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RESEARCH

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A genome-wide association study of production traits in a commercial population of Large White pigs: evidence of haplotypes affecting meat quality

Marie-Pierre Sanchez^{1,2*}, Thierry Tribout^{1,2}, Nathalie Iannuccelli³, Marcel Bouffaud⁴, Bertrand Servin³, Amabel Tenghe^{1,2}, Patrice Dehais³, Nelly Muller⁴, Maria Pilar Del Schneider^{1,2}, Marie-José Mercat⁵, Claire Rogel-Gaillard^{1,2}, Denis Milan³, Jean-Pierre Bidanel^{1,2} and H  l  ne Gilbert³

Abstract

Background: Numerous quantitative trait loci (QTL) have been detected in pigs over the past 20 years using microsatellite markers. However, due to the low density of these markers, the accuracy of QTL location has generally been poor. Since 2009, the dense genome coverage provided by the Illumina PorcineSNP60 BeadChip has made it possible to more accurately map QTL using genome-wide association studies (GWAS). Our objective was to perform high-density GWAS in order to identify genomic regions and corresponding haplotypes associated with production traits in a French Large White population of pigs.

Methods: Animals (385 Large White pigs from 106 sires) were genotyped using the PorcineSNP60 BeadChip and evaluated for 19 traits related to feed intake, growth, carcass composition and meat quality. Of the 64 432 SNPs on the chip, 44 412 were used for GWAS with an animal mixed model that included a regression coefficient for the tested SNPs and a genomic kinship matrix. SNP haplotype effects in QTL regions were then tested for association with phenotypes following phase reconstruction based on the Sscrofa10.2 pig genome assembly.

Results: Twenty-three QTL regions were identified on autosomes and their effects ranged from 0.25 to 0.75 phenotypic standard deviation units for feed intake and feed efficiency (four QTL), carcass (12 QTL) and meat quality traits (seven QTL). The 10 most significant QTL regions had effects on carcass (chromosomes 7, 10, 16, 17 and 18) and meat quality traits (two regions on chromosome 1 and one region on chromosomes 8, 9 and 13). Thirteen of the 23 QTL regions had not been previously described. A haplotype block of 183 kb on chromosome 1 (six SNPs) was identified and displayed three distinct haplotypes with significant ($0.0001 < P < 0.03$) associations with all evaluated meat quality traits.

Conclusions: GWAS analyses with the PorcineSNP60 BeadChip enabled the detection of 23 QTL regions that affect feed consumption, carcass and meat quality traits in a LW population, of which 13 were novel QTL. The proportionally larger number of QTL found for meat quality traits suggests a specific opportunity for improving these traits in the pig by genomic selection.

* Correspondence: marie-pierre.sanchez@jouy.inra.fr

¹INRA, UMR1313 G  n  tique Animale et Biologie Int  grative, F-78350 Jouy-en-Josas, France

²INRA, AgroParisTech, UMR1313 G  n  tique Animale et Biologie Int  grative, F-78350 Jouy-en-Josas, France

Full list of author information is available at the end of the article

Background

Over the past 20 years, several whole-genome scans using mainly microsatellite markers have revealed quantitative trait loci (QTL) for a large number of traits in pigs. To date, a total of some 8300 QTL for more than 600 overlapping phenotypic traits have been reported in the PigQTLdb (<http://www.genome.iastate.edu/cgi-bin/QTLdb/SS/index>, March 21, 2013; [1]). However, because of the low density of microsatellite markers, these QTL are generally located with poor accuracy and additional long-term fine-mapping experiments are necessary to refine their positions and determine causative variants (e.g. [2,3]). Moreover, QTL linkage designs have in most cases been based on a limited number of families from crosses between divergent populations [4], resulting in limited mapping accuracy and QTL results that are not directly transferable to commercial populations.

The high-throughput genotyping of single nucleotide polymorphisms (SNPs) has become available for pigs with the Illumina PorcineSNP60 BeadChip [5]. The dense genome coverage provided by this chip makes it possible to exploit the linkage disequilibrium (LD) between SNPs and QTL through genome-wide association studies (GWAS). GWAS have been shown to be more powerful for accurate QTL mapping than linkage-based methods (see for example [6] in livestock). Since the PorcineSNP60 BeadChip has become available, several GWAS have been performed in commercial pig populations and have revealed significant associations for economically important traits such as boar taint [7], reproductive characteristics [8], body composition [9], pathogen susceptibility [10], hematological traits [11], feed efficiency [12,13], and meat quality [14]. GWAS have also been performed in a Large White × Minzhu inter-cross population for meat quality [15] and hematological traits [16]. However, to date results on dense GWAS are not available for other major traits in commercial pig populations such as growth rate.

The objective of this work was to perform a high-density genome-wide association study with the PorcineSNP60 BeadChip in the French Large White (LW) breed in order to map the genomic regions that are associated with growth efficiency, carcass and meat quality traits, and to identify haplotypes that may be suitable for inclusion in selection strategies.

Methods

Ethical statement

The animals involved in this study were reared and slaughtered in compliance with national regulations applicable to animal research and commercial slaughtering.

Animals and traits

A total of 385 French LW castrated male pigs from 29 LW nucleus herds were performance-tested at the French

national test station in Le Rheu (UETP, Le Rheu, Ille et Vilaine, France). Pigs descended from 106 sires and 313 dams and originated from 335 litters. The piglets entered UETP at 4 to 6 weeks of age and were placed in group pens in a post-weaning unit until they were 10 weeks old. They were then transferred to a fattening unit and placed in group pens for 12 animals equipped with Acema 64 single-place electronic feeders (Acemo, Pontivy, France). All pigs were weighed at the beginning (around 35 kg live weight) and end (around 110 kg live weight) of the performance test. During the test period, they were fed *ad libitum* with pellets composed of cereals and soybean meal containing 9.5 MJ net energy and 156 g crude protein per kg, with a minimum of 0.87 g digestible lysine per MJ of net energy. The average daily gain (ADG), daily feed intake (DFI) and feed conversion ratio (FCR) during the test period were calculated using individual weight measurements and data collected from the electronic feeders. Residual feed intake (RFI) was computed as the difference between DFI and a “theoretical” feed intake predicted from maintenance and production requirements using a phenotypic multiple linear regression method, as described in [17]. When pigs reached 110 kg live weight, they were fasted for a minimum of 16 hours and then transported for about 35 minutes to be slaughtered in a commercial abattoir (Cooperl, Montfort-sur-Meu).

Carcass weight and length were measured shortly after slaughter. Carcass length (CL) was determined from the atlas to the anterior edge of the pubian symphysis. Average carcass backfat thickness (CBF) was computed as the mean of carcass fat depths at the shoulder, the last rib, and the hip joint. Carcasses with head and feet but without kidney fat were then chilled in a cooling room at 4°C for 24 hours. Dressing percentage (DP) was defined as the ratio of cold carcass weight to slaughter weight measured after the fasting period prior to transportation. The day after slaughter, the right half-carcass (RHC) (without head) was weighed and then cut based on a standardized procedure [18]. The ham, loin, belly, shoulder and backfat were weighed separately (HAM, LOIN, BELLY, SHOULDER and BACKFAT, respectively). Lean meat content was then estimated using the cut weights as $ELMC (\%) = 25.08 - (1.23 \times (100 \times BACKFAT/RHC)) + (0.87 \times (100 \times LOIN/RHC)) + (0.73 \times (100 \times (100 \times HAM/RHC)))$ [19].

Several meat quality traits were recorded on the day after slaughter. Ultimate pH was measured on the *semi-membranosus* (pH24 SM) muscle at 4°C using a Knick Portaness 910 pH meter (Knick GmbH & Co., Berlin, Germany) equipped with a Mettler Toledo Probe (Mettler-Toledo International Inc., Urdorf, Switzerland). Meat color was assessed on the *gluteus superficialis* (GS) muscle using the three coordinates, L*, a* and b*, of the CIE-LAB color space with a Minolta CR-300 chromameter

(Konica Minolta, Tokyo, Japan) under the D65 illuminant option and an 11-mm orifice (L*GS, a*GS and b*GS). Under this system, L* indicates how light the meat color is (a lower value being associated with darker meat) while a* represents the degree of green-redness (redder meat for a higher a* value) and b* reflects the degree of blue-yellowness of the meat (yellower meat for a greater b* value).

The water holding capacity (WHC) of GS was measured as the time necessary for a piece of filter paper (about 1 cm²) to become wet when placed on the freshly cut surface of the muscle [20], a higher value being associated with a lower ability to lose water (1 point = 10 s; maximum 20 points). A meat quality index (MQI) was computed as a linear combination of pH24 SM, WHC GS and L* GS: MQI (%) = 34 + (11.04 × pH24 SM) + (0.105 × WHC GS) – (0.231 × L* GS). This equation has been defined as a predictor of technological yield (ratio of the weight of cooked ham to the weight of defatted and boneless fresh ham) during cured-cooked ham processing [21].

Genotyping and genotype quality control

Blood was sampled from the test pigs at a live weight of about 60 kg. The DNA was extracted from blood samples of 491 animals (385 piglets and their 106 sires) and genotyped using the Illumina PorcineSNP60 BeadChip (San Diego, CA, USA) containing 62 163 SNPs [5] at the Centre National de Génotypage (Evry, France). The order of the SNPs was based on the Pig Sscrofa10.2 assembly released by the International Swine Genome Sequencing Consortium [22], combined with RH mapping information [23]. Quality control was done considering genotyping of the 106 sires that were more representative of the LW population. The *check.marker* function of the GenABEL R package [24] was applied. It excluded 5390 SNPs with call rates lower than 97%, 12 077 SNPs with minor allele frequencies less than 5%, and 1051 SNPs with a *P*-value of a χ^2 test for a Hardy-Weinberg equilibrium lower than 1.10^{-5} . After applying these quality control measures, 42 272 SNPs located on autosomes and 2140 SNPs that were not located on the Pig Sscrofa 10.2 assembly were retained for association analyses, i.e. an average of 15 SNPs per Mb. After filtering, the number of SNPs per chromosome ranged from 1047 (SSC18 i.e. *Sus scrofa* chromosome 18) to 5155 (SSC1). The call rate across the retained SNPs was higher than 90% for all animals and so all animals were retained for analyses.

Statistical analyses

Adjustment of data for systematic environmental effects

Prior to GWAS, the phenotypes of the 385 LW animals were analyzed jointly with the phenotypes of their batch mates (3030 animals from five breeds) to adjust the data

for systematic environmental effects. All traits were corrected using a linear model (GLM procedure, SAS Inst., Inc., Cary, NC [25]), which included the fixed effects of breed (five levels), the combination of test year, test station and contemporary group (slaughter date for meat quality traits or fattening batch for other traits; 20 and 97 levels, respectively) and body weight of the animal at the start of the test (for traits recorded during growth) or at slaughter (for traits recorded at the abattoir), nested within breed, as a linear covariate. The residuals from these analyses were then used as trait phenotypes for GWAS.

Genome-wide association studies

The GWAS were performed using the GenABEL R package [24]. For each trait, SNP effects were tested with the FASTA (FAMily-based Score Test for Association) method [26] based on a mixed animal model (1) that included the genomic kinship matrix **G** (*ibs* procedure in GenABEL) to account for relatedness in the sampled population:

$$Y_j = \mu + b_{ij}M_i + u_j + e_{ij} \quad (1)$$

with Y_j = the phenotype corrected for systematic environmental effects; μ = the overall mean; b_{ij} = the genotype score (0, 1 or 2) of the i^{th} SNP for the j^{th} individual; M_i = the additive effect of the i^{th} SNP; u_j = the random polygenic effect of the j^{th} individual, with covariance structure $u_j \sim N(0, G\sigma_u^2)$, where **G** is the genomic kinship matrix and σ_u^2 is the polygenic variance; and e_{ij} = the random residual effect with $e_{ij} \sim N(0, I\sigma_e^2)$, where **I** is an identity matrix and σ_e^2 is the residual variance. As a first step, the variance components σ_u^2 and σ_e^2 were estimated using the genomic kinship matrix in an animal mixed model without a marker effect. These variance components were used in a second step to jointly estimate all the effects included in model (1).

To avoid inflation in the test statistic due to potential deviations from its assumptions, the consistency with χ^2 tests for the distribution of the *P*-values was controlled by regressing the observed *P*-values of each GWAS against the expected *P*-values of a χ^2 test. The *P*-values were then corrected by multiplying observed *P*-values by the regression factor λ , using the genomic control method [27]. This correction assumes that the number of SNPs with an effect on the trait is very small compared to the total number of SNPs tested.

As proposed in Teyssèdre et al. [28], three *P*-value thresholds were used to identify and describe regions of interest. The most stringent threshold was 5.10^{-6} , which corresponds to approximately 10 000 independent tests corrected with Bonferroni. A less stringent threshold of 5.10^{-5} was also applied to detect moderate associations,

as proposed by the Wellcome Trust Case Control Consortium [29]. Finally, in order to take into account effects of QTL on correlated traits, SNPs with a P -value threshold of 5.10^{-4} that were located in the vicinity of the QTL regions were also reported. A QTL region was considered by grouping SNPs with P -values lower than 5.10^{-4} in a 10 Mb interval.

Haplotype blocks and multiple regression analyses

Haplotypes transmitted by a parent to each of its offspring were inferred based on informative SNPs, using a similar procedure to that of Coop et al. [30] and described in Tortereau et al. [31]. Briefly, the haplotype reconstruction procedure is based on three steps. First, within each half-sib family, a partial haplotype phase was reconstructed for the father based on the genotypes of the offspring. This was done by first partially reconstructing the haplotype transmitted by the father based on markers that are homozygous in the offspring. Then, paternally transmitted haplotypes of all offspring were combined to reconstruct the haplotypes of the father and the haplotype transmitted to each offspring by its mother. Second, the model of Scheet and Stephens [32] was fitted to the partial haplotypes of fathers and mothers across families to increase the level of haplotype reconstruction. Finally, given the phase information in the fathers and for each offspring, segregation indicators that describe which of the paternal alleles was transmitted at each SNP, were reconstructed.

In regions that contained at least one SNP with significant effects on a trait (P -value $< 5.10^{-6}$), the LD between SNPs was calculated as r^2 , using Haploview (V4.2; [33]). LD blocks were generated for SNPs that were separated by less than 500 kb, as proposed by Gabriel et al. [34]. In regions that contained at most 10 SNPs, the haplotypes of the progeny were inferred from their genotypes and phased genotypes of their parents and additional haplotype analyses were performed using the following multiple regression mixed model for each region:

$$Y_{ijk} = \mu + \sum_{i=1}^t \beta_{ij} H_i + S_j + e_{ijk} \quad (2)$$

where Y_{ijk} = the phenotype corrected for systematic environmental effects; μ = the overall mean; β_{ij} = the haplotype score (0, 1 or 2) of the i^{th} haplotype for the j^{th} individual, with t = the number of haplotypes segregating in the population for that region; H_i = the effect of the i^{th} haplotype; S_j = the random sire effect and e_{ijk} = the random residual effect. Model (2) was tested using the Mixed procedure of the SAS software (SAS Inst., Inc., Cary, NC [25]). The overall effect of the haplotypes in a region and contrasts between two haplotype effects were tested using the CONTRAST and ESTIMATE functions, respectively.

Results

For the 19 traits analyzed, the number of records, means and standard deviations are in Table 1. Phenotypic correlations between traits corrected for systematic environmental effects are in Figure 1.

Genome-wide association studies

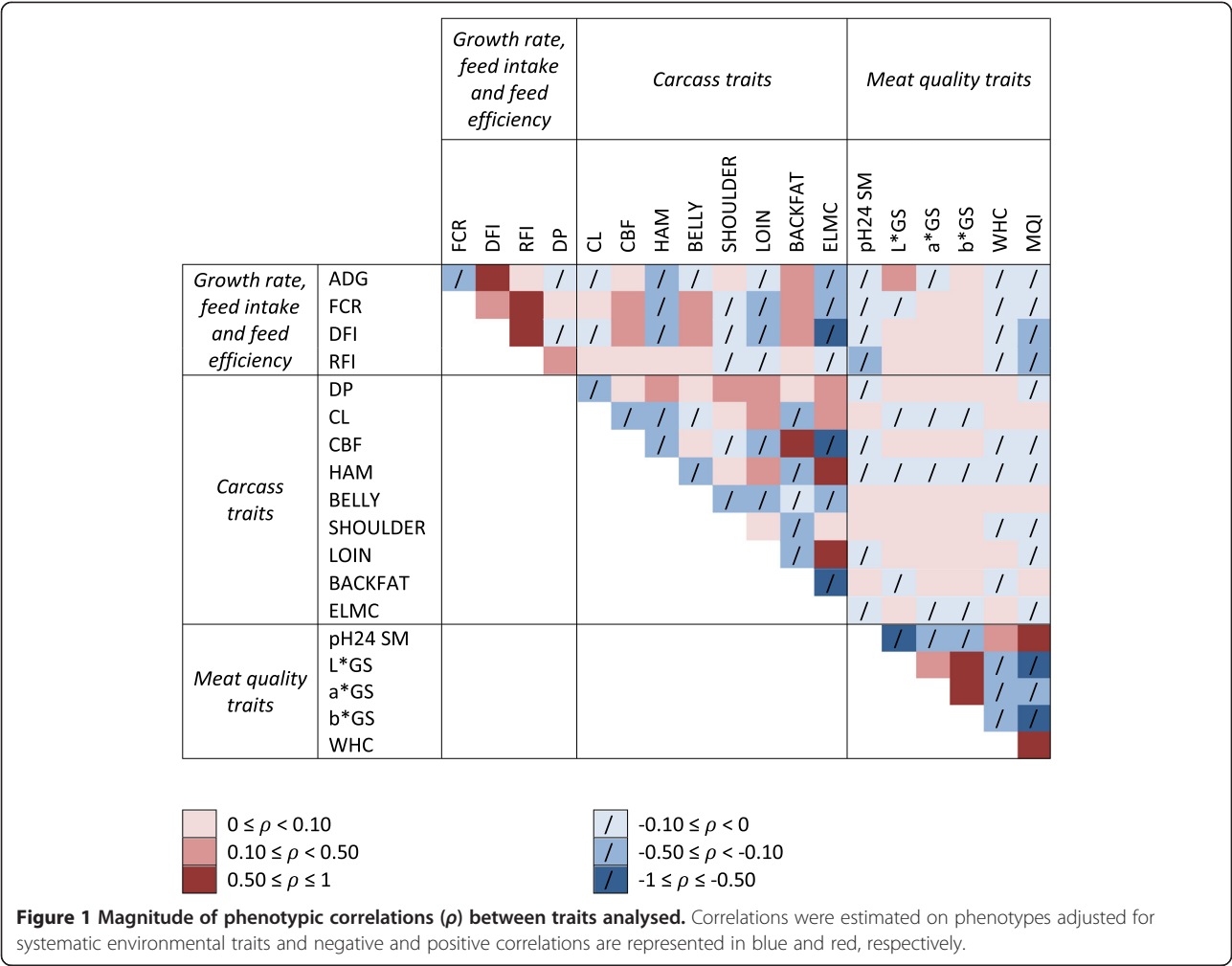
After checking that no structure was present in our population by applying multidimensional scaling available in the GenABEL R package [24], GWAS analyses were performed for the 19 traits. The average inflation factor of P -values was 1.11 ± 0.12 , with a minimum of 1 (for five of the 19 traits) and a maximum of 1.39 (for BACKFAT), indicating relatively good concordance between the observed and assumed distributions of the test statistics.

Seventeen trait $\times$ SNP tests, involving 16 distinct SNPs, were significant at the threshold of 5.10^{-6} and 52 tests, involving 48 distinct SNPs, were significant at the

Table 1 Descriptive statistics and abbreviations for the traits analyzed

Trait	Abbreviation	N	Mean	STD
Growth, feed intake and feed efficiency				
Average daily gain during the test period (kg.d ⁻¹)	ADG	385	0.974	0.089
Daily feed intake (kg.d ⁻¹)	DFI	385	2.61	0.25
Residual feed intake (kg.d ⁻¹)	RFI	385	0.00	0.13
Feed conversion ratio (kg.kg ⁻¹ BW)	FCR	385	2.68	0.21
Carcass traits				
Dressing percentage (%)	DP	385	78.7	1.4
Carcass length (mm)	CL	385	1010	29
Mean carcass backfat thickness (mm)	CBF	385	23.5	3.2
Ham weight (kg)	HAM	385	9.71	0.65
Belly weight (kg)	BELLY	385	5.14	0.53
Shoulder weight (kg)	SHOULDER	385	9.46	0.66
Loin weight (kg)	LOIN	385	10.90	0.88
Backfat weight (kg)	BACKFAT	385	3.42	0.65
Lean meat content calculated with cut weights (%)	ELMC	385	56.0	3.1
Meat quality traits				
Ultimate pH of <i>semimembranosus</i> muscle	pH24 SM	385	5.70	0.17
L* of <i>gluteus superficialis</i> muscle	L*GS	384	50.9	4.0
a* of <i>gluteus superficialis</i> muscle	a*GS	384	9.4	1.6
b* of <i>gluteus superficialis</i> muscle	b*GS	384	5.5	1.8
Water holding capacity of <i>gluteus superficialis</i> muscle (10s)	WHC	385	13.5	6.3
Meat Quality Index	MQI	384	86.9	2.8

N = number of records, STD = phenotypic standard deviation.



threshold of 5.10^{-5} . Only two of these SNPs were not located on the Pig Sscrofa10.2 draft. In total, 23 QTL regions were identified, including all autosomes except SSC2, 5 and 12 (Figure 2). At least one QTL region was identified for each trait, except ADG, DFI, SHOULDER and WHC. The magnitude of the estimated SNP effects was expressed in trait phenotypic standard deviation (STD) units, which was calculated based on phenotypes adjusted for systematic environmental effects (Table 2).

Ten QTL regions were identified at the most stringent threshold (P -value $< 5.10^{-6}$), i.e. five for carcass traits, five for meat quality traits and none for growth rate, feed intake and feed efficiency. Two of the five carcass QTL, on SSC7 and SSC17, had an effect on carcass length (from 0.28 to 0.42 STD) only. The three other carcass QTL, on SSC10, 16 and 18, affected carcass cut weight and backfat thickness (0.27 - 0.45 STD). The five QTL for meat quality traits were located on SSC1 (two QTL), SSC8, 9 and 13. Each of these QTL affected several meat quality traits, with effects ranging from 0.27 to 0.75 STD. Most of the detected QTL regions were composed

of a single SNP that was significant at the 5.10^{-6} threshold, except for the QTL region on SSC17 for CL (three SNPs), and on SSC1d (four SNP) and SSC9a (two SNP) for meat quality traits. However, for each of the 10 most significant QTL regions, other trait $\times$ SNP combinations were found at the 5.10^{-5} and 5.10^{-4} thresholds.

Based on the moderate threshold (P -value $< 5.10^{-5}$), 13 additional QTL regions were identified. Three QTL, on SSC3, SSC11 and SSC18, had effects of about 0.30 STD on FCR, and one QTL on SSC6 had effects ranging from 0.41 to 0.46 STD for RFI. Seven QTL with effects on different carcass composition traits (0.25 - 0.59 STD) were detected on SSC1, 3, 6, 9, 14, 15 and 16. For meat quality traits, two additional QTL regions (on SSC1b and SSC4) were identified, with moderate effects (0.27 - 0.38) on meat color and ultimate pH.

Relative to the number of traits analyzed, a larger number of tests with a P -value $< 5.10^{-5}$ was obtained for meat quality traits (on average 3.5 significant tests per trait) than for growth rate and feed intake (1.5 significant tests per trait) and for carcass traits (2.8 significant

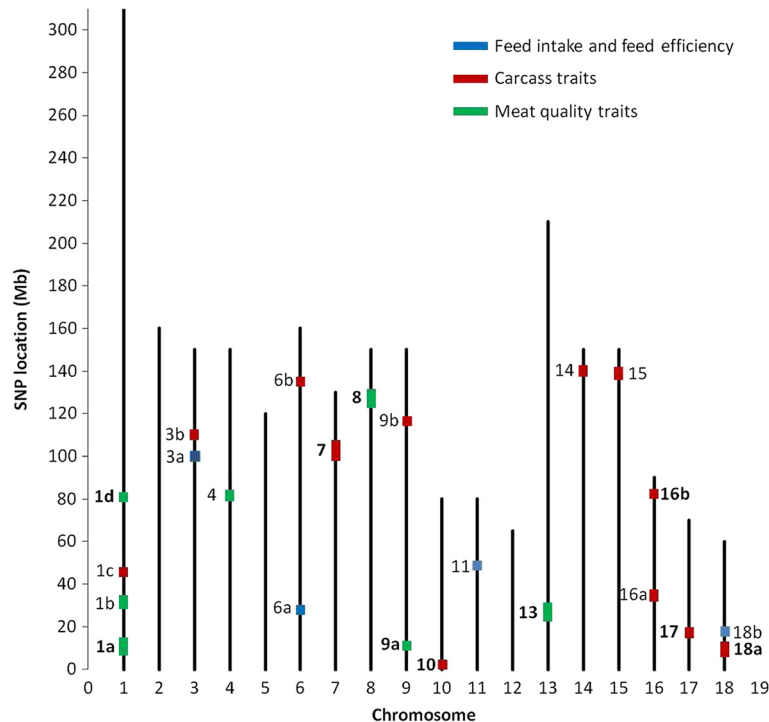


Figure 2 Distribution of SNPs with P -values $< 5.0E-05$ for each group of traits. The letters distinguish different QTL regions on the same chromosome; when at least one SNP in the region was significant at the threshold of 5.10^{-6} , names of the QTL regions are in bold (see Table 2).

tests per trait). With the criteria that were used to define QTL regions, no QTL region was shown to simultaneously affect different groups of traits (growth, feed intake and feed efficiency, carcass and meat quality traits). However, within meat quality traits for example, some peaks tended to be shared, as for pH24 SM and MQI on SSC8, or for b* GS and L* GS on SSC1 (Figure 3).

Haplotype analyses

The 10 most significant QTL regions were subjected to haplotype analyses. First, SNPs that had significant effects at the threshold of 5.10^{-5} and that were in high LD within a QTL region were grouped together in haplotype blocks according to the criteria specified by Gabriel et al. [34]. Using these criteria, a haplotype block could be identified for only one of the QTL regions, i.e. the 183 kb SSC1d region, with six SNPs that were associated with meat quality traits (Table 2). The r^2 between SNPs in this region ranged from 0.30 to 1 (Figure 4a). Three distinct haplotypes (ACTCTA, ACTTTC and GTCTCC, referred to as Haplo1, Haplo2 and Haplo3 hereinafter) were identified in the SSC1d region with frequencies of 57%, 24% and 19%, respectively (Figure 4b). Their effects on the 19 traits were evaluated for the 376 phased progeny in order to evaluate potential pleiotropic effects of the region. Parental phases were unavailable for nine animals, which were discarded from the haplotype

analysis. The haplotypes had significant ($0.0001 < P$ -value < 0.01) effects on all six meat quality traits analyzed (Table 3). The highest significance levels were obtained for b*GS and L*GS and for the meat quality index (P -value $< 1.10^{-4}$), i.e. the traits that displayed significant results in the GWAS (Table 2). For these three traits and for pH24 SM, Haplo1 and Haplo3 showed significant and opposite effects. Haplo1 was associated with favorable effects (e.g. for MQI: +0.5 point for Haplo1 vs -0.6 for Haplo3). Haplo2 had effects that were similar to Haplo1 for b*GS and L*GS, but had no significant effect on MQI and pH24 SM. For the two other traits, Haplo1 was the only haplotype that had a significant effect on a*GS, while Haplo1 and Haplo2 had, respectively, favorable (+8 s) and unfavorable (-8 s) effects on WHC. As a consequence, Haplo1 had a consistently favorable effect on all meat quality traits recorded, and Haplo3 displayed an almost systematically opposite effect on these traits. The effects of Haplo2 were either intermediate or not significantly different from Haplo1.

Discussion

The PorcineSNP60 BeadChip has become available quite recently [5]. A GWAS method testing individual SNP effects was chosen because previous studies had demonstrated that single-marker tests produced similar or even greater power than haplotype-based approaches [35,36]. Moreover, testing individual SNP effects does not depend

Table 2 QTL regions with at least one SNP with a P -value less than 5.10^{-5}

	QTL ¹	Thresholds ²			Positions of flanking markers (kb)	Traits ³	Effects in STD ⁴ (min-max)
		5.10 ⁻⁴	5.10 ⁻⁵	5.10 ⁻⁶			
<i>Growth rate, feed intake, feed efficiency</i>	3a	2	1	0	100 098–100 119	FCR	0.29 – 0.33
	6a	4	3	0	27 650–28 138	RFI	0.41 – 0.46
	11	1	1	0	48 555	FCR	0.36
	18b	4	1	0	17 724–17 817	FCR	0.29 – 0.33
<i>Carcass traits</i>	1c	4	4	0	45 536–45 998	BELLY	0.57 – 0.59
	3b	5	3	0	109 719–110 511	BELLY	0.43 – 0.55
	6b	7	1	0	134 691–135 078	BACKFAT- ELMC- HAM	0.25 – 0.32
	7	10	2	1	100 145–105 315	CL	0.28 – 0.40
	9b	2	1	0	116 328–116 390	LOIN	0.33 – 0.42
	10	3	1	1	2 015 – 2 080	BELLY	0.28 – 0.35
	14	5	1	0	139 323–140 810	BACKFAT- ELMC - CBF - CL	0.29 – 0.39
	15	3	1	0	137 725–139 857	DP	0.30 – 0.52
	16a	6	2	0	34 003–35 190	HAM	0.30 – 0.39
	16b	17	5	1	82 092–82 664	BACKFAT- ELMC	0.30 – 0.45
	17	4	3	3	16 788–17 549	CL	0.28 – 0.42
	18a	9	1	1	7 813 – 11 122	HAM- ELMC - CBF - BELLY	0.27 – 0.40
<i>Meat quality traits</i>	1a	14	3	1	8 715–12 869	a*GS - b*GS - L*GS	0.27 – 0.35
	1b	7	2	0	30 337–32 684	pH24 SM - b*GS - MQI	0.28 – 0.32
	1d	16	8	4	80 701–80 884	b*GS - L*GS - MQI	0.30 – 0.49
	4	5	2	0	80 963–82 252	a*GS	0.27 – 0.38
	8	6	2	2	124 786–129 300	pH24 SM - b*GS - MQI	0.29 – 0.45
	9a	7	3	2	11 043–11 445	L*GS - MQI	0.27 – 0.37
	13	3	1	1	24 391–29 002	L*GS - MQI	0.29 – 0.75

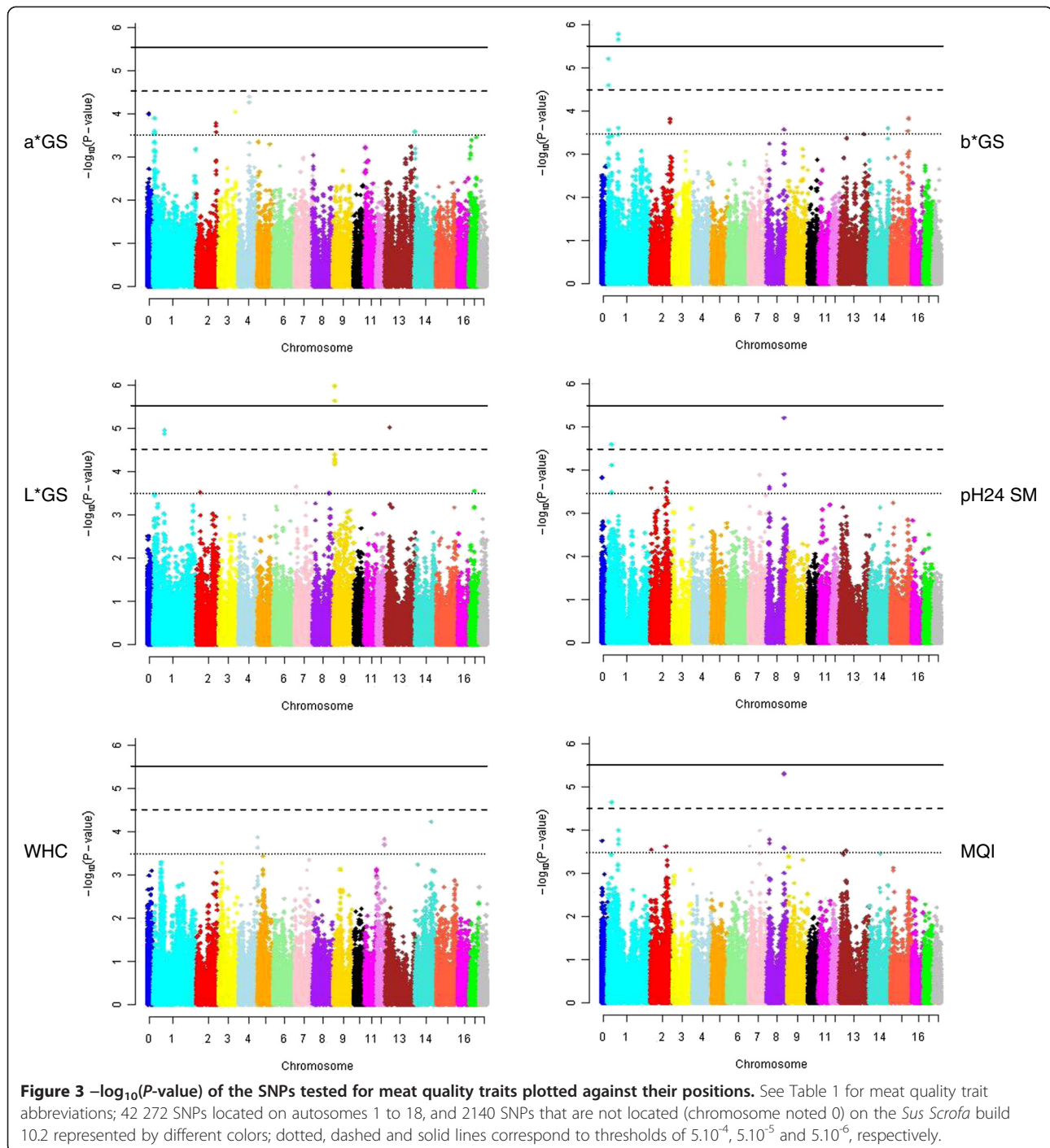
Lines in bold are regions where at least one SNP had a P -value $< 5.10^{-6}$; ¹QTL region as defined by the chromosome number and location (letters are ordered by positions on chromosomes, see Figure 2); ²For each QTL region, the number of SNP $\times$ trait combinations with a P -value lower than the corresponding threshold; ³see Table 1 for trait abbreviations; ⁴STD = phenotypic standard deviation estimated on phenotypes adjusted for systematic environmental effects.

on SNP positions and haplotype reconstruction. None of the detected QTL regions displayed significant effects on more than one group of traits, despite the presence of significant genetic correlations [17], such as between growth rate and body composition. During preliminary simulation studies, the relatively limited size of the data set (about three half-sib progeny from about 100 sires) was shown to enable detection of only relatively large QTL (with effects greater than 0.5 STD) for traits with a heritability of 0.5, and power was less than 30% when the effect of the QTL or trait heritability was reduced [37]. With such limited power, it is therefore not surprising that only large QTL were detected, and that no region was identified to affect multiple traits with moderate genetic correlations. Nevertheless, a relatively large number of significant QTL was found for meat quality traits, although these traits are generally less heritable than growth, feed intake or carcass traits [17].

LD blocks were used to identify haplotypes associated with phenotypes in our study. Only one region could be

dissected based on this approach, since the other regions displayed no LD blocks for the SNPs with significant effects. This small number of LD blocks might be due to the distance between SNPs with significant effects or to local inaccuracies in the published sequence or to the presence of limited LD between adjacent SNPs in our population. However, in QTL regions for which LD blocks were identified, it was possible to identify haplotypes that significantly affected the traits.

Alignment of the genetic and physical maps on the most recent porcine genome sequence assembly (Sscrofa10.2) in PigQTLdb [1] allowed our results to be compared with previously reported QTL locations. Ten of the QTL regions found in our study were consistent with QTL locations previously detected by linkage analyses. However, most of the QTL regions detected in our study (13 of the 23 QTL regions) were not previously described. In particular, none of the regions detected coincided with the QTL regions reported by Tribout et al. [38], who used a grand-daughter design with microsatellite markers in the

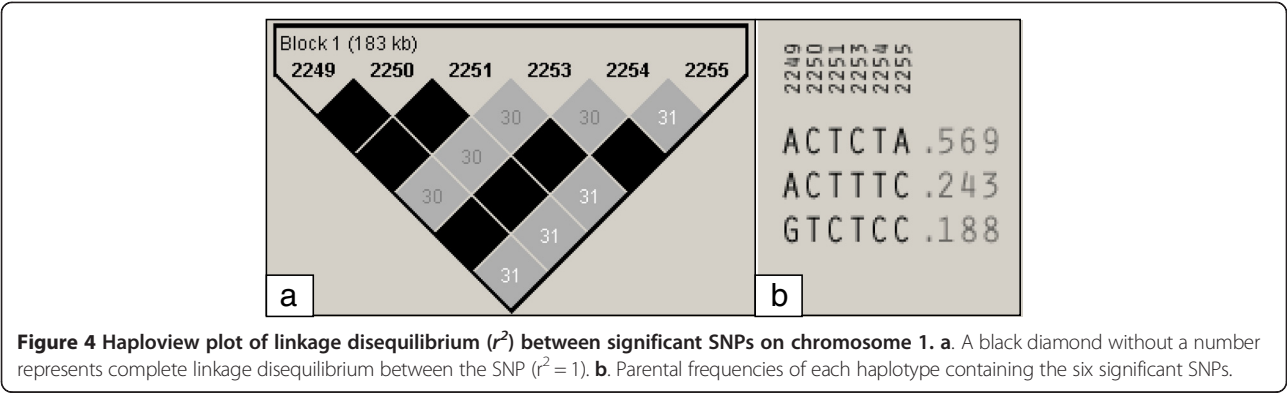


same population. The discrepancies between population structures, methods of analysis, and density and informativity of markers could explain the differences between the results of Tribout et al. [38] and our study.

QTL for growth rate, feed intake and feed efficiency

Four QTL regions had a significant effect ($P\text{-value} < 5.10^{-5}$) on RFI (SSC6a) or FCR (SSC3a, 11 and 18b). Markers

flanking the SSC6a QTL region are located in the *FTO* gene (*fat mass and obesity associated*), where a polymorphism has previously found to be associated with RFI in Yorkshire pigs [39], but which was subsequently not reported as significantly associated with RFI in a GWAS in that same population [13]). Among the three QTL that affected FCR, two (SSC3a and SSC18b) were not previously reported in the literature. These two QTL regions



were small (about 21 and 92 kb respectively) and they had an effect of about 1/3 STD on FCR. In the vicinity of the SSC11 region, a QTL for FCR was previously described in a F2 Meishan × Large White population [40]. Another recent GWAS for FCR in a Duroc population [12] identified no common QTL regions with our LW pigs.

QTL for carcass traits

Most earlier QTL linkage analyses included carcass traits and a very large number of QTL have been reported for these traits. Nevertheless, to our knowledge, eight of the 12 QTL regions found for carcass traits in our study were not previously described, *i.e.* on SSC1c (at 46 Mb), SSC3b (at 110 Mb), SSC6b (at 135 Mb), SSC9b (at 116 Mb), SSC10 (at 2 Mb), SSC14 (at 139 Mb), SSC15 (at 138 Mb) and SSC16a (at 34 Mb). Belly weight and carcass length displayed the largest number of significant associations (four distinct QTL regions for belly weight and three for carcass length). This might be because these two traits are not included in the French LW breeding objectives, so that QTL that affect these traits are less likely to have been fixed by selection than QTL for traits included in selection. In a purebred commercial population, our study confirmed the presence of four QTL that were previously detected in crossbred populations. Of these, a QTL on SSC7b that influences

carcass length was previously described in several crossbred populations [41-43]. Liu et al. [44] also reported a QTL with an effect on backfat thickness in the SSC16b region. The most significant association found in our study (P -value = 7.10^{-7}), *i.e.* the QTL on SSC17 (at 17 Mb) for carcass length, is located in a region where a QTL was previously described by Karlskov-Mortensen et al. [45] in crossbred Hampshire and Landrace pigs. Finally, in the SSC18 QTL region (at 11 Mb), a QTL was previously described in an F2 Berkshire × Yorkshire population [46].

These results show that several QTL that affect carcass traits with moderate to strong effects (from 0.3 to 0.6 STD) are still segregating in this LW population, despite more than 30 generations of selection for these traits. However, during the past two decades, this population has also been selected for reproductive traits, so that part of the selection pressure is applied to maternal abilities and prolificacy. This may explain why some QTL with relatively strong effects are still segregating in this population, either because selection pressure has not been sufficient to fix them, or because they exert antagonistic effects on production or reproduction traits. As a consequence, although the effects of our QTL must be confirmed in a larger population, estimates of their effects on reproductive traits are also required, in order to decipher how they can actually be

Table 3 Haplotype effects of the SSC1d QTL for meat quality traits (N = 376 pigs)

Trait ¹	STD	P-value ²	Haplo1 ACTCTA		Haplo2 ACTTTC		Haplo3 GTCTCC	
			β ³	P-value ⁴	β ³	P-value ⁴	β ³	P-value ⁴
pH24 SM	0.15	0.0007	0.16 ^a	0.0010	0.03 ^a	0.6438	-0.19 ^b	0.0021
L*GS	3.49	< 0.0001	-0.16 ^a	0.0008	-0.14 ^a	0.0282	0.30 ^b	< 0.0001
a*GS	1.54	0.0252	-0.13 ^a	0.0075	0.04 ^{ab}	0.4994	0.09 ^b	0.1508
b*GS	1.43	< 0.0001	-0.18 ^a	0.0002	-0.13 ^a	0.0505	0.31 ^b	< 0.0001
WHC (s)	59.6	0.0113	0.14 ^a	0.0042	-0.13 ^b	0.0472	-0.02 ^{ab}	0.80
MQI (%)	2.45	< 0.0001	0.20 ^a	< 0.0001	0.03 ^a	0.5838	-0.23 ^b	0.0002

¹See Table 1 for trait abbreviations; ²P-value of the haplotype effect in a linear mixed model including a mean, a random sire effect and multiple regression on the haplotypes; ³estimates of regression coefficients (β) in phenotypic standard deviation (STD) units of the traits estimated using phenotypes adjusted for systematic environmental traits; for a given trait, values with different superscripts (^a or ^b) were significantly different ($P < 0.05$); ⁴P-values for the test of $\beta \neq 0$.

used in the context of future marker-assisted selection strategies.

QTL for meat quality traits

Three of the seven QTL regions detected for meat quality traits were not previously described in the literature: the SSC1a region (at about 8–13 Mb), the SSC9a region (at about 11 Mb) and the SSC13 region (24–29 Mb). The first two QTL regions have moderate effects on meat color traits (about 0.33 STD), while the SSC13 QTL has an effect of 0.75 STD on meat lightness. Three QTL for meat quality traits were previously reported for the SSC1b, SSC4 and SSC8 regions by Ponsuksilii et al. [47] and van Wijk et al. [48] in crossbred populations, while the SSC1d QTL region has been shown to influence meat quality traits in Landrace pigs [49].

In the SSC1d region, a cluster of six SNPs was identified, and the three corresponding haplotypes had significant effects on all meat quality traits analyzed in our study, but not on other production traits. In this region of 183 kb, no functional candidate gene based on the present draft of the pig sequence could be identified. The 106 half-sib families in our study are representative of this LW population at the time of sampling, so that the estimated haplotype frequencies are expected to be close to those in the whole population. Assuming random mating, with the frequency of the favorable haplotype estimated at 57%, only 32% of the animals are expected to carry two copies of the most favorable haplotype. Thus, 68% of the animals are carriers of at least one unfavorable haplotype and almost 4% of the pigs carry two copies of the most unfavorable haplotype. In addition, although meat quality traits had only moderate phenotypic correlations with growth or intake traits in our study, meat quality traits are known to have antagonistic relationships with feed efficiency traits [50]. Such antagonistic effects were, however, not found for these haplotypes or for any of the significant SNPs for meat quality and either FCR or RFI. The lack of adverse influences of the favorable haplotype on correlated production traits and on traits that were not evaluated in this study, such as reproduction traits, will therefore require specific tests and validation before the haplotype is used in selection. Finally, a survey of haplotypes that segregate in other commercial populations (Landrace, Piétrain, Duroc, etc.) and estimation of their effects on meat quality traits, might be necessary to identify the underlying causal polymorphisms.

Conclusions

This study in a major French commercial pig population confirmed the segregation of several QTL affecting production and meat quality traits. Some of these QTL had not been reported before, while others were detected in crossbred populations using microsatellite markers. These

findings demonstrate that using relatively dense SNP arrays within a purebred population makes it possible to detect QTL regions that were not detected by linkage analyses. Given the number of traits analyzed, the largest number of significant associations was obtained for meat quality traits. As selection has until recently tended to focus on growth or carcass traits rather than meat quality, genes with moderate to strong effects are more likely to be still segregating for these traits. However, our results also show that some QTL with moderate to strong effects on feed efficiency and carcass traits continue to segregate in this LW population. Moreover, the QTL detected did not affect multiple types of traits, which suggests that SNP could be used to improve growth, feed intake, feed efficiency and carcass traits without degrading meat quality traits and, reciprocally, to improve meat quality traits without affecting other production traits. This needs further validation to overcome the relatively limited power of our design.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

MPS performed GWAS and haplotype analyses and drafted the manuscript. TT supervised the performance testing and performed the statistical correction of phenotypes. NI was responsible for blood sampling, DNA extractions and the coordination of genotyping. MB and NM were responsible for performance testing in the experimental unit. BS carried out the parental phase reconstruction. PD was responsible for the genotyping database. MdPS and AT performed the preliminary simulations and GWAS analyses. MJM participated in conducting the experiment, as representative of BIOPORC breeding organizations. HG and JPB supervised the overall analysis and helped to draft the manuscript. DM and CRG had scientific responsibility for the Delisus and Immopig projects, respectively. All authors read and approved the final manuscript.

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Author details

¹INRA, UMR1313 Génétique Animale et Biologie Intégrative, F-78350 Jouy-en-Josas, France. ²INRA, AgroParisTech, UMR1313 Génétique Animale et Biologie Intégrative, F-78350 Jouy-en-Josas, France. ³INRA, UMR444 Laboratoire de Génétique Cellulaire, F-31326 Castanet-Tolosan, France. ⁴INRA, UE450 Testage Porcs, F-35651 Le Rheu, France. ⁵IFIP, Pôle génétique, F-35651 Le Rheu, France.

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