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A Ruellan, Alain Guinault, Cyrille Sollogoub, G Chollet, A Ait-Mada, et al.. Industrial vegetable oil by-products increase the ductility of polylactide. Express Polymer Letters, 2015, 9 (12), pp.1087-1103. $10.3144/{\rm expresspolymlett.}2015.98$. hal-01218107

HAL Id: hal-01218107

https://hal.science/hal-01218107

Submitted on 20 Oct 2015

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A RUELLAN, Alain GUINAULT, Cyrille SOLLOGOUB, G CHOLLET, A AIT-MADA, V DUCRUET, S DOMENEK - Industrial vegetable oil by-products increase the ductility of polylactide - Express Polymer Letters - Vol. 9, p.1087-1103 - 2015

Industrial vegetable oil by-products increase the ductility of polylactide

A. Ruellan^{1,2}, A. Guinault¹, C. Sollogoub¹, G. Chollet³, A. Ait-Mada⁴, V. Ducruet^{2,4}, S. Domenek^{2,4*}

Abstract. The use of industrial by-products of the vegetable oil industry as ductility increasing additives of polylactide (PLA) was investigated. Vegetable oil deodorization condensates were melt-blended by twin-screw extrusion up to a maximum inclusion quantity of 20 wt% without preliminary purification. Sample films were obtained by single screw cast extrusion. Compounded PLA films featured largely improved ductility in tensile testing with an elongation at break up to 180%. The glass transition temperature remained higher than room temperature. The native mixture of molecules, which composed the deodorization condensates, had superior performance compared to a synthetic mixture of main compounds. The investigation of the correlation between composition of the additives and the ductility of the PLA blends by Principal Component Analysis showed synergy in property improvement between fatty acids having a melting point below and beyond the room temperature. Furthermore, a compatibilizing effect of molecules present in the native mixture was evidenced. Oil deodorization condensates, which are a price competitive by-product of the vegetable oil industry, are therefore a very promising biobased and biodegradable additive for improving the ductility of PLA.

Keywords: biodegradable polymers, poly(lactic acid), ductility, vegetable oil by-products, crazing

1. Introduction

Biodegradable and/or biobased polymers are widely studied to replace petroleum-based materials with the aim of contributing to higher sustainability of plastics. However, in many cases these new products still do not meet economical or functional requirements to be competitive in commodity applications. Polylactide (PLA) is one of the polymers which can potentially enter high volume markets, mostly in the food packaging [1–3] or textile [4] sector. Polylactide is a biobased and compostable polymer [5–7] issued from polymerization of lactic acid produced by fermentation of starch, potatoes or

beets [7, 8]. Although it has been known before [9], production cost reduction in the 1990s and 2000s thanks to breakthroughs in the polymerization technology [10, 11], made it economically competitive with petroleum-based materials. PLA offers many interests such as a glass transition higher than room temperature, ease of processing, high transparency, printability, glossy aspect [1, 12, 13]. However, its mechanical properties still remain an obstacle for many applications. In fact, PLA features high tensile strength and modulus but also high brittleness [14]. In order to enhance the PLA ductility, melt-blending with rubbery materials as poly(ether)urethane [15,

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16], polyamide elastomer [17], acrylonitrile-butadiene-styrene copolymer, or various impact modifiers [18–26] is effective. Furthermore, plasticizers such as citrate esters [27–37], polyethylene glycols [27, 33, 38–44] or manifold other molecules [29, 43, 45–48] can be used. Unfortunately, impact modifiers are often not biodegradable nor biobased and plasticizers are generally derived from fossil resources, degrading one important environmental advantage of employing PLA. Therefore biobased and biodegradable additives have been and are investigated for toughening PLA. Chemically modified vegetable oils have received important research interest, because ester or epoxy groups can be degraded by microorganisms [49], maintaining the biodegradability of blends with PLA. Polymerized soybean oil derivatives prepared by crosslinking double bonds of alkyl chains afforded ductility increase after meltblending with PLA and using a compatibilizer [50]. Improvements were also obtained with conjugated soybean oil which was reactively compatibilized with PLA by unsaturated triglycerides [51]. Epoxidized soybean oil [52-55] and epoxidized palm oil [56–58] also showed some positive effects on PLA toughness. Another approach consists in using rigid particles, which usually increased stiffness but is ineffective, or even detrimental for the ductility. Nonetheless, NatureWorks [59] reported the use of EMforce[®], a mineral additive, able to change the polymer failure mode from brittle to ductile. In fact, the efficient crack initiation in glassy PLA, which was provided by the well-dispersed additive with good interfacial adhesion to the matrix, enhanced the PLA elongation at break.

Expertise from plasticizing polyvinylchloride teaches that mixtures of different plasticizers are efficient for increasing toughness, because they make use of different mechanisms available for ductility improvement [49]. For example, a mixture of tributyl citrate (TBC) and a block copolymer PLA-g-polyethyleneglycol yielded a material with high elongation at break and satisfying stress at yield [30]. Al-Mulla et al. [55] successfully used modified nanoclays in combination with epoxidized soybean oil to increase stiffness and ductility. The combination of PLA with rubber and compatibilizers has also been investigated by several authors, showing important gains in ductility, with an elongation at break of up to 160% for high rubber concentrations [60] or bicontinous phases [61], and up to 200% when using

compatibilizers [62, 63]. However, these solutions often require the inclusion of several additives, chemically modified and costly components that increase the price of the final formulation, which is inappropriate for a large volume production. Finding a low cost biobased and biodegradable toughening agent would therefore help PLA to conquer new markets.

Vegetable oils contain a number of potentially interesting molecules for PLA toughening and the refinery of vegetable oils gives rise to several by-products. Among those, oil deodorization condensates represent a high-volume by-product, which is chemically close to the vegetable oil. They are obtained by distillation in the aim of purifying the raw oil from odorous compounds, which are negative for the sensorial properties of vegetable oils. Deodorization condensates contain free fatty acids, glycerides and unsaponifiable molecules. The objective of this study was to use vegetable oils and the byproduct deodorization condensate as produced for toughening PLA. The individual molecules which make up oils and deodorization condensates were tested on the mechanical properties and several vegetable oils were screened.

2. Experimental

2.1. Materials

PLA 4060D was supplied by NatureWorks (U.S.A.) and consists of $89\pm1\%$ L-lactic acid and $11\pm1\%$ D-lactic acid units, making it unable to crystallize under common conditions [1, 64]. The glass transition temperature ($T_{\rm g}$) obtained from differential scanning calorimetry (DSC) measurements was 56°C. Average molecular weights obtained from size exclusion chromatography (SEC) measurements were $M_{\rm w} = 236\,800~{\rm g\cdot mol^{-1}}$, $M_{\rm n} = 103\,700~{\rm g\cdot mol^{-1}}$ and dispersity $M_{\rm w}/M_{\rm n} = 2,28$.

Hydrogenated palm oil (HPO), Hydrogenated Copra Oil (HCO), Rapeseed Oil Deodorization Condensate (RODC), Soybean Oil Deodorization Condensate (SODC), Olive Oil Deodorization Condensate (OODC) and Palm Oil Deodorization Condensate (PODC) were supplied by ITERG (Bordeaux, France).

Palmitic acid (purity >98%) *i.e.* hexadecanoic acid (C16:0), oleic acid (purity >96%) *i.e.* 9-octadecenoic acid (C18:1), squalene (purity >98%) *i.e.* (6E,10E,14E,18E)-2,6,10,15,19,23-hexamethyltetracosa-2,6,10,14,18,22-hexaene, and alpha-toco-

pherol (purity >96%) *i.e.* (2R)-2,5,7,8-tetramethyl-2-[(4R,8R)-(4,8,12-trimethyltridecyl)]-6-chromanol, were supplied by Sigma-Aldrich (France).

2.2. Film fabrication

Prior to melt-blending, PLA pellets were dried at 60°C for 24 h under dried air using a SOMOS 60L. Relative humidity of dried pellets was controlled to be lower than 350 ppm using an Aboni FMX Hydrotracer (France). Melt-mixing of PLA with or without additives was carried out using a twin screw extruder (Thermo Haake Ptw 16-40D, France), having a screw diameter of 16 mm and a length to diameter ratio (*L/D*) 40:1. A temperature of 180°C was used to process by twin-screw extrusion the PLA pellets as reference material, while the temperature profile of the 7 heating zones decreased from 180 to about 150/130°C along the extrusion flow, depending on the nature and the amount of additive. Screw

speed was 300 rpm. The exact temperature profile for each formulation is given in Table 1. To properly control the feed rate of additives, which are solid at room temperature, a home-made apparatus consisting in a heated syringe was used. Obtained pellets were stored into hermetic sealed metalized bags to avoid PLA rehydration.

Films of about 0.8 mm thickness containing palmitic acid, oleic acid, hydrogenated palm oil, hydrogenated copra oil, squalene, α -tocopherol and some combinations of these products were obtained using a single screw extruder (Scamex Rheoscam, France), mounted with a screw of 20 mm diameter and a length to diameter ratio (L/D) 12:1 and a flat die of 40 mm width and 1 mm thickness. Films were stretched and cooled with chill rolls. Due to the short length of the screw, an increasing temperature profile from 180 to 195°C over the 3 heating zones was used for PLA, while it was decreasing from 180

Table 1. Processing conditions of melt-mixing twin screw extrusion

	Formulation	Temperature profile [°C]
Neat PI	LA	175/180/190/190/190/190/180
PLA+	10 wt% C16:0	175/180/180/170/170/170/170
PLA+	10 wt% C18:1	175/180/180/170/170/170/170
PLA+	10 wt% [50 wt% C16:0 + 50 wt% C18:1]	175/180/180/170/170/170/170
PLA+	10 wt% [95 wt% (50 wt% C16:0 + 50 wt% C18:1) + 5 wt% HPO]	175/180/180/170/170/170/170
	5 wt% HCO	175/180/180/180/180/170/170
PLA+	10 wt% HCO	175/180/180/180/180/170/170
	15 wt% HCO	175/180/180/180/170/170/160
-	5 wt% HPO	175/180/180/180/180/170/170
PLA+	10 wt% HCO	175/180/180/180/180/170/170
	15 wt% HCO	175/180/180/180/180/170/170
	5 wt% α-tocopherol	175/180/180/180/180/180/180
PLA+	10 wt% α-tocopherol	175/180/180/180/180/180/170
	15 wt% α-tocopherol	175/180/180/180/180/170/170
	5 wt% squalene	175/180/180/180/180/180/180
PLA+	10 wt% squalene	175/180/180/180/180/170/170
	15 wt% squalene	175/180/180/180/180/170/170
	5 wt% RODC	175/180/180/180/180/175/170
PLA+	10 wt% RODC	175/180/180/180/180/170/160
TLA	15 wt% RODC	175/180/180/180/180/170/160
	20 wt% RODC	175/180/180/180/170/160/150
	5 wt% SODC	175/180/180/180/180/175/170
PLA +	10 wt% SODC	175/180/180/180/180/170/160
TLA	15 wt% SODC	175/180/180/180/180/170/160
	20 wt% SODC	175/180/180/180/170/160/150
	5 wt% OODC	175/180/180/170/170/170/170
PLA+	10 wt% OODC	175/180/180/170/170/170/170
PLA +	15 wt% OODC	175/180/180/170/170/160/160
	20 wt% OODC	175/180/180/170/160/150/150
	5 wt% PODC	175/180/180/180/180/175/170
PLA +	10 wt% PODC	175/180/180/180/170/160/155
ı LA	15 wt% PODC	175/180/180/180/170/160/150
	20 wt% PODC	175/180/180/170/160/150/135

Table 2. Processing conditions of cast films extrusion

	Formulation	Temperature profile [°C]	Die temperature [°C]
Neat PI	A	180/190/195	185
PLA +	10 wt% C16:0	180/180/175	170
PLA +	10 wt% C18:1	180/180/175	170
PLA +	10 wt% [50 wt% C16:0 + 50 wt% C18:1]	180/180/175	170
PLA +	10 wt% [95 wt% (50 wt% C16:0 + 50 wt% C18:1) + 5 wt% HPO]	180/180/175	170
	5 wt% HCO	180/180/180	180
PLA+	10 wt% HCO	180/170/170	170
	15 wt% HCO	180/170/170	170
	5 wt% HPO	180/180/180	170
PLA+	10 wt% HPO	180/170/170	170
	15 wt% HPO	180/170/170	170
	5 wt% α-tocopherol	180/180/180	180
PLA+	10 wt% α-tocopherol	180/180/180	180
	15 wt% α-tocopherol	180/175/170	170
	5 wt% squalene	180/180/180	180
PLA+	10 wt% squalene	180/180/175	170
	15 wt% squalene	180/180/170	170

to about 170°C for blends. The specific temperatures depending on the nature and the amount of additive are given in Table 2.

In the case of the deodorization condensates of the vegetable oils (RODC, SODC, OODC, PODC), the available quantity was insufficient for carrying out a single screw extrusion in the aim to obtain films. In that case PLA sheets of 1 mm thickness were thermo-moulded by compression at 220 bar (Laboratory Press Gibitre Instruments 20 tons, Italy). For this, pellets were pre-melted at 180°C without pressure during 180 seconds then heating plates were closed with progressive increase in pressure during 120 seconds to eliminate air bubbles.

2.3. Chemical and physical chemical characterization of vegetable oils and deodorization condensates

Glyceride composition of fats was determined according to the IUPAC 6.002 and EN 14105 standards using a Shimadzu GC-2010 Plus gas chromatograph (France) equipped with a Zebron ZB 5 HT Inferno (15 m, 0.25 mm, 0.1 μm) column and a flame ionization detector set at 380°C. The vector gas was H₂ at a flow rate of 1.17 mL·min⁻¹. Both the injector and the oven temperature were set at 60°C for 3 min, then raised to 370°C at 10°C·min⁻¹ and held at 370°C for 12 min. Direct on-column injection was performed.

Fatty acid composition was determined according to the ISO 12966-2 standard using a Shimadzu GC-

2010 Plus gas chromatograph (France) equipped with a BPX70 (50 m, 0.22 mm, 0.25 μ m) column, and a flame ionization detector set at 250°C. The vector gas was H₂ at a flow rate of 0.32 mL·min⁻¹. The oven temperature was set at 60 C for 2 min, raised to 170°C at 20°C·min⁻¹, held at 170°C for 25 min, raised to 230°C at 4°C·min⁻¹ and held at 230°C for 10 min. The injector temperature was set at 250°C and a split ratio of 200 was used.

Acid value was determined according to the ISO 660 standard using a mixture of ethanol 95% and diethylic ether as solvent, potassium hydroxide 0.5 mol·L⁻¹ in ethanol 95% as titrant and alkali blue 6B as indicator.

Saponification value was determined according to the ISO 3657 standard. Samples were saponified with potassium hydroxide 0.5 mol·L⁻¹ in ethanol 95% boiled under reflux during 2 h. Hydrochloric acid 0.5 mol·L⁻¹ was used as titrant and alkali blue 6B as indicator.

Water content was measured using a Mettler Toledo HB43 S Halogen Moisture Analyzer set (France) at 103°C.

Melting point of free fatty acids was determined from literature [65]. Estimation of mono, di- and triglycerides average chemical structure of oil deodorization condensates was based on the averages unsaturation number and the alkyl chains length of the corresponding free fatty acids composition profile (Table 3). Average glycerides melting point was estimated calculating a weighted average of the

Table 3. Comp	position of hydro	genated vegetable	oils and vegetable	oil deodorization condensates

Type			HPO	НСО	RODC	SODC	OODC	PODC
	Free fatty acids		0.0	0.0	39.0	43.1	39.2	95.4
	Monoglycerides		0.2	0.0	11.2	2.9	2.0	1.7
ide	Diglycerides		10.6	0.0	2.7	9.0	8.4	2.2
Glyceride composition [%]	Triglycerides		88.9	100	12.2	16.8	33.5	0.7
Januar Ja	Sterols (α-tocopher	rol)	0.0	0.0	25.7	9.8	1.6	0.0
	Hydrocarbons (Squ	ialene)	0.0	0.0	0.0	13.8	13.0	0.0
	Unidentified		0.3	0.0	9.2	4.6	2.3	0.0
Acid value [m	ig KOH·g ^{−1}]		0.1	0.1	65.3	68.4	47.7	201.0
Saponification	n value [mg KOH·g-	1]	198.6	255.2	125.1	157.9	162.2	205.7
Water content	[%]		0.11	0.10	0.55	0.34	0.37	0.16
	Caproic acid	C6:0	0.0	0.5	0.0	0.0	0.0	0.0
	Caprylic acid	C8:0	0.0	6.8	0.0	0.0	0.0	0.0
	Capric acid	C10:0	0.0	5.7	0.0	0.0	0.1	0.0
	Lauric acid	C12:0	0.5	47.5	0.0	0.8	0.0	0.4
l uc	Myristic acid	C14:0	1.2	18.1	0.0	0.4	0.0	1.3
siti	Palmitic acid	C16:0	43.6	9.4	7.4	12.3	11.3	49.8
od	Palmitoleic acid	C16:1	0.0	0.0	0.0	0.0	0.0	0.2
om []	Stearic acid	C18:0	53.8	10.8	3.4	4.1	2.5	4.1
d cor	Oleic acid	C18:1	0.0	1.0	27.3	21.7	69.5	35.2
aci	Linoleic acid	C18:2	0.0	0.0	42.4	49.7	10.9	7.8
Fatty acid composition [%]	Linolenic acid	C18:3	0.0	0.0	1.5	6.4	0.6	0.3
	Arachidic acid	C20:0	0.5	0.1	0.5	0.3	0.4	0.3
	Eicosenoic acid	C20:1	0.0	0.0	0.3	0.2	0.4	0.1
	Behenic acid	C22:0	0.1	0.0	1.0	0.5	0.0	0.0
	Lignoceric acid	C24:0	0.1	0.0	0.5	0.2	0.1	0.0
	Unidentified		0.5	0.1	0.2	15.7	13.4	4.2
Average alkyl	chain carbon quanti	ty	17.1	13.0	17.9	17.7	17.8	16.9
Average alkyl	chain unsaturation of	quantity	0.0	0.0	1.4	1.5	1.0	0.5

specific melting points of the contained molecules taken from references [65–67]. Physical properties of molecules are given in Table 4.

2.4. Calculation of solubility parameters

Molar volumes and molar attraction constants of polylactide and additives were determined according to the van Krevelen and Hoftyzer atomic group contribution method [68]. Average molar volumes and average molar attraction constants of mono, di and triglycerides of oil deodorization condensates were estimated based on average chemical structures previously determined.

Hansen Solubility parameters were calculated using Equations (1), (2) and (3) [69, 70]. Used molar constant values [68] are presented in Table 5:

$$\delta_{\rm d} = \frac{\sum F_{\rm dt}}{\sum V_{\rm i}} \tag{1}$$

$$\delta_{\mathbf{p}} = \frac{\sqrt{\sum F_{\mathbf{Pi}}^2}}{\sum V_{\mathbf{i}}} \tag{2}$$

$$\delta_{\rm h} = \frac{\sqrt{\sum E_{\rm hi}}}{\sum V_{\rm i}} \tag{3}$$

where $\delta_{\rm d}$ is the dispersion component of the solubility parameter in J^{1/2}·cm^{-3/2}, $\delta_{\rm p}$ the polar component of the solubility parameter in J^{1/2}·cm^{-3/2}, $\delta_{\rm h}$ the hydrogen bonding component of the solubility parameter in J^{1/2}·cm^{-3/2}, $F_{\rm d}$ the dispersion contribution of the molar attraction constant in (J^{1/2}·cm^{-3/2})·mol⁻¹, $F_{\rm p}$ the polar contribution of the molar attraction constant in (J^{1/2}·cm^{-3/2})·mol⁻¹, $E_{\rm h}$ the hydrogen bonding energy contribution of the molar attraction constant in J·mol⁻¹ and V the molar volume contribution of the chemical group involved in cm³·mol⁻¹.

The solubility of the molecules in PLA (Table 4) was assessed using the HSP Relative Energy Difference (RED) from Equations (4) and (5):

$$Distance = \sqrt{4(\delta_{\text{dmolec}} - \delta_{\text{dPLA}})^2 + (\delta_{\text{pmolec}} - \delta_{\text{pPLA}})^2 + (\delta_{\text{hmolec}} - \delta_{\text{hPLA}})^2}$$
(4)

$$RED = \frac{Distance}{Radius} \tag{5}$$

Table 4. Physical properties and solubility parameters of fatty acids, mono-, di-, and triglycerides contained in the hydrogenated vegetable oils and oil deodorization condensates

				Molor woight	Molon wolumo	Melting		Hansen Solt	Hansen Solubility Parameters (HSP)	(HSP)	
ceach Choice - - - - - (mides) 0.99 6.0 (mides) in each CRO 116 1.5 1.5 1.5 1.6 1.6 2.65 7.24 8.89 8.89 in cach CRO 117 1.9 1.6 1.65 2.65 7.24 9.05 9.4 in cach CRO 2.0 2.7 2.5 1.64 1.46 5.00 9.4 9.05 in cach CRO 2.2 2.5 5.5 1.64 1.46 5.00 9.4 9.05 9.4 9.05 9.4 9.05 9.4 9.05 9.05 9.4 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05 9.05				[g·mol ⁻¹]	[cm ³ ·mol ⁻¹]	point [°C]	$\delta_{ m d}$ $[{f J}^{1/2}{ m cm}^{-3/2}]$	$\begin{bmatrix} \delta_{\rm p} \\ [J^{1/2}\text{-cm}^{-3/2}] \end{bmatrix}$	$\delta_{\rm h}$ $[{\bf J}^{1/2}{\cdot}{\bf cm}^{-3/2}]$	Distance with PLA	RED with PLA
Capprise and C 6 0 114 126 -3° 160 23.2 88.9 88.9 Capprise and C 100 C 100 114 126 -3° 16.00 2.65 7.94 8.90 Capprise and C 100 C 112 191 146 16.30 2.05 7.94 8.90 Aystric and C 120 2.08 2.25 2.87 16.40 1.46 5.90 9.46 Palminic and C 180 2.85 2.87 6.49 1.46 5.90 9.46 Palminic and C 180 2.85 2.87 6.49 1.46 5.90 9.49 Palminic and C 180 2.85 2.87 2.87 1.64 1.13 5.90 9.49 Palminic and C 180 2.85 3.04 -4° 16.35 1.13 5.90 9.86 Anachic and C 200 3.13 3.85 3.44 8.49 16.45 1.14 5.95 9.87 Posteric and C 200 3.13 3.45 3.44 1.65 1.19 4	PLA	1 4060D		242 800	ı	ı	18.6	6.6	0.9	(radius: 10.7)	ı
Capyin acid CROPA 114 1159 116 16.30 2.65 794 8.91 Capyin acid CROPA 114 1159 116 16.30 2.05 724 9.05 Lauric acid C120 228 225 535 16.40 1.65 6.70 9.04 Myristic acid C120 226 2285 225 13 16.40 1.65 1.90 9.04 Philmiotic acid C181 254 228 226 1.74 16.40 1.30 559 9.85 Plantinotic acid C181 228 3.04 -1.4* 16.49 1.31 559 9.85 Olici acid C181 228 3.04 -1.4* 16.40 1.14 9.71 Raminic acid C182 228 3.04 -1.4* 16.54 1.14 9.71 Colici acid C182 228 3.04 -1.4* 16.54 1.14 16.94 9.74 Li		Caproic acid	C6:0	116	126	-3a	16.06	3.32	8.89	8.80	0.82
Caming and C100 T172 191 31* 16.30 2.00 7.24 9.05 Caming and C140 C100 172 191 31* 16.30 2.00 9.30 Palmits and C140 228 255 55* 65* 146 5.90 9.46 Palmits and C160 16.0 224 225 255 149 16.40 1146 5.90 9.46 Palmits and C181 283 320 77* 16.27 1146 5.90 9.46 Palmits and C182 283 320 77* 16.23 113 5.59 9.46 Debracada C182 280 340 14 15.24 113 5.69 9.46 Inchore and C201 313 341 347 342 16.22 113 5.69 9.46 Elocasons and C201 31 341 347 344 344 344 344 344 344 344 344 344 344 344		Caprylic acid	C8:0	144	159	16 ^a	16.20	2.65	7.94	8.91	0.83
Mystic acid C12.0 200 223 44* 16.77 188 6.70 9.20 Mystic acid C14.0 256 287 63* 164 1.46 5.90 9.44 Pulmite acid C16.0 256 287 63* 164 1.46 5.90 9.46 Shanti acid C16.1 284 320 79* 164.9 1.34 5.95 9.46 Shanti acid C18.1 283 314 13* 16.49 1.34 5.96 9.46 Linoles acid C18.1 283 314 13* 16.49 1.34 5.64 9.71 Linoles acid C18.1 283 314 13* 1.4* 15.54 1.36 5.64 9.71 Linoles acid C18.1 283 314 4.4* 16.54 1.19 5.54 9.71 Linoles acid C18.1 31 34 34 4.4* 16.54 1.19 5.94 9.8 </td <th></th> <td>Capric acid</td> <td>C10:0</td> <td>172</td> <td>191</td> <td>31a</td> <td>16.30</td> <td>2.20</td> <td>7.24</td> <td>9.05</td> <td>0.85</td>		Capric acid	C10:0	172	191	31a	16.30	2.20	7.24	9.05	0.85
Mysicia acid C140 228 255 559 1642 1.65 6.05 9.34 Palminicatic acid C160 256 227 63° 1649 1.16 5.09 946 Palminicatic acid C18.0 285 287 17° 16.49 1.31 5.59 9.58 Steam card C18.1 283 349 19° 16.49 1.34 5.59 9.58 Linoletic acid C18.1 283 349 14° 16.49 1.34 5.69 9.86 Linoletic acid C18.2 228 349 14° 16.42 1.14 5.99 9.86 Linoletic acid C18.2 228 16.44 1.15 5.69 9.86 Linoletic acid C18.2 239 16.44 1.15 5.93 9.86 Inchestic acid C220 341 347 289 16.53 1.10 4.90 9.86 Bebenic acid C220 341 346 <th></th> <td>Lauric acid</td> <td>C12:0</td> <td>200</td> <td>223</td> <td>44ª</td> <td>16.37</td> <td>1.88</td> <td>6.70</td> <td>9.20</td> <td>98.0</td>		Lauric acid	C12:0	200	223	44ª	16.37	1.88	6.70	9.20	98.0
Palminic and C16.0 C16.0 256 287 63° 1646 1.46 5.59 946 Palminic and C18.0 C18.0 254 222 1° 1627 1.49 5.59 9.62 Steare and C18.1 C18.0 283 320 77° 16.47 1.34 5.59 9.62 Ober and C18.1 283 320 77° 16.27 1.34 5.59 9.62 Ober and C18.3 287 320 77° 16.23 1.34 5.56 9.73 Linolevia and C18.3 278 340 -14° 15.55 1.38 5.74 10.08 Anachida and C22.0 341 342 278 16.52 1.19 5.37 9.68 Linolevia and C22.0 341 346 84 16.54 1.19 5.37 9.68 Linolevia and C22.0 440 16.52 1.11 4.99 9.73 Behanic and C24.0 340 416 84-76 11.14 1.24 <t< td=""><th></th><td>Myristic acid</td><td>C14:0</td><td>228</td><td>255</td><td>55a</td><td>16.42</td><td>1.65</td><td>6.26</td><td>9.34</td><td>0.87</td></t<>		Myristic acid	C14:0	228	255	55a	16.42	1.65	6.26	9.34	0.87
Particle of Cl61 254 222 1° 1627 149 595 962 Particle of Cl81 Cl80 285 320 70° 1627 1131 559 962 Olete acid Cl81 283 334 19° 1632 1131 559 962 Olete acid Cl82 280 349 17 4 559 962 Linolec acid CR0 313 349 19° 1632 1134 559 962 Linolec acid CR0 4 18° 16.32 1.34 569 978 Anachidic acid CR0 4 18° 16.32 1.34 574 10.03 Anachidic acid C20.0 311 347 22° 16.36 1.13 574 9.86 Anachidic acid C20.0 31 34 34 34° 34° 16.36 1.11 5.37 9.86 Bebraic acid C20.0 34 34	sbi	Palmitic acid	C16:0	256	287	63a	16.46	1.46	5.90	9.46	0.88
Steartic and Cl R80 Cl R80 285 314 170* 1644 1.31 5.59 9.58 Linableic acid Cl R21 Cl R21 283 314 170* 16.14 5.69 9.71 Linableic acid Cl R23 CR R3 378 -9* 16.14 1.36 5.69 9.71 Amchilein acid Cl CR Cl R33 313 32.4 75* 16.36 1.13 5.69 9.71 Amchilein acid Cl CR	ge.	Palmitoleic acid	C16:1	254	282	1a	16.27	1.49	5.95	9.62	06.0
Obleic acid CR81 283 314 13° 1632 1.34 564 971 Obleic acid CR81 283 309 -9° 1614 1.36 564 971 Lunolenic acid CR82 288 309 -9° 1614 1.36 564 971 Lunolenic acid CR90 313 352 75° 16.52 1.19 5.74 10.08 Eicocanolic acid C200 341 347 23° 16.54 1.19 5.37 9.68 Bebraic acid C210 341 344 88° 16.54 1.19 5.37 9.68 Bobbenic acid C220 341 344 88° 16.54 1.10 490 9.78 Bobbenic acid C240 11.1 2.44 11.65 9.88 9.88 Bobbenic acid C240 11.1 3.44 3.44 3.44 3.44 1.14 2.44 11.09 5.14 10.00	YIIE	Stearic acid	C18:0	285	320	70ª	16.49	1.31	5.59	9.58	0.89
Linolice acid C18.2 280 309 -9* 16.14 1.36 5.69 9.86 Linolice acid C18.3 278 304 -14* 15.95 1.19 5.59 9.86 Linoler acid C20.1 311 347 23* 16.54 1.09 5.37 9.80 Eiczeenore acid C20.1 341 344 80* 16.54 1.01 4.90 9.77 Bebenic acid C24.0 369 416 84* 16.54 1.01 4.90 9.80 Bebenic acid C24.0 369 416 84* 16.54 1.01 4.90 9.77 Bebenic acid C24.0 16.0 4.90 3.40 4.6 8.76 1.71 2.49 11.66 9.77 RODC (HO)-Gl-Cl-Cl-Cl-Qi-Cl-Cl-Si-LS) 351 352 25-22* 16.98 2.44 11.65 9.83 RODC (HO)-Gl-Cl-Cl-Cl-Gi-Gi-Gi-Gi-Gi-Gi-Gi-Gi-Gi-Gi-Gi-Gi-Gi-	3 ə	Oleic acid	C18:1	283	314	13a	16.32	1.34	5.64	9.71	0.91
Linolenic acid C18.3 278 304 -14° 15.95 1.38 574 1003 Acarchide acid C20.0 313 352 75° 16.52 1.19 5.33 968 Acarchide acid C20.0 311 347 23° 16.36 1.19 5.33 968 Bebenic acid C22.0 341 384 80° 16.54 1.09 5.30 988 Lignoceric acid C22.0 369 416 88-76 17.14 2.49 11.66 9.77 HPO (HO)-CGIA-CI7.3:1.5 351 384 88-18° 16.56 2.45 11.66 9.87 SODC (HO)-CGIA-CI7.8:1.5 351 348 8-18° 16.56 2.47 11.66 9.84 SODC (HO)-CGIA-CI7.10:0 354 35.2 16.59 1.34 10.00 8.94 SODC (HO)-CGIA-CI7.10:0 35.2 22-12° 16.59 1.34 9.84 RODC (HO)-C	Fre	Linoleic acid	C18:2	280	309	-9a	16.14	1.36	5.69	98.6	0.92
Anethide acid CDO 313 352 75° 16.52 119 5.33 968 Eleosenoi acid C20:1 311 347 32° 16.36 1191 5.37 968 Behnine acid C20:1 341 384 84° 16.36 1.01 4.90 9.77 Behnine acid C20:0 346 68-76° 17.14 2.49 11.66 9.77 HPO (HO)-Gly-(C17.5:1.5) 354 351 10-22° 16.92 2.45 11.66 9.77 SODC (HO)-Gly-(C17.5:1.5) 354 354 354 352 2.45 11.66 9.77 SODC (HO)-Gly-(C17.8:1.0) 354 354 354 352 2.45 11.65 9.83 PODC (HO)-Gly-(C17.8:1.0) 342 340 45-55°c 17.07 2.53 10.00 8.93 RODC (HO)-Gly-(C17.8:1.0) 61 63 65-12°c 1.85 1.33 7.27 9.56		Linolenic acid	C18:3	278	304	-14ª	15.95	1.38	5.74	10.03	0.94
Eicosenoic acid C20-1 341 347 23° and 16.54 10.1 5.37 980 Ligheneric acid C22:0 369 416 84° and 16.55 11.0 4.00 9.87 Lighoceric acid C22:0 369 416 84° and 16.55 1.09 5.10 9.80 RODC (HO)r-Gly-C(17.51.5) 354 354 354 354 354 11.56 9.89 SODC (HO)r-Gly-C(17.51.5) 354 354 354 354 354 354 11.56 9.89 SODC (HO)r-Gly-C(17.51.5) 354 354 354 354 354 354 354 354 354 368 368 344 11.65 247 11.66 9.89 374 9.89 374 9.81 9.84 9.84 9.84 11.65 247 11.66 9.89 9.84 9.84 9.84 11.65 11.65 9.89 9.84 9.84 11.65 11.65 11.65 11.65		Arachidic acid	C20:0	313	352	75ª	16.52	1.19	5.33	89.6	06.0
Behenic acid C22.0 341 384 80° and a control of cont		Eicosenoic acid	C20:1	311	347	23ª	16.36	1.21	5.37	9.80	0.92
Lignoceric acid C24.0 369 416 84° 16.55 1.01 4.90 9.85 HPO (HO)z-Gly-(C17.1.0) 346 346 68-76° 17.14 2.49 11.66 9.77 RODC (HO)z-Gly-(C17.9.1.4) 354 351 10-22° 16.92 2.45 11.66 9.77 RODC (HO)z-Gly-(C17.8.1.6) 342 342 342 16.82 2.47 11.65 9.89 PODC (HO)z-Gly-(C17.8.1.0) 342 342 32.32 25-32° 16.82 2.47 11.63 9.89 PODC (HO)z-Gly-(C17.8.1.0) 342 342 342 45-55° 17.07 2.33 10.00 8.93 PODC (HO)z-Gly-(C17.1.0) 60 630 65-27° 16.82 1.33 7.34 9.34 RODC (HO)-Gly-(C17.8.1.0) 61 642 2-12° 16.83 1.33 7.41 9.44 RODC (HO)-Gly-(C17.8.1.0) 881 7.48° 16.4		Behenic acid	C22:0	341	384	80^{a}	16.54	1.09	5.10	9.77	0.91
HPO (HO)z-Gly-(C17.1:0) 346 346 68-76b 17.14 2.49 11.66 9.77 RODC (HO)z-Gly-(C17.9:1.4) 354 351 10-22b 16.92 2.45 11.56 9.89 RODC (HO)z-Gly-(C17.8:1.0) 354 352 25-32b 16.92 2.47 11.56 9.84 PODC (HO)z-Gly-(C17.8:1.0) 352 25-32b 16.92 2.47 11.56 9.84 PODC (HO)z-Gly-(C17.1:0) 600 630 65-72b 16.82 1.36 7.34 9.35 RODC (HO)z-Gly-(C17.1:0) 617 642 2-12b 16.59 1.33 6.49 9.48 RODC (HO)-Gly-(C17.9:1.4) 611 634 4-8b 16.57 1.33 7.27 9.50 RODC (HO)-Gly-(C17.8:1.0) 615 643 10-20b 16.57 1.33 7.41 9.41 RODC (HO)-Gly-(C17.8:1.0) 854 91 4-8b 16.54 1.33 7.41		Lignoceric acid	C24:0	369	416	84 ^a	16.55	1.01	4.90	9.85	0.92
RODC (HO) ₂ -Gly-C17.9:1.4) 354 351 10-22 ^b 16.92 2.45 11.56 9.89 SODC (HO) ₂ -Gly-C17.8:1.0) 351 348 8-18 ^b 16.56 2.47 11.63 10.17 SODC (HO) ₂ -Gly-C17.8:1.0) 354 352 340 45-55 ^a c 17.07 2.53 10.00 8.93 PODC (HO) ₂ -Gly-C17.8:1.0) 600 630 65-72 ^b 16.82 1.36 7.34 9.48 RODC (HO)-Gly-C17.1:0,2 617 643 10-20 ^b 16.59 1.35 7.27 9.56 RODC (HO)-Gly-C17.1:0,2 615 643 10-20 ^b 16.66 1.33 7.27 9.50 PODC (HO)-Gly-C17.1:0,3 681 717 8-42 16.66 1.33 7.41 9.41 HCO Gly-C17.1:0,3 854 915 62-66 16.70 0.93 4.79 9.50 HCO Gly-C17.1:0,3 877 87 93 1.44	səp		$(HO)_2$ -Gly- $(C17.1:0)$	346	346	68–76 ^b	17.14	2.49	11.66	9.77	0.91
SODC (HO)2-Gly-(C17.5:1.5) 351 348 8-18b 16.56 2.47 11.63 10.17 OODC (HO)2-Gly-(C17.8:1.0) 354 352 25-32b 16.58 2.44 11.56 9.84 PODC (HO)2-Gly-(C17.8:1.0) 600 630 65-72b 16.82 1.36 7.34 9.35 RODC (HO)-Gly-(C17.2:1.0) 611 642 2-12b 16.59 1.33 6.49 9.48 SODC (HO)-Gly-(C17.2:1.5) 615 643 10-20b 16.59 1.33 7.27 9.50 PODC (HO)-Gly-(C17.8:1.0) 552 619 42-48b 16.74 1.38 7.41 9.41 PODC (HO)-Gly-(C17.8:1.0) 854 915 62-66 16.74 1.38 7.41 9.41 HOO Gly-(C17.9:0.5) 854 915 62-66 16.70 0.93 4.79 9.82 RODC Gly-(C17.9:1.4)s 877 932 1.24-106 16.44 0.91 <th>ilə</th> <td></td> <td>$(HO)_2-Gly-(C17.9:1.4)$</td> <td>354</td> <td>351</td> <td>10-22^b</td> <td>16.92</td> <td>2.45</td> <td>11.56</td> <td>68.6</td> <td>0.92</td>	ilə		$(HO)_2-Gly-(C17.9:1.4)$	354	351	10-22 ^b	16.92	2.45	11.56	68.6	0.92
OODC (HO)2-GIy-(C17.8:1.0) 354 352 25-32b and color (HO)2-GIY-(C17.8:1.0) 11.56 9.84 and color (HO)2-GIY-(C17.8:1.0) 9.85 and color (HO)2-GIY-(C17.8:1.0) 9.87 and color (HO)2-GIY-(C17.8:1.0) 9.81 and color (HO)2-GIY-(C17.8:1.0) 9.82	λĮλ		$(HO)_2-Gly-(C17.5:1.5)$	351	348	8-18 ^b	16.56	2.47	11.63	10.17	0.95
PODC (HO)2-GIy-(C16.9.0.5) 342 340 45-55° c 17.07 2.53 10.00 8.93 HPO HO-GIy-(C17.1:0)2 600 630 65-72° 16.82 1.36 7.34 9.35 RDDC (HO)-GIy-(C17.1:0)2 611 642 2-12° 16.59 1.33 7.27 9.48 SODC (HO)-GIy-(C17.1:0)2 615 643 10-20° 16.66 1.33 7.27 9.56 OODC (HO)-GIy-(C17.8:1.0)2 592 619 4.248° 16.74 1.38 7.41 9.41 PODC (HO)-GIy-(C18.0.5)2 592 619 4.248° 16.74 1.38 7.41 9.41 HCO GIy-(C13.0.1)3 854 915 62-66° 16.74 0.93 4.79 9.82 RODC GIy-(C17.1.0)3 870 921 -3026° 16.44 0.92 4.78 10.04 SODC GIy-(C17.8.1.0)3 877 933 -1410° 16.51 0.91	Bou	_	$(HO)_2-GIy-(C17.8:1.0)$	354	352	25–32 ^b	16.98	2.44	11.56	9.84	0.92
HPO HO-Gly-(C17.1:0) ₂ 600 630 65-72 ^b 16.82 1.36 7.34 9.35 RODC (HO)-Gly-(C17.9:1.4) ₂ 617 642 2-12 ^b 16.59 1.33 6.49 9.48 SODC (HO)-Gly-(C17.9:1.4) ₂ 611 643 10-20 ^b 16.66 1.33 7.27 9.50 PODC (HO)-Gly-(C17.8:1.0) ₂ 592 619 42-48 ^b 16.66 1.33 7.27 9.50 PODC (HO)-Gly-(C17.8:1.0) ₂ 592 619 42-48 ^b 16.68 1.18 7.41 9.41 HCO Gly-(C13.0) ₃ 884 915 62-66 16.70 0.93 4.79 9.82 RODC Gly-(C17.9:1.4) ₃ 879 921 -3228 ^c 16.46 0.91 4.74 9.98 RODC Gly-(C17.8:1.0) ₃ 877 933 -1410 ^c 16.51 0.91 4.74 9.87 Q-to-(C17.8:1.0) ₃ 842 898 15-25 ^c 16.61 0.99 <th>юМ</th> <td></td> <td>$(HO)_2$-Gly-$(C16.9:0.5)$</td> <td>342</td> <td>340</td> <td>45–55^{a, c}</td> <td>17.07</td> <td>2.53</td> <td>10.00</td> <td>8.93</td> <td>0.83</td>	юМ		$(HO)_2$ -Gly- $(C16.9:0.5)$	342	340	45–55 ^{a, c}	17.07	2.53	10.00	8.93	0.83
RODC (HO)-GJy-(C17.9:1.4)2 617 642 2-12b 16.59 1.33 6.49 9.48 SODC (HO)-GJy-(C17.7:1.5)2 611 634 4-8b 16.57 1.35 7.27 9.56 OODC (HO)-GJy-(C17.8:1.0)2 615 643 10-20b 16.66 1.33 7.27 9.50 PODC (HO)-GJy-(C16.9:0.5)2 619 42-48b 16.66 1.33 7.41 9.41 HCO GJy-(C13:0)3 681 717 8-42 16.74 1.38 7.41 9.41 HPO GJy-(C17.1:0)3 854 915 62-66 16.70 0.93 4.79 9.82 RODC GJy-(C17.1:0)3 879 932 -3228c 16.44 0.92 4.78 10.03 SODC GJy-(C17.1:0)3 877 933 -1410c 16.51 0.95 4.84 9.87 PODC GJy-(C16.9:0.5)3 842 898 15-25c 16.61 0.99 4.84 9.87 <th>S</th> <td>HPO</td> <td>HO-Gly-(C17.1:0)₂</td> <td>009</td> <td>630</td> <td>65-72^b</td> <td>16.82</td> <td>1.36</td> <td>7.34</td> <td>9.35</td> <td>0.87</td>	S	HPO	HO-Gly-(C17.1:0) ₂	009	630	65-72 ^b	16.82	1.36	7.34	9.35	0.87
SODC (HO)-Gly-(C17.71.5)2 611 634 4-8b 16.57 1.35 7.32 9.56 OODC (HO)-Gly-(C17.81.0)2 615 643 10-20b 16.66 1.33 7.27 9.50 PODC (HO)-Gly-(C17.81.0)2 592 619 42-48b 16.66 1.38 7.41 9.41 HCO Gly-(C13:0)3 854 915 62-66c 16.70 0.93 4.79 9.82 HPO Gly-(C17.1.0)3 879 932 -30-26c 16.46 0.91 4.75 10.03 SODC Gly-(C17.1.1.5)3 870 921 -32-28c 16.44 0.92 4.78 10.04 OODC Gly-(C17.8.1.0)3 842 889 15-25c 16.61 0.95 4.74 9.98 PODC Gly-(C17.8.1.0)3 842 898 15-25c 16.61 0.95 4.74 9.87 A-tocopherol - 411 477 -7a 16.10 0.00 0.00 <t< td=""><th>əbin</th><td>RODC</td><td>$(HO)-Gly-(C17.9:1.4)_2$</td><td>617</td><td>642</td><td>$2-12^{b}$</td><td>16.59</td><td>1.33</td><td>6.49</td><td>9.48</td><td>0.89</td></t<>	əbin	RODC	$(HO)-Gly-(C17.9:1.4)_2$	617	642	$2-12^{b}$	16.59	1.33	6.49	9.48	0.89
OODC (HO)-Gly-(C17.8:1.0)2 615 643 10-20b 16.66 1.33 7.27 9.50 PODC (HO)-Gly-(C16.9:0.5)2 592 619 42-48b 16.66 1.38 7.41 9.41 HCO Gly-(C13:0)3 681 717 8-42 16.68 1.18 5.41 9.54 HCO Gly-(C17.1:0)3 854 915 62-66 16.70 0.93 4.79 9.82 RODC Gly-(C17.1:0)3 879 921 -3026° 16.46 0.91 4.75 10.03 SODC Gly-(C17.7:1.5)3 877 921 -3228° 16.44 0.92 4.74 9.98 PODC Gly-(C17.8:1.0)3 842 898 15-25° 16.61 0.95 4.84 9.87 squatene - 431 477 -7a 16.10 0.00 0.00 0.00 12.61	iəə/	SODC	$(HO)-Gly-(C17.7:1.5)_2$	611	634	-4-8 ^b	16.57	1.35	7.32	9.56	0.89
PODC (HO)-Gly-(C16.9:0.5) ₂ 592 619 42-48 ^b 16.74 1.38 7.41 9.41 HCO Gly-(C13:0) ₃ 681 717 8-42 16.68 1.18 5.41 9.54 HCO Gly-(C17.1:0) ₃ 854 915 62-66° 16.70 0.93 4.79 9.82 HPO Gly-(C17.1:0) ₃ 879 932 -3026° 16.46 0.91 4.75 10.03 SODC Gly-(C17.7:1.5) ₃ 870 921 -3228° 16.44 0.92 4.78 10.04 OODC Gly-(C17.8:1.0) ₃ 877 933 -1410° 16.53 0.91 4.74 9.98 PODC Gly-(C16.9:0.5) ₃ 842 898 15-25° 16.61 0.95 4.84 9.87 a-tocopherol - 431 477 -7a 16.10 0.00 0.00 0.00 12.61	(lgi	00DC	$(HO)-Gly-(C17.8:1.0)_2$	615	643	$10-20^{b}$	16.66	1.33	7.27	9.50	0.89
HCO Gly-(C13:0) ₃ 681 717 8-42 16.68 1.18 5.41 9.54 HPO Gly-(C17.1:0) ₃ 854 915 62-66° 16.70 0.93 4.79 9.82 RODC Gly-(C17.1:0) ₃ 879 932 -3026° 16.46 0.91 4.75 10.03 SODC Gly-(C17.7:1.5) ₃ 870 921 -3228° 16.44 0.92 4.78 10.04 OODC Gly-(C17.8:1.0) ₃ 877 933 -1410° 16.53 0.91 4.74 9.98 PODC Gly-(C16.9:0.5) ₃ 842 898 15-25° 16.61 0.95 4.84 9.87 α-tocopherol - 431 436 2a 17.63 1.47 7.27 8.74 Squalene - - - - - - - - - - 16.10 0.00 0.00 0.00 12.61 12.61 12.61 12.61 12.61	D	PODC	(HO)-Gly-(C16.9:0.5) ₂	592	619	42–48 ^b	16.74	1.38	7.41	9.41	0.88
HPO Gly-(C17.1:0)3 854 915 62-66° 16.70 0.93 4.79 9.82 RODC Gly-(C17.9:1.4)3 879 932 -3026° 16.46 0.91 4.75 10.03 SODC Gly-(C17.7:1.5)3 870 921 -3228° 16.44 0.92 4.78 10.04 OODC Gly-(C17.8:1.0)3 842 898 15-25° 16.61 0.95 4.84 9.87 PODC Gly-(C16.9:0.5)3 431 436 2a 17.63 0.95 4.84 9.87 a-tocopherol - 431 477 -7a 16.10 0.00 0.00 0.00 12.61		НСО	Gly-(C13:0) ₃	681	717	8-42	16.68	1.18	5.41	9.54	0.89
RODC Gly-(C17.9:1.4)3 879 932 -3026° 16.46 0.91 4.75 10.03 SODC Gly-(C17.7:1.5)3 870 921 -3228° 16.44 0.92 4.78 10.04 OODC Gly-(C17.8:1.0)3 877 933 -1410° 16.53 0.91 4.74 9.98 PODC Gly-(C16.9:0.5)3 842 898 15-25° 16.61 0.95 4.84 9.87 a-tocopherol - 431 436 2a 17.63 1.47 7.27 8.74 Squalene - 411 477 -7a 16.10 0.00 0.00 12.61	səp	HPO	$Gly-(C17.1:0)_3$	854	915	$62-66^{\circ}$	16.70	0.93	4.79	9.82	0.92
SODC Gly-(C17.7.1.5) ₃ 870 921 -3228° lo.44 16.44 0.92 4.78 10.04 OODC Gly-(C17.8.1.0) ₃ 877 933 -1410° lo.53 0.91 4.74 9.98 PODC Gly-(C16.9:0.5) ₃ 842 898 15-25° lo.61 0.95 4.84 9.87 a-tocopherol - 431 436 2ª 17.63 1.47 7.27 8.74 Squalene -7ª 16.10 0.00 0.00 12.61 12.61	nəc	RODC	Gly-(C17.9:1.4) ₃	879	932	-3026°	16.46	0.91	4.75	10.03	0.94
OODC Gly-(C17.8:1.0) ₃ 877 933 -1410 ^c 16.53 0.91 4.74 9.98 PODC Gly-(C16.9:0.5) ₃ 842 898 15-25 ^c 16.61 0.95 4.84 9.87 a-tocopherol - 431 436 2 ^a 17.63 1.47 7.27 8.74 Squalene - 411 477 -7 ^a 16.10 0.00 0.00 12.61	λĮδ	SODC	$Gly-(C17.7:1.5)_3$	870	921	-3228°	16.44	0.92	4.78	10.04	0.94
PODC Gly-(C16.9:0.5) ₃ 842 898 15-25c 16.61 0.95 4.84 9.87 a-tocopherol - 431 436 2a 17.63 1.47 7.27 8.74 Squalene 411 477 -7a 16.10 0.00 0.00 12.61	irī	00DC	$Gly-(C17.8:1.0)_3$	877	933	-1410°	16.53	0.91	4.74	86.6	0.93
α -tocopherol - 431 436 2^a 17.63 1.47 7.27 8.74 Squalene 411 477 -7a 16.10 0.00 0.00 12.61	,	PODC	$Gly-(C16.9:0.5)_3$	842	868	15–25°	16.61	0.95	4.84	9.87	0.92
Squalene 411 477 -7a 16.10 0.00 0.00 12.61	ers	α-tocopherol	I	431	436	2ª	17.63	1.47	7.27	8.74	0.82
	ЧιО	Squalene		411	477	_7a	16.10	0.00	0.00	12.61	1.18

 a from [65], b from [66], c from [67]

Table 5.	Used gr	up contribution	molar constants
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Group	F _d	F_p^2	E _h	Molar volume
Group	$[(J^{1/2} \cdot cm^{-3/2}) \cdot mol^{-1}]$	[(J·cm ⁻³)·mol ⁻¹]	[J⋅mol ⁻¹]	[cm ³ ·mol ⁻¹]
-CH ₃	420	0	0	33.5
-CH ₂ -	270	0	0	16.1
=CH-	200	0	0	13.5
>CH-	80	0	0	-1.0
>C<	-70	0	0	-1
=C<	70	0	0	-5.5
-COO-	390	240 100	7 000	18
-СООН	530	176400	10 000	28.5
-OH	210	250 000	20 000	10
Ring	190	0	0	16

where δ_d , δ_p , and δ_h are the components of the solubility parameter of the molecules and the PLA obtained from Equations (1), (2) and (3), where the radius value is the maximal distance obtained from [71] beyond which the molecules are not miscible anymore with the polymer. Therefore, the closer the RED value to zero, the better the compatibility. A RED value higher than 1 means a theoretical non-miscibility of the additive with PLA.

2.5. Physical-chemical characterization PLA blends

Differential Scanning Calorimetry (DSC) analyses were performed using a Mettler Toledo (France) DSC1 STARe System under nitrogen atmosphere (50 mL·min⁻¹) in 40 μL standard Aluminum pans (Mettler). Calibration of the device was done using Indium and Zinc standards. Calorimetric scans of additives were performed from -80 to 220°C at a heating rate of 2°C·min⁻¹. Experiments were carried out in duplicate. Calorimetric scans of blended PLA samples were done at a heating/cooling rate of 10°C⋅min⁻¹. The first heating scan for sample rejuvenation was performed from 25 to 75°C with an isotherm at 75°C during 2 min. Then, the cooling scan was from 75 to -20°C and the second heating scan from -20 to 100°C. The glass transition temperature (T_g) was taken at the midpoint of the second heating scan. Experiments were done in triplicate.

Tensile properties were investigated at 23°C, relative humidity (RH) 50±10 % and cross-head speed of 25 mm·min⁻¹, using an universal tensile machine (Instron model 4301, France) equipped with a load cell 1000±1 N and without extensiometer. The dog bone shaped samples (ISO 527-2, type 5A) were directly cut from the materials. Prior to tensile testing, samples were conditioned at 23°C and 50±10%

RH for at least 72 h. Each mechanical characteristic value is an average of 8 measurements.

Uniaxial deformation under an optical microscope was done in cutting rectangular samples of 20 mm length, 4 mm width, and 0.8 mm thickness from blank PLA, PLA + 10 wt% C16:0, PLA + 10 wt% C18:1 and PLA + 10 wt% PODC films. They were stretched at 5 mm·min⁻¹ using a homemade tensile machine placed under an Olympus Japan optical microscope mounted with WHK 10×/20 L ocular lens and MD Plan 10 0.25 objective lens. Observations were performed in optical transmission light mode using a Sony CCD-IRIS Model DXC-107P camera (France).

The morphology of the dispersed phase of PLA + 10 wt% C16:0, PLA + 10 wt% C18:1 and PLA + 10 wt% PODC materials before and after being stretched was observed with Scanning Electron Microscopy (SEM) (Hitachi 4800 II, France). Observations were directly conducted on the longitudinal surface of the dog bone shaped samples without any previous preparation.

2.6. Principal component analysis (PCA)

PCA was carried out considering the contents of each free fatty acid, mono, di and triglyceride types, hydrocarbons, sterols and water, the acid and saponification values and the measured elongation at break [%], Young modulus and $T_{\rm g}$ values of the PLA formulations tested. PCA was done with XLstat software.

3. Results and discussion

3.1. Effects of free fatty acids, glycerides,

α-tocopherol and squalene on PLA ductility Individual components present in vegetable oils, which could feature toughening ability for PLA, as free fatty acids, glycerides, and the unsaponifiable components as α-tocopherol and squalene were tested. In order to achieve the most efficient formulation, the solubility of a molecule in the polymer matrix is one determinant for its plasticizing power [37]. Therefore the Hansen Solubility Parameters (HSP) of each compound was calculated. HSP (Table 4) shows low solubility of the different compounds in the PLA matrix. All the obtained values are close to one, which marks the solubility limit. The highest solubility (although still modest) was displayed by small fatty acids and α-tocopherol. The mechanical and thermal properties of the corresponding compounds are given in Table 6. The raw data of typical stress/strain curves of blends are presented in Figure 1. The DSC thermograms of neat additives in Figure 2 and of blends in Figure 3 for further information.

Literature studies on the use of vegetable oil as toughening agent of PLA showed that generally chemical modification is required to obtain positive effects [50, 52-54, 56-58, 72]. Not surprisingly due to their low solubility in PLA, the impact of the tested fatty acids (C16:0, C18:1), vegetable oils (HPO and HCO) and unsaponifiable compounds on T_g was negligible. However, despite the low solubility of the compounds, PLA ductility was increased in some cases. In particular, HCO, which contains mainly triglycerides, but with short chain length, improved the PLA ductility and lowered the apparent Young modulus, stress and elongation at yield. HPO, which mostly contains triglycerides and some diglycerides with higher chain length did not bring significant ductility enhancement nor $T_{\rm g}$ decrease. HSP of di- and triglycerides from HPO and triglycerides from HCO

Table 6. Mechanical and thermal properties of formulations

	Formulation	Elongation at break [%]	app. Young modulus ^a [MPa]	Elongation at yield [%]	Stress at yield [MPa]	T _g [°C]
PLA (tv	win screw pellets)	6±1	1760±90	4.2±0.4	64±5	56.3±0.2
PLA+	10 wt% C16:0	4±1	1720±60	3.2±0.1	50±3	49.3±0.1
PLA+	10 wt% C18:1	16±5	1440±50	2.2±0.1	28±2	45.2±0.2
PLA +	10 wt% [50 wt% C16:0 + 50 wt% C18:1]	45±12	1590±80	2.2±0.1	31±2	40.7±0.3
	10 wt% [95 wt% (50 wt% C16:0 + 50 wt% :1) + 5 wt% HPO]	74±15	1550±120	2.2±0.2	25±4	39.8±0.4
	5 wt% HCO	9±3	1680±70	3.6±0.2	51±1	54.1±0.2
PLA+	10 wt% HCO	9±6	1540±80	3.6±0.2	42±2	54.1±0.1
	15 wt% HCO	11±5	1460±70	3.8±0.2	38±2	54.8 ± 0.3
	5 wt% HPO	55±20	1610±50	2.7±0.1	38±1	52.6±0.2
PLA+	10 wt% HPO	85±15	1420±70	2.6±0.2	26±2	54.4±0.4
	15 wt% HPO	91±14	1300±80	2.4±0.1	26±1	53.9 ± 0.3
	5 wt% α-tocopherol	6±2	1150±90	3.8±0.2	41±4	51.3±0.2
PLA+	10 wt% α-tocopherol	5±2	1070±60	4.0±0.4	31±3	49.8 ± 0.2
	15 wt% α-tocopherol	8±3	990±70	3.9±0.3	29±4	47.1±0.2
	5 wt% squalene	38±7	1180±90	3.3±0.4	36±7	51.4±0.2
PLA+	10 wt% squalene	51±9	990±60	3.1±0.9	28±3	49.5±0.2
	15 wt% squalene	29±7	890±80	2.9±0.7	19±2	49.4±0.1
	5 wt% RODC	22±3	1530±80	3.2±0.2	40±3	52.6±0.2
PLA+	10 wt% RODC	55±7	1490±140	2.4±0.2	28±4	49.1±0.1
PLA +	15 wt% RODC	65±15	1380±155	2.3±0.3	23±4	48.8 ± 0.1
	20 wt% RODC	73±20	1270±150	2.2±0.3	20±4	48.5±0.3
	5 wt% SODC	29±8	1550±110	3.3±0.3	41±7	50.6±0.1
DT A	10 wt% SODC	73±18	1510±90	2.3±0.2	25±5	49.4±0.3
PLA+	15 wt% SODC	80±13	1340±80	2.2±0.3	23±3	48.7 ± 0.1
	20 wt% SODC	52±15	1220±100	2.1±0.4	16±4	47.5±0.1
	5 wt% OODC	49±6	1430±90	3.1±0.3	34±4	49.5±0.2
DI A	10 wt% OODC	85±17	1490±90	2.6±0.2	31±2	49.0±0.1
PLA+	15 wt% OODC	88±16	1360±70	2.1±0.2	20±3	48.5±0.1
	20 wt% OODC	67±16	1370±110	2.2±0.2	21±3	48.7±0.1
	5 wt% PODC	51±17	1680±110	2.5±0.2	36±2	45.3±0.1
, .	10 wt% PODC	132±18	1460±40	2.1±0.2	24±1	39.2±0.2
PLA+	15 wt% PODC	179±15	1180±130	2.3±0.3	18±2	35.1±0.2
	20 wt% PODC	84±31	1130±60	2.0±0.4	14±3	33.8±0.2

^aapparent Young modulus for comparison reasons obtained without extensiometer

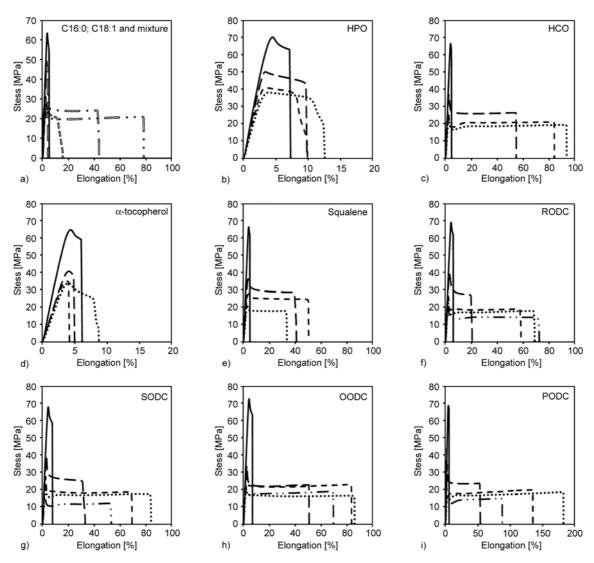


Figure 1. Stress/strain curves of PLA blends with free fatty acids, hydrogenated vegetable oils (HPO and HCO), unsaponifiable compounds (squalene, α-tocopherol), and vegetable oil deodorization condensates (palm oil PODC, olive oil OODC, soybean oil SODC and rapeseed oil RODC); (—) neat PLA; (== ==) PLA + 10 wt% C16:0; (= =) PLA + 10 wt% C18:1; (== •) PLA + 10 wt% [50 wt% C16:0 + 50 wt% C18:1]; (== •) PLA + 10 wt% [95 wt% (50 wt% C16:0 + 50 wt% C18:1) + 5 wt% HPO]; (——) PLA + 5 wt% additive; (- - -) PLA + 10 wt% additive; (• •) PLA + 15 wt% additive; (—• •) PLA + 20 wt% additive

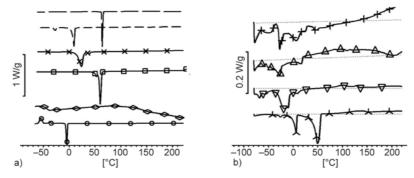


Figure 2. Thermograms (first heating scan) of free fatty acids, hydrogenated vegetable oils (HPO and HCO), unsaponifiable compounds (squalene, α-tocopherol), and vegetable oil deodorization condensates (palm oil PODC, olive oil OODC, soybean oil SODC and rapeseed oil RODC); (— —) C16:0; (− –) C18:1; (×) HCO; (□) HPO; (◊) α-tocopherol; (⋄) squalene; (+) RODC; (△) SODC; (▽) OODC; (▲) PODC

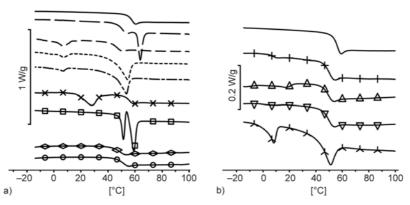


Figure 3. Thermograms (second heating scan) of PLA blends with free fatty acids, hydrogenated vegetable oils (HPO and HCO), unsaponifiable compounds (squalene, α-tocopherol), and vegetable oil deodorization condensates (palm oil PODC, olive oil OODC, soybean oil SODC and rapeseed oil RODC); (—) neat PLA; (— —) PLA + 10 wt% C16:0; (— –) PLA + 10 wt% C18:1; (— - –) PLA + 10 wt% [50 wt% C16:0 + 50 wt% C18:1]; (— -) PLA + 10 wt% [95 wt% (50 wt% C16:0 + 50 wt% C18:1) + 5 wt% HPO]; (×) PLA + 10 wt% HCO; (□) PLA + 10 wt% HPO; (◊) PLA + 10 wt% α-tocopherol; (⋄) PLA + 10 wt% squalene; (+) PLA + 10 wt% RODC; (Δ) PLA + 10 wt% SODC; (∇) PLA + 10 wt% OODC; (Λ) PLA + 10 wt% PODC

were close, while the average molar volume and molecular weight of triglycerides from HCO are smaller than the ones from HPO (Table 2). Testing of single fatty acids with longer chain length (C16:0, C18:1) showed that they were not able to increase PLA ductility. This is coherent with existing literature. For example, Jacobsen and Fritz [73] studied PLA blends with fatty acid esters. No significant ductility improvement was observed. The blending with the unsaponifiable compound α -tocopherol showed small gains in elongation at break, which extends already existing knowledge. It was already shown that the use of α -tocopherol as a natural antioxidant in PLA at small quantities (<4 wt%) induced a slightly $T_{\rm g}$ decrease [74] but no increase in elongation at break [75]. Here, no elongation at break improvement upon was found using greater α-tocopherol amounts, despite solubility was assessed to be higher than other molecules (such as HPO and HCO, Table 4) and a $T_{\rm g}$ decrease was observed (Table 3). Squalene, which is a natural tri-terpene found in vegetable oils, showed some toughening abilities (Table 6) despite its low solubility in PLA (Table 4). In fact, the creation of a dispersed phase promoting crazing was most probably responsible for this effect [37].

The most interesting result was obtained upon mixing different compounds. A mix of C18:1 and C16:0 fatty acids increased the PLA ductility much more than using them separately. The Young modulus and the $T_{\rm g}$ remained higher than using only C18:1 likely because of its lower content involved, while the stress and elongation at yield were lessened as

much as adding only C18:1. Mixture made of [95 wt% (50 wt% C16:0 + 50 wt% C18:1) + 5 wt% HPO] at a total content of 10 wt% in PLA, further enhanced the ductility. There was apparently a synergistic effect between compounds, which can be possibly exploited.

3.2. Properties of PLA blends with vegetable oil deodorization condensates

Deodorization oil condensates are mixtures of different molecules contained in vegetable oils, the composition of which depends on the botanic source and oil refinery process. Four kinds of oil deodorization condensates (RODC, SODC, OODC and PODC) were blended with PLA, each at four concentrations (5, 10, 15 and 20 wt%). As shown in Table 3, content in free fatty acids, mono di or triglycerides, sterols (mainly alpha-tocopherol) and hydrocarbons (mainly squalene) was highly variable, as well as unsaturation degree of fatty acids. From the composition, an average HSP was calculated and is given in Table 4. As observed in Table 4, the longer and/or more unsaturated the alkyl chain is, the lower the solubility in PLA. However, except for very short fats alkyl chains and organic compounds as α-tocopherol or squalene, HSP of involved molecules appear to be rather similar. Mechanical and thermal properties of PLA/ oil deodorization condensates blends are given in Table 6 and typical curves of raw data are shown in the Figure 1. Significant increases in elongation at break were obtained, especially with the PODC. Addition of too much additive, i.e. about 20 wt%, induced a stagnation or decrease in the elongation at break. All the deodorization condensates led to a

lowering in the stress and elongation at yield. Interestingly, the apparent Young modulus of materials remained high. The stress/strain curves (Figure 1) showed that the yielding peak shrunk and flattened with the increase in additive content, as if the yield critical stress would tend to meet the plateau value where the stress remains constant with the strain. There is thus an important gain of using native mixtures of fatty acids present in deodorisation condensates for PLA ductility increase.

3.3. Study of the deformation mechanisms involved in PLA/oil deodorization condensates blends

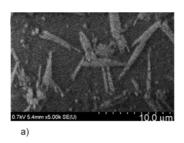
In all samples the $T_{\rm g}$ remained higher than the measurement temperature, the main deformation mechanism was thus crazing of the glassy polymer [37]. The yield process in glassy amorphous polymers can be described as a stress induced glass-transition. In fact, because PLA Poisson's ratio is less than $0.5 (v_{PLA} = 0.36)$ [76], the volume of PLA increases when subjected to tensile stress. Correspondingly, samples experienced important stress whitening, which appeared simultaneously with yielding. Stress whitening is caused by the formation of sizeable microvoids in the polymer matrix due to cavitation and crazes [77]. Crazes can also be initiated inside non-miscible inclusions in the polymer matrix or on the interface between inclusion and polymer in the case of low compatibility. The addition of oil deodorization condensates initiated crazes at stress levels substantially below those of the brittle failure of neat PLA, which propagated perpendicularly to the stretching direction. Morphological analysis of the form and size of inclusions of PODC, C16:0 and C18:1 was done to help interpretation. Figure 4 shows SEM micrographs of [PLA/C16:0], [PLA/ C18:1] and [PLA/PODC] 90 wt%/ 10 wt% blends. [PLA/C16:0] blends exhibit rods of about 5 to 10 μm length and 1 to 2 μm width, corresponding to fatty acid crystals. [PLA/C18:1] showed small spherical domains of fatty acids of about $0.5~\mu m$ diameters and some aggregates of about 3 to $5~\mu m$ diameters. The [PLA/PODC] blend micrograph showed both rods and aggregates of droplets, where the droplets appear to be distributed all around the crystals.

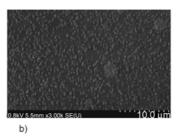
Figure 5 shows optical micrographs in transmission mode of stretched samples under the microscope taken at different percentages of elongation. Many small cracks started appear in neat PLA when approaching the yield peak, which is common for glassy amorphous polymers. Addition of C16:0, *i.e.* rod like crystals, increased both the occurrence and the length of cracks. Cracks quantity upon stretching caused the material to become opaque.

Failure of the blend revealed PLA fibrils. C18:1, *i.e.* small spherical liquid domains, induced fewer but much larger cracks as if their extensibility was eased. PODC, *i.e.* mostly a combination of C16:0 and C18:1 and some minor constituents, showed a high number of cracks but with smaller width as if using only C18:1. The mixture of palmitic acid crystals and liquid inclusions of oleic acid had thus a synergistic effect allowing for efficient craze initiation by the crystals and by cavitation inside the liquid inclusions. The superior performance of the PODC blends seems thus to be linked to the chemical composition in fatty acids and the presence of the minor constituents.

3.4. Role of the chemical composition of deodorization condensates in PLA ductility improvement

A Principal Components Analysis (PCA) was carried out including composition data and physicochemical characteristics of the deodorization condensates and of the PLA samples. Figure 6 shows the principal components projection plot of F1 and F2 of the data set. Only 50% of the total variance in these PLA samples is extracted according to F1 and F2 axes. In fact, scores of formulations containing





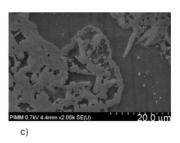


Figure 4. SEM micrographs of (a) C16:0, (b) C18:1 and (c) PODC blends with PLA at 10 wt%

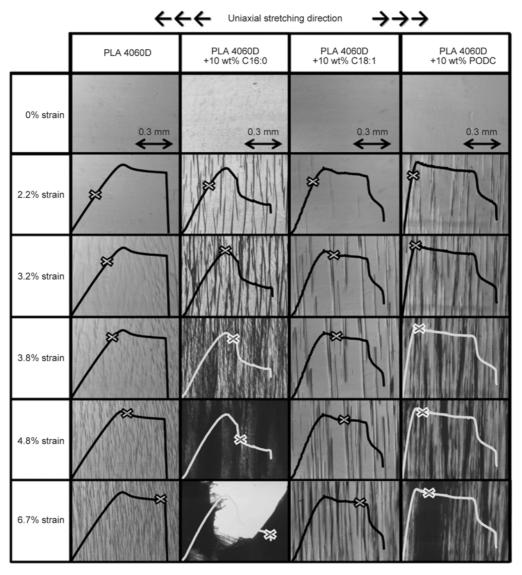


Figure 5. Optical micrographs of uniaxially stretched PLA and PLA blends with free fatty acids (C16:0, C18:1) and palm oil deodorization condensate (PODC) at different strain

squalene, α-tocopherol, HPO, HCO and mixtures of C16:0, C18:1 and HPO are not discriminated. However discrimination of PLA samples containing deodorization condensates of each kind of vegetable oils and according to their respective amount is obtained. Corresponding loadings are plotted in Figure 7. The positive part of F1 axis appears to be mainly governed by the elongation at break and as opposed to its negative part with the $T_{\rm g}$. The F2 axis mostly separates fats as a function of the unsaturation and length of their alkyl chains. Looking at the elongation at break improvement, unsaturation of alkyl chains appears to be required, but the lower the ratio, the larger the enhancement was. Medium alkyl chain length, i.e. from lauric (C12) to stearic (C18) acid, also tends to be preferable than the long

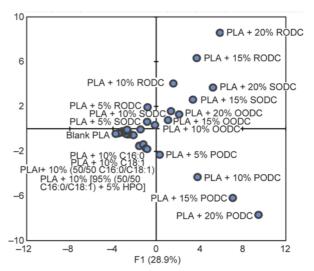


Figure 6. Principal components projection plot of F1 and F2 of the data set

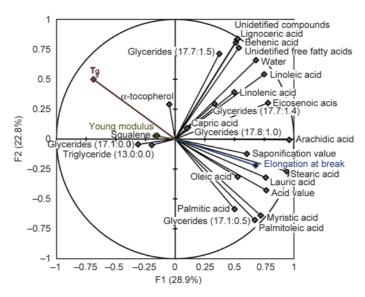


Figure 7. Loadings plot of the Principal Components Analysis of the dataset

ones as behenic (C22) or lignoceric (C24) acids. Both the unsaturation and the length of alkyl chains are main physical factors governing the melting point of fats. One common point is that unsaturation decreases melting points while length increases it. Therefore, efficiency of fats in toughening PLA appeared to be linked to medium melting point properties. A mixture of crystalline and liquid fatty acids at room temperature seemed favorable. This could explain the ability of HCO triglycerides to increase the PLA ductility while HPO did not, although their HSP were similar, as their ability to depress the $T_{\rm g}$ of PLA. In fact, due to short alkyl chains, HCO triglycerides are waxy at room temperature. The DSC thermogram (Figure 2) shows a broad melting peak going from 8 to 42°C of HCO while it is beyond 55°C for HPO. Literature shows that combining liquid and solid additives can sometimes be effective for PLA [78]. Acid and saponification values are related to the elongation at break improvement (Figure 7). Therefore, the higher the esterified fats and/or the free fatty acid contents, the better the efficiency of the additive. In opposite, unidentified compounds, i.e. mostly unsaponifiable compounds, are not favorable to the ductility increase. Therefore, the superiority in toughening abilities of the PODC compared to the alike mixture made of [95 wt% (50 wt% C16:0 + 50 wt% C18:1) + 5 wt% HPO] can be attributed to the complex and favorable mixture of fatty acids having different chain length.

4. Conclusions

By-products of the vegetable oils industry, namely deodorization condensates, were investigated as biobased additives for ductility improvement of PLA. The deodorization condensates improved substantially the elongation at break of PLA up to 180%. The glass transition was merely decreased which brought ductility to the still glassy polymer. The deformation mechanism was efficient craze initiation delaying failure. Most importantly, it was shown that the industrial by-product of the palm oil refinery had superior properties compared to mixtures of fatty acids and vegetable oils because it contained a mixture of fatty acids with melting points below and beyond room temperature. This makes deodorization condensates efficient and low price additives for PLA able to be used in commodity applications such as food packaging.

Acknowledgements

This work was supported by Brodart Packaging and the BIP-ADEME French project CREABioM. The authors greatly thank the laboratory Institut des Corps Gras (ITERG – Bordeaux, France) for its technical support.

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