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Seasonal Variations in Maternal Provisioning of *Crepidula fornicata* (Gastropoda): Fatty Acid Composition of Females, Embryos and Larvae

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Abstract

Recruitment success of marine invertebrate populations not only depends on the number of recruits but also on their quality which affects their survival. In species characterized by a mixed development (encapsulated embryonic development and release of planktotrophic larvae), the offspring quality depends on both maternal provisioning and larval feeding. Here, we investigated potential changes of maternal provisioning over the whole reproductive period in a gastropod with a mixed development: *Crepidula fornicata*. In its introduction area, *C. fornicata* reproduces from February to October, which implies that both adults and larvae are exposed to different food availabilities. Maternal provisioning was assessed by measuring the fatty acid (FA) composition of females, encapsulated embryos and larvae, in February, May, July and September 2009. FA are essential resources for the development of embryos and larvae, and are key biomarkers of offspring quality. Our results showed differences in FA composition between muscles, visceral masses, and encapsulated embryos. In particular, FA composition of embryos was similar to that of the visceral mass. Seasonal variations in FA composition were observed: in the middle of the reproductive season (May and July), female tissues and embryos showed a higher proportion of polyunsaturated fatty acids and especially $\omega 3$, as compared to the beginning and end of the reproductive season (February and September). This showed that through maternal provisioning the quality of *C. fornicata* offspring was higher in the middle of the reproductive season. Whether this would result in an increase of recruitment success and juvenile performance would require further investigations.

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Introduction

In most marine invertebrates, larval life history traits (developmental mode, growth, survival...) are intimately linked to the egg size and nutrient content provided by the mother (e.g. [1–4]). This is not only true for species with lecithotrophic development which mostly rely on maternally-provided resources and do not need to feed, but also for species producing planktotrophic larvae which need to feed in order to complete their development. Whereas lecithotrophic eggs have a greater weight-specific energy content to achieve complete development [5], in planktotrophs, offspring performance depends on both endogenous reserves derived from the parents and exogenous resources provided through larval nutrition [4,6–8]. Assessing the relative contribution of these two sources is of primary importance for the understanding of larval

development and juvenile performance. In some species, parental provisioning has been shown to influence larval growth and survival (e.g. [1,6]), being in some cases even more important than larval feeding [8], whereas in some others increase in maternal investment in eggs has a positive effect on larval size but not on larval development at natural food concentrations [9]. Maternal supply to embryos is influenced by numerous factors: mother body size, maternal feeding, and maternal habitat which includes food availability for parents [1,2,6,9–12]. This latter factor is crucial for species with a long reproductive period, with both parents and offspring facing seasonal variations of the environmental conditions, including food abundance and composition. In these species, the offspring condition may thus vary seasonally (e.g. [13]), with potential consequences on dispersal abilities and juvenile performance.

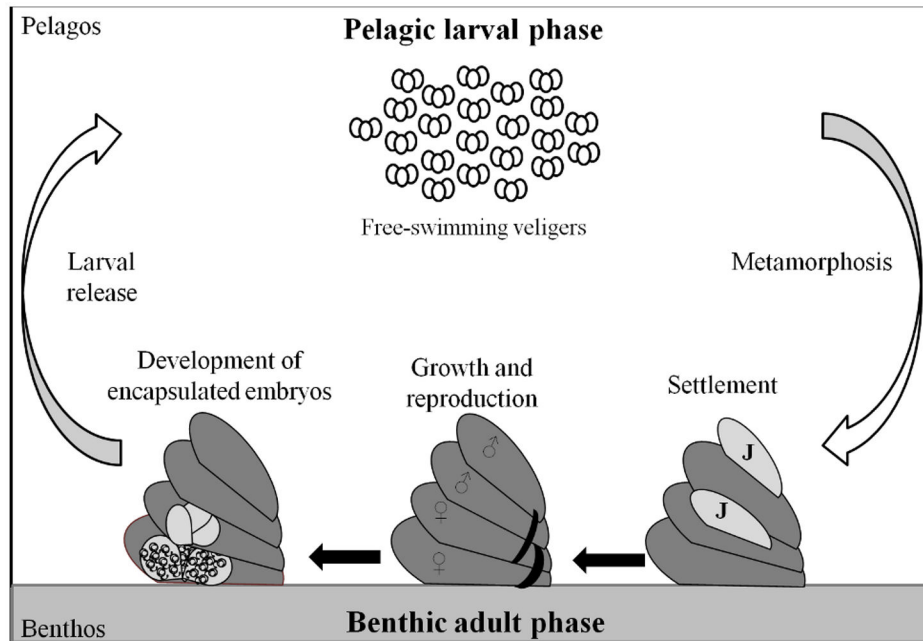


Figure 1. Schematic of the benthopelagic life cycle of *Crepidula fornicata*.

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Maternal investment is not only important in terms of quantity, but the biochemical composition of nutrients provided by females is also fundamental for the condition of offspring [14]. In particular, lipids are the major metabolic energy reserve in most marine animals, including marine invertebrate larvae [15]. They play a central role in the embryo development as an energy resource [15–17] and influence the hatching rate, the larval survival and the settlement success of many invertebrate species [18,19]. Fatty acids (FA) are the fundamental structural components of almost all lipid forms [20]. They are known to play a major role in embryogenesis and larval development, especially the polyunsaturated fatty acids (PUFA). In particular, PUFA with a terminal end omega-3 (ω 3) are mostly a source of energy (with the exception of the docosahexaenoic acid (DHA; 22:6 ω 3), which has a role in membrane integrity), and omega-6 (ω 6) PUFA are structural components of cell membranes. The PUFA EPA (20:5 ω 3; eicosapentaenoic acid) and DHA were shown to increase hatching success and larval survival in bivalves [21–24] and gastropods [25]. Therefore, FA composition of eggs and larval tissues may be used to assess the offspring condition. Besides, FA composition of eggs and larvae depends on both parental and larval nutrition [26] and may be considered as trophic biomarkers allowing the identification of assimilated food sources (e.g. [20,27–30]).

In this paper, we investigated the seasonal variations in maternal investment in embryos of the gastropod *Crepidula fornicata* Linnaeus 1758 (Calyptreaeidae) and its consequences on the quality of released larvae. *C. fornicata* broods its embryos for about one month within thin-walled capsules located between the neck and the propodium of the mother, before releasing them as planktotrophic larvae [31,32] (Figure 1). Each female broods between 25 and 75 capsules, each

containing 300 to 500 embryos, all at the same developmental stage [33,34]. To our knowledge, in *C. fornicata* there is no potential for nutritive exchange between females and their associated broods, and encapsulated embryos have a limited access to external dissolved organic matter [35], so encapsulated embryos mainly or even exclusively rely on resources provided by the mother. Resources mostly consist of yolk [16] and intracapsular albumen [33], investment in the form of extra-embryonic protection (e.g. egg capsules) being low (2–8% of total energy supply in closely-related calyptraeid species [36,37]). Finally, nurse eggs have not been observed in *C. fornicata* [38] although cannibalism on damaged eggs or embryos has been reported [39].

Native from the East Coast of North America, *C. fornicata* has been introduced in Europe where it has become invasive in many bays and estuaries [40]. In its native and introduced areas, *C. fornicata* has a long reproductive period. For example, in the North Coast of France, *C. fornicata* reproduces from February to October and larvae are frequently observed from March to November [41–43]. This suggests that resource allocation to offspring may vary along the reproductive period, which is especially important in the context of the introduction of this species, reproduction success being a key feature for successful invasion.

Seasonal variations in maternal investment in embryos of *Crepidula fornicata* were investigated by studying the variations of the FA composition of females, encapsulated embryos at four developmental stages, and pelagic larvae during one reproductive season.

Materials and Methods

Sample collection

Stacks of *Crepidula fornicata* were collected with a grab in the bay of Morlaix (48°40' N, 3°53' W; Fig. 2), Brittany, France. In this species, the reproductive period extends from February to October [41–43]. To cover the whole period, sampling was performed at four occasions in 2009: 26th February, 5th May, 2nd July and 18th September. Individuals were starved in 2-liter tanks filled with filtered seawater (0.45 µm) during 24 hours before tissue and brood sampling, in order to empty the gut. The water was changed three times during this period. Stacks were then opened in order to collect the broods located between the neck and the propodium of the females. Without prior knowledge of changes that might occur in FA composition during embryo development, we first aimed to compare the FA composition of early embryos. Second, because evidencing potential seasonal differences in the FA composition of early embryos would not necessarily imply seasonal differences in the FA composition of released larvae, we investigated the potential changes occurring in subsequent developmental stages, at each sampling period. Using a dissecting microscope four developmental stages were identified according to Chipperfield [44]: early embryo (I; one cell to gastrula; orange to yellow), trochophore (II; ciliated embryo; light yellow), early veliger (III; developing velum, well-formed shell, dark color due to the presence of yolk), and ready-to-hatch veliger (IV; prior to hatching; well-formed shell, well-developed and strongly ciliated velum, translucent color due to the lack of reserves). Embryos were excapsulated and filtered on pre-combusted GF/F filters. Females corresponding to each brood were dissected in two parts: foot (muscle), and visceral mass (which contains viscera and gonad). All samples were freeze-dried and stored at -20°C until fatty acids extraction.

The number of replicates for each developmental stage depended on their availability at time of sampling. Five females were collected in February with their associated broods (all at stage I); twelve were collected both in May and July, with three replicates for each developmental stage; eight females with their associated broods were collected in September with three replicates for early veliger stage, two replicates for early embryo and ready-to-hatch veliger, and one replicate for trochophore. To compare the fatty acid composition of broods at the four sampling occasions, only the early embryos (stage I) were used. Fatty acid contents of the three other developmental stages were then evaluated in May, July and September.

Pelagic larvae of *C. fornicata* were collected monthly by horizontal plankton tows (WP2 net, 200 µm mesh size [45]) at the same site from May to October 2009 (six samples: 12th May; 4th June; 1st July; 31st July; 28th August; 6th October). About 300 larvae were identified following Werner [46] and Thiriou-Quiévreux and Scheltema [47], sorted alive and filtered on pre-combusted GF/F filters. In addition, surface and bottom seawater was sampled with a Niskin bottle to collect particulate organic matter (POM). For this purpose, 3 to 4 L of seawater were pre-filtered on 20 and 200 µm mesh size and filtered on

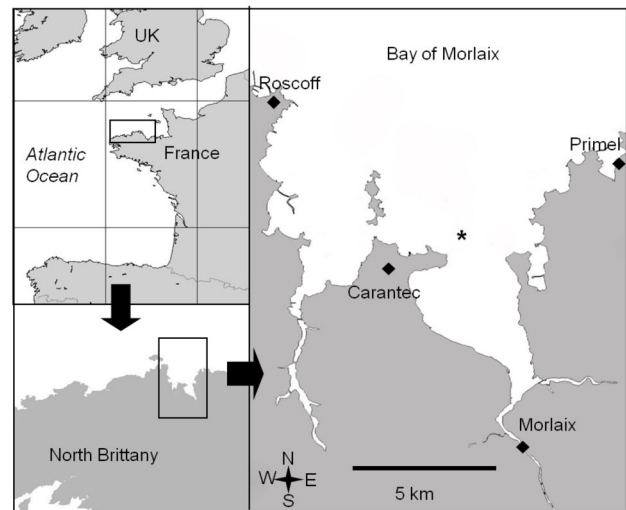


Figure 2. The sampling site. Map showing the location of the sampling site (black cross) in the bay of Morlaix (48°40' N, 3°53' W), North Brittany, France.

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pre-combusted GF/F filters. All filters were freeze-dried and stored at -20°C until FA extraction.

No specific permissions were required for sampling at the selected location, as it is not privately-owned or protected. Field sampling did not involve endangered or protected species.

Fatty acids extraction and analysis

The lipids from female tissues, excapsulated embryos and larvae of *Crepidula fornicata* were extracted following the method of Bligh and Dyer [48] modified by Meziane et al. [30]. Briefly, lipids were extracted by homogenization for two minutes followed by ultrasonication for 20 minutes with a mixture of distilled water: methanol: chloroform (1:2:1, v:v:v). The addition of distilled water: chloroform mixture (1:1, v:v) formed an aqueous-organic two-layer system enhanced by centrifugation (3000 rpm, 5 min). The lower chloroform phase, comprising the lipids, was retained and concentrated by rotary evaporation. The lipid extracts were fully dried under nitrogen and then saponified under reflux (1h30, 80°C) with a 2 M NaOH solution in methanol and distilled water (2:1,v:v). Saponification and methylation were carried out according to Meziane and Tsuchiya [49] in order to collect total FA. After this extraction step, FAME (Fatty Acid Methyl Esters) were separated and quantified by gas chromatography (Varian, 3800) equipped with a flame ionization detector. In order to allow the calculation of fatty acids concentration after analysis by gas chromatography, a defined amount of an internal standard (23: 0 FA) was added into the samples before extraction. Hydrogen was used as a carrier gas. At the injection time, the oven temperature was 60°C. It was raised to 150°C at 40°C min⁻¹, and remained at this temperature for three minutes. Then the temperature increased at 240°C at the rate of 3°C

min⁻¹ and remained constant again for three minutes. Most FAME peaks were identified by comparing their retention times with those of standards (Supelco). When needed, some peaks of FA were identified with gas chromatography coupled with mass spectrometry (ThermoFinnigan TRACE DSQ GC-MS).

Fatty acids were designed as X:Y ω Z, where X is the number of carbon atoms, Y is the number of double bonds and Z is the position of the ultimate double bond from the terminal methyl group.

The concentration of each FA (C_{FA}) was calculated according to Schomburg [50]:

$$C_{FA} = (A_S / A_{IS}) \times (C_{IS} / W_S)$$

Where A_S was the peak area of a given fatty acid on the chromatogram, A_{IS} was the peak area of the internal standard, C_{IS} was the quantity of the internal standard (23: 0) (mg), W_S was the dry weight of the sample (g), and C_{FA} was expressed in mg_{FA} / g_{dry weight}.

To study the changes in FA composition during embryonic development (from stage I to stage IV), we only considered the FA relative abundances (%). Although FA contents were measured for early embryos, they were not estimated in shelled embryos (stage III and IV) because dry weight included shell weight, thus leading to underestimated FA concentrations. Thus, a potential decrease in FA concentration might be only due to an increase in shell weight.

Statistical Analysis

Data were expressed as the mean $\pm$ standard deviation. The non-parametric Kruskal-Wallis test [51] was used to test for differences in the relative abundances of the FA classes in the different tissues and embryos between the four occasions. When significant differences were observed, the Student-Newman-Keuls (SNK) post-hoc test was applied.

The PRIMER software [52] was used for multivariate analyses. The data matrices (fatty acid contributions (% of total fatty acids) in different groups (foot, visceral mass, embryo (stage I to IV), larva and POM) and at different sampling dates) were used to create triangular similarity matrices, based on the Bray-Curtis similarity coefficient, followed by non-metric multidimensional scaling (n-MDS) and hierarchical clustering analyses. Stress values inferior to 0.2 were considered robust following the recommendation of Clarke [53]. No transformation was performed on the data. Differences in fatty acid compositions between the different sampling occasions and the different groups were tested using a one-way analysis of similarity (ANOSIM). The statistical test (R statistic and associated *p*-value), was computed with 5000 permutations [52]. One-way ANOSIM was applied to groups with more than four replicates. When differences (between sampling dates, developmental stages, or groups) in the fatty acid composition were detected, similarity of percentages (SIMPER, a module of PRIMER) analyses were used to determine which fatty acids drive the observed differences.

Results

Seasonal variations of FA composition in female tissues and encapsulated embryos (stage I)

45 individual FA were identified in female tissues, and 43 in stage I embryos (Tables 1 and 2). The major FA were polyunsaturated FA (PUFA; 41.8 to 54.0%), followed by saturated FA (SFA; 20.2 to 32.0%), monounsaturated FA (MUFA; 16.0 to 20.8%), branched FA (BFA; 4.0 to 8.3%) and other FA (1.4 to 3.8%). The PUFA 20:5 ω 3, 22:6 ω 3, 20:4 ω 6, 22:5 ω 3 and 18:4 ω 3, the SFA 16: 0, 18: 0 and 14: 0, and the MUFA 16:1 ω 7, 18:1 ω 7, 18:1 ω 9, and 20:1 ω 9 were the most abundant FA. At each sampling date, FA composition of foot, visceral mass and embryos were statistically different (ANOSIM, *p*<0.01). N-MDS analysis (data not shown) further showed that embryos had a FA composition closer to that of the visceral mass.

Total fatty acid concentrations were different between the female tissues and embryos (Table 1, Figure 3), ranging from 5.41 ± 1.6 mg g⁻¹ dry weight (*n*=37) in the foot to 75.47 ± 24.5 mg g⁻¹ dry weight (*n*=13) in stage I embryos.

Seasonal variations in FA contents for each class of FA were tested with the non-parametric Kruskal-Wallis test (Figure 3). Except for PUFA, ω 3 and ω 6, all FA classes in foot samples showed a significant difference between dates (*p*<0.05). For total FA, SFA, MUFA and BFA, foot content was significantly higher in May, July and September than in February (SNK post-hoc test; *p*<0.01). In the visceral mass, except for BFA and ω 6, all FA classes showed a significant difference between dates (*p*<0.05), with significantly higher total FA, PUFA (especially ω 3), SFA, and MUFA concentrations in May and July than in February and September (SNK post-hoc test, *p*<0.05). In stage I embryos, although values were variable, no significant differences between dates were observed (*p*>0.05).

FA composition (relative abundance, in %) of foot tissues were statistically different between sampling dates (one-way ANOSIM, *p*<0.05). Hierarchical clustering (data not shown) and n-MDS ordination (Figure 4A) clearly demonstrated a grouping of samples based on their FA composition: 1) February and May (similar at 84%), and 2) May, July and September (similar at 88%). For visceral mass, one-way ANOSIM showed significant differences in FA composition between sampling dates (*p*<0.05), which were well separated in the hierarchical clustering (data not shown) and n-MDS ordination (Figure 4B) which grouped samples from February and September (similar at 88%), and samples from May and July (similar at 88.5%). Except for May and July which showed a strong intra-group variability (*R*=0.121), the *R* statistic of other groups was between 0.54 and 1 which indicated a weak intra-group variability. Similarly for stage I embryos, n-MDS ordination (Figure 4C) and hierarchical clustering (data not shown) separated the samples into two groups at the 89% similarity level: February and September (similar at 91%) and May and July (similar at 91%).

For each class of FA, differences in relative abundance between sampling dates were tested with the non-parametric Kruskal-Wallis test. In the foot, except for MUFA, all FA classes showed significant temporal differences (*p*<0.01; Figure 5A and

Table 1. Relative contribution (%) of fatty acids in female tissues (foot and visceral mass) and embryos at stage I of *Crepidula fornicata*.

FA	Foot				Visceral mass				Embryos at stage I			
	February	May	July	September	February	May	July	September	February	May	July	September
Saturated												
12:0	0.00 ± 0.0	0.01 ± 0.0	0.03 ± 0.1	0.05 ± 0.1	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0
14:0	1.17 ± 0.2	2.04 ± 0.4	2.84 ± 0.3	2.72 ± 0.4	1.27 ± 0.1	2.20 ± 0.3	2.95 ± 0.5	2.28 ± 0.6	2.17 ± 0.3	2.48 ± 0.2	3.38 ± 0.1	3.08 ± 0.2
15:0	0.42 ± 0.1	0.55 ± 0.3	0.72 ± 0.1	0.65 ± 0.1	0.60 ± 0.0	0.51 ± 0.1	0.57 ± 0.1	0.57 ± 0.1	0.64 ± 0.1	0.47 ± 0.0	0.58 ± 0.0	0.63 ± 0.0
16:0	7.36 ± 0.6	10.46 ± 2.3	12.65 ± 1.0	13.30 ± 1.2	10.86 ± 0.6	16.15 ± 1.7	17.70 ± 2.1	14.03 ± 2.1	17.39 ± 1.2	19.86 ± 0.3	20.85 ± 0.6	20.61 ± 0.4
17:0	1.23 ± 0.3	1.07 ± 0.2	1.29 ± 0.1	1.26 ± 0.1	1.47 ± 0.1	1.00 ± 0.1	1.05 ± 0.1	1.12 ± 0.1	1.51 ± 0.2	0.91 ± 0.0	1.04 ± 0.1	1.30 ± 0.1
18:0	8.44 ± 0.4	8.16 ± 0.7	8.33 ± 0.5	8.00 ± 0.4	7.23 ± 0.2	6.64 ± 0.5	6.28 ± 0.4	6.18 ± 0.4	5.93 ± 1.0	5.95 ± 0.7	5.38 ± 0.3	5.60 ± 0.7
19:0	0.00 ± 0.0	0.05 ± 0.1	0.11 ± 0.0	0.12 ± 0.0	0.04 ± 0.1	0.03 ± 0.0	0.04 ± 0.1	0.00 ± 0.0	0.12 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0
20:0	1.34 ± 1.2	1.14 ± 1.0	0.67 ± 0.4	1.00 ± 0.4	0.44 ± 0.4	0.64 ± 0.5	0.24 ± 0.3	0.38 ± 0.2	0.38 ± 0.4	0.46 ± 0.2	0.16 ± 0.1	0.08 ± 0.1
22:0	0.21 ± 0.1	0.17 ± 0.1	0.20 ± 0.1	0.23 ± 0.0	0.07 ± 0.1	0.13 ± 0.2	0.11 ± 0.1	0.02 ± 0.1	0.00 ± 0.0	0.40 ± 0.1	0.07 ± 0.1	0.09 ± 0.1
24:0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.09 ± 0.3	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0
Sum %	20.17 ± 0.7	23.66 ± 2.6	26.84 ± 1.3	27.31 ± 1.3	21.97 ± 1.6	27.38 ± 3.8	28.93 ± 3.6	24.57 ± 3.5	28.14 ± 0.4	30.53 ± 0.8	31.46 ± 0.6	31.40 ± 0.9
Monounsaturated												
15:1	0.58 ± 0.1	0.44 ± 0.2	0.40 ± 0.2	0.21 ± 0.1	0.27 ± 0.0	0.11 ± 0.1	0.14 ± 0.1	0.18 ± 0.2	0.04 ± 0.1	0.07 ± 0.1	0.04 ± 0.1	0.07 ± 0.1
16:1ω5	0.11 ± 0.2	0.98 ± 0.6	1.49 ± 0.7	2.21 ± 0.3	0.02 ± 0.0	0.53 ± 0.5	0.59 ± 0.6	0.45 ± 0.3	0.03 ± 0.1	0.00 ± 0.0	0.09 ± 0.1	0.10 ± 0.1
16:1ω7	1.86 ± 0.7	2.35 ± 0.8	2.52 ± 0.2	2.45 ± 0.2	2.08 ± 0.3	3.83 ± 0.5	4.01 ± 0.5	2.30 ± 1.0	3.38 ± 0.1	4.29 ± 0.1	4.35 ± 0.1	3.84 ± 0.1
16:1ω9	0.18 ± 0.2	0.49 ± 0.6	0.42 ± 0.1	0.38 ± 0.0	0.51 ± 0.0	0.39 ± 0.1	0.44 ± 0.1	0.61 ± 0.6	0.62 ± 0.1	0.51 ± 0.0	0.66 ± 0.0	0.60 ± 0.0
17:1	4.17 ± 1.5	2.26 ± 1.3	2.09 ± 1.1	0.85 ± 0.6	3.56 ± 0.1	1.55 ± 1.1	1.76 ± 1.2	3.10 ± 1.4	0.91 ± 0.4	0.62 ± 0.5	0.62 ± 0.4	1.05 ± 0.3
18:1ω11	0.16 ± 0.1	0.18 ± 0.1	0.26 ± 0.0	0.28 ± 0.1	0.16 ± 0.1	0.29 ± 0.1	0.28 ± 0.0	0.20 ± 0.1	0.20 ± 0.1	0.33 ± 0.1	0.36 ± 0.1	0.28 ± 0.0
18:1ω5	0.11 ± 0.0	0.17 ± 0.1	0.22 ± 0.0	0.23 ± 0.0	0.30 ± 0.0	0.23 ± 0.1	0.21 ± 0.0	0.16 ± 0.1	0.32 ± 0.1	0.27 ± 0.0	0.20 ± 0.0	0.24 ± 0.0
18:1ω7	1.43 ± 0.2	2.30 ± 0.4	2.28 ± 0.3	2.01 ± 0.2	2.11 ± 0.3	3.56 ± 0.4	3.44 ± 0.4	2.49 ± 0.4	3.72 ± 0.4	4.96 ± 0.3	4.32 ± 0.2	4.31 ± 0.1
18:1ω9	4.53 ± 0.5	4.75 ± 0.7	4.65 ± 0.7	5.08 ± 0.5	3.55 ± 0.2	2.83 ± 0.5	2.77 ± 0.4	2.98 ± 0.2	3.32 ± 0.2	2.29 ± 0.2	2.36 ± 0.2	2.93 ± 0.0
20:1ω9	5.21 ± 0.5	4.52 ± 0.4	5.38 ± 0.4	5.96 ± 0.3	4.37 ± 0.4	4.13 ± 0.3	4.05 ± 0.6	5.13 ± 0.7	3.86 ± 0.6	3.13 ± 0.3	2.58 ± 0.1	3.28 ± 0.2
20:1	1.59 ± 0.4	1.31 ± 0.3	1.08 ± 0.2	1.00 ± 0.2	1.67 ± 0.2	1.02 ± 0.8	0.73 ± 0.1	0.81 ± 0.1	1.25 ± 0.2	0.64 ± 0.0	0.66 ± 0.0	0.65 ± 0.0
Sum %	19.93 ± 1.2	19.74 ± 1.5	20.79 ± 0.8	20.67 ± 1.0	18.61 ± 2.6	18.46 ± 4.3	18.40 ± 4.0	18.41 ± 4.9	17.65 ± 1.0	17.10 ± 0.4	16.22 ± 0.6	17.35 ± 0.3
Branched												
15:0a	0.00 ± 0.0	0.31 ± 0.2	0.44 ± 0.2	0.60 ± 0.1	0.00 ± 0.0	0.10 ± 0.1	0.09 ± 0.1	0.04 ± 0.1	0.01 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0
15:0i	0.05 ± 0.1	0.48 ± 0.3	0.69 ± 0.2	0.93 ± 0.2	0.11 ± 0.1	0.17 ± 0.1	0.24 ± 0.2	0.24 ± 0.1	0.23 ± 0.0	0.06 ± 0.0	0.19 ± 0.0	0.28 ± 0.1
16:0i	0.59 ± 0.1	0.45 ± 0.2	0.65 ± 0.1	0.65 ± 0.1	0.88 ± 0.1	0.54 ± 0.1	0.72 ± 0.1	0.69 ± 0.1	1.25 ± 0.1	0.58 ± 0.1	0.93 ± 0.1	1.06 ± 0.0
17:0a	1.32 ± 0.2	1.46 ± 0.2	1.57 ± 0.2	1.68 ± 0.2	1.72 ± 0.2	1.68 ± 0.2	1.36 ± 0.2	1.39 ± 0.1	2.39 ± 0.2	1.64 ± 0.2	1.59 ± 0.1	1.92 ± 0.1
17:0i	2.51 ± 0.5	2.41 ± 0.4	2.80 ± 0.2	2.77 ± 0.4	3.10 ± 0.4	2.12 ± 0.2	2.44 ± 0.2	2.52 ± 0.3	4.38 ± 0.5	2.26 ± 0.3	2.96 ± 0.2	3.69 ± 0.1
Sum %	4.48 ± 0.6	5.12 ± 1.0	6.16 ± 0.9	6.62 ± 0.7	5.81 ± 0.7	4.61 ± 0.7	4.85 ± 0.7	4.89 ± 0.7	8.27 ± 0.8	4.53 ± 0.5	5.68 ± 0.3	6.95 ± 0.2
Polyunsaturated												
16:2	2.13 ± 0.4	1.82 ± 0.5	2.01 ± 0.4	1.82 ± 0.3	1.04 ± 0.1	0.59 ± 0.2	0.59 ± 0.2	0.73 ± 0.2	0.17 ± 0.1	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0
16:3	2.90 ± 2.9	3.00 ± 1.9	2.40 ± 1.0	3.73 ± 0.8	1.05 ± 1.0	1.78 ± 1.3	1.20 ± 1.0	1.47 ± 0.7	1.12 ± 0.8	0.95 ± 0.5	0.93 ± 0.3	0.66 ± 0.2
18:2ω6	3.13 ± 0.3	3.22 ± 0.4	3.47 ± 0.3	3.57 ± 0.8	2.78 ± 0.3	2.39 ± 0.4	2.67 ± 0.4	2.82 ± 0.4	2.13 ± 0.2	1.50 ± 0.1	1.86 ± 0.1	1.98 ± 0.0
18:3ω3	0.88 ± 0.1	0.96 ± 0.1	0.93 ± 0.1	0.94 ± 0.1	1.06 ± 0.1	1.18 ± 0.1	1.26 ± 0.2	1.10 ± 0.1	1.56 ± 0.2	1.49 ± 0.0	1.71 ± 0.1	1.49 ± 0.1
18:3ω6	0.10 ± 0.1	0.04 ± 0.1	0.10 ± 0.0	0.11 ± 0.0	0.03 ± 0.0	0.09 ± 0.1	0.19 ± 0.1	0.12 ± 0.1	0.19 ± 0.0	0.59 ± 0.6	0.26 ± 0.0	0.32 ± 0.0
18:4ω3	1.69 ± 0.3	2.93 ± 0.7	2.50 ± 0.5	1.98 ± 0.3	2.16 ± 0.4	4.37 ± 0.7	3.71 ± 0.6	3.72 ± 0.4	4.10 ± 0.7	6.05 ± 0.3	5.21 ± 0.4	4.70 ± 0.1
20:2	3.09 ± 0.4	2.39 ± 0.3	2.23 ± 0.3	2.12 ± 0.3	2.76 ± 0.4	1.68 ± 0.3	1.51 ± 0.3	2.00 ± 0.5	0.82 ± 0.1	0.52 ± 0.1	0.44 ± 0.0	0.43 ± 0.0
20:3	0.34 ± 0.3	0.16 ± 0.2	0.29 ± 0.2	0.28 ± 0.1	0.31 ± 0.2	0.01 ± 0.1	0.02 ± 0.1	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0

Table 1 (continued).

FA	Foot				Visceral mass				Embryos at stage I			
	February	May	July	September	February	May	July	September	February	May	July	September
20:4ω6	11.94 ± 0.9	9.29 ± 1.4	8.19 ± 0.7	9.30 ± 0.6	9.23 ± 0.6	4.98 ± 1.1	5.37 ± 1.0	9.54 ± 1.4	3.13 ± 0.2	1.37 ± 0.1	1.82 ± 0.1	4.55 ± 0.2
20:5ω3	10.18 ± 1.2	11.22 ± 1.5	9.12 ± 0.8	7.62 ± 0.6	13.97 ± 1.0	15.85 ± 1.5	15.41 ± 1.7	12.06 ± 1.4	16.72 ± 1.6	20.48 ± 0.3	18.89 ± 0.1	15.91 ± 0.6
22:3	1.24 ± 0.2	1.05 ± 0.2	1.05 ± 0.1	1.08 ± 0.1	1.34 ± 0.1	1.34 ± 0.2	1.07 ± 0.2	1.27 ± 0.1	0.89 ± 0.2	1.40 ± 0.1	0.86 ± 0.0	1.03 ± 0.1
22:4ω6	0.35 ± 0.1	0.20 ± 0.1	0.21 ± 0.1	0.23 ± 0.0	0.44 ± 0.1	0.18 ± 0.1	0.19 ± 0.1	0.31 ± 0.0	0.32 ± 0.0	0.09 ± 0.1	0.10 ± 0.1	0.23 ± 0.0
22:5ω3	6.17 ± 0.3	4.58 ± 0.9	3.82 ± 0.4	4.05 ± 0.5	4.59 ± 0.4	3.30 ± 0.6	3.25 ± 0.7	4.66 ± 0.8	2.22 ± 0.3	2.02 ± 0.2	2.02 ± 0.2	1.86 ± 0.1
22:6ω3	9.89 ± 0.7	7.98 ± 1.5	6.85 ± 0.7	6.96 ± 0.7	9.57 ± 0.2	8.28 ± 1.0	8.54 ± 1.3	10.25 ± 0.9	9.36 ± 0.6	7.93 ± 0.8	9.27 ± 0.7	8.59 ± 0.5
Sum %	54.02 ± 1.3	48.85 ± 4.3	43.16 ± 1.4	43.79 ± 1.9	50.33 ± 4.8	46.04 ± 7.6	44.97 ± 7.6	50.04 ± 6.9	42.72 ± 2.1	44.40 ± 0.5	43.37 ± 0.9	41.75 ± 0.9
DHA/EPA	0.98 ± 0.1	0.71 ± 0.1	0.75 ± 0.1	0.92 ± 0.1	0.69 ± 0.2	0.52 ± 0.7	0.55 ± 0.8	0.85 ± 0.6	0.56 ± 0.0	0.39 ± 0.0	0.49 ± 0.0	0.54 ± 0.0
Sum ω6	15.53 ± 1.0	12.75 ± 1.5	11.96 ± 0.7	13.22 ± 0.6	12.47 ± 0.9	7.64 ± 1.6	8.42 ± 1.5	12.78 ± 1.9	5.77 ± 0.1	3.56 ± 0.5	4.03 ± 0.2	7.07 ± 0.2
Sum ω3	28.79 ± 2.2	27.67 ± 3.6	23.22 ± 1.9	21.54 ± 1.6	31.35 ± 2.1	32.99 ± 4.0	32.16 ± 4.4	31.79 ± 3.6	33.96 ± 1.6	37.98 ± 1.0	37.11 ± 0.5	32.55 ± 0.9
ω3/ω6	1.87 ± 0.2	2.20 ± 0.4	1.95 ± 0.2	1.63 ± 0.1	2.52 ± 0.2	4.46 ± 1.0	3.91 ± 0.7	2.53 ± 0.38	5.89 ± 0.4	10.95 ± 1.8	9.22 ± 0.4	4.61 ± 0.2
Sum n.i. %	1.40 ± 0.3	2.63 ± 0.9	3.05 ± 1.2	1.61 ± 0.8	3.29 ± 0.6	3.52 ± 0.5	2.84 ± 0.3	2.10 ± 0.4	3.22 ± 0.5	3.43 ± 0.5	3.27 ± 0.0	2.55 ± 0.1
16:1ω7/16:0	0.26 ± 0.1	0.23 ± 0.1	0.20 ± 0.0	0.19 ± 0.0	0.19 ± 0.0	0.24 ± 0.0	0.23 ± 0.0	0.16 ± 0.1	0.20 ± 0.0	0.22 ± 0.0	0.21 ± 0.0	0.19 ± 0.0
Total mg g ⁻¹	3.63 ± 0.8	5.60 ± 1.4	5.73 ± 1.4	5.76 ± 1.7	12.89 ± 2.6	20.69 ± 7.7	20.35 ± 6.8	13.07 ± 3.3	60.29 ± 8.5	91.15 ± 25.7	81.97 ± 23.1	80.13 ± 24.8

Total amount (mg g⁻¹) of FA is also given.

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5D), particularly with a significant increase in SFA contribution and a significant decrease in PUFA contribution from February to September (SNK post hoc test; $p < 0.01$, except between July and September where $p > 0.05$). In visceral mass, except for MUFA and ω3, all FA classes showed significant temporal differences ($p < 0.05$; Figure 5B and 5E), with a significantly higher SFA contribution (SNK post-hoc test; $p < 0.01$, except between May and July where $p > 0.05$) and a significantly lower PUFA contribution (due to a significant decrease of ω6) in May and July (SNK post-hoc test; $p < 0.01$, except between May and July ($p > 0.05$) and between February and September ($p > 0.05$)). In stage I embryos, except for PUFA and MUFA, all FA classes showed a significant temporal difference ($p < 0.05$; Figure 5C and 5F). They showed a lower contribution of SFA in February (SNK post-hoc test; $p < 0.01$, no significant difference being observed between May, July and September ($p > 0.05$)). These differences in the relative abundance (%) of FA between sampling dates in foot, visceral mass, and embryos at stage I were mostly due to the PUFA 18:4ω3, 20:5ω3 (EPA), 20:4ω6 (AA), and 22:6ω3 (DHA), the SFA 16:0, the MUFA 17:1, and the BFA 17:0 iso (SIMPER analysis).

Altogether, the above results showed temporal variations in both FA content and composition of female tissues and embryos (stage I). In particular, they showed different compositions in May-July and February-September.

As a result of variations in abundance of individual FA, variations in the ω3/ω6 ratio were also observed between tissues (Kruskal-Wallis test, $p < 0.01$): it was approximately twice higher in visceral mass compared to foot, and twice higher in early embryos than in visceral mass (Figure 6). Within each sample type (foot, visceral mass and stage I embryos), the ω3/ω6 ratio varied between sampling dates (Kruskal-Wallis test, $p < 0.05$) (Figure 6). It was almost twice higher in May and July (middle of the reproductive season) than in February and September (beginning and end of the reproductive season). ω3 contribution in the foot decreased from 28.79% ± 2.2% in February to 21.54% ± 1.6% ($n = 37$) in September (SNK post-hoc test, $p < 0.01$); while ω6 contribution was higher in February (15.53% ± 1.0%, $n = 5$) than at the three other dates (12.57% ± 1.2%, $n = 32$) (SNK post-hoc test, $p < 0.01$; Figure 5D). As a consequence, ω3/ω6 varied from 1.63 ± 0.1 to 2.20 ± 0.4 ($n = 37$; Figure 6). In the visceral mass, ω3/ω6 ratios were 2.53 ± 0.3 ($n = 13$) and 4.18 ± 0.9 ($n = 24$) in February and September,

Table 2. Relative contribution (%) of fatty acids in embryos (stages I to IV) of *Crepidula fornicata*.

FA	February	May	July				September						
	Stage I	Stage I	Stage II	Stage III	Stage IV	Stage I	Stage II	Stage III	Stage IV	Stage I	Stage II	Stage III	Stage IV
Saturated													
14:0	2.17 ± 0.3	2.48 ± 0.2	2.35 ± 0.2	2.12 ± 0.2	2.11 ± 0.2	3.38 ± 0.1	3.03 ± 0.3	2.05 ± 0.1	2.05 ± 0.1	3.08 ± 0.2	3.90	3.31 ± 0.3	2.13 ± 0.4
15:0	0.64 ± 0.1	0.47 ± 0.0	0.55 ± 0.0	0.51 ± 0.0	0.61 ± 0.2	0.58 ± 0.0	0.57 ± 0.1	0.50 ± 0.0	0.45 ± 0.1	0.63 ± 0.0	0.59	0.54 ± 0.0	0.40 ± 0.1
16:0	17.39 ± 1.2	19.86 ± 0.3	19.11 ± 0.3	18.26 ± 0.8	18.68 ± 0.8	20.85 ± 0.6	20.95 ± 0.5	18.95 ± 1.0	15.49 ± 1.3	20.61 ± 0.4	20.32	20.39 ± 0.6	16.00 ± 1.0
17:0	1.51 ± 0.2	0.91 ± 0.0	1.19 ± 0.0	1.24 ± 0.1	1.33 ± 0.1	1.04 ± 0.1	1.07 ± 0.1	1.14 ± 0.0	1.13 ± 0.1	1.30 ± 0.1	1.10	1.27 ± 0.1	0.57 ± 0.6
18:0	5.93 ± 1.0	5.95 ± 0.7	6.29 ± 0.7	6.42 ± 0.5	6.31 ± 0.4	5.38 ± 0.3	5.55 ± 0.7	5.72 ± 0.3	6.87 ± 0.2	5.60 ± 0.7	5.38	6.00 ± 0.4	6.20 ± 0.4
19:0	0.12 ± 0.0	0.00 ± 0.0	0.03 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00	0.00 ± 0.0	0.00 ± 0.0
20:0	0.38 ± 0.4	0.46 ± 0.2	0.31 ± 0.1	0.33 ± 0.1	0.24 ± 0.2	0.16 ± 0.1	0.18 ± 0.1	0.15 ± 0.2	0.06 ± 0.1	0.08 ± 0.1	0.33	0.46 ± 0.1	0.32 ± 0.3
22:0	0.00 ± 0.0	0.40 ± 0.1	0.12 ± 0.1	0.03 ± 0.0	0.04 ± 0.1	0.07 ± 0.1	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.09 ± 0.1	0.18	0.00 ± 0.0	0.00 ± 0.0
Sum %	28.14 ± 0.4	30.53 ± 0.8	29.95 ± 1.0	28.90 ± 1.1	29.32 ± 0.2	31.46 ± 0.6	31.34 ± 0.7	28.50 ± 0.8	26.06 ± 1.4	31.40 ± 0.9	31.80	31.97 ± 0.7	25.62 ± 0.6
Monounsaturated													
15:1	0.04 ± 0.1	0.07 ± 0.1	0.05 ± 0.0	0.06 ± 0.0	0.04 ± 0.1	0.04 ± 0.1	0.08 ± 0.1	0.06 ± 0.1	0.15 ± 0.1	0.07 ± 0.1	0.06	0.04 ± 0.1	0.07 ± 0.1
16:1ω5	0.03 ± 0.1	0.00 ± 0.0	0.09 ± 0.1	0.16 ± 0.1	0.25 ± 0.2	0.09 ± 0.1	0.23 ± 0.2	0.00 ± 0.0	0.00 ± 0.0	0.10 ± 0.1	0.30	0.28 ± 0.2	0.31 ± 0.3
16:1ω7	3.38 ± 0.1	4.29 ± 0.1	3.50 ± 0.1	3.26 ± 0.1	2.77 ± 0.1	4.35 ± 0.1	4.00 ± 0.1	3.01 ± 0.2	1.29 ± 0.9	3.84 ± 0.1	3.99	4.15 ± 0.1	2.41 ± 0.0
16:1ω9	0.62 ± 0.1	0.51 ± 0.0	0.61 ± 0.1	0.66 ± 0.1	0.37 ± 0.3	0.66 ± 0.0	0.60 ± 0.1	0.60 ± 0.0	0.83 ± 1.2	0.60 ± 0.0	0.54	0.28 ± 0.2	0.38 ± 0.1
17:1	0.91 ± 0.4	0.62 ± 0.5	0.78 ± 0.4	0.99 ± 0.3	1.28 ± 0.6	0.62 ± 0.4	0.61 ± 0.5	1.51 ± 0.3	4.52 ± 0.7	1.05 ± 0.3	0.26	1.01 ± 0.3	2.78 ± 0.3
18:1ω11	0.20 ± 0.1	0.33 ± 0.1	0.31 ± 0.1	0.25 ± 0.0	0.11 ± 0.1	0.36 ± 0.1	0.33 ± 0.0	0.26 ± 0.0	0.16 ± 0.1	0.28 ± 0.0	0.21	0.25 ± 0.0	0.30 ± 0.0
18:1ω5	0.32 ± 0.1	0.27 ± 0.0	0.33 ± 0.0	0.30 ± 0.0	0.19 ± 0.1	0.20 ± 0.0	0.25 ± 0.0	0.16 ± 0.1	0.11 ± 0.1	0.24 ± 0.0	0.13	0.24 ± 0.0	0.22 ± 0.0
18:1ω7	3.72 ± 0.4	4.96 ± 0.3	4.05 ± 0.1	3.82 ± 0.3	3.53 ± 0.2	4.32 ± 0.2	4.36 ± 0.5	4.01 ± 0.4	3.01 ± 0.2	4.31 ± 0.1	4.15	3.45 ± 0.1	3.24 ± 0.3
18:1ω9	3.32 ± 0.2	2.29 ± 0.2	2.82 ± 0.4	3.59 ± 0.3	3.25 ± 0.4	2.36 ± 0.2	2.51 ± 0.3	2.63 ± 0.3	2.30 ± 0.2	2.93 ± 0.0	2.48	2.85 ± 0.4	2.70 ± 0.2
20:1ω9	3.86 ± 0.6	3.13 ± 0.3	3.87 ± 0.3	4.09 ± 0.5	4.86 ± 0.1	2.58 ± 0.1	3.34 ± 0.3	4.49 ± 0.7	6.69 ± 0.4	3.28 ± 0.2	3.25	4.74 ± 0.1	6.02 ± 0.2
20:1	1.25 ± 0.2	0.64 ± 0.0	0.92 ± 0.1	1.27 ± 0.2	1.38 ± 0.2	0.66 ± 0.0	0.77 ± 0.1	0.94 ± 0.1	0.70 ± 0.0	0.65 ± 0.0	0.71	0.66 ± 0.1	1.13 ± 0.1
Sum %	17.65 ± 1.0	17.10 ± 0.4	17.31 ± 0.3	18.46 ± 0.7	18.02 ± 1.3	16.22 ± 0.6	17.08 ± 0.5	17.66 ± 0.1	19.77 ± 0.5	17.35 ± 0.3	16.08	17.95 ± 0.5	19.55 ± 0.8
Branched													
15:0a	0.01 ± 0.0	0.00 ± 0.0	0.02 ± 0.0	0.00 ± 0.0	0.04 ± 0.1	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00	0.00 ± 0.0	0.00 ± 0.0
15:0i	0.23 ± 0.0	0.06 ± 0.0	0.12 ± 0.0	0.11 ± 0.0	0.24 ± 0.0	0.19 ± 0.0	0.20 ± 0.0	0.05 ± 0.1	0.00 ± 0.0	0.28 ± 0.1	0.26	0.21 ± 0.0	0.16 ± 0.0
16:0i	1.25 ± 0.1	0.58 ± 0.1	0.78 ± 0.1	0.86 ± 0.1	1.08 ± 0.1	0.93 ± 0.1	1.00 ± 0.2	0.80 ± 0.1	0.57 ± 0.1	1.06 ± 0.0	0.88	0.80 ± 0.0	0.39 ± 0.4
17:0a	2.39 ± 0.2	1.64 ± 0.2	1.64 ± 0.1	1.78 ± 0.2	2.14 ± 0.1	1.59 ± 0.1	1.83 ± 0.3	1.50 ± 0.1	1.19 ± 0.2	1.92 ± 0.1	1.62	1.43 ± 0.1	1.21 ± 0.3

Table 2 (continued).

FA	February	May	July							September			
	Stage I	Stage I	Stage II	Stage III	Stage IV	Stage I	Stage II	Stage III	Stage IV	Stage I	Stage II	Stage III	Stage IV
17:0i	4.38 ± 0.5	2.26 ± 0.3	2.87 ± 0.2	3.04 ± 0.4	3.78 ± 0.2	2.96 ± 0.2	3.28 ± 0.4	2.79 ± 0.2	2.27 ± 0.1	3.69 ± 0.1	3.08	2.73 ± 0.1	2.29 ± 0.5
Sum %	8.27 ± 0.8	4.53 ± 0.5	5.43 ± 0.4	5.78 ± 0.7	7.29 ± 0.3	5.68 ± 0.3	6.31 ± 0.9	5.14 ± 0.4	4.04 ± 0.3	6.95 ± 0.2	5.84	5.17 ± 0.3	4.05 ± 1.1
Polyunsaturated													
16:2	0.17 ± 0.1	0.00 ± 0.0	0.09 ± 0.1	0.19 ± 0.2	0.23 ± 0.2	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.05 ± 0.1	0.00 ± 0.0	0.00	0.00 ± 0.0	0.00 ± 0.0
16:3	1.12 ± 0.8	0.95 ± 0.5	0.77 ± 0.1	0.97 ± 0.2	1.18 ± 0.5	0.93 ± 0.3	0.91 ± 0.3	0.68 ± 0.5	0.53 ± 0.2	0.66 ± 0.2	1.39	1.81 ± 0.2	0.96 ± 1.0
18:2ω6	2.13 ± 0.2	1.50 ± 0.1	1.84 ± 0.2	1.81 ± 0.2	2.18 ± 0.2	1.86 ± 0.1	1.88 ± 0.1	2.22 ± 0.3	2.80 ± 0.3	1.98 ± 0.0	1.79	2.04 ± 0.1	2.42 ± 0.4
18:3ω3	1.56 ± 0.2	1.49 ± 0.0	1.61 ± 0.1	1.44 ± 0.1	1.10 ± 0.1	1.71 ± 0.1	1.51 ± 0.3	1.53 ± 0.1	1.20 ± 0.2	1.49 ± 0.1	1.20	1.06 ± 0.0	1.18 ± 0.3
18:3ω6	0.19 ± 0.0	0.59 ± 0.6	0.05 ± 0.1	0.08 ± 0.1	0.04 ± 0.1	0.26 ± 0.0	0.19 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.32 ± 0.0	0.26	0.20 ± 0.0	0.00 ± 0.0
18:4ω3	4.10 ± 0.7	6.05 ± 0.3	4.94 ± 0.1	4.03 ± 0.4	2.84 ± 0.2	5.21 ± 0.4	4.49 ± 0.8	4.53 ± 0.4	4.34 ± 0.6	4.70 ± 0.1	3.84	2.86 ± 0.1	3.44 ± 1.3
20:2	0.82 ± 0.1	0.52 ± 0.1	0.70 ± 0.1	0.83 ± 0.1	0.94 ± 0.2	0.44 ± 0.0	0.58 ± 0.1	0.96 ± 0.2	1.43 ± 0.2	0.43 ± 0.0	0.50	0.80 ± 0.0	1.16 ± 0.1
20:3	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00 ± 0.0	0.00	0.00 ± 0.0	0.00 ± 0.0
20:4ω6	3.13 ± 0.2	1.37 ± 0.1	1.88 ± 0.3	2.50 ± 0.3	3.76 ± 0.5	1.82 ± 0.1	2.04 ± 0.2	2.29 ± 0.3	4.97 ± 2.4	4.55 ± 0.2	5.08	6.26 ± 0.1	5.74 ± 2.8
20:5ω3	16.72 ± 1.6	20.48 ± 0.3	19.84 ± 0.7	19.01 ± 0.2	17.19 ± 1.2	18.89 ± 0.1	18.75 ± 0.6	18.91 ± 0.9	16.85 ± 0.5	15.91 ± 0.6	15.81	15.63 ± 0.8	19.01 ± 0.9
22:3	0.89 ± 0.2	1.40 ± 0.1	1.06 ± 0.2	0.96 ± 0.1	0.78 ± 0.1	0.86 ± 0.0	0.85 ± 0.1	0.94 ± 0.1	0.79 ± 0.3	1.03 ± 0.1	1.28	1.11 ± 0.0	0.84 ± 0.0
22:4ω6	0.32 ± 0.0	0.09 ± 0.1	0.14 ± 0.1	0.21 ± 0.0	0.26 ± 0.0	0.10 ± 0.1	0.12 ± 0.1	0.23 ± 0.0	0.08 ± 0.1	0.23 ± 0.0	0.27	0.21 ± 0.0	0.09 ± 0.1
22:5ω3	2.22 ± 0.3	2.02 ± 0.2	2.13 ± 0.2	2.58 ± 0.2	2.98 ± 0.2	2.02 ± 0.2	2.23 ± 0.2	3.11 ± 0.4	3.82 ± 0.3	1.86 ± 0.1	2.38	3.25 ± 0.3	4.18 ± 0.7
22:6ω3	9.36 ± 0.6	7.93 ± 0.8	8.44 ± 0.0	8.65 ± 0.7	9.14 ± 1.7	9.27 ± 0.7	8.79 ± 0.5	10.36 ± 0.2	9.95 ± 0.6	8.59 ± 0.5	10.44	7.32 ± 0.3	9.06 ± 1.3
Sum %	42.72 ± 2.1	44.40 ± 0.5	43.49 ± 0.1	43.26 ± 1.4	42.62 ± 3.0	43.37 ± 0.9	42.34 ± 1.7	45.76 ± 0.4	46.82 ± 1.9	41.75 ± 0.9	44.24	42.55 ± 1.3	48.07 ± 1.2
DHA/EPA	0.56 ± 0.0	0.39 ± 0.0	0.43 ± 0.0	0.46 ± 0.0	0.53 ± 0.1	0.49 ± 0.0	0.47 ± 0.0	0.55 ± 0.0	0.59 ± 0.0	0.54 ± 0.0	0.66	0.47 ± 0.0	0.47 ± 0.1
Sum ω6	5.77 ± 0.1	3.56 ± 0.5	3.91 ± 0.5	4.59 ± 0.3	6.25 ± 0.7	4.03 ± 0.2	4.23 ± 0.4	4.73 ± 0.6	7.85 ± 2.3	7.07 ± 0.2	7.40	8.71 ± 0.1	8.25 ± 3.1
Sum ω3	33.96 ± 1.6	37.98 ± 1.0	36.95 ± 0.3	35.72 ± 1.1	33.25 ± 2.8	37.11 ± 0.5	35.77 ± 1.3	38.43 ± 1.1	36.17 ± 1.9	32.55 ± 0.9	33.67	30.12 ± 1.2	36.87 ± 3.0
ω3/ω6	5.89 ± 0.4	10.95 ± 1.8	9.58 ± 1.2	7.81 ± 0.5	5.39 ± 0.8	9.22 ± 0.4	8.53 ± 0.9	8.28 ± 1.3	4.99 ± 1.3	4.61 ± 0.2	4.55	3.46 ± 0.2	5.34 ± 2.4
Sum n.i. %	3.22 ± 0.5	3.43 ± 0.5	3.82 ± 0.5	3.60 ± 0.1	2.76 ± 0.8	3.27 ± 0.0	2.92 ± 0.3	2.95 ± 0.2	3.31 ± 0.3	2.55 ± 0.1	2.04	2.36 ± 0.3	2.72 ± 0.2
16:1ω7/16:0	0.20 ± 0.0	0.22 ± 0.0	0.18 ± 0.0	0.18 ± 0.0	0.15 ± 0.0	0.21 ± 0.0	0.19 ± 0.0	0.16 ± 0.0	0.09 ± 0.1	0.19 ± 0.0	0.2	0.20 ± 0.0	0.15 ± 0.0

Values are means ± standard deviations

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and in May and July, respectively (Figure 6), with ω3 remaining stable ($32.24 \pm 2.8\%$; $n=37$) at the four occasions (Kruskal-Wallis test, $p > 0.05$; Figure 5E); consequently, changes in the

ω3/ω6 values were only due to changes in ω6 value which was $8.03 \pm 1.3\%$ ($n=24$) in the middle of the reproductive season and $12.66 \pm 1.4\%$ ($n=13$) at the beginning and at the end of the

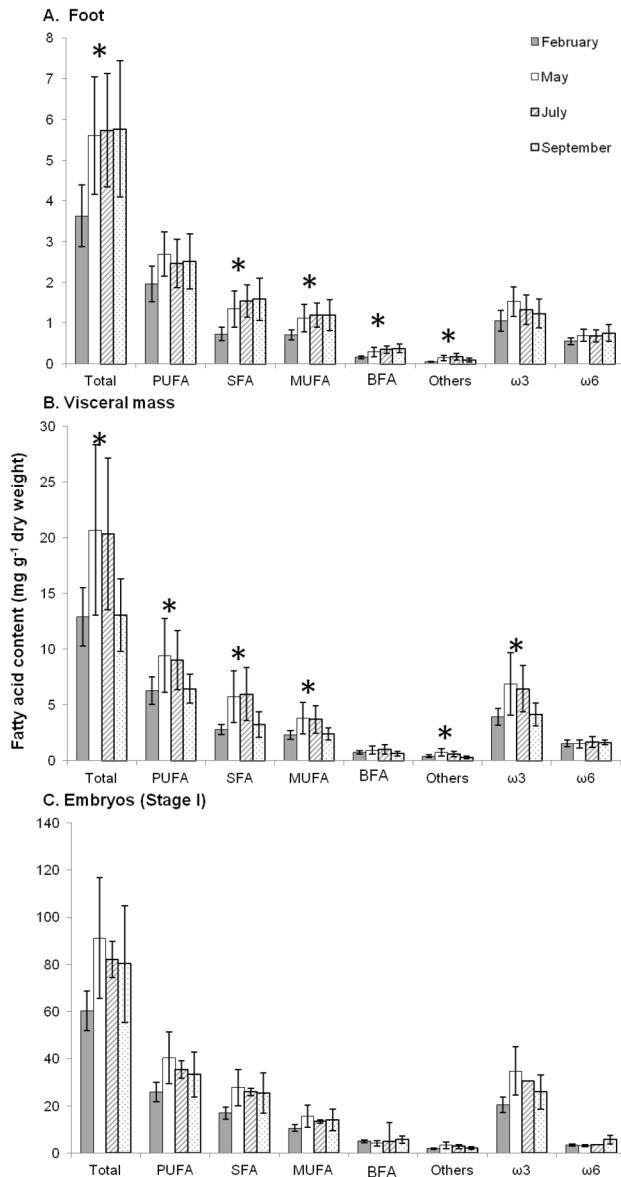


Figure 3. Seasonal variations of fatty acid content of females and embryos (stage I). Seasonal variations of fatty acid content (Total FA, PUFA, SFA, MUFA, BFA, Others, $\omega 3$ and $\omega 6$) at four periods of the reproductive season of *Crepidula fornicata* (February, May, July and September). A: foot; B: visceral mass; C: embryos (stage I). Histograms represent means and error bars correspond to standard deviations. Black stars represent significant differences between periods (Kruskal-Wallis test, $p < 0.05$).

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reproductive season (SNK post-hoc test; $p < 0.01$). In early embryos, the ratio was 5.52 ± 0.7 ($n=7$) at the beginning and at the end of the reproductive season and 10.08 ± 1.7 ($n=6$) in the middle of the reproductive season (Figure 6). $\omega 3$ contributions varied significantly between $37.54 \pm 1.0\%$ ($n=6$) in the middle of the reproductive season and $33.55 \pm 1.7\%$ ($n=7$) at the

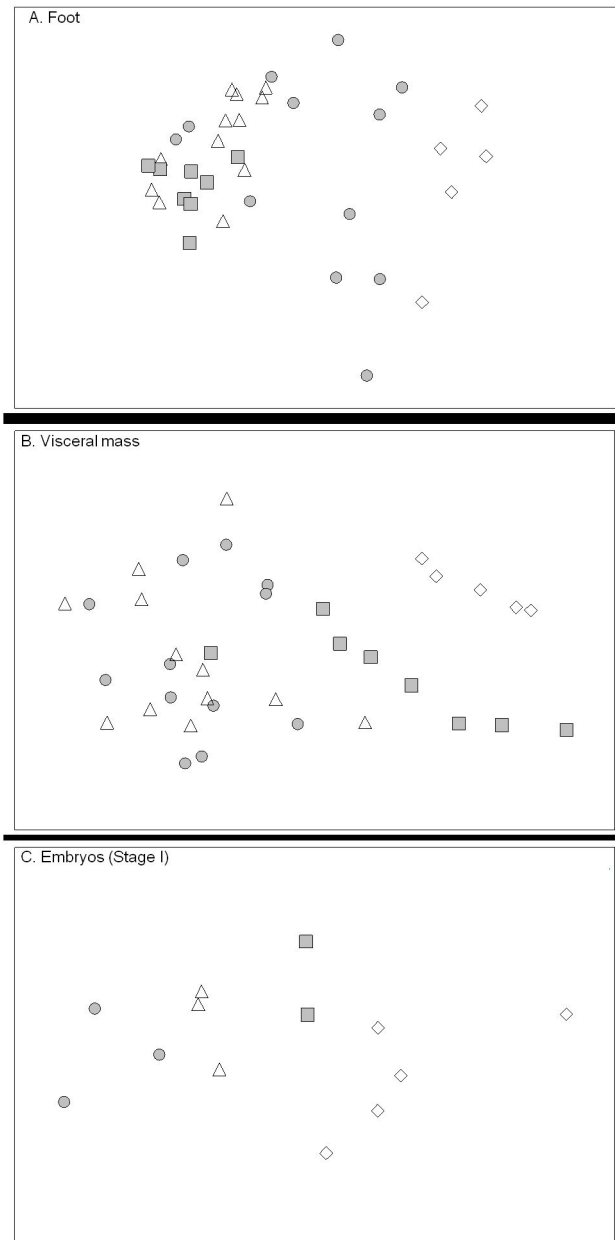


Figure 4. n-MDS ordination plot of females and embryos (stage I). n-MDS ordination plot of Bray-Curtis similarities based on FA composition of A: foot, B: visceral mass, and C: embryos at stage I of *Crepidula fornicata*. The degree of stress (0.13, 0.1, 0.04, for foot, visceral mass, and embryos at stage I, respectively) is within the range of values recommended by Clarke and Warwick [52] for robust grouping. Symbols corresponded to replicates collected in February (open diamond), May (filled circle), July (open triangle), and September (filled square).

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beginning and at the end of the reproductive season (SNK post-hoc test; $p < 0.01$); $\omega 6$ contributions varied between $6.14 \pm$

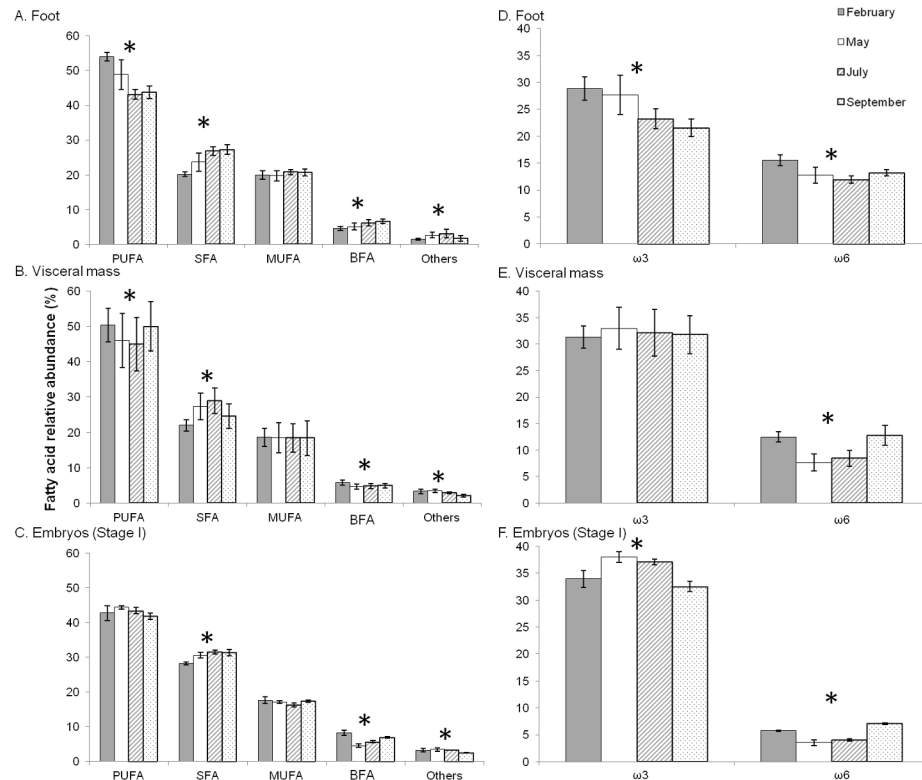


Figure 5. Seasonal variations of relative abundance of FA of females and embryos (stage I). Seasonal variations of relative abundance of different classes of FA (PUFA, SFA, MUFA, BFA, Others, $\omega 3$ and $\omega 6$) at four periods of the reproductive season of *Crepidula fornicata* (February, May, July, and September). A and D: foot; B and E: visceral mass; and C and F: embryos (stage I). Histograms represent means and error bars correspond to standard deviations. Black stars represent significant differences between periods (Kruskal-Wallis test, $p < 0.05$).

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0.7% ($n=7$) for February and September and $3.80 \pm 0.5\%$ ($n=6$) for May and July (SNK post-hoc test; $p < 0.01$; Figure 5F), and were half than those measured in the visceral masses.

To summary, the above results showed that female tissues and embryos at stage I have a higher $\omega 3/\omega 6$ ratio in May and July as compared to February and September.

Changes in fatty acid composition during embryo development

The n-MDS analyses of the FA composition of embryos at the four stages showed, for the three dates, differences from stage I to stage IV (Figure 7). Hierarchical clustering analysis (data not shown) confirmed this trend and demonstrated a clear separation of stages I, III, and IV. Stage II occupies an intermediate place, closer to stage I and/or III depending on the sampling date (Figure 7). Hierarchical clustering analysis further showed that stages I, II and III had similar FA composition with more than 90% similarity. Embryos at stage IV were more dissimilar (80 to 85% similarity for July and September, and more than 85% for May).

For all developmental stages, the major FA class was the PUFA (41.75 to 48.07%), followed by SFA (25.62% to 31.97%), MUFA (16.08 to 19.77%), BFA (4.04 to 7.29%) and other FA

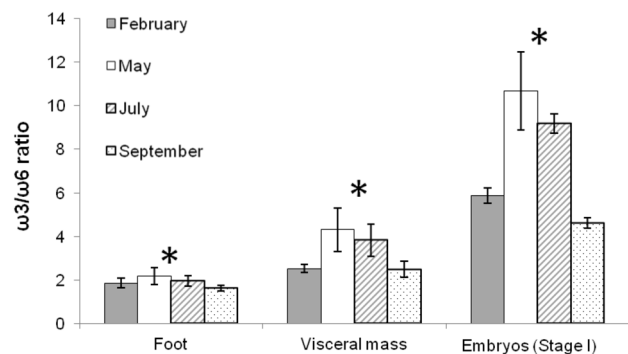


Figure 6. Seasonal variations of the quality index of females and embryos (stage I). Seasonal variations of the $\omega 3/\omega 6$ ratio at four periods of the reproductive season of *Crepidula fornicata* (February, May, July, and September) in foot, visceral mass, and embryos (stage I). Histograms represent means and error bars correspond to standard deviations. Black stars represent significant differences between periods (Kruskal-Wallis test, $p < 0.05$).

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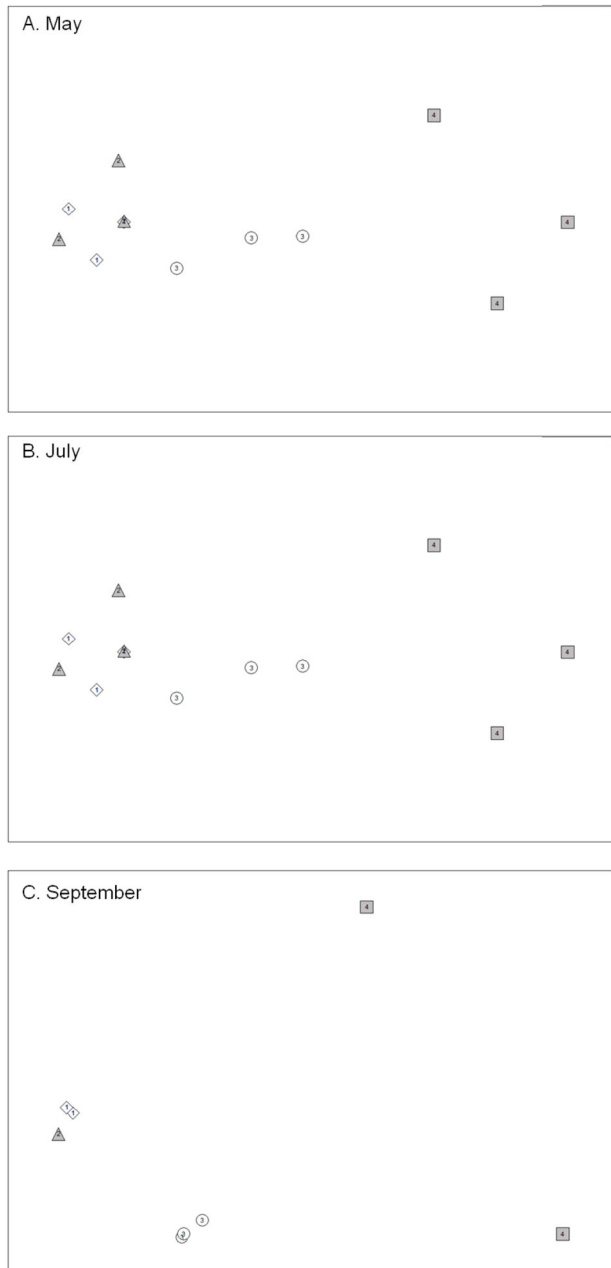


Figure 7. n-MDS ordination plot of embryos at stages I to IV. n-MDS ordination plot of Bray-Curtis similarities based on FA composition of embryos (stage I to IV) of *Crepidula fornicata*. A: May; B: July; and C: September. The degree of stress (0.05, 0.03, and 0.01, for May, July, and September, respectively) is within the range of values recommended by Clarke and Warwick [52] for robust grouping. Symbols corresponded to replicates of stages I (1; open diamond), II (2; filled triangle), III (3; open circle), and IV (4; filled square).
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(2.04 to 3.82%) (Figure 8). The FA 14:0, 16:0, 18:0, 17:0 iso, 16:1 ω 7, 18:1 ω 7, 20:1 ω 11, 18:4 ω 3, 20:5 ω 3, and 22:6 ω 3 were

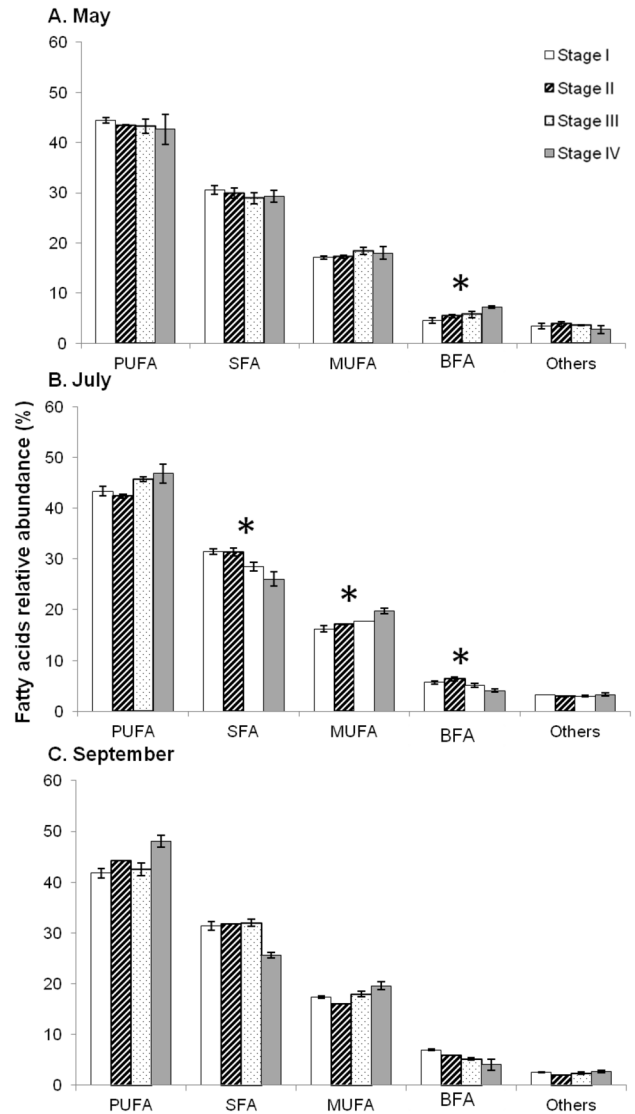


Figure 8. Seasonal variations of FA composition of embryos at stages I to IV. Seasonal variations of relative abundance of different classes of FA (PUFA, SFA, MUFA, BFA, and Others) at four developmental stages (I to IV) of *Crepidula fornicata*. A: May; B: July; and C: September. Histograms represent means and error bars correspond to standard deviations. Black stars represent significant differences between developmental stages (Kruskal-Wallis test, $p < 0.05$).
doi: 10.1371/journal.pone.0075316.g008

the major FA found in embryos and contributed up to 79.7–84.1% of the total FA content.

Comparisons of the relative contribution of the different FA classes (PUFA, SFA, MUFA, BFA, and others) between the four developmental stages at each sampling date with a Kruskal-Wallis test, revealed that in May, except for BFA and ω 6 ($p < 0.05$), there was no significant difference ($p > 0.05$); in July, except for BFA, SFA and MUFA ($p < 0.05$), all other FA

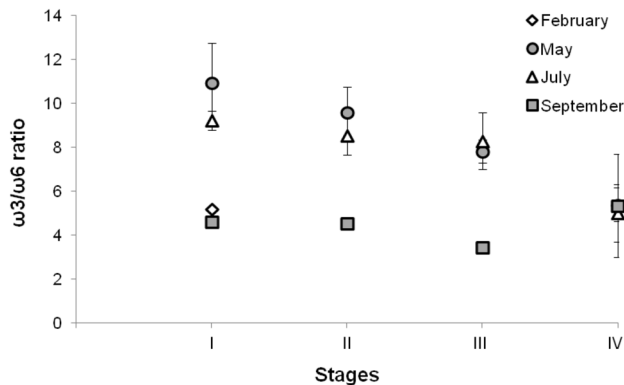


Figure 9. Variations of the quality index of embryos at stages I to IV. Variations of ω_3/ω_6 ratio in embryos at stages I to IV, at four periods of the reproductive season of *Crepidula fornicata* (February, May, July, and September). Points represent means and error bars correspond to standard deviations.

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classes showed no significant difference ($p>0.05$); in September, all FA classes were not significantly different ($p>0.05$) between stages. The observed differences were mostly due to the PUFA 18:4 ω_3 , 20:5 ω_3 (EPA), 20:4 ω_6 (AA), and 22:6 ω_3 (DHA), the SFA 16: 0, and the MUFA 20:1 ω_9 (SIMPER analysis).

Looking more specifically at PUFA in May, the relative concentration of ω_3 decreased from stage I to stage IV embryos (from 37.98 to 33.25%) whereas total ω_6 FA increased (3.56 to 6.25%), leading to a decrease of the ω_3/ω_6 ratio from 11.0 to 5.4 (Figure 9). The same pattern was observed in July, with a decrease of the ω_3/ω_6 ratio from 9.2 to 5.0 (Figure 9). A different pattern was observed in September, with an increase of the relative concentration of ω_3 (from 32.55 to 36.87%), and ω_6 (from 7.07 to 8.71%) between stage I and IV, leading to an increase in the ω_3/ω_6 ratio between 3.5 and 5.3 (Figure 9).

Seasonal variations in the FA composition of pelagic larvae of *C. fornicata*

35 FA were identified in pelagic larvae (Table 3). FA composition was highly variable depending on the sampling date (Figure 10), and no clear seasonal pattern was evidenced. Total FA in larvae were mostly SFA (38.23 to 56.87%), followed by PUFA (19.48% to 37.90%), MUFA (16.40 to 18.85%), and BFA (1.29 to 3.09%) (Figure 10, Table 3), except in larvae collected on the 28th August, in which PUFA dominated (38.98%). Individual FA that most contributed to the larval FA pool (between 71 to 82% of the total FA content) were the SFA 16: 0, 18: 0 and 14: 0, the PUFA 20:5 ω_3 , 22:6 ω_3 , 22:5 ω_3 and 20:4 ω_6 , the MUFA 20:1 ω_9 , 18:1 ω_7 , 16:1 ω_7 , and the BFA 17: 0*iso*.

A one-way ANOSIM showed significant differences in FA composition of pelagic larvae and embryos at stage IV ($p<0.01$). The two groups were well separated after hierarchical

clustering (data not shown) and n-MDS ordination (Figure 11). The n-MDS analysis (Figure 11) clearly demonstrated a grouping of the larval samples (similar at 80.2%), and of the samples of embryos at stage IV (similar at 87.3%). Differences between these two groups were mostly due to the SFA (12: 0, 14: 0, 16: 0, 17: 0 and 18: 0) and the PUFA (20:5 ω_3 , 20:4 ω_6 , and 22:6 ω_3) (SIMPER). The larval ω_3/ω_6 ratios (between 3.94 and 5.34) were similar to those observed in embryos at stage IV, except at two sampling dates: June (ω_3/ω_6 = 8.04) and October (ω_3/ω_6 = 2.39).

Seasonal variations in the FA composition of POM

35 FA were identified in POM. Whatever the sampling date, depth, or pre-filtration mesh-size, POM was always dominated by SFA (56.41 to 67.56%), followed by MUFA (25.02 to 33.20%), PUFA (4.19 to 10.88%), and BFA (2.46 to 5.05%) (Table 4). The fatty acids 14: 0, 15: 0, 16: 0, 18: 0, 16:1 ω_7 , 16:1 ω_9 , 18:1 ω_7 , and 18:1 ω_9 were the major FA found in POM samples.

Discussion

Seasonal variations in maternal supply to early embryos

Through its role on larval and even postlarval performance [1,3,6,54], maternal provisioning to offspring may influence dispersal and recruitment success in marine invertebrates. In species with a long reproductive period, parents may have to cope with seasonal variations of the environmental conditions, including food abundance and composition, which may in turn change resource allocation to offspring. Ultimately this may lead to seasonal changes in offspring condition (e.g. [13]), with potential consequences on recruitment success. Assessing the variations in maternal provisioning along the reproductive cycle of a given species is thus essential.

In the slipper limpet *Crepidula fornicata*, which reproduces from February to October, the foot and visceral mass of females showed different fatty acid compositions, as previously reported in other gastropod species [55,56]. Furthermore, both tissues showed different seasonal patterns along the reproductive period (Figure 4A and B). These differences likely highlighted the different metabolic patterns in both tissues. The foot samples clearly showed a seasonal gradient from February to July and September, with the samples from May having a high intra-group variability (Figure 4A). These latter could be divided into three subgroups: 1) samples which displayed a winter signature, similar to those of February, 2) samples which were closer to those of the summer (July and September), and 3) samples showing an intermediate position between those from February and July. In the opposite, visceral masses did not show a seasonal gradient, but a separation between females from the middle of the reproductive period (i.e. May and July) and those from the beginning and end of the reproductive period (Figure 4B).

As reported earlier by Gardner and Riley [57], unsaturated FA are dominant (up to 75% of total FA) in both female tissues. Even if we have to be cautious in comparing our results with these data - they were obtained from composite samples of

Table 3. Relative contribution (%) of fatty acids in pelagic larvae of *Crepidula fornicata*.

FA	Larvae					
	12/05/2009	04/06/2009	01/07/2009	31/07/2009	28/08/2009	06/10/2009
Saturated						
12:0	1.25	1.04	3.85	3.49	1.32	5.38
14:0	3.38	3.77	5.74	5.46	4.02	6.55
15:0	0.74	1.09	1.07	1.04	1.21	2.06
16:0	16.56	26.85	33.54	22.04	19.35	22.72
17:0	6.51	4.38	1.14	1.03	1.48	7.34
18:0	8.68	10.41	11.53	8.52	8.96	12.14
22:0	0.54	0.00	0.00	0.00	0.00	0.00
24:0	0.58	0.72	0.00	0.19	0.16	0.33
Sum %	38.23	48.25	56.87	41.77	36.50	56.51
Monounsaturated						
16:1 ω 7	2.02	4.55	2.41	3.15	2.29	1.59
16:1 ω 9	0.91	0.88	0.52	0.92	1.62	2.81
17:1	3.01	0.64	1.42	0.85	1.19	1.11
18:1 ω 7	2.53	3.99	2.42	2.95	2.62	1.24
18:1 ω 9	3.92	2.56	3.62	2.83	2.89	6.33
20:1 ω 9	5.09	4.63	6.04	4.92	5.70	2.84
20:1	1.37	0.87	0.94	0.90	1.07	0.47
Sum %	18.85	18.12	17.38	16.51	17.38	16.40
Branched						
15:0i	0.00	0.00	0.00	0.00	0.25	0.00
16:0i	0.56	0.00	0.00	0.49	0.50	0.00
17:0a	0.90	0.81	0.87	0.67	0.80	0.28
17:0i	1.82	1.69	1.91	1.93	2.05	1.01
Sum %	3.28	2.50	2.78	3.09	3.60	1.29
Polyunsaturated						
16:2	0.00	0.00	0.00	0.00	0.00	1.57
16:3	0.00	0.00	0.00	0.00	0.34	0.63
18:2 ω 6	1.88	1.58	1.87	1.83	1.79	2.17
18:3 ω 3	0.95	0.99	0.65	0.91	0.87	1.25
18:3 ω 6	0.52	0.00	0.00	0.00	0.00	0.84
18:4 ω 3	2.83	2.35	1.28	1.65	1.68	0.87
20:2	0.96	0.70	0.69	0.82	0.93	0.51
20:4 ω 6	3.68	1.47	1.82	3.48	5.73	3.44
20:5 ω 3	14.49	12.38	7.30	14.95	14.69	6.15
22:3	1.20	0.72	0.00	0.54	0.54	0.00
22:5 ω 3	3.40	2.71	1.85	3.09	4.09	2.35
22:6 ω 3	8.00	6.13	4.02	7.77	8.27	4.75
Sum %	37.90	29.02	19.48	35.03	38.92	24.52
DHA/EPA	0.55	0.50	0.55	0.52	0.56	0.77
Sum ω 6	6.08	3.05	3.69	5.31	7.51	6.44
Sum ω 3	29.67	24.55	15.10	28.37	29.60	15.37
ω 3/ ω 6	4.88	8.04	4.10	5.34	3.94	2.39
Sum n.i. %	1.74	2.11	3.49	3.59	3.60	1.28
16:1 ω 7/16:0	0.12	0.17	0.07	0.14	0.12	0.07

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whole tissues from several animals collected in November, and for three lipid classes (triglycerides, phospholipids and sterol esters) – some differences may be highlighted. First, both female tissues were characterized by high levels of C20 and C22 mono- and polyunsaturated fatty acids, most of which being essential in marine animals including molluscs, and

known to play major roles in their growth, survival and reproduction ([26,56,58], and references herein). Among them, the 20:4 ω 6 arachidonic acid (AA), DHA and EPA are more abundant in both foot and visceral mass than in Gardner and Riley's samples. In particular, AA showed an interesting seasonal pattern in the visceral mass, decreasing in May and

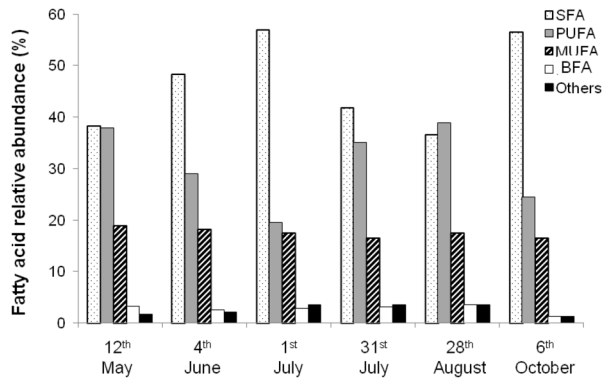


Figure 10. Seasonal variations of FA composition of pelagic larvae. Seasonal variations of relative abundance of different classes of FA (SFA, PUFA, MUFA, BFA, and Others) in pelagic larvae of *Crepidula fornicata* in 2009.

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Figure 11. n-MDS ordination plot of embryos at stage IV and pelagic larvae. n-MDS ordination plot of Bray-Curtis similarities based on FA composition of embryos at stage IV (open squares) and pelagic larvae (filled squares) of *Crepidula fornicata*. The degree of stress (0.06) is within the range of values recommended by Clarke and Warwick [52] for robust grouping.

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July (ca. 5%, compared to ca. 9% in February and September), most likely in relation with the reproductive activity of *C. fornicata*, as previously reported in abalone [56]. Indeed, AA is known to play an important role in gametogenesis as a precursor of prostaglandins (e.g. [24]). Second, as expected from their feeding regime, most marine FA biomarkers (e.g. [20]) are found in various amounts in adult tissues of *C. fornicata*. Indeed, adults of *C. fornicata* are filter-feeders which are considered omnivorous: they feed on various sizes of phytoplankton, from picoplankton to large diatoms and dinoflagellates, both directly or within particle aggregates (e.g. [59–62]), without any selection [63,64]. A noticeable difference with Gardner and Riley's data is the absence in our study of the

Table 4. Relative contribution (%) of fatty acids in the particulate organic matter (POM).

FA	POM
Saturated	
12:0	2.37 ± 0.74
14:0	10.88 ± 1.83
15:0	3.80 ± 1.11
16:0	30.72 ± 2.62
17:0	2.10 ± 1.11
18:0	8.38 ± 1.58
20:0	0.16 ± 0.25
22:0	1.08 ± 0.80
24:0	1.36 ± 0.48
Sum %	60.85 ± 3.12
Monounsaturated	
15:1	1.01 ± 0.45
16:1ω5	0.36 ± 0.21
16:1ω7	7.06 ± 2.90
16:1ω9	7.35 ± 2.63
17:1	0.63 ± 0.57
18:1ω5	0.28 ± 0.21
18:1ω7	3.50 ± 0.98
18:1ω9	7.77 ± 1.43
20:1ω9	0.30 ± 0.48
Sum %	28.25 ± 2.62
Branched	
15:0a	1.27 ± 0.32
15:0i	0.93 ± 0.19
16:0i	0.34 ± 0.29
17:0a	0.22 ± 0.27
17:0i	1.03 ± 0.38
Sum %	3.79 ± 0.98
Polyunsaturated	
16:2	0.20 ± 0.28
16:3	0.93 ± 0.85
18:2ω6	1.22 ± 0.45
18:3ω3	0.77 ± 0.39
18:3ω6	0.26 ± 0.38
18:4ω3	0.87 ± 0.72
20:2	0.11 ± 0.19
20:4ω6	0.09 ± 0.15
20:5ω3	1.19 ± 0.85
22:3	0.37 ± 0.56
22:5ω3	0.03 ± 0.09
22:6ω3	0.42 ± 0.42
Sum %	6.46 ± 2.19
DHA/EPA	0.31 ± 0.29
Sum ω6	1.57 ± 0.75
Sum ω3	3.27 ± 2.00
ω3/ω6	2.56 ± 2.24
Sum n.i. %	0.65 ± 0.49
16:1ω7/16:0	0.23 ± 1.11

Data represent means and standard deviations of 13 samples.

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cetoleic acid (22:1 ω 11), a biomarker of predation on zooplankton (mainly copepods [65]). Feeding on zooplankton is plausible in *C. fornicata* especially when phytoplankton is not abundant [66], like in the winter months. These observed differences are thus likely explained by seasonal differences in food availability (less phytoplankton in November).

As food available to parents may influence the energy status of the embryos which mainly or even exclusively rely on resources provided by the mother, we studied the FA composition of the embryos. The FA composition of the early encapsulated embryos was closer to that of the visceral mass than to that of the foot. This observation is in agreement with previous results on the gastropod *Haliotis fulgens* by Nelson et al. [56] who showed that lipids are transferred from the hepatopancreas to the gonad to the larvae. In addition, the seasonal variations observed in the FA composition of the visceral mass were also observed in embryos at stage I, which suggested that parental feeding markedly influence the FA composition of the early embryos, as reported in other species [26,67].

Although females supplied the same proportion of total PUFA whatever the season to their embryos, we observed a preferential allocation of ω 3 to early embryos in the middle of the reproductive season, as shown by the higher proportion of ω 3 in embryos at stage I than in the visceral mass of the females, with the opposite trend for ω 6 (Figure 5E and F). The major PUFA in early embryos and visceral masses were EPA, DHA and 18:4 ω 3 for ω 3 and AA for ω 6. Among them, EPA was accumulated in early embryos as compared to visceral mass (Table 1). Although several invertebrates, including larval stages, are able to synthesize EPA from 18:3 ω 3 [68–70], our observation is more likely a preferential allocation as suggested by the similarity of the relative abundance of 18:3 ω 3 in visceral mass and early embryo (Table 1). Specific accumulation of EPA, known to be used as an energy source for development in molluscs, has already been observed in eggs of nudibranch species, irrespective of its presence in diet [26], but also in eggs of non-molluscan taxa (e.g. *Daphnia* [71]).

This preferential allocation of ω 3 to early embryos led to high ω 3/ ω 6 ratios. Even if PUFA requirements for embryo development in *C. fornicata* are not known, it is widely recognized that the essential ω 3 PUFA (especially EPA and DHA) are crucial for hatching success and larval growth and survival in bivalves [21–24,72–74], gastropods [25], and sea urchins [69,70,75]. Accumulation of ω 3 over ω 6 was reported for several species and the ω 3/ ω 6 ratio was considered an index of egg quality in several fishes and invertebrates [76–79]. Following such assumptions, our results showed that in early embryos of *C. fornicata*, the ω 3/ ω 6 ratio was twice higher in May and July as compared to February and September, and thus strongly suggested that early embryos had a higher quality in the middle of the reproductive season. Seasonal variations in egg quality have been demonstrated in some fish species with potential consequences on larval survival [79,80]. Although our data cannot allow us to assess such consequences, the observed changes in FA composition over the spawning season are expected to impact hatching success and larval performance in *C. fornicata*.

Several complementary hypotheses may explain the seasonal variations in maternal provisioning and propagule quality in *C. fornicata*, and more especially explain the lower quality at the beginning and end of the reproductive period. First, the food available for maternal provisioning varies seasonally, both quantitatively and qualitatively: at our study site, between September and March chlorophyll *a* concentration is low [41] with low abundances of nanoplankton and pelagic diatoms, rich in essential PUFA [81], and dominance of bacteria and cyanobacteria which are recognized as low quality food [28,81]. Second, *C. fornicata* females can reproduce several times during one reproductive season [31,42]. One may hypothesize that at the end of the reproductive season females will invest less energy to reproduce. This still has to be demonstrated in *C. fornicata*, but it has been shown that egg size and energy content may vary with brood order (e.g. [82,83]). Last, temperature might also influence maternal provisioning as has been shown through variations in egg size and hatching size in other calyptraeid species [84].

The above conclusions were drawn from a single reproductive period. To confirm these observations of the variations of maternal provisioning during the whole reproductive season would benefit from additional sampling at the same study site.

Variations in FA composition during intracapsular development

Lipids are the major source of energy used for embryonic and larval development in many marine invertebrates, including crustaceans [85–87] and molluscs [4,88]. In *C. fornicata* Pandian [16] estimated that lipids provided 65.3% of the energy required for early development. We thus expected changes in the FA composition of its embryos during intracapsular development. Embryos from stage I to IV have been analyzed at three sampling dates (May, July and September) and their FA composition varied gradually during their intracapsular development (Figures 7 and 8). These changes most likely occurred through the oxidative degradation of embryonic FA as source of energy, as in *C. fornicata* exogenous feeding is not likely because (1) the intracapsular fluid and capsular wall only contain low amounts of lipids in many gastropod species, including calyptraeids ([89–91], Table S1), and (2) feeding on extracapsular organic matter is not likely [35]. In addition, molluscs have a limited capacity to elongate or desaturate the PUFA 18:2 ω 6, 18:3 ω 3, 20:4 ω 6, 20:5 ω 3 and 22:6 ω 3 [23,26,29,92–94], suggesting that *de novo* synthesis is not likely.

During intracapsular development, we observed a decrease in all ω 3 except 22:5 ω 3 (DPA; docosapentaenoic acid) and 22:6 ω 3 in May and July (where we hypothesized a preferential allocation of ω 3, see above), leading to the embryos at stage IV reaching approximately the same ω 3/ ω 6 ratio (5.23 ± 1.6 ; $n=8$) at the three sampling dates. This highlighted the role of energy-supply of many ω 3 PUFA as reported previously [95]. In the opposite, DHA, DPA and ω 6 PUFA increased in proportion, suggesting they had a major structural function. In particular, this resulted in a decrease of the EPA/AA ratio

(except in September where it is low) as previously reported through embryonic development of crustaceans [96]. Similar results were observed during embryonic development of the bivalve *Patinopecten yessoensis*, Dall 1898, with a linear decrease in the content of the ω 3 PUFA, with the exception of 22:6 ω 3 [97]. In several molluscan species, 22:4 ω 6 and 22:5 ω 3 (DPA) have been reported to increase during embryogenesis and to be retained by larvae during starvation [26,95]. In our study, between 1 to 4% of 22:5 ω 3 (DPA) were found in embryos and larvae of *C. fornicata*, whereas 22:4 ω 6 was found in small amounts (0.09% to 0.32%), only in embryos. The need for these particular FAs is, however, species-dependent. For example, *Tapes semidecussata* (= *Venerupis philippinarum*, Reeve 1864) and *Mercenaria mercenaria*, Linnaeus 1758, needed 22:6 ω 3 (DHA) while *Crassostrea* sp., Sacco 1897, showed a fundamental need for 20:5 ω 3 (EPA) [95,98]. EPA has also been reported to decrease during embryonic and larval development in many bivalve species [4,26,95,99–101]. Finally, we observed a linear decrease (from approximately 5.0% to 3.0% in May, July, and September) of the MUFA 18:1 ω 7 which suggested a nutritive role, contrary to the observation of its linear increase during embryonic development of the pectinid *Crassadoma gigantea* by Whyte et al. [4], which suggested an essential structural role for this FA.

Larval feeding and seasonal variations in fatty acid composition of the larvae

The FA composition of pelagic larvae of *Crepidula fornicata* was highly variable depending on the sampling date and no clear seasonal pattern emerged (Table 3). Larvae sampled at our site mostly had a small size close to the hatching size. Thus we expected these larvae to exhibit a FA composition similar to that of late embryos (stage IV). However, larvae showed markedly different compositions, mostly due to the relative contributions of SFA and PUFA. The dominance of SFA, especially the palmitic (16: 0) and stearic (18: 0) acids in pelagic larvae has also been observed in larvae of other molluscan species [56,73] and has been assumed to indicate a non-optimal condition [102], despite the dietary interest of short SFA [103,104]. Whether the pelagic larvae of *C. fornicata* were in sub-optimal condition is not known. Newly-hatched veligers are able to feed on particles immediately upon release. They are also able to use dissolved organic matter, but free fatty acids have not been tested [33,35]. Thus it is unlikely that larvae of *C. fornicata* were starving. Alternatively, the higher 16: 0 and other SFA levels may indicate that larvae were feeding on SFA-rich particulate organic matter (POM). FA composition of the POM at the sampling site was indeed dominated by SFA (with more than 50% of total FA). An explanation for high levels of SFA in POM would be that

bacteria and cyanobacteria (*Synechococcus* sp.), both rich in SFA [20,105] are a dominant component of the microbial community in these waters [106]. Cyanobacteria are well recognized as low quality food for zooplankton [28,105,107] being depleted in PUFAs and sterols [74,105,108].

In conclusion, the FA compositions of female tissues and embryos of *C. fornicata* varied between seasons, with embryos of higher quality in May and July (middle of the reproductive period). Our study suggested a preferential allocation of ω 3 in May and July to embryos. In addition, the FA composition of embryos varied during intracapsular development (from stage I to stage IV), with different patterns depending on sampling dates. One consequence was that the latest stage reached the same quality regarding the ω 3/ ω 6 ratio. This suggested that the excess of ω 3 was consumed when reaching stage IV and that it favored only the intracapsular development. Replicating this sampling during longer term studies would allow confirming these observations. Future work might also include feeding experiments with various controlled food sources (e.g. different levels of PUFA) which would allow better understanding FA allocation to the developing embryos. Pelagic larvae sampled at our site showed FA profiles clearly different from those of embryos at stage IV, with a dominance of SFA as compared to early embryos and females. The SFA dominance might be due to availability of low quality food for larvae but this needs to be confirmed by experimental studies. Besides, the FA composition of larvae varied between the six sampling dates and further investigations are needed to link this composition to their available sources.

Supporting Information

Table S1. FA content (mg g⁻¹) of the capsule wall of *Crepidula fornicata*. Only FA representing 1% or more of the total FA content are indicated. Results were obtained from pooled capsule walls of a single female. (DOCX)

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Author Contributions

Conceived and designed the experiments: FL TM PR TC. Analyzed the data: FL TM PR TC. Wrote the manuscript: FL TM PR TC.

References

1. George SB (1990) Population and seasonal differences in egg quality of *Arbacia lixula* (Echinodermata: Echinoidea). Invertebr Reprod Dev 17: 111-121. doi:10.1080/07924259.1990.9672098.
2. George SB, Cellario C, Fenaux L (1990) Population differences in egg quality of *Arbacia lixula* (Echinodermata: Echinoidea): proximate composition of eggs and larval development. J Exp Mar Biol Ecol 141: 107-118. doi:10.1016/0022-0981(90)90217-Z.
3. Marshall DJ, Keough MJ (2008) The relationship between offspring size and performance in the sea. Am Nat 171: 214-224. doi:10.1086/524954. PubMed: 18197774.
4. Whyte JNC, Bourne N, Ginther NG (1990) Biochemical and energy changes during embryogenesis in the rock scallop *Crassadoma gigantea*. Mar Biol 106: 239-244. doi:10.1007/BF01314806.
5. Jaekle WB (1995) Variation in the size, energy content, and biochemical composition of invertebrate eggs: correlates to the mode of larval development. In: L McEdward. Ecology of marine invertebrate larvae. Boca Raton: CRC Press. pp. 49-77.
6. Bertram DF, Strathmann RR (1998) Effects of maternal and larval nutrition on growth and form of planktotrophic larvae. Ecology 79: 315-327. doi:10.1890/0012-9658(1998)079[0315:EOMALN]2.0.CO;2.
7. Olson RR, Olson MH (1989) Food limitation of planktotrophic marine invertebrate larvae: does it control recruitment success? Annu Rev Ecol Syst 20: 225-247. doi:10.1146/annurev.ecolsys.20.1.225.
8. Poorbagher H, Lamare MD, Barker MF (2010) The relative importance of parental nutrition and population versus larval diet on development and phenotypic plasticity of *Sclerasterias mollis* larvae. J Mar Biol Assoc UK 90: 527-536. doi:10.1017/S0025315409990907.
9. Smith BL, Bolton TF (2007) Consequences of natural variation in maternal investment and larval nutrition on larval growth of a marine polychaete. Mar Freshw Res 58: 1002-1007. doi:10.1071/MF06103.
10. de Jong-Westman M, Qian P-Y, March BE, Carefoot TH (1995) Artificial diets in sea urchin culture: effects of dietary protein level and other additives on egg quality, larval morphometrics, and larval survival in the green sea urchin, *Strongylocentrotus droebachiensis*. Can J Zool 73: 2080-2090. doi:10.1139/z95-245.
11. Gehman A-LM, Bingham BL (2010) Maternal diet and juvenile quality in the sea star *Leptasterias aequalis*. J Exp Mar Biol Ecol 386: 86-93. doi:10.1016/j.jembe.2010.01.017.
12. George SB (1999) Egg quality, larval growth and phenotypic plasticity in a forcipulate seastar. J Exp Mar Biol Ecol 237: 203-224. doi:10.1016/S0022-0981(98)00205-6.
13. Jarrett JN (2003) Seasonal variation in larval condition and postsettlement performance of the barnacle *Semibalanus balanoides*. Ecology 84: 384-390. doi:10.1890/0012-9658(2003)084[0384:SVILCA]2.0.CO;2.
14. Byrne M, Sewell MA, Prowse TAA (2008) Nutritional ecology of sea urchin larvae: influence of endogenous and exogenous nutrition on echinopluteal growth and phenotypic plasticity in *Triploneustes gratilla*. Funct Ecol 22: 643-648. doi:10.1111/j.1365-2435.2008.01427.x.
15. Sewell MA (2005) Utilization of lipids during early development of the sea urchin *Evechinus chloroticus*. Mar Ecol Prog Ser 304: 133-142. doi:10.3354/meps304133.
16. Pandian TJ (1969) Yolk utilization in the gastropod *Crepidula fornicata*. Mar Biol 3: 117-121. doi:10.1007/BF00353430.
17. Rosa R, Calado R, Andrade AM, Narciso L, Nunes ML (2005) Changes in amino acids and lipids during embryogenesis of European lobster, *Homarus gammarus* (Crustacea: Decapoda). Comp Biochem Physiol B 140: 241-249. doi:10.1016/j.cbpb.2005.01.002. PubMed: 15649771.
18. Tremblay R, Olivier F, Bourget E, Rittschof D (2007) Physiological condition of *Balanus amphitrite* cyprid larvae determines habitat selection success. Mar Ecol Prog Ser 340: 1-8. doi:10.3354/meps340001.
19. Utting SD, Millican PF (1997) Techniques for the hatchery conditioning of bivalve broodstocks and the subsequent effect on egg quality and larval viability. Aquaculture 155: 45-54. doi:10.1016/S0044-8486(97)00108-7.
20. Dalsgaard J, St John M, Kattner G, Müller-Navarra D, Hagen W (2003) Fatty acid trophic markers in the pelagic marine environment. Adv Mar Biol 46: 225-340. doi:10.1016/S0065-2881(03)46005-7. PubMed: 14601414.
21. Caers M, Utting SD, Coutteau P, Millican PF, Sorgeloos P (2002) Impact of the supplementation of a docosahexaenoic acid-rich emulsion on the reproductive output of oyster broodstock, *Crassostrea gigas*. Mar Biol 140: 1157-1166. doi:10.1007/s00227-002-0798-5.
22. Hendriks IE, van Duren LA, Herman PMJ (2003) Effect of dietary polyunsaturated fatty acids on reproductive output and larval growth of bivalves. J Exp Mar Biol Ecol 296: 199-213. doi:10.1016/S0022-0981(03)00323-X.
23. Nevejan N, Saez I, Gajardo G, Sorgeloos P (2003) Supplementation of EPA and DHA emulsions to a *Dunaliella tertiolecta* diet: effect on growth and lipid composition of scallop larvae, *Argopecten purpuratus* (Lamarck, 1819). Aquaculture 217: 613-632. doi:10.1016/S0044-8486(02)00585-9.
24. Soudant P, Marty Y, Moal J, Robert R, Quéré C et al. (1996) Effect of food fatty acid and sterol quality on *Pecten maximus* gonad composition and reproduction process. Aquaculture 143: 361-378. doi:10.1016/0044-8486(96)01276-8.
25. Bautista-Teruel MN, Millamena OM, Fermin AC (2001) Reproductive performance of hatchery-bred donkey's ear abalone, *Haliotis asinina*, Linne, fed natural and artificial diets. Aquac Res 32: 249-254. doi:10.1046/j.1355-557x.2001.00022.x.
26. Martínez-Pita I, García F, Pita M-L (2005) Fatty acid composition and utilization in developing eggs of some marine nudibranchs (Mollusca: Gastropoda: Opisthobranchia) from southwest Spain. J Shellfish Res 24: 1209-1216.
27. Ackman RG, Tocher CS, McLachlan J (1968) Marine phytoplankter fatty acids. J Fish Res Board Can 25: 1603-1620. doi:10.1139/f68-145.
28. Desvillettes C, Bourdier G, Amblard C, Barth B (1997) Use of fatty acids for the assessment of zooplankton grazing on bacteria, protozoans and microalgae. Freshw Biol 38: 629-637. doi:10.1046/j.1365-2427.1997.00241.x.
29. Dunstan GA, Volkman JK, Barrett SM, Leroi J-M, Jeffrey SW (1993) Essential polyunsaturated fatty acids from 14 species of diatom (Bacillariophyceae). Phytochemistry 35: 155-161. doi:10.1016/S0031-9422(00)90525-9.
30. Meziane T, Bodineau L, Retière C, Thoumelin G (1997) The use of lipid markers to define sources of organic matter in sediment and food web of the intertidal salt-marsh ecosystem of Mont-Saint-Michel Bay, France. J Sea Res 38: 47-58. doi:10.1016/S1385-1101(97)00035-X.
31. Conklin EG (1897) The embryology of *Crepidula*, a contribution to the cell lineage and early development of some marine gastropods. J Morphol 13: 1-226. doi:10.1002/jmor.1050130102.
32. Orton JH (1912) An account of the natural history of the slipper-limpet (*Crepidula fornicata*), with some remarks on its occurrence on the oyster grounds on the Essex coast. J Mar Biol Assoc UK 9: 437-443. doi:10.1017/S0025315400048372.
33. Brante A, Fernández M, Viard F (2009) Limiting factors to encapsulation: the combined effects of dissolved protein and oxygen availability on embryonic growth and survival of species with contrasting feeding strategies. J Exp Biol 212: 2287-2295. doi:10.1242/jeb.016329. PubMed: 19561219.
34. Hoagland KE (1986) Patterns of encapsulation and brooding in the Calyptraeidae (Prosobranchia: Mesogastropoda). Am Malacol Bull 4: 173-183.
35. Leroy F, Comtet T, Brante A, Leroux C, Riera P (2012) Can encapsulated embryos of *Crepidula fornicata* (L.) use extracapsular dissolved organic matter? An experimental study with a 13 C-enriched amino acid. J Molluscan Stud 78: 100-104. doi:10.1093/mollus/eyr044.
36. Chaparro OR, Oyarzun RF, Vergara AM, Thompson RJ (1999) Energy investment in nurse eggs and egg capsules in *Crepidula dilatata* Lamarck (Gastropoda, Calyptraeidae) and its influence on the hatching size of the juvenile. J Exp Mar Biol Ecol 232: 261-274. doi:10.1016/S0022-0981(98)00115-4.
37. Chaparro OR, Flores ML (2002) Reproductive output of *Crepidula fecunda* females: distribution of energy in the production of gametes and capsular walls. N Z J Mar Freshwat Res 36: 661-673. doi:10.1080/00288330.2002.9517122.
38. Collin R (2003) Worldwide patterns in mode of development in calyptraeid gastropods. Mar Ecol Prog Ser 247: 103-122. doi:10.3354/meps247103.
39. Hoagland KE (1979) The behavior of three sympatric species of *Crepidula* (Gastropoda: Prosobranchia) from the Atlantic, with implications for evolutionary ecology. Nautilus 94: 143-149.
40. Blanchard M (, Mar1997) Spread of the slipper limpet *Crepidula fornicata* (L., 1758) in Europe: current state and consequences. Sci, Mar 61: 109-118.
41. Leroy F (2011) Influence des conditions trophiques sur le développement larvaire de l'espèce invasive *Crepidula fornicata* - Conséquences sur ses capacités de dispersion. PhD thesis Paris: Université Pierre et Marie Curie. 210 p
42. Richard J, Huet M, Thouzeau G, Paulet Y-M (2006) Reproduction of the invasive slipper limpet, *Crepidula fornicata*, in the Bay of Brest, France. Mar Biol 149: 789-801. doi:10.1007/s00227-005-0157-4.
43. Rigal F (2009) Dynamique spatio-temporelle du nuage larvaire du gastéropode introduit *Crepidula fornicata* au sein d'une baie mégatidale, la baie de Morlaix (France). PhD thesis Paris: Université Pierre et Marie Curie. 160 p

44. Chipperfield PNJ (1951) The breeding of *Crepidula fornicata* (L.) in the river Blackwater, Essex. J Mar Biol Assoc UK 30: 49-71. doi:10.1017/S0025315400012571.
45. UNESCO (1968) Zooplankton sampling; Paris: UNESCO.. p. 174.
46. Werner B (1955) Über die Anatomie, die Entwicklung und Biologie des Veligers und der Veliconcha von *Crepidula fornicata* L. (Gastropoda Prosobranchia). Helgol Wiss Meeresunters 5: 169-217. doi:10.1007/BF01610508.
47. Thiriot-Quievreux C, Scheltema RS (1982) Planktonic larvae of New England gastropods. V. *Bittium alternatum*, *Triphora nigrocincta*, *Cerithiopsis emersoni*, *Lunatia heros* and *Crepidula plana*. Malacologia 23: 37-46.
48. Bligh EG, Dyer WJ (1959) A rapid method of total lipid extraction and purification. Can J Biochem Physiol 37: 911-917. doi:10.1139/o59-099. PubMed: 13671378.
49. Mezziane T, Tsuchiya M (2002) Organic matter in a subtropical mangrove-estuary subjected to wastewater discharge: origin and utilisation by two macrozoobenthic species. J Sea Res 47: 1-11. doi:10.1016/S1385-1101(01)00092-2.
50. Schomburg G (1987) Gaschromatographie: Grundlagen, Praxis, Kapillartechnik. Weinheim: Wiley-VCH Verlag.
51. Scherrer B (1984) Biostatistique. Chicoutimi: Gaëtan Morin éditeur: 850.
52. Clarke KR, Warwick RM (2001) Change in marine communities: an approach to statistical analysis and interpretation. Plymouth: PRIMER-E Ltd.
53. Clarke KR (1993) Non-parametric multivariate analyses of changes in community structure. Aust J Ecol 18: 117-143. doi:10.1111/j.1442-9993.1993.tb00438.x.
54. Marshall DJ, Bolton TF, Keough MJ (2003) Offspring size affects the post-metamorphic performance of a colonial marine invertebrate. Ecology 84: 3131-3137. doi:10.1890/02-0311.
55. Mateos HT, Lewandowski PA, Su XQ (2010) Seasonal variations of total lipid and fatty acid contents in muscle, gonad and digestive glands of farmed Jade Tiger hybrid abalone in Australia. Food Chem 123: 436-441. doi:10.1016/j.foodchem.2010.04.062.
56. Nelson MM, Leighton DL, Phleger CF, Nichols PD (2002) Comparison of growth and lipid composition in the green abalone, *Haliotis fulgens*, provided specific macroalgal diets. Comp Biochem Physiol B 131: 695-712. doi:10.1016/S1096-4959(02)00042-8. PubMed: 11923083.
57. Gardner D, Riley JP (1972) The component fatty acids of the lipids of some species of marine and freshwater molluscs. J Mar Biol Assoc UK 52: 827-838. doi:10.1017/S0025315400040571.
58. Pernet F, Bricelj VM, Parrish CC (2005) Effect of varying dietary levels of w6 polyunsaturated fatty acids during the early ontogeny of the sea scallop, *Placopecten magellanicus*. J Exp Mar Biol Ecol 327: 115-133. doi:10.1016/j.jembe.2005.06.008.
59. Jørgensen CB, Kiørboe T, Møhlenberg F, Riisgård HU (1984) Ciliary and mucus-net filter feeding, with special reference to fluid mechanical characteristics. Mar Ecol Prog Ser 15: 283-292. doi:10.3354/meps015283.
60. Kach DJ, Ward JE (2008) The role of marine aggregates in the ingestion of picoplankton-size particles by suspension-feeding molluscs. Mar Biol 153: 797-805. doi:10.1007/s00227-007-0852-4.
61. Lesser MP, Shumway SE, Cucci T, Smith J (1992) Impact of fouling organisms on mussel rope culture: interspecific competition for food among suspension-feeding invertebrates. J Exp Mar Biol Ecol 165: 91-102. doi:10.1016/0022-0981(92)90291-H.
62. Newell RC, Kofoed LH (1977) The energetics of suspension-feeding in the gastropod *Crepidula fornicata* L. J Mar Biol Ass U K 57: 161-180.
63. Beninger PG, Decottignies P, Guiheneuf F, Barillé L, Rincé Y (2007) Comparison of particle processing by two introduced suspension feeders: selection in *Crepidula fornicata* and *Crassostrea gigas*. Mar Ecol Prog Ser 334: 165-177. doi:10.3354/meps334165.
64. Decottignies P, Beninger PG, Rincé Y, Riera P (2007) Trophic interactions between two introduced suspension-feeders, *Crepidula fornicata* and *Crassostrea gigas*, are influenced by seasonal effects and qualitative selection capacity. J Exp Mar Biol Ecol 342: 231-241. doi:10.1016/j.jembe.2006.10.005.
65. Iverson SJ (2009) Tracing aquatic food webs using fatty acids: from qualitative indicators to quantitative determination. In: MT ArtsMT BrettM Kainz. Lipids in aquatic ecosystems. New York: Springer Verlag. pp. 281-307.
66. Pechenik JA, Blanchard M, Rotjan R (2004) Susceptibility of larval *Crepidula fornicata* to predation by suspension-feeding adults. J Exp Mar Biol Ecol 306: 75-94. doi:10.1016/j.jembe.2004.01.004.
67. Soudant P, Marty Y, Moal J, Samain J-F (1996) Fatty acids and egg quality in great scallop. Aquac Int 4: 191-200. doi:10.1007/BF00117381.
68. Kelly JR, Scheibling RE, Iverson SJ, Gagnon P (2008) Fatty acid profiles in the gonads of the sea urchin *Strongylocentrotus droebachiensis* on natural algal diets. Mar Ecol Prog Ser 373: 1-9. doi:10.3354/meps07746.
69. Liu H, Kelly MS, Cook EJ, Black K, Orr H et al. (2007) The effect of diet type on growth and fatty-acid composition of sea urchin larvae, I. *Paracentrotus lividus* (Lamarck, 1816) (Echinodermata). Aquaculture 264: 247-262. doi:10.1016/j.aquaculture.2006.12.021.
70. Liu H, Kelly MS, Cook EJ, Black K, Orr H et al. (2007) The effect of diet type on growth and fatty acid composition of the sea urchin larvae, II. *Psammechinus miliaris* (Gmelin). Aquaculture 264: 263-278. doi:10.1016/j.aquaculture.2006.12.022.
71. Becker C, Boersma M (2005) Differential effects of phosphorus and fatty acids on *Daphnia magna* growth and reproduction. Limnol Oceanogr 50: 388-397. doi:10.4319/lo.2005.50.1.0388.
72. Bernisson KM, Jonsson PR, Wängberg S-Å, Carlsson AS (1997) Effects of broodstock diets on fatty acid composition, survival and growth rates in larvae of the European flat oyster, *Ostrea edulis*. Aquaculture 154: 139-153. doi:10.1016/S0044-8486(97)00041-0.
73. Leonardos N, Lucas IAN (2000) The use of larval fatty acids as an index of growth in *Mytilus edulis* L. larvae. Aquaculture 184: 155-166. doi:10.1016/S0044-8486(99)00320-8.
74. Vanderploeg HA, Liebig JR, Gluck AA (1996) Evaluation of different phytoplankton for supporting development of zebra mussel larvae (*Dreissena polymorpha*): the importance of size and polyunsaturated fatty acid content. J Great Lakes Res 22: 36-45. doi:10.1016/S0380-1330(96)70932-2.
75. Carboni S, Vignier J, Chiantore M, Tocher DR, Migaud H (2012) Effects of dietary microalgae on growth, survival and fatty acid composition of sea urchin *Paracentrotus lividus* throughout larval development. Aquaculture 324-325: 250-258. doi:10.1016/j.aquaculture.2011.10.037.
76. Faleiro F, Narciso L (2010) Lipid dynamics during early development of *Hippocampus guttulatus* sea horses: searching for clues on fatty acid requirements. Aquaculture 307: 56-64. doi:10.1016/j.aquaculture.2010.07.005.
77. Lanes CFC, Bizuayehu TT, Bolla S, Martins C, de Oliveira Fernandes JM et al. (2012) Biochemical composition and performance of Atlantic cod (*Gadus morhua* L.) eggs and larvae obtained from farmed and wild broodstocks. Aquaculture 324-325: 267-275. doi:10.1016/j.aquaculture.2011.10.036.
78. Pettersen AK, Turchini GM, Jahangard S, Ingram BA, Sherman CDH (2010) Effects of different dietary microalgae on survival, growth, settlement and fatty acid composition of blue mussel (*Mytilus galloprovincialis*) larvae. Aquaculture 309: 115-124. doi:10.1016/j.aquaculture.2010.09.024.
79. Yanes-Roca C, Rhody N, Nystrom M, Main KL (2009) Effects of fatty acid composition and spawning season patterns on egg quality and larval survival in common snook (*Centropomus undecimalis*). Aquaculture 287: 335-340. doi:10.1016/j.aquaculture.2008.10.043.
80. Bachan MM, Fleming IA, Trippel EA (2012) Maternal allocation of lipid classes and fatty acids with seasonal egg production in Atlantic cod (*Gadus morhua*) of wild origin. Mar Biol 159: 2281-2297. doi:10.1007/s00227-012-2014-6.
81. Brett MT, Müller-Navarra DC (1997) The role of highly unsaturated fatty acids in aquatic foodweb processes. Freshw Biol 38: 483-499. doi:10.1046/j.1365-2427.1997.00220.x.
82. Collin R (2010) Repeatability of egg size in two marine gastropods: brood order and female size do not contribute to intraspecific variation. Mar Ecol Prog Ser 410: 89-96. doi:10.3354/meps08638.
83. Ito K (1997) Egg-size and -number variations related to maternal size and age, and the relationship between egg size and larval characteristics in an annual marine gastropod, *Halio japonica* (Opisthobranchia; Cephalaspidea). Mar Ecol Prog Ser 152: 187-195. doi:10.3354/meps152187.
84. Collin R, Salazar MZ (2010) Temperature-mediated plasticity and genetic differentiation in egg size and hatching size among populations of *Crepidula* (Gastropoda: Calyptraeidae). Biol J Linn Soc 99: 489-499. doi:10.1111/j.1095-8312.2009.01388.x.
85. Pandian TJ (1967) Changes in chemical composition and caloric content of developing eggs of the shrimp *Crangon crangon*. Helgol Wiss Meeresunters 16: 216-224. doi:10.1007/BF01611705.
86. Pandian TJ, Schumann KH (1967) Chemical composition and caloric content of egg and zoea of the hermit crab *Eupagurus bernhardus*. Helgol Wiss Meeresunters 16: 225-230. doi:10.1007/BF01611706.
87. Pandian TJ (1970) Ecophysiological studies on the developing eggs and embryos of the European lobster *Homarus gammarus*. Mar Biol 5: 154-167. doi:10.1007/BF00352599.

88. Gallager SM, Mann R, Sasaki GC (1986) Lipid as an index of growth and viability in three species of bivalve larvae. *Aquaculture* 56: 81-103. doi:10.1016/0044-8486(86)90020-7.
89. Miloslavich P (1996) Biochemical composition of prosobranch egg capsules. *J Molluscan Stud* 62: 133-135. doi:10.1093/mollus/62.1.133.
90. Ojeda JA, Chaparro OR (2004) Morphological, gravimetric, and biochemical changes in *Crepidula fecunda* (Gastropoda: Calyptraeidae) egg capsule walls during embryonic development. *Mar Biol* 144: 263-269. doi:10.1007/s00227-003-1194-5.
91. Sullivan CH, Mangel TK (1984) Formation, organization, and composition of the egg capsule of the marine gastropod, *Ilyanassa obsoleta*. *Biol Bull* 167: 378-389. doi:10.2307/1541284.
92. Delaunay F, Marty Y, Moal J, Samain J-F (1993) The effect of monospecific algal diets on growth and fatty acid composition of *Pecten maximus* (L.) larvae. *J Exp Mar Biol Ecol* 173: 163-179.
93. Laing I, Child AR, Janke A (1990) Nutritional value of dried algae diets for larvae of Manila clam (*Tapes philippinarum*). *J Mar Biol Assoc UK* 70: 1-12. doi:10.1017/S0025315400034147.
94. Waldock MJ, Holland DL (1984) Fatty acid metabolism in young oyster *Crassostrea gigas*: polyunsaturated fatty acids. *Lipids* 19: 332-336. doi:10.1007/BF02534783.
95. Labarta U, Fernández-Reiriz MJ, Pérez-Camacho A (1999) Dynamics of fatty acids in the larval development, metamorphosis and post-metamorphosis of *Ostrea edulis* (L.). *Comp Biochem Physiol A* 123: 249-254. doi:10.1016/S1095-6433(99)00054-9.
96. Torres P, Penha-Lopes G, Narciso L, Macia A, Paula J (2008) Fatty acids dynamics during embryonic development in genus *Uca* (Brachyura: Ocypodidae), from the mangroves of Inhaca Island, Mozambique. *Est Coast Shelf Sci* 80: 307-313. doi:10.1016/j.ecss.2008.07.018.
97. Whyte JNC, Bourne N, Hodgson CA (1989) Influence of algal diets on biochemical composition and energy reserves in *Patinopecten yessoensis* (Jay) larvae. *Aquaculture* 78: 333-347. doi:10.1016/0044-8486(89)90110-5.
98. Helm MM, Laing I (1987) Preliminary observations on the nutritional value of 'Tahiti *Isochrysis*' to bivalve larvae. *Aquaculture* 62: 281-288. doi:10.1016/0044-8486(87)90170-0.
99. Marty Y, Delaunay F, Moal J, Samain J-F (1992) Changes in the fatty acid composition of *Pecten maximus* (L.) during larval development. *J Exp Mar Biol Ecol* 163: 221-234.
100. Soudant P, Marty Y, Moal J, Masski H, Samain J-F (1998) Fatty acid composition of polar lipid classes during larval development of scallop *Pecten maximus* (L.). *Comp Biochem Physiol A* 121: 279-288.
101. Whyte JNC, Bourne N, Ginther NG (1991) Depletion of nutrient reserves during embryogenesis in the scallop *Patinopecten yessoensis* (Jay). *J Exp Mar Biol Ecol* 149: 67-79. doi:10.1016/0022-0981(91)90117-F.
102. Helm MM, Holland DL, Utting SD, East J (1991) Fatty acid composition of early non-feeding larvae of the European flat oyster *Ostrea edulis*. *J Mar Biol Assoc UK* 71: 675-691.
103. Pernet F, Tremblay R (2004) Effect of varying levels of dietary essential fatty acid during early ontogeny of the sea scallop *Placopecten magellanicus*. *J Exp Mar Biol Ecol* 310: 73-86. doi:10.1016/j.jembe.2004.04.001.
104. Thompson PA, Guo M, Harrison PJ (1993) The influence of irradiance on the biochemical composition of three phytoplankton species and their nutritional value for larvae of the Pacific oyster (*Crassostrea gigas*). *Mar Biol* 117: 259-268. doi:10.1007/BF00345671.
105. Martin-Creuzburg D, von Elert E (2009) Good food versus bad food: the role of sterols and polyunsaturated fatty acids in determining growth and reproduction of *Daphnia magna*. *Aquat Ecol* 43: 943-950. doi:10.1007/s10452-009-9239-6.
106. Leroy F, Riera P, Jeanthon C, Edmond F, Leroux C et al. (2012) Importance of bacterivory and preferential selection toward diatoms in larvae of *Crepidula fornicata* (L.) assessed by a dual stable isotope (¹³C, ¹⁵N) labeling approach. *J Sea Res* 70: 23-31. doi:10.1016/j.seares.2012.02.006.
107. Müller-Navarra DC, Brett MT, Liston AM, Goldman CR (2000) A highly unsaturated fatty acid predicts carbon transfer between primary producers and consumers. *Nature* 403: 74-77. doi:10.1038/47469. PubMed: 10638754.
108. Patil V, Källqvist T, Olsen E, Vogt G, Gislerød HR (2007) Fatty acid composition of 12 microalgae for possible use in aquaculture feed. *Aquac Int* 15: 1-9. doi:10.1007/s10499-006-9060-3.