



**HAL**  
open science

## How do species, population and active ingredient influence insecticide susceptibility in [i]Culicoides[/i] biting midges (Diptera: Ceratopogonidae) of veterinary importance?

Roger Venail, Jonathan Lhoir, Moussa Fall, Ricardo del Rio, Sandra Talavera, Karien Labuschagne, Miguel Miranda, Nonito Pagès, Gert J. Venter, Ignace Rakotoarivony, et al.

### ► To cite this version:

Roger Venail, Jonathan Lhoir, Moussa Fall, Ricardo del Rio, Sandra Talavera, et al.. How do species, population and active ingredient influence insecticide susceptibility in [i]Culicoides[/i] biting midges (Diptera: Ceratopogonidae) of veterinary importance?. *Parasites & Vectors*, 2015, 8, 9 p. 10.1186/s13071-015-1042-8 . hal-01231832

**HAL Id: hal-01231832**

**<https://hal.science/hal-01231832v1>**

Submitted on 20 Nov 2015

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

RESEARCH

Open Access



# How do species, population and active ingredient influence insecticide susceptibility in *Culicoides* biting midges (Diptera: Ceratopogonidae) of veterinary importance?

Roger Venail<sup>1\*</sup>, Jonathan Lhoir<sup>2</sup>, Moussa Fall<sup>3</sup>, Ricardo del Río<sup>4</sup>, Sandra Talavera<sup>5</sup>, Karien Labuschagne<sup>6</sup>, Miguel Miranda<sup>4</sup>, Nonito Pagès<sup>5</sup>, Gert Venter<sup>6</sup>, Ignace Rakotoarivony<sup>2</sup>, Xavier Allène<sup>2</sup>, Bethsabée Scheid<sup>1</sup>, Laëtitia Gardès<sup>2</sup>, Geoffrey Gimonneau<sup>2,3</sup>, Renaud Lancelot<sup>2</sup>, Claire Garros<sup>2</sup>, Catherine Cêtre-Sossah<sup>2</sup>, Thomas Balenghien<sup>2</sup>, Simon Carpenter<sup>7</sup> and Thierry Baldet<sup>2</sup>

## Abstract

**Background:** *Culicoides* biting midges are biological vectors of internationally important arboviruses of livestock and equines. Insecticides are often employed against *Culicoides* as a part of vector control measures, but systematic assessments of their efficacy have rarely been attempted. The objective of the present study is to determine baseline susceptibility of multiple *Culicoides* vector species and populations in Europe and Africa to the most commonly used insecticide active ingredients. Six active ingredients are tested: three that are based on synthetic pyrethroids (alpha-cypermethrin, deltamethrin and permethrin) and three on organophosphates (phoxim, diazinon and chlorpyrifos-methyl).

**Methods:** Susceptibility tests were conducted on 29,064 field-collected individuals of *Culicoides obsoletus* Meigen, *Culicoides imicola* Kieffer and a laboratory-reared *Culicoides nubeculosus* Meigen strain using a modified World Health Organization assay. Populations of *Culicoides* were tested from seven locations in four different countries (France, Spain, Senegal and South Africa) and at least four concentrations of laboratory grade active ingredients were assessed for each population.

**Results:** The study revealed that insecticide susceptibility varied at both a species and population level, but that broad conclusions could be drawn regarding the efficacy of active ingredients. Synthetic pyrethroid insecticides were found to inflict greater mortality than organophosphate active ingredients and the colony strain of *C. nubeculosus* was significantly more susceptible than field populations. Among the synthetic pyrethroids, deltamethrin was found to be the most toxic active ingredient for all species and populations.

**Conclusions:** The data presented represent the first parallel and systematic assessment of *Culicoides* insecticide susceptibility across several countries. As such, they are an important baseline reference to monitor the susceptibility status of *Culicoides* to current insecticides and also to assess the toxicity of new active ingredients with practical implications for vector control strategies.

**Keywords:** *Culicoides imicola*, *Culicoides obsoletus*, *Culicoides nubeculosus*, Insecticide susceptibility, Pyrethroids, Organophosphates, Vector control

\* Correspondence: rvenail@yahoo.com

<sup>1</sup>EID Méditerranée, 34184 Montpellier, France

Full list of author information is available at the end of the article

## Background

*Culicoides* Latreille (Diptera: Ceratopogonidae) are small haematophagous insects implicated worldwide as primary biological vectors of arboviruses causing important diseases of livestock [1]. These arboviruses include blue-tongue (BTV), African horse sickness (AHSV), epizootic haemorrhagic disease (EHDV) and Schmallenberg (SBV) viruses [2]. *Culicoides*-borne arboviruses have a severe economic impact through direct losses due to the morbidity and mortality that occurs in susceptible animals. Additional losses also occur, however, due to the imposition of animal movement restrictions to limit BTV spread that inhibit animal trade [3, 4] and the indirect costs of monitoring and surveillance measures during outbreaks. *Culicoides* are also notorious as a biting nuisance in some regions, causing discomfort in humans and livestock and seasonal recurrent allergic dermatitis in horses [5–7].

In attempts to control *Culicoides*-borne arboviruses such as BTV and AHSV outside of their endemic range, compulsory vaccination campaigns and livestock movement restrictions are usually employed as the most effective way of controlling outbreaks [8]. Where safe and effective vaccines to *Culicoides*-borne viruses are either not initially available or economically unviable, control measures against *Culicoides* have been recommended by veterinary authorities to reduce host-vector contact and thus mitigate against arbovirus transmission. The use of insecticide residual spraying within stables and during transport when livestock is moved outside a restricted movement zone has been recommended in protecting animals with high economic value (e.g. prize rams and racehorses). Additional physical measures have also been suggested to reduce *Culicoides* populations such as the mechanical removal and/or reduction of larval breeding sites on farms and housing livestock during periods of high *Culicoides* activity [9].

To date, no insecticidal products have been authorized specifically against *Culicoides* in the European Union (EU), although a wide range of products are available, licensed and in use against other arthropods of veterinary importance [9]. Worldwide, the most commonly used method to protect livestock from *Culicoides* is the application of insecticides to livestock at risk of infection. Synthetic pyrethroid (SP) active ingredients are most often used in this role, but certain organophosphate (OP) products are also still available and licensed for use in Europe [7]. Insecticidal pour-on products exert their effect in two ways: primarily they are highly toxic to insects landing on the treated animal, often killing them within minutes of their landing on the host; secondarily they exert a contact irritation that may reduce the probability of the insect successfully initiating or completing a blood meal from the host. While some effort has been

made to assess the efficiency of pour-on products against *Culicoides*, results vary greatly between studies according to different experimental designs [7]. Methodologies used include the exposure of *Culicoides* to hair clippings from treated animals [10–12] or direct exposure to a treated animal [13]. The variability of results across studies highlights the importance of using a standardized method to obtain comparable and reliable data.

Following the European Food Safety Authority's recommendation to assess susceptibility of *Culicoides* to insecticides using standardized procedures [9], a World Health Organisation (WHO) standardized technique in adult mosquitoes has been adapted for use with *Culicoides* [13, 14]. This baseline information is essential for recommending the most effective insecticides and in detecting and monitoring the development of resistance. Insecticide resistance to earlier classes of insecticides including organochlorine-based larval treatments such as dieldrin and lindane was documented in *Culicoides* in the late 1950's [15], but has not been examined for either OP or SP use. This risk exists considering that products based on single classes of insecticide have been used on a wide scale on livestock to control other arthropods including ticks, horn flies and stable flies in addition to often being used on crops.

Standardised information concerning the susceptibility of *Culicoides* to insecticides is at present limited to small scale studies [13, 14]. This study aims to assess the susceptibility of multiple populations of *Culicoides* species in different countries to the most frequently used insecticide active ingredients in Europe (SP: alpha-cypermethrin, deltamethrin and permethrin; OP: diazinon/dimpylate and phoxim) and Africa/Latin America (OP: chlorpyrifos-methyl). The main objective of the study is to generate reference baseline data regarding the efficiency of insecticidal products in killing *Culicoides* under laboratory conditions. Implementation of such insecticidal treatments into control programmes against *Culicoides*-borne diseases is then discussed.

## Methods

### *Culicoides* collection and identification

Susceptibility tests were performed on three *Culicoides* species. Laboratory-reared *Culicoides nubeculosus* Meigen were provided from a colony maintained in an insectary (temperature: 24 °C ± 1 °C; relative humidity: 70 ± 10 %; light:dark: 12:12) at Cirad (Montpellier, France). This colony was established in Cirad during 2012 from eggs and larvae provided by The Pirbright Institute (UK). Field populations of *Culicoides obsoletus* Meigen and *Culicoides imicola* Kieffer were collected from multiple locations in two European countries (France, Spain) and two African countries (Senegal, South Africa) (Additional file 1: Table S1). Collection sites were privately owned

farms characterized by abundant populations of *Culicoides* target species and reduced use of insecticides on the animals or pesticides on crops.

*Culicoides* were collected using a modified suction UV-light trap (OVI model, South Africa) [16] with the collection beaker replaced by a fine mesh netted cage to enable live collections. To prevent desiccation of *Culicoides* during the collection period, wet paper was placed on aluminium foil and rolled around the mesh cages. Traps were set before sunset and retrieved at dawn. *Culicoides* collection cages were stored in an isothermal container with an ice pack during transport to the insecticide trials room. Following completion of insecticide trials, field-collected individuals were morphologically identified to species level for *C. imicola* or to *Obsoletus* complex [17] using a binocular microscope and *Obsoletus* complex individuals were further identified to species level using a diagnostic multiplex polymerase chain reaction (PCR) assay [18].

#### **Selection of insecticides and production of impregnated papers**

Insecticide active ingredients were selected from those used most frequently in pour-on formulations within Europe. All active ingredients were used at > 98 % purity (Pestanal®, a registered trademark Sigma-Aldrich Laborchemikalien GmbH, London, UK). Test papers (Whatman n°1 filter paper, 90 g/m<sup>2</sup>, 12 × 15 cm) were impregnated following training provided by a WHO collaborative centre (LIN-IRD, France). Insecticide active ingredients were applied at different concentrations (Additional file 2: Table S2) to papers in a silicone oil as the carrier agent (2 ml per paper, 67 % acetone, and 33 % silicone oil). Control papers were impregnated with 2 ml of acetone-silicone oil mix only. Impregnations were conducted by the same person (RV) to ensure consistency. Papers were impregnated a few days before the testing period, wrapped in aluminium foil and then stored at 4 °C. Impregnated papers were sent to each country in a polystyrene cooler box with ice cooler packs for maintaining the temperature at 4 °C during transport. Each paper was used three times in assays and stored at 4 °C between trials.

#### **Insecticide susceptibility tests**

Insecticide susceptibility tests were performed following the standardized WHO protocol for adult mosquito bioassay using test tubes (WHO/VBC/81.806) [19]) adapted for *Culicoides* [13]. Because insecticide susceptibility could be age specific [20], and physiological status and sex dependant [21, 22], bioassays were performed with 2–3 day old laboratory-reared *C. nubeculosus* unfed females. Due to the difficulties of colonizing *C. obsoletus* and *C. imicola*, adults were collected from the field and used one day after collection in bioassays. For field-collected

*Culicoides*, only unpigmented females that were believed to have not previously taken a blood meal were used in data analysis, as determined through observation of abdominal pigmentation [23].

*Culicoides* were exposed for 1 h to either insecticide-impregnated papers or a control paper with the carrier compound only. For each replicate carried out on field-collected *Culicoides*, approximately 100 unsorted individuals were placed in each tube to obtain at least 25 unfed females of the target population. Following this exposure period, all *Culicoides* (including incapacitated individuals) were transferred using a motorised aspirator from exposure to observation tubes. Observation tubes were then stored vertically for 24 h and *Culicoides* within tubes were given access to a 10 % sugar solution provided on cotton wool pads through the top of the tube. Following the observation period, live and dead individuals were recorded and placed in 96 % ethanol. A replicate within the trials consisted of one complete set of serial dilutions and one negative control (untreated paper). Four replicates were performed for each active ingredient and target population. All susceptibility tests were performed in each country in a dedicated laboratory by the same trained person following standard protocols and at a temperature of 21 ± 3 °C and relative humidity of 70 ± 10 %.

#### **Statistical analysis**

Dose–response analysis for *Culicoides* mortality followed WHO recommendations [22]. Mortality rates were calculated by pooling the total number of dead *Culicoides* by active ingredient concentration across all replicates and expressed as a percentage of the total number of exposed individuals. When control mortality exceeded 20 % of the *Culicoides* exposed the replicate was discarded, while at rates of 5 to 20 % control mortality, rates were corrected using Abbott's method (corrected mortality = 100 × (% observed mortality - % control mortality)/(100 - % control mortality) [24]). Abbott's method reduces the estimated mortality effect of the treatment by the non-treatment mortality, as measured by the control. Data were analysed by a probit regression analysis [25] using PriProbit ver. 1.63 to obtain susceptibility values (LC<sub>50</sub> and LC<sub>90</sub>) and sigmoidal curves of dose–response estimations of each insecticide active ingredient for each target population.

A second insecticide susceptibility analysis was performed to determine the effect of species origin (country and population), active ingredient concentration and their interactions. The two families of insecticide active ingredients were analysed separately as the concentrations used in testing differed. Initially, a generalised linear model with a binomial distribution was fitted, leading to an over dispersion of data (goodness of fit,  $p < 0.05$ ). To assess the fixed effects (species and origin), the differences in deviation between the complete model including fixed effects

(species, origin and doses without interaction) and without the fixed effect were calculated, taking into account the dispersion factor. R freeware (R Development Core Team 2012) and additional packages (aods3, lattice) were used for data analysis and graphics [26].

## Results

A total of 29,064 unpigmented females were used in bioassays (11,761 *C. nubeculosus*, 11,975 *C. imicola* and 5,328 *C. obsoletus*). Among the 5,516 individuals collected belonging to the *Obsoletus* group: 5,328 (96.6 %) were molecularly identified as *C. obsoletus*, 152 (2.8 %) as *C. scoticus* and 36 specimens (0.6 %) were not identified and were excluded from the analysis. Amongst the 4,973 individuals from *Obsoletus* group collected in Corrèze, France; 4,815 (96.8 %) were identified as *C. obsoletus*, 149 (3.0 %) as *C. scoticus* and 9 (0.2 %) were unidentified; from the 543 individuals collected in Mallorca Island, Spain, 513 (94.5 %) were identified as *C. obsoletus*, 3 (0.5 %) as *C. scoticus* and 27 (5.0 %) were not identified. Data analysis was only performed with *C. obsoletus* as numbers of *C. scoticus* were not sufficient for examination.

Mortality recorded 24 h after 1 h exposure to insecticide active ingredients indicated that all *Culicoides* populations were susceptible to the active ingredients tested. The lethal concentrations (LC<sub>50</sub> and LC<sub>90</sub>) calculated by probit analysis are presented in percentage of active ingredient (Tables 1, 2, 3, 4) and in its equivalent in mg/m<sup>2</sup> (Additional file 3: Tables S3, Additional file 4: Table S4, Additional file 5: Table S5a, S5b). Within SPs, permethrin elicited the highest LC<sub>50</sub> and LC<sub>90</sub> values indicating less sensitivity, whereas deltamethrin gave the lowest values for all the populations studied. Among the OPs, chlorpyrifos-methyl produced the lowest LC<sub>50</sub> and LC<sub>90</sub> values, whereas diazinon gave the highest values.

Sigmoidal curves of dose–response obtained after data analysis indicated that SPs were 1–3 log-fold more effective/unit weight than OPs (Fig. 1). Within SPs, there was no significant impact according to the species tested for deltamethrin ( $p = 0.14$ ), alpha-cypermethrin ( $p = 0.26$ ) or permethrin ( $p = 0.65$ ). In contrast, within OPs tested significant species effects were recorded in all three cases ( $p < 0.001$ ). Chlorpyrifos-methyl and diazinon elicited LC<sub>50</sub> values in *C. nubeculosus* which were 61.5–73.7 and 44.5–54.6 % higher than the other two species tested, whereas the phoxim LC<sub>50</sub> of *C. obsoletus* was 74.4–82.1 % lower than the other two species (Fig. 1). Diagnostic concentrations (defined as twice the value of LC<sub>99</sub>) for the insecticides tested are presented in Table 5 and Additional file 6: S6. In addition, the statistical analysis showed that there was no difference between species in the diagnostic concentrations for deltamethrin ( $p = 0.98$ ), alpha-cypermethrin ( $p = 0.26$ ) or permethrin ( $p = 0.16$ ). In contrast, OP active ingredients were significantly different

**Table 1** Susceptibility values (LC<sub>50</sub> and LC<sub>90</sub> expressed in % of active ingredient) of *Culicoides nubeculosus* from French colony to different active ingredients. Mortality was recorded 24 h after 1 h exposure to different concentrations

Active ingredient	No. test (n)	LC <sub>50</sub> % (95 % CI)	LC <sub>90</sub> % (95 % CI)
Deltamethrin	4 (2,528)	0.0003 (0.0001–0.0004)	0.0019 (0.0012–0.0043)
Alpha-cypermethrin	3 (1,883)	0.0016 NA	0.0199 NA
Permethrin	3 (2,055)	0.0102 (0.0080–0.0117)	0.1045 (0.0849–0.1337)
Chlorpyrifos-methyl	4 (1,803)	0.0725 NA	0.1661 NA
Phoxim	3 (1,965)	0.1532 (0.1143–0.2091)	0.2759 (0.2036–0.5761)
Diazinon	4 (1,527)	0.1839 (0.0800–0.2747)	0.3317 (0.2347–0.4333)

No test number of tests performed, n number of individuals tested, CI confidence interval, NA confidence interval not computed due to a large variability in the dose/response effect

in species effects ( $p < 0.001$ ) between chlorpyrifos-methyl, phoxim and diazinon.

When the effect of the variable origin (country) was tested, a significant effect was found for deltamethrin ( $p < 0.001$ ) and permethrin ( $p < 0.001$ ). LC<sub>50</sub> and LC<sub>90</sub> values of field-collected *Culicoides* populations from France to deltamethrin were significantly lower than those from other countries (Tables 1, 2, 3, 4). Similarly, when the effect of origin population was tested, a significant effect was found for deltamethrin ( $p < 0.001$ ) and permethrin ( $p < 0.001$ ) responses with the French population of *C. obsoletus* and the Senegal population of *C. imicola* eliciting lower LC<sub>50</sub> and LC<sub>90</sub> values for deltamethrin and permethrin, respectively (Figs. 1 and 2).

A steeper slope of sigmoidal curves (Fig. 1), and the small gap between LC<sub>50</sub> and LC<sub>90</sub> for deltamethrin and permethrin (Fig. 2), demonstrated the efficacy of pyrethroids to induce higher mortality in *Culicoides* by only a slight increase in dose. Despite the general robustness and low variability of the response within each population against each active ingredient, which can be inferred from the small size of gaps between lower and upper 95 % confidence intervals (95 % CI) (length of LC lines in Fig. 2), the results from the *C. obsoletus* population in Mallorca (Spain) were highly variable.

## Discussion

This study presents the first systematic survey of insecticide susceptibility of *Culicoides* species of veterinary interest on a wide geographic scale in Europe and Africa. More than 25,000 *Culicoides* were tested to obtain a



**Table 2** Susceptibility values (LC<sub>50</sub> and LC<sub>90</sub> expressed in % of active ingredient) of different populations of *Culicoides obsoletus* to different active ingredients. Mortality was recorded 24 h after 1 h exposure to different concentrations

Active ingredient	<i>C. obsoletus</i> (Corrèze, France)			<i>C. obsoletus</i> (Mallorca, Spain)		
	No. test (n)	LC <sub>50</sub> % (95 % CI)	LC <sub>90</sub> % (95 % CI)	No. test (n)	LC <sub>50</sub> % (95 % CI)	LC <sub>90</sub> % (95 % CI)
Deltamethrin	3 (1,491)	0.0001 (0.0000–0.0002)	0.0008 (0.0005–0.0018)	4 (382)	0.0005 (0.0002–0.0011)	0.0032 (0.0013–0.1246)
Alpha-cypermethrin	3 (502)	0.0012 NA	0.0102 NA			
Permethrin	2 (527)	0.0207 (0.0188–0.0229)	0.0668 (0.0565–0.0822)	4 (131)	0.0147 (0.0101–0.0203)	0.0840 (0.0502–0.2445)
Chlorpyrifos-methyl	5 (615)	0.0182 (0.0142–0.0222)	0.0769 (0.0632–0.0980)			
Phoxim	6 (763)	0.0273 (0.0238–0.0312)	0.1177 (0.0944–0.1564)			
Diazinon	2 (917)	0.0848 NA	0.2944 NA			

No test number of tests performed, n number of individuals tested, CI confidence interval, NA confidence interval not computed due to a large variability in the dose/response effect

robust and reliable dataset demonstrating that species within the genus varied significantly in their susceptibility to SP and OP active ingredients according to their origin. The use of the standardized method enabled the assessment of the *Culicoides* susceptibility to current insecticides, providing important baseline information, including reference values for the laboratory reared-species, *C. nubeculosus*. SPs were more toxic to the three *Culicoides* species tested than OPs, confirming previous studies conducted in wind tunnel trials carried out in USA

with other *Culicoides* vector species [27–29]. This differential toxicity between insecticide families has also previously been highlighted in other insect species such as mosquitoes [30–32] and termites [33].

The most toxic insecticides tested against *Culicoides* in the study were deltamethrin and alpha-cypermethrin, both of which are synthetic 2nd generation type II  $\alpha$ -cyano SPs. During our trials, no excito-repellency effect was observed, even when *Culicoides* were exposed to the highest concentration of permethrin, which has a documented

**Table 3** Susceptibility values (LC<sub>50</sub> and LC<sub>90</sub> expressed in % of active ingredient) of different European populations of *Culicoides imicola* to different active ingredients. Mortality was recorded 24 h after 1 h exposure to different concentrations

Active ingredient	Corsica, France			Catalonia, Spain		
	No. test (n)	LC <sub>50</sub> % (95 % CI)	LC <sub>90</sub> % (95 % CI)	No. test (n)	LC <sub>50</sub> % (95 % CI)	LC <sub>90</sub> % (95 % CI)
Deltamethrin	3 (2,525)	0.0002 (0.0001–0.0002)	0.0008 (0.0005–0.0014)	3 (378)	0.0003 (0.0002–0.0004)	0.0023 (0.0016–0.0037)
Alpha-cypermethrin	4 (2,084)	0.0008 NA	0.0034 NA			
Permethrin	4 (1,562)	0.0194 (0.0171–0.0219)	0.0812 (0.0695–0.0975)	2 (123)	0.0175 NA	0.1071 NA
Chlorpyrifos-methyl	1 (122)	0.0274 (0.0176–0.0388)	0.1273 (0.0784–0.3535)			
Phoxim	2 (2,207)	0.1053 NA	0.2441 NA			
Diazinon	2 (1,704)	0.1031 NA	0.3424 NA			

No test number of tests performed, n number of individuals tested, CI confidence interval, NA confidence interval not computed; due to a large variability in the dose/response effect

**Table 4** Susceptibility values (LC<sub>50</sub> and LC<sub>90</sub> expressed in % of active ingredient) of different African populations of *Culicoides imicola* to different active ingredients. Mortality was recorded 24 h after 1 h exposure to different concentrations

Active ingredient	Rufisque, Senegal			Pretoria, South Africa		
	No. test (n)	LC <sub>50</sub> % (95 % CI)	LC <sub>90</sub> % (95 % CI)	No. test (n)	LC <sub>50</sub> % (95 % CI)	LC <sub>90</sub> % (95 % CI)
Deltamethrin	4 (458)	0.0005 (0.0004–0.0005)	0.0015 (0.0012–0.0018)	3 (291)	0.0003 NA	0.0020 NA
Permethrin	3 (521)	0.0031 (0.0018–0.0045)	0.0168 (0.0105–0.0431)			

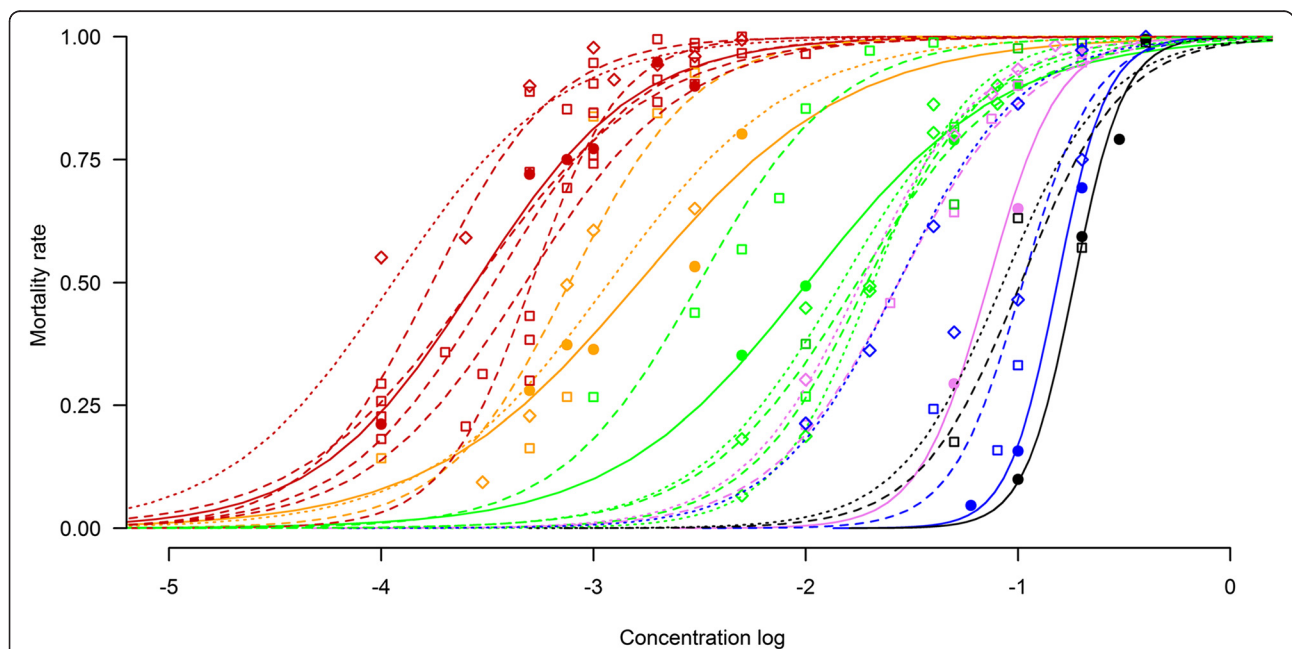
No test number of tests performed, n number of individuals tested, CI confidence interval, NA confidence interval not computed; due to a large variability in the dose/response effect

repellence effect in mosquitoes [31]. Further work is required, however, to define methods of testing the impact of repellency versus toxicity in *Culicoides* as this factor might influence their efficacy in the field.

The WHO recommendations for adult mosquito susceptibility tests advises the use of insects of standardized ages for analysis [19, 22]. This restricts material in use to either adult females derived from larval collections (the preferred option) or, if larval collections are not possible, the F<sub>1</sub> progeny of field collected females. These cohorts were not used in the present study as colonization of *C. imicola* or *C. obsoletus* has not been achieved and larval rearing does not provide sufficient numbers of adults for use [34]. As a more logistically feasible alternative to

circumvent this issue, field-collected unpigmented females were assumed to be of a similar age [23]. While this raises the issues of repeatability of the study when a population with a different age structure is assessed, the restriction to unpigmented individuals at least reduces this source of error to a likely variation of days rather than weeks, which could occur if both unpigmented and pigmented individuals were used. These issues highlight the requirement for further development of accurate age grading methods for *Culicoides*, which, at present, are lacking.

The classical approach (probit regression analysis) for assessment of dose–response data as LC<sub>50</sub>, LC<sub>90</sub> and their 95 % confidence intervals has been used for decades [19].



**Fig. 1** Sigmoidal curves of concentration-response estimations of different *Culicoides* populations exposed to different insecticides active ingredients. Dots represent pooled data obtained for each tested concentration (filled circle = *C. nubeculosus*; square = *C. obsoletus*; diamond = *C. imicola*) and lines represent the logistic regression of each population (straight: *C. nubeculosus*; dotted: *C. obsoletus*; dashed: *C. imicola*) for each active ingredient (red = deltamethrin; orange = alpha-cypermethrin; green = permethrin; purple = chlorpyrifos-methyl; blue = phoxim; black = diazinon). Data was analysed with PriProbit ver. 1.63, based on the mortality recorded at 24 h after 1 h exposure to different concentrations of active ingredients

**Table 5** Insecticide diagnostic concentrations expressed in % of active ingredient of different populations of *Culicoides* to different active ingredients

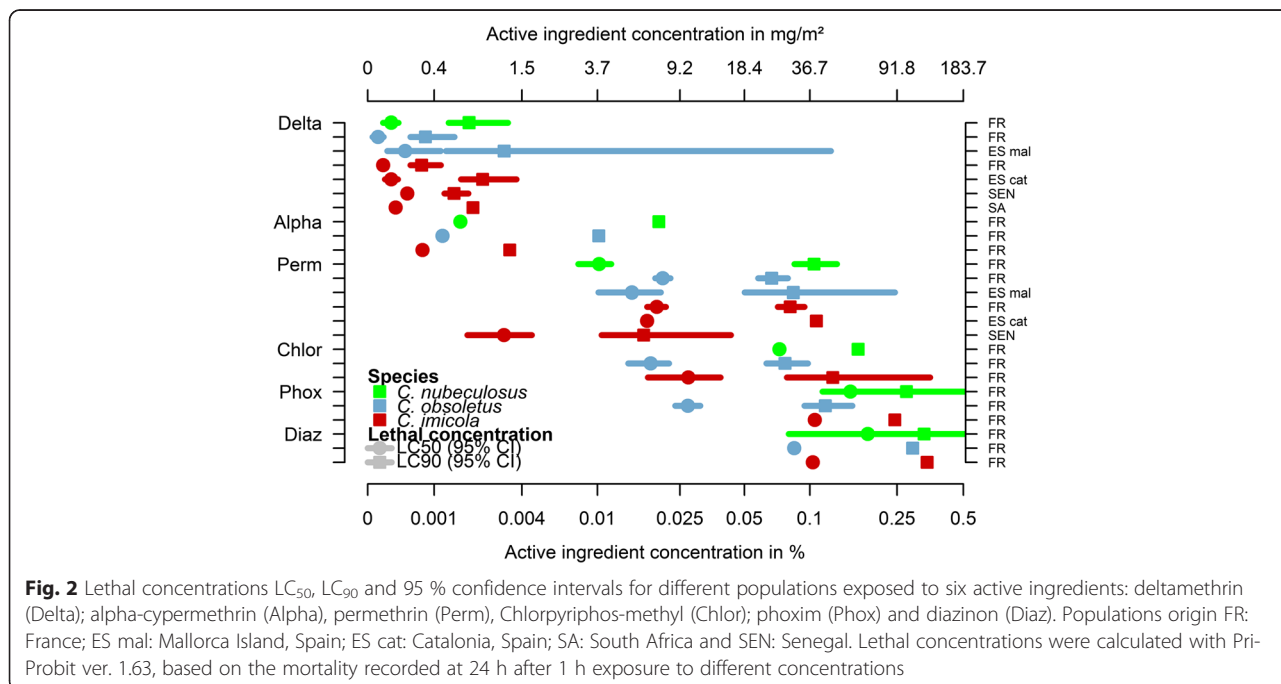
Active ingredient	Population (origin)						
	<i>C. nubeculosus</i> (Cirad, FR)	<i>C. obsoletus</i> (Corrèze, FR)	(Mallorca, ES)	<i>C. imicola</i> (Corsica, FR)	(Catalonia, ES)	(Rufisque, SEN)	(Pretoria,SA)
<b>Pyrethroids</b>							
Deltamethrin	0.03	0.01	0.04	0.01	0.04	0.01	0.03
Alpha-cypermethrin	0.61	0.21		0.03			
Permethrin	2.65	0.48	1.12	0.77	1.55	0.20	
<b>Organophosphates</b>							
Chlorpyrifos-methyl	0.82	0.74		1.36			
Phoxim	1.05	1.16		1.22			
Diazinon	1.26	2.29		2.54			

Most of diagnostic concentrations likely overestimated due to classical dose/response analysis (see discussion)  
FR France, ES Spain, SEN Senegal, SA South Africa

However, the probit regression method used in the current study for data analysis of mortality data has limitations. Mainly, this method doesn't take into account over-dispersion of the data and when this occurs it cannot calculate associated confidence intervals. The probit method also predicts beyond data observed limits and outputs reference values as LC<sub>90</sub> or LC<sub>99</sub> using this prediction, which may underestimate the effect of insecticides. Two examples illustrate this point: i) using 0.005 % of deltamethrin, 99 % of exposed *C. obsoletus* died, but the prediction for the LC<sub>99</sub> was 0.007 % (1.4-fold greater than the observed data); ii) a 99 % mortality was observed with 0.4 % of permethrin, but the prediction was LC<sub>99</sub> = 1.32 % (3.3-fold greater than the observed data). Thirdly,

results obtained after analyses are aggregated and comparison between individual trials is difficult. This is in part due to the fact that the probit regression method was designed to demonstrate if a field population of a given species is less sensitive than a susceptible reference population of the same species when both populations are exposed to the same range of concentrations of an active ingredient.

The advantage of the second method of analysis used was that over-dispersion in the datasets is taken into account in the calculation method. In addition, the predictions remain inside the data limits, no pre-determined shape was imposed on the regressions and results are more detailed. This approach highlighted no species-





specific differences in toxicity of the three SP active ingredients tested. Results with OPs highlighted species effects, however, suggesting natural species-specific susceptibility as previously reported for mosquitoes [35, 36]. This could also represent specific resistance mechanisms, although diagnostic concentrations are similar to those of susceptible mosquito strains of *Anopheles gambiae* [22].

Resistance to insecticides has been reported in the New World for *Haematobia irritans irritans* Linnaeus (Diptera: Muscidae), a large biting fly which has been specifically targeted by insecticide treatment of ruminants using pour-on applications. This resistance has been demonstrated to occur through several complex resistance mechanisms, including target site insensitivity [37, 38] and metabolic detoxification [39]. In order to detect the development of resistance mechanisms in *Culicoides*, as in mosquitoes, it is recommended to test the vector susceptibility by bioassay over time throughout the year to assess temporal trends in resistance, and also to compare multiple sites in order to assess geographical distribution of resistance. Similar surveys of broadly distributed species such as *C. imicola* and *C. obsoletus* could provide helpful information about resistance according to the different insecticide pressure and environmental context across countries. While *Culicoides* populations investigated in the current study present no evidence of resistance, low variability between results was observed except for the Mallorca (Spain) population of *C. obsoletus*. A key consideration is that multicentric insecticide trials are subject to extrinsic factors that can influence results, misleading the real effect of an insecticide to a given population [22, 40]. In the current study significant efforts were made to control conditions during testing including ambient temperature and humidity, the origin of impregnated papers and the bioassay procedure. However, one could not exclude the possibility that some differences in the procedures persist between countries.

## Conclusions

In conclusion, this study has defined the baseline susceptibility status of different *Culicoides* vector populations against six SP and OP insecticide active ingredients. The information regarding lethal concentrations will be used in future studies with the aim of testing the efficiency of insecticidal products applied directly on animals (e.g. pour-on and baths/dips), or insecticide impregnated materials (e.g. nets and paints) and also to monitor the potential emergence of resistance in field populations. This will improve our understanding of the efficacy of control measures against *Culicoides* in the field and enable better policy recommendations for their use in Europe and Africa.

## Additional files

**Additional file 1: Table S1.** *Culicoides* collections sites and date of collection during trials. (DOCX 16 kb)

**Additional file 2: Table S2.** Serial dilutions of insecticides used in trials to assess susceptibility of different populations of *Culicoides*. (DOCX 16 kb)

**Additional file 3: Table S3.** Susceptibility values ( $LC_{50}$  and  $LC_{90}$  expressed in  $mg/m^2$  of active ingredient) of *Culicoides nubeculosus* from French colony to different active ingredients. Mortality recorded 24 h after 1 h exposure to different concentrations. (DOCX 15 kb)

**Additional file 4: Table S4.** Susceptibility values ( $LC_{50}$  and  $LC_{90}$  expressed in  $mg$  of active ingredient/ $m^2$ ) of different populations of *Culicoides obsoletus* to different active ingredients. Mortality was recorded 24 h after a 1 h exposure to insecticides. (DOCX 16 kb)

**Additional file 5: Table S5a.** Susceptibility values ( $LC_{50}$  and  $LC_{90}$  expressed in  $mg$  of active ingredient/ $m^2$ ) of different European populations of *Culicoides imicola* to different active ingredients. Mortality was recorded 24 h after 1 h exposure to different concentrations. Table S5b. Susceptibility values ( $LC_{50}$  and  $LC_{90}$  expressed in  $mg$  of active ingredient/ $m^2$ ) of different African populations of *Culicoides imicola* to different active ingredients. Mortality was recorded 24 h after 1 h exposure to different concentrations. (DOC 54 kb)

**Additional file 6: Table S6.** Insecticide diagnostic concentrations expressed in  $mg$  of active ingredient/ $m^2$  of different populations of *Culicoides* to different active ingredients. (DOCX 16 kb)

## Competing interests

The authors declare that they have no competing interests.

## Authors' contributions

RV, ThiB, ThoB, SC designed the study. RV, JL, MF, RdR, ST, KL, MM, NP, GV, IG, XA, BS, LG, GG, MM carried out the bioassays. RV, ThiB, GG, ThoB, RL, CG, CC-S analysed the data. RV, ThiB, GG, ThoB, SC wrote the manuscript, which was revised by all the authors. All authors read and approved the final manuscript.

## Acknowledgments

This study was funded by the "Direction générale de l'alimentation du Ministère français en charge de l'Agriculture" and the EU grant FP7-261504. This manuscript is catalogued by the EDENext steering committee as EDENext404 (<http://www.edenext.eu>).

The authors are grateful to farmers: Jacques Virolles and family (Corrèze, France), Albert Jungen and family (Corsica, France), Joan Capó and family (Catalonia, Spain), Andreu Cosme Oliver Ramón and family (Mallorca Island, Spain), Samba Coura Diop and family (Rufisque, Senegal) and the ARC-OVI and OBP (Onderstepoort Biological Products) (Pretoria, South Africa), who permitted us to enter into their farms and made their stables available for *Culicoides* collection. Authors are also grateful to Marie Laure Setier-Rio from EID-Med (France) and Lassana Konaté from LEVP/UCAD (Senegal) for technical help.

## Author details

<sup>1</sup>EID Méditerranée, 34184 Montpellier, France. <sup>2</sup>Cirad, UMR15 CMAEE; INRA, UMR1309 CMAEE, 34398 Montpellier, France. <sup>3</sup>Institut sénégalais de recherches agricoles (ISRA), BP 1386 Dakar, Senegal. <sup>4</sup>Laboratory of Zoology, University of the Balearic Islands (UIB), Km 7.5, Palma de Mallorca, Spain. <sup>5</sup>Centre de Recerca en Sanitat Animal (CRESA), UAB-IRTA, Campus de la Universitat Autònoma de Barcelona, 08193 Bellaterra, Barcelona, Spain. <sup>6</sup>Agricultural Research Council – ARC- Onderstepoort Veterinary Institute, Private Bag X5, Onderstepoort 0110, Republic of South Africa. <sup>7</sup>The Pirbright Institute, Vector-borne Viral Disease Programme, Woking GU24 0NF, UK.

Received: 13 January 2015 Accepted: 12 August 2015

Published online: 28 August 2015

## References

- Purse BV, Carpenter S, Venter GJ, Bellis G, Mullens BA. Bionomics of temperate and tropical culicoides midges: knowledge gaps and

- consequences for transmission of culicoides-borne viruses. *Annu Rev Entomol.* 2015;60:373–92.
2. Carpenter S, Groschup MH, Garros C, Felipe-Bauer ML, Purse BV. *Culicoides* biting midges, arboviruses and public health in Europe. *Antiviral Res.* 2013;100:102–13.
  3. Sinclair M, Bührmann G, Gummow B. An epidemiological investigation of the African horsesickness outbreak in the Western Cape Province of South Africa in 2004 and its relevance to the current equine export protocol. *J S Afr Vet Assoc.* 2006;77:191–6.
  4. Hoogendam K. International study on the economic consequences of outbreaks of bluetongue serotype 8 in north-western Europe. University of Van Hall-Larenstein. Leewarden, Netherlands; 2007.
  5. Riek RF. Studies on allergic dermatitis ("Queensland itch") of the horse. *Aust Vet J.* 1953;29:177–84.
  6. Anderson GS, Belton P, Kleider N. The hypersensitivity of horses to *Culicoides* bites in British Columbia. *Can Vet J.* 1988;29:718–23.
  7. Carpenter S, Mellor PS, Torr SJ. Control techniques for *Culicoides* biting midges and their application in the U.K. and northwestern Palaearctic. *Med Vet Entomol.* 2008;22:175–87.
  8. Savini G, al e. Vaccines against bluetongue in Europe. 2007.
  9. EFSA. Scientific opinion of the panel on animal health and welfare on a request from the European commission (DG SANCO) on bluetongue. *The EFSA Journal.* 2008;7:35:1–69.
  10. Schmahl G, Walldorf V, Klimpel S, Al-Quraishy S, Mehlhorn H. Efficiency of Oxyfly on *Culicoides* species -the vectors of Bluetongue virus- and other insects. *Parasitol Res.* 2008;103:1101–3.
  11. Mehlhorn H, Schmahl G, D'Haese J, Schumacher B. Butox <sup>®</sup> 7.5 pour on: a deltamethrin treatment of sheep and cattle: pilot study of killing effects on *Culicoides* species (Ceratopogonidae). *Parasitol Res.* 2008;102:515–8.
  12. Papadopoulos E, Bartram D, Carpenter S, Mellor PS, Wall R. Efficacy of alphacypermethrin applied to cattle and sheep against the biting midge *Culicoides nubeculosus*. *Vet Parasitol.* 2009;163:110–4.
  13. Venail R, Mathieu B, Setier-Rio ML, Borba C, Alexandre M, Viudes G, et al. Laboratory and field-based tests of deltamethrin insecticides against adult *Culicoides* biting midges. *J Med Entomol.* 2011;48:351–7.
  14. del Rio R, Venail R, Calvete C, Barceló C, Baldet T, Lucientes J, et al. Sensitivity of *Culicoides obsoletus* (Meigen) (Diptera: Ceratopogonidae) to deltamethrin determined by an adapted WHO standard susceptibility test. *Parasitology.* 2014;141:542–6.
  15. Smith CN, Davis AN, Weidhaas DE, Seabrook EL. Insecticide resistance in the salt-marsh sand fly *Culicoides furens*. *J Econ Entomol.* 1959;52:352–3.
  16. Venter GJ, Labuschagne K, Hermanides KG, Boikanyo SNB, Majatladi DM, Morey L. Comparison of the efficiency of five suction light traps under field conditions in South Africa for the collection of *Culicoides* species. *Vet Parasitol.* 2009;166:299–307.
  17. Delécolle JC, De La Rocque S. Contribution à l'étude des Culicoides de Corse. Liste des espèces recensées en 2000/2001 et redescription du principal vecteur de la fièvre catarrhale ovine : *C. imicola* Kieffer, 1913 (Diptera, Ceratopogonidae). *Bull Soc Entomol Fr.* 2002;107:371–9.
  18. Nolan V, Carpenter S, Barber J, Mellor PS, Dallas J, Mordue JA, et al. Rapid diagnostic PCR assays for members of the *Culicoides obsoletus* and *Culicoides pulicaris* species complexes, implicated vectors of bluetongue virus in Europe. *Vet Microbiol.* 2007;124:82–94.
  19. WHO. World Health Organization. Instructions for determining the susceptibility or resistance of adult mosquitoes to organochlorine, organophosphate and carbamate insecticides. Diagnostic test. Geneva: WHO/VBC; 1981. p. 81.806.
  20. Rajatileka S, Burhani J, Ranson H. Mosquito age and susceptibility to insecticides. *Trans R Soc Trop Med Hyg.* 2011;105:247–53.
  21. Chareonviriyaphap T, Kongmee M, Bangs MJ, Sathantriphop S, Meunworn V, Parbaripai A, et al. Influence of nutritional and physiological status on behavioral responses of *Aedes aegypti* (Diptera: Culicidae) to deltamethrin and cypermethrin. *J Vector Ecol.* 2006;31:89–101.
  22. WHO. Test procedures for insecticide resistance monitoring in malaria vector mosquitoes. 2013.
  23. Dyce A. The recognition of nulliparous and parous *Culicoides* (Diptera: Ceratopogonidae) without dissection. *Aust J Entomol.* 1969;8:11–5.
  24. Abbott WS. A method of computing the effectiveness of an insecticide. *J Econ Entomol.* 1925;18:265–7.
  25. Finney D. Probit analysis: a statistical treatment of the sigmoid response curve. Oxford, England: Macmillan; 1947.
  26. R Development Core Team (2013). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL <http://www.R-project.org>
  27. Kline D, Haile D, Baldwin K. Wind tunnel tests with seven insecticides against adult *Culicoides mississippiensis* Hoffman. *Mosq News.* 1981;41:745–7.
  28. Floore T. Laboratory wind tunnel tests of nine insecticides against adult *Culicoides* species. *Fla Entomol.* 1985;68:678–82.
  29. Holbrook FR. Wind tunnel evaluations of insecticides applied to colonized *Culicoides variipennis* (Diptera: Ceratopogonidae). *J Fla Anti-Mosq Assoc.* 1986;57:1–3.
  30. Bansal SK, Singh KV. Efficacy of different organophosphate and synthetic pyrethroid insecticides to the larvae of malaria vector *Anopheles stephensi*, Liston. *J Environ Biol.* 2004;25:485–8.
  31. Moshia F, Lyimo I, Oxborough R, Matowo J, Malima R, Feston E, et al. Comparative efficacies of permethrin-, deltamethrin- and alpha-cypermethrin-treated nets, against *Anopheles arabiensis* and *Culex quinquefasciatus* in northern Tanzania. *Ann Trop Med Parasitol.* 2008;102:367–76.
  32. Juntarajumnong W, Pimnon S, Bangs MJ, Thanispong K, Chareonviriyaphap T. Discriminating lethal concentrations and efficacy of six pyrethroids for control of *Aedes aegypti* in Thailand. *J Am Mosq Control Assoc.* 2012;28:30–7.
  33. Su N-Y, Scheffrahn RH, Ban PM. Barrier efficacy of pyrethroid and organophosphate formulations against subterranean termites (Isoptera : Rhinotermitidae). *J Econ Entomol.* 1993;86:772–6.
  34. Nayduch D, Cohnstaedt LW, Sasaki C, Lawson D, Kersey P, Fife M, et al. Studying *Culicoides* vectors of BTV in the post-genomic era: Resources, bottlenecks to progress and future directions. *Virus Res.* 2014;182:43–9.
  35. Somboon P, Prapanthadara LA, Suwonkerd W. Insecticide susceptibility tests of *Anopheles minimus* s.l., *Aedes aegypti*, *Aedes albopictus*, and *Culex quinquefasciatus* in northern Thailand. *Southeast Asian J Trop Med Public Health.* 2003;34:87–93.
  36. Pridgeon JW, Pereira RM, Becnel JJ, Allan SA, Clark GG, Linthicum KJ. Susceptibility of *Aedes aegypti*, *Culex quinquefasciatus* Say, and *Anopheles quadrimaculatus* Say to 19 pesticides with different modes of action. *J Med Entomol.* 2008;45:82–7.
  37. Foil LD, Guerrero F, Alison MW, Kimball MD. Association of the kdr and superkdr sodium channel mutations with resistance to pyrethroids in Louisiana populations of the horn fly, *Haematobia irritans irritans* (L.). *Vet Parasitol.* 2005;129:149–58.
  38. Oyarzún MP, Li AY, Figueroa CC. High levels of insecticide resistance in introduced horn fly (Diptera: Muscidae) populations and implications for management. *J Econ Entomol.* 2011;104:258–65.
  39. Szalanski AL, Black WC, Broce AB. Esterase staining activity in pyrethroid-resistant horn flies (Diptera: Muscidae). *J Kans Entomol Soc* 1991;68:303–312.
  40. WHO. World Health Organization. Test procedures for insecticide resistance monitoring in malaria vectors, bio-efficacy and persistence of insecticides on treated surfaces. 1998.

**Submit your next manuscript to BioMed Central and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
[www.biomedcentral.com/submit](http://www.biomedcentral.com/submit)

