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# Industrial vegetable oil by-products increase the ductility of polylactide

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**Abstract.** The use of industrial by-products of the vegetable oil industry as ductility increasing additives of polylactide (PLA) was investigated. Vegetable oil deodorization condensates were melt-blended by twin-screw extrusion up to a maximum inclusion quantity of 20 wt% without preliminary purification. Sample films were obtained by single screw cast extrusion. Compounded PLA films featured largely improved ductility in tensile testing with an elongation at break up to 180%. The glass transition temperature remained higher than room temperature. The native mixture of molecules, which composed the deodorization condensates, had superior performance compared to a synthetic mixture of main compounds. The investigation of the correlation between composition of the additives and the ductility of the PLA blends by Principal Component Analysis showed synergy in property improvement between fatty acids having a melting point below and beyond the room temperature. Furthermore, a compatibilizing effect of molecules present in the native mixture was evidenced. Oil deodorization condensates, which are a price competitive by-product of the vegetable oil industry, are therefore a very promising biobased and biodegradable additive for improving the ductility of PLA.

Keywords: biodegradable polymers, poly(lactic acid), ductility, vegetable oil by-products, crazing

### 1. Introduction

Biodegradable and/or biobased polymers are widely studied to replace petroleum-based materials with the aim of contributing to higher sustainability of plastics. However, in many cases these new products still do not meet economical or functional requirements to be competitive in commodity applications. Polylactide (PLA) is one of the polymers which can potentially enter high volume markets, mostly in the food packaging [1–3] or textile [4] sector. Polylactide is a biobased and compostable polymer [5–7] issued from polymerization of lactic acid produced by fermentation of starch, potatoes or

beets [7, 8]. Although it has been known before [9], production cost reduction in the 1990s and 2000s thanks to breakthroughs in the polymerization technology [10, 11], made it economically competitive with petroleum-based materials. PLA offers many interests such as a glass transition higher than room temperature, ease of processing, high transparency, printability, glossy aspect [1, 12, 13]. However, its mechanical properties still remain an obstacle for many applications. In fact, PLA features high tensile strength and modulus but also high brittleness [14]. In order to enhance the PLA ductility, melt-blending with rubbery materials as poly(ether)urethane [15,

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16], polyamide elastomer [17], acrylonitrile-butadiene-styrene copolymer, or various impact modifiers [18–26] is effective. Furthermore, plasticizers such as citrate esters [27–37], polyethylene glycols [27, 33, 38–44] or manifold other molecules [29, 43, 45–48] can be used. Unfortunately, impact modifiers are often not biodegradable nor biobased and plasticizers are generally derived from fossil resources, degrading one important environmental advantage of employing PLA. Therefore biobased and biodegradable additives have been and are investigated for toughening PLA. Chemically modified vegetable oils have received important research interest, because ester or epoxy groups can be degraded by microorganisms [49], maintaining the biodegradability of blends with PLA. Polymerized soybean oil derivatives prepared by crosslinking double bonds of alkyl chains afforded ductility increase after meltblending with PLA and using a compatibilizer [50]. Improvements were also obtained with conjugated soybean oil which was reactively compatibilized with PLA by unsaturated triglycerides [51]. Epoxidized soybean oil [52-55] and epoxidized palm oil [56–58] also showed some positive effects on PLA toughness. Another approach consists in using rigid particles, which usually increased stiffness but is ineffective, or even detrimental for the ductility. Nonetheless, NatureWorks [59] reported the use of EMforce<sup>®</sup>, a mineral additive, able to change the polymer failure mode from brittle to ductile. In fact, the efficient crack initiation in glassy PLA, which was provided by the well-dispersed additive with good interfacial adhesion to the matrix, enhanced the PLA elongation at break.

Expertise from plasticizing polyvinylchloride teaches that mixtures of different plasticizers are efficient for increasing toughness, because they make use of different mechanisms available for ductility improvement [49]. For example, a mixture of tributyl citrate (TBC) and a block copolymer PLA-g-polyethyleneglycol yielded a material with high elongation at break and satisfying stress at yield [30]. Al-Mulla et al. [55] successfully used modified nanoclays in combination with epoxidized soybean oil to increase stiffness and ductility. The combination of PLA with rubber and compatibilizers has also been investigated by several authors, showing important gains in ductility, with an elongation at break of up to 160% for high rubber concentrations [60] or bicontinous phases [61], and up to 200% when using

compatibilizers [62, 63]. However, these solutions often require the inclusion of several additives, chemically modified and costly components that increase the price of the final formulation, which is inappropriate for a large volume production. Finding a low cost biobased and biodegradable toughening agent would therefore help PLA to conquer new markets.

Vegetable oils contain a number of potentially interesting molecules for PLA toughening and the refinery of vegetable oils gives rise to several by-products. Among those, oil deodorization condensates represent a high-volume by-product, which is chemically close to the vegetable oil. They are obtained by distillation in the aim of purifying the raw oil from odorous compounds, which are negative for the sensorial properties of vegetable oils. Deodorization condensates contain free fatty acids, glycerides and unsaponifiable molecules. The objective of this study was to use vegetable oils and the byproduct deodorization condensate as produced for toughening PLA. The individual molecules which make up oils and deodorization condensates were tested on the mechanical properties and several vegetable oils were screened.

### 2. Experimental

### 2.1. Materials

PLA 4060D was supplied by NatureWorks (U.S.A.) and consists of  $89\pm1\%$  L-lactic acid and  $11\pm1\%$  D-lactic acid units, making it unable to crystallize under common conditions [1, 64]. The glass transition temperature ( $T_{\rm g}$ ) obtained from differential scanning calorimetry (DSC) measurements was 56°C. Average molecular weights obtained from size exclusion chromatography (SEC) measurements were  $M_{\rm w} = 236\,800~{\rm g\cdot mol^{-1}}$ ,  $M_{\rm n} = 103\,700~{\rm g\cdot mol^{-1}}$  and dispersity  $M_{\rm w}/M_{\rm n} = 2,28$ .

Hydrogenated palm oil (HPO), Hydrogenated Copra Oil (HCO), Rapeseed Oil Deodorization Condensate (RODC), Soybean Oil Deodorization Condensate (SODC), Olive Oil Deodorization Condensate (OODC) and Palm Oil Deodorization Condensate (PODC) were supplied by ITERG (Bordeaux, France).

Palmitic acid (purity >98%) *i.e.* hexadecanoic acid (C16:0), oleic acid (purity >96%) *i.e.* 9-octadecenoic acid (C18:1), squalene (purity >98%) *i.e.* (6E,10E,14E,18E)-2,6,10,15,19,23-hexamethyltetracosa-2,6,10,14,18,22-hexaene, and alpha-toco-

pherol (purity >96%) *i.e.* (2R)-2,5,7,8-tetramethyl-2-[(4R,8R)-(4,8,12-trimethyltridecyl)]-6-chromanol, were supplied by Sigma-Aldrich (France).

### 2.2. Film fabrication

Prior to melt-blending, PLA pellets were dried at 60°C for 24 h under dried air using a SOMOS 60L. Relative humidity of dried pellets was controlled to be lower than 350 ppm using an Aboni FMX Hydrotracer (France). Melt-mixing of PLA with or without additives was carried out using a twin screw extruder (Thermo Haake Ptw 16-40D, France), having a screw diameter of 16 mm and a length to diameter ratio (*L/D*) 40:1. A temperature of 180°C was used to process by twin-screw extrusion the PLA pellets as reference material, while the temperature profile of the 7 heating zones decreased from 180 to about 150/130°C along the extrusion flow, depending on the nature and the amount of additive. Screw

speed was 300 rpm. The exact temperature profile for each formulation is given in Table 1. To properly control the feed rate of additives, which are solid at room temperature, a home-made apparatus consisting in a heated syringe was used. Obtained pellets were stored into hermetic sealed metalized bags to avoid PLA rehydration.

Films of about 0.8 mm thickness containing palmitic acid, oleic acid, hydrogenated palm oil, hydrogenated copra oil, squalene,  $\alpha$ -tocopherol and some combinations of these products were obtained using a single screw extruder (Scamex Rheoscam, France), mounted with a screw of 20 mm diameter and a length to diameter ratio (L/D) 12:1 and a flat die of 40 mm width and 1 mm thickness. Films were stretched and cooled with chill rolls. Due to the short length of the screw, an increasing temperature profile from 180 to 195°C over the 3 heating zones was used for PLA, while it was decreasing from 180

Table 1. Processing conditions of melt-mixing twin screw extrusion

	Formulation	Temperature profile
Neat PL	A	175/180/190/190/190/190/180
PLA + 1	10 wt% C16:0	175/180/180/170/170/170/170
PLA + 1	10 wt% C18:1	175/180/180/170/170/170/170
PLA + 1	10 wt% [50 wt% C16:0 + 50 wt% C18:1]	175/180/180/170/170/170/170
	10 wt% [95 wt% (50 wt% C16:0 + 50 wt% C18:1) + 5 wt% HPO]	175/180/180/170/170/170/170
	5 wt% HCO	175/180/180/180/180/170/170
PLA+	10 wt% HCO	175/180/180/180/180/170/170
	15 wt% HCO	175/180/180/180/170/170/160
	5 wt% HPO	175/180/180/180/180/170/170
PLA+	10 wt% HCO	175/180/180/180/180/170/170
	15 wt% HCO	175/180/180/180/180/170/170
	5 wt% α-tocopherol	175/180/180/180/180/180/180
PLA+	10 wt% α-tocopherol	175/180/180/180/180/180/170
	15 wt% α-tocopherol	175/180/180/180/180/170/170
	5 wt% squalene	175/180/180/180/180/180/180
PLA+	10 wt% squalene	175/180/180/180/180/170/170
	15 wt% squalene	175/180/180/180/180/170/170
	5 wt% RODC	175/180/180/180/180/175/170
PLA+	10 wt% RODC	175/180/180/180/180/170/160
PLA +	15 wt% RODC	175/180/180/180/180/170/160
	20 wt% RODC	175/180/180/180/170/160/150
	5 wt% SODC	175/180/180/180/180/175/170
PLA+	10 wt% SODC	175/180/180/180/180/170/160
rla +	15 wt% SODC	175/180/180/180/180/170/160
	20 wt% SODC	175/180/180/180/170/160/150
	5 wt% OODC	175/180/180/170/170/170/170
PLA+	10 wt% OODC	175/180/180/170/170/170/170
ILAF	15 wt% OODC	175/180/180/170/170/160/160
	20 wt% OODC	175/180/180/170/160/150/150
	5 wt% PODC	175/180/180/180/180/175/170
PLA+	10 wt% PODC	175/180/180/180/170/160/155
ILA	15 wt% PODC	175/180/180/180/170/160/150
	20 wt% PODC	175/180/180/170/160/150/135

Table 2. Processing conditions of cast films extrusion

	Formulation	Temperature profile [°C]	Die temperature [°C]
Neat PI	A	180/190/195	185
PLA +	10 wt% C16:0	180/180/175	170
PLA +	10 wt% C18:1	180/180/175	170
PLA +	10 wt% [50 wt% C16:0 + 50 wt% C18:1]	180/180/175	170
PLA +	10 wt% [95 wt% (50 wt% C16:0 + 50 wt% C18:1) + 5 wt% HPO]	180/180/175	170
	5 wt% HCO	180/180/180	180
PLA+	10 wt% HCO	180/170/170	170
	15 wt% HCO	180/170/170	170
	5 wt% HPO	180/180/180	170
PLA+	10 wt% HPO	180/170/170	170
	15 wt% HPO	180/170/170	170
	5 wt% α-tocopherol	180/180/180	180
PLA+	10 wt% α-tocopherol	180/180/180	180
	15 wt% α-tocopherol	180/175/170	170
	5 wt% squalene	180/180/180	180
PLA+	10 wt% squalene	180/180/175	170
	15 wt% squalene	180/180/170	170

to about 170°C for blends. The specific temperatures depending on the nature and the amount of additive are given in Table 2.

In the case of the deodorization condensates of the vegetable oils (RODC, SODC, OODC, PODC), the available quantity was insufficient for carrying out a single screw extrusion in the aim to obtain films. In that case PLA sheets of 1 mm thickness were thermo-moulded by compression at 220 bar (Laboratory Press Gibitre Instruments 20 tons, Italy). For this, pellets were pre-melted at 180°C without pressure during 180 seconds then heating plates were closed with progressive increase in pressure during 120 seconds to eliminate air bubbles.

## 2.3. Chemical and physical chemical characterization of vegetable oils and deodorization condensates

Glyceride composition of fats was determined according to the IUPAC 6.002 and EN 14105 standards using a Shimadzu GC-2010 Plus gas chromatograph (France) equipped with a Zebron ZB 5 HT Inferno (15 m, 0.25 mm, 0.1 μm) column and a flame ionization detector set at 380°C. The vector gas was H<sub>2</sub> at a flow rate of 1.17 mL·min<sup>-1</sup>. Both the injector and the oven temperature were set at 60°C for 3 min, then raised to 370°C at 10°C·min<sup>-1</sup> and held at 370°C for 12 min. Direct on-column injection was performed.

Fatty acid composition was determined according to the ISO 12966-2 standard using a Shimadzu GC-

2010 Plus gas chromatograph (France) equipped with a BPX70 (50 m, 0.22 mm, 0.25  $\mu$ m) column, and a flame ionization detector set at 250°C. The vector gas was H<sub>2</sub> at a flow rate of 0.32 mL·min<sup>-1</sup>. The oven temperature was set at 60 C for 2 min, raised to 170°C at 20°C·min<sup>-1</sup>, held at 170°C for 25 min, raised to 230°C at 4°C·min<sup>-1</sup> and held at 230°C for 10 min. The injector temperature was set at 250°C and a split ratio of 200 was used.

Acid value was determined according to the ISO 660 standard using a mixture of ethanol 95% and diethylic ether as solvent, potassium hydroxide 0.5 mol·L<sup>-1</sup> in ethanol 95% as titrant and alkali blue 6B as indicator.

Saponification value was determined according to the ISO 3657 standard. Samples were saponified with potassium hydroxide 0.5 mol·L<sup>-1</sup> in ethanol 95% boiled under reflux during 2 h. Hydrochloric acid 0.5 mol·L<sup>-1</sup> was used as titrant and alkali blue 6B as indicator.

Water content was measured using a Mettler Toledo HB43 S Halogen Moisture Analyzer set (France) at 103°C.

Melting point of free fatty acids was determined from literature [65]. Estimation of mono, di- and triglycerides average chemical structure of oil deodorization condensates was based on the averages unsaturation number and the alkyl chains length of the corresponding free fatty acids composition profile (Table 3). Average glycerides melting point was estimated calculating a weighted average of the

Table 3. Comp	position of hydro	genated vegetable	oils and vegetable	oil deodorization condensates

Type			HPO	НСО	RODC	SODC	OODC	PODC
	Free fatty acids		0.0	0.0	39.0	43.1	39.2	95.4
	Monoglycerides		0.2	0.0	11.2	2.9	2.0	1.7
ide itio	Diglycerides		10.6	0.0	2.7	9.0	8.4	2.2
ceri posi [%]	Triglycerides		88.9	100	12.2	16.8	33.5	0.7
Glyceride composition [%]	Sterols (α-tocopher	rol)	0.0	0.0	25.7	9.8	1.6	0.0
5 8	Hydrocarbons (Squ	ialene)	0.0	0.0	0.0	13.8	13.0	0.0
	Unidentified		0.3	0.0	9.2	4.6	2.3	0.0
Acid value [m	g KOH·g <sup>-1</sup> ]		0.1	0.1	65.3	68.4	47.7	201.0
Saponification	n value [mg KOH·g-	1]	198.6	255.2	125.1	157.9	162.2	205.7
Water content	[%]		0.11	0.10	0.55	0.34	0.37	0.16
	Caproic acid	C6:0	0.0	0.5	0.0	0.0	0.0	0.0
	Caprylic acid	C8:0	0.0	6.8	0.0	0.0	0.0	0.0
	Capric acid	C10:0	0.0	5.7	0.0	0.0	0.1	0.0
	Lauric acid	C12:0	0.5	47.5	0.0	0.8	0.0	0.4
l g	Myristic acid	C14:0	1.2	18.1	0.0	0.4	0.0	1.3
 sitic	Palmitic acid	C16:0	43.6	9.4	7.4	12.3	11.3	49.8
od	Palmitoleic acid	C16:1	0.0	0.0	0.0	0.0	0.0	0.2
Fatty acid composition [%]	Stearic acid	C18:0	53.8	10.8	3.4	4.1	2.5	4.1
	Oleic acid	C18:1	0.0	1.0	27.3	21.7	69.5	35.2
	Linoleic acid	C18:2	0.0	0.0	42.4	49.7	10.9	7.8
	Linolenic acid	C18:3	0.0	0.0	1.5	6.4	0.6	0.3
	Arachidic acid	C20:0	0.5	0.1	0.5	0.3	0.4	0.3
	Eicosenoic acid	C20:1	0.0	0.0	0.3	0.2	0.4	0.1
	Behenic acid	C22:0	0.1	0.0	1.0	0.5	0.0	0.0
	Lignoceric acid	C24:0	0.1	0.0	0.5	0.2	0.1	0.0
	Unidentified		0.5	0.1	0.2	15.7	13.4	4.2
Average alkyl	chain carbon quanti	ty	17.1	13.0	17.9	17.7	17.8	16.9
Average alkyl	chain unsaturation	quantity	0.0	0.0	1.4	1.5	1.0	0.5

specific melting points of the contained molecules taken from references [65–67]. Physical properties of molecules are given in Table 4.

### 2.4. Calculation of solubility parameters

Molar volumes and molar attraction constants of polylactide and additives were determined according to the van Krevelen and Hoftyzer atomic group contribution method [68]. Average molar volumes and average molar attraction constants of mono, di and triglycerides of oil deodorization condensates were estimated based on average chemical structures previously determined.

Hansen Solubility parameters were calculated using Equations (1), (2) and (3) [69, 70]. Used molar constant values [68] are presented in Table 5:

$$\delta_{\rm d} = \frac{\sum F_{\rm dt}}{\sum V_{\rm i}} \tag{1}$$

$$\delta_{\rm p} = \frac{\sqrt{\sum F_{\rm Pi}^2}}{\sum V_{\rm i}} \tag{2}$$

$$\delta_{\rm h} = \frac{\sqrt{\sum E_{\rm hi}}}{\sum V_{\rm i}} \tag{3}$$

where  $\delta_{\rm d}$  is the dispersion component of the solubility parameter in J<sup>1/2</sup>·cm<sup>-3/2</sup>,  $\delta_{\rm p}$  the polar component of the solubility parameter in J<sup>1/2</sup>·cm<sup>-3/2</sup>,  $\delta_{\rm h}$  the hydrogen bonding component of the solubility parameter in J<sup>1/2</sup>·cm<sup>-3/2</sup>,  $F_{\rm d}$  the dispersion contribution of the molar attraction constant in (J<sup>1/2</sup>·cm<sup>-3/2</sup>)·mol<sup>-1</sup>,  $F_{\rm p}$  the polar contribution of the molar attraction constant in (J<sup>1/2</sup>·cm<sup>-3/2</sup>)·mol<sup>-1</sup>,  $E_{\rm h}$  the hydrogen bonding energy contribution of the molar attraction constant in J·mol<sup>-1</sup> and V the molar volume contribution of the chemical group involved in cm<sup>3</sup>·mol<sup>-1</sup>.

The solubility of the molecules in PLA (Table 4) was assessed using the HSP Relative Energy Difference (RED) from Equations (4) and (5):

Distance = 
$$\sqrt{4(\delta_{\text{dmolec}} - \delta_{\text{dPLA}})^2 + (\delta_{\text{pmolec}} - \delta_{\text{pPLA}})^2 + (\delta_{\text{hmolec}} - \delta_{\text{hPLA}})^2}$$
 (4)

$$RED = \frac{Distance}{Radius} \tag{5}$$

Table 4. Physical properties and solubility parameters of fatty acids, mono-, di-, and triglycerides contained in the hydrogenated vegetable oils and oil deodorization condensates

				Molor woight	Molar voluma	Melting		Hansen Sol	Hansen Solubility Parameters (HSP)	(HSP)	
Particle   Particle				[g·mol <sup>-1</sup> ]	[cm <sup>3</sup> ·mol <sup>-1</sup> ]	point [°C]	$\delta_{ m d}$ $[{f J}^{1/2}{f \cdot}{f cm}^{-3/2}]$	$\delta_{ m p}$ $[{ m J}^{1/2}.{ m cm}^{-3/2}]$	$\delta_{\rm h}$ $[{\bf J}^{1/2}.{\bf cm}^{-3/2}]$	Distance with PLA	RED with PLA
Cappole said         C60         116         126         -34         160         3.22         8.89         880           Cappole said         C80         114         126         -34         16.00         2.66         7.94         8.90           Cappole acid         C100         20         114         159         134         16.20         2.26         7.94         8.90           Cappole acid         C120         228         2.25         547         16.46         1.46         5.90         9.46           Palmitic acid         C160         2.85         2.87         16.45         1.46         5.90         9.46           Palmitic acid         C180         2.85         2.87         16.45         1.46         5.90         9.46           Palmitic acid         C182         2.85         3.20         770         16.49         1.34         5.94         9.80           Palmitic acid         C182         2.87         3.40         1.94         16.52         1.34         5.94         9.80           Delace acid         C182         2.82         3.40         1.94         1.64         1.14         5.94         9.80           Delace acid <th< th=""><th>PL/</th><th>1 4060D</th><th></th><th>242 800</th><th>ı</th><th>1</th><th>18.6</th><th>6.6</th><th>0.9</th><th>(radius: 10.7)</th><th>1</th></th<>	PL/	1 4060D		242 800	ı	1	18.6	6.6	0.9	(radius: 10.7)	1
Capyris and CR0         (144)         159         116         16.0         2.65         7.94         8.91           Capyris and CR0         (120)         278         116         16.30         2.26         7.94         8.91           Lauric and Cl2.0         (120)         228         2.25         44°         16.37         1.18         6.70         9.05           Palminole and Cl4.1         256         2.87         2.87         13         16.49         1.34         6.25         9.34           Palminole and Cl6.1         2.85         2.87         6.87         16.49         1.31         5.95         9.85           Palminole and Cl8.1         2.88         3.94         -1,4°         16.49         1.31         5.95         9.82           Dole and Cl8.2         2.89         3.94         -1,4°         16.49         1.34         5.95         9.82           Limberic and Cl8.2         2.89         3.94         -1,4°         16.54         1.19         5.64         9.77           Limberic and Cl8.2         2.89         3.94         -1,4°         16.54         1.34         5.95         9.88           Limberic and Cl8.2         2.89         3.84         3.84 <td< td=""><td></td><td>Caproic acid</td><td>C6:0</td><td>116</td><td>126</td><td>_3a</td><td>16.06</td><td>3.32</td><td>8.89</td><td>8.80</td><td>0.82</td></td<>		Caproic acid	C6:0	116	126	_3a	16.06	3.32	8.89	8.80	0.82
Campin and C1000         C1000         T72         191         31°         16.30         2.00         7.24         9.05           Chair and C1000         C1000         172         255         555         555         16.6         6.0         9.30           Pulmine and C160         C160         226         225         555         146         5.0         9.46           Pulmine and C161         C180         226         226         1.0         1.0         9.0         9.46           Pulmine and C181         228         329         1.0         16.4         1.46         5.90         9.46           Pulminoles and C182         228         320         1.0         16.4         1.36         5.60         9.58           Discussion and C182         228         340         -9         16.4         1.36         5.60         9.58           Lingloucera and C220         1.0         341         342         1.2         1.5         1.34         5.60         9.77           Robert and C220         341         342         342         1.6         1.0         9.77         9.66           Behem and C220         341         345         346         1.6         1.6 </td <td></td> <td>Caprylic acid</td> <td>C8:0</td> <td>144</td> <td>159</td> <td>16a</td> <td>16.20</td> <td>2.65</td> <td>7.94</td> <td>8.91</td> <td>0.83</td>		Caprylic acid	C8:0	144	159	16a	16.20	2.65	7.94	8.91	0.83
Mystic acid         C12-0         200         223         44*         16.77         188         6.70         9.20           Mystic acid         C16-0         226         287         63*         164         1.46         5.90         9.46           Palmitic acid         C16-1         226         287         63*         164         1.46         5.90         9.46           Plannic acid         C16-1         226         287         282         70*         1649         1.34         5.59         9.46           Steams acid         C18.1         283         3.94         -14*         16.22         1.34         5.54         9.71           Linoletic acid         C18.1         283         3.44         -14*         15.55         1.19         5.54         9.71           Linoletic acid         C18.2         3.74         1.44         15.54         1.19         5.54         9.71           Linoletic acid         C18.2         3.44         4.44         15.54         1.19         5.54         9.71           Linoletic acid         C18.2         3.44         4.44         15.54         1.19         5.54         9.71           Linoletic acid         C18.2 </td <td></td> <td>Capric acid</td> <td>C10:0</td> <td>172</td> <td>191</td> <td>31a</td> <td>16.30</td> <td>2.20</td> <td>7.24</td> <td>9.05</td> <td>0.85</td>		Capric acid	C10:0	172	191	31a	16.30	2.20	7.24	9.05	0.85
Mystics and C140         228         255         55°         1642         1.65         6.0         94           Plannitic and C160         256         227         63°         1642         1.16         5.0         946           Plannitic and C180         258         227         63°         1649         1.31         5.99         9.46           Plannicles and C181         283         340         178         16.49         1.31         5.59         9.58           Linolecie and C182         283         344         174         16.49         1.34         5.59         9.80           Linolecie and C182         283         344         14*         16.42         1.34         5.59         9.80           Linolecie and C182         283         344         14*         16.42         1.14         5.99         9.80           Linolecie and C182         283         344         14*         16.35         1.19         5.74         10.00           Anachidic acid         C182         346         346         84**         16.55         1.10         4.90         9.80           Eccosenor and C200         4100-C04-C17.1.40         354         346         84**         16.55		Lauric acid	C12:0	200	223	44ª	16.37	1.88	6.70	9.20	98.0
Palminoles acid         C166         256         287         63³         1646         1.46         5.59         9.46           Palminoles acid         C181         254         282         1³         1627         1.49         5.59         9.46           Steat acid         C18.1         285         320         70°         1627         1.19         5.59         9.62           Oleta cand         C18.1         283         320         70°         1622         1.34         5.59         9.73           Oleta cand         C18.1         280         390         -9         16.27         1.19         5.59         9.73           Innoles acid         C18.3         278         394         -14*         15.55         1.38         5.74         10.09           Amenhic acid         C200         31         34         34         27*         16.52         1.19         5.37         9.68           Amenhic acid         C200         341         344         346         346         346         346         346         347         4.99         9.83           Behanic acid         C200         340         446         87*         16.54         1.10         5			C14:0	228	255	55a	16.42	1.65	6.26	9.34	0.87
Palminotic acid         Cl6.1         254         222         1*         16.27         1.49         5.95         9.62           Palminotic acid         Cl8.0         285         320         7°         16.24         1.31         5.95         9.62           Olete acid         Cl8.1         283         314         13*         16.24         1.34         5.59         9.62           Olete acid         Cl8.2         288         314         13*         16.34         1.34         5.69         9.85           Linoleca acid         CR0.0         313         32.2         73*         16.35         1.13         5.69         9.85           Arachdic acid         CR0.0         31         347         23*         16.36         1.13         5.74         10.03           Arachdic acid         C20.0         31         347         34*         32*         16.36         1.13         5.74         10.03           Behavic acid         C20.0         31         34         34*         34*         35*         1.04         3.74         9.88           Behavic acid         C20.0         41         34         34*         35*         44*         16.25         11.10 <td>sbi</td> <td></td> <td>C16:0</td> <td>256</td> <td>287</td> <td>63a</td> <td>16.46</td> <td>1.46</td> <td>5.90</td> <td>9.46</td> <td>0.88</td>	sbi		C16:0	256	287	63a	16.46	1.46	5.90	9.46	0.88
Steams and CIRRO         CIRRO         285         314         170*         1649         1.31         5.59         9.58           Linoleic acid CIRRI         CIRRI         288         314         170*         1632         1.34         5.69         9.71           Linoleic acid CIRRI         CIRRI         288         399         -9*         16.14         1.36         5.69         9.81           Linoleic acid CRO         CRO         311         352         75*         16.35         1.19         5.69         9.81           Arachidic acid CRO         CRO         311         347         27*         16.36         1.10         5.37         9.80           Bebenic acid CRO         CRO         341         344         84*         16.54         1.10         4.90         9.87           HOO         HOO-GLACIT.SLI)         346         346         88*         16.54         1.10         4.90         9.87           BODC         HOO-GLACIT.SLI)         344         84*         16.56         2.47         11.66         9.87           SODC         HOO-GLACIT.SLI)         354         352         25-29*         10.98         2.44         11.56         9.80	ge.		C16:1	254	282	1a	16.27	1.49	5.95	9.62	06.0
Oble acid         CIRS1         233         314         13°s         16.32         1.34         5.64         9.71           Oble acid         CIRS1         238         309         -9°s         16.14         1.36         5.64         9.71           Linolenic acid         CIRS2         288         309         -14°s         15.92         1.38         5.74         10.03           Linolenic acid         CRO         313         35.2         73°s         16.52         1.19         5.53         9.68           Ecosenoic acid         C20.0         341         347         23°s         16.54         1.19         5.74         10.03           Behenic acid         C22.0         341         344         80°s         16.54         1.19         5.37         9.68           Behenic acid         C22.0         341         344         80°s         16.54         1.10         4.90         9.83           Behenic acid         C24.0         11.1         2.45         1.10         4.90         9.83           Behenic acid         C24.0         11.1         3.44         3.44         3.44         3.44         3.44         3.44         3.44         3.44         3.44	YHE		C18:0	285	320	70a	16.49	1.31	5.59	9.58	0.89
Linoletic acid         C18.2         280         309         -9*         16.14         136         5.69         9.86           Linoletic acid         C18.3         278         309         -9*         16.14         136         5.69         9.86           Arachidic acid         C20.1         311         347         324         16.55         1.19         5.37         9.86           Belevisence acid         C22.0         341         384         87         16.54         1.01         4.90         9.77           Belevisence acid         C24.0         346         416         84*         16.54         1.01         4.90         9.87           Belevisence acid         C24.0         346         416         84*         16.54         1.01         4.90         9.77           Belevisence acid         C24.0         1.01         344         346         84*         16.54         1.01         4.90         9.77           RDO         (HO)-GIV-CIV-CIV-SI-1.5         351         346         45-58*         16.98         2.44         11.56         9.84           NDO         (HO)-GIV-CIV-CIV-SI-1.5         351         340         45-58*         16.98         2.44         11.5	3 ə		C18:1	283	314	13a	16.32	1.34	5.64	9.71	0.91
Linolemic acid         C18:3         374         144         15.55         1.38         5.74         1003           Amachidic acid         C20-0         313         32.2         1.34         15.5         1.19         5.33         968           Eignoceric acid         C20-0         311         347         234         16.54         1.09         5.31         968           Eignoceric acid         C20-0         341         384         804         16.54         1.09         5.30         968           Lignoceric acid         C22-0         346         346         68-76         17.14         2.49         11.66         9.77           HPO         (HO)-EGIA-CIT.3-1.9         351         348         8-18*         16.56         2.44         11.66         9.77           SODC         (HO)-EGIA-CIT.8-1.0         354         354         348         8-18*         16.56         2.44         11.66         9.77           SODC         (HO)-EGIA-CIT.8-1.0         354         354         34.0         42-58*         16.56         2.44         11.56         9.84           SODC         (HO)-EGIA-CIT.8-1.0         354         34.0         42-48*         16.57         1.35         <	Fre		C18:2	280	309	9a	16.14	1.36	5.69	98.6	0.92
Amerbidic acid         C200         313         352         75°         16.52         1.19         5.33         968           Eleosenoir acid         C20.1         341         347         23°         16.54         1.19         5.37         9.80           Behnic acid         C22.0         341         344         84°         16.54         1.01         4.90         9.77           Behnic acid         C22.0         34         346         84°         16.54         1.01         4.90         9.77           HPO         (HO)-Gly-(C17.1.0)         346         346         88-76°         17.14         2.49         11.66         9.77           RODC         (HO)-Gly-(C17.1.0)         354         351         1.02°         16.98         2.44         11.56         9.84           RODC         (HO)-Gly-(C17.8.1.0)         354         352         25-32°         16.98         2.44         11.56         9.84           RODC         (HO)-Gly-(C17.8.1.0)         342         340         45-55°         16.98         2.44         11.56         9.84           RODC         (HO)-Gly-(C17.8.1.0)         354         42.9         16.53         1.33         7.24         11.56			C18:3	278	304	-14 <sup>a</sup>	15.95	1.38	5.74	10.03	0.94
Eirocenoic acid         C20:1         311         347         23*         16.36         1.21         5.37         9.80           Bignoenic acid         C20:0         369         364         384         80*         16.54         1.09         5.10         9.77           Bignoenic acid         C20:0         369         416         84*         16.54         1.09         4.90         9.87           BoDC         (HO)r-Gly-C(17.51.3)         354         354         346         88-76*         17.14         2.49         11.66         9.77           RODC         (HO)r-Gly-C(17.51.3)         354         354         32.2         2.45*         11.66         9.89           SODC         (HO)r-Gly-C(17.81.0)         354         32.2         2.25*         10.00         8.93           PODC         (HO)r-Gly-C(17.81.0)         342         45-55*c         17.07         2.53         10.00         8.93           RODC         (HO)-Gly-C(17.1.0)         342         44-8*         16.56         1.36         7.34         9.36           RODC         (HO)-Gly-C(17.1.0)         61         642         2.12*         16.59         1.36         7.34         9.36           RODC		Arachidic acid	C20:0	313	352	75a	16.52	1.19	5.33	89.6	06.0
Behenic acid         C22.0         341         384         80°         16.54         1.09         5.10         9.77           Ligmoceric acid         C24.0         369         416         84°         16.55         1.01         4.90         9.77           HDO         (HOD-GIA-(CT7.1.0)         346         346         68-76         17.14         2.49         11.66         9.77           SDDC         (HOD-GIA-(CT7.8.1.0)         354         351         348         8-18b         16.56         2.47         11.66         9.77           SDDC         (HOD-GIA-(CT7.8.1.0)         354         352         25-32b         16.98         2.47         11.66         9.89           PODC         (HOD-GIA-(CT7.8.1.0)         354         352         25-32b         16.98         2.47         11.66         9.84           PODC         (HOD-GIA-CT7.1.0)         342         45-55°         16.98         1.35         1.30         9.84           RDD         (HOD-GIA-CT7.7.1.0)         617         642         2-7b         16.89         1.35         1.34         9.48           SDDC         (HOD-GIA-CT7.7.1.0)         617         642         2-7b         1.653         1.35         1.41 <td></td> <td>Eicosenoic acid</td> <td>C20:1</td> <td>311</td> <td>347</td> <td>23a</td> <td>16.36</td> <td>1.21</td> <td>5.37</td> <td>08.6</td> <td>0.92</td>		Eicosenoic acid	C20:1	311	347	23a	16.36	1.21	5.37	08.6	0.92
Lignoceric acid         C24:0         369         416         84*         1655         1.01         4.90         9.85           HPO         (HO)L-Gly-(C17.1:0)         346         346         68-76*         17.14         2.49         11.66         9.77           RODC         (HO)L-Gly-(C17.9:14)         354         354         351         10-22*         16.92         2.45         11.56         9.89           RODC         (HO)L-Gly-(C17.1:0)         354         352         25-32*         16.98         2.47         11.56         9.89           PODC         (HO)L-Gly-(C17.1:0)         342         342         45.58*         17.07         2.33         10.00         8.93           PODC         (HO)L-Gly-(C17.1:0)         342         340         45-53*         10.00         8.93           RODC         (HO)L-Gly-(C17.1:0)         61         642         2-12*         16.89         1.33         7.34         9.38           RODC         (HO)-Gly-(C17.3:1.0)         61         642         1.62*         1.38         7.41         9.48           RODC         (HO)-Gly-(C17.3:1.0)         61         643         10-20*         16.66         1.33         7.41         9.41		Behenic acid	C22:0	341	384	$80^{a}$	16.54	1.09	5.10	9.77	0.91
HOD         (HO)z-GIy-(CI7.1:0)         346         346         68-76b         17.14         2.49         11.66         9.77           RODC         (HO)z-GIy-(CI7.9:14)         354         351         10-22b         16.92         2.45         11.56         9.89           RODC         (HO)z-GIy-(CI7.8:1.0)         354         352         25-32b         16.82         2.47         11.66         9.84           OODC         (HO)z-GIy-(CI7.8:1.0)         354         340         45-55a (17.0)         1.00         8.93           PODC         (HO)z-GIy-(CI7.1:0)z         600         630         65-72b         16.82         1.36         7.34         9.38           RODC         (HO)-GIy-(CI7.1:0)z         617         642         2-12b         16.59         1.33         6.49         9.48           RODC         (HO)-GIy-(CI7.3:1.4)z         611         634         4-8b         16.57         1.33         7.27         9.50           RODC         (HO)-GIy-(CI7.3:1.4)z         611         634         42-48b         16.54         1.33         7.41         9.41           RODC         (HO)-GIy-(CI7.3:0.1)z         851         619         42-48b         16.74         1.33         7.41		Lignoceric acid	C24:0	369	416	84ª	16.55	1.01	4.90	9.85	0.92
RODC         (HO)z-Gly-(C17.9:1.4)         354         351         10-22b         16.92         2.45         11.56         9.89           SODC         (HO)z-Gly-(C17.8:1.0)         354         351         348         8-18b         16.56         2.47         11.63         10.17           SODC         (HO)z-Gly-(C17.8:1.0)         354         352         352         340         45-55a*         17.07         2.53         10.00         8.93           PODC         (HO)z-Gly-(C17.1.0)z         610         630         65-72b         16.82         1.36         7.34         9.35           RDC         (HO)-Gly-(C17.1.0)z         617         643         10-20b         16.59         1.33         7.27         9.50           SODC         (HO)-Gly-(C17.8:1.0)z         615         643         10-20b         16.66         1.33         7.27         9.50           SODC         (HO)-Gly-(C17.8:1.0)z         615         643         10-20b         16.66         1.33         7.41         9.41           HCO         Gly-(C17.8:1.0)z         884         915         62-66         16.74         1.38         7.41         9.41           HCO         Gly-(C17.1:0)z         884         870	səp		$(HO)_2$ -Gly-(C17.1:0)	346	346	68–76 <sup>b</sup>	17.14	2.49	11.66	9.77	0.91
SODC         (HO)2-GIy-(C17.5:1.5)         351         348         8-18th only         16.56         2.47         11.63         10.17           PODC         (HO)2-GIy-(C17.8:1.0)         354         352         25-32th only         16.98         2.44         11.66         9.84           PODC         (HO)2-GIy-(C17.1:0)2         600         630         65-72th only         16.59         1.33         6.49         9.48           RODC         (HO)-GIY-(C17.1:1.0)2         611         642         1-2th only         16.59         1.33         6.49         9.48           SODC         (HO)-GIY-(C17.1:1.0)2         611         642         1-2th only         16.59         1.33         6.49         9.48           SODC         (HO)-GIY-(C17.1:1.0)2         615         643         10-20th only         16.56         1.33         7.27         9.50           PODC         (HO)-GIY-(C17.1:1.0)2         681         717         8-42         16.74         1.38         7.41         9.41           HCO         GIY-(C13.0)3         854         915         62-66         16.70         0.93         4.79         9.82           RODC         GIY-(C17.9.1.4)3         877         932         1.24-10	inə		$(HO)_2-Gly-(C17.9:1.4)$	354	351	10-22 <sup>b</sup>	16.92	2.45	11.56	68.6	0.92
OODC         (HO)2-GJy-(C17.8:1.0)         354         352         25-32b to the component of the co	βJλc		$(HO)_2-Gly-(C17.5:1.5)$	351	348	8-18 <sup>b</sup>	16.56	2.47	11.63	10.17	0.95
PODC         (HO)GIy(CIO.9:0.5)         342         340         45-55°° (a)         17.07         2.53         10.00         8.93           HPO         HO-GIy(CI7.1:0)2         600         630         65-72°         16.82         1.36         7.34         9.35           RODC         (HO)GIy(CI7.1:0)2         611         642         2-12°         16.59         1.33         6.49         9.48           SODC         (HO)GIy(CI7.2:0)2         615         643         10-20°         16.66         1.33         7.27         9.56           OODC         (HO)GIy(CI7.3:0)3         681         717         8-42         16.68         1.18         7.41         9.41           PODC         GIy(CI3.0)3         681         717         8-42         16.68         1.18         5.41         9.41           HCO         GIy(CI3.0)3         884         915         62-66         16.70         0.93         4.79         9.82           RODC         GIy(CI7.1:0)3         879         921         -23-28°         16.44         0.92         4.78         10.04           RODC         GIy(CI7.3:1.0)3         877         933         -14-10°         16.51         0.91 <t< td=""><td>gon</td><td></td><td><math>(HO)_2-Gly-(C17.8:1.0)</math></td><td>354</td><td>352</td><td>25-32<sup>b</sup></td><td>16.98</td><td>2.44</td><td>11.56</td><td>9.84</td><td>0.92</td></t<>	gon		$(HO)_2-Gly-(C17.8:1.0)$	354	352	25-32 <sup>b</sup>	16.98	2.44	11.56	9.84	0.92
HPO         HO-Gly-C17.1:0 <sub>2</sub> 600         630         65-72 <sup>b</sup> 16.82         1.36         7.34         9.35           RODC         (HO)-Gly-(C17.1:0 <sub>2</sub> )         617         642         2-12 <sup>b</sup> 16.59         1.33         6.49         9.48           SODC         (HO)-Gly-(C17.7:1.5) <sub>2</sub> 611         634         4-8 <sup>b</sup> 16.57         1.35         7.27         9.50           OODC         (HO)-Gly-(C17.8:1.0) <sub>2</sub> 615         643         10-20 <sup>b</sup> 16.66         1.33         7.27         9.50           PODC         (HO)-Gly-(C17.8:1.0) <sub>2</sub> 592         619         42-48 <sup>b</sup> 16.68         1.38         7.41         9.41           HCO         Gly-(C13.0) <sub>3</sub> 681         717         8-42         16.68         1.18         5.41         9.54           HPO         Gly-(C17.1:0) <sub>3</sub> 854         915         62-66         16.70         0.93         4.79         9.82           RODC         Gly-(C17.9:1.4) <sub>3</sub> 870         921         -32-28 <sup>c</sup> 16.44         0.92         4.78         10.04           SODC         Gly-(C17.8:1.0) <sub>3</sub> 842         884         898         15-25 <sup>c</sup> 16.6	юΜ		$(HO)_2$ -Gly-(C16.9:0.5)	342	340	45–55 <sup>a, c</sup>	17.07	2.53	10.00	8.93	0.83
RODC         (HO)-Gly-(C17.9:1.4)2         617         642         2-12b to the composition of the composi	S		HO-Gly-(C17.1:0) <sub>2</sub>	009	630	65-72 <sup>b</sup>	16.82	1.36	7.34	9.35	0.87
SODC         (HO)-Gly-(C17.71.5)2         611         634         4-8b         16.57         1.35         7.32         9.56           OODC         (HO)-Gly-(C17.81.0)2         615         643         10-20b         16.66         1.33         7.27         9.50           PODC         (HO)-Gly-(C17.81.0)2         592         619         42-48b         16.68         1.18         7.41         9.41           HCO         Gly-(C13.0)3         881         717         8-42         16.68         1.18         5.41         9.41           HCO         Gly-(C17.10)3         854         915         62-66         16.70         0.93         4.79         9.82           HPO         Gly-(C17.11.0)3         879         921         -30-26         16.46         0.91         4.75         10.03           SODC         Gly-(C17.71.5)3         870         921         -32-28c         16.44         0.92         4.78         9.98           PODC         Gly-(C17.81.0)3         842         898         15-25c         16.61         0.95         4.84         9.87           q-tocopherol         -         431         477         -7a         16.10         0.00         0.00         0.00 <td>əbir</td> <td></td> <td><math>(HO)-Gly-(C17.9:1.4)_2</math></td> <td>617</td> <td>642</td> <td>2-12<sup>b</sup></td> <td>16.59</td> <td>1.33</td> <td>6.49</td> <td>9.48</td> <td>0.89</td>	əbir		$(HO)-Gly-(C17.9:1.4)_2$	617	642	2-12 <sup>b</sup>	16.59	1.33	6.49	9.48	0.89
OODC         (HO)-Gly-(C17.8:1.0)2         615         643         10-20b look         16.66         1.33         7.27         9.50           PODC         (HO)-Gly-(C16.9:0.5)2         592         619         42-48b look         16.68         1.18         7.41         9.41           HCO         Gly-(C13:0)3         681         717         8-42         16.68         1.18         5.41         9.54           HFO         Gly-(C17.1:0)3         854         915         62-66         16.70         0.93         4.79         9.82           RODC         Gly-(C17.9:1.4)3         879         921         -3228°         16.44         0.91         4.75         10.03           SODC         Gly-(C17.8:1.0)3         877         933         -1410°         16.51         0.95         4.84         9.87           PODC         Gly-(C16.9:0.5)3         842         898         15-25°         16.61         0.95         4.84         9.87           a-tocopherol         -         431         477         -7a         16.10         0.00         0.00         0.00         12.61	iəə/		$(HO)-Gly-(C17.7:1.5)_2$	611	634	-4-8b	16.57	1.35	7.32	9.56	0.89
PODC         (HO)-Gly-(C16.9:0.5) <sub>2</sub> 592         619         42-48 <sup>b</sup> 16.74         1.38         7.41         9.41         9.41           HCO         Gly-(C13:0) <sub>3</sub> 681         717         8-42         16.68         1.18         5.41         9.54           HCO         Gly-(C13:0) <sub>3</sub> 854         915         62-66°         16.70         0.93         4.79         9.82           RODC         Gly-(C17.1.0) <sub>3</sub> 879         932         -3026°         16.46         0.91         4.75         10.03           SODC         Gly-(C17.1.1.5) <sub>3</sub> 877         921         -3228°         16.44         0.92         4.78         10.04           OODC         Gly-(C17.8:1.0) <sub>3</sub> 877         933         -1410°         16.53         0.91         4.74         9.98           PODC         Gly-(C16.9:0.5) <sub>3</sub> 842         898         15-25°         16.61         0.95         4.84         9.87           A-tocopherol         -         431         477         -7a         16.10         0.00         0.00         0.00         0.00         0.99         4.84         9.87	(lgi		$(HO)-Gly-(C17.8:1.0)_2$	615	643	10-20 <sup>b</sup>	16.66	1.33	7.27	9.50	0.89
HCO         Gly-(C13:0) <sub>3</sub> 681         717         8-42         16.68         1.18         5.41         9.54           HPO         Gly-(C17.1:0) <sub>3</sub> 854         915         62-66°         16.70         0.93         4.79         9.82           RODC         Gly-(C17.1:0) <sub>3</sub> 879         932         -3026°         16.46         0.91         4.75         10.03           SODC         Gly-(C17.7:1.5) <sub>3</sub> 870         921         -3228°         16.44         0.92         4.78         10.04           OODC         Gly-(C17.8:1.0) <sub>3</sub> 877         933         -1410°         16.51         0.91         4.74         9.98           PODC         Gly-(C16.9:0.5) <sub>3</sub> 842         898         15-25°         16.61         0.95         4.84         9.87           Squalenc         -         431         436         2a         17.63         0.00         0.00         0.00         12.61	D		(HO)-Gly-(C16.9:0.5) <sub>2</sub>	592	619	42-48 <sup>b</sup>	16.74	1.38	7.41	9.41	0.88
HPO         Gly-(C17.1:0)3         854         915         62-66°         16.70         0.93         4.79         9.82           RODC         Gly-(C17.9:1.4)3         879         932         -3026°         16.46         0.91         4.75         10.03           SODC         Gly-(C17.7:1.5)3         870         921         -3228°         16.44         0.92         4.78         10.04           OODC         Gly-(C17.8:1.0)3         877         933         -1410°         16.53         0.91         4.74         9.98           PODC         Gly-(C16.9:0.5)3         842         898         15-25°         16.61         0.95         4.84         9.87           Squalence         -         431         477         -7a         16.10         0.00         0.00         0.00         12.61			Gly-(C13:0) <sub>3</sub>	681	717	8-42	16.68	1.18	5.41	9.54	0.89
RODC         Gly-(C17.9:1.4)3         879         932         -3026°         16.46         0.91         4.75         10.03           SODC         Gly-(C17.7:1.5)3         870         921         -3228°         16.44         0.92         4.78         10.04           OODC         Gly-(C17.8:1.0)3         877         933         -1410°         16.53         0.91         4.74         9.98           PODC         Gly-(C16.9:0.5)3         842         898         15-25°         16.61         0.95         4.84         9.87           a-tocopherol         -         431         436         2a         17.63         1.47         7.27         8.74           Squalene         -         411         477         -7a         16.10         0.00         0.00         0.00         12.61	səp		Gly-(C17.1:0) <sub>3</sub>	854	915	62–66°	16.70	0.93	4.79	9.82	0.92
SODC         Gly-(C17.7.1.5)₃         870         921         -3228¢ ODC         16.44         0.92         4.78         10.04           OODC         Gly-(C17.8.1.0)₃         877         933         -1410¢ 16.53         0.91         4.74         9.98           PODC         Gly-(C16.9:0.5)₃         842         898         15-25¢ 16.61         0.95         4.84         9.87           a-tocopherol         -         431         436         2³         17.63         1.47         7.27         8.74           Squalene         -         411         477         -7³         16.10         0.00         0.00         12.61	inəc		$Gly-(C17.9:1.4)_3$	628	932	-3026°	16.46	0.91	4.75	10.03	0.94
OODC         Gly-(C17.8:1.0) <sub>3</sub> 877         933         -1410 <sup>c</sup> 16.53         0.91         4.74         9.98           PODC         Gly-(C16.9:0.5) <sub>3</sub> 842         898         15-25 <sup>c</sup> 16.61         0.95         4.84         9.87           a-tocopherol         -         431         436         2 <sup>a</sup> 17.63         1.47         7.27         8.74           Squalene         -         411         477         -7 <sup>a</sup> 16.10         0.00         0.00         12.61	λĮδ		$Gly-(C17.7:1.5)_3$	820	921	-3228°	16.44	0.92	4.78	10.04	0.94
PODC         Gly-(C16.9:0.5)₃         842         898         15–25°         16.61         0.95         4.84         9.87           α-tocopherol         -         431         436         2³         17.63         1.47         7.27         8.74           Squalene         411         477         -7³         16.10         0.00         0.00         12.61	irī		$Gly-(C17.8:1.0)_3$	877	933	$-1410^{c}$	16.53	0.91	4.74	86.6	0.93
$\alpha$ -tocopherol         -         431         436 $2^a$ 17.63         1.47         7.27         8.74           Squalene         411         477         -7a         16.10         0.00         0.00         12.61	_		$Gly-(C16.9:0.5)_3$	842	868	15–25°	16.61	0.95	4.84	9.87	0.92
Squalene A11 477 -7a 16.10 0.00 0.00 12.61	GLS	a-tocopherol	I	431	436	2ª	17.63	1.47	7.27	8.74	0.82
	Ч1Ο	Squalene		411	477	_7a	16.10	0.00	0.00	12.61	1.18

<sup>a</sup>from [65], <sup>b</sup>from [66], <sup>c</sup>from [67]

Table 5.	Used grou	p contribution	molar constants
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Group	F <sub>d</sub>	$F_p^2$	Eh	Molar volume
- · · · I	[(J <sup>1/2</sup> ·cm <sup>-3/2</sup> )·mol <sup>-1</sup> ]	[(J·cm <sup>-3</sup> )·mol <sup>-1</sup> ]	[J·mol <sup>-1</sup> ]	[cm <sup>3</sup> ·mol <sup>-1</sup> ]
-CH <sub>3</sub>	420	0	0	33.5
-CH <sub>2</sub> -	270	0	0	16.1
=CH-	200	0	0	13.5
>CH-	80	0	0	-1.0
>C<	-70	0	0	-1
=C<	70	0	0	-5.5
-COO-	390	240 100	7 000	18
-СООН	530	176 400	10 000	28.5
-OH	210	250 000	20 000	10
Ring	190	0	0	16

where  $\delta_d$ ,  $\delta_p$ , and  $\delta_h$  are the components of the solubility parameter of the molecules and the PLA obtained from Equations (1), (2) and (3), where the radius value is the maximal distance obtained from [71] beyond which the molecules are not miscible anymore with the polymer. Therefore, the closer the RED value to zero, the better the compatibility. A RED value higher than 1 means a theoretical non-miscibility of the additive with PLA.

### 2.5. Physical-chemical characterization PLA blends

Differential Scanning Calorimetry (DSC) analyses were performed using a Mettler Toledo (France) DSC1 STARe System under nitrogen atmosphere (50 mL·min<sup>-1</sup>) in 40 μL standard Aluminum pans (Mettler). Calibration of the device was done using Indium and Zinc standards. Calorimetric scans of additives were performed from -80 to 220°C at a heating rate of 2°C·min<sup>-1</sup>. Experiments were carried out in duplicate. Calorimetric scans of blended PLA samples were done at a heating/cooling rate of 10°C⋅min<sup>-1</sup>. The first heating scan for sample rejuvenation was performed from 25 to 75°C with an isotherm at 75°C during 2 min. Then, the cooling scan was from 75 to -20°C and the second heating scan from -20 to 100°C. The glass transition temperature  $(T_g)$  was taken at the midpoint of the second heating scan. Experiments were done in triplicate.

Tensile properties were investigated at 23°C, relative humidity (RH) 50±10 % and cross-head speed of 25 mm·min<sup>-1</sup>, using an universal tensile machine (Instron model 4301, France) equipped with a load cell 1000±1 N and without extensiometer. The dog bone shaped samples (ISO 527-2, type 5A) were directly cut from the materials. Prior to tensile testing, samples were conditioned at 23°C and 50±10%

RH for at least 72 h. Each mechanical characteristic value is an average of 8 measurements.

Uniaxial deformation under an optical microscope was done in cutting rectangular samples of 20 mm length, 4 mm width, and 0.8 mm thickness from blank PLA, PLA + 10 wt% C16:0, PLA + 10 wt% C18:1 and PLA + 10 wt% PODC films. They were stretched at 5 mm·min<sup>-1</sup> using a homemade tensile machine placed under an Olympus Japan optical microscope mounted with WHK 10×/20 L ocular lens and MD Plan 10 0.25 objective lens. Observations were performed in optical transmission light mode using a Sony CCD-IRIS Model DXC-107P camera (France).

The morphology of the dispersed phase of PLA + 10 wt% C16:0, PLA + 10 wt% C18:1 and PLA + 10 wt% PODC materials before and after being stretched was observed with Scanning Electron Microscopy (SEM) (Hitachi 4800 II, France). Observations were directly conducted on the longitudinal surface of the dog bone shaped samples without any previous preparation.

### 2.6. Principal component analysis (PCA)

PCA was carried out considering the contents of each free fatty acid, mono, di and triglyceride types, hydrocarbons, sterols and water, the acid and saponification values and the measured elongation at break [%], Young modulus and  $T_{\rm g}$  values of the PLA formulations tested. PCA was done with XLstat software.

### 3. Results and discussion

### 3.1. Effects of free fatty acids, glycerides,

α-tocopherol and squalene on PLA ductility Individual components present in vegetable oils, which could feature toughening ability for PLA, as free fatty acids, glycerides, and the unsaponifiable components as α-tocopherol and squalene were tested. In order to achieve the most efficient formulation, the solubility of a molecule in the polymer matrix is one determinant for its plasticizing power [37]. Therefore the Hansen Solubility Parameters (HSP) of each compound was calculated. HSP (Table 4) shows low solubility of the different compounds in the PLA matrix. All the obtained values are close to one, which marks the solubility limit. The highest solubility (although still modest) was displayed by small fatty acids and α-tocopherol. The mechanical and thermal properties of the corresponding compounds are given in Table 6. The raw data of typical stress/strain curves of blends are presented in Figure 1. The DSC thermograms of neat additives in Figure 2 and of blends in Figure 3 for further information.

Literature studies on the use of vegetable oil as toughening agent of PLA showed that generally chemical modification is required to obtain positive effects [50, 52-54, 56-58, 72]. Not surprisingly due to their low solubility in PLA, the impact of the tested fatty acids (C16:0, C18:1), vegetable oils (HPO and HCO) and unsaponifiable compounds on  $T_g$  was negligible. However, despite the low solubility of the compounds, PLA ductility was increased in some cases. In particular, HCO, which contains mainly triglycerides, but with short chain length, improved the PLA ductility and lowered the apparent Young modulus, stress and elongation at yield. HPO, which mostly contains triglycerides and some diglycerides with higher chain length did not bring significant ductility enhancement nor  $T_{\rm g}$  decrease. HSP of di- and triglycerides from HPO and triglycerides from HCO

**Table 6.** Mechanical and thermal properties of formulations

	Formulation	Elongation at break [%]	app. Young modulus <sup>a</sup> [MPa]	Elongation at yield [%]	Stress at yield [MPa]	T <sub>g</sub> [°C]
PLA (tv	win screw pellets)	6±1	1760±90	4.2±0.4	64±5	56.3±0.2
PLA+	10 wt% C16:0	4±1	1720±60	3.2±0.1	50±3	49.3±0.1
PLA+	10 wt% C18:1	16±5	1440±50	2.2±0.1	28±2	45.2±0.2
PLA +	10 wt% [50 wt% C16:0 + 50 wt% C18:1]	45±12	1590±80	2.2±0.1	31±2	40.7±0.3
	10 wt% [95 wt% (50 wt% C16:0 + 50 wt% :1) + 5 wt% HPO]	74±15	1550±120	2.2±0.2	25±4	39.8±0.4
	5 wt% HCO	9±3	1680±70	3.6±0.2	51±1	54.1±0.2
PLA+	10 wt% HCO	9±6	1540±80	3.6±0.2	42±2	54.1±0.1
	15 wt% HCO	11±5	1460±70	3.8±0.2	38±2	$54.8 \pm 0.3$
	5 wt% HPO	55±20	1610±50	2.7±0.1	38±1	52.6±0.2
PLA+	10 wt% HPO	85±15	1420±70	2.6±0.2	26±2	54.4±0.4
	15 wt% HPO	91±14	1300±80	2.4±0.1	26±1	$53.9 \pm 0.3$
	5 wt% α-tocopherol	6±2	1150±90	3.8±0.2	41±4	51.3±0.2
PLA+	10 wt% α-tocopherol	5±2	1070±60	4.0±0.4	31±3	$49.8\pm0.2$
	15 wt% α-tocopherol	8±3	990±70	3.9±0.3	29±4	47.1±0.2
	5 wt% squalene	38±7	1180±90	3.3±0.4	36±7	51.4±0.2
PLA+	10 wt% squalene	51±9	990±60	3.1±0.9	28±3	49.5±0.2
	15 wt% squalene	29±7	890±80	2.9±0.7	19±2	49.4±0.1
	5 wt% RODC	22±3	1530±80	3.2±0.2	40±3	52.6±0.2
PLA+	10 wt% RODC	55±7	1490±140	2.4±0.2	28±4	49.1±0.1
PLA +	15 wt% RODC	65±15	1380±155	2.3±0.3	23±4	$48.8 \pm 0.1$
	20 wt% RODC	73±20	1270±150	2.2±0.3	20±4	48.5±0.3
	5 wt% SODC	29±8	1550±110	3.3±0.3	41±7	50.6±0.1
DT A	10 wt% SODC	73±18	1510±90	2.3±0.2	25±5	49.4±0.3
PLA+	15 wt% SODC	80±13	1340±80	2.2±0.3	23±3	$48.7 \pm 0.1$
	20 wt% SODC	52±15	1220±100	2.1±0.4	16±4	47.5±0.1
	5 wt% OODC	49±6	1430±90	3.1±0.3	34±4	49.5±0.2
DI A	10 wt% OODC	85±17	1490±90	2.6±0.2	31±2	49.0±0.1
PLA+	15 wt% OODC	88±16	1360±70	2.1±0.2	20±3	48.5±0.1
	20 wt% OODC	67±16	1370±110	2.2±0.2	21±3	48.7±0.1
	5 wt% PODC	51±17	1680±110	2.5±0.2	36±2	45.3±0.1
, .	10 wt% PODC	132±18	1460±40	2.1±0.2	24±1	39.2±0.2
PLA+	15 wt% PODC	179±15	1180±130	2.3±0.3	18±2	35.1±0.2
	20 wt% PODC	84±31	1130±60	2.0±0.4	14±3	33.8±0.2

<sup>&</sup>lt;sup>a</sup>apparent Young modulus for comparison reasons obtained without extensiometer

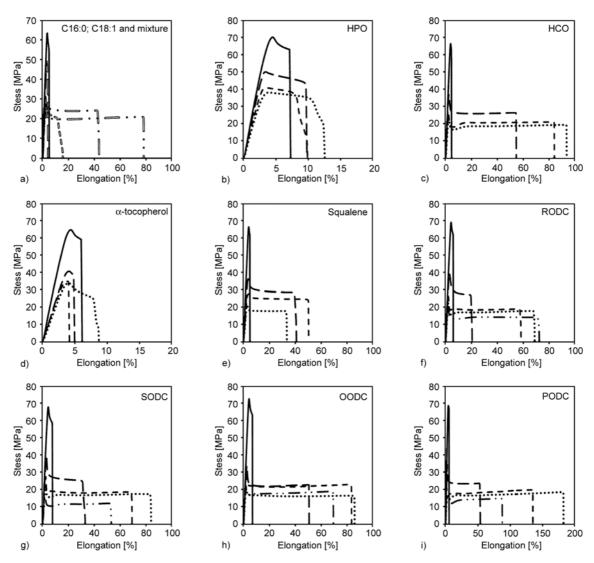
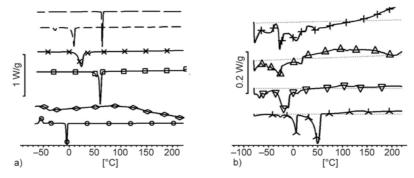


Figure 1. Stress/strain curves of PLA blends with free fatty acids, hydrogenated vegetable oils (HPO and HCO), unsaponifiable compounds (squalene, α-tocopherol), and vegetable oil deodorization condensates (palm oil PODC, olive oil OODC, soybean oil SODC and rapeseed oil RODC); (—) neat PLA; (== ==) PLA + 10 wt% C16:0; (= =) PLA + 10 wt% C18:1; (== •) PLA + 10 wt% [50 wt% C16:0 + 50 wt% C18:1]; (== •) PLA + 10 wt% [95 wt% (50 wt% C16:0 + 50 wt% C18:1) + 5 wt% HPO]; (——) PLA + 5 wt% additive; (- - -) PLA + 10 wt% additive; (• •) PLA + 15 wt% additive; (—• •) PLA + 20 wt% additive



**Figure 2.** Thermograms (first heating scan) of free fatty acids, hydrogenated vegetable oils (HPO and HCO), unsaponifiable compounds (squalene, α-tocopherol), and vegetable oil deodorization condensates (palm oil PODC, olive oil OODC, soybean oil SODC and rapeseed oil RODC); (— —) C16:0; (− –) C18:1; (×) HCO; (□) HPO; (◊) α-tocopherol; (⋄) squalene; (+) RODC; (△) SODC; (▽) OODC; (▲) PODC

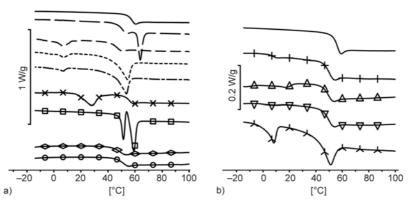


Figure 3. Thermograms (second heating scan) of PLA blends with free fatty acids, hydrogenated vegetable oils (HPO and HCO), unsaponifiable compounds (squalene, α-tocopherol), and vegetable oil deodorization condensates (palm oil PODC, olive oil OODC, soybean oil SODC and rapeseed oil RODC); (—) neat PLA; (— —) PLA + 10 wt% C16:0; (— –) PLA + 10 wt% C18:1; (— - –) PLA + 10 wt% [50 wt% C16:0 + 50 wt% C18:1]; (— -) PLA + 10 wt% [95 wt% (50 wt% C16:0 + 50 wt% C18:1) + 5 wt% HPO]; (×) PLA + 10 wt% HCO; (□) PLA + 10 wt% HPO; (◊) PLA + 10 wt% α-tocopherol; (⋄) PLA + 10 wt% squalene; (+) PLA + 10 wt% RODC; (Δ) PLA + 10 wt% SODC; (∇) PLA + 10 wt% OODC; (Λ) PLA + 10 wt% PODC

were close, while the average molar volume and molecular weight of triglycerides from HCO are smaller than the ones from HPO (Table 2). Testing of single fatty acids with longer chain length (C16:0, C18:1) showed that they were not able to increase PLA ductility. This is coherent with existing literature. For example, Jacobsen and Fritz [73] studied PLA blends with fatty acid esters. No significant ductility improvement was observed. The blending with the unsaponifiable compound  $\alpha$ -tocopherol showed small gains in elongation at break, which extends already existing knowledge. It was already shown that the use of  $\alpha$ -tocopherol as a natural antioxidant in PLA at small quantities (<4 wt%) induced a slightly  $T_{\rm g}$  decrease [74] but no increase in elongation at break [75]. Here, no elongation at break improvement upon was found using greater α-tocopherol amounts, despite solubility was assessed to be higher than other molecules (such as HPO and HCO, Table 4) and a  $T_{\rm g}$  decrease was observed (Table 3). Squalene, which is a natural tri-terpene found in vegetable oils, showed some toughening abilities (Table 6) despite its low solubility in PLA (Table 4). In fact, the creation of a dispersed phase promoting crazing was most probably responsible for this effect [37].

The most interesting result was obtained upon mixing different compounds. A mix of C18:1 and C16:0 fatty acids increased the PLA ductility much more than using them separately. The Young modulus and the  $T_{\rm g}$  remained higher than using only C18:1 likely because of its lower content involved, while the stress and elongation at yield were lessened as

much as adding only C18:1. Mixture made of [95 wt% (50 wt% C16:0 + 50 wt% C18:1) + 5 wt% HPO] at a total content of 10 wt% in PLA, further enhanced the ductility. There was apparently a synergistic effect between compounds, which can be possibly exploited.

## 3.2. Properties of PLA blends with vegetable oil deodorization condensates

Deodorization oil condensates are mixtures of different molecules contained in vegetable oils, the composition of which depends on the botanic source and oil refinery process. Four kinds of oil deodorization condensates (RODC, SODC, OODC and PODC) were blended with PLA, each at four concentrations (5, 10, 15 and 20 wt%). As shown in Table 3, content in free fatty acids, mono di or triglycerides, sterols (mainly alpha-tocopherol) and hydrocarbons (mainly squalene) was highly variable, as well as unsaturation degree of fatty acids. From the composition, an average HSP was calculated and is given in Table 4. As observed in Table 4, the longer and/or more unsaturated the alkyl chain is, the lower the solubility in PLA. However, except for very short fats alkyl chains and organic compounds as α-tocopherol or squalene, HSP of involved molecules appear to be rather similar. Mechanical and thermal properties of PLA/ oil deodorization condensates blends are given in Table 6 and typical curves of raw data are shown in the Figure 1. Significant increases in elongation at break were obtained, especially with the PODC. Addition of too much additive, i.e. about 20 wt%, induced a stagnation or decrease in the elongation at break. All the deodorization condensates led to a

lowering in the stress and elongation at yield. Interestingly, the apparent Young modulus of materials remained high. The stress/strain curves (Figure 1) showed that the yielding peak shrunk and flattened with the increase in additive content, as if the yield critical stress would tend to meet the plateau value where the stress remains constant with the strain. There is thus an important gain of using native mixtures of fatty acids present in deodorisation condensates for PLA ductility increase.

## 3.3. Study of the deformation mechanisms involved in PLA/oil deodorization condensates blends

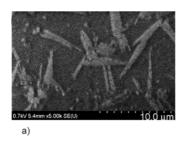
In all samples the  $T_{\rm g}$  remained higher than the measurement temperature, the main deformation mechanism was thus crazing of the glassy polymer [37]. The yield process in glassy amorphous polymers can be described as a stress induced glass-transition. In fact, because PLA Poisson's ratio is less than  $0.5 (v_{PLA} = 0.36)$  [76], the volume of PLA increases when subjected to tensile stress. Correspondingly, samples experienced important stress whitening, which appeared simultaneously with yielding. Stress whitening is caused by the formation of sizeable microvoids in the polymer matrix due to cavitation and crazes [77]. Crazes can also be initiated inside non-miscible inclusions in the polymer matrix or on the interface between inclusion and polymer in the case of low compatibility. The addition of oil deodorization condensates initiated crazes at stress levels substantially below those of the brittle failure of neat PLA, which propagated perpendicularly to the stretching direction. Morphological analysis of the form and size of inclusions of PODC, C16:0 and C18:1 was done to help interpretation. Figure 4 shows SEM micrographs of [PLA/C16:0], [PLA/ C18:1] and [PLA/PODC] 90 wt%/ 10 wt% blends. [PLA/C16:0] blends exhibit rods of about 5 to 10 μm length and 1 to 2 μm width, corresponding to fatty acid crystals. [PLA/C18:1] showed small spherical domains of fatty acids of about  $0.5~\mu m$  diameters and some aggregates of about 3 to  $5~\mu m$  diameters. The [PLA/PODC] blend micrograph showed both rods and aggregates of droplets, where the droplets appear to be distributed all around the crystals.

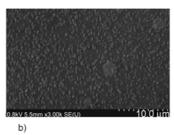
Figure 5 shows optical micrographs in transmission mode of stretched samples under the microscope taken at different percentages of elongation. Many small cracks started appear in neat PLA when approaching the yield peak, which is common for glassy amorphous polymers. Addition of C16:0, *i.e.* rod like crystals, increased both the occurrence and the length of cracks. Cracks quantity upon stretching caused the material to become opaque.

Failure of the blend revealed PLA fibrils. C18:1, *i.e.* small spherical liquid domains, induced fewer but much larger cracks as if their extensibility was eased. PODC, *i.e.* mostly a combination of C16:0 and C18:1 and some minor constituents, showed a high number of cracks but with smaller width as if using only C18:1. The mixture of palmitic acid crystals and liquid inclusions of oleic acid had thus a synergistic effect allowing for efficient craze initiation by the crystals and by cavitation inside the liquid inclusions. The superior performance of the PODC blends seems thus to be linked to the chemical composition in fatty acids and the presence of the minor constituents.

## 3.4. Role of the chemical composition of deodorization condensates in PLA ductility improvement

A Principal Components Analysis (PCA) was carried out including composition data and physicochemical characteristics of the deodorization condensates and of the PLA samples. Figure 6 shows the principal components projection plot of F1 and F2 of the data set. Only 50% of the total variance in these PLA samples is extracted according to F1 and F2 axes. In fact, scores of formulations containing





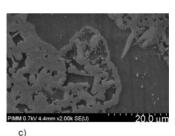
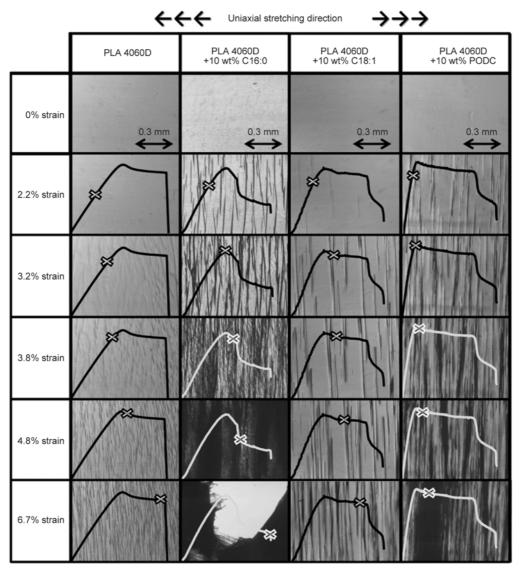
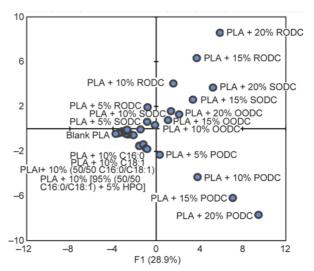


Figure 4. SEM micrographs of (a) C16:0, (b) C18:1 and (c) PODC blends with PLA at 10 wt%



**Figure 5.** Optical micrographs of uniaxially stretched PLA and PLA blends with free fatty acids (C16:0, C18:1) and palm oil deodorization condensate (PODC) at different strain

squalene, α-tocopherol, HPO, HCO and mixtures of C16:0, C18:1 and HPO are not discriminated. However discrimination of PLA samples containing deodorization condensates of each kind of vegetable oils and according to their respective amount is obtained. Corresponding loadings are plotted in Figure 7. The positive part of F1 axis appears to be mainly governed by the elongation at break and as opposed to its negative part with the  $T_{\rm g}$ . The F2 axis mostly separates fats as a function of the unsaturation and length of their alkyl chains. Looking at the elongation at break improvement, unsaturation of alkyl chains appears to be required, but the lower the ratio, the larger the enhancement was. Medium alkyl chain length, i.e. from lauric (C12) to stearic (C18) acid, also tends to be preferable than the long



**Figure 6.** Principal components projection plot of F1 and F2 of the data set

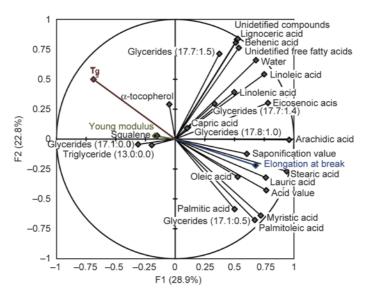


Figure 7. Loadings plot of the Principal Components Analysis of the dataset

ones as behenic (C22) or lignoceric (C24) acids. Both the unsaturation and the length of alkyl chains are main physical factors governing the melting point of fats. One common point is that unsaturation decreases melting points while length increases it. Therefore, efficiency of fats in toughening PLA appeared to be linked to medium melting point properties. A mixture of crystalline and liquid fatty acids at room temperature seemed favorable. This could explain the ability of HCO triglycerides to increase the PLA ductility while HPO did not, although their HSP were similar, as their ability to depress the  $T_{\rm g}$  of PLA. In fact, due to short alkyl chains, HCO triglycerides are waxy at room temperature. The DSC thermogram (Figure 2) shows a broad melting peak going from 8 to 42°C of HCO while it is beyond 55°C for HPO. Literature shows that combining liquid and solid additives can sometimes be effective for PLA [78]. Acid and saponification values are related to the elongation at break improvement (Figure 7). Therefore, the higher the esterified fats and/or the free fatty acid contents, the better the efficiency of the additive. In opposite, unidentified compounds, i.e. mostly unsaponifiable compounds, are not favorable to the ductility increase. Therefore, the superiority in toughening abilities of the PODC compared to the alike mixture made of [95 wt% (50 wt% C16:0 + 50 wt% C18:1) + 5 wt% HPO] can be attributed to the complex and favorable mixture of fatty acids having different chain length.

#### 4. Conclusions

By-products of the vegetable oils industry, namely deodorization condensates, were investigated as biobased additives for ductility improvement of PLA. The deodorization condensates improved substantially the elongation at break of PLA up to 180%. The glass transition was merely decreased which brought ductility to the still glassy polymer. The deformation mechanism was efficient craze initiation delaying failure. Most importantly, it was shown that the industrial by-product of the palm oil refinery had superior properties compared to mixtures of fatty acids and vegetable oils because it contained a mixture of fatty acids with melting points below and beyond room temperature. This makes deodorization condensates efficient and low price additives for PLA able to be used in commodity applications such as food packaging.

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#### References

[1] Auras R., Harte B., Selke S.: An overview of polylactides as packaging materials. Macromolecular Bioscience, 4, 835–864 (2004).

DOI: 10.1002/mabi.200400043

- [2] Auras R. A., Singh S. P., Singh J. J.: Evaluation of oriented poly(lactide) polymers vs. existing PET and oriented PS for fresh food service containers. Packaging Technology and Science, 18, 207–216 (2005). DOI: 10.1002/pts.692
- [3] Petersen K., Væggemose Nielsen P., Bertelsen G., Lawther M., Olsen M., Nilsson N., Mortensen G.: Potential of biobased materials for food packaging. Trends in Food Science and Technology, 10, 52-68 (1999). DOI: 10.1016/S0924-2244(99)00019-9
- [4] Solarski S., Ferreira M., Devaux E., Fontaine G., Bachelet P., Bourbigot S., Delobel R., Coszach P., Murariu M., Da Silva Ferreira A., Alexandre M., Degee P., Dubois P.: Designing polylactide/clay nanocomposites for textile applications: Effect of processing conditions, spinning, and characterization. Journal of Applied Polymer Science, 109, 841-851 (2008). DOI: 10.1002/app.28138
- [5] Zhang J-F., Sun X.: Mechanical and thermal properties of poly(lactic acid)/starch blends with dioctyl maleate. Journal of Applied Polymer Science, 94, 1697–1704 (2004).DOI: 10.1002/app.21078
- [6] Södergård A., Stolt M.: Properties of lactic acid based polymers and their correlation with composition. Progress in Polymer Science, 27, 1123–1163 (2002). DOI: 10.1016/S0079-6700(02)00012-6
- [7] Amass W., Amass A., Tighe B.: A review of biodegradable polymers: Uses, current developments in the synthesis and characterization of biodegradable polyesters, blends of biodegradable polymers and recent advances in biodegradation studies. Polymer International, 47, 89–144 (1998).
  - DOI: 10.1002/(SICI)1097-0126(1998100)47:2<89:: AID-PI86>3.0.CO;2-F
- [8] Kaplan D. L.: Biopolymers from renewable resources. Springer, Heidelberg (1998). DOI: 10.1007/978-3-662-03680-8
- [9] Lowe C.: Preparation of high molecular weight polyhydroxyacetic ester. U.S. Patent 2668162 A, USA (1954).
- [10] Gruber P. R., Hall E. S., Kolstad J. J., Iwen M. L., Benson R. D., Borchardt R. L.: Continuous process for the manufacture of lactide and lactide polymers. U.S. Patent 6326458, USA (2001).
- [11] Gruber P. R., Hall E. S., Kolstad J. J., Iwen M. L., Benson R. D., Borchardt R. L.: Continuous process for manufacture of lactide polymers with purification by distillation. U.S. Patent 5357035, USA (1994).
- [12] Avérous L.: Monomers, polymers and composites from renewable resources. in 'Polylactic acid: Synthesis, properties and applications' Elsevier, Amsterdam, 433-450 (2008).
- [13] Siracusa V., Rocculi P., Romani S., Rosa M. D.: Biodegradable polymers for food packaging: A review. Trends in Food Science and Technology, 19, 634–643

DOI: 10.1016/j.tifs.2008.07.003

- [14] Liu H., Zhang J.: Research progress in toughening modification of poly(lactic acid). Journal of Polymer Science Part B: Polymer Physics, 49, 1051–1083 (2011). DOI: 10.1002/polb.22283
- [15] Li Y., Shimizu H.: Toughening of polylactide by melt blending with a biodegradable poly(ether)urethane elastomer. Macromolecular Bioscience, 7, 921–928 (2007). DOI: 10.1002/mabi.200700027
- [16] NatureWorks. Technology focus report: Toughened PLA. Ver.3/1/2007 (2007).
- [17] Zhang W., Chen L., Zhang Y.: Surprising shape-memory effect of polylactide resulted from toughening by polyamide elastomer. Polymer, **50**, 1311–1315 (2009). DOI: 10.1016/j.polymer.2009.01.032
- [18] Byrne F., Ward P. G., Kennedy J., Imaz N., Hughes D., Dowling D. P.: The effect of masterbatch addition on the mechanical, thermal, optical and surface properties of poly(lactic acid). Journal of Polymers and the Environment, 17, 28-33 (2009). DOI: 10.1007/s10924-009-0119-x
- [19] Murariu M., Da Silva Ferreira A., Degée P., Alexandre M., Dubois P.: Polylactide compositions. Part 1: Effect of filler content and size on mechanical properties of PLA/calcium sulfate composites. Polymer, 48, 2613-2618 (2007).
  - DOI: 10.1016/j.polymer.2007.02.067
- [20] Murariu M., Da Silva Ferreira A., Duquesne E., Bonnaud L., Dubois P.: Polylactide (PLA) and highly filled PLA - Calcium sulfate composites with improved impact properties. Macromolecular Symposia, 272, 1-12 (2008).
  - DOI: 10.1002/masy.200851201
- [21] Hughes P. A., Becraft M. L., Opusko S.: Polymeric blend comprising polylactic acid. U.S. Patent 20070255013 A1, USA (2007).
- [22] Brulé B., Fine T., Flat J-J., Yasuda M.: Composite based on polylactic acid and polyamide, having improved impact resistance, its manufacturing process and use. World Patent WO2007144543 (2007).
- [23] Markarian J.: Biopolymers present new market opportunities for additives in packaging. Plastics, Additives and Compounding, 10, 22-25 (2008). DOI: 10.1016/S1464-391X(08)70091-6
- [24] Jin H-J., Chin I-J., Kim M-N., Kim S-H., Yoon J-S.: Blending of poly(L-lactic acid) with poly(cis-1,4-isoprene). European Polymer Journal, 36, 165–169 (2000). DOI: 10.1016/S0014-3057(99)00041-5
- [25] Li Y., Shimizu H.: Improvement in toughness of poly (L-lactide) (PLLA) through reactive blending with acrylonitrile-butadiene-styrene copolymer (ABS): Morphology and properties. European Polymer Journal, **45**, 738–746 (2009). DOI: 10.1016/j.eurpolymj.2008.12.010
- [26] McCarthy S. P., Gross R. A., Ma W.: Polylactic acid based blends. U.S. Patent 5883199 A, USA (1999).

- [27] Baiardo M., Frisoni G., Scandola M., Rimelen M., Lips D., Ruffieux K., Wintermantel E.: Thermal and mechanical properties of plasticized poly(L-lactic acid). Journal of Applied Polymer Science, 90, 1731–1738 (2003).
  - DOI: 10.1002/app.12549
- [28] Ljungberg N., Andersson T., Wesslén B.: Film extrusion and film weldability of poly(lactic acid) plasticized with triacetine and tributyl citrate. Journal of Applied Polymer Science, 88, 3239–3247 (2003). DOI: 10.1002/app.12106
- [29] Murariu M., Da Silva Ferreira A., Alexandre M., Dubois P.: Polylactide (PLA) designed with desired end-use properties: 1. PLA compositions with low molecular weight ester-like plasticizers and related performances. Polymers for Advanced Technologies, 19, 636–646 (2008). DOI: 10.1002/pat.1131
- [30] Lemmouchi Y., Murariu M., Dos Santos A. M., Amass A. J., Schacht E., Dubois P.: Plasticization of poly(lactide) with blends of tributyl citrate and low molecular weight poly(D,L-lactide)-b-poly(ethylene glycol) copolymers. European Polymer Journal, 45, 2839– 2848 (2009).
  - DOI: <u>10.1016/j.eurpolymj.2009.07.006</u>
- [31] Wang R. Y., Wan C. Y., Wang S. F., Zhang Y.: Morphology, mechanical properties, and durability of poly (lactic acid) plasticized with di(isononyl) cyclohexane-1,2-dicarboxylate. Polymer Engineering and Science, 49, 2414–2420 (2009).
  DOI: 10.1002/pen.21490
- [32] Sierra J., Noriega M., Cardona E., Ospina S.: Relationship between properties, citrate content and postproduction time for a plasticized polylactic acid. in 'Proceedings of ANTEC 2010, Orlando, USA' 127–132 (2010).
- [33] Courgneau C., Domenek S., Guinault A., Averous L., Ducruet V.: Analysis of the structure-properties relationships of different multiphase systems based on plasticized poly(lactic acid). Journal of Polymers and the Environment, 19, 362–371 (2011). DOI: 10.1007/s10924-011-0285-5
- [34] Labrecque L. V., Kumar R. A., Davé V., Gross R. A., McCarthy S. P.: Citrate esters as plasticizers for poly (lactic acid). Journal of Applied Polymer Science, 66, 1507–1513 (1997).
  - DOI: <u>10.1002/(SICI)1097-4628(19971121)66:8<1507</u> ::AID-APP11>3.0.CO;2-0
- [35] Ljungberg N., Andersson T., Wesslén B.: Film extrusion and film weldability of poly(lactic acid) plasticized with triacetine and tributyl citrate. Journal of Applied Polymer Science, 88, 3239–3247 (2003). DOI: 10.1002/app.12106
- [36] Yu J., Wang N., Ma X.: Fabrication and characterization of poly(lactic acid)/acetyl tributyl citrate/carbon black as conductive polymer composites. Biomacromolecules, 9, 1050–1057 (2008).
  DOI: 10.1021/bm7012857

- [37] Ruellan A., Guinault A., Sollogoub C., Ducruet V., Domenek S.: Solubility factors as screening tools of biodegradable toughening agents of polylactide. Journal of Applied Polymer Science, in press (2015). DOI: 10.1002/APP.42476
- [38] Kulinski Z., Piorkowska E., Gadzinowska K., Stasiak M.: Plasticization of poly(L-lactide) with poly(propylene glycol). Biomacromolecules, 7, 2128–2135 (2006). DOI: 10.1021/bm060089m
- [39] Kulinski Z., Piorkowska E.: Crystallization, structure and properties of plasticized poly(L-lactide). Polymer, 46, 10290–10300 (2005). DOI: 10.1016/j.polymer.2005.07.101
- [40] Hu Y., Hu Y. S., Topolkaraev V., Hiltner A., Baer E.: Crystallization and phase separation in blends of high stereoregular poly(lactide) with poly(ethylene glycol). Polymer, 44, 5681–5689 (2003). DOI: 10.1016/S0032-3861(03)00609-8
- [41] Jacobsen S., Fritz H. G.: Plasticizing polylactide The effect of different plasticizers on the mechanical properties. Polymer Engineering and Science, 39, 1303– 1310 (1999). DOI: 10.1002/pen.11517
- [42] Martin O., Averous L.: Poly(lactic acid): Plasticization and properties of biodegradable multiphase systems. Polymer, **42**, 6209–6219 (2001). DOI: 10.1016/S0032-3861(01)00086-6
- [43] Pillin I., Montrelay N., Grohens Y.: Thermo-mechanical characterization of plasticized PLA: Is the miscibility the only significant factor? Polymer, 47, 4676–4682 (2006).
  - DOI: 10.1016/j.polymer.2006.04.013
- [44] Sheth M., Kumar R. A., Davé V., Gross R. A., McCarthy S. P.: Biodegradable polymer blends of poly(lactic acid) and poly(ethylene glycol). Journal of Applied Polymer Science, 66, 1495–1505 (1997).
  - DOI: 10.1002/(SICI)1097-4628(19971121)66:8<1495 ::AID-APP10>3.0.CO;2-3
- [45] Martino V., Ruseckaite R., Jiménez A.: Thermal and mechanical characterization of plasticized poly(*L*-lactide-*co-D,L*-lactide) films for food packaging. Journal of Thermal Analysis and Calorimetry, **86**, 707–712 (2006).
  - DOI: 10.1007/s10973-006-7897-3
- [46] Martino V. P., Jiménez A., Ruseckaite R. A.: Processing and characterization of poly(lactic acid) films plasticized with commercial adipates. Journal of Applied Polymer Science, **112**, 2010–2018 (2009). DOI: 10.1002/app.29784
- [47] Ljungberg N., Wesslén B.: Thermomechanical film properties and aging of blends of poly(lactic acid) and malonate oligomers. Journal of Applied Polymer Science, **94**, 2140–2149 (2004).

  DOI: 10.1002/app.21100
- [48] Ren Z., Dong L., Yang Y.: Dynamic mechanical and thermal properties of plasticized poly(lactic acid). Journal of Applied Polymer Science, 101, 1583–1590 (2006). DOI: 10.1002/app.23549

- [49] Rahman M., Brazel C. S.: The plasticizer market: An assessment of traditional plasticizers and research trends to meet new challenges. Progress in Polymer Science, 29, 1223–1248 (2004).
  - DOI: 10.1016/j.progpolymsci.2004.10.001
- [50] Robertson M. L., Chang K., Gramlich W. M., Hillmyer M. A.: Toughening of polylactide with polymerized soybean oil. Macromolecules, 43, 1807–1814 (2010). DOI: 10.1021/ma9022795
- [51] Gramlich W. M., Robertson M. L., Hillmyer M. A.: Reactive compatibilization of poly(l-lactide) and conjugated soybean oil. Macromolecules, 43, 2313–2321 (2010).

DOI: 10.1021/ma902449x

- [52] Ali F., Chang Y-W., Kang S. C., Yoon J. Y.: Thermal, mechanical and rheological properties of poly (lactic acid)/epoxidized soybean oil blends. Polymer Bulletin, 62, 91–98 (2009). DOI: 10.1007/s00289-008-1012-9
- [53] Xu Y-Q., Qu J-P.: Mechanical and rheological properties of epoxidized soybean oil plasticized poly(lactic acid). Journal of Applied Polymer Science, 112, 3185–3191 (2009).
  DOI: 10.1002/app.29797
- [54] Xiong Z., Yang Y., Feng J., Zhang X., Zhang C., Tang Z., Zhu J.: Preparation and characterization of poly(lactic acid)/starch composites toughened with epoxidized soybean oil. Carbohydrate Polymers, 92, 810–816 (2012).
  DOI: 10.1016/j.carbpol.2012.09.007
- [55] Al-Mulla E. A. J., Suhail A. H., Aowda S. A.: New biopolymer nanocomposites based on epoxidized soybean oil plasticized poly(lactic acid)/fatty nitrogen compounds modified clay: Preparation and characterization. Industrial Crops and Products, 33, 23–29 (2010). DOI: 10.1016/j.indcrop.2010.07.022
- [56] Silverajah V. S. G., Ibrahim N. A., Zainuddin N., Yunus W., Abu Hassan H.: Mechanical, thermal and morphological properties of poly(lactic acid)/epoxidized palm olein blend. Molecules, 17, 11729–11747 (2012). DOI: 10.3390/molecules171011729
- [57] Silverajah V. S. G., Ibrahim N. A., Yunus W., Abu Hassan H., Woei C. B.: A comparative study on the mechanical, thermal and morphological characterization of poly(lactic acid)/epoxidized palm oil blend. International Journal of Molecular Sciences, 13, 5878–5898 (2012).

DOI: 10.3390/ijms13055878

- [58] Al-Mulla E. A. J., Yunus W. M. Z. W., Ibrahim N. A. B., Ab Rahman M. Z.: Properties of epoxidized palm oil plasticized polytlactic acid. Journal of Materials Science, 45, 1942–1946 (2010). DOI: 10.1007/s10853-009-4185-1
- [59] NatureWorks: Technology focus report: Polylactic acid containing fillers and fibres (2007).

- [60] Bitinis N., Fortunati E., Verdejo R., Bras J., Kenny J. M., Torre L., Lopez-Manchado M. A.: Poly(lactic acid)/ natural rubber/cellulose nanocrystal bionanocomposites. Part II: Properties evaluation. Carbohydrate Polymers, 96, 621–627 (2013). DOI: 10.1016/j.carbpol.2013.03.091
- [61] Yuan D., Chen K., Xu C., Chen Z., Chen Y.: Cross-linked bicontinuous biobased PLA/NR blends via dynamic vulcanization using different curing systems. Carbohydrate Polymers, 113, 438–445 (2014). DOI: 10.1016/j.carbpol.2014.07.044
- [62] Thepthawat A., Srikulkit K.: Improving the properties of polylactic acid by blending with low molecular weight polylactic acid-g-natural rubber. Polymer Engineering and Science, 54, 2770–2776 (2014). DOI: 10.1002/pen.23835
- [63] Ayutthaya W. D. N., Poompradub S.: Thermal and mechanical properties of poly(lactic acid)/natural rubber blend using epoxidized natural rubber and poly (methyl methacrylate) as *co*-compatibilizers. Macromolecular Research, 22, 686–692 (2014). DOI: 10.1007/s13233-014-2102-1
- [64] Hughes J., Thomas R., Byun Y., Whiteside S.: Improved flexibility of thermally stable poly-lactic acid (PLA). Carbohydrate Polymers, **88**, 165–172 (2012). DOI: 10.1016/j.carbpol.2011.11.078
- [65] Bradley J-C., Lang A., Williams A.: Jean-Claude Bradley double plus good (highly curated and validated) melting point dataset (2014). DOI: 10.6084/m9.figshare.1031638
- [66] Vereeckeen J., Foubert I., Meeussen W., Lesaffer A., Dewettinck K.: Fat structuring with partial acylglycerols: Effect on solid fat profiles. European Journal of Lipid Science and Technology, 111, 259–272 (2009). DOI: 10.1002/ejlt.200800151
- [67] Sato K.: Kinetic aspects in polymorphic crystallization and transformation of fats mixed and dispersed systems. in 'Physical properties of fats, oils, and emulsifiers' (ed.: Widlak N.) The American Oil Chemists Society, Champaign, 33–48 (1999).
- [68] Van Krevelen D. W., Te Nijenhuis K.: Cohesive properties and solubility. in 'Properties of polymers' (eds.: Van Krevelen D. W., Te Nijenhuis K.) Elsevier, Amsterdam, 189–228 (2009).
- [69] van Krevelen D. W.: Chemical structure and properties of coal. XXVIII. Coal constitution and solvent extraction. Fuel, 44, 229–242 (1965).
- [70] Van Krevelen D. W., Hoftyzer P. J.: Practical evaluation of the [η]–*M* relationship. III. Estimation of the exponent A. Journal of Applied Polymer Science, 11, 2189–2200 (1967).
  DOI: 10.1002/app.1967.070111109
- [71] Abbott S. J.: Chemical compatibility of poly (lactic acid): A practical framework using hansen solubility parameters. in 'Poly(lactic acid): Synthesis, structures, properties, processing, and applications' (eds.: Auras R., Lim L. T., Selke S., Tsuji H.) Wiley, Hoboken, 83–93 (2010).

- [72] Robertson M. L., Paxton J. M., Hillmyer M. A.: Tough blends of polylactide and castor oil. ACS Applied Materials and Interfaces, 3, 3402–3410 (2011). DOI: 10.1021/am2006367
- [73] Jacobsen S., Fritz H. G.: Plasticizing polylactide The effect of different plasticizers on the mechanical properties. Polymer Engineering and Science, 39, 1303– 1310 (1999).

DOI: 10.1002/pen.11517

[74] Manzanarez-López F., Soto-Valdez H., Auras R., Peralta E.: Release of α-tocopherol from poly(lactic acid) films, and its effect on the oxidative stability of soybean oil. Journal of Food Engineering, **104**, 508–517 (2011).

DOI: 10.1016/j.jfoodeng.2010.12.029

[75] Hwang S. W., Shim J. K., Selke S. E. M., Soto-Valdez H., Matuana L., Rubino M., Auras R.: Poly(L-lactic acid) with added α-tocopherol and resveratrol: Optical, physical, thermal and mechanical properties. Polymer International, 61, 418–425 (2012).

DOI: 10.1002/pi.3232

[76] Jamshidian M., Tehrany E. A., Imran M., Jacquot M., Desobry S.: Poly-lactic acid: Production, applications, nanocomposites, and release studies. Comprehensive Reviews in Food Science and Food Safety, 9, 552–571 (2010).

DOI: 10.1111/j.1541-4337.2010.00126.x

- [77] Lampman S.: Fracture and fractography. in 'Characterization and failure analysis of plastics' (ed.: Lampman S.) ASM International, Materials Park, 404–416 (2003).
- [78] Notta-Cuvier D., Odent J., Delille R., Murariu M., Lauro F., Raquez J. M., Bennani B., Dubois P.: Tailoring polylactide (PLA) properties for automotive applications: Effect of addition of designed additives on main mechanical properties. Polymer Testing, 36, 1–9 (2014).

DOI: 10.1016/j.polymertesting.2014.03.007