The influence of phenolic hydroxy substitution on the electron transfer and anti-cancer properties of compounds based on the 2-ferrocenyl-1-phenyl-but-1-ene motif

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The ferrocenyl compound 2-ferrocenyl-1,1-bis(4-hydroxyphenyl)-but-1-ene (**3**), is very cytotoxic against breast cancer cells (IC_{50} = 0.44 μM against MDA-MB-231). We now report the synthesis of a new series of *para*- and *meta*- substituted mono- and di- ferrocenyl phenols [2-ferrocenyl-1-(3-hydroxyphenyl)-1-phenyl-but-1-ene (**6**), 2-ferrocenyl-1-(3-hydroxyphenyl)-1-(4-hydroxyphenyl)-but-1-ene (**7**), 1,2-di-ferrocenyl-1-(4-hydroxyphenyl)-but-1-ene (**8**), and 1,2-di-ferrocenyl-1-(3-hydroxyphenyl)-but-1-ene (**9**)] and their electrochemical and biochemical properties, especially in comparison to the previously reported "standard" compounds [2-ferrocenyl-1-(4-hydroxyphenyl)-1-phenyl-but-1-ene (**2**) and (**3**)]. We also report the synthesis and characterization of the diphenyl analogue, 2-ferrocenyl-1,1-diphenyl-but-1-ene (**5**). This structure-activity relationship study was motivated by our hypothesis that the cytotoxicity of **3** is related to its ability to form a quinone methide structure after two *in situ* 1-electron oxidations, a process which requires the presence of at least one *p*-phenol. The mono-ferrocenyl compounds (including those previously reported) are reasonably well recognized by the oestrogen receptors α (RBAs = 0.9-9.6%) and β (RBAs = 0.28-16.3%), although the bulkier di-ferrocenyl compounds show very little affinity. *In vitro*, the cytotoxic effects of the phenolic complexes are related to the positioning of the hydroxyl group (*para*- superior to *meta*-), and to the number of ferrocenyl groups (one superior to two), with IC_{50} values against the MDA-MB-231 cell line ranging

from 0.44-3.5 μM. On the hormone-dependent breast cancer cell line MCF-7, the observed effect seems to be the result of two components, one cytotoxic (antiproliferative) and one estrogenic (proliferative). Electrochemical studies show that only the compounds with a *p*-phenol engage in proton-coupled intramolecular electron transfer.

Introduction

Although the anti-cancer properties of ferrocene-containing molecules were first studied in the late 1970s,[1] systematic investigations were not carried out until Köpf-Meyer and Neuse established anti-tumour activity for ferricenium salts in 1984.[2] This work led to the proposal that ferrocenyl compounds could be activated in the cell by biooxidation, and that both -ferricenium and ferrocene- (in a water soluble form) containing compounds could give rise to cytotoxic effects.[3] To this end ferrocene has been incorporated into water soluble polymers,[4] tethered to a DNA intercalator,[5] phosphino compounds,[6] vitamin B1,[7] and other biomolecules.[8] Diferrocene compounds[9] and ferrocene-bearing transition metal ligands,[10] and a variety of other small ferrocenyl molecules[11] have also been investigated for anti-cancer activity. Cytotoxic pathways involving DNA have been suggested for the activity of ferrocenyl compounds.[12]

Our laboratory has been studying the effects on the proliferation of breast cancer cells of ferrocenyl phenols, especially those based on the 1,1-diphenyl-but-1-ene motif.[13,14] Some small organic phenols have been shown to possess oestrogen receptor modulating properties,[15] and, at higher concentrations, cytotoxic properties.[16] We have been trying to enhance the cytotoxicity of these types of compounds by the addition of ferrocene, which we hope will be oxidized to ferricenium within the cell. We routinely test the proliferative/antiproliferative effects of our new compounds on MCF-7 (oestrogen receptor positive) and MDA-MB-231 (oestrogen receptor negative) breast cancer cell lines, and have had varying degrees of success. The most promising compounds to date are shown in Chart 1, all of which exhibit IC_{50} values at low micromolar or submicromolar concentrations for the MDA-MB-231 cell line. The MCF-7 cell line is the standard for hormone dependant breast cancers, and the proliferative/antiproliferative effects connected to the estrogenicity/antiestrogenicity of molecules at concentrations of 10^{-5}-10^{-7} M are primarily mediated by the oestrogen receptor alpha (ERα), a nuclear receptor present in these cells. It is known that the dimethyl amino chain of the active metabolite of the breast cancer drug tamoxifen, and also present in compound **1**, interacts with ERα in such a way as to prevent

DNA transcription and cell replication.[17] The other compounds, lacking the amino chain, would be expected to be estrogenic and proliferative on MCF-7 cells. However, compounds **2-4** have shown antiproliferative effects in both the MCF-7 (ER+, hormone dependent) and MDA-MB-231 (ER-, hormone independent) cell lines, which can be attributed only to the innate cytotoxicity of the molecule. It is important to note that **1-4** conform to a particular structural motif, where the ferrocenyl group is located on carbon 2 of the but-1-ene group, the phenol group(s) resides on carbon 1, and a conjugated π-system exists between the ferrocenyl and phenol groups. It appears that this motif is directly related to the cytotoxic effects shown by these compounds. In a previous study we discovered that compounds **1-4** exhibit cyclic voltammograms characteristic of an interesting structural rearrangement due to an intramolecular electron transfer from the phenolic donor to the electrochemically generated ferricenium acceptor in basic conditions.[18] We have proposed that the final outcome of this process is the formation of a reactive quinone methide-type (QM) structure after two one-electron oxidations and the loss of two protons.

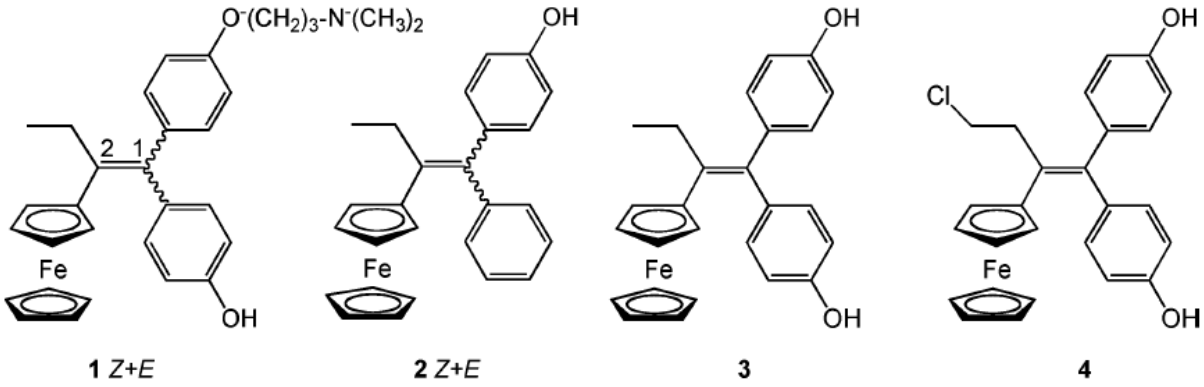


Chart 1 Previously reported cytotoxic ferrocenyl phenolic compounds **1**,[14] **2**,[19] **3**,[13] **4**.[20]

Donor-acceptor assemblies possessing a ferrocene donor have been extensively studied, especially in terms of their non-linear optical properties,[21] and structural rearrangements as a result of proton- or metal ion-coupled intramolecular electron transfer processes.[22] However, the possibility of a ferricenium moiety acting as an acceptor has only recently been explored.[23,24] In particular, a paper by Nishihara and co-workers in Tokyo described the rearrangement of 2-(2-ferrocenylvinyl)hydroquinone to a novel allene quinonoid structure *via* two one-electron oxidations and intramolecular electron transfer to a ferricenium acceptor.[25]

In order to test our hypothesis that QM-generation is related to the observed cancer cell death, and to further study this novel mechanism of formation, we have synthesized a number

of new compounds, including those where the hydroxyl group of the phenol is in the *meta*-position. For example, in compound **6**, the ferrocene-π-system-phenol motif is maintained, but it carries a *m*-phenol, so that a QM structure is not accessible. Compound **7** is similar to **6**, with the addition of a *p*-phenol group; in this case we expect that the *m*-phenol will act as a spectator, and the *p*-phenol might engage in QM formation. We have also synthesized the diferrocenyl *p*-phenol (**8**), and *m*-phenol (**9**) to evaluate the importance of the steric effect on the biological efficacy of the compounds. We here report the synthetic, electrochemical, and biochemical results for the new compounds **5-9** shown in Chart 2, and compare these results to those of previously reported compounds **2** and **3** as appropriate.

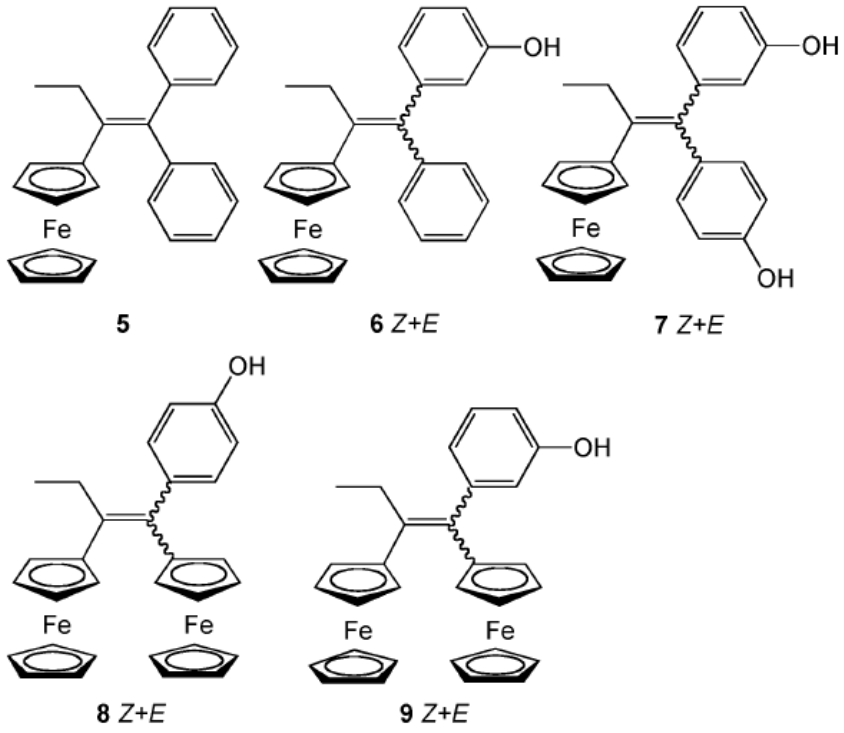
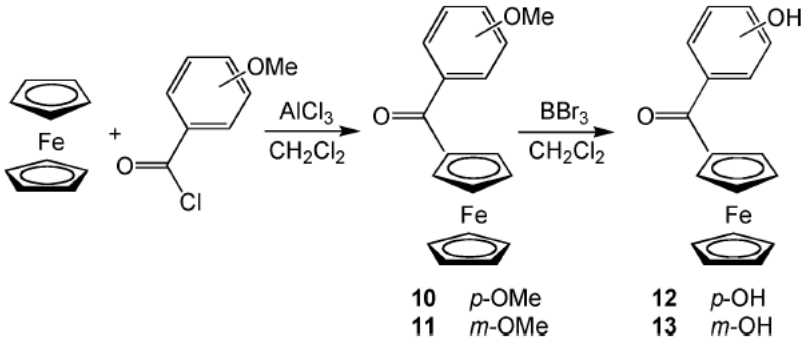


Chart 2 New compounds studied in this report.

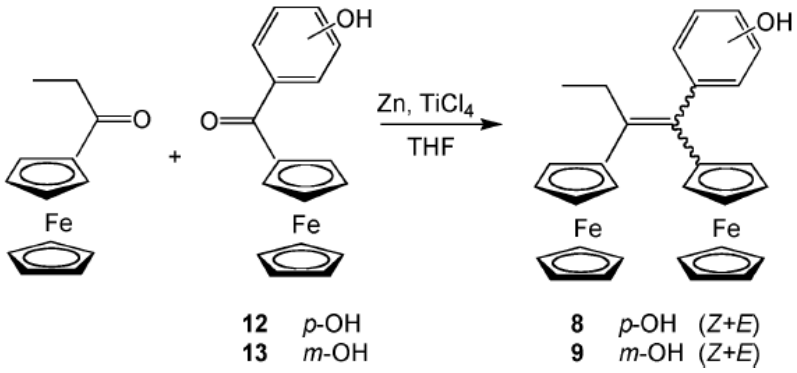


Scheme 1

Results and discussion

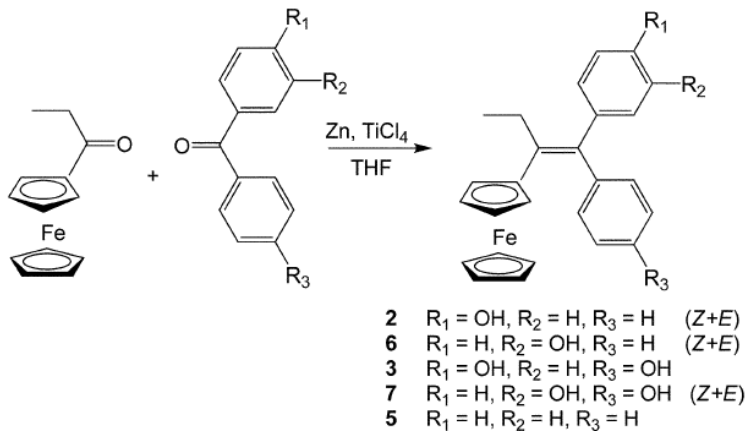
Synthesis

The formation of the new compounds **5-9** was generally accomplished by a Friedel-Craft acylation of ferrocene, followed by a McMurry cross-coupling of the ferrocenyl ketone with the appropriate benzophenone. Thus, to synthesize compounds **8** and **9**, we first prepared the known ketone **10**[26] (yield 62%) and the new ketone **11** (yield 82%) by a Friedel-Craft reaction of the corresponding acyl chlorides with ferrocene, as shown in Scheme 1. The ketones were demethyled with boron tribromide in dichloromethane to give the phenolic compounds **12** and **13** with yields of 74% and 76%, respectively. The yield for the formation of **12** using boron tribromide was higher than the previously reported demethylation of **10** with aluminium trichloride (32%),[26] but the reaction time was longer (unoptimized 12 h *vs.* 1.25 h). The McMurry reaction of these ketones with propionyl ferrocene **27** gave the cross-coupled diferrocenyl compounds **8** and **9** with a yield of 39% for each, Scheme 2.



Scheme 2

In the same manner, the McMurry reaction of propionyl ferrocene with dibenzophenone gave compound **5** with a yield of 25%, Scheme 3. This compound was impossible to separate by silica gel column chromatography from the two homo-coupled compounds, due to the similar (low) polarities of these compounds lacking any phenol group. Compound **5** was directly isolated by preparative HPLC of the crude mixture.



Scheme 3

The mono-ferrocenyl compounds **6** and **7** were synthesized from propionyl ferrocene by a McMurry reaction using the corresponding phenolic ketones (commercially available for **6**, and known from the literature for **7**[28,29]) to give a mixture of *Z* and *E* isomers in an approximately 1 : 1 ratio.

Compounds **6** and **7** were purified on a silica gel column with dichloromethane or a dichloromethane-acetone solution as the eluent, and then were re-purified *via* HPLC. *Z* and *E* isomers of compounds **6** and **7** were impossible to separate, while isomers of compounds **8** and **9** were partially separated but rapidly (within a few hours) isomerised to give a 1 : 1 mixture of *Z* and *E* isomers.

Cyclic voltammetry.

Compounds **5-9** were studied by cyclic voltammetry in methanol and methanol-pyridine solutions. In methanolic solutions, all of the compounds gave rise to the expected reversible ferrocene/ferricenium redox waves, with **5**, **6**, and **7** giving rise to one wave, and compounds **8** and **9** displaying 2 one-electron waves. The separation of observed redox potentials ($\Delta E^{\circ\prime}$) for the two waves generated by compounds **8** and **9** was not significantly different: 162 and 159 mV, respectively. These values are slightly lower than that of *trans*-Fc(CH=CH)Fc, which has been reported as 170 mV in CH_2Cl_2,[30] although the disparity can be accounted for by the decrease in Coulombic repulsion due to the solvent cage. The presence of two one-electron oxidation waves instead of one two-electron oxidation wave signifies a stabilization of the mixed valence species (Fc^{II}, Fc^{III}), the extent of which is often expressed as the comproportionation constant, *K*c. For **8** and **9**, *K*c = 550 and 490, respectively, (using the equation $\Delta E^{\circ\prime} = (RT / F)\ln Kc$)[30] and thus these compounds can be considered Robin-Day

Class II mixed-valent complexes, with moderate electron coupling between the oxidized and reduced centres.[31] Oxidation potentials *vs.* SCE are given in Table 1.

Table 1 Formal oxidation and redox potentials for compounds **2**, **5**-**9** *vs.* SCE.[a]

	Solvent	$E_{p,o}$ (Fc) $E_{p,o}$ (*other*)	$E^{\circ\prime}$ (Fc/Fc^{+})[c]	Electron transfer?
2[b]	MeOH	0.397(2)	0.357(2)	Yes
		0.97(*1*)[d]		
	MeOH-py	0.423(4)[d,e]	[d]	
		0.510(*3*)[c]		
5[b]	MeOH	0.421(3)	0.380(3)	No
	MeOH-py	0.442(3)	0.400(3)	
6	MeOH	0.403(3)	0.365(3)	No
	MeOH-py	0.408(3)	0.375(3)	
7	MeOH	0.418(3)	0.386(3)	Yes
		1.01(*1*)[d]		
	MeOH-py	0.43(1)[d,e]	[d]	
		0.528(*3*)[d]		
8	MeOH	0.334(3) 0.504(3)	0.300(3) 0.462(3) [d]	Yes
	MeOH-py	0.34(1)[d,e]	0.649(3)	
		0.444(*3*)[d]		
		0.703(3)		
9	MeOH	0.344(3) 0.503(3)	0.311(3) 0.470(3)	Yes
	MeOH-py	0.346(3) 0.523(3)	0.313(3) 0.488(3)	

[a] Scan rate = 0.1 V s^{-1}. [b] Values reported from reference 18. [c] $E^{\circ\prime}$/V is the average of the anodic and cathodic peak potentials. [d] Irreversible. [e] Shoulder.

In terms of the interaction of the electrochemically generated cations with the added pyridine, the compounds can be divided into two categories. For compounds **5**, **6** and **9**, which do not possess *p*-phenols, no significant difference in the electrochemical behaviour was observed upon the addition of pyridine, Fig. 1a,b. Conversely, for those compounds carrying a *p*-phenol, **7** and **8**, the addition of pyridine altered the CV substantially, especially in view of the reversibility of the ferrocene redox couple and the emergence of a new peak slightly higher in potential than the ferrocene oxidation. We have previously observed this

electrochemical behaviour for compounds **1-4**, and have attributed it to an intramolecular electron transfer from the organic skeleton to the ferricenium moiety coupled with deprotonation of the phenol by pyridine, eventually resulting in a QM structure.[18] While the electrochemical behaviour of compound **7** is similar to that of compounds **1-4**, that of **8** is complicated by the presence of a second ferrocenyl group. The voltammetry of **8** in the presence and absence of pyridine shows that the intramolecular electron transfer leading ultimately to the quinone methide formation occurs at the level of the first oxidation wave. Since the ferricinium group attached to the C1 carbon atom of the central double-bond (see Scheme 4) cannot be conjugated directly to the phenol moiety, one must conclude that the intramolecular phenol oxidation occurs *via* the C2 ferricinium. Yet since both ferrocene units give rise to a significant electron transfer interaction, one cannot strictly differentiate between the two under electrochemical conditions. Nevertheless, this suggests that the C1 ferrocene moiety is oxidized *after* the electron transfer to the C2 ferricenium moiety and rearrangement to the quinonoid, as shown in Scheme 4.

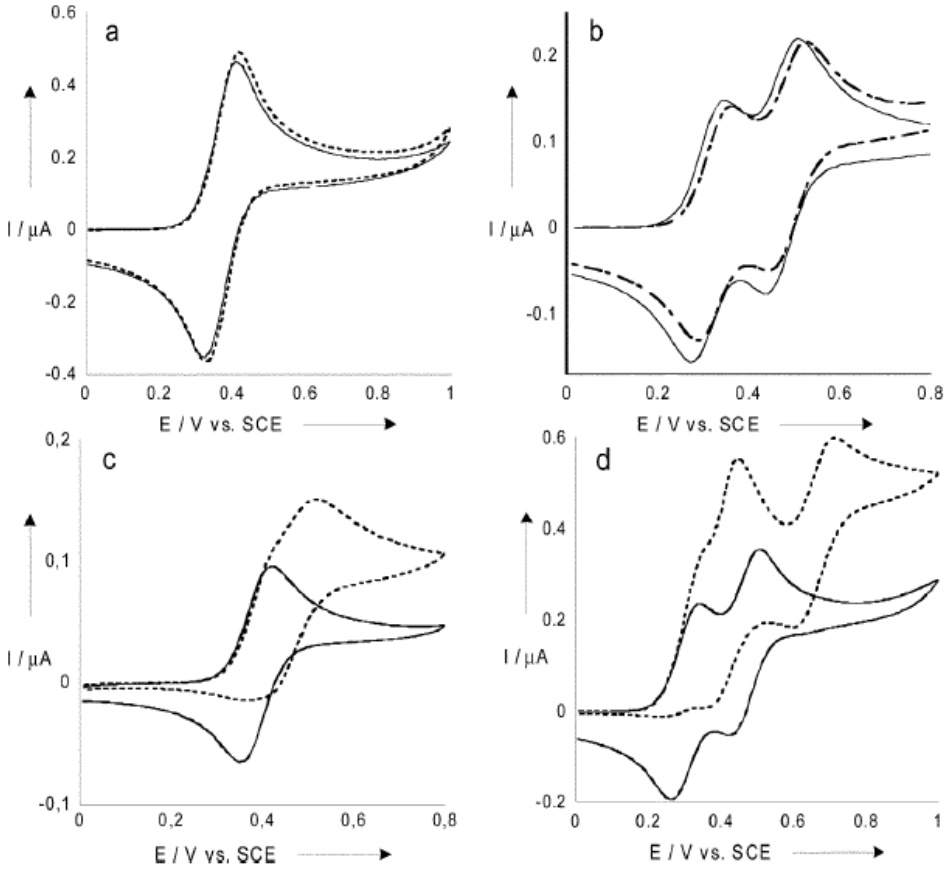
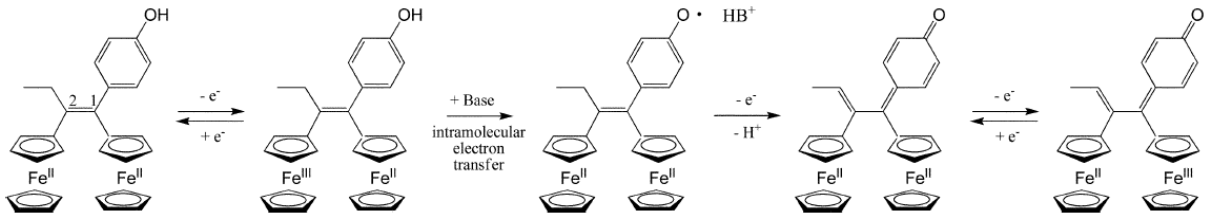


Fig. 1 Cyclic voltammograms of **6** (a), **9** (b), **7** (c), and **8** (d) in 0.1 M Bu_4NBF_4/MeOH in absence (solid line) and presence (dashed line) of pyridine in a 1 : 6 volume ratio. Scan rate

0.1 V s^{-1}. Pt electrode of 0.5 mm diameter. The CV of compound **5** has been previously published in reference 18.



Scheme 4

Thus, the electronic environment around the C1 ferrocene is expected to be substantially changed when it is finally oxidized. This is confirmed by the large anodic shift of its oxidation potential in MeOH-py compared to that in MeOH (0.703 *vs.* 0.504 V, respectively). The influence of nearby quinonoids on the destabilization of the ferricenium cation has been previously observed, and has been attributed to partial electron donation from the ferrocene to the quinonoid moiety.[24]

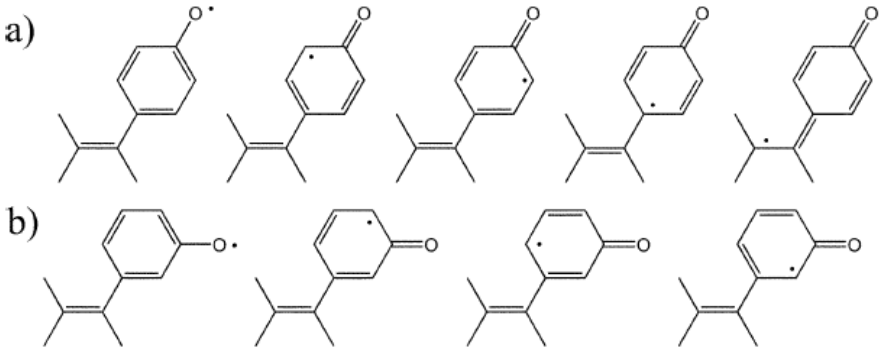


Chart 3 Electron delocalization on a) *p*-phenol and b) *m*-phenol.

We asked ourselves why only the compounds possessing a *p*-phenol engaged in electron transfer, given the ostensible similarities in molecular and electronic structure between these compounds. We have previously shown in these types of compounds that in the initial cation the radical is localized on the ferricenium group, and that there is little electronic delocalization with the phenol group prior to deprotonation.[18] This is supported by the comparison of the ferrocene oxidation potentials in MeOH for **2** (0.397 V), **5** (0.421 V), and **6** (0.403 V), which show little difference, and hence little stabilization of the radical by the phenol group. The answer may be found instead in considering the stability of the neutral

radical species which is generated after electron transfer and deprotonation. By drawing the canonical structures contributing to the delocalization of the radical, Chart 3, we can see that the *p*-phenol is able to delocalize the radical onto the C1,C2-alkene, while the radical in the *m*-phenol can only be delocalized over the phenol itself. Thus, complexes with a *para*-substituted phenol benefit from greater resonance stabilization and are therefore energetically more accessible intermediates than their *m*-phenol analogues.

Biochemistry

RBA and lipophilicity values. The affinities for the oestrogen receptor of the newly synthesized complexes were measured on the two isoforms of the oestrogen receptor, ERα and ERβ, and are reported as relative binding affinity (RBA) values in Table 2. All the compounds were recognized by both forms of the ER but the RBA values are quite different ranging from high (about 5%) to low values (less than 1%). In the mono-ferrocenyl series the change of the OH substituent from the *para*- to the *meta*-position induced only a slight decrease of the RBA value for the alpha form of the estrogen receptor (4.6% *versus* 3.6% for the monophenols, 9.6% *versus* 5.4% for the diphenols). On the contrary, the presence of an *m*-OH substituent dramatically decreased the affinity of the complexes for the beta form of the oestrogen receptor with an RBA ratio ERβ/ERβ of 21 for the monophenols **2** and **6**, and 6.8 for the diphenols **3** and **7**. Quite surprisingly, the diphenyl complex **5**, *i.e.* the compound with no OH, has a non-zero RBA value of 0.9%. A similar RBA value (0.8%) was previously reported for the corresponding organic molecule, 1,1,2-triphenylbut-1-ene.[32] Finally, the RBA values found for the di-ferrocenyl derivatives are quite low for both forms of the oestrogen receptor. This is probably due to the presence of the two ferrocenyl units which are bulkier than a phenyl substituent. Regarding the log $P_{o/w}$ values, the change of the OH group from the *para*- to the *meta*- position plays no role, and, as expected, the complexes with no hydroxyl groups or with two ferrocenyl substituents are more hydrophobic with log $P_{o/w}$ values around 6.4.

Effect of the compounds on the growth of breast cancer cells.

The effect of these complexes at a concentration of 1×10^{-6} M was studied on hormone-independent (MDA-MB-231) and hormone-dependent (MCF-7) breast cancer cells and the results are displayed in Fig. 2. The antiproliferative effect observed on the hormone-independent breast cancer cells can be attributed only to a cytotoxic effect potentially induced

by the ferrocenyl unit. On these cells, complexes **6**, **8**, and **9** show the lowest antiproliferative effects with IC_{50} values between 2.7 and 3.5 µM (Table 2); compounds **2** and **7** are more cytotoxic (IC_{50} values around 1 µM); while **3** with an IC_{50} of 0.44 µM is the most cytotoxic of the series. These results show that the repositioning of one OH group from the *para*- to the *meta*-position significantly lowers the cytotoxicity of the complex (ratio of IC_{50} values of **6** *versus* **2** and **7** *versus* **3** being respectively 2.4 and 2.3). The presence of a second ferrocenyl unit decreases the cytotoxicity of the complexes (IC_{50} of 2.8 µM for **8** and 1.13 µM for **2**; 3.5 µM for **9** and 2.7 µM for **6**); here also the complex with a *m*-OH group is less cytotoxic than the one with a *p*-OH. Finally, as expected, estradiol has no effect on these cells with no ERα.

On the contrary, the effect observed on hormone-dependent breast cancer cells is the result of the estrogenic (proliferative) effect expected for these compounds that all show an affinity for the alpha form of the ER minus its cytotoxic component observed on the hormone-independent cancer cells. At a concentration of 1 µM and in a medium without phenol red, which is best suited to the expression of the estrogenic component of a molecule, only the most cytotoxic complexes **2** and **3** are able to reverse the strong estrogenic effect shown by estradiol. As expected complexes **5** and **6** which are the less cytotoxic on the MDA-MB-231 cells show a clear proliferative effect.

Experimental

General remarks

The synthesis of all compounds was performed under an argon atmosphere, using standard Schlenk techniques. Anhydrous THF was obtained by distillation from sodium-benzophenone. Thin layer chromatography was performed on silica gel 60 GF254. Infrared spectra were obtained on an IRFT BOMEM Michelson-100 spectrometer equipped with a DTGS detector as a KBr plate. ^{1}H and ^{13}C NMR spectra were recorded on a 300 MHz Bruker spectrometer. Mass spectrometry was performed with a Nermag R 10-10C spectrometer. High resolution mass spectrometry (HRMS) was performed on a JEOL MS 700 instrument. Melting points were measured with a Kofler device. Elemental analyses were performed by the microanalysis service of CNRS at Gif sur Yvette. The semi-preparative HPLC separations were performed on a Shimadzu apparatus with a Kromasil C18 column (length of 25 cm, diameter of 2 cm, particles size of 10 µm).

Cyclic voltammograms were obtained utilizing an Autolab PG-Stat20 potentiostat, driven by GPES software (General Purpose Electrochemical System, Version 4.8, EcoChemie B.V., Utrecht, the Netherlands), a platinum wire counter electrode, a 500 µM platinum disc working electrode, and an aqueous standard calomel reference electrode. Analyte solutions were 1-2 mM in MeOH with 0.1 M Bu_4NBF_4 supporting electrolyte. Solvent ratios were 6 : 1 MeOH : py, except for compound **5**, where the ratio was 3 : 1 MeOH : py. Unfortunately, the insolubility of these compounds in water prevented the preparation of aqueous samples at suitable concentrations. Solvents were spectrometric grade and used as received.

Synthesis and characterization

Compound 10. The synthesis is described in reference 26.

Compound 11. Ferrocene (10.2 g, 54.8 mmol) was dissolved in dry dichloromethane (400 mL). Aluminium trichloride (7.30 g, 54.8 mmol) was added in portions over 15 min. Then, *m*-anisoyl chloride (7.79 g, 45.7 mmol) was added slowly over 20 min and the stirring was continued overnight. The solution was slowly poured into a mixture of water and ice and decanted. The aqueous layer was extracted with dichloromethane and the combined organic layers were washed with water, dried over magnesium sulfate and concentrated under reduced pressure. The mixture was chromatographed on a silica gel column with a solution of dichloromethane-petroleum ether 50 : 50 as the eluent. The pure product **11** was obtained as an oil (11.92 g, 82%). ^{1}H NMR (300 MHz, $CDCl_3$) δ 3.81 (s, 3 H, OCH_3), 4.14 (s, 5 H, Cp), 4.51 (t, J = 1.9 Hz, 2 H, C_5H_4), 4.85 (t, J = 1.9 Hz, 2H, C_5H_4), 7.02 (ddd, J = 8.0, 3.6, 1.0 Hz, 1 H, H_{arom}), 7.30 (t, J = 8.0 Hz, 1 H, H_{arom}), 7.35 (m, 1 H, H_{arom}), 7.42 (dt, J = 8.0, 1.0 Hz, 1 H, H_{arom}). ^{13}C NMR ($CDCl_3$) δ 55.5 (OCH_3), 70.2 (5 CH Cp), 71.5 (2 CH C_5H_4), 72.6 (2 CH C_5H_4), 78.1 (C C_5H_4), 113.1 (CH_{arom}), 117.6 (CH_{arom}), 120.6 (CH_{arom}), 129.2 (CH_{arom}), 141.1 (C), 159.5 (C), 198.9 (CO). IR: 3098, 2938, 2836 (CH_3), 1638 (CO) cm^{-1}. MS (CI, NH_3) *m*/*z*: 321 $[MH]^{+\bullet}$.

Demethylation of compounds 10 and 11. Compound **10** or **11** (3.2 g, 10 mmol), was dissolved in dry dichloromethane (60 mL) at 0 °C and boron tribromide (2.84 mL, 30 mmol) was added. The mixture was stirred overnight at room temperature. The solution was slowly poured into a mixture of water and ice and extracted with ethyl acetate. The combined organic layers were washed with water, dried over magnesium sulfate and concentrated under reduced pressure. The mixture was chromatographed on a silica gel column with dichloromethane as

the eluent to yield the phenol compounds which were recrystallized from an ether-pentane solution.

Formation of compound 12, anisoylferrocene. This compound is described in the literature and was synthesized *via* the demethylation of **10** using $AlCl_3$ (yield 32%) in place of BBr_3, the latter being used in our case. Compound **12** was obtained as a dark orange solid (2.24 g, 74% yield). The characteristics were identical to those described in the literature (mp 186 °C, 186-188 °C,[26] 190-191 °C[27]). Anal. Calcd for $C_{17}H_{14}FeO_2$: C, 66.69; H, 4.60. Found: C, 66.77; H, 4.45.

Formation of compound 13. This compound was obtained as an orange dark solid (2.3 g, 76% yield). ^{1}H NMR (300 MHz, $CDCl_3$) δ 4.12 (s, 5 H, Cp), 4.53 (s, 2 H, C_5H_4), 4.87 (s, 2 H, C_5H_4), 7.02 (d, J = 6.8 Hz, 1 H, H_{arom}), 7.12-7.41 (m, 3 H, H_{arom}), 7.45 (s, 1 H, OH). ^{13}C NMR ($CDCl_3$) δ 70.4 (5 CH Cp), 71.7 (2 CH C_5H_4), 73.0 (2 CH C_5H_4), 77.7 (C C_5H_4), 115.1 (CH_{arom}), 119.2 (CH_{arom}), 120.3 (CH_{arom}), 129.4 (CH_{arom}), 140.8 (C), 156.4 (C), 200.3 (CO). IR: 3423 (OH), 2928, 2856 (CH_3), 1614 (CO) cm^{-1}. MS (CI, NH_3) *m*/*z*: 307 $[MH]^{+\bullet}$, 324 $[M + NH_4]^+$. Anal. Calcd for $C_{17}H_{14}FeO_2$: C, 66.69; H, 4.60. Found: C, 66.83; H, 4.57.

General procedure for formation of compounds 5, 6, 7, 8, 9.

Titanium tetrachloride (3.6 mL, 33 mmol) was added dropwise to a suspension of zinc powder (4 g, 61 mmol) in 80 mL of dry THF at 0 °C. The mixture was heated at reflux for 2 h. A second solution was prepared by dissolving propionyl ferrocene (2.42 g, 10 mmol) and the corresponding ketones (10 mmol) in 50 ml of dry THF. This latter solution was added dropwise to the first solution and then the reflux was continued for 2 h. After cooling to room temperature, the mixture was stirred with water and dichloromethane. The mixture was acidified with diluted hydrochloric acid until the dark colour disappeared and was decanted. The aqueous layer was extracted with dichloromethane and the combination of organic layers was dried over magnesium sulfate. After concentration under reduced pressure, the crude product was chromatographed on a silica gel column with dichloromethane or a solution of dichloromethane-acetone 95 : 5 as the eluent (**5** excepted). For the biological tests, each products were re-purified on semi-preparative HPLC with acetonitrile-water or acetonitrile as the eluent to give pure **5**, **6**, **7**, **8**, and **9**. The isomers, if any, were either inseparable, or partially separated but remixed in the same flask (because of rapid isomerisation) before evaporation of acetonitrile under reduced pressure. The mixture was extracted with

dichloromethane and water, decanted, dried over magnesium sulfate, and concentrated under reduced pressure. The mixture of isomers, if any, was recrystallized in the appropriate solvent.

2-Ferrocenyl-1,1-di-phenyl-but-1-ene, 5. The reaction was performed with 1.82 g (10 mmol) of commercially available benzophenone. The crude product was directly purified on HPLC with acetonitrile as eluent to yield **5** as an orange solid (0.96 g, 25% yield). The compound was recrystallized from acetonitrile. Mp: 160 °C. ^{1}H NMR (300 MHz, $CDCl_3$) δ 0.97 (t, J = 7.5 Hz, 3 H, CH_3), 2.50 (q, J = 7.5 Hz, 2 H, CH_2), 3.80 (t, J = 1.9 Hz, 2 H, C_5H_4), 3.99 (t, J = 1.9 Hz, 2 H, C_5H_4), 4.04 (s, 5 H, Cp), 6.95-7.36 (m, 10 H, H_{arom}). ^{13}C NMR ($CDCl_3$) δ 15.5 (CH_3), 27.8 (CH_2), 68.2 (2 CH C_5H_4), 69.2 (5 CH Cp), 69.3 (2 CH C_5H_4), 86.5 (C C_5H_4), 126.2 (2 CH_{arom}), 128.2 (2 CH_{arom}), 128.3 (2 CH_{arom}), 129.3 (2 CH_{arom}), 129.8 (2 CH_{arom}), 137.4 (C), 138.0 (C), 144.5 (C), 144.7 (C). IR: 3078, 3044, 2963, 2930, 2869 (CH_2, CH_3) cm^{-1}. MS (EI, 70 eV) *m*/*z*: 392[M]$^{+\bullet}$, 363 [M - Et]$^{+}$, 327 [M - Cp]$^{+}$, 121 [CpFe]$^{+}$. HRMS (EI, 70 eV, $C_{26}H_{24}Fe$:M^{+}) calcd: 392.1228, found: 392.1218. Anal. Calcd for $C_{26}H_{24}Fe$: C, 79.59; H, 6.16. Found: C, 79.37; H, 6.11.

2-Ferrocenyl-1-(3-hydroxyphenyl)-1-phenyl-but-1-ene, 6. The reaction was performed with 1.98 g (10 mmol) of commercially available 3-hydroxybenzophenone. The crude product was chromatographed with CH_2Cl_2 as eluent. The compound was purified on HPLC with acetonitrile-water 80 : 20 as eluent to yield **6** as an orange solid (0.753 g, 19% yield). The mixture of isomers was recrystallized from heptane. ^{1}H NMR (300 MHz, $CDCl_3$) δ 1.03 and 1.07 (t, J = 7.4 Hz, 3 H, CH_3), 2.56 and 2.60 (q, J = 7.4 Hz, 2 H, CH_2), 3.87 and 3.94 (t, J = 1.9 Hz, 2 H, C_5H_4), 4.07 and 4.09 (t, J = 1.9 Hz, 2 H, C_5H_4), 4.11 and 4.12 (s, 5 H, Cp), 4.53 and 4.62 (s, 1 H, OH), 6.54-6.88 (m, 3 H, H_{arom}), 7.06-7.28 (m, 5 H, H_{arom}), 7.33 (t, J = 7.5 Hz, 1 H, H_{arom}). ^{13}C NMR ($CDCl_3$) δ 15.5 and 15.6 (CH_3), 27.8 (CH_2), 68.2 and 68.3 (2 CH C_5H_4), 69.2 (5 CH Cp), 69.3 (2 CH C_5H_4), 86.3 (C C_5H_4), 113.2 (CH_{arom}), 116.2 and 116.6 (CH_{arom}), 121.9 and 122.5 (CH_{arom}), 126.2 (CH_{arom}), 128.2 and 128.3 (2 CH_{arom}), 129.3 and 129.8 (2 CH_{arom}), 129.4 and 129.5 (CH_{arom}), 137.4 (C), 137.5 and 137.6 (C), 144.3 and 144.4 (C), 146.2 and 146.3 (C), 155.4 (C). IR: 3498, 3422 (OH), 3078, 3052, 3021, 2959, 2927, 2870 (CH_2, CH_3) cm^{-1}. MS (EI, 70 eV) *m*/*z*: 408 [M]$^{+\bullet}$, 379 [M - Et]$^{+}$, 343 [M - Cp]$^{+}$, 121 [CpFe]$^{+}$. HMRS (EI, 70 eV, $C_{26}H_{24}FeO$: M^{+}) calcd: 408, 1177; found: 408.1172. Anal. Calcd for $C_{26}H_{24}FeO$: C, 76.48; H, 5.92. Found: C, 76.39; H, 5.81.

2-Ferrocenyl-1-(3-hydroxyphenyl)-1-(4-hydroxyphenyl)-but-1-ene, 7. The reaction was performed with 2.14 g (10 mmol) of the known 3,4'-dihydroxybenzophenone.[29] The crude product was chromatographed with a solution of CH_2Cl_2-acetone 95 : 5 as eluent. The compound was purified on HPLC with acetonitrile-water 70 : 30 as eluent to yield **7** as an orange solid (2.43 g, 58% yield). The mixture of isomers was recrystallized from ethanol. ^{1}H NMR (300 MHz, CD_3COCD_3) δ 1.07 and 1.09 (t, J = 7.4 Hz, 3 H, CH_3), 2.64 and 2.65 (q, J = 7.4 Hz, 2 H, CH_2), 3.96 and 3.97 (t, J = 2.0 Hz, 2 H, C_5H_4), 4.10 and 4.11 (t, J = 2.0 Hz, 2 H, C_5H_4), 4.15 and 4.16 (s, 5 H, Cp), 6.59-6.65 (m, 1 H, H_{arom}), 6.66-7.05 (m, 5 H, H_{arom}), 7.05-7.25 (m, 2 H, H_{arom}), 8.12 and 8.21 (s, 1 H, OH), 8.24 and 8.27 (s, 1 H, OH). ^{13}C NMR (CD_3COCD_3) δ 15.9 and 16.0 (CH_3), 28.2 and 28.3 (CH_2), 68.8 (2 CH C_5H_4), 69.9 (5 CH Cp), 70.0 (2 CH C_5H_4), 87.2 and 87.4 (C C_5H_4), 113.8 and 113.9 (CH_{arom}), 115.8 and 115.9 (2 CH_{arom}), 116.9 and 117.3 (CH_{arom}), 121.2 and 121.7 (CH_{arom}), 129.9 and 130.0 (CH_{arom}), 131.0 and 131.5 (2 CH_{arom}), 136.8 and 136.9 (C), 137.2 and 137.4 (C), 138.7 (C), 147.4 and 147.6 (C), 156.7 and 156.8 (C), 158.1 and 158.2 (C). IR: 3284, 3397 (OH), 2872, 2931, 2977, 3092 (CH_2, CH_3) cm^{-1}. MS (EI, 70 eV) m/z: 424 $[M]^{+\bullet}$, 395 $[M - Et]^+$, 359 $[M - Cp]^+$, 121 $[CpFe]^+$. HRMS (EI, 70 eV, $C_{26}H_{24}FeO_2$:M^+) calcd: 424.1126, found: 424.1129. Anal. Calcd for $C_{26}H_{24}FeO_2$: C, 73.59; H, 5.70. Found: C, 73.72; H, 5.85.

1,2-di-Ferrocenyl-1-(4-hydroxyphenyl)-but-1-ene, 8. The reaction was performed with 3.06 g (10 mmol) of the known compound **12**.[26] The crude product was chromatographed with CH_2Cl_2 as eluent. The compound was purified on HPLC with acetonitrile-water 90 : 10 as eluent to yield **8** as an orange solid (1.98 g, 39% yield). The mixture of isomers was recrystallized from an ether-pentane solution. ^{1}H NMR (300 MHz, $CDCl_3$) δ 1.12 and 1.38 (t, J = 7.4 Hz, 3 H, CH_3), 2.32 and 2.81 (q, J = 7.4 Hz, 2 H, CH_2), 3.65 and 3.69 (t, J = 1.9 Hz, 2 H, C_5H_4), 3.83 and 4.00 (s, 5 H, Cp), 4.02 and 4.09 (s, 5H, Cp), 4.00-4.25 (m, 6 H, C_5H_4), 5.10 and 5.14 (s, 1 H, OH), 6.89 and 6.90 (d, J = 8.5 Hz, 2 H, H_{arom}), 7.08 and 7.17 (d, J = 8.5 Hz, 2 H, H_{arom}). ^{13}C NMR ($CDCl_3$) δ 15.4 and 15.5 (CH_3), 26.6 and 30.9 (CH_2), 67.3 and 68.1 (2 CH C_5H_4), 67.4 and 68.3 (2 CH C_5H_4), 68.9 and 69.0 (2 CH C_5H_4), 69.1 (2 × 5 CH Cp), 69.6 and 70.1 (2 CH C_5H_4), 86.8 and 89.6 (C C_5H_4), 88.8 and 89.0 (C C_5H_4), 114.6 and 115.2 (2 CH_{arom}), 130.2 and 131.1 (2 CH_{arom}), 132.5 and 134.8 (C), 136.5 and 136.6 (C), 137.2 and 137.3 (C), 154.0 and 154.3 (C). IR: 3438 (OH), 3092, 2967, 2929, 2872 (CH_2, CH_3) cm^{-1}. MS (EI, 70 eV) m/z: 516 $[M]^{+\bullet}$, 451 $[M - Cp]^+$, 121 $[CpFe]^+$. HRMS (EI, 70 eV, $C_{30}H_{28}Fe_2O$: M^+)

calcd: 516.0840, found: 516.0820. Anal. Calcd for $C_{30}H_{28}Fe_2O$: C, 69.79; H, 5.46. Found: C, 69.91; H, 5.39.

1,2-di-Ferrocenyl-1-(3-hydroxyphenyl)-but-1-ene, 9. The reaction was performed with 3.06 g (10 mmol) of compound **13**. The crude product was chromatographed with CH_2Cl_2 as eluent. The compound was purified on HPLC with acetonitrile-water 90 : 10 as eluent to yield **9** as an orange solid (2.00 g, 39% yield). The mixture of isomers was recrystallized from an ether-pentane solution. ^{1}H NMR (300 MHz, $CDCl_3$) δ 1.14 and 1.37 (t, J = 7.4 Hz, 3 H, CH_3), 2.32 and 2.81 (q, J = 7.4 Hz, 2 H, CH_2), 3.69 and 3.74 (t, J = 1.9 Hz, 2 H, C_5H_4), 3.82 and 4.00 (s, 5 H, Cp), 4.04 and 4.09 (s, 5 H, Cp), 4.00-4.25 (m, 6 H, C_5H_4), 4.84 and 4.87 (s, 1 H, OH), 6.65-6.95 (m, 3 H, H_{arom}), 7.30 (t, J = 7.9 Hz, 1 H, H_{arom}). ^{13}C NMR ($CDCl_3$) δ 14.5 (CH_3), 25.5 and 29.6 (CH_2), 66.4 and 67.1 (2 CH C_5H_4), 66.5 and 67.4 (2 CH C_5H_4), 67.9 and 68.0 (2 CH C_5H_4), 68.2 (2 × 5 CH Cp), 68.6 and 69.0 (2 CH C_5H_4), 85.6 and 88.3 (C C_5H_4), 87.4 and 88.5 (C C_5H_4), 112.0 and 112.4 (CH_{arom}), 114.9 and 116.0 (CH_{arom}), 120.3 and 121.3 (CH_{arom}), 127.7 and 128.2 (CH_{arom}), 131.7 and 133.5 (C), 135.3 and 135.4 (C), 145.2 and 145.3 (C), 154.5 and 155.0 (C). IR: 3407 (OH), 3094, 2967, 2975, 2927, 2866 (CH_2, CH_3) cm^{-1}. HRMS (CI, NH_3, $C_{30}H_{29}Fe_2O$: MH^+) calcd: 517.0918, found: 517.0922. Anal. Calcd for $C_{30}H_{28}Fe_2O$: C, 69.79; H, 5.46. Found: C, 69.67; H, 5.48.

Biochemical experiments

Materials

Stock solutions (1×10^{-3} M) of the ferrocenyl complexes **2**, **3**, and **5-9** to be tested were prepared in DMSO and were kept at 4 °C in the dark; under these conditions they are stable at least two months. Serial dilutions in DMSO were prepared just prior to use. A stock solution (1×10^{-3} M) of 17β-estradiol was prepared in ethanol. Dulbecco's modified eagle medium (DMEM) was purchased from Gibco BRL, fetal calf serum from Dutscher, Brumath, France, glutamine, estradiol and protamine sulfate were from Sigma. MCF-7 and MDA-MB-231 cells were from the Human Tumor Cell Bank. Sheep uteri weighing approximately 7 g were obtained from the slaughterhouse at Mantes-la-Jolie, France. They were immediately frozen and kept in liquid nitrogen prior to use.

Determination of the Relative Binding Affinity (RBA) of the compounds for ERα and ERβ. RBA values were measured on ERα from lamb uterine cytosol and on ERβ purchased in solution from Pan Vera (Madison, WI, USA). Sheep uterine cytosol prepared in buffer A (0.05 M Tris-HCL, 0.25 M sucrose, 0.1% β-mercaptoethanol, pH 7.4 at 25 ◦C) as described previously,[14] was used as a source of ERα. For ERβ, 10 µl of the solution containing 3500 pmol ml^{-1} were added to 16 ml of buffer B (10% glycerol, 50 mM Bis-Tris-Propane pH = 9, 400 mM KCl, 2 mM DTT, 1 mM EDTA, 0.1% BSA) in a silanized flask. Aliquots (200 µl) of ERα in glass tubes or ERβ in polypropylene tubes were incubated for 3 h at 0 °C with [6,7-^{3}H]-estradiol (2×10^{-9} M, specific activity 1.62 TBq $mmol^{-1}$, NEN Life Science, Boston MA) in the presence of nine concentrations of the ferrocenyl complexes **2**, **3**, and **5-9** to be tested (between 6×10^{-7} M and 6×10^{-9} M for the complexes with RBA values higher than 5% and between 6×10^{-6} M and 6×10^{-8} M for the compounds with RBA values lower than 5%) or of 17β-estradiol (between 8×10^{-8} M and 7.5×10^{-10} M). At the end of the incubation period, the fractions of [^{3}H]-estradiol bound to the estrogen receptors (Y values) were precipitated by addition of a 200 µl of a cold solution of protamine sulfate (1 mg mL^{-1} in water). After a 10 min period of incubation at 4 °C, the precipitates were recovered by filtration on 25 mm circle glass microfibre filters (GF/C) using a Millipore 12 well filtration ramp. The filters were rinsed twice with cold phosphate buffer and then transferred in 20 ml plastic vials. After addition of 5 ml of scintillation liquid (BCS Amersham) the radioactivity of each fraction was counted in a Packard tri-carb 2100TR liquid scintillation analyzer. The concentration of unlabeled steroid required to displace 50% of the bound [^{3}H]-estradiol was calculated for 17β-estradiol and for each complex by plotting the logit values of *Y* (logit $Y = \ln(Y/100 - Y)$ *versus* the mass of the competing complex. The RBA was calculated as follows: RBA of a compound = concentration of estradiol required to displace 50% of [^{3}H]-estradiol × 100/concentration of the compound required to displace 50% of [^{3}H]-estradiol. The RBA value of estradiol is by definition equal to 100%.

Measurement of octanol/water partition coefficient (log $P_{o/w}$) of the compounds. The log $P_{o/w}$ values of the compounds were determined by reverse-phase HPLC on a C-8 column (Nucleosil 5 C8, from Macherey Nagel, France) according to the method previously described by Minick[33] and Pomper.[34] Measurement of the chromatographic capacity factors (*kN*) for each compounds was performed at various concentrations in the range 85%-60% methanol (containing 0.25% octanol) and an aqueous phase consisting of 0.15% n-decylamine in 0.02 M MOPS (3-morpholinopropanesulfonic acid) buffer pH 7.4 (prepared in 1-octanol-saturated

water). These capacity factors (*kN*) are extrapolated to 100% of the aqueous component given the value of *k*'w. log $P_{o/w}(y)$ is then obtained by the formula: $y = 0.13418 + 0.98452 \times \log k'w$.

Culture conditions

Cells were maintained in monolayer in DMEM with phenol red supplemented with 8-9% fetal calf serum and 2 mM glutamine at 37 °C in a 5% CO_2 air humidified incubator. For proliferation assays, cells were plated in 24-well sterile plates at a density of 1.1×10^4 cells for MDA-MB-231 and of 3×10^4 cells for MCF-7 in 1 mL of DMEM medium without phenol red, supplemented with 10% decomplemented and hormone-depleted fetal calf serum and 2 mM glutamine and incubated. The following day (D0), 1 ml of the same medium containing the compounds to be tested was added to the plates (final volumes of DMSO: 0.1%; 4 wells for each conditions). After 3 days (D3) the incubation medium was removed and fresh medium containing the compounds was added. After 5 days (D5) the total protein content of the plate was analyzed by methylene blue staining as follows. Cell monolayers were fixed for 1 h in methanol, stained for 1 h with methylene blue (1 mg mL^{-1}) in PBS, then washed thoroughly with water. One ml of HCl (0.1 M) was then added and the absorbance of each well was measured at 620 nm with a Biorad spectrophotometer. The results are expressed as the percentage of proteins *versus* the control.

Conclusions

Our results show that the presence of a *p*-phenol has a significant influence on both the electrochemistry and on the biological efficacy of these ferrocenyl phenol compounds, supporting our hypothesis that oxidative activation to a QM structure may be a key to their biological activity. Compounds **5** and **6** which do not possess a *p*-phenol also did not show intramolecular electron transfer and rearrangement to a QM structure in the electrochemical experiments, as expected. Compound **5**, lacking any kind of phenolic group, showed no appreciable cytotoxicity, while **6**, possessing only a *m*-phenol, was much less efficacious than its *para*-substituted analogue **2**. Similarly, compound **7**, carrying one *m*- and one *p*-phenol did not show nearly as strong an activity as that of **3**, which possesses two *p*-phenols. In fact, the biological activity of **7** is much closer to that of **2** (with IC_{50} values of 1.13 *vs.* 1.03 µM, respectively) than that of **3** (0.44 µM).

The diferrocenyl compounds **8** and **9** follow the same trend, with higher biological activity observed for *p*-phenol **8**, than *m*-phenol **9**. As expected, electron transfer from the phenol to the ferricenium moiety is only electrochemically observed for compound **8**. However, the activity of these compounds cannot be solely attributed to their electronic and structural rearrangements. There also exists the steric effect of the additional ferrocene group, which has a negative effect on the biological activity of **8** and **9**. Whether carrying a *p*- or *m*- phenol, the diferrocenyl compounds are less efficacious than their mono-ferrocenyl analogues **2** and **6**.

Although each of the compounds carrying a *p*-phenol behaves similarly from an electrochemical perspective, the steric effect suggests that the QMs are formed, or react, *via* one or several specific biological pathways. For example, QMs are alkylating agents which can react with O-, N-, or S- nucleophiles, especially glutathione, *via* a Michael 1,4-addition.[35] The depletion of glutathione can then lead to the alkylation of proteins by an excess of quinone. Alternatively, quinonoids are known to undergo redox cycling in cells resulting in the production of ROS, *via* such enzymes as NADPH:cytochrome P450 reductase, NADPH:cytochrome b_5 reductase, NADPH:ubiquinone oxidoreductase, and NAD(P)H:quinone oxidoreductase, among others,[36] and iron-containing compounds are known to produce ROS *via* the Fenton reaction and H_2O_2. We are currently investigating the ROS production of these types of compounds, and we hope that future use of specific ROS probes and enzyme-inhibitors will allow us to begin to unravel the metabolic fate of this new class of cytotoxic ferrocenyl phenolic compounds.

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