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**Retention of ciliates and flagellates by the oyster *Crassostrea gigas* in French
Atlantic coastal ponds: protists as a trophic link between bacterioplankton
and benthic suspension-feeders.**

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Abstract

In French Atlantic coastal ponds of Charente, oysters can grow under conditions where phytoplankton production is limited by nutrients exhaustion. Such ponds typically show a high concentration of ciliates and flagellates during the growing season (1×10^4 to 3×10^5 cells l^{-1} in June 1997). In order to evaluate the importance of the "protozoan trophic link" for energy transfer from the "microbial food web" to large benthic suspension feeders, we offered a coastal pond community of ciliates and flagellates as potential prey to the oyster, *Crassostrea gigas*. Clearance rate, filtered particles and relative retention efficiency were evaluated. In the grazing experiment, 94 % of ciliates and 86 % of flagellates (size between 4 and 72 μm), were retained by the oyster. Whatever their size, protists were similarly retained by the oyster gills. In terms of carbon, oyster retain on average 126 μg carbon (C) $h^{-1} g^{-1}$ dry weight, a value over 4 times higher than reported for phytoplankton. These results indicate that a field community of protists can contribute in coastal oyster rearing ponds to the energy requirements of the oyster *Crassostrea gigas*. We report here the first experimental evidence of a significant retention of a protist community by oysters, supporting the role of protists as a trophic link between picoplankton and benthic filter-feeding bivalves.

Key-words

Bivalve, oyster, food source, coastal pond, microbial food web, protist, picoplankton, trophic link

INTRODUCTION

Oysters obtain energy resources by filtering particles from sea water and their growth depends upon the nutritive value of the retained seston (Berg & Newell, 1986) and the trophic capacity of coastal waters (Héral, 1987). The natural habitats of the oyster *Crassostrea gigas* are open coastal ecosystems, rocky shores or mud flats. Charente Maritime, on the French Atlantic coast, is the most important European oyster farming area. Shellfish culture has developed in muddy bays (rearing areas of 4 800 ha) and in semi-closed coastal ponds (3 000 ha), characterized by relative confinement and low water renewal rates.

The importance of phytoplankton in the nutrition of oysters is well documented (Héral, 1987; Pastoureaud et al., 1996). However, in oyster rearing environments, such as the particularly light-limited turbid estuary of Marennes-Oléron, or in coastal ponds of Charente where nutrients are quickly exhausted, phytoplankton cannot entirely account for the energy requirements of oysters (Héral, 1987).

In the oceans, more than 50 % of the primary production is due to unicellular organisms less than 3 μm in size (Platt et al., 1983; Li et al., 1983; Glover et al., 1986), which constitutes a nutrient source of particulate and dissolved organic matter for heterotrophic organisms. Dissolved organic matter (DOM) present in coastal waters (Pomeroy and Wiebe, 1993) provides a potential for high bacterial production. Thus, in the Atlantic coastal ponds, bacterioplankton constitutes 50 % of the planktonic carbon biomass (Frikha et al., 1987). Such heterotrophic bacterioplankters, with typically high growth rates and growth efficiencies, represent a significant energy pathway by recycling DOM into particles potentially available to upper trophic levels (Pomeroy, 1974; Azam et al., 1983; Fenchel, 1988).

However, small sized bacteria and autotrophic picoplankton are not retained by gills of bivalves, particularly oysters (Shumway et al., 1985; Héral, 1987; Riisg rd, 1988; Barillé et al., 1993). Flagellate and ciliate protists, which consume bacteria and phytoplankton, are abundant in coastal ecosystems (Revelante & Gilmartin, 1983; Sherr et al., 1986 a; Fenchel, 1988; Leakey et al., 1992) and are preyed upon by numerous organisms of zooplankton, particularly copepods (Berk et al., 1977; Jonsson & Tiselius, 1990; Gifford & Dagg, 1991; Hartmann et al., 1993). Protozoa have been suggested as a major trophic link between picoplankton and micro or macroplankton (Porter et al., 1979; Conover, 1982; Sherr et al., 1986 b; Stoecker & Capuzzo, 1990).

Likewise, protists might represent a trophic link between bacteria and filter-feeding bivalves. Some data support this assumption. Tintinnids were observed in the stomachs of oysters (Paulmier, 1972). Moreover, filter-feeding benthic molluscs retain protists, as exemplified by contaminations of bivalves by toxic flagellates (Sournia et al., 1991). In a mixed cell suspension of phytoplankton and dinoflagellates, six different species of bivalves were able to selectively clear and digest dinoflagellates (Shumway et al., 1985). Recently, Bardouil et al. (1996) showed that *Crassostrea gigas* easily consumes a non toxic dinoflagellate and Kreeger and Newell (1996) clearly demonstrated in mussels the ingestion and assimilation of bacterial carbon *via* heterotrophic flagellates. Le Gall *et al.* (1997) reported experimental significant retention and ingestion of cultured bacterivorous ciliates, *Uronema* sp., by the oyster *Crassostrea gigas*. Heterotrophic protists, which are abundant in coastal ecosystems, may thus constitute an alternative or complementary food resource for benthic filter feeders allowing the indirect recuperation of DOM and picoplanktonic production, otherwise not accessible to them.

We present evidence of oyster grazing on protists: a ciliate and flagellate community from a coastal oyster rearing pond was offered to oysters in a laboratory experimental system. Clearance rate, filtered particles and relative retention efficiency were determined by following the taxonomic composition and relative abundance of the protist community over time in presence or absence of actively filtering oysters.

MATERIALS AND METHODS

Oyster collection and acclimation

Oysters were collected in June 1997 from our oyster pond research facility " Marais du Plomb" (L'Houmeau, near La Rochelle, French Atlantic coast). Twenty adult *Crassostrea gigas* (1 year old, shell length 5 cm and mean dry tissue weight 1.64 ± 0.29 g) were transported to the laboratory, removed of epibionts and acclimated overnight at the ambient field temperature of 18°C, in GF/C (Whatman) filtered coastal pond water. Just before the experiment, 10 actively filtering oysters were selected and placed in 1 liter pyrex rectangular trays containing 800 ml of GF/C (Whatman) filtered coastal pond water.

Protist community: sampling and enumeration

The field planktonic community provided as potential food to the experimental oysters came from the coastal pond. Natural unfiltered oyster pond water was collected, using a sampling 2.5 l "Van Doorn" bottle (Wildco) and held in the laboratory at 18 °C in an opaque carboy until use. Ciliates and flagellates were fixed, stained and enumerated according to methods modified from Haas (1982), Caron (1983) and Sherr et al. (1994). For ciliate examination, 20 ml samples were stained live for 10 minutes by adding proflavin hemisulfate solution (Sigma, 0.033 % w/v, final concentration, 0.00066 %): preliminary comparative experiments showed that live staining had no deleterious effects on the ciliate community. Ciliates were then preserved by adding glutaraldehyde (Sigma electron microscopy grade, 25 % v/v in 0.2 µm filtered seawater, final concentration 1 %). The cells were enumerated in Utermöhl settling chambers (Hydro-Bios combined plate chambers), using a reverse epifluorescence microscope (Leitz DMIRB, 100 W mercury lamp and blue light excitation). Ciliate taxons were enumerated and identified under combined epifluorescence and interference contrast illumination (magnification: x 400 or x 630). Sizes of all cells (length and width) were measured through a calibrated ocular micrometer. Mean cell volume of each ciliate taxon was calculated by equating the shape to standard geometric configurations. The cell volume was converted into carbon units, using a theoretical carbon/volume ratio of 0.17 pg Carbon (C) µm⁻³ (Putt and Stoecker, 1989), corrected for glutaraldehyde fixative, according to Leakey et al. (1994)).

For flagellate counting, 20 ml samples were preserved with formaldehyde (paraformaldehyde powder Sigma, 8 % w/v in 0.2 µm filtered seawater, final concentration 1%); each sample was concentrated to 10 ml in a filtration tower mounted with black a 0.6 µm-pore polycarbonate membrane (Nuclepore) and a cellulosic backing filter (Whatman 1 µm) and stained by primulin (direct yellow 59 from Sigma; working solution was according to Sherr et al. (1994): 250 µg primulin in 100 ml of 0.1 M Trizma HCl at pH 4.0; 50 µg ml⁻¹ final concentration). The primulin method allows observation of cell outlines and permits distinguishing autotrophic from heterotrophic flagellates by repeated interchange of the filter sets (Caron, 1983): phototrophic cells (crimson under UV 365 nm excitation and red colored under green 450-490 nm excitation) and heterotrophic cells (blue under UV excitation and invisible under green excitation) were separately enumerated. Fields were viewed first for primulin fluorescence to locate flagellates, and then for chlorophyll *a* fluorescence (by changing the filter set) to confirm which of these cells were

pigmented. Length and width of 100 flagellates were measured (observation under UV 365 nm excitation and magnification x 630) from triplicate samples. However, the presence of the black Nuclepore filter did not allow any observation of the flagellates under light microscopy and thus prevented identification of taxon or species.

Experimental protocol for the study of protist retention

The possible influence of oyster filtration upon the natural protist community was studied during 90 minutes in an experimental chamber at 18°C by comparing the evolution of protist abundances in triplicate suspensions with or without filtering oysters. At the start of the feeding period, 6 oysters were transferred to individual 500 ml pyrex rectangular trays containing 400 ml natural unfiltered oyster pond water, gently homogenized with a magnetic rod to prevent sedimentation. As protists are fragile organisms, only a moderate homogenization was processed in order to avoid cell damage; because of this restriction, the volume of the protist suspension could not be more than 400 ml, to maintain an homogenous concentration of living protists.

Two experimental treatments were performed each in triplicate: the natural ciliate and flagellate suspensions were (1) allowed to evolve as controls, in presence of 3 living but non filtering oysters, tightly tied up by a knotted string (controls for physical sedimentation of the suspension), or (2) delivered to 3 actively filtering oysters. It is to be noticed that, at natural food concentration used in this study, there was no visible production of pseufoeces. Dry tissue weight of each oyster was recorded at the end of the experiment and clearance rates and filtered particles were expressed per g of oyster dry tissue.

Calculation of clearance rate, filtered particles and relative retention efficiency

In order to control the normality of oyster filtration in our laboratory experiments, the clearance rate was estimated and compared to literature data. Defined as the theoretical water volume entirely cleared from particles (assuming 100 % retention) per unit time and per oyster dry tissue weight ($\text{l h}^{-1} \text{g}^{-1}$) (Bayne & Widdows, 1978), the clearance rate was calculated from the evolution in ciliate or flagellate cell concentration in the triplicate suspensions with filtering oysters. During the first five minutes of the experiment, individual variations in the settling of a regular oyster filtration prevented any reliable study of the protist abundance evolution in the triplicate suspensions: therefore, we selected the subsequent sampling time (15 mn) as the most appropriate "standard" time

in our clearance experiment. Assuming exponential decline of the retained cells, the clearance rate was calculated according to Coughlan (1969) during the first 15 minutes:

$$F = \frac{\text{Ln}C_0 - \text{Ln}C_t}{t - t_0} \times V$$

where F = clearance rate (l h^{-1}), V = volume of the suspension (l), C_0 = initial concentration of the suspension (cells l^{-1}), C_t = concentration at time t (cells l^{-1}) and $(t-t_0)$ = time interval (h).

Taking into account that the weight specific filtration decreases with increasing body size, standardized clearance rates were calculated according to Riisg rd (1988): F/W^b

where F = clearance rate (l h^{-1}), W = dry tissue weight (g) and $b = 0.73$ for *Crassostrea virginica* (Riisg rd, 1988).

The filtered particles, which is the number of each protist taxon retained per unit time and per g of oyster dry tissue ($\text{cells h}^{-1} \text{g}^{-1}$), was calculated directly from the difference of cells present between t_0 and t_{15} minutes.

To investigate the possibility of a differential grazing by the oyster among the various protist taxons, we compared the relative retention efficiencies for each ciliate taxon and each ciliate and flagellate order. Defined as the number of a specific cell type retained during 15 mn, related to the initial available number of the same cell type at the beginning of the experiment, each relative retention efficiency was calculated from the difference in abundances at t_0 and t_{15} minutes, related as a percentage to the abundance at t_0 :

$$\text{Er (\%)} = 100 \times [(C_0 - C_t)/C_0]$$

where Er = relative retention efficiency, C_0 = initial particle concentration (cells l^{-1}) at t_0

and C_t = particle concentration (cells l^{-1}) at 15 minutes.

Initial ciliate and flagellate abundances from the triplicate experiments with filtering or closed oyster were compared using a t-Student test (data were previously tested for normality by Kolmogorov-Smirnov test). Evolution during the 90 minutes experiment of ciliate and flagellate abundances in triplicate controls was followed by comparing the 5 time points sampled (0, 5, 15, 45 and 90 minutes) with a regression test.

RESULTS

Taxonomic composition and standing stocks of protists in the coastal oyster pond in June 1997

In the summer period of the experiment, the ciliate community of the coastal pond was abundant ($23\,700 \pm 3\,600$ cells l^{-1}) and dominated by members of the subclass Choreotrichia, mainly represented by the order Choreotrichida, with *Tintinnopsis* spp. ($10\,000$ to $11\,200$ cells l^{-1}) and by the order Oligotrichida, dominated by *Strombidium* spp. ($5\,700$ to $8\,500$ cells l^{-1}). Other common taxa from the subclass Haptoria and order Haptorida (*Mesodinium* sp., *Askenasia* sp.) were also representative of the assemblage ($3\,400$ to $5\,700$ cells l^{-1}). Ciliate sizes ranged from $8\ \mu\text{m}$ length for a *Mesodinium* sp., to $72\ \mu\text{m}$ for *Strombidium conicum* (table 1). Prevalant ciliate cell lengths were between 16 and $48\ \mu\text{m}$.

Table 1 near here

Flagellate abundances in the coastal pond varied from 4.2 to 6.7×10^6 cells l^{-1} and flagellates accounted for about 99.5% of the protists enumerated in water samples. Mean flagellate sizes ranged from $4\ \mu\text{m}$ for heterotrophic to $6\ \mu\text{m}$ for autotrophic flagellates.

Tintinnina biovolumes as well as cell carbon were much higher than Oligotrichida and Haptorida ones (for the most abundant taxon in each order, $19\,181\ \mu\text{m}^3$ for *Tintinnopsis* sp. ($48\ \mu\text{m}$ - $24\ \mu\text{m}$), $5\,579\ \mu\text{m}^3$ for *Strombidium* sp. ($32\ \mu\text{m}$ - $24\ \mu\text{m}$) and $2\,145\ \mu\text{m}^3$ for *Mesodinium* sp. ($16\ \mu\text{m}$ - $16\ \mu\text{m}$)). By multiplying the taxon abundances at the beginning of experiment by the individual carbon content of each ciliate taxon, we estimated the quantity of ciliate carbon available for oysters: on average, $63.5\ \mu\text{g C } l^{-1}$. In this study, the flagellate carbon was not evaluated because flagellate taxonomy and biovolumes could not be determined.

Grazing experiments

The initial concentration in the natural suspension sampled for the grazing experiment was $25\,000 \pm 3\,900$ ciliates l^{-1} and $4.5 \times 10^6 \pm 1.12 \times 10^6$ flagellates l^{-1} . Since all suspensions originated from the same coastal pond sample, initial protist abundances in the experimental trays showed no significant difference between controls and oyster treatments (t Student test, $n = 6$, $p \gg 0.05$). In the 3 control suspensions, ciliate and flagellate

abundances remained relatively constant over 90 minutes (figure 1) with regression test ($r^2 = 0.17$, $p \gg 0.05$ for ciliates and $r^2 = 0.23$, $p \gg 0.05$ for flagellates). **Figure 1 here**

In the 3 experimental trays with filtering oysters, ciliates whose size was between 20 and 40 μm were 100% retained; the relative retention efficiency in the experimental suspension within 15 minutes was 96 % for Haptorida and Tintinnina (figure 2-a and -b)) and 91 % for Oligotrichida, (figure 2 c). Similarly, flagellates decreased by 86 % within 15 minutes in the trays with the filtering oyster (figure 2-d). At the end of the experiment (90 minutes), virtually all ciliates and 96 % of the flagellates had been retained by the bivalves. **Figure 2 here**

The relative retention efficiency for each protist taxon, related to the protist sizes present in the suspension remained constant in the studied size range (figure 3), unless a slight decrease was noticed for the smaller and larger taxons: only 84 % of the 4 μm particles and 88 % of the 72 μm particles were retained. For concentrations below the pseudofeces threshold, all protist from 4 to 72 μm were similarly retained by the oyster gills. **Fig 3 here**

Clearance rates, filtered particles

Table 2 near here

Clearance rates of oysters averaged $4.0 \pm 1.3 \text{ l h}^{-1} \text{ g}^{-1}$ for flagellates and $7.2 \pm 3.5 \text{ l h}^{-1} \text{ g}^{-1}$ for Oligotrichida ciliates (table 2). The filtered particles, calculated between 0 and 15 minutes (table 3) were dependent on protist taxon. Tintinnina were more retained ($27\,000 \pm 11\,500 \text{ cells h}^{-1} \text{ g}^{-1}$) than Haptorida ($8\,800 \pm 4\,400 \text{ cells h}^{-1} \text{ g}^{-1}$) or Oligotrichida ($19\,600 \pm 10\,200 \text{ cells h}^{-1} \text{ g}^{-1}$). By multiplying filtered particles ($\text{cells h}^{-1} \text{ g}^{-1}$) by the individual carbon content of each taxon, we obtained the quantity of ciliate carbon retained per hour per gram oyster dry weight ($\mu\text{g C h}^{-1} \text{ g}^{-1}$), which averaged $126 \mu\text{g C h}^{-1} \text{ g}^{-1}$ (table 3). **Table 3 near here**

DISCUSSION

Marine planktonic protists (ciliates and flagellates) have recently been shown to be abundant in Atlantic coastal ponds: our estimations of protist abundances in our coastal pond at the time of the grazing experiment were respectively $23\,700 \pm 3\,600 \text{ ciliate l}^{-1}$ and $4.5 \times 10^6 \pm 1.12 \times 10^6 \text{ flagellates l}^{-1}$. These protist abundances fell within the

range estimated in the same pond by Robin (pers.com.), 10 000 to 30 000 cells l⁻¹ for ciliates and 53 x 10⁴ to 2.2 x 10⁶ flagellates l⁻¹.

In absence of published report on ciliate abundances in the Atlantic coastal ecosystem near the coastal pond, we compared our data to results from distant estuaries and bays. In other temperate estuaries, ciliate abundances were in the same range, from 200 to 19 000 cells l⁻¹ (Saint-Laurent estuary, Sime Ngando et al., 1995) and from 220 cells l⁻¹ to 56 000 cells l⁻¹ (Northern Adriatic, River Po estuary, Revelante & Gilmartin, 1983). However, in the Gulf of Maine, ciliate abundances were higher: 350 000 to 6 000 000 cells l⁻¹ (Montagnes et al., 1988).

In our study, the ciliate community was dominated by the order Choreotrichida with *Tintinnopsis* spp. (10 000 to 11 200 cells l⁻¹) and by the order Oligotrichida with *Strombidium* spp. (5 700 to 8 500 cells l⁻¹). Robin (pers. com.) observed up to 300 000 Tintinnina l⁻¹ in June 1996 in the same coastal pond of L'Houmeau. Tintinnina are also abundant in the Mediterranean Sea: 10 000 ciliate l⁻¹ in Villefranche-sur-mer (Rassoulzadegan & Gostan, 1976) and 8 000 cells l⁻¹ in the south-eastern mediterranean Alger Bay (Vitiello, 1964). On the opposite, in a Northern Mediterranean coastal lagoon (Etang de Thau), Tintinnina abundance was only 75 cells l⁻¹ (Lam-hoai et al., 1997), a value much lower than our. Oligotrichida abundances (5 700 to 8 500 cells l⁻¹) were in the range of values collected by Robin (pers. com.) during the spring of 1996 (4 300 to 11 500 cells l⁻¹) but lower than abundances (90 000 cells l⁻¹) during the summer in Mediterranean Sea (Rassoulzadegan, 1977).

Our values for flagellate abundances were close to those obtained in the Saint-Laurent estuary: 1.9 x 10⁶ to 6 x 10⁶ cells l⁻¹ (Lovejoy et al., 1993) or in the marine shallow-water Limfjorden in Denmark: 2 x 10⁶ cells l⁻¹ (Andersen & Sorensen, 1986).

The wide range of these data shows the natural variability of protist abundances in the field. Moreover, since the coastal ponds are periodically closed systems in which the plankton community undergoes rapid fluctuations, it remains difficult to establish valid criteria for comparisons with open coastal systems. Nevertheless, in terms of potential carbon resources available to the oysters, the amounts calculated for ciliates (63.5 µg C l⁻¹) were at the

high level for protozoan carbon levels in coastal waters (Saint-Laurent Estuary: 0.23 to 51.6 $\mu\text{g C l}^{-1}$, Sime Ngando et al, 1995).

When a coastal pond planktonic community was provided as potential food, clearance rates of oysters for protists ($4.0 \pm 1.3 \text{ l h}^{-1} \text{ g}^{-1}$ for flagellates and $7.2 \pm 3.5 \text{ l h}^{-1} \text{ g}^{-1}$ for Oligotrichida ciliates) were in a range similar to values measured for phytoplankton by Gerdes (1983): $4.8 \text{ l h}^{-1} \text{ g}^{-1}$, Deslous-Paoli et al. (1987): $4.7 \text{ l h}^{-1} \text{ g}^{-1}$, Riisg rd (1988): $6.8 \text{ l h}^{-1} \text{ g}^{-1}$ and Soletchnik et al. (1991): 3 to 4 $\text{ l h}^{-1} \text{ g}^{-1}$. However, in our experimental closed system, the concentration of particles is rapidly declining during the experiment (figure 2); the standard time for our clearance experiment (15 mn), selected to avoid drawbacks related to the irregular settling of oyster filtration during the first five minutes, is tardily to allow accurate evaluation of clearance rates. Nevertheless, the possible negative effects of our suboptimal laboratory experimental conditions on the bivalve filter pump efficiency (Jorgensen, 1996) may have only resulted in the underestimation of our experimental values: field clearance rates of oysters for protists might be even higher.

The relative retention efficiency was 94 % for the ciliates and 86 % for the flagellates within 15 minutes from 400 ml suspensions. This finding supports the results of Le Gall *et al.* (1997), who demonstrated that the oyster *Crassostrea gigas* retained *Uronema* sp., a cultured ciliate isolated from the oyster pond, with a 85 % relative retention efficiency when present at a concentration close to field ciliate abundances. It also corroborates the observations by Paulmier (1972), who reported Tintinnids to be abundant in the stomachs of wild oysters from the Atlantic coast. Likewise, Kreeger & Newell (1996) estimated that 58 % and 44 % respectively of the cultured heterotrophic nanoflagellates were retained by *Geukensia demissa* and *Mytilus edulis*, compared to values of 66 % and 77 %, respectively, for the autotroph *Isochrysis galbana*. Ciliates and flagellates thus represent a potentially valuable food source and might be a significant component in the natural diet of suspension-feeding bivalves, given that their relative abundance is sufficiently high in the available seston.

To investigate the possible influence of particle size on oyster retention, we separately followed the evolution of each protist taxon abundance in the experimental suspensions. In our experiments, ciliates and flagellates in a size range from 4 to 72 μm were retained by the oyster, but the smallest heterotrophic flagellates (4 μm) and the largest ciliates (*Strombidium conicum*, 72 μm x 32 μm) displayed a slightly lower relative retention efficiency than the

ciliates, whose size was between 20 and 40 μm . Indeed, the flagellate sizes in our suspensions were at the lower end of the particle size spectrum, known to be retained by *Crassostrea gigas*. Barillé et al. (1993) showed that this oyster has a limited capacity to retain small particles: 4 μm particles (equivalent spherical diameter, ESD) were retained with 100% retention efficiency when sestonic load was low, but the limit increased to 12 μm for higher sestonic loads; for particles below these thresholds, retention efficiency quickly decreased. Similarly Deslous-Paoli et al. (1987) demonstrated that the oyster is not able to retain small particles. Bougrier et al. (1997) reported that the selection of algae by the oyster, *Crassostrea gigas* was independent of the size, volume or carbon content of each species (size between 3.65 and 9 μm ESD). They observed, nevertheless, that some algae were preferentially filtered or rejected, due to cell shape and flexibility.

Conversely, mussels are able to retain even picoplankton-size particles (Kemp et al., 1990; Kreeger & Newell, 1996): in particular, in high-quality food suspensions (expressed as percent of particles with chlorophyll fluorescence) prey retention and selection in *Mytilus edulis* is not dependent on the prey size (Newell et al., 1989). Reduction in food quality induced a drop in the ability to select living cells from silt particles, independently of size. These investigations demonstrated however, as for oysters, a selectivity based on cell shape (Newell et al., 1989). Since unlike the mussel, *Crassostrea gigas* cannot retain picoplankton-size particles at natural concentrations, the picoplankton-protzoa trophic pathway (Le Gall et al., 1997) may represent a significant energy source for the oyster (figure 4).

Ciliates are more nutritious prey than phytoplankton cells. They are relatively rich in nitrogen (C:N ratio near 4, Putt & Stoecker, 1989; Ohman & Snyder, 1991, as compared to > 5 for phytoplankton, Heinbokel et al., 1978; Burkhardt et al., 1997), and contain more carbon per cell than phytoplankton : our estimations of cell carbon contents, which are comparable to values previously reported in literature (3 100 pg cell^{-1} for *Strombidium* sp. (43 μm - 42 μm), Stoecker & Egloff, 1987; 1 100 pg C cell^{-1} for *Strombidium* sp. (43 μm - 30 μm), Jonsson & Tiselius, 1990) were much higher than phytoplankton carbon content per cell from 10 to 21 pg C cell^{-1} for *Skeletonema costatum* (Strathmann, 1967; Burkhardt et al., 1997; Bougrier et al., 1997), 1.61 pg C cell^{-1} for *Phaedactylum tricoratum* (Fiala-Medioni et al., 1983) and 10.3 pg C cell^{-1} for *Navicula filata* (Bougrier et al., 1997). In our experiment, on average, oysters retained 126 $\mu\text{g ciliate C h}^{-1} \text{g}^{-1}$ for a ciliate concentration of $25\,000 \pm 3\,900 \text{ cells l}^{-1}$.

¹. Fiala-Mediono et al. (1983) estimated that oyster filtering *Phaedactylum tricornatum* retained $27.5 \mu\text{g C h}^{-1} \text{g}^{-1}$ for a phytoplankton concentration of 1×10^6 cells l^{-1} . Ciliates may thus contribute to the carbon requirements of *Crassostrea gigas* in the same way as do heterotrophic flagellates for the mussels *Geukensia demissa* and *Mytilus edulis* (Kreeger & Newell, 1996).

Most studies that have examined the nutritional importance of protist as a "trophic link" have focused on pelagic consumers, such as zooplankton (Berk et al., 1977; Porter et al., 1979; Sherr et al., 1986 b, Jonsson and Tiselius, 1990; Gifford & Dagg, 1991; Hartmann et al., 1993). However, only few studies have done the same for benthic consumers (Kreeger & Newell, 1996 ; Le Gall et al, 1997). Trophic coupling between pelagic protists and benthic suspension-feeders is poorly documented in aquatic food models (e.g. see Legendre & Le Fèvre, 1995).

In open water oyster beds, primary producers, in particular phytoplankton and resuspended microphytobenthos can be considered important food sources for bivalves suspension feeders (Blanchard et al., 1997). Otherwise in coastal ponds, the resuspension of microphytobenthos is low, due a lack of turbulence: even if it may attain up to 25 times higher levels than phytoplankton (Zanette, 1980; Robert, 1983), the microphytobenthos is unlikely to be an important direct resource. However, the DOM released by these autotrophs contributes to the important bacterial biomass that develops in coastal ponds: bacterioplankton constitutes 50 % of the planktonic C biomass in oyster ponds of the Charente (Frikha et al., 1987). The bacteria, in turn, will be primary food for heterotrophic/mixotrophic ciliates and flagellates which develop biomasses comparable to phytoplankton: in our coastal pond, the protist biomass was similar to the phytoplankton biomass of coastal oyster ponds from Bourgneuf Bay (Robert, 1983). Since bacterivorous ciliates have a gross growth efficiency of about 40 % (Johnson et al., 1982; Ohman & Snyder, 1991), relatively large amounts of bacterial C must be recovered by oysters *via* the protist trophic link.

In coastal pond habitats, bivalve molluscs are abundant and may be the dominant consumers of seston. Oysters are most likely opportunist omnivores, balancing their C (and N) requirements by utilizing a wide variety of living and dead material (Riera & Richard, 1996) including protists. In addition to phytoplankton which cannot entirely account for the energy requirements of *Crassostrea gigas* (Héral, 1987), oysters can derive nutrition from microzooplankton, in particular from protists. Our experiment presents the first data about oyster nutrition on a field community of protists. These results clearly show that suspension-feeding bivalves feed on ciliates and

flagellates. Such a trophic relationship could be of primary importance for the transfer of C, and probably N, from the microbial food web to higher trophic levels in the benthos.

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Order	Suborder	Family	Taxon	Species	Length (µm)	Width (µm)	Biovolume (µm ³)	Carbon per cell (pg C cell ⁻¹)			
Choreotrichida	Tintinnina	Codonellidae	<i>Tintinnopsis</i>	sp.	35	24	13994	2379			
			<i>Tintinnopsis</i>	sp.	40	24	15984	2717			
			<i>Tintinnopsis</i>	sp.	48	24	19181	3261			
			<i>Tintinnopsis</i>	sp.	48	40	53281	9058			
			<i>Tintinnopsis</i>	sp.*	51	40	56632	9627			
			<i>Tintinnopsis</i>	sp.	56	24	22378	3804			
		Oligotrichida		Codonellopsidae	<i>Stenosemella</i>	sp.*	24	22	7603	1293	
					Halteriidae	<i>Halteria</i>	sp.	27	19	3024	514
				Strombidiidae		<i>Strombidium</i>	sp. *	24	19	2617	445
						<i>Strombidium</i>	sp.	25	22,5	4307	732
						<i>Strombidium</i>	sp.	32	24	5579	948
<i>Strombidium</i>	sp.					35	25,6	6863	1167		
<i>Strombidium</i>	sp.				40	28,5	9569	1627			
<i>Strombidium</i>	<i>conicum</i>	72	32	20642	3509						
Haptorida		Didiniidae	<i>Didinium</i>	sp.*	64	54	97716	16612			
			Mesodiniidae	non identified		16	16	2145	365		
		non identified			27	14	2771	471			
		<i>Mesodinium</i>		sp.*	8	8	268	45			
		<i>Mesodinium</i>		sp.	16	16	2145	365			
		<i>Mesodinium</i>		<i>pulex</i>	14	10	733	125			
		<i>Askenasia</i>		sp.	24	16	3217	547			
		Autotrophic flagellate					6,2	4,2			
		Heterotrophic flagellate					4,1	3,5			

Table 1: Taxonomic composition, sizes, biovolumes and carbon content per cell of the protist community in the coastal pond in June 1997. Taxa printed in bold type were abundant and represented in all samples. Taxa identified by an * were rare and/or not present in all samples. When species unidentified, taxa were typified by their size.

Taxon (length-width in μm)	Cell abundances (cells l^{-1}) at To in experimental suspension with an actively filtering oyster		Standardized clearance rate ($\text{l h}^{-1}\text{g}^{-1}$)	
	Mean	SD	Mean	SD
Haptorida				
<i>Mesodinium sp.</i> (16-16)	1178	237	3.6	0.7
<i>Mesodiniidae</i> (16-16)	1748	776	7.9	4.3
<i>Mesodinium pulex</i> (14-10)	2736	3178	4.1	4.5
<i>Askenasia</i> (24-16)	76	132	2.5	4.4
Haptorida average	5738	4192	5.5	2.3
Oligotrichida				
<i>Strombidium sp.</i> (25-22,5)	1026	1777	3.7	6.5
<i>Strombidium sp.</i> (32-24)	3648	3288	4.9	6.4
<i>Strombidium sp.</i> (35-25,6)	2318	4015	4.1	7.1
<i>Strombidium sp.</i> (40-28,5)	114	197	2.7	4.7
<i>Strombidium conicum</i> (72-32)	1064	628	4.9	3.5
<i>Halteria sp.</i> (27-19)	76	132	0.0	0.0
Oligotrichida average	8284	2028	7.2	3.5
Tintinnina				
<i>Tintinnopsis sp.</i> (35-24)	152	174	4.9	4.3
<i>Tintinnopsis sp.</i> (40-24)	1102	1425	8.7	2.2
<i>Tintinnopsis sp.</i> (48-24)	9082	3933	6.5	5.8
<i>Tintinnopsis sp.</i> (48-40)	760	1316	3.6	6.2
<i>Tintinnopsis sp.</i> (56-24)	114	114	4.7	4.1
Tintinnina average	11210	2146	7.8	1.5
Flagellates				
Autotrophic flagellate	1.38E+06	1.07E+06	4.9	3.0
Heterotrophic flagellate	3.55E+06	5.00E+04	3.1	1.8
Flagellate average	4.93E+06	2.47E+06	4.0	1.3

Table 2: Cell abundances in experimental suspensions (cells l^{-1} at To) and standardized clearance rates by *Crassostrea gigas* ($\text{l h}^{-1}\text{g}^{-1}$) for the different ciliate and flagellate taxa (mean \pm SD, n=3). When species unidentified, taxa were typified by their size by their size (length and width in μm).

Taxon (length-width in μm)	Filtered particles (cells $\text{h}^{-1}\text{g}^{-1}$)		Carbon per cell (pg C cell^{-1})	Filtered particles ($\text{ng C h}^{-1}\text{g}^{-1}$)	
	Mean	SD		Mean	SD
Haptorida					
<i>Mesodinium sp.</i> (16-16)	2707	823	365	988	301
<i>Mesodiniidae</i> (16-16)	4441	2933	365	1621	1070
<i>Mesodinium pulex</i> (14-10)	1488	1394	125	186	174
<i>Askenasia</i> (24-16)	227	393	547	124	215
Haptorida sum	8863	4390		2919	1583
Oligotrichida					
<i>Strombidium sp.</i> (25-22,5)	3063	5305	732	2242	3883
<i>Strombidium sp.</i> (32-24)	7811	7739	948	7405	7337
<i>Strombidium sp.</i> (35-25,6)	6919	11985	1167	8075	13986
<i>Strombidium sp.</i> (40-28,5)	276	479	1627	450	779
<i>Strombidium conicum</i> (72-32)	2154	1126	3509	7560	3953
<i>Halteria sp.</i> (27-19)	158	274	514	81	141
Oligotrichida sum	19643	10285		25812	8643
Tintinnina					
<i>Tintinnopsis sp.</i> (35-24)	432	528	2379	1029	1256
<i>Tintinnopsis sp.</i> (40-24)	2382	2899	2717	6472	7876
<i>Tintinnopsis sp.</i> (48-24)	22770	14798	3261	74254	48257
<i>Tintinnopsis sp.</i> (48-40)	1583	2742	9058	14342	24841
<i>Tintinnopsis sp.</i> (56-24)	318	342	3804	1210	1303
Tintinnina sum	27487	11584		97307	32081
Mean sum for all ciliates	55993			126038	

Table 3: Retention of various ciliate taxa by *Crassostrea gigas* expressed as filtered particles per unit time and unit oyster dry weight (cell $\text{h}^{-1}\text{g}^{-1}$ or $\text{ng C h}^{-1}\text{g}^{-1}$, mean \pm SD, n = 3). When species unidentified, taxa were typified by their size (length and width in μm).

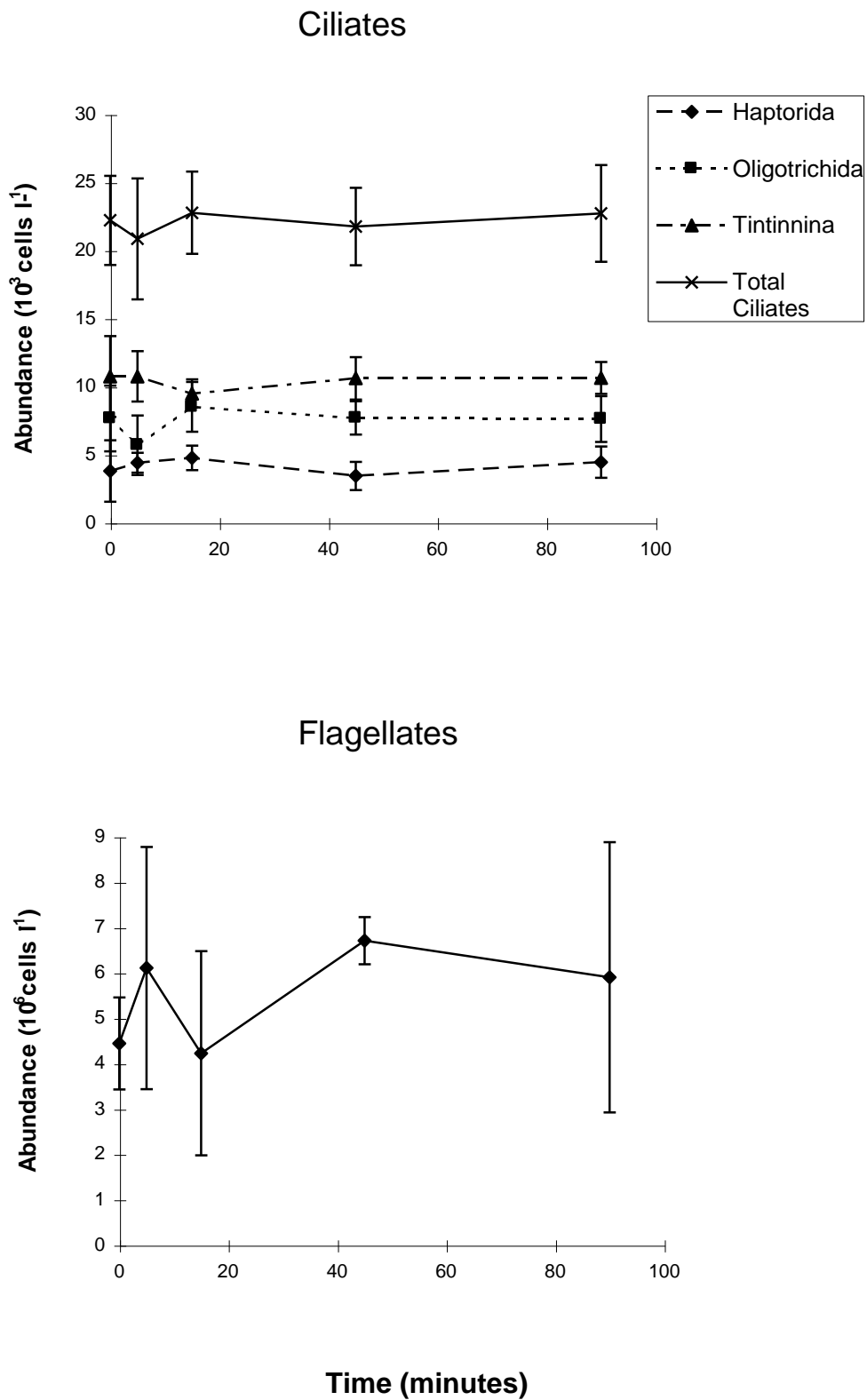


Figure 1: Evolution of ciliate and flagellate abundances in control suspensions. Abundance data (mean \pm SD, n = 3) were collected from 3 separate experiments, performed with a closed non-filtering oyster in 400 ml suspension of coastal pond water.

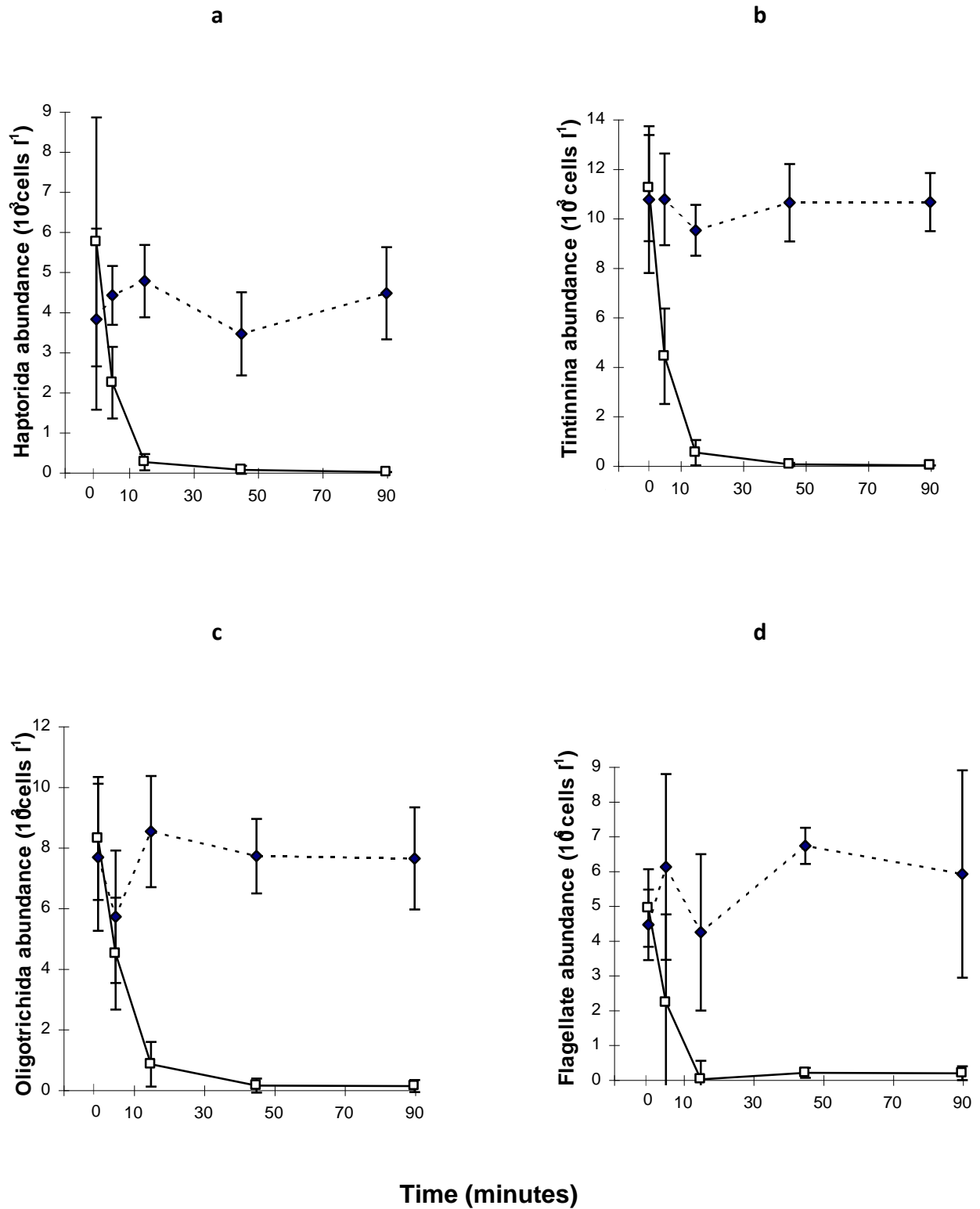


Figure 2: Retention of various protist taxa by the oyster *Crassostrea gigas* (Haptorida (a), Tintinnina (b) and Oligotrichida (c) and flagellates (d). Protist abundance data (mean \pm SD, n = 3) were collected from 6 separate experiments performed in 400 ml marine pond water suspensions with a closed non-filtering oyster (♦ and dashed lines) or with a filtering oyster (□ and solid lines).

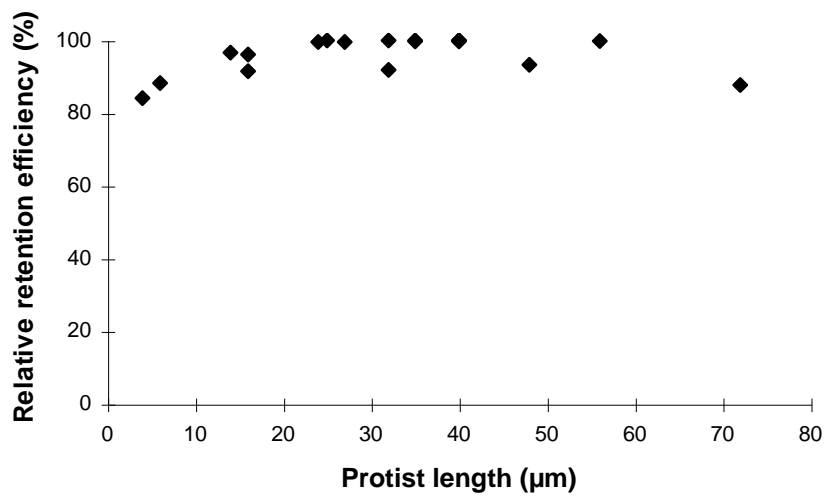


Figure 3: Relative retention efficiencies of protists related to their size class.

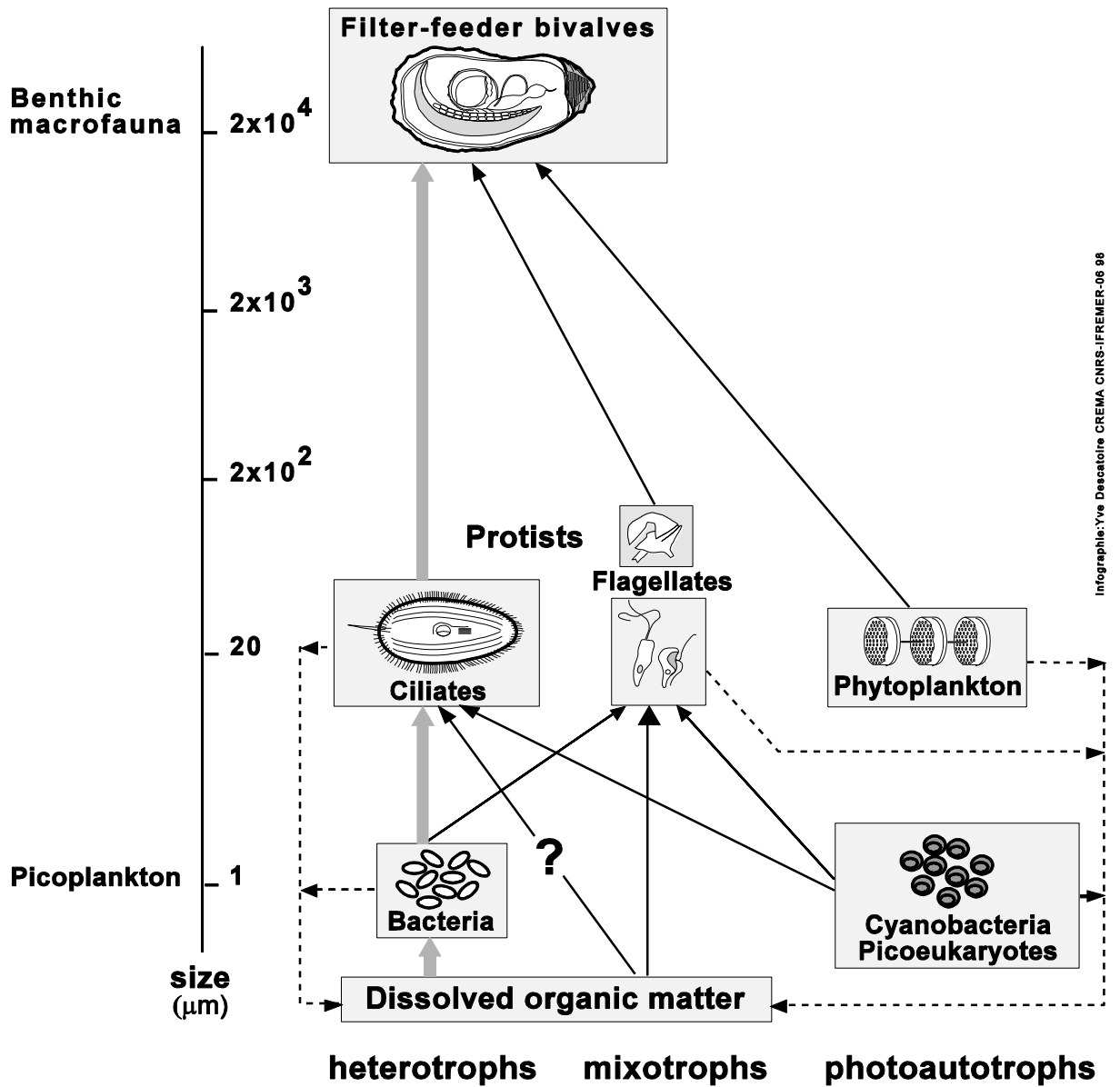


Figure 4: Hypothetical microbial food web in an oyster growing area

(modified from Le Gall et al., 1997)