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Metallic oxide nanoparticle translocation across the human bronchial epithelial barrier

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Footnote :

† Electronic supplementary information (ESI) available: 1. NP synthesis. 2. NP characterization. 3. NP cytotoxicity and pro-inflammatory response. 4. Modulation of cellular permeability after NP treatment and NP retention inside the Calu-3 monolayer after 1 week of exposure. 5. NP translocation through TF in the absence of cells as a function of NP concentration, composition or corona. 6. Confocal microscopy observations of NCI-H292 cells exposed to fresh and conditioned media. 7. Quantification of NP internalization by Calu-3 cells by flow cytometry. See DOI: [10.1039/c4nr07079h](https://doi.org/10.1039/c4nr07079h)

Inhalation is the most frequent route of unintentional exposure to nanoparticles (NPs). Our aim was to quantify the translocation of different metallic NPs across human bronchial epithelial cells and to determine the factors influencing this translocation. Calu-3 cells forming a tight epithelial barrier when grown on a porous membrane in a two compartment chamber were exposed to fluorescently labelled NPs to quantify the NP translocation. NP translocation and uptake by cells were also studied by confocal and transmission electron microscopy. Translocation was characterized according to NP size (16, 50, or 100 nm), surface charge (negative or positive SiO₂), composition (SiO₂ or TiO₂), presence of proteins or phospholipids and in an inflammatory context. Our results showed that NPs can translocate through the Calu-3 monolayer whatever their composition (SiO₂ or TiO₂), but this translocation was increased for the smallest and negatively charged NPs. Translocation was not associated with an alteration of the integrity of the epithelial monolayer, suggesting a transcytosis of the internalized NPs. By modifying the NP corona, the ability of NPs to cross the epithelial barrier differed depending on their intrinsic properties, making positively charged NPs more prone to translocate. NP translocation can be amplified by using agents known to open tight junctions and to allow paracellular passage. NP translocation was also modulated when mimicking an inflammatory context frequently found in the lungs, altering the epithelial integrity and inducing transient tight junction opening. This in vitro evaluation of NP translocation could be extended to other inhaled NPs to predict their biodistribution.

Introduction

During the last few decades the widespread use of nanomaterials (NMs) in many fields such as cosmetics, electronics or textile industries has raised questions about their potential adverse health effects.¹ Nanoparticles (NPs) can enter human organisms through different pathways such as lungs, the digestive

tract, or skin,² and their unique properties are suspected to favour their ability to easily cross protective biological barriers. The respiratory tract, as an interface organ, is a major route of NP entry after involuntary exposure and NPs can deposit all along the airways as a function of their physicochemical properties. Conducting airways, composed of a mucociliary epithelium, contribute to the particle clearance through a “mucociliary escalator”. The mucus lining the epithelium helps to trap the particles present in the inhaled air and is expelled by the activity of ciliated cells.^{2,3} Moreover the presence of a belt of tight junctions on the apical side of the cells prevents the passage of exogenous particles through the paracellular space, thus restricting their access to the interstitium.⁴⁻⁶ In the alveolar region, the most potent mechanism for solid particle clearance is mediated by alveolar macrophages through phagocytosis of deposited particles that is, however, less efficient for NPs.^{2,3}

Despite all these protective mechanisms *in vivo* studies have shown important lung retention. Previous studies have revealed that cerium,⁷ gold⁸⁻¹⁰ or iridium¹¹ NPs could be trapped inside rat or mice lungs. This NP pulmonary retention is associated with NP translocation to the systemic circulation followed by a bio-distribution into different organs.^{7,10,12,13} *In vivo* studies having provided evidence of translocation, *in vitro* models of biological barriers are needed to characterize the NP determinants favouring their capacity to go through protective epithelium and to determine the underlying mechanisms. Numerous *in vitro* studies have investigated the mechanisms of NP internalization and have determined that NPs can enter cells by active transport via macropinocytosis,^{14,15} caveolae-mediated processes,^{16,17} or clathrin-dependant pathways.¹⁸⁻²⁰ However, very few *in vitro* studies have questioned the mechanism of NP translocation through the pulmonary barrier.

The physicochemical characteristics of NPs involved in their fate have not yet been clearly identified. In addition to their intrinsic features such as shape, size, composition, or surface charges, the biological medium encountered by the NPs could influence their behaviour and interaction with cells due to modifications of their agglomeration and the formation of a corona.^{21,22} In the respiratory tract, this corona could include proteins of the mucus layer and of the epithelial lining fluids and molecules of the alveolar surfactant.^{23,24} In the context of respiratory diseases, translocation could be increased. People suffering from chronic inflammatory diseases such as asthma or COPD (chronic obstructive pulmonary disease) could be more susceptible to NP penetration.

In this study our aim was the characterization and the quantification of the translocation of NPs across an *in vitro* model of the pulmonary barrier according to the physicochemical properties of the NPs and the integrity of the bronchial epithelium. For this purpose, we used different metal oxide NPs: SiO₂-NPs of three different sizes (16, 50, 100 nm) with negative or positive surface charges, and TiO₂-coated SiO₂-NPs with a size of 140 nm. These NPs were fluorescently labelled for facilitating their detection after cellular uptake or to follow their translocation.^{16,25,26} We performed our experiments with the Calu-3 cell line seeded onto a Transwell Filter (TF) because of their *in vitro* abilities to develop a confluent and tight monolayer.²⁷⁻³⁰ Qualitative experiments by confocal and transmission electron microscopy (TEM) were carried out to determine NP internalization by Calu-3 monolayers. Furthermore fluorescence measurements of media inside the apical and basolateral chambers allowed the reliable quantification of NP translocation across the Calu-3 epithelium. NP translocation was compared according to their size, charge and composition as well as treatment conditions (the presence or absence of foetal calf serum (FCS) or dipalmitoyl lecithin (DPL) which has been suspected to form a protein or lipid corona respectively). To mimic inflammatory pathological conditions, the epithelial monolayer was previously exposed to the pro-inflammatory mediator tumor necrosis factor-alpha (TNF- α) or to lipopolysaccharide (LPS) to stimulate an inflammatory response.

Briefly, we have demonstrated a NP translocation based on NP physicochemical characteristics without cytotoxicity. NP translocation followed NP uptake by Calu-3 cells which highlights transcytosis

mechanisms. NP translocation could be modulated by the formation of a protein or lipid corona. Mimicking pathological contexts frequently observed in the lungs, we noted a paracellular NP translocation in addition to the transcytosis pathway.

Results and discussion

NP characterization

The NP synthesis process was validated by Transmission Electron Microscopy (TEM) observations (ESI S1A[†]) showing the expected NP sizes and by Diffuse Reflectance Infrared Fourier Transform (DRIFT) spectroscopy (ESI S2A[†]). The different NPs were compared for their isoelectric point by laser Doppler interferometry showing the same surface features for fluorescent and non-fluorescent NPs (IEP, ESI S2B[†]). Evaluation of the ratio between Si and Ti elements measured by ICP-OES for the TiO₂-coated SiO₂-NPs of 140 nm size allowed confirmation of the estimated 2 nm thickness of the TiO₂ (ESI S2C[†]).

Characterization of NPs suspended in DMEM/F-12 medium at 5 µg cm⁻² was performed by measurements of Dynamic Light Scattering (DLS) and zeta Potential (Pz) (ESI S1B[†]). These experiments have shown small agglomerates for all NPs. We could note that SiO₂-FITC-NPs⁺ have formed larger agglomerates than SiO₂-FITC-NPs. Furthermore a negative surface charge in DMEM/F12 culture medium was observed for each batch of NPs. This low absolute charge could explain the colloidal instability. It was previously described for polymer NPs of different surface charges.³¹

Calu-3 monolayer characterization

We chose to use Calu-3 cell lines as these cells were able to form a confluent (Fig. 1A) and tight epithelium after 14 days of growth on a TF of 3 µm porosity. Liquid covered cultures were chosen for this study as they develop tighter junctions and a less permeable cell layer compared to air-liquid interface cultures³² and because exposure was performed with NPs in the liquid phase. These cells kept their capacities to secrete mucus,^{33,34} and more specifically the MUC5AC mucin (Fig. 1B). The immunodetection of zonula-occludens 1 (ZO-1) and claudin-4 (CLD-4) proteins (Fig. 1C and 1D) known for their binding to tight junctions, and labelling of the actin filaments (Fig. 1E) revealed the formation of a tight and confluent epithelium. Moreover we have previously demonstrated that Calu-3 cells developed a strong trans-epithelial electric resistance (TEER) of 1200 mV after 14 days of culture when the passage of a fluorescent marker, the Lucifer Yellow (452 Da), was prevented.²⁸ Our results are in accordance with those of Florea and collaborators obtained for Calu-3 cells seeded onto a 0.4 µm pore TF.³⁵ Altogether these characteristics make Calu-3 cells an appropriated cell line to study NP translocation across a pulmonary epithelium^{27,28,36,37} by comparison with other human pulmonary cell lines such as NCI-H292 and A549 cells that did not form a tight epithelium.^{28,38-40}

NP exposure did not alter the epithelial monolayer viability and integrity

As determined by the WST-1 metabolic cytotoxicity assay, NPs were not cytotoxic for Calu-3 cells exposed to 16, 50, 100 nm SiO₂-FITC-NPs or 140 nm TiO₂-coated SiO₂-FITC-NPs until 50 µg cm⁻² for 24 h (ESI Fig. S3A[†]). The absence of SiO₂-NP cytotoxicity has previously been shown for several cell types such as fibroblasts, but for other cells like macrophages SiO₂-NPs have induced an important toxicity but only at very high concentrations up to 1000 µg mL⁻¹.^{41,42} This discrepancy could be attributed to SiO₂-NP characteristics or to cellular sensitivity.

In our study we limited NP treatments to 5 and 10 $\mu\text{g cm}^{-2}$ to ensure complete absence of cytotoxicity. Moreover at these concentrations no, or a low, pro-inflammatory response characterized by IL-8 release was observed after NP treatment (ESI Fig. S3B[†]). NP fluorescence was easily detected and it could be considered as non-excessive exposure concentrations. In addition we have evaluated whether NP treatment could alter epithelial barrier integrity by measuring TEER values before and after 24 h of NP treatment. After 24 h of exposure at 5 and 10 $\mu\text{g cm}^{-2}$ we observed an increase of TEER. This result indicated that NPs did not alter the integrity of tight junctions and suggested that NPs could not cross the cellular monolayer through the paracellular pathway (ESI Fig. S4A[†]).

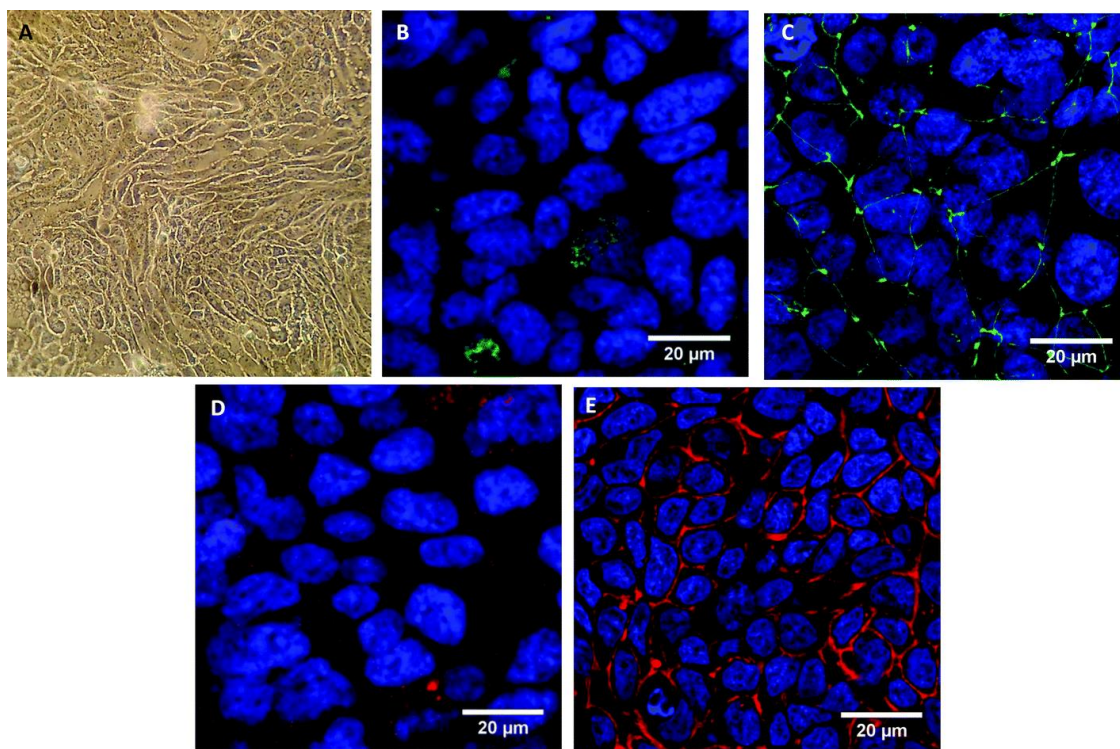


Fig. 1 Morphological characterization of the Calu-3 epithelium without any treatment. Control Calu-3 cells were observed by optical microscopy (A) or confocal microscopy (B–E) after 14 days of TF-culture. The (x;y) images correspond to the z-projection of a stack of 5 to 10 images acquired. Staining of the cells is as follows: blue – DAPI-stained nuclei, green – (B) MUC5AC protein labelled with Alexa-488-anti mouse antibody or (C) ZO-1 protein labelled with Alexa-488-antirabbit antibody, red – (D) CLD-4 protein labelled with Alexa-568-antigoat antibody, or (E) phalloidin-stained actin filaments. The scale bar corresponds to 20 μm .

NP uptake into the epithelial monolayer

To support the hypothesis of a NP transcytosis, we assessed NP uptake by Calu-3 cells (Fig. 2A). Confocal microscopy observations allowed the demonstration of NP internalization and that NP uptake did not modify actin organization and ZO-1 distribution to tight junctions. Orthogonal views clearly demonstrated that NPs were located inside the cellular monolayer and very few NPs adsorbed on the top of the cells. The pictures revealed a heterogeneous NP distribution and NP agglomerates inside the cellular monolayer whatever the NP size.

It has been known that NPs could be internalized by cells through different mechanisms, among which is clathrin-mediated or caveolae-mediated endocytosis.^{20,43,44} Despite NP endocytosis in Calu-3 cells was very poorly documented, a study has shown that Calu-3 cells exposed to amphiphilic glycopolymer NPs could internalize NPs by clathrin-mediated endocytosis or lipid raft/caveolae-mediated endocytosis but not by macropinocytosis.⁴⁵ Other NP types have been shown to be internalized by Calu-3 cells as CeO_2 -

NPs as a function of their effective density and so their sedimentation speed²⁹ or as PLGA (poly(lactide-co-glycolide)) NPs coated by using PVA (poly(vinyl alcohol)) or poloxamer F68.^{46,47}

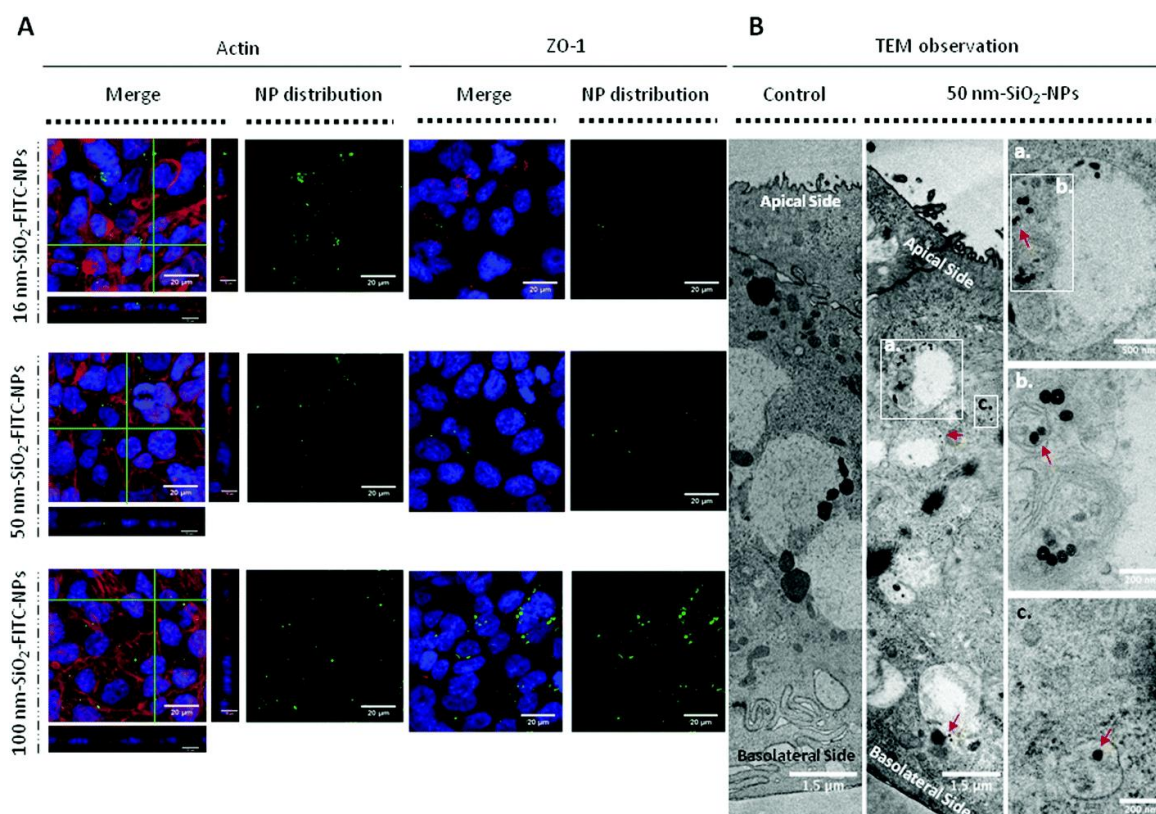


Fig. 2 Internalization of SiO₂-FITC-NPs inside the Calu-3 monolayer. (A) Cells were exposed for 24 h to 16, 50, and 100 nm SiO₂-FITC-NPs at 5 $\mu\text{g cm}^{-2}$. The (x,y) left images correspond to the z-projection of a stack of 5 to 10 images acquired by confocal microscopy. Orthogonal views represent (x,z)- and (y,z)-slices of the section indicated by the green lines. Staining of the cells is as follows: blue – DAPI-stained nuclei, green – SiO₂-FITC-NPs, red – phalloidin-stained actin filaments or ZO-1 protein labelled with Alexa-568-antirabbit antibody. The (x,y) right images corresponding to the z-projection of a stack of 5 to 10 images acquired by using only the green channel for a better observation of SiO₂-FITC-NP distribution inside the cellular monolayer. The scale bar corresponds to 20 μm . (B) Cells exposed, or not, for 24 h to 50 nm SiO₂-FITC-NPs at 5 $\mu\text{g cm}^{-2}$ were observed by TEM. The 50 nm SiO₂-FITC-NPs were found on the apical and basolateral side of the cell inside vesicles with a simple or double membrane. The scale bar corresponds to 200 nm and NPs are indicated by red arrows.

TEM observations revealed individualized 50 nm SiO₂-FITC-NPs at the apical and basolateral sides of the cell (Fig. 2B) whereas no similar signal was observed for non-treated cells control (Fig. 2B). Interestingly no NPs have been found inside cell nuclei in TEM observations in contrast to confocal microscopy results obtained in a previous study performed on human oral buccal mucosa cells treated with SiO₂-NPs.⁹⁴ NPs were visible inside large double membrane vesicles containing membrane or organelle fragments (Fig. 2B) that could be specific to a lysosome-based degradative pathway, the autophagy.⁴⁸ Recently several studies established that some NPs such as gold NPs,^{49,50} fullerenes^{51,52} or quantum dots^{53,54} could induce autophagy and thus NPs may become a novel class of autophagy activators.^{50,55} Immunolabelling with anti-LC3-II to specifically identify autophagosomes was performed on Calu-3 treated with 16 nm SiO₂-FITC-NPs at 5 $\mu\text{g cm}^{-2}$ for 24 h (Fig. 3). Cells exhibited a LC3-II-autophagosome accumulation after treatment compared to controls (data not shown) that could be either due to the increased production or blocking of the degradation of the autophagosome. Moreover Pearson coefficient (PC) calculation provided evidence of a co-localisation of NPs inside autophagosomes (Fig. 3). Although a low NP signal was detected, PC reflects the difference between the fluorescence level of NPs and LC3-II staining and could not be due to background noise.⁹³

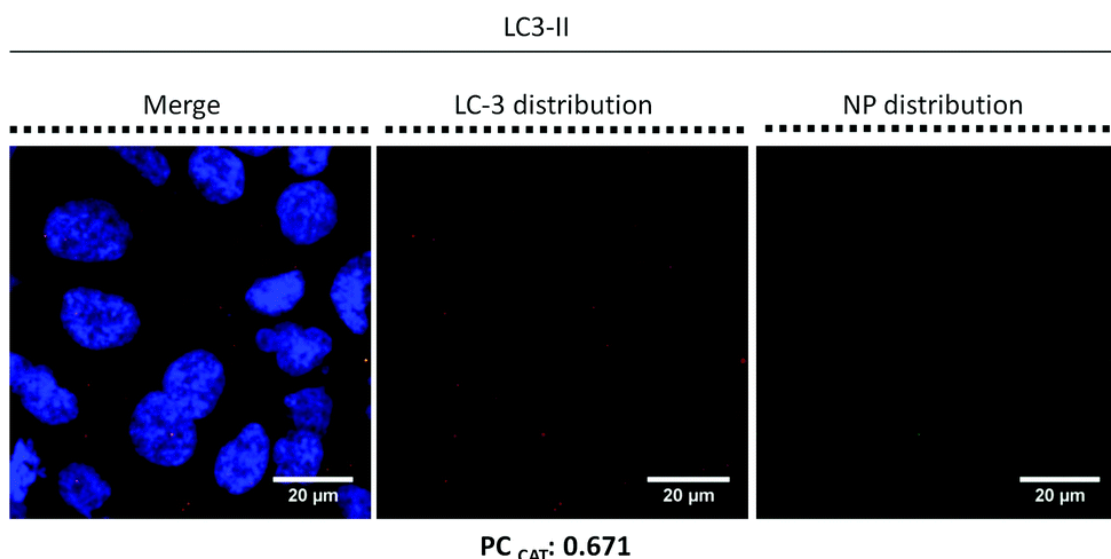


Fig. 3 Detection of autophagy inside NP-treated Calu-3 cells. Calu-3 cells were exposed to 16 nm SiO₂-FITC-NPs at 5 µg cm⁻² for 24 h. Confocal microscopy experiments were performed after cell fixation. The (x;y) left images corresponding to the z-projection of a stack of 5 to 10 images acquired. Staining of the cells is as follows: blue – DAPI-stained nuclei, green – 16 nm SiO₂-FITC-NPs, red – LC3-II proteins labelled with Alexa-568-antirabbit antibody. The (x;y) central images corresponding to the z-projection of a stack of 5 to 10 images acquired by using only the red channel for a better observation of LC3-II expression. The (x;y) right images corresponding to the z-projection of a stack of 5 to 10 images acquired by using only the green channel for a better observation of 16 nm SiO₂-FITC-NP distribution inside the cellular monolayer. The scale bar corresponds to 20 µm. Pearson coefficient (PC) was calculated by Image J software; PC (LC3-II) = 0.116. To improve PC a Costes' Automatic Threshold (PCCAT) was performed and new PC was obtained,⁹³ PCCAT (LC3-II) = 0.671.

NP translocation across the Calu-3 epithelium

To determine if NPs could translocate depending on their size, Calu-3 cells were treated for 24 h with 16, 50, 100 nm SiO₂-FITC-NPs at 5 and 10 µg cm⁻². By measuring fluorescence in the apical and basolateral compartments, NP concentrations were determined using a standard curve and compared to the initial applied concentration. NP translocation across TF in the absence of cells was firstly determined to ensure that 3 µm porosity filters were not a limitation to our experiments (ESI Fig. S5†).

Regardless of NP size, fluorescence can be detected inside the basolateral compartment, suggesting that a NP translocation occurred for all NPs (Fig. 4A). However the rate of NP translocation was lower after treatment at 10 µg cm⁻² than that after treatment at 5 µg cm⁻²; for instance 16 nm SiO₂-FITC-NP translocation was determined at 17.9% and 8.6% corresponding to 0.89 µg cm⁻² or 0.86 µg cm⁻² (respectively for initial concentration at 5 and 10 µg cm⁻²). NP translocation tended to be more important for the smaller NPs at low concentration. For example, after initial NP treatment at 5 µg cm⁻² we determined a NP translocation of 17.9%/14.4%/12.1% respectively for 16/50/100 nm SiO₂-FITC-NPs. Low levels of fluorescence were revealed in apical compartments, suggesting a low retention unlike the very high retention inside the cellular & filter compartment. While the NP exposure was generally limited to 24 h, we observed that NPs could be retained inside the cellular monolayer even 1 week after treatment (ESI Fig. S4B†).

Several studies underlined that one disadvantage of fluorescently labelled NPs could be the potential leakage of the fluorochrome from the NPs.^{56,57} To ascertain that the fluorescence measurement in the basolateral side was really related to NP translocation and not to a leakage of fluorescence, two control experiments were performed. Initial NP suspension, control basolateral media, and basolateral media recovered after exposure to 50 nm SiO₂-FITC-NPs were observed by TEM and analyzed by X-ray (Fig. 3B). TEM observation of initial and basolateral suspensions revealed the presence of 50 nm SiO₂-FITC-NPs

while no NPs were detected inside the control media. X-ray analysis attested the absence of silicon (but a calcium phosphate peak) in control media whereas a peak of SiO₂ was detected in initial and basolateral suspensions. These results provided evidence that NPs have translocated across the Calu-3 epithelium. Moreover these basolateral media were used to expose another cell line, NCI-H292 cells for 6 h showing fluorescent agglomerates not observed in cells exposed to free FITC (ESI Fig. S6[†]), confirming NP translocation.

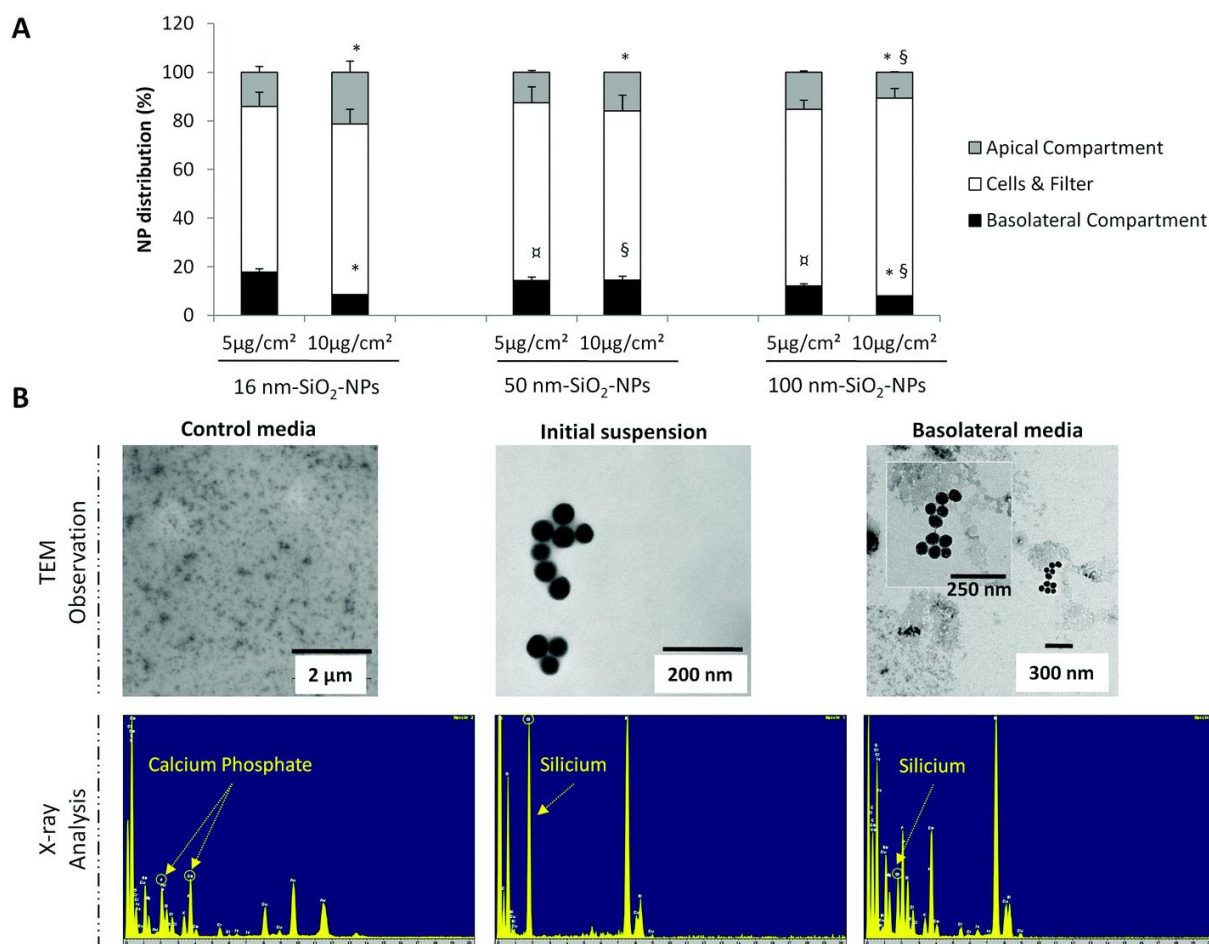


Fig. 4 NP translocation across the Calu-3 monolayer as a function of the size and the concentration. Calu-3 cells were treated for 24 h with 16, 50, and 100 nm SiO₂-FITC-NPs at 5 and 10 µg cm⁻². (A) Quantification of NP distribution in percentage inside the apical (grey bar), basolateral (black bar) and the cells & filter (white bar) compartments. Data were expressed as mean ± SEM, n = 4. *: different from 5 µg cm⁻² value (p < 0.05); ∓: different from 16 nm SiO₂-FITC-NPs value at 5 µg cm⁻² (p < 0.05); §: different from 16 nm SiO₂-FITC-NPs value at 10 µg cm⁻² (p < 0.05). (B) For 50 nm SiO₂-FITC-NPs: observation of basolateral media of control cell cultures, initial suspension and Calu-3 basolateral media after 24 h of exposure to 50 nm SiO₂-FITC-NPs by TEM, and the corresponding X-ray analysis.

NP translocation was dependent on NP surface charges

To study the impact of surface charge of the NPs on cellular uptake and translocation, SiO₂-FITC NPs were synthesized with a positive surface charge (SiO₂-FITC-NPs⁺) and compared with SiO₂-FITC-NPs. Confocal microscopy observations demonstrated that these positive NP were taken up without modifying actin filaments and ZO-1 distribution to tight junctions, as already observed for negatively charged SiO₂-FITC-NPs (Fig. 2A). SiO₂-FITC-NPs⁺ exhibited a heterogeneous distribution within the cellular monolayer that was more diffuse and seemed more important (Fig. 5A) than for NPs with a negative surface charge (Fig. 2A). Many in vivo studies provided evidence that positively charged NPs were absorbed more efficiently than negatively or neutrally charged NPs^{58,59} that led to a better blood clearance and a higher accumulation in the lung and the liver.⁶⁰ Enhancement by a positive surface charge of cellular uptake has

been demonstrated for selenium NPs,⁶¹ gold NPs,⁶² or iron NPs^{63,64} compared to their respective anionic NPs. Cellular uptake mechanisms were also governed by NP surface charge as reported in the overview presented by Fröhlich.⁶⁵

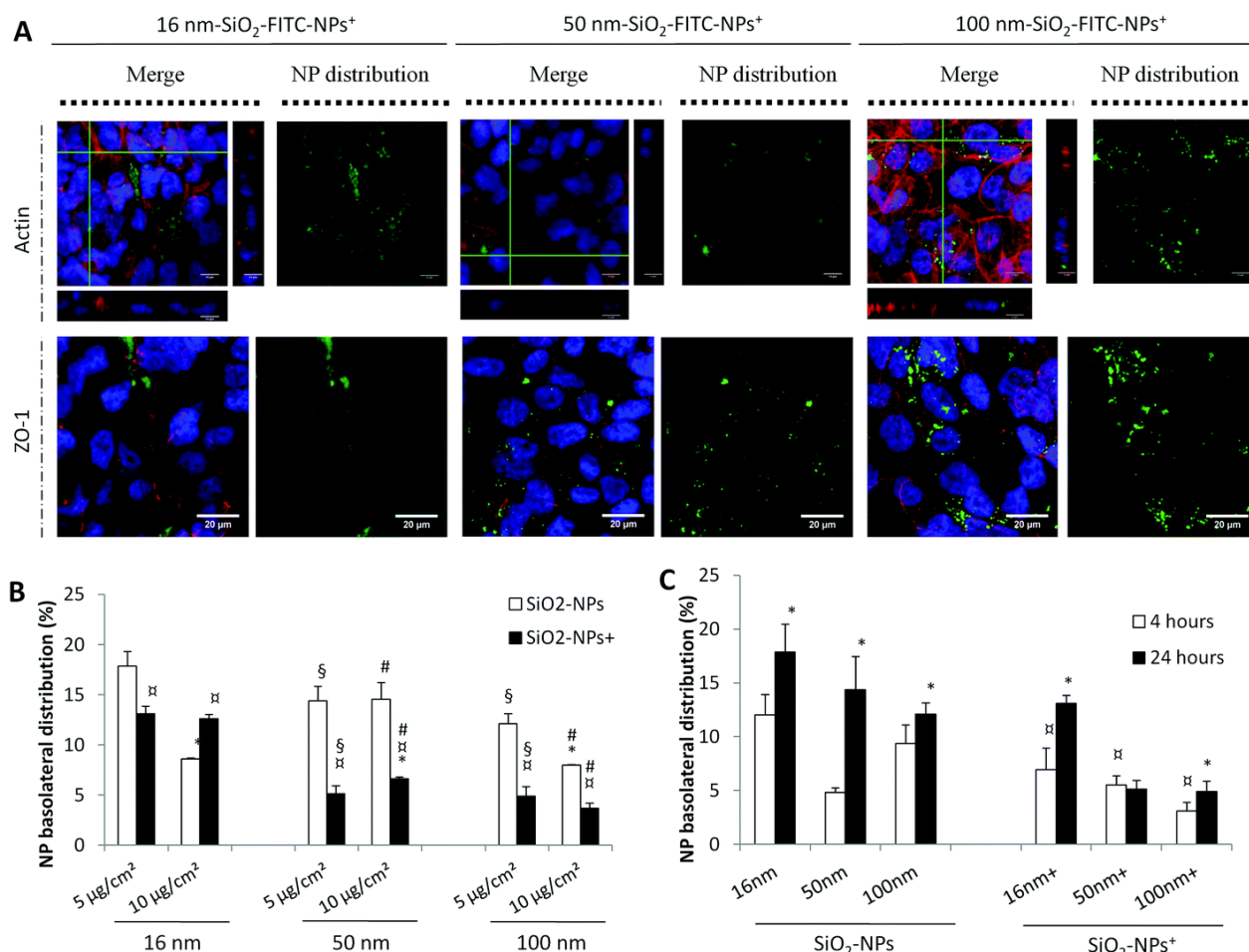


Fig. 5 Internalization of SiO₂-FITC-NPs⁺ by Calu-3 cells and their translocation across the monolayer in comparison with SiO₂-FITC-NPs. Calu-3 cells were exposed to 16, 50, and 100 nm SiO₂-FITC-NPs⁺ at 5 µg cm⁻² for 24 h before being rinsed and fixed for (A) confocal microscopy experiments. The (x,y) left images corresponding to the z-projection of a stack of 5 to 10 images acquired. Orthogonal views represent (x,z)- and (y,z)-slices of the section indicated with the green lines. Staining of the cells is as follows: blue – DAPI-stained nuclei, green – SiO₂-FITC-NPs⁺, red – phalloidin-stained actin filaments or ZO-1 protein labelled with Alexa-568-antirabbit antibody. The (x,y) right images corresponding to the z-projection of a stack of 5 to 10 images acquired by using only the green channel for a better observation of SiO₂-FITC-NP⁺ distribution inside the cellular monolayer. The scale bar corresponds to 20 µm. (B) Quantification of NP distribution in percentage inside the basolateral compartment as a function of the surface charge of the NP, in white SiO₂-FITC-NP and in black SiO₂-FITC-NP⁺. Data were expressed as mean ± SEM, n = 3. *: different from 5 µg cm⁻² value (p < 0.05); ‡: different from SiO₂-FITC-NPs (p < 0.05); §: different from 16 nm-SiO₂-FITC-NPs value at 5 µg cm⁻² (p < 0.05); #: different from 16 nm-SiO₂-FITC-NPs value at 10 µg cm⁻² (p < 0.05). (C) Calu-3 cells were exposed to 16, 50, and 100 nm SiO₂⁻ or SiO₂⁺-FITC-NPs at 5 µg cm⁻² for 4 h (white bar) or 24 h (black bar) before quantification of NP distribution in percentage inside the basolateral compartment as a function of the time of exposure. Data were expressed as mean ± SEM, n = 3. *: different from 4 h-treatment (p < 0.05); ‡: different from SiO₂-FITC-NPs value after 4 h of treatment (p < 0.05).

Comparison between SiO₂-FITC-NPs and SiO₂-FITC-NPs⁺ revealed that the translocation of NPs with a negative surface charge was the most important (Fig. 5B). The low translocation of SiO₂-FITC-NPs⁺ was likely not related to a low internalization. Flow cytometry analysis demonstrated that NPs were taken up by Calu-3 cells in the same amount (ESI Fig. S7†). These results were only based on NP internalization without taking into account NP adsorption onto the cellular membrane thanks to the addition of trypan blue during flow cytometry measurements. Moreover, a time-course study revealed that after 4 h of exposure translocation of SiO₂-FITC-NPs was already important whereas SiO₂-FITC-NPs⁺ translocation seemed delayed. The results also show that SiO₂ NPs crossed increasingly over time whatever the NP size

or surface charge (Fig. 5B). Quantification of SiO₂-FITC-NPs⁺ translocation across the Calu-3 epithelium revealed that 16 nm SiO₂-FITC-NPs⁺ translocated more easily than larger SiO₂-FITC-NPs⁺.

Besides the influence of NP size on NP translocation, several studies have underlined the fact that the negative or positive surface charge of the NP could modulate their translocation across the air–blood barrier.³ This could be explained by the modification of NP speed and extent of agglomeration due to the NP attraction or repulsion. NPs with the same surface charge repel each other, while neutral NPs tend more to agglomerate.⁶⁶ In vivo studies went further and elucidated NP passage across the lung barrier as a function of their surface charge. Quantification of translocation in rat models of NIR fluorescent NPs which varied in surface charge (zwitterionic, polar, anionic or cationic) revealed that negatively charged NPs could easily translocate.¹²

NP translocation was dependent on NP composition

In order to determine whether NP composition mattered, the translocation of 140 nm TiO₂-coated SiO₂-FITC-NPs was compared to 100 nm SiO₂ and SiO₂⁺-FITC-NPs. As for SiO₂⁺-FITC-NPs, TiO₂-coated SiO₂-FITC-NPs were shown to be internalized without altering the actin network and ZO-1 distribution to tight junctions and also exhibited heterogeneous distribution of large agglomerates (Fig. 6A).

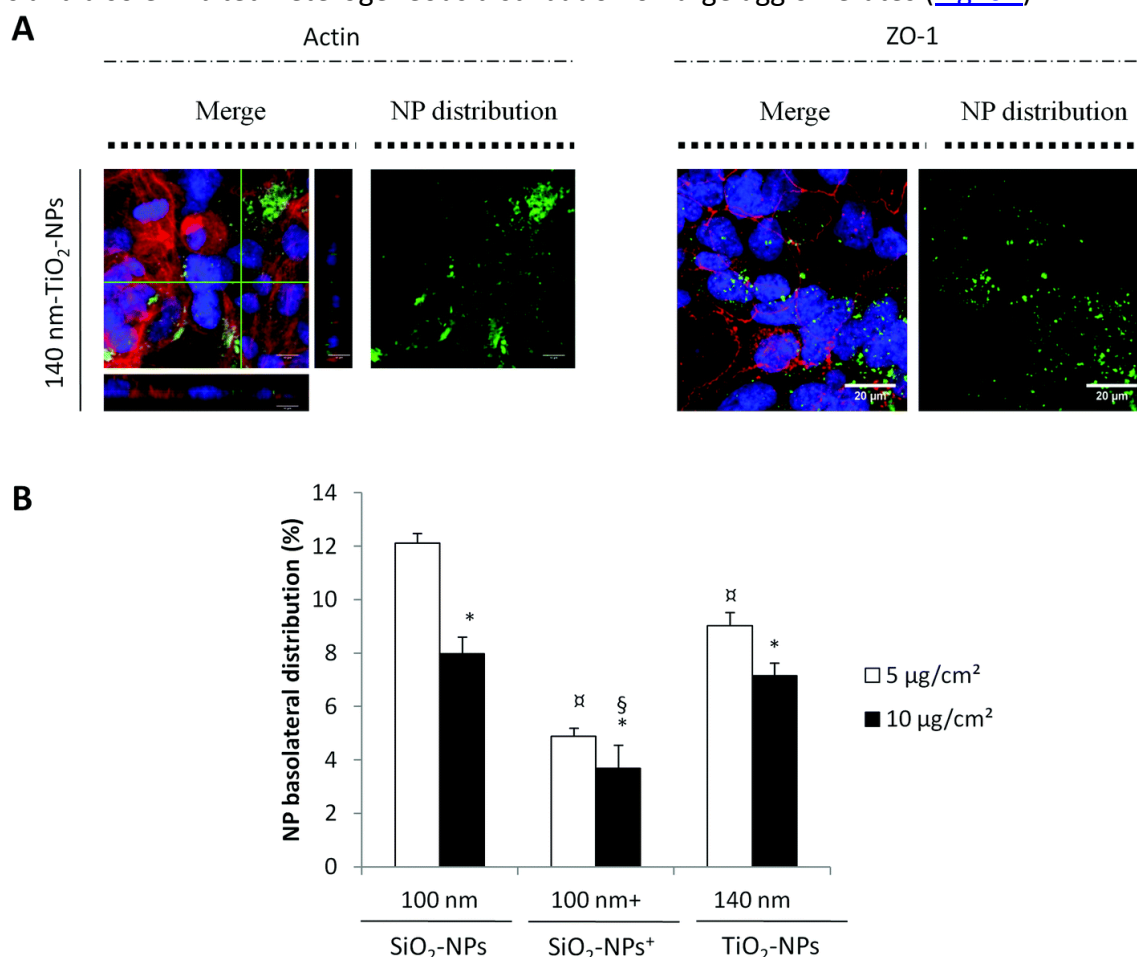


Fig. 6 Internalization and translocation of 140 nm TiO₂-coated SiO₂-FITC-NPs by Calu-3 cells. (A) Calu-3 cells were exposed to 140 nm TiO₂-coated SiO₂-FITC-NPs at 5 µg cm⁻² for 24 h before being rinsed and fixed for confocal microscopy observations. The (x,y) left images corresponding to the z-projection of a stack of 5 to 10 images acquired. Orthogonal views represent (x,z)- and (y,z)-slices of the section indicated with the green lines. Staining of the cells is as follows: blue – DAPI-stained nuclei, green – 140 nm-TiO₂-coated SiO₂-FITC-NPs, red – phalloidin-stained actin filaments or ZO-1 protein labelled with Alexa-568-antirabbit antibody. The (x,y) right images corresponding to the z-projection of 5 to 10 images acquired by using only the green channel for a better observation of 140 nm TiO₂-coated SiO₂-FITC-NP distribution inside the cellular monolayer. The scale bar corresponds to 20 µm. (B) Quantification of NP distribution in percentage inside the basolateral compartment for 140 nm TiO₂-coated SiO₂-FITC-NPs in comparison with SiO₂⁻ and SiO₂⁺-FITC-NPs at the same size, at 5 (white bar) and 10 µg cm⁻² (black bar). Data were expressed as mean ± SEM, n = 6. *: Different from 5 µg

cm^{-2} value ($p < 0.05$); ¤ : different from $\text{SiO}_2\text{-FITC-NPs}$ value at $5 \mu\text{g cm}^{-2}$ ($p < 0.05$); § : different from $\text{SiO}_2\text{-FITC-NPs}$ value at $10 \mu\text{g cm}^{-2}$ ($p < 0.05$).

Again we observed that the efficiency of NP translocation was higher for the lower initial NP concentration. 140 nm TiO_2 -coated $\text{SiO}_2\text{-FITC-NPs}$ translocated better than 100 nm $\text{SiO}_2\text{-FITC-NPs}^+$ and less than 100 nm $\text{SiO}_2\text{-FITC-NPs}$. This ranking is in accordance with the NP iso-electric point (IEP), $\text{IEP}_{(\text{SiO}_2)} < \text{IEP}_{(\text{TiO}_2\text{-coated SiO}_2)} < \text{IEP}_{(\text{SiO}_2^+)}$ (ESI Fig. S2A[†]), which could play a role in the NP retention inside the cell after internalization. Moreover, at physiological pH $\text{SiO}_2\text{-FITC-NPs}$ and TiO_2 -coated $\text{SiO}_2\text{-FITC-NPs}$ were both negatively charged but TiO_2 -coated $\text{SiO}_2\text{-FITC-NP}$ translocation was less important than those of $\text{SiO}_2\text{-FITC-NPs}$.

Quantification of translocation in rat models of NIR fluorescent NPs revealed no difference in translocation for either organic or inorganic NPs¹² of the same size and charge. In vitro studies performed on the intestinal barrier using the Caco-2 cell line seeded onto TF have demonstrated that NP translocation could be modulated by the NP composition. Indeed recent studies have shown that translocation of gold⁶⁷ or 12 nm anatase titanium NPs⁶⁸ across the gut epithelium varied as a function of their physico-chemical characteristics.

Protein and lipid corona increased translocation of NPs with a positive surface charge

NPs were rapidly covered with a corona when they interact with physiological fluids.²² This phenomenon could modulate NP–cell interactions and induce changes in cell responses.²¹

To investigate whether the presence of a protein corona could modify NP translocation, cells were treated with 16, 50, 100 nm SiO_2^- or $\text{SiO}_2\text{-FITC-NPs}^+$ at $5 \mu\text{g cm}^{-2}$ for 24 h in the presence or absence of FCS (2%). Confocal microscopy observation revealed no modification of the actin network or changes in ZO-1 distribution (data not shown). The uptake of 50 nm $\text{SiO}_2\text{-FITC-NPs}$ seemed reduced after FCS treatment and their distribution was more diffused inside the cellular monolayer (Fig. 7A) compared to cells treated with NPs in the absence of FCS. After FCS treatment, $\text{SiO}_2\text{-FITC-NP}$ translocation decreased while $\text{SiO}_2\text{-FITC-NPs}^+$ translocation increased (Fig. 7A). This was in contrast to the results obtained in the absence of the Calu-3 monolayer when we observed NP translocation across TF after co-exposure with FCS (ESI Fig. S5[†]). $\text{SiO}_2\text{-FITC-NP}$ translocation was enhanced by FCS by a factor of 3 whereas $\text{SiO}_2\text{-FITC-NP}^+$ translocation was not modified. Altogether these data suggest that the nature of the protein corona is quite different depending on the initial surface properties of NPs, which greatly contributes to modifying the fate of NPs.

Different proteins could form the corona such as plasma proteins^{69,70} or mucus proteins.²³ The NP interaction with secreted mucus of Calu-3 cells has shown to decrease the penetration of positively and neutrally charged PLGA-NPs, while PLGA-NPs coated with a hydrophilic compound (PF68) could diffuse more easily across the mucus barrier.⁴⁶ While protein corona formation is possible for any NPs, Lesniak et al. provided evidence that the formation of a protein corona around silica NPs could alter their level of uptake and the intracellular fate of NPs in A549 cells.²¹ Impacts were dependent on surface properties and NP size.⁷⁰ The uptake was also NP composition-dependent, and for instance the mechanisms involved were different for polystyrene spheres,⁷¹ gold NPs,⁷² silica NPs,⁷³ titanium NPs^{74,75} or carbon nanotubes.⁷⁶

To determine whether the presence of a lipid corona could modify NP translocation, cells were treated with 16, 50, 100 nm SiO_2^- or $\text{SiO}_2\text{-FITC-NPs}^+$ at $5 \mu\text{g cm}^{-2}$ for 24 h in the presence or absence of DPL (0.004%), a pulmonary synthetic surfactant phospholipid. Confocal microscopy observation revealed no modification of the actin network or changes in ZO-1 distribution (data not shown). We note the formation of large agglomerates of 50 nm $\text{SiO}_2\text{-FITC-NPs}$ within the Calu-3 epithelium in the presence of DPL (Fig. 7B). During co-treatment with DPL, $\text{SiO}_2\text{-FITC-NP}$ translocation was not significantly modified while $\text{SiO}_2\text{-FITC-NP}^+$ translocation was significantly increased.

FITC-NP⁺ translocation greatly increased (Fig. 7B). However these results were in contrast to those obtained in the absence of cells (ESI Fig. S5†). SiO₂-FITC-NP translocation was enhanced by DPL by a factor of 10 whereas DPL exposure did not modulate SiO₂-FITC-NP⁺ translocation.

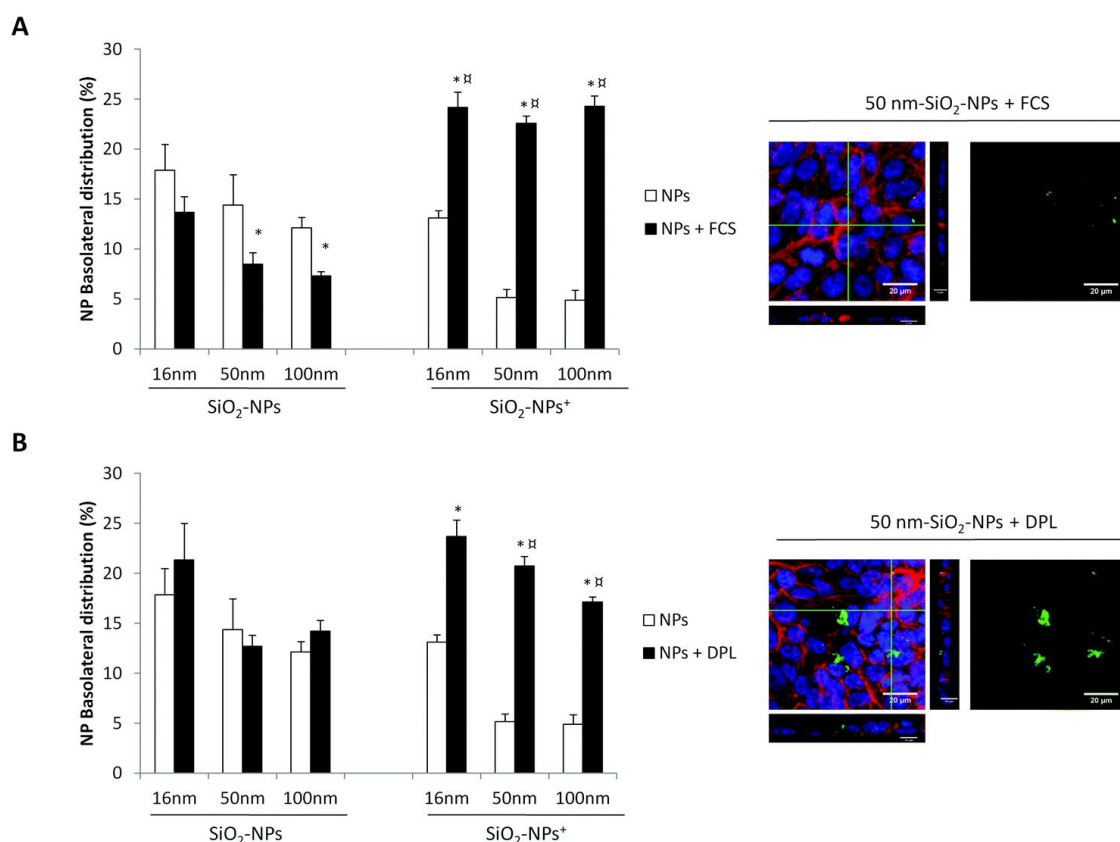


Fig. 7 Modulation of the NP translocation in the presence of FCS or DPL. Calu-3 cells were exposed simultaneously with (A) FCS at 2% or (B) DPL at 0.004% and with SiO₂⁻ or SiO₂⁺-FITC-NPs at 5 $\mu\text{g cm}^{-2}$ for 24 h. NP distribution in percentage was determined for the basolateral compartment as a function of the size and the surface charge of NPs, in white the NP translocation without additives, in black in the presence of FCS or DPL. Data were expressed as mean \pm SEM, $n = 6$. *: different from NP treatment value without modification of cellular environment ($p < 0.05$); \boxtimes : different from SiO₂-FITC-NPs value ($p < 0.05$). Cells were then fixed for confocal microscopy experiments. The (x;y) left images corresponding to the z-projection of 5 to 10 images acquired. Orthogonal views represent (x,z)- and (y,z)-slices of the section indicated with the green lines. Staining of the cells is as follows: blue – DAPI-stained nuclei, green – 50 nm-SiO₂-FITC-NPs, red – phalloidin-stained actin filaments. The (x;y) right images corresponding to the z-projection of 5 to 10 images acquired by using only the green channel for a better observation of 50 nm SiO₂-FITC-NP distribution inside the cellular monolayer. The scale bar corresponds to 20 μm .

It has been previously reported that NPs could interact with the pulmonary surfactant. These interactions were governed by NP physicochemical properties. Several studies using alveolar macrophages have shown an increase of NP uptake in the presence of surfactants²⁴ potentially mediated by the surfactant protein A.²⁷ The surfactant protein D could also play an important role in NP uptake because of its co-localization in A549 cells.²⁴ Moreover, the physiological functions of the lung surfactant could also be modified after NP exposure. A recent study has provided evidence that hydrophilic NPs could more easily translocate through the surfactant film compared to hydrophobic NPs.⁷⁸ It was also demonstrated that gold NP⁷⁹ or TiO₂-NPs⁸⁰ could impede the structure and function of the lung surfactant according to NP size or surface area.

Inflammatory context enhanced NP translocation

The lung is a site of frequent inflammations related to the exposure to particles, toxins or pathogens and can show chronic inflammation⁶⁶ in case of pulmonary diseases like asthma or COPD.

It is well established that the Calu-3 cell line expresses the TLR-4 (Toll like receptor-4)⁸¹ proteins involved in the pro-inflammatory response, in order to protect the airway epithelium from endotoxins. To investigate whether NP translocation could be modulated by pathological conditions, and more particularly by an inflammatory context, the Calu-3 monolayer was pre-treated for 4 h with TNF- α (1 ng mL⁻¹), a pro-inflammatory mediator, or LPS (25 ng mL⁻¹), an endotoxin known as a strong inducer of inflammation by the activation of TLR-4. Thereafter cells were exposed to 16, 50, 100 nm SiO₂ or SiO₂⁺-FITC-NPs or 140 nm TiO₂-coated SiO₂-FITC-NPs at 5 μ g cm⁻² for 24 h. TNF- α as well as LPS-exposure clearly induced a decrease of TEER (loss of 32% and 36% respectively), an increase of Lucifer Yellow passage (by a factor of 4 and 3 respectively) after the end of a 4 h exposure (Fig. 8A and 8C), and alterations of ZO-1 distribution (Fig. 8B) which was not exacerbated by NP treatment. These results underlined the abilities of TNF- α and LPS to alter tight junctions. It confirms the observations of Hermanns and colleagues showing the ability of TNF- α to open tight junction in an in vitro alveolar–capillary barrier.⁸²

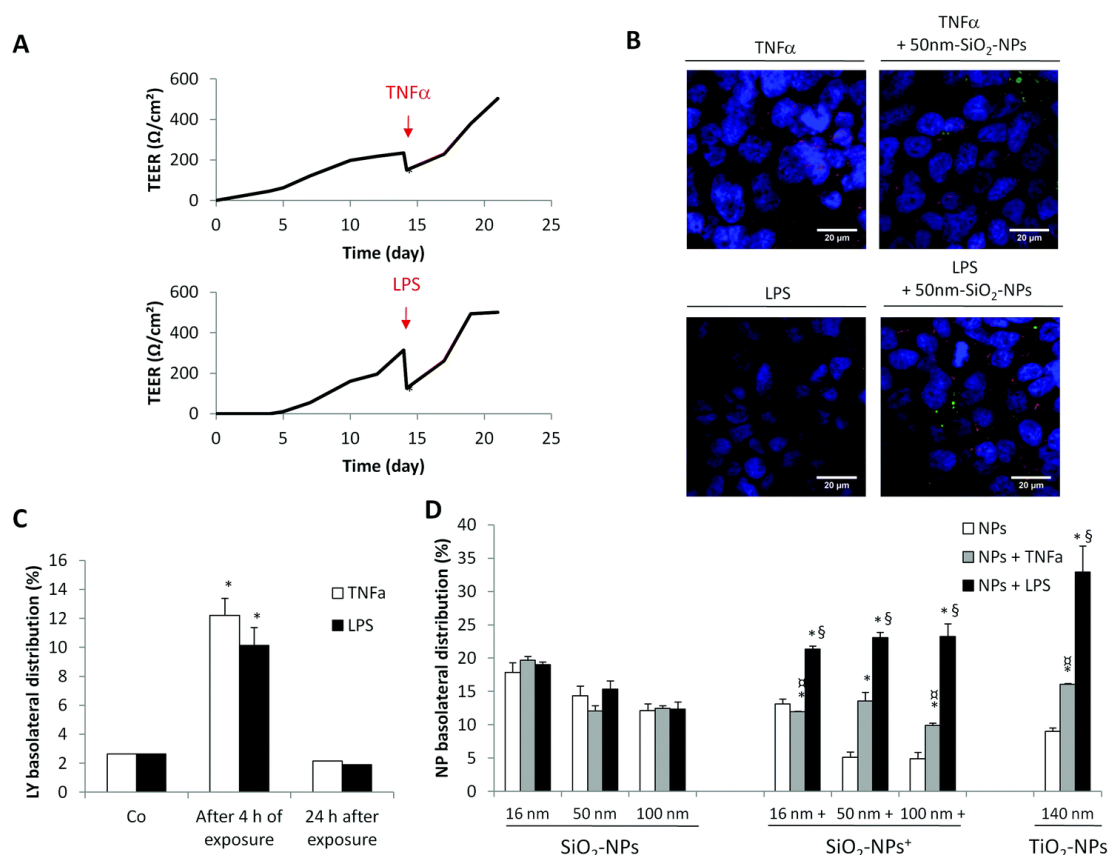


Fig. 8 Modulation of NP translocation in a pro-inflammatory context. Calu-3 cells were initially exposed during 4 h to TNF- α at 1 ng mL⁻¹, or LPS at 25 ng mL⁻¹, then rinsed and treated for 24 h with 16, 50, 100 nm SiO₂ or SiO₂⁺-FITC-NPs or 140 nm TiO₂-coated SiO₂-FITC-NPs at 5 μ g cm⁻². (A) TEER values were determined over time, before and after exposure to TNF- α or LPS. *: different from control ($p < 0.05$). (B) Cells were then fixed for confocal microscopy experiments. The (x;y) images corresponding to the z-projection of 5 to 10 images acquired. Staining of the cells is as follows: blue – DAPI-stained nuclei, green – SiO₂⁺-FITC-NPs of 50 nm, red – ZO-1 protein labelled with Alexa-568-antirabbit antibody. The scale bar corresponds to 20 μ m. (C) Quantification of Lucifer Yellow (LY) distribution was determined in percentage as a function of the pro-inflammatory treatment, without (white bar), with TNF- α (grey bar), or with LPS (black bar) just after pre-treatment, or after NP treatment. Data were expressed as mean \pm SEM, $n = 3$. *: different from control ($p < 0.05$). (D) NP distribution was determined in percentage as a function of the NP composition, in white the NP translocation in a normal case, in grey in the presence of TNF- α and in black in the presence of LPS. Data were expressed as mean \pm SEM, $n = 3$. *: different from NP treatment value in the absence of pro-inflammatory context ($p < 0.05$); ¶: different from SiO₂-FITC-NPs value in the presence of TNF- α ($p < 0.05$); §: different from SiO₂-FITC-NPs value in the presence of LPS ($p < 0.05$).

Quantification of the basolateral distribution of NPs after TNF- α pre-exposure has shown that 50, 100 nm SiO₂⁺ and 140 nm TiO₂-coated SiO₂-FITC-NP translocation were increased by a factor of 2.6/2/ and 1.8,

respectively. However, NP translocation was not modified for SiO₂-FITC-NPs and 16 nm SiO₂⁺-FITC-NPs. In addition, LPS pre-exposure improved 16, 50, 100 nm SiO₂⁺ and 140 nm TiO₂-coated SiO₂-FITC-NP translocation by a factor of 1.7/4.5/4.8/ and 3.7, respectively, but had no effect on SiO₂-FITC-NP translocation. These results allowed us to hypothesize that NP translocation could be enhanced by a pro-inflammatory context.

In vivo experiments have demonstrated that NP biodistribution was altered in the presence of LPS. Oropharyngeal aspiration of gold NPs has resulted in high retention of NPs in the lung treated with LPS, but in the absence of LPS, NPs were observed in the spleen.⁸³ Tight junction opening and pro-inflammatory response induced by LPS exposure could explain the fact that NPs have translocated in greater amounts compared to NP-treated Calu-3 cells without LPS-pretreatment.

Mechanisms of NP translocation

We demonstrated the translocation of the different NPs with an efficacy depending on their own properties. This raises the question of the mechanism involved in their passage. The observation of an internalization, the absence of cytotoxicity, and the stability of the TEER suggest a transcellular passage. In contrast, mimicking an inflammatory context alters the epithelial integrity and an increased translocation is observed for some NPs that could be related to a paracellular transport as the TEER is transiently reduced. To ascertain that the opening of tight junctions could increase NP translocation through the paracellular route, EGTA and CS (produced by deacetylation of chitin) were used for their ability to open tight junctions.^{5,6,84,85} Calu-3 cells were pre-exposed for 1 h to EGTA (20 mM)⁸⁶ and CS (0.5 mg mL⁻¹)⁸⁷ and then treated with 16, 50, 100 nm SiO₂-FITC-NPs at 5 µg cm⁻² for 24 h. NP translocation significantly increased after EGTA and CS treatment (Fig. 9A and 9B). For both compounds, NP translocation was modulated in a size-dependent manner. 16 nm-SiO₂-FITC-NP translocation was enhanced by 48% and 45%, 50 nm SiO₂-FITC-NPs NP translocation by 33% and 18%, 100 nm SiO₂-FITC-NPs NP translocation by 72% and 69% by EGTA and CS pre-exposure, respectively. NP translocation was more important for larger NPs. Altogether these results demonstrate that the translocation of SiO₂-FITC-NPs can occur by the paracellular route. A previous study provided evidence that NPs could pass through the cellular barrier. They demonstrate that TiO₂-NPs could interact with the junctional VE-cadherin protein of endothelial cells by physical interactions.⁸⁸

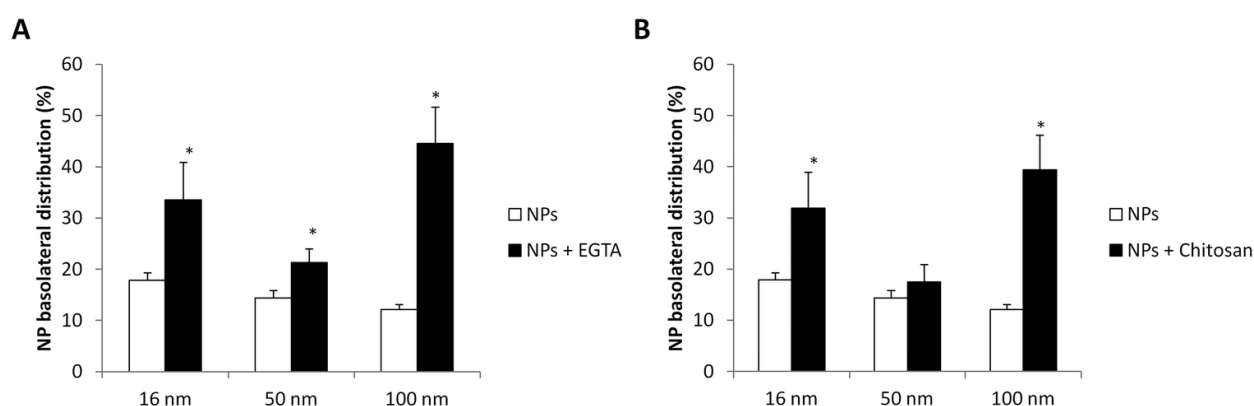


Fig. 9 Modulation of NP translocation after treatment with EGTA or chitosan. Calu-3 cells were exposed to 16, 50, 100 nm SiO₂-FITC-NPs at 5 µg cm⁻² for 24 h. Quantification of NP translocation after 1 h of pre-exposure to (A) EGTA at 20 mM or (B) chitosan at 0.5 mg mL⁻¹. Data were expressed as mean ± SEM, n = 3. *: different from NP treatment value without opening tight junctions (p < 0.05).

Interestingly NP translocation was more important in the presence of the Calu-3 monolayer compared to NP translocation across the TF in the absence of cells. These data confirm that SiO₂-FITC-NP passage to the basolateral side was facilitated by the modification of the NP surface during cellular interactions. The

mechanism involved in the transcytosis is still an open and unsolved question and the role of the cytoskeleton has to be addressed as it has been shown that NPs can bind and disturb the microtubule cytoskeleton.⁹⁵ We consider that the intracellular trafficking of internalized NPs is likely different according to the NP type as we have seen for positive SiO₂-FITC-NPs showing a similar amount of NP internalized but with a less effective mechanism of translocation.

Conclusion

In conclusion our results have shown that NPs can translocate through the Calu-3 monolayer whatever their composition (SiO₂ or TiO₂), but this translocation is increased for the smallest and negatively charged NPs. The translocation is not associated with an alteration of the integrity of the epithelial monolayer, suggesting a transcytosis of the internalized NP. By modifying the NP corona, the ability of NPs to cross the epithelial barrier differs depending on their intrinsic properties, making positively charged NPs more prone to translocate. NP translocation can be amplified by using agents known to open tight junctions and to allow paracellular passage. The translocation of NPs was also modulated when mimicking an inflammatory context, frequently found in the lungs, that alters the epithelial integrity and induces transient tight junction opening. These results held promise for supplying PBPK models with NP absorption data.

Methods

Nanoparticle synthesis

Size-controlled fluorescent silica nanoparticle (NP) production. SiO₂-NPs of three size classes, 16, 50 and 100 nm were produced and covalently fluorescently labelled for this study by incorporation of fluorescein into the silica NP according to a procedure modified from van Blaaderen et al.⁸⁹ These NPs were named SiO₂-FITC-NPs. For more details please refer to ESI (ESI Fig. S1A[†]).

Chemical surface modification of SiO₂-FITC-NPs by EDPS. Surface modifications of SiO₂-FITC-NPs were carried out by adding in the medium an excess of 10 equivalents of N-(trimethoxysilylpropyl)ethylenediamine (EDPS), sufficient to provide 2–3 monolayer coatings of the SiO₂-FITC-NPs. These NPs were named SiO₂-FITC-NPs⁺. For more details please refer to ESI (ESI Fig. S1A[†]).

Production of TiO₂-coated fluorescent silica nanoparticles. This TiO₂ coating was performed according to a modified procedure from Imhof used to coat a TiO₂ layer on poly(styrene) nanospheres.^{90,91} These NPs were named TiO₂-coated SiO₂-FITC-NPs. For more details please refer to ESI (ESI Fig. S1A[†]). Initial NP suspensions were observed by TEM (Tecnai 12 microscope) at 80 kV with a CCD camera (SiS 1k × 1k Keen View), by drop depositing and drying on a TEM grid (ESI Fig. S1A[†]).

NP characterization

NP suspensions in DMEM/F-12 medium diluted at 5 µg cm⁻² (10 µg mL⁻¹) were characterized by DLS in intensity and zeta potential measurements (Zetasizer, nano ZS, Malvern Instruments, USA). Additional NP characterization methods are described in ESI.[†]

Cell culture

Human epithelial cell lines. Human lung adenocarcinoma cells (Calu-3) were purchased from the American Type Culture Collection (ATCC, Sigma-Aldrich) and grown in DMEM/F12 (Dulbecco's modified Eagle's medium) culture medium with phenol red (Life Technologies, Saint Aubin, France), containing 1% GlutaMAX (Life Technologies) and 10% foetal calf serum (FCS, Life Technologies), 1% penicillin–streptomycin (PS, Life Technologies), and 0.5% amphotericin B (Life Technologies), subsequently referred

to as complete cell culture medium. All experiments were performed with these cells from passages 25 to 33. Cells were grown in T75-flasks (Costar, Corning, New York, USA) for cellular expansion and seeded at 500 000 cells cm^{-2} ([ref. 28,30,35,92](#)) onto 3 μm pore size polycarbonate Transwell Filters (TF, Costar) in two compartment chambers (12 mm in diameter) for translocation experiments. 500 μL of complete culture medium were added in the apical chamber and 1500 μL in the basolateral chamber.

Calu-3 translocation Studies

Without modification of cellular environment. After 14 days of culture, TF-confluent Calu-3 cultures were rinsed with PBS to eliminate trace amounts of FCS. NP stock solutions were shortly vortexed before making final dilutions at 5 and 10 $\mu\text{g cm}^{-2}$ (corresponding respectively to 10 $\mu\text{g mL}^{-1}$ and 20 $\mu\text{g mL}^{-1}$) in cell culture medium without phenol red, GlutaMAX or FCS. Cells were apically treated with NPs for 4 or 24 h. After treatments, apical and basolateral media were removed and kept for fluorescence measurements, Transmission Electron Microscopy (TEM) or Enzyme-Linked ImmunoSorbent Assay (ELISA). Cells were rinsed two times with PBS and fixed for TEM and confocal microscopy observations.

With alterations of cellular environment. Calu-3 cells were treated with NPs \pm FCS (Life Technologies) or NPs \pm DPL (Sigma Aldrich) respectively at 2% and 0.004% final in culture medium for 24 h. Pro-inflammatory context was induced by a 4 h exposure to 2 different pro-inflammatory mediators such as Tumor Necrosis Factor alpha (TNF- α , 1 ng mL^{-1} final in culture medium, R&D Systems, Abington, UK), or a strong inducer of inflammation, the lipopolysaccharide (LPS, 25 ng mL^{-1} final in culture medium, Enzo, Life Science, Villeurbanne, France). In some experiments the epithelial monolayer was altered by a 1 h pre-treatment with Ethylene Glycol Tetraacetic Acid (EGTA) at 20 mM, a Ca^{2+} chelator inducing tight junction opening, or chitosan at 0.5 mg mL^{-1} known to open tight junctions. After all pre-treatments, mediators were removed and cells were rinsed two times with PBS and then treated for 24 h by NPs. After NP treatments apical and basolateral media were removed, cells were rinsed two times with PBS and fixed for confocal microscopy experiments. To ascertain tight junction permeability the LY passage was measured, and cells were rinsed with PBS (Life Technologies, supplemented with Ca^{2+} and Mg^{2+}) and apically incubated for 1 h at 37 °C with 500 μL of LY (0.2 $\mu\text{g mL}^{-1}$ final in PBS) before quantification of LY in the apical and basal compartment by spectrophotometric analysis.^{[30](#)}

Quantification of NP translocation. Serial dilutions of NPs were performed (in a concentration range from 0 to 10 $\mu\text{g cm}^{-2}$) in culture media to establish a standard curve. After 24 h of treatment, 100 μL of each apical and basolateral media as well as the serial NP dilutions were deposited inside a white 96-well plate with a clear bottom (Greiner, Courtaboeuf, France). FITC fluorescence (488 nm/521 nm) coupled to NPs was quantified using FluoStar Galaxy (BMG, Ortenberg, Germany). The fluorescence background value (medium without NPs) was subtracted from sample fluorescence. Fluorescence values were converted in final NP concentrations in each compartment by using a standard curve, and then in percentage of the initial NP concentration. For the “filter” and “cells/filter” compartments, values were determined by subtraction of the apical and basolateral media from the initial NP concentration applied.

Immunolabelling and confocal microscopy observations

After NP treatment cells were fixed using 4% paraformaldehyde in PBS (Santa-Cruz, Heidelberg, Germany) over 20 min at RT, then incubated with NH_4Cl (50 mM, Sigma-Aldrich) for 10 min and permeabilized in 0.05% Tween 20 (Sigma-Aldrich) in PBS for 5 min. Fixed cells were incubated overnight at 4 °C with human microtubule-associated protein 1A/1B-light chain 3 (LC3-II) polyclonal primary antibody (Abcam, Paris, France) diluted to 1 : 200 produced in rabbit; human Claudin-4 (CLD-4) polyclonal primary antibodies (Santa Cruz Biotechnology, Heidelberg, Germany) diluted to 1 : 100 produced in goat; human MUC5AC monoclonal primary antibodies (Life Technologies) diluted to 1 : 100 produced in mouse or with human ZO-1 polyclonal primary antibody (Sigma Aldrich) diluted to 1 : 500 produced in rabbit. This incubation was followed by incubation with secondary antibodies stained with Alexa 488 or 568 fluorochrome diluted

to 1 : 500 (Life Technologies) for 2 h at RT. For actin filaments staining, fixed cells were incubated for 40 min with rhodamine–phalloidin (final concentration at 0.9 nM in PBS, Life Technologies). Cell nuclei were stained for 2 min with DAPI (4',6-diamidino-2-phenylindole dihydrochloride, final concentration at 0.25 $\mu\text{g mL}^{-1}$ in PBS, Life Technologies). Cells were examined under a Zeiss LSM710 confocal microscope using an objective 63 \times with a 1.5 zoom after mounting in polyvinyl alcohol mounting medium with DABCO (Sigma-Aldrich). Considering optical laws the theoretical resolution was calculated and instrument settings adapted to obtain the best possible resolution in our images. Image treatment and co-localisation study were done with Image J software (Image J 1.42 NIH, USA) by the addition of LOCI and Jacol plugins respectively. Imaris software (Imaris 7.6, Bitplane, Zurich, Switzerland) was used for the three-dimensional (3D) visualisation. Z-stack merges were obtained with the merging of 5 to 10 z-stacks with a stack-spread of 0.45 μm .

Transmission electron microscopy

Calu-3 cell preparation. Cells were fixed using 1.6% (v/v) glutaraldehyde and 1% (v/v) tannic acid in phosphate buffer 0.1 M, pH 7.4 for 1 h and post-fixed using 1% (v/v) osmium tetroxide for 60 min. The membranes were then stripped from the insert, dehydrated in ascending series of ethanol dilutions and impregnated in ascending dilutions of resin in ethanol, left in pure resin overnight (Epon, Inland Europe), embedded and polymerized at 60 °C for 48 h. Ultra-thin sections (70 nm) performed on ultramicrotome (RMC, PowerTome PC) were collected on Butvar-coated single-slot copper grids, and stained with 2% (v/v) uranyl acetate for 30 min and with lead citrate for 2 min. For characterization of the basal medium, solution at 40 $\mu\text{g mL}^{-1}$ was deposited on a carbon coated grid previously submitted to a glow discharge (Elmo, Cordouan Technologies). Grids were examined by TEM (CM120, FEI) and the images were acquired using a digital 2k \times 2k Gatan camera.

X-Ray analysis. With a view to checking the size and chemical nature of NP, to assess the effectiveness of translocation and the NP agglomeration, several observations and chemical analyses were performed with a Transmission Electron Microscope (TEM, JEOL JEM 1400 ST) fitted with an Energy Dispersive X-Ray Spectrometer (EDS, Oxford Inca Energy TEM 350-EDS 7215) and with a digital camera (Gatan Erlangshen ES 500W-782). Observations were performed at a magnification between $\times 15\,000$ and $\times 80\,000$ at 120 kV, and X-ray data were analyzed with INCA software.

Statistical analysis

Every experiment was repeated at least twice with triplicates for each condition. Data are represented as means \pm SEM and are analyzed with SigmaStat commercially available software (Version 3.0, Systat software Inc., San Jose, California, USA) using analysis of variance (one-way ANOVA) followed by Dunnett's test for multiple comparisons with $p < 0.05$ (two tailed) considered as significant.

Conflict of interest: The authors declare no competing financial interest.

Author contribution: IG contributed to the study design, did the experimental work, analyzed data and wrote the manuscript. GN prepared Calu-3 cells for TEM experiments and acquired TEM pictures. VC helped in the co-localization study and did 3D reconstructions of confocal microscopic images. SM performed the synthesis of the different batches of NPs. KB and LM performed X-ray analysis of NP suspensions. SB and OL critically reviewed the manuscript and gave intellectual input. ABS contributed to the study design, analyzed data, coordinated the work and wrote the manuscript. All the authors have read and approved the final manuscript.

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