



HAL
open science

Imaging and quantitative analysis of size and distribution of sperical bodies, [i]e.g[/i]. embryonic oil bodies

Martine Miquel, Ghassen Trigui, Alain Trubuil, Bertrand Dubreucq

► To cite this version:

Martine Miquel, Ghassen Trigui, Alain Trubuil, Bertrand Dubreucq. Imaging and quantitative analysis of size and distribution of sperical bodies, [i]e.g[/i]. embryonic oil bodies. Bio-protocol , 2015, 5 (1), pp.1-6. hal-01204094

HAL Id: hal-01204094

<https://hal.science/hal-01204094>

Submitted on 27 May 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Imaging and Quantitative Analysis of Size and Distribution of Spherical Bodies, e.g. Embryonic Oil Bodies

Martine Miquel^{1*}, Ghassen Trigui^{2,3}, Alain Trubuil² and Bertrand Dubreucq^{1*}

¹Institut National de la Recherche Agronomique (INRA), Institut Jean-Pierre Bourgin, Versailles, France; AgroParisTech, Institut Jean-Pierre Bourgin, Versailles, France; Centre National de la Recherche Scientifique, Institut Jean-Pierre Bourgin, Versailles, France; ²Institut National de la Recherche Agronomique (INRA), Jouy en Josas, France; ³Present address: Geves: Groupe d'Étude et de contrôle des Variétés Et des Semences, rue Georges Morel, Beaucozézé, France

*For correspondence: martine.miquel@versailles.inra.fr; bertrand.dubreucq@versailles.inra.fr

[Abstract] Oil bodies (OBs) are seed-specific lipid storage organelles that allow the accumulation of neutral lipids that sustain plantlet development after the onset of germination. Using fluorescent dyes and confocal microscopy, we monitored the dynamics of OBs in living *Arabidopsis thaliana* embryos at different stages of development (Miquel *et al.*, 2014). Image acquisition was followed by a detailed statistical analysis of OB size and distribution during seed development in the four dimensions (x , y , z , and t).

Materials and Reagents

1. Plant material
 - a. *Arabidopsis thaliana* plants, wild type or mutant or transgenic plants.
 - b. Developing siliques between 6 and 11 days after pollination of plants grown in a greenhouse under the following conditions (13 h of light, diurnal temperature of 25 °C, and nocturnal temperature of 17 °C), and irrigated twice per week with mineral nutrient solution.
2. Nile Red (a neutral lipid stain, at 2 g/ml final in acetone) (Sigma-Aldrich, catalog number: 72485)

Equipment

1. Confocal microscope

Note: In this study an inverted LEICA SP2-AOBS spectral confocal laser microscope (Leica Microsystems) equipped with an HCX PL APO CS 40 x 1.25 objective and a multiline argon laser was used.
2. Forceps (Dumont No.5) (Sigma-Aldrich)

3. Scalpel (11 P, blade) (Swann Norton)
4. Glass slides or glass-bottom dish, cover slips
5. Binocular (Nikon Corporation, model: SMZ1000) (Champigny sur Marne with a led ring - Shott easyled™ for illumination)

Software

1. ND-SAFIR (<http://serpico.rennes.inria.fr/doku.php?id=software:nd-safir:index>)
2. AVIZO® FIRE (<http://www.vsg3d.com/avizo/fire>)

Procedure

A. Sample preparation

1. Remove developing seeds from 2-3 siliques under the binocular with a sharp scalpel and forceps (DUMONT N°5) and place them on a glass slide or in a glass-bottom dish.
2. Add a small volume (between 20 and 30 µl) of Nile Red solution and cover with a cover slip. Remove embryos from the seed teguments by gently pressing seeds between the slide and cover slip with the rounded end of the handle of the scalpel and observe after 30 min of incubation in the dark at room temperature (see Video 1).

Video 1. Dissecting embryos out of Arabidopsis developing siliques



Note: You can use a 60% (v/v) glycerol solution or 0.4 % (w/v) of low melting agarose as mounting medium for your staining solution to better immobilize the embryos for long observation periods.

B. Confocal microscopy

1. Find appropriate embryos and adjust the focus on cotyledon edges using a 40x objective.
2. Set optical parameters to optimize signal to noise ratio.

- a. Excitation laser: the argon laser is set at 10% of 488 nm for Nile Red excitation.
- b. Emission band of 550 to 650 nm.
- c. Capture the images (90 to 100 optical sections of 0.16 μm thickness) with a 40x/1.25 oil objective and 8x digital zooming. Each section is the average of two scans conducted at the resolution of 512 x 512 pixels with a spatial resolution of 0.09 mm x 0.09 mm x 0.16 μm in the x, y, z referential.

Notes:

- i. *The Nile Red stain is highly fluorescent when associated with neutral lipids. Nevertheless you can bleach it if using too high laser power for excitation. According to our experience, reducing the laser power to its minimal value and laser excitation to 10% of laser power is far enough for excellent observation and preserves fluorescence for up to one night.*
- ii. *As the fluorescence signal is usually very strong, you can decrease the pinhole's size to 0.7 Airy to reduce the thickness of the optical section.*
- iii. *Increasing scanning speed as well as using bidirectional-scanning mode will speed up acquisition.*

C. Quantitative analysis

1. Obtain images at a similar location from 3 different embryos for statistical analysis.
2. Analyze the images using the following pipeline:
 - a. The crude image stacks present several characteristics that disturb direct automated counting of the oil bodies (Figure 1). To circumvent this problem, the procedures for image segmentation by watershed and data extraction were implemented in AVIZO[®] FIRE in a semi-automated pipeline. A specific module (WshedBinSeparate) has been developed in TCL language (Tool Command Language). The outline of the procedure is described in Figure 2. A movie shows the sequence of operations (Video 2).

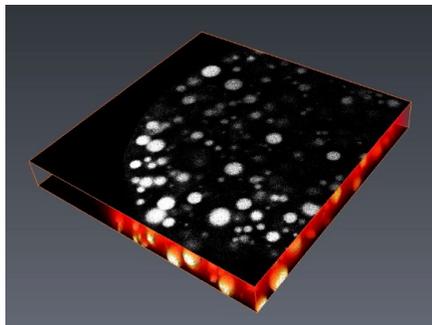


Figure 1. 2D/3D view of an image stack with various problems preventing direct use in image analysis. A. Images with high noise; B. Non homogenous objects in size; C. Variable grey levels; D. Objects with touching edges.

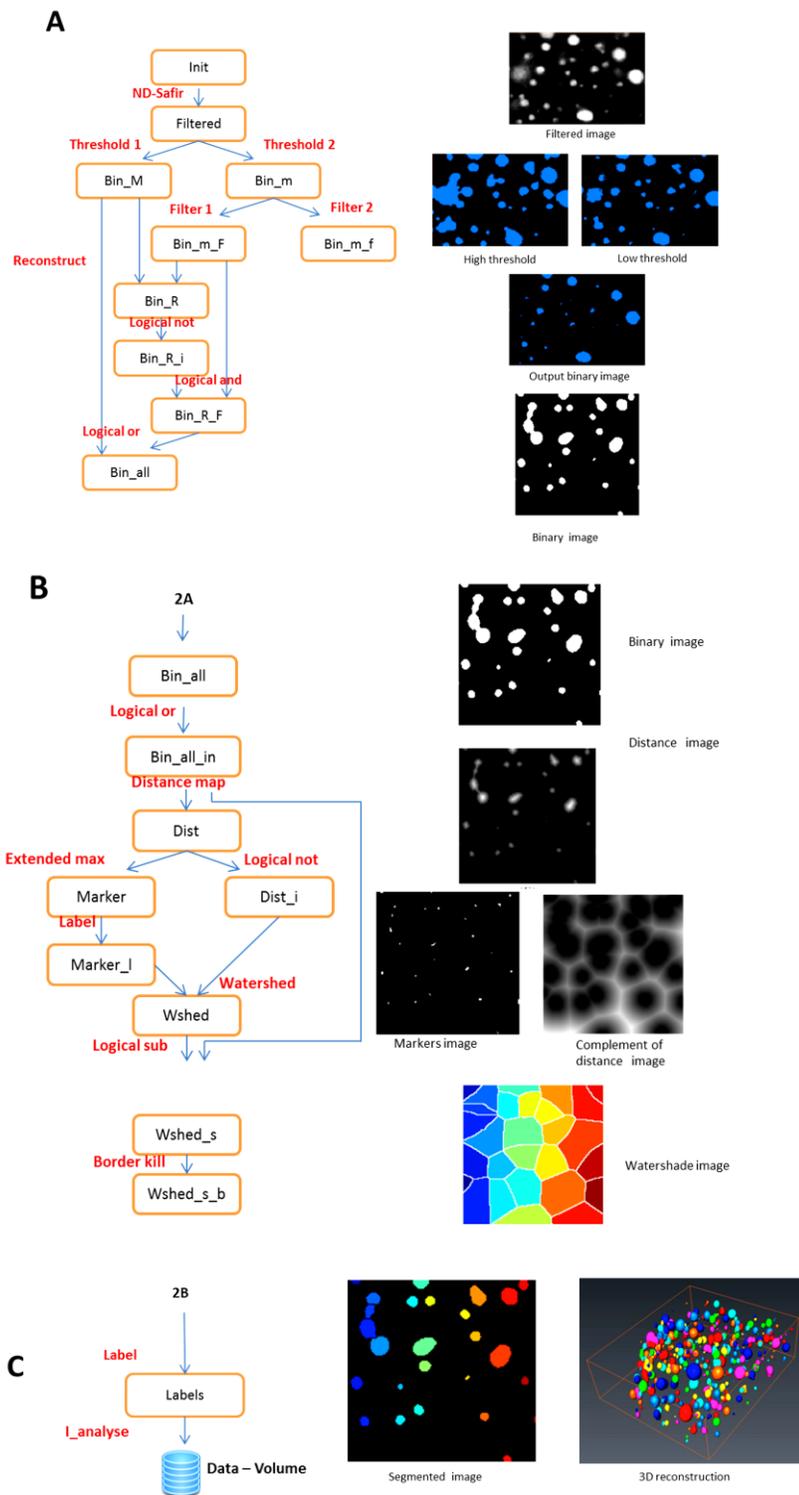
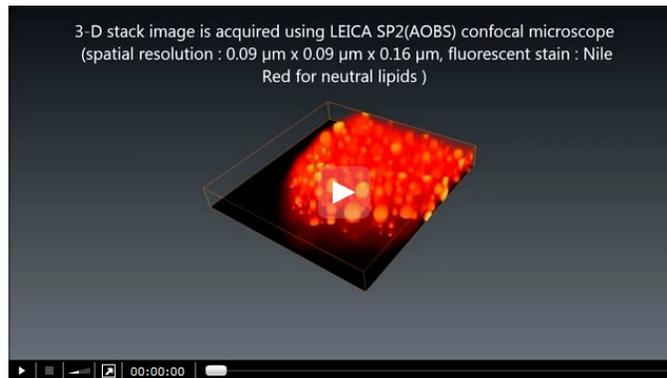


Figure 2. Image analysis pipeline. A. Generating binary images with double thresholding; B. Generating watersheded images; C. Individual labelling of objects and 3D rendering. The segmentation chain in Avizo (left) and the resulting images (right).

Video 2. OB image processing pipeline



- b. The WshedBinSeparate module allows segmentation of oil bodies images stacks by watershed methods.
- c. The input image is a 3D stack in grey scale. The images are first de-noised with a Gaussian filter using ND-SAFIR. Two thresholding operations are then performed with threshold values manually indicated by the operator. The best value is chosen by visual test of various threshold values.
- d. The high threshold allows the detection of large objects without worrying about over- or under-estimating them but without consideration of small objects.
- e. The low threshold allows the detection of small objects without worrying about the over- or under-estimation of the large objects. The images are combined to produce a thresholded “binary image”.
- f. The 3D distance transform of this binary image is computed (a value is given for each pixel of the image according to its distance to the border of the closest object in the vicinity).
- g. The maximum of the distance function produces the “marker image” *i.e.* the centre of the segmented quasi-spherical filled object. The complement of the distance transform (CDT), with the marker image is used to create the “watershed lines”: the watershed algorithm expands the markers (like inflating a balloon) toward increasing value of the CDT. The watershed lines represent the boundaries separating the objects. Finally, the watershed lines are subtracted from the binary image to obtain separated and individualized objects (“segmented image”). The image of separated objects is then converted to a spread sheet document where each item in the spread sheet is a labeled object volume.

Acknowledgments

This work was funded by INRA and, in particular, by the divisions BAP (Plant Biology and Breeding) and MIA (Applied Mathematics and Informatics) for G. Trigui's Ph.D. This work was adapted from Miquel *et al.* (2014).

References

1. Miquel, M., Trigui, G., d'Andrea, S., Kelemen, Z., Baud, S., Berger, A., Deruyffelaere, C., Trubuil, A., Lepiniec, L. and Dubreucq, B. (2014). [Specialization of oleosins in oil body dynamics during seed development in Arabidopsis seeds](#). *Plant Physiol* 164(4): 1866-1878.
2. Pierre, S. (2003). Morphological image analysis: Principles and Applications. 2nd edition. Springer.