been
214 identified in two families affected by **spondyloepiphyseal dysplasia type Kimberley** (SEDK)
215 [40] and OD [41] respectively. These two mutations affect the C-type lectin G3 domain of
216 aggrecan, which mediates interactions with other proteins in the ECM [42]. However, a
217 homozygous missense mutation of *ACAN* has been identified in a family presenting a recessive
218 form of spondyloepimetaphyseal dysplasia without manifestation of OA phenotype [43]. This
219 mutation affects also the C-type lectin G3 domain of aggrecan. Thus, disruption of this domain
220 does not inevitably lead to an EO-OA phenotype. Yet, a 7-bp deletion in the *ACAN* gene has
221 been found to cause certain forms of chondrodysplasia in *cmd* mouse [44]. So, mutations in
222 *ACAN* seems not to be directly causal for primary EO-OA but rather to skeletal disorders
223 leading to secondary EO-OA.

224

225 Genes that encode other non-collagenous proteins of the ECM have also been screened for
226 mutations in EO-OA patients. The cartilage oligomeric matrix protein gene (*COMP*) encodes a
227 pentameric ECM glycoprotein that catalyzes collagen assembly and promotes the formation of
228 well-defined fibrils [45]. Thus, it may play a role in the structural integrity of cartilage. *COMP*
229 gene mutations were initially identified in familial and sporadic **pseudoachondroplasia**
230 (PSACH) cases [46,47] and in sporadic MED cases [47] and were subsequently validated by
231 numerous studies [48–52]. In vivo, the tetracycline-inducible D469del-*COMP* mouse model
232 most closely mimics the PSACH pathology [53]. Genotype-to-phenotype correlations of
233 *COMP* gene mutations in these chondrodysplasias were studied for 300 *COMP* mutations [54].

234 In 2011, a missense *COMP* mutation (c.2152C>T, p.Arg718Trp) was identified in a large
235 extended six-generation Taiwanese kindred with familial EO-OA, negative for *COL2A1*
236 mutations [55]. Twenty-five relatives had radiologically diagnoses OA with a mean age onset
237 of 30 years, and most of the secondary causes of OA were ruled out by physical and laboratory
238 findings. On 26 mutation carriers, 21 had EO-OA and five were asymptomatic with a mean age
239 of 18 years, possibly explaining the absence of OA. Four cases with clinical and radiological
240 EO-OA did not carry the mutation, suggesting a multifactorial inheritance of OA. The mutation
241 has been previously described in MED patients [49,56,57]. Thus, the authors conclude that the
242 mutation causes a mild form MED mimicking the EO-OA phenotype.

243

244 Matrilin-3 gene (*MATN3*) mutations have been identified in patients with MED [58–60]. All
245 are missense mutations and mainly affect conserved residues within the beta-sheet of the single
246 A-domain of matrilin-3. These mutations cause protein misfolding, and preventing its secretion
247 from the rough endoplasmic reticulum, both *in vitro* and *in vivo* [61–63]. In mammalian cells
248 overexpressing *MATN3* mutant (CHO-B2 cell line or primary bovine articular chondrocytes),
249 mutant protein were retained and accumulated within the cells, in particular within the rough
250 endoplasmic reticulum due to unfolded conformation [61,62]. These observations were also
251 found in a murine model of epiphyseal dysplasia generated by knocking-in a *Matn3* mutation.
252 The phenotype of mice homozygous for the mutation is consistent to human multiple epiphyseal
253 dysplasia. In contrast, a missense variation affecting the epidermal growth factor–like domain
254 of matrilin-3 cosegregated with hand OA in several families [64]. This variation does not appear
255 to interfere with secretion but rather with extracellular assembly of matrix structures [65]. The
256 variation frequency is slightly more than 2% in patients with hand OA in the Icelandic
257 population [64]. It appears to contribute to the susceptibility to adult-onset OA affecting hand
258 joints preferentially, and is not associated with an early-onset or unusual form of OA [64]. The

259 phenotype of transgenic mouse models are in contradiction. For instance, in one study, *Matn3*
260 null mice appeared normal, were fertile, and showed no obvious skeletal malformations [66].
261 This suggested functional redundancy among matrilins, indicating that MED disorder
262 phenotypes might not be caused by the absence of matrilin-3 in ECM [66]. However, in a
263 second study, aged *Matn3* null mice were much more predisposed to develop severe OA than
264 their wild-type littermates. Also, the lack of *Matn3* did not lead to postnatal chondrodysplasia
265 but yet accounted for higher incidence of late-onset OA [67].

266

267 **Mutations in the *SMAD3* gene: involvement in the TGF- β pathway**

268 Members of the **transforming growth factor-beta** (TGF- β) superfamily represent a major
269 signaling pathway in joints and are involved in numerous cellular processes in cartilage [2].
270 Mice harboring mutations in members of the TGF- β signaling pathway display phenotypes
271 similar to human osteoarthritis. For instance, mice overexpressing a dominant-negative form of
272 *Tgfb2* [68] and *Smad3*^{-/-} mice [69] have shown severe progressive osteoarthritis-like disease.
273 To date, only the *SMAD3* gene has been screened for mutations in human OA patients. This
274 gene encodes a transcriptional modulator activated by transforming growth factor-beta (TGF-
275 β) that plays a role in the anabolic pathway in cartilage [70].

276 Based on the phenotype observed in an animal model, *SMAD3* exons were screened in 32
277 patients with sporadic knee OA [71]. It is important to note that this cohort was not evaluated
278 for other anomalies. A missense mutation (c.590A>T, p.Asn197Ile) was identified in one
279 patient, with higher expression of pro-MMP-2 and pro-MMP-9 in serum relative to controls
280 and to non-carrier OA patients. This suggests that this mutation may lead to upregulation of
281 MMPs and eventually to the development of OA. However, (i) this mutation is located in the
282 linker region of *SMAD3* which is not well conserved, (ii) only one sporadic OA case carried a
283 *SMAD3* mutation (no replication cohorts were screened to identify a *SMAD3* mutation in

284 additional cases), and (iii) neither the prematurity nor the age of onset of OA were specified.
285 Subsequently, *SMAD3* mutations were identified in patients with **aneurysms-osteoarthritis**
286 **syndrome** (AOS)[72]. The evaluation of the clinical phenotype of 45 patients from eight
287 different AOS families with eight different *SMAD3* mutations showed that radiologically
288 validated OA in 96% of the 26 examined patients, with 75% having two or more affected joints
289 [73]. Early-onset joint abnormalities, including OA, intervertebral disc degeneration, OD and
290 meniscal anomalies were present in almost all patients with AOS, in contrast to other forms of
291 thoracic aortic aneurysms and dissections in which these anomalies are rarely described. This
292 establishes early-onset joint abnormalities as a key feature of this new syndrome. The causal
293 role of heterozygous loss of function mutations in the *SMAD3* gene were further validated by
294 independent studies in human [74,75]. Recently, the clinical spectrum of *SMAD3* mutations
295 were completed on 50 mutation carriers [74]. Osteoarticular manifestations were recorded in
296 all patients. Joint involvement could be severe, requiring surgery in young patients (at a mean
297 age of 33), with unusual localization such as tarsus, shoulder and carpus, or mimicking
298 chondrocalcinosis. In contrast, other cohorts did not report OA, joint pain or deformities in
299 some *SMAD3* mutation carriers [76,77]. However, either the individuals did not undergo
300 imaging, with the diagnosis of OA and joint pain based on medical records/interviews to assess
301 joint pain and potentially leading to the underestimation of OA cases [76], or, mutation carriers
302 were young, which could explain the absence of radiographic signs of OA [77]. In addition, as
303 mutations were mostly described in the context of cardiac disease, the apparent absence of OA
304 could be explained by incomplete clinical investigation or description (X-rays are not
305 systematic). Recently, the vascular phenotypes of *Smad3* knockout mice were investigated,
306 showing that this mouse model also replicates clinical vascular aspects of AOS, marked by the
307 progressive development of aneurysms [78,79].

308 **Mutations in genes encoding proteins involved in other functions and pathways**

309 The trafficking protein particle complex 2 gene (*TRAPPC2*) encodes the sedlin protein that
310 plays a role in vesicular transport from endoplasmic reticulum to Golgi. *TRAPPC2* mutations
311 are causal for **spondyloepiphyseal dysplasia tarda** (SED_T)[80]. Distinctive radiological signs
312 include mild to moderate epiphyseal dysplasia which usually leads to severe premature OA of
313 large joints frequently necessitating hip joint replacement in the third or fourth decade [81]. As
314 a majority of mutations identified in *TRAPPC2* lead to protein truncation, and based on the
315 speculation that less severe missense mutations may have different phenotypic effects such as
316 EO-OA alone, a cohort of 37 male patients with early end-stage primary hip or knee OA were
317 screened for *TRAPPC2* mutations [82]. Secondary OA risk factors were excluded. No
318 *TRAPPC2* mutations were identified in the entire coding region indicating that *TRAPPC2*
319 mutations are not a common cause of early primary OA in men. However, neither non-coding
320 regions nor introns of *TRAPPC2* were screened, and other known causal genes in EO-OA were
321 not investigated.

322

323 In the last few years, whole-exome sequencing has been successfully applied to identify the
324 molecular basis of numerous Mendelian diseases. In 2015, Ramos *et al.* used this approach for
325 the first time in patients with EO-OA and particularly in two distant family members with
326 dominantly inherited primary EO-OA at multiple joint sites with radiographic
327 **chondrocalcinosis** [5]. The age of onset of OA in the family varies between 30 years and 50
328 years, and OA arises in the absence of mild or severe chondrodysplasia. Thanks to this high-
329 throughput technique, the causal mutation for the familial EO-OA phenotype was identified in
330 the *TNFRSF11B* gene (tumor necrosis factor receptor superfamily, member 11b gene) that
331 encodes the osteoprotegerin (OPG) protein, a member of the tumor necrosis factor receptor
332 superfamily. This protein plays an important role in bone remodeling. The power of this method

333 is highlighted by the fact that candidate gene screening and genetic linkage analysis previously
334 failed to reveal the molecular basis of OA development in this large family. The heterozygous
335 stop loss mutation (c.1205A>T, p.Ter402Leu) cosegregates with the phenotype of OA in the
336 family and is absent in 744 controls. Functional analyses showed that this mutation establishes
337 a gain of function of the encoding protein OPG. The authors hypothesize that the mutation acts
338 *via* an unfavorable interplay between subchondral bone and cartilage towards enhanced matrix
339 mineralization, which is a major hallmark of OA disease process [83].

340

341 **Concluding Remarks**

342 Primary EO-OA is a genetically heterogeneous disease. Several mutations in genes encoding
343 components of the ECM or involved in distinct biological pathways (TGF- β , vesicular transport
344 or bone remodeling) have been identified in idiopathic EO-OA patients or in patients having
345 syndromic forms of EO-OA. Mutations in these genes have only been reported in a few studies
346 in human due to the rarity of EO-OA cases. In addition, there is little genetic overlap between
347 early- and late-onset OA, with only *COL11A1* and *SMAD3* genes that harbor mutation or
348 polymorphism for both types of OA [84,85]. This suggests that EO-OA and common late-onset
349 OA are two very different types of OA from a molecular genetics perspective.

350 The genetic heterogeneity observed in primary EO-OA can be explained in part by various
351 clinical features associated with EO-OA, mainly observed in syndromic diseases. In the latter,
352 OA is not systematically the key feature studied, leading to a lack of phenotypic information.
353 In addition, the range of clinical and radiological definitions of OA used in genetic studies
354 results in difficulties when interpreting the causal role of mutations in OA determinism, thus
355 complicating a clear genotype-to-phenotype description.

356

357 Further studies are required to clarify the molecular basis of EO-OA (Outstanding Questions).
358 First, only *COL2A1* mutations are clearly involved in EO-OA determinism. The causal role of
359 other genes needs to be confirmed by independent studies in idiopathic EO-OA patients.
360 Second, assessment of the prevalence of gene mutations in idiopathic EO-OA is needed to
361 determine their respective share in EO-OA development. Third, establishment of a genotype-
362 to-phenotype correlation in large cohorts of both idiopathic and syndromic forms of EO-OA
363 may give clues as to which classes of mutations or relevant protein domains are specific to a
364 particular phenotype. For this purpose, the use of an international recognized definition for EO-
365 OA is important as well as a complete description of the patients' phenotypes. Of note, imaging
366 is absolutely necessary in establishing OA diagnosis, as the disease can be asymptomatic,
367 particularly in young individuals.

368

369 To highlight a new molecular basis for EO-OA, future studies should focus on patients with
370 severe phenotypes, as these are the most informative cases. Clear molecular basis, prevalence
371 data, and genotype-to-phenotype correlations are essential to improve EO-OA diagnosis,
372 management and therapy. For this end, the ExoRhum project (NCT01999166) aims to identify
373 new gene mutations involved in EO-OA pathogenesis, and to estimate the prevalence of
374 mutations in known causal genes (<https://clinicaltrials.gov/ct2/show/NCT01999166>). This
375 genetic study will be performed on patients with idiopathic EO-OA, based on a strict definition
376 of OA and excluding known causes of secondary OA. It will rely on whole exome sequencing,
377 an approach that has been successfully applied to identify the molecular basis of numerous
378 Mendelian diseases, and which has recently enabled to reveal a gain of function mutation in the
379 *TNFRSF11B* gene in two distant family members with idiopathic EO-OA [5]. The use of exome
380 sequencing should identify candidate genes harboring nucleotidic variants or copy number
381 variations that have never been studied in the context of EO-OA.

382

383 In conclusion, to-date studies have identified twelve genes potentially causal for EO-OA, that
384 could be targeted for OA treatment. Thus, a gain of function mutation was recently identified
385 in *TNFRSF11B*, thanks to a strict selection of cases and controls and to whole-exome
386 sequencing. Enhanced function of osteoprotegerin, encoded by *TNFRSF11B*, could be the
387 direct underlying cause of OA and hence, a potential therapeutic target for OA.

388

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393

394 **References**

- 395 1 Lawrence, R.C. *et al.* (2008) Estimates of the prevalence of arthritis and other
396 rheumatic conditions in the United States. Part II. *Arthritis Rheum.* 58, 26–35
- 397 2 Baugé, C. *et al.* (2014) Regulation and Role of TGF β Signaling Pathway in Aging and
398 Osteoarthritis Joints. *Aging Dis.* 5, 394–405
- 399 3 Sandell, L.J. (2012) Etiology of osteoarthritis: genetics and synovial joint
400 development. *Nat. Rev. Rheumatol.* 8, 77–89
- 401 4 Min, J.L. *et al.* (2007) Mutation analysis of candidate genes within the 2q33.3 linkage
402 area for familial early-onset generalised osteoarthritis. *Eur. J. Hum. Genet. EJHG* 15, 791–
403 799

404 5 Ramos, Y.F.M. *et al.* (2015) A gain of function mutation in TNFRSF11B encoding
405 osteoprotegerin causes osteoarthritis with chondrocalcinosis. *Ann. Rheum. Dis.* 74, 1756–
406 1762

407 6 Stecher, R. (1941) , Heberden’s nodes: heredity in hypertrophic arthritis of the finger
408 joints. , *Am J Med Sci*

409 7 Kellgren, J.H. *et al.* (1963) Genetic Factors in Generalized Osteo-Arthrosis. *Ann.*
410 *Rheum. Dis.* 22, 237–255

411 8 Spector, T.D. *et al.* (1996) Genetic influences on osteoarthritis in women: a twin
412 study. *BMJ* 312, 940–943

413 9 Tsezou, A. (2014) Osteoarthritis year in review 2014: genetics and genomics.
414 *Osteoarthr. Cartil. OARS Osteoarthr. Res. Soc.* 22, 2017–2024

415 10 Reynard, L.N. and Loughlin, J. (2013) Insights from human genetic studies into the
416 pathways involved in osteoarthritis. *Nat. Rev. Rheumatol.* 9, 573–583

417 11 Reynard, L.N. and Loughlin, J. (2013) The genetics and functional analysis of primary
418 osteoarthritis susceptibility. *Expert Rev. Mol. Med.* 15, e2

419 12 Chapman, K. and Valdes, A.M. (2012) Genetic factors in OA pathogenesis. *Bone* 51,
420 258–264

421 13 Loughlin, J. (2015) Genetic contribution to osteoarthritis development: current state of
422 evidence. *Curr. Opin. Rheumatol.* 27, 284–288

423 14 Panoutsopoulou, K. and Zeggini, E. (2013) Advances in osteoarthritis genetics. *J.*
424 *Med. Genet.* 50, 715–724

425 15 Ramos, Y.F.M. and Meulenbelt, I. (2015) Implementation of Functional Genomics for
426 Bench-to-Bedside Transition in Osteoarthritis. *Curr. Rheumatol. Rep.* 17, 53

427 16 Palotie, A. *et al.* (1989) Predisposition to familial osteoarthrosis linked to type II
428 collagen gene. *Lancet Lond. Engl.* 1, 924–927

429 17 Meulenbelt, I. *et al.* (1997) Genetic linkage analysis of 14 candidate gene loci in a
430 family with autosomal dominant osteoarthritis without dysplasia. *J. Med. Genet.* 34, 1024–
431 1027

432 18 Ingvarsson, T. *et al.* (2001) A large Icelandic family with early osteoarthritis of the hip
433 associated with a susceptibility locus on chromosome 16p. *Arthritis Rheum.* 44, 2548–2555

434 19 Jakkula, E. *et al.* (2005) The role of sequence variations within the genes encoding
435 collagen II, IX and XI in non-syndromic, early-onset osteoarthritis. *Osteoarthritis Cartilage*
436 13, 497–507

437 20 Nakata, K. *et al.* (1993) Osteoarthritis associated with mild chondrodysplasia in
438 transgenic mice expressing alpha 1(IX) collagen chains with a central deletion. *Proc. Natl.*
439 *Acad. Sci. U. S. A.* 90, 2870–2874

440 21 Fang, H. and Beier, F. (2014) Mouse models of osteoarthritis: modelling risk factors
441 and assessing outcomes. *Nat. Rev. Rheumatol.* 10, 413–421

442 22 Staal, B. *et al.* (2014) Cartilage-specific deletion of Mig-6 results in osteoarthritis-like
443 disorder with excessive articular chondrocyte proliferation. *Proc. Natl. Acad. Sci. U. S. A.*
444 111, 2590–2595

445 23 Husar-Memmer, E. *et al.* (2013) Premature osteoarthritis as presenting sign of type II
446 collagenopathy: a case report and literature review. *Semin. Arthritis Rheum.* 42, 355–360

447 24 Nishimura, G. *et al.* (2005) The phenotypic spectrum of COL2A1 mutations. *Hum.*
448 *Mutat.* 26, 36–43

449 25 Knowlton, R.G. *et al.* (1990) Genetic Linkage of a Polymorphism in the Type II
450 Procollagen Gene (COL2A1) to Primary Osteoarthritis Associated with Mild
451 Chondrodysplasia. *N. Engl. J. Med.* 322, 526–530

452 26 Ala-Kokko, L. *et al.* (1990) Single base mutation in the type II procollagen gene
453 (COL2A1) as a cause of primary osteoarthritis associated with a mild chondrodysplasia. *Proc.*
454 *Natl. Acad. Sci. U. S. A.* 87, 6565–6568

455 27 Ritvaniemi, P. *et al.* (1995) Identification of COL2A1 gene mutations in patients with
456 chondrodysplasias and familial osteoarthritis. *Arthritis Rheum.* 38, 999–1004

457 28 Lankisch, P. *et al.* (2014) COL2A1 mutation as a cause of premature osteoarthritis in a
458 13-year-old child. *Joint Bone Spine* 81, 83–85

459 29 Horton, W.A. *et al.* (1992) Characterization of a type II collagen gene (COL2A1)
460 mutation identified in cultured chondrocytes from human hypochondrogenesis. *Proc. Natl.*
461 *Acad. Sci. U. S. A.* 89, 4583–4587

462 30 Kannu, P. *et al.* (2012) Clinical phenotypes associated with type II collagen mutations.
463 *J. Paediatr. Child Health* 48, E38–43

464 31 Säämänen, A.-M.K. *et al.* (2000) Osteoarthritis-like lesions in transgenic mice
465 harboring a small deletion mutation in type II collagen gene. *Osteoarthritis Cartilage* 8, 248–
466 257

467 32 Holt, D.W. *et al.* (2012) Osteoarthritis-like changes in the heterozygous sedc mouse
468 associated with the HtrA1-Ddr2-Mmp-13 degradative pathway: a new model of osteoarthritis.
469 *Osteoarthr. Cartil. OARS Osteoarthr. Res. Soc.* 20, 430–439

470 33 Macdonald, D.W. *et al.* (2013) Structural variations in articular cartilage matrix are
471 associated with early-onset osteoarthritis in the spondyloepiphyseal dysplasia congenita (sedc)
472 mouse. *Int. J. Mol. Sci.* 14, 16515–16531

473 34 Muragaki, Y. *et al.* (1996) A mutation in the gene encoding the alpha 2 chain of the
474 fibril-associated collagen IX, COL9A2, causes multiple epiphyseal dysplasia (EDM2). *Nat.*
475 *Genet.* 12, 103–105

476 35 Paassilta, P. *et al.* (1999) COL9A3: A third locus for multiple epiphyseal dysplasia.
477 *Am. J. Hum. Genet.* 64, 1036–1044

478 36 Czarny-Ratajczak, M. *et al.* (2001) A mutation in COL9A1 causes multiple epiphyseal
479 dysplasia: further evidence for locus heterogeneity. *Am. J. Hum. Genet.* 69, 969–980

480 37 Fässler, R. *et al.* (1994) Mice lacking alpha 1 (IX) collagen develop noninflammatory
481 degenerative joint disease. *Proc. Natl. Acad. Sci. U. S. A.* 91, 5070–5074

482 38 Jeong, C. *et al.* (2014) Novel COL9A3 mutation in a family diagnosed with multiple
483 epiphyseal dysplasia: a case report. *BMC Musculoskelet. Disord.* 15, 371

484 39 Xu, L. *et al.* (2003) Osteoarthritis-like changes and decreased mechanical function of
485 articular cartilage in the joints of mice with the chondrodysplasia gene (cho). *Arthritis Rheum.*
486 48, 2509–2518

487 40 Gleghorn, L. *et al.* (2005) A Mutation in the Variable Repeat Region of the Aggrecan
488 Gene (AGC1) Causes a Form of Spondyloepiphyseal Dysplasia Associated with Severe,
489 Premature Osteoarthritis. *Am. J. Hum. Genet.* 77, 484–490

490 41 Stattin, E.-L. *et al.* (2010) A missense mutation in the aggrecan C-type lectin domain
491 disrupts extracellular matrix interactions and causes dominant familial osteochondritis
492 dissecans. *Am. J. Hum. Genet.* 86, 126–137

493 42 Aspberg, A. (2012) The different roles of aggrecan interaction domains. *J. Histochem.*
494 *Cytochem. Off. J. Histochem. Soc.* 60, 987–996

495 43 Tompson, S.W. *et al.* (2009) A recessive skeletal dysplasia, SEMD aggrecan type,
496 results from a missense mutation affecting the C-type lectin domain of aggrecan. *Am. J. Hum.*
497 *Genet.* 84, 72–79

498 44 Watanabe, H. *et al.* (1994) Mouse cartilage matrix deficiency (cmd) caused by a 7 bp
499 deletion in the aggrecan gene. *Nat. Genet.* 7, 154–157

500 45 Acharya, C. *et al.* (2014) Cartilage oligomeric matrix protein and its binding partners
501 in the cartilage extracellular matrix: Interaction, regulation and role in chondrogenesis. *Matrix*
502 *Biol.* 37, 102–111

503 46 Hecht, J.T. *et al.* (1995) Mutations in exon 17B of cartilage oligomeric matrix protein
504 (COMP) cause pseudoachondroplasia. *Nat. Genet.* 10, 325–329

505 47 Briggs, M.D. *et al.* (1995) Pseudoachondroplasia and multiple epiphyseal dysplasia
506 due to mutations in the cartilage oligomeric matrix protein gene. *Nat. Genet.* 10, 330–336

507 48 Jackson, G.C. *et al.* (2012) Pseudoachondroplasia and Multiple Epiphyseal Dysplasia:
508 A 7-Year Comprehensive Analysis of the Known Disease Genes Identify Novel and
509 Recurrent Mutations and Provides an Accurate Assessment of Their Relative Contribution.
510 *Hum. Mutat.* 33, 144–157

511 49 Kennedy, J. *et al.* (2005) COMP mutation screening as an aid for the clinical diagnosis
512 and counselling of patients with a suspected diagnosis of pseudoachondroplasia or multiple
513 epiphyseal dysplasia. *Eur. J. Hum. Genet. EJHG* 13, 547–555

514 50 Briggs, M.D. and Chapman, K.L. (2002) Pseudoachondroplasia and multiple
515 epiphyseal dysplasia: mutation review, molecular interactions, and genotype to phenotype
516 correlations. *Hum. Mutat.* 19, 465–478

517 51 Unger, S. and Hecht, J.T. (2001) Pseudoachondroplasia and multiple epiphyseal
518 dysplasia: New etiologic developments. *Am. J. Med. Genet.* 106, 244–250

519 52 Briggs, M.D. *et al.* (1998) Diverse mutations in the gene for cartilage oligomeric
520 matrix protein in the pseudoachondroplasia-multiple epiphyseal dysplasia disease spectrum.
521 *Am. J. Hum. Genet.* 62, 311–319

522 53 Posey, K.L. *et al.* (2014) Pseudoachondroplasia/COMP — translating from the bench
523 to the bedside. *Matrix Biol. J. Int. Soc. Matrix Biol.* 0, 167–173

524 54 Briggs, M.D. *et al.* (2014) Genotype to phenotype correlations in cartilage oligomeric
525 matrix protein associated chondrodysplasias. *Eur. J. Hum. Genet.* 22, 1278–1282

526 55 Mu, S.-C. *et al.* (2009) A large kindred of early-onset osteoarthritis of the knee and
527 hip: excluding the link to COL2A1 gene. *Rheumatology* 48, 371–374

528 56 Jakkula, E. *et al.* (2003) A recurrent R718W mutation in COMP results in multiple
529 epiphyseal dysplasia with mild myopathy: clinical and pathogenetic overlap with collagen IX
530 mutations. *J. Med. Genet.* 40, 942–948

531 57 Mabuchi, A. *et al.* (2003) Novel types of COMP mutations and genotype-phenotype
532 association in pseudoachondroplasia and multiple epiphyseal dysplasia. *Hum. Genet.* 112, 84–
533 90

534 58 Chapman, K.L. *et al.* (2001) Mutations in the region encoding the von Willebrand
535 factor A domain of matrilin-3 are associated with multiple epiphyseal dysplasia. *Nat. Genet.*
536 28, 393–396

537 59 Mabuchi, A. *et al.* (2004) Novel and recurrent mutations clustered in the von
538 Willebrand factor A domain of MATN3 in multiple epiphyseal dysplasia. *Hum. Mutat.* 24,
539 439–440

540 60 Mäkitie, O. *et al.* (2004) Clinical and radiographic findings in multiple epiphyseal
541 dysplasia caused by MATN3 mutations: description of 12 patients. *Am. J. Med. Genet. A.*
542 125A, 278–284

543 61 Cotterill, S.L. *et al.* (2005) Multiple epiphyseal dysplasia mutations in MATN3 cause
544 misfolding of the A-domain and prevent secretion of mutant matrilin-3. *Hum. Mutat.* 26, 557–
545 565

546 62 Otten, C. *et al.* (2005) Matrilin-3 mutations that cause chondrodysplasias interfere
547 with protein trafficking while a mutation associated with hand osteoarthritis does not. *J. Med.*
548 *Genet.* 42, 774–779

549 63 Leighton, M.P. *et al.* (2007) Decreased chondrocyte proliferation and dysregulated
550 apoptosis in the cartilage growth plate are key features of a murine model of epiphyseal
551 dysplasia caused by a *matn3* mutation. *Hum. Mol. Genet.* 16, 1728–1741

552 64 Stefánsson, S.E. *et al.* (2003) Genomewide Scan for Hand Osteoarthritis: A Novel
553 Mutation in Matrilin-3. *Am. J. Hum. Genet.* 72, 1448–1459

554 65 Otten, C. *et al.* (2010) A matrilin-3 mutation associated with osteoarthritis does not
555 affect collagen affinity but promotes the formation of wider cartilage collagen fibrils. *Hum.*
556 *Mutat.* 31, 254–263

557 66 Ko, Y. *et al.* (2004) Matrilin-3 is dispensable for mouse skeletal growth and
558 development. *Mol. Cell. Biol.* 24, 1691–1699

559 67 van der Weyden, L. *et al.* (2006) Functional knockout of the matrilin-3 gene causes
560 premature chondrocyte maturation to hypertrophy and increases bone mineral density and
561 osteoarthritis. *Am. J. Pathol.* 169, 515–527

562 68 Serra, R. *et al.* (1997) Expression of a Truncated, Kinase-Defective TGF- β Type II
563 Receptor in Mouse Skeletal Tissue Promotes Terminal Chondrocyte Differentiation and
564 Osteoarthritis. *J. Cell Biol.* 139, 541–552

565 69 Yang, X. *et al.* (2001) TGF-beta/Smad3 signals repress chondrocyte hypertrophic
566 differentiation and are required for maintaining articular cartilage. *J. Cell Biol.* 153, 35–46

567 70 Zhu, Y. *et al.* (2015) Transforming growth factor- β 1 induces type II collagen and
568 aggrecan expression via activation of extracellular signal-regulated kinase 1/2 and Smad2/3
569 signaling pathways. *Mol. Med. Rep.* 12, 5573–5579

570 71 Yao, J.-Y. *et al.* (2003) Mutation analysis of the Smad3 gene in human osteoarthritis.
571 *Eur. J. Hum. Genet.* 11, 714–717

572 72 van de Laar, I.M.B.H. *et al.* (2011) Mutations in SMAD3 cause a syndromic form of
573 aortic aneurysms and dissections with early-onset osteoarthritis. *Nat. Genet.* 43, 121–126

574 73 van de Laar, I.M.B.H. *et al.* (2012) Phenotypic spectrum of the SMAD3-related
575 aneurysms–osteoarthritis syndrome. *J. Med. Genet.* 49, 47–57

576 74 Aubart, M. *et al.* (2014) Early-Onset Osteoarthritis, Charcot-Marie-Tooth Like
577 Neuropathy, Autoimmune Features, Multiple Arterial Aneurysms and Dissections: An
578 Unrecognized and Life Threatening Condition. *PLoS ONE* 9,

579 75 Zhang, W. *et al.* (2015) A Novel Mutation of SMAD3 Identified in a Chinese Family
580 with Aneurysms-Osteoarthritis Syndrome. *BioMed Res. Int.* 2015,

581 76 Regalado, E.S. *et al.* (2011) Exome Sequencing Identifies SMAD3 Mutations as a
582 Cause of Familial Thoracic Aortic Aneurysm and Dissection with Intracranial and Other
583 Arterial Aneurysms. *Circ. Res.* 109, 680–686

584 77 Wischmeijer, A. *et al.* (2013) Thoracic Aortic Aneurysm in Infancy in Aneurysms–
585 Osteoarthritis Syndrome Due to a Novel SMAD3 Mutation: Further Delineation of the
586 Phenotype. *Am. J. Med. Genet. A.* 161, 1028–1035

587 78 Ye, P. *et al.* (2013) GM-CSF contributes to aortic aneurysms resulting from SMAD3
588 deficiency. *J. Clin. Invest.* 123, 2317–2331

589 79 Tan, C.K. *et al.* (2013) SMAD3 Deficiency Promotes Inflammatory Aortic Aneurysms
590 in Angiotensin II–Infused Mice Via Activation of iNOS. *J. Am. Heart Assoc. Cardiovasc.*
591 *Cerebrovasc. Dis.* 2,

592 80 Gedeon, A.K. *et al.* (1999) Identification of the gene (SEDL) causing X-linked
593 spondyloepiphyseal dysplasia tarda. *Nat. Genet.* 22, 400–404

594 81 Bannerman, R.M. *et al.* (1971) X-linked spondyloepiphyseal dysplasia tarda: clinical
595 and linkage data. *J. Med. Genet.* 8, 291–301

596 82 Fiedler, J. *et al.* (2002) Mutations in the X-linked spondyloepiphyseal dysplasia tarda
597 (SEDL) coding sequence are not a common cause of early primary osteoarthritis in men. *Clin.*
598 *Genet.* 62, 94–95

599 83 Goldring, M.B. and Goldring, S.R. (2010) Articular cartilage and subchondral bone in
600 the pathogenesis of osteoarthritis. *Ann. N. Y. Acad. Sci.* 1192, 230–237

601 84 Rodriguez-Fontenla, C. *et al.* (2014) Assessment of osteoarthritis candidate genes in a
602 meta-analysis of nine genome-wide association studies. *Arthritis Rheumatol. Hoboken NJ* 66,
603 940–949

604 85 Valdes, A.M. *et al.* (2010) Genetic variation in the SMAD3 gene is associated with hip
605 and knee osteoarthritis. *Arthritis Rheum.* 62, 2347–2352

606 86 Moon, S. *et al.* (2015) A genome-wide association study of copy-number variation
607 identifies putative loci associated with osteoarthritis in Koreans. *BMC Musculoskelet. Disord.*
608 16, 76

609 87 Panoutsopoulou, K. *et al.* (2011) Insights into the genetic architecture of osteoarthritis
610 from stage 1 of the arcOGEN study. *Ann. Rheum. Dis.* 70, 864–867

611 88 Mu, S.-C. *et al.* (2011) A mutation in cartilage oligomeric matrix protein (COMP)
612 causes early-onset osteoarthritis in a large kindred study. *Ann. Hum. Genet.* 75, 575–583

613 89 Myllyharju, J. (2014) Extracellular Matrix and Developing Growth Plate. *Curr.*
614 *Osteoporos. Rep.* 12, 439–445

615

616 **Figure legends**

617 **Figure 1: Radiographic phenotype of osteoarthritis in humans**

618 (A) Normal knee joint. (B) Osteoarthritis of the knee located in the medial femoro-tibial
619 compartment: joint space narrowing, tibial and femoral **osteophytosis**, **subchondral bone**
620 **geode** of the medial tibial plateau.

621

622 **Figure 2, Key Figure: Proteins involved in early-onset osteoarthritis pathogenesis**

623 (A) *Articular cartilage: composition and metabolism.* Chondrocytes are surrounded by ECM
624 which confers stiffness and elasticity to cartilage. The TGF- β pathway has been implicated in
625 promoting cartilage synthesis. Briefly, TGF- β binds to TGFBR2 leading to the recruitment of
626 TGFBR1, which subsequently phosphorylates SMAD2 or SMAD3. Activated SMAD2 or
627 SMAD3 complexes with SMAD4, translocating into the nucleus. The Smad complex binds to
628 the promoter regions of TGF- β target genes, interacting with other transcription factors and co-
629 factors, and inducing the transcription of target genes encoding EMC components. Components
630 of ECM are secreted by chondrocytes by vesicle-mediated transport. Sedlin is a part of the
631 TRAPP complex, required for the endoplasmic reticulum export of procollagen. ECM consists
632 of collagens (type II, IX and XI), proteoglycans (aggrecan, biglycan, decorin, fibromodulin),
633 other non-collagenous proteins (link protein, fibronectin, COMP, matrilin-3, integrin) and
634 hyaluronan.

635 (B) *Bone resorption.* OPG is a soluble decoy receptor which inhibits **osteoclastogenesis** by
636 competing with RANK (expressed on the membrane of pre-**osteoclasts**) in binding its ligand,
637 RANKL. Gain of function mutations of OPG lead to a more efficient antagonism of
638 osteoclastogenesis.

639 Proteins encoded by genes involved in EO-OA pathogenesis are shown in red.

640 Abbreviations: co-F, cofactors; COMP, cartilage oligomeric matrix protein; ECM, extracellular
641 matrix; OPG, osteoprotegerin; RANK, receptor activator of the nuclear factor- κ B; RANKL,
642 RANK ligand; TF, transcription factor; TGF- β , transforming growth factor- β ; TGFBR1/2,
643 TGF- β receptor type-1/2; TRAPP, trafficking protein particle.

644

645 **Glossary**

646 **Aneurysms-osteoarthritis syndrome (AOS):** syndromic form of autosomal dominant thoracic
647 aortic aneurysms and dissections characterised by the presence of arterial aneurysms and
648 tortuosity, mild craniofacial, skeletal and cutaneous anomalies and EO-OA.

649 **Animal model:** laboratory animals used to study disease mechanisms. Results can be
650 extrapolated to human disorders.

651 **Chondrocalcinosis:** deposition of calcium salts in the cartilage of joints.

652 **Chondrocyte:** cell found in cartilage, that produces extra-cellular matrix components.

653 **Chondrodysplasia:** bone disease characterized by abnormal growth of the epiphyses.

654 **Cleft palate:** opening in the roof of the mouth.

655 **Early-onset osteoarthritis (EO-OA):** osteoarthritis that occurs before 50 years old, rare and
656 mainly familial.

657 **Epiphyse:** end of a long bone.

658 **Heberden's nodes:** bony enlargement of the terminal joint of a finger.

659 **Late-onset osteoarthritis:** osteoarthritis that occurs after 50 years old, frequent.

660 **Multiple epiphyseal dysplasia (MED):** relatively mild and clinically variable
661 osteochondrodysplasia, characterized by delayed and irregular ossification of the epiphyses and
662 EO-OA.

663 **Non-ophthalmic Stickler syndrome:** clinically variable disorder characterized by auditory,
664 skeletal and orofacial abnormalities.

665 **Osteoarthritis (OA):** degenerative joint disease, mainly characterized by the gradual loss of
666 articular cartilage, particularly affecting the knee, hip, spine, hand and foot.

667 **Osteochondritis dissecans (OD):** skeletal disorder defined as a separation of cartilage and
668 subchondral bone from the surrounding tissue which primarily affects the knee, ankle and elbow
669 joints.

670 **Osteochondrodysplasia:** abnormal growth or development of cartilage and bone.

671 **Osteoclast:** cell involved in bone resorption.

672 **Osteoclastogenesis:** development of osteoclasts.

673 **Osteophytosis:** presence of one or more osteophytes which are bony growths that develop on
674 bone extremities.

675 **Otospondylomegaepiphyseal dysplasia (OSMED):** characterized by sensorineural hearing
676 loss, enlarged epiphyses, disproportionate shortness of the limbs, abnormalities in vertebral
677 bodies and typical facial features.

678 **Platyspondyly:** radiographic feature referring to flattened vertebral bodies throughout the axial
679 skeleton.

680 **Polyphen *in silico* analysis:** tool which predicts possible impact of an amino acid substitution
681 on the structure and function of a human protein using straightforward physical and
682 comparative considerations.

683 **Pseudoachondroplasia (PSACH):** osteochondrodysplasia characterized by disproportionate
684 short stature, deformity of the lower limbs, brachydactyly, loose joints and ligamentous laxity.

685 **Robin sequence:** set of abnormalities affecting the head and face (micrognathia, glossoptosis
686 and cleft palate).

687 **Spondyloepiphyseal dysplasia:** disorder of bone growth that results in short stature, skeletal
688 abnormalities and problems with vision and hearing.

689 **Spondyloepiphyseal dysplasia, Kimberley type (SEDK):** characterized by short stature and
690 premature degenerative arthropathy.

691 **Spondyloepiphyseal dysplasia tarda (SEDT):** rare X-linked recessive
692 osteochondrodysplasia, characterized by disproportionate short stature.

693 **Stickler syndrome (STL):** clinically variable disorder characterized by ocular, auditory,
694 skeletal and orofacial abnormalities.

695 **Subchondral bone geode:** lytic lesion in the periarticular surfaces.

696 **Syndrome:** group of signs and symptoms that occur together and characterize a particular
697 abnormality.

698 **Transforming growth factor-beta (TGF- β) pathway:** major signaling pathway in joints,
699 involved in numerous cellular processes in cartilage.

700

701

702 **Table 1: Pathologies associated with secondary early-onset osteoarthritis**
703 **development**

| Pathology | Phenotype MIM number | Clinical features |
|--|----------------------------|---|
| OA secondary to excessive biomechanical constraints | | |
| Post-traumatic OA | | |
| Osteochondritis dissecans (OD) | #165800 | skeletal disorder defined as a separation of cartilage and subchondral bone from the surrounding tissue which primarily affects the knee, ankle, and elbow joints. Familial OD is characterized by multiple osteochondritic lesions in knees and/or hips and/or elbows, |

| | | |
|---|---------|--|
| | | disproportionate short stature, and early-onset osteoarthritis. |
| Pseudoachondroplasia (PSACH) | #177170 | osteochondrodysplasia characterized by disproportionate short stature, deformity of the lower limbs, brachydactyly, loose joints, and ligamentous laxity. Vertebral anomalies, present in childhood, usually resolve with age, but osteoarthritis is progressive and severe. Exhibits a similar, but more severe phenotype than MED. |
| Spondyloepiphyseal dysplasia, Kimberley type (SEDK) | #608361 | skeletal disorder with proportionate short stature and early-onset progressive osteoarthropathy |
| Spondyloepimetaphyseal dysplasia (SEMD) | #612813 | skeletal dysplasia with a predominant epiphyseal component leading to premature osteoarthritis of the weight-bearing large joints |
| OA secondary to weakening of the cartilage, induced by metabolic or inflammatory pathologies | | |

| | | |
|--|---------|--|
| Alkaptonuria (AKU) | #203500 | metabolic disorder characterized by accumulation of homogentisic acid, leading to darkened urine, pigmentation of connective tissue (ochronosis), joint and spine arthritis, and destruction of the cardiac valves |
| Ankylosing spondylitis (AS) | #106300 | common inflammatory rheumatic disease that affects the axial skeleton. AS is the major subtype and a main outcome of an inter-related group of rheumatic diseases now named spondyloarthropathies . |
| Chondrocalcinosis (CCAL) | #600668 | cartilage calcification which is a common condition that usually results from deposition of crystals of calcium pyrophosphate dihydrate (CPPD) in articular hyaline and fibro-cartilage |
| Rheumatoid arthritis (RA) | #180300 | inflammatory disease, primarily of the joints, with autoimmune features |
| OA secondary to joint deformities caused by collagen defect | | |

| | | |
|---|--|--|
| Avascular necrosis of the femoral head (ANFH) in adults | #608805 | debilitating disease that usually leads to destruction of the hip joint in the third to fifth decade of life |
| Legg-Calve-Perthes disease (LCPD) in children | #150600 | form of ANFH in growing children |
| Aneurysms-osteoarthritis syndrome (AOS) also known as Loeys-Dietz syndrome type III | #613795 | aortic aneurysm syndrome with widespread systemic involvement, which is associated with early-onset osteoarthritis |
| Czech dysplasia | #609162 | characterized by normal height, early-onset osteoarthritis, platyspondyly , short metatarsals, but with absence of ophthalmological complications or cleft palate |
| Kniest dysplasia | #156550 | disorder of bone growth characterized by short stature (dwarfism) with other skeletal abnormalities and problems with vision and hearing. |
| Multiple epiphyseal dysplasia (MED) | #132400 #600204 #600969 #607078 | skeletal disorder characterized by short stature and early-onset osteoarthritis. |

| | | |
|---|--|---|
| | #614135 | Similar, but milder phenotype than PSACH |
| Otospondylomegaepiphyseal dysplasia (OSMED) | #215150 | characterized by sensorineural hearing loss, enlarged epiphyses, disproportionate shortness of the limbs, abnormalities in vertebral bodies, and typical facial features |
| Spondyloepiphyseal dysplasia tarda (SEDT) | #313400 | skeletal disorder associated with early-onset osteoarthritis |
| Spondyloperipheral dysplasia | #271700 | disorder that impairs bone growth |
| Stickler syndrome (STL) | #108300 #609508 #604841 #184840 #614134 #614284 | clinically variable disorder characterized by ocular, auditory, skeletal, and orofacial abnormalities. Additional findings may include mild spondyloepiphyseal dysplasia, and early-onset osteoarthritis. |

Table 2: Mutated genes identified in idiopathic early-onset osteoarthritis

| Gene symbol | Approved name | Locus | Mutation ^a | Phenotype | Reference |
|---|----------------------------|-------------|--|--|-----------|
| Genes encoding collagen proteins | | | | | |
| <i>COL2A1</i> | collagen, type II, alpha 1 | 12q12-q13.2 | NM_001844.4:c.823C>T (p.Arg275Cys) | - Familial EO-OA - AO: 12-40y - Bilateral joint sites simultaneously: hip and/or knee | [28] |
| | | | NM_001844.4:c.2155C>T (p.Arg719Cys) | - Familial generalized EO-OA - AO: 2 nd -3 rd decade - Multiple joint sites simultaneously: hip, knee, shoulder, wrist, hand | [26] |
| | | | NM_001844.4:c.2659G>A (p.=) ^b | - Familial EO-OA - AO: 30-50y - Multiple joint sites | [19] |

| | | | | | |
|--|---|-------------|---|--|------|
| | | | | simultaneously: hip, knee, hand | |
| <i>COL11A2</i> | collagen, type XI, alpha 2 | 6p21.3 | NM_080680.2:c.1615 C>T (p.Arg539Trp) | - Familial EO-OA - AO: 38y (proband), <50y (family members) - Multiple joint sites simultaneously: hip, knee, hand, spine | [19] |
| | | | NM_080680.2:c.3151- 2delA (p.?) | - Familial EO-OA - AO: 17-20y - Multiple joint sites simultaneously: knee, hip, spine | [19] |
| Gene encoding other component of the cartilage extracellular matrix | | | | | |
| <i>COMP</i> | cartilage oligomeric matrix protein | 19p13. 1 | NM_000095.2:c.2152 C>T (p.Arg718Trp) | - Familial EO-OA - AO: 29.7 ± 16.1y - Classical OA joint sites involved | [88] |

| Gene encoding protein involved in bone remodeling | | | | | |
|---|--|------|--------------------------------------|---|-----|
| <i>TNFRSF11B</i> | tumor necrosis factor receptor superfamily, member 11b | 8q24 | NM_002546.3:c.1205 A>T (p.Ter402Leu) | - Familial generalized EO-OA with chondrocalcinosis - AO: 30-50 y - Multiple joint sites simultaneously: hand, knee, hip, spine | [5] |

706 ^aMutation at cDNA and protein level according to HGVS nomenclature.

707 ^bSplice mutation associated with decreased mRNA level in carrier.

708 AO, age of onset; EO, early-onset; OA, osteoarthritis

709

710 **Table 3: Causal genes identified in early-onset osteoarthritis forms associated**
 711 **with other clinical features**

| Gene symbol | Approved name | Locus | Associated disease presenting EO-OA ^a |
|--|----------------------------|-------------|--|
| Genes encoding collagen proteins | | | |
| <i>COL2A1</i> | collagen, type II, alpha 1 | 12q12-q13.2 | <ul style="list-style-type: none"> - Familial EO-OA - Avascular necrosis of femoral head - Legg-Calvé-Perthes disease - Czech dysplasia - Kniest dysplasia - Otopondylomegaepiphyseal dysplasia - Spondyloperipheral dysplasia - Stickler syndrome, type I |
| <i>COL9A1</i> | collagen, type IX, alpha 1 | 6q13 | <ul style="list-style-type: none"> - Multiple epiphyseal dysplasia, type VI - Stickler syndrome, type IV |
| <i>COL9A2</i> | collagen, type IX, alpha 2 | 1p33-p32 | <ul style="list-style-type: none"> - Multiple epiphyseal dysplasia, type II - Stickler syndrome, type V |
| <i>COL9A3</i> | collagen, type IX, alpha 3 | 20q13.3 | <ul style="list-style-type: none"> - Multiple epiphyseal dysplasia, type III - Stickler syndrome |
| <i>COL11A1</i> | collagen, type XI, alpha 1 | 1p21 | <ul style="list-style-type: none"> - Stickler syndrome, type II |
| <i>COL11A2</i> | collagen, type XI, alpha 2 | 6p21.3 | <ul style="list-style-type: none"> - Familial EO-OA - Otopondylomegaepiphyseal dysplasia - Stickler syndrome, type III |
| Genes encoding other components of the cartilage extracellular matrix | | | |

| | | | |
|---|--|-----------|---|
| <i>ACAN</i> | aggrecan | 15q26.1 | <ul style="list-style-type: none"> - Osteochondritis dissecans - Spondyloepimetaphyseal dysplasia - Spondyloepiphyseal dysplasia, Kimberley type |
| <i>COMP</i> | cartilage oligomeric matrix protein | 19p13.1 | <ul style="list-style-type: none"> - Familial EO-OA - Multiple epiphyseal dysplasia, type I - Pseudoachondroplasia |
| <i>MATN3</i> | matrilin 3 | 2p24-p23 | <ul style="list-style-type: none"> - Multiple epiphyseal dysplasia, type V |
| Gene encoding protein involved in the TGF-β pathway | | | |
| <i>SMAD3</i> | SMAD family member 3 | 15q21-q22 | <ul style="list-style-type: none"> - Aneurysms-osteoarthritis syndrome |
| Gene encoding protein involved in vesicular transport | | | |
| <i>TRAPPC2</i> | trafficking protein particle complex 2 | Xp22 | <ul style="list-style-type: none"> - Spondyloepiphyseal dysplasia tarda |
| Gene encoding protein involved in bone remodeling | | | |
| <i>TNFRSF11B</i> | tumor necrosis factor receptor superfamily, member 11b | 8q24 | <ul style="list-style-type: none"> - Familial EO-OA |

713 **Box 1: Description of the collagen types**

714 Collagens are structural components of the ECM secreted by **chondrocytes** [89]. They bring
715 traction resistance to cartilage. The major collagen protein is the fibril-forming type II collagen,
716 other collagen types being the type XI and IX collagens. Type IX collagen fibrils are typically
717 found within the core of type II collagen fibrils which help maintain the spacing and diameter
718 of type II collagen fibrils, while type IX collagen resides at the surface of type II/XI fibrils. So
719 mutations in genes encoding collagens lead to loss of resistance to traction, absorption and
720 distribution of forces in cartilage which affect the structure of the cartilage.

721

722 **Box 2: Strict inclusion criteria for primary EO-OA genetic studies**

723 1. No known causes of secondary OA development (e.g. negative trauma history)

724 2. Body mass index (BMI) ≤ 30

725 3. Radiologically proven OA (Kellgren and Lawrence grade ≥ 2)

726 4. Age at onset ≤ 40 years and ≥ 1 joint site involved

727 **or** Age at onset ≤ 50 years **and** multiple joints sites involved (≥ 2 joints)

728 **or** Age at onset ≤ 50 years **and** 1 joint site involved **and** positive family history

729 **or** Age at onset ≤ 50 years **and** 1 unusual joint site involved (shoulder, elbow, wrist, ankle)

730 Criteria adapted from [16]

731

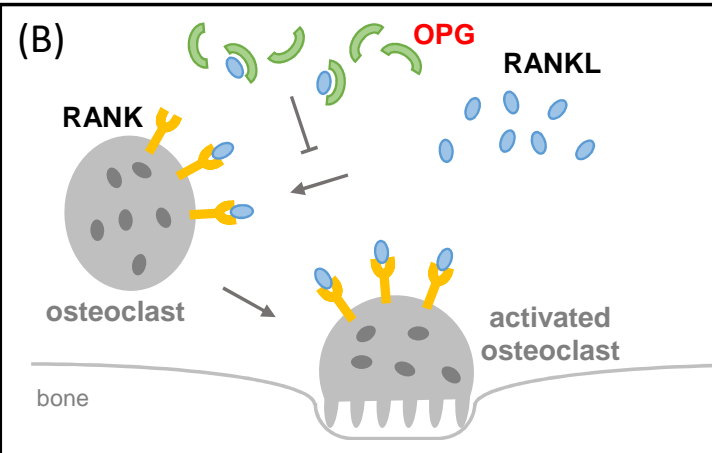
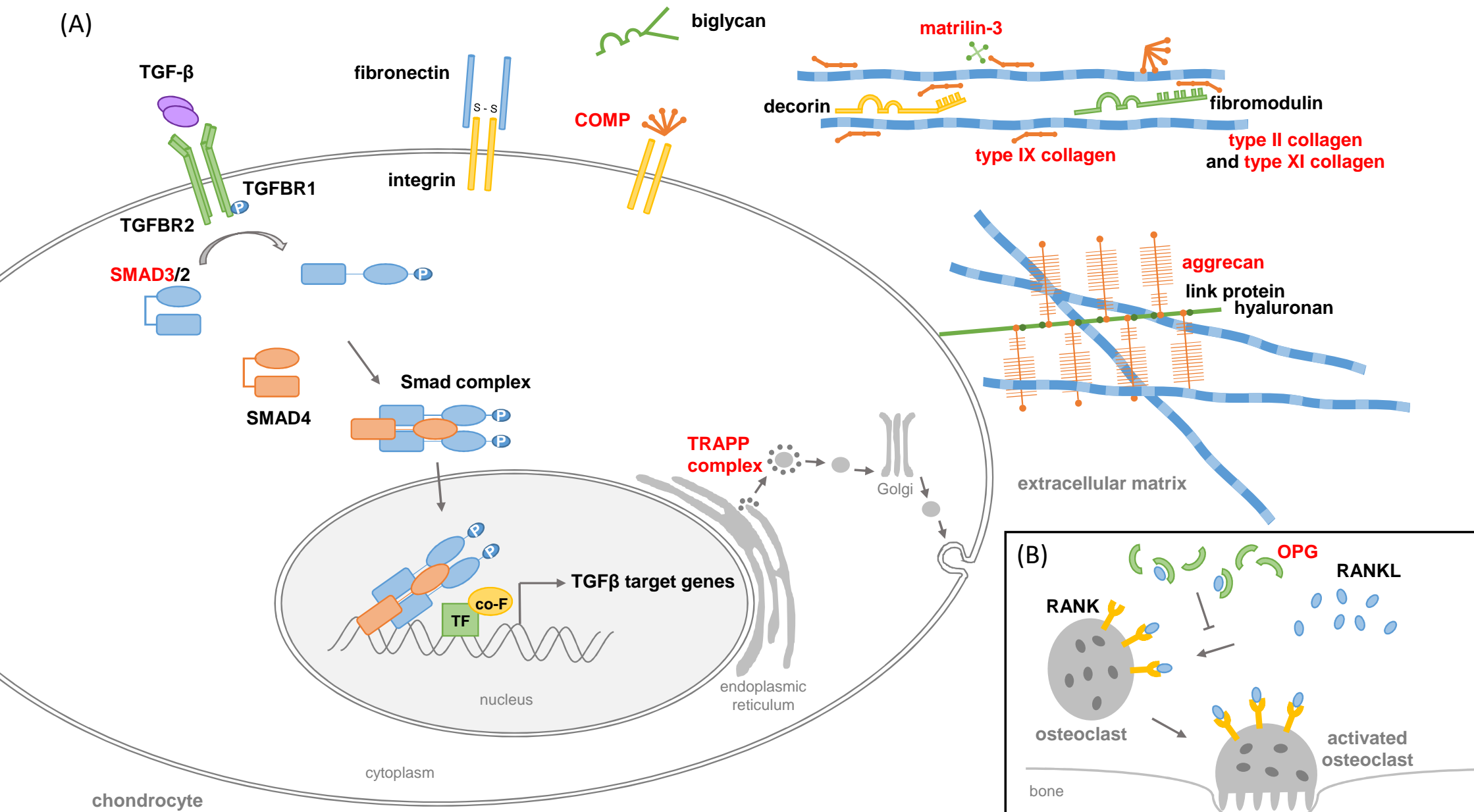
(A)



(B)



(A)



Outstanding questions box

- What is the prevalence of mutations in known causal genes for EO-OA cases? Determining the respective share of each causal gene in EO-OA development will be essential to improve EO-OA diagnosis, in particular to prioritize gene mutations screening.
- Are there genotype-to-phenotype correlations in patients with EO-OA? These correlations will improve EO-OA diagnosis as well as management.
- Are others genes and pathways involved in EO-OA pathogenesis than those yet established? Identification of new genes or pathways could open new therapeutic perspectives as highlighted with the recent description of a gain of function mutation in *TNFRSF11B* gene encoding osteoprotegerin which leads the authors to advocate that agents counteracting the function of osteoprotegerin could be a promising therapy for OA [5].
- What is the contribution of copy number variations in the genetic determinism to EO-OA? Rare copy number variations are involved in the pathogenesis of various diseases such as neurological diseases or cancers. Recently, a genome-wide association study identified putative loci associated with osteoarthritis in Koreans [86]. However, their contribution to OA EO-OA determinism has never been studied and will lead to a better understanding of the pathogenesis of OA.
- Are some idiopathic EO-OA cases oligogenic? Genome-wide association scan data suggests that common osteoarthritis is a highly polygenic disease with multiple risk variants conferring small effects[87]. So we can hypothesis that some idiopathic EO-OA cases harbor few genes mutations and that their combination is causal for the observed phenotype.

Trends box

- By contrast to late-onset OA, idiopathic EO-OA are rare Mendelian forms, mainly including other clinical features such as ocular, auditory or aortic abnormalities.
- Genetic heterogeneity is observed in EO-OA patients. To date, only *COL2A1* mutations are clearly causal for EO-OA. Eleven additional genes need to be further validated, including components of the ECM or proteins involved in the TGF- β pathway, vesicular transport or bone remodeling.
- The rise of high-throughput sequencing technologies give new opportunities, at a reasonable cost, to identify genes or pathways that could be new therapeutic target for OA. Thus, in 2015, a gain of function mutation was identified in *TNFRSF11B* which segregates with a familial form of EO-OA. Enhanced function of osteoprotegerin, encoded by *TNFRSF11B*, could be the direct underlying cause of OA and hence, a potential therapeutic target for OA.