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DMRT1 turns on testicular differentiation during XX sex-reversal in the goat

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Background

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In mammals, ovaries (XX) and testes (XY) differentiate from a common bipotential gonad. Testicular differentiation depends on the activation of *SRY* located on the Y chromosome. Numerous studies realized in the mouse demonstrated that SRY activates *SOX9* and the testis pathway. In females, the absence of *SRY* and the activation of several pro-ovarian genes (*ie. FOXL2, RSPO1, WNT4*) leads to ovarian differentiation.

• XX gonads lacking FOXL2 differentiate into testes in the goat

In goats, the Polled Intersex Syndrome (PIS) mutation is a natural mutation responsible for hornlessness and female-to-male sex-reversal [1]. Using a zinc-finger nuclease technology, we demonstrated that the XX sex-reversal is due to FOXL2 silencing in the differentiating XX PIS^{-} gonad. Thus, the XX gonads lacking FOXL2 differentiate into testes instead of ovaries [2]. To highlight (i) the morphogenetic mechanisms occuring downstream of FOXL2, and (ii) its targets implicated in ovarian differentiation and anti-testis action, we performed RNA-sequencing at the onset of gonadal switch in the goat (36 dpc), using XY testes, XX ovaries and XX PIS^{-} intersex (IS) gonads (**Fig. 1**).

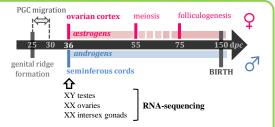
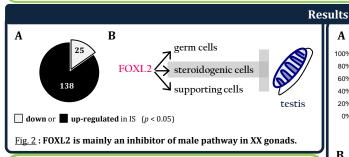


Fig. 1: Identification of FOXL2 target genes by RNA-sequencing at 36dpc.



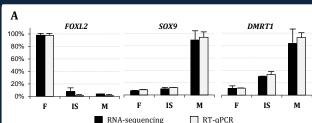
• FOXL2 acts mainly as a repressor in the XX-developping ovary.

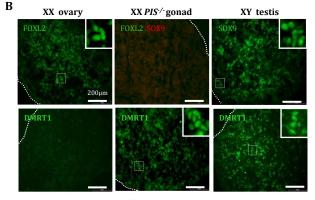
The RNA-sequencing results showed that 163 genes were deregulated in the IS gonads comparing to ovaries (**Fig.2**). 25 of them (15%) were down-regulated and may correspond to genes activated by FOXL2, whereas 138 genes were up-regulated and may correspond to genes inhibited by FOXL2 (**Fig. 2A**). So FOXL2 appeared to act mainly as a repressor in the developping ovary. Further analyses revealed that this repression impact all the cellular levels in XX gonads (**Fig. 2B**). For instance, FOXL2 indirectly protect XX germ cells fate and permit them to enter meiosis by repressing *CYP26B1*. At the steroidogenic level, FOXL2 avoids androgens synthesis by inhibiting *176HSD3* and it promotes estrogens production as it directly activates *CYP19* [3]. At least, FOXL2 prevents the expression of testes supporting cell-markers, such as *DMRT1*.

• Caracterization of maleness progression: role of DMRT1. Whereas neither FOXL2 nor SOX9 were expressed in the IS gonad

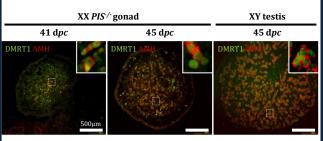
at 36 dpc (**Fig. 3A&B**), DMRT1 was already up-regulated. *SOX9* expression increases few days later [4], around 40 dpc. When we looked at maleness progression in XX *PIS* gonads (**Fig. 4**), we found that Sertoli-like cells (AMH and so SOX9-positive cells) first appeared at the center of the gonad, in DMRT1-expressing cells. Then at 45 dpc, AMH-staining and so Sertoli-like cells differentiation spread from the center to the

DMRT1-expressing cells. Then at 45 dpc, AMH-staining and so Sertoli-like cells differentiation spread from the center to the cortex of the gonad. Taking together, these results strongly suggested that DMRT1 is at the onset of maleness activation during sex-reversal in the goat.





 $\underline{\text{Fig. 3}}$: DMRT1 is up-regulated before SOX9 in XX gonads lacking FOXL2.



<u>Fig. 4</u>: Sertoli-like cells differentiate from DMRT1-expressing cells in XX PIS $^{\prime}$ gonads.

Conclusion and working model



In mice, Foxl2 conditionnal knock-out leads to ovary-to-testis transdifferentation at adulhood, starting with Sox9 up-regulation then followed by Dmrt1 expression [5]. In the goat, the absence of FOXL2 in the XX-developping gonad leads to sex-reversal as soon as fetal life [1,2,4]. Our results showed that contrary to the mouse, the primary event of this sex-reversal is DMRT1 up-regulation, followed by SOX9 expression. This supports that, in the goat (i) FOXL2 is a sex-determining gene, (ii) FOXL2 antagonizes male pathway in the developping ovary, and (iii) DMRT1 can be an activator of SOX9 and the male pathway.

<u>References</u> :

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