



Phenolic indeno[1,2-b]indoles as ABCG2-selective potent and non-toxic inhibitors stimulating basal ATPase activity.

Gustavo Jabor Gozzi, Zouhair Bouaziz, Evelyn Winter, Nathalia Daflon-Yunes, Mylène Honorat, Nathalie Guragossian, Christelle Marminon, Glaucio Valdameri, Andre Bollacke, Jean Guillon, et al.

► To cite this version:

Gustavo Jabor Gozzi, Zouhair Bouaziz, Evelyn Winter, Nathalia Daflon-Yunes, Mylène Honorat, et al.. Phenolic indeno[1,2-b]indoles as ABCG2-selective potent and non-toxic inhibitors stimulating basal ATPase activity.. Drug Design, Development and Therapy, 2015, 9, pp.3481-3495. 10.2147/DDDT.S84982 . hal-01179426

HAL Id: hal-01179426

<https://hal.science/hal-01179426>

Submitted on 27 Nov 2023

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Phenolic indeno[1,2-*b*]indoles as ABCG2-selective potent and non-toxic inhibitors stimulating basal ATPase activity

Gustavo Jabor Gozzi^{1,2}Zouhair Bouaziz³Evelyn Winter^{1,4}Nathalia Daflon-Yunes¹Mylène Honorat¹Nathalie Guragossian³Christelle Marminon³Glaucio Valdameri^{1,2}Andre Bollacke⁵Jean Guillon⁶Noël Pinaud⁷Mathieu Marchivie⁸Silvia M Cadena²Joachim Jose⁵Marc Le Borgne³Attilio Di Pietro¹

¹Equipe Labellisée Ligue 2014, BMSSI UMR5086 CNRS/Lyon I University, IBCP, Lyon, France; ²Department of Biochemistry and Molecular Biology, Federal University of Paraná, Curitiba, Paraná, Brazil; ³Faculty of Pharmacy – ISPB, EA 4446 Biomolecules, Cancer and Chemoresistance, Health SFR of East Lyon CNRS UMS3453 - INSERM U57, University of Lyon, Lyon I University, Lyon Cedex 8, France; ⁴Department of Pharmaceutical Sciences, PGFAR, Federal University of Santa Catarina, Florianopolis, Santa Catarina, Brazil; ⁵Institute of Pharmaceutical and Medicinal Chemistry, University of Münster, Münster, Germany; ⁶ARNA Laboratory, Pharmaceutical Sciences UFR, INSERM U869, University of Bordeaux, Bordeaux Cedex, France; ⁷ISM – CNRS UMR 5255, University of Bordeaux Cedex, France; ⁸ICMCB CNRS-UPR 9048, University of Bordeaux, Pessac Cedex, France

Correspondence: Attilio Di Pietro
Equipe Labellisée Ligue 2014, BMSSI UMR5086
CNRS/Lyon I University, IBCP, Passage du
Vercors 7, 69367 Lyon Cedex 07, France
Tel +33 4 72 72 26 29
Fax +33 4 72 72 26 04
Email a.dipietro@ibcp.fr

Abstract: Ketonic indeno[1,2-*b*]indole-9,10-dione derivatives, initially designed as human casein kinase II (CK2) inhibitors, were recently shown to be converted into efficient inhibitors of drug efflux by the breast cancer resistance protein ABCG2 upon suited substitutions including a *N*-phenethyl on C-ring and hydrophobic groups on D-ring. A series of ten phenolic and seven *p*-quinonic derivatives were synthesized and screened for inhibition of both CK2 and ABCG2 activities. The best phenolic inhibitors were about threefold more potent against ABCG2 than the corresponding ketonic derivatives, and showed low cytotoxicity. They were selective for ABCG2 over both P-glycoprotein and MRP1 (multidrug resistance protein 1), whereas the ketonic derivatives also interacted with MRP1, and they additionally displayed a lower interaction with CK2. Quite interestingly, they strongly stimulated ABCG2 ATPase activity, in contrast to ketonic derivatives, suggesting distinct binding sites. In contrast, the *p*-quinonic indenoindoles were cytotoxic and poor ABCG2 inhibitors, whereas a partial inhibition recovery could be reached upon hydrophobic substitutions on D-ring, similarly to the ketonic derivatives.

Keywords: multidrug resistance, cancer cells, ABCG2/BCRP, indenoindole inhibitors, structure–activity relationships, ATPase activity

Introduction

A major obstacle of tumor treatment by chemotherapy is cancer cell multidrug resistance, which may be caused by several factors including overexpression of multidrug ATP-binding cassette (ABC) transporters. These transmembrane proteins work as efflux pumps reducing the intracellular concentration of drugs.¹ Among the 48 human genes encoding ABC transporters, only three are recognized to be associated with low prognostic in cancer patients: ABCB1/Pgp (P-glycoprotein), ABCC1/MRP1 (multidrug resistance protein 1) and ABCG2/BCRP (breast cancer resistance protein).² Pgp was the first multidrug ABC transporter to be discovered, and has been extensively studied,³ while MRP1 was later associated with multidrug resistance,⁴ and ABCG2 was more recently identified.^{5–7}

One of the strategies aimed at eliminating resistant tumors is to use inhibitors of the multidrug ABC transporters. Combination of inhibitors with anticancer drugs should increase the intracellular accumulation of drugs and their availability to cellular targets. A number of Pgp inhibitors have been optimized in vitro, up to third-/fourth-generation compounds, but their intrinsic toxicity and low in vivo activity prevented the achievement of clinical trials.^{8,9}

ABCG2 is a “half-transporter” of 655 amino acids, containing one cytosolic nucleotide-binding domain and one transmembrane domain with six α -helical spans,

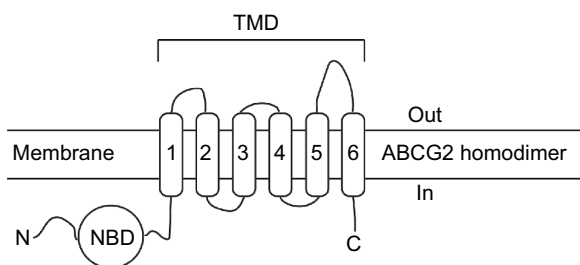


Figure 1 Structural arrangement of ABCG2.

Abbreviations: NBD, nucleotide-binding domain; TMD, transmembrane domain.

which needs to at least dimerize to be functional (Figure 1). This transporter is present in various membrane barriers protecting sensitive organs, as well as in many types of cancer cells.^{10,11} Selective inhibitors are interesting for two main reasons: 1) *in vitro* to study the specific role of the transporter and 2) *in vivo*, hopefully up to clinical trials, to use very low concentrations to inhibit each targeted transporter because most potent Pgp inhibitors, such as elacridar or tariquidar, also inhibited ABCG2 but at much higher concentrations expected to induce cytotoxicity and related side effects. The first specific ABCG2 inhibitor, of natural origin, was fumitremorgin C that unfortunately displayed serious neurotoxicity.¹² Synthetic derivatives were developed, resulting in the highly potent Ko143 inhibitor, which however still retained significant residual toxicity.¹³ Screening of different classes of flavonoids identified interesting inhibitors such as hydrophobic flavones, acridones, chromones, asymmetric chalcones, and symmetric *bis*-chalcones,¹⁴ some of them being active *in vivo* in mouse models.^{15,16}

A different type of ABCG2-selective inhibitors was recently developed as a series of ketonic indenoindoles, upon appropriate substitutions of potent inhibitors of casein kinase II (CK2), such as the replacement of isopropyl by phenethyl at *N*⁵ position of C-ring, and the addition of hydrophobic substituents on D-ring.¹⁷ The present work was aimed at further modifying the D-ring, by replacing the ketone by either a phenol or a *p*-quinone, and reinvestigating the effects of substituents (Figure 2). The results showed that phenolic

indenoindoles, independently of hydrophobic substituents, were better inhibitors of ABCG2-mediated drug efflux than the previously described ketonic derivatives, through higher potency and selectivity, and the ability to strongly stimulate ATPase activity. In contrast, the *p*-quinonic derivatives displayed reduced inhibition capacity and significant cytotoxicity.

Materials and methods

Chemistry

Melting points were determined on an Electrothermal 9200 capillary apparatus. The infrared (IR) spectra were recorded on a Perkin Elmer Spectrum Two IR Spectrometer. The ¹H and ¹³C nuclear magnetic resonance (NMR) spectra were recorded at 400 MHz on a Bruker DRX 400 spectrometer. Chemical shifts are expressed in ppm (δ) downfield from internal tetramethylsilane and coupling constants *J* are reported in hertz (Hz). The following abbreviations are used: s: singlet; bs: broad singlet; d: doublet; t: triplet; dd: doubled doublet; dt: doubled triplet; q: quartet; m: multiplet; Cquat: quaternary carbons. The mass spectra were performed by direct ionization (electron ionization or chemical ionization) on a ThermoFinnigan MAT 95 XL apparatus. Chromatographic separations were performed on silica gel columns by column chromatography (Kieselgel 300–400 mesh). All reactions were monitored by thin-layer chromatography on GF254 plates that were visualized under a UV lamp (254 nm). Evaporation of solvent was performed in vacuum with rotating evaporator. The purity of the final compounds (>95%) was determined by ultra high performance liquid chromatography/mass spectrometry on an Agilent 1290 system using a Agilent 1290 Infinity ZORBAX Eclipse Plus C18 column (2.1×50 mm, 1.8 μm particle size) with a gradient mobile phase of H₂O/CH₃CN (90:10, v/v) with 0.1% of formic acid to H₂O/CH₃CN (10:90, v/v) with 0.1% of formic acid at a flow rate of 0.5 mL/min, with UV monitoring at the wavelength of 254 nm with a runtime of 10 minutes.

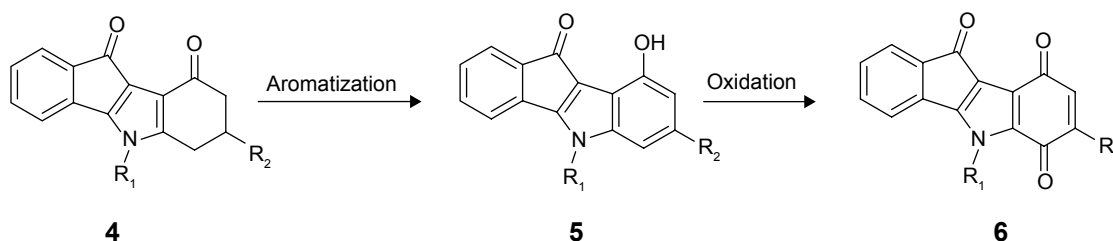


Figure 2 Preparation of the target compounds 5 and 6 from ketones 4.

General procedure for the synthesis of compounds 5

To a solution of compound **4** (2.4 mmol) in Ph₂O (15 mL) was added 0.48 g of 10% Pd-C. Then, the mixture was heated to reflux for 6 hours. After cooling, 25 mL of MeOH was added and the solution filtered on celite. Evaporation of the solvent left a residue which was purified by silica gel column chromatography using ethyl acetate (EtOAc)/cyclohexane (1:2, v/v) as the eluent.

Compound 5c: 9-Hydroxy-5-(2-phenylethyl)-5H-indeno[1,2-b]indole-10-one

Red solid. Yield 53%. mp 144°C. IR (ν cm⁻¹): 3,232, 1,660, 1,602, 1,581. ¹H NMR (CDCl₃, 400 MHz) δ: 7.31 (m, 1H, Harom), 7.25–7.12 (m, 3H, Harom), 7.10–7.03 (m, 5H, Harom), 6.75–6.72 (m, 2H, Harom), 6.67 (dd, 1H, *J*₁=0.4 Hz, *J*₂=7.8 Hz, Harom), 6.45 (bs, 1H, OH), 4.36 (t, 2H, *J*=7.1 Hz, CH₂N), 3.14 (t, 2H, *J*=7.1 Hz, CH₂Ph). ¹³C NMR + DEPT (CDCl₃, 100 MHz) δ: 186.20 (Cquat), 157.04 (Cquat), 150.29 (Cquat), 143.65 (Cquat), 140.55 (Cquat), 137.40 (Cquat), 135.91 (Cquat), 132.41 (CH), 129.62 (CH), 129.19 (2CH), 129.10 (2CH), 127.55 (CH), 125.72 (CH), 123.56 (CH), 118.72 (CH), 115.14 (Cquat), 113.48 (Cquat), 108.33 (CH), 103.41 (CH), 47.92 (CH₂), 36.71 (CH₂). HRMS calcd for C₂₃H₁₇NNaO₂ [M + Na]⁺ 362.1151; found, 362.1140.

Compound 5d: 9-Hydroxy-5-(3-phenylpropyl)-5H-indeno[1,2-b]indole-10-one

Red solid. Yield 64%. mp 146°C. IR (ν cm⁻¹): 3,441, 1,665, 1,603. ¹H NMR (DMSO-*d*₆, 400 MHz) δ: 9.53 (bs, 1H, OH), 7.34–7.23 (m, 8H, Harom), 7.10–6.96 (m, 3H, Harom), 6.62 (m, 1H, Harom), 4.39 (t, 2H, *J*=7.3 Hz, NCH₂CH₂CH₂Ph), 2.74 (t, 2H, *J*=7.5 Hz, NCH₂CH₂CH₂Ph), 2.12 (m, 2H, NCH₂CH₂CH₂Ph). ¹³C NMR + DEPT (DMSO-*d*₆, 100 MHz) δ: 183.62 (Cquat), 157.76 (Cquat), 152.13 (Cquat), 144.96 (Cquat), 141.82 (Cquat), 141.17 (Cquat), 134.98 (Cquat), 133.25 (CH), 130.36 (CH), 129.36 (2CH), 129.31 (2CH), 126.98 (CH), 125.34 (CH), 123.31 (CH), 119.76 (CH), 114.35 (Cquat), 113.59 (Cquat), 109.01 (CH), 103.67 (CH), 45.28 (CH₂), 32.98 (CH₂), 32.61 (CH₂). HRMS calcd for C₂₄H₁₉NNaO₂ [M + Na]⁺ 376.1308; found, 376.1299.

Compound 5f: 9-Hydroxy-7-methyl-5-(2-phenylethyl)-5H-indeno[1,2-b]indole-10-one

Dark red solid. Yield 44%. mp 154°C. IR (ν cm⁻¹): 3,433, 1,667, 1,646, 1,602. ¹H NMR (DMSO-*d*₆, 400 MHz) δ: 9.41 (s, 1H, OH), 7.27–7.12 (m, 9H, Harom), 6.82 (s, 1H, H-6 or H-8), 6.45 (s, 1H, H-6 or H-8), 4.57 (t, 2H, *J*=7.0 Hz, CH₂N), 3.11 (t, 2H, *J*=7.0 Hz, CH₂Ph), 2.54 (s, 3H,

CH₃-7). ¹³C NMR + DEPT (DMSO-*d*₆, 100 MHz) δ: 183.70 (Cquat), 157.84 (Cquat), 151.60 (Cquat), 144.88 (Cquat), 140.92 (Cquat), 138.58 (Cquat), 135.11 (Cquat), 134.98 (Cquat), 132.98 (CH), 129.97 (2CH), 129.90 (CH), 129.24 (2CH), 127.52 (CH), 122.91 (CH), 119.45 (CH), 114.17 (Cquat), 111.51 (Cquat), 110.56 (CH), 103.97 (CH), 47.30 (CH₂), 36.45 (CH₂), 22.48 (CH₃). HRMS calcd for C₂₄H₁₉N-NaO₂ [M + Na]⁺ 376.1308; found, 376.1295.

Compound 5g: 9-Hydroxy-7-phenyl-5-(2-phenylethyl)-5H-indeno[1,2-b]indole-10-one

Dark red solid. Yield 55%. mp 168°C. IR (ν cm⁻¹): 3,428, 1,665, 1,633, 1,600. ¹H NMR (CDCl₃, 400 MHz) δ: 7.53 (d, 2H, *J*=7.3 Hz, Harom), 7.44 (t, 2H, *J*=7.3 Hz, Harom), 7.38–7.03 (m, 9H, Harom), 6.91 (s, 1H, H-6 or H-8), 6.82 (s, 1H, H-6 or H-8), 6.72 (d, 1H, *J*=6.5 Hz, Harom), 6.42 (bs, 1H, OH), 4.36 (t, 2H, *J*=7.1 Hz, CH₂N), 3.14 (t, 2H, *J*=7.1 Hz, CH₂Ph). ¹³C NMR + DEPT (CDCl₃, 100 MHz) δ: 186.11 (Cquat), 157.42 (Cquat), 150.08 (Cquat), 144.05 (Cquat), 141.75 (Cquat), 140.47 (Cquat), 139.64 (Cquat), 137.41 (Cquat), 135.82 (Cquat), 132.44 (CH), 129.59 (CH), 129.17 (2CH), 129.12 (2CH), 129.06 (2CH), 127.55 (4CH), 123.50 (CH), 118.73 (CH), 114.91 (Cquat), 112.58 (Cquat), 108.09 (CH), 102.29 (CH), 47.81 (CH₂), 36.74 (CH₂). HRMS calcd for C₂₉H₂₂NO₂ [M + H]⁺ 416.1645; found, 416.1635.

Compound 5h: 9-Hydroxy-5-[2-(2-methoxyphenyl)ethyl]-5H-indeno[1,2-b]indole-10-one

Red solid. Yield 51%. mp 166°C. IR (ν cm⁻¹): 3,408, 1,666, 1,642, 1,603. ¹H NMR (CDCl₃, 400 MHz) δ: 7.34 (dd, 1H, *J*₁=1.0 Hz, *J*₂=7.0 Hz, Harom), 7.21–6.99 (m, 6H, Harom), 6.85–6.80 (m, 3H, Harom), 6.67 (d, 1H, *J*=7.8 Hz, Harom), 6.45 (bs, 1H, OH), 4.37 (t, 2H, *J*=7.6 Hz, CH₂N), 3.86 (s, 3H, OCH₃), 3.16 (t, 2H, *J*=7.6 Hz, CH₂Ph). ¹³C NMR + DEPT (CDCl₃, 100 MHz) δ: 186.19 (Cquat), 157.83 (Cquat), 157.16 (Cquat), 150.24 (Cquat), 143.95 (Cquat), 140.82 (Cquat), 136.09 (Cquat), 132.39 (CH), 131.04 (CH), 129.65 (CH), 129.03 (CH), 125.60 (CH), 125.53 (Cquat), 123.55 (CH), 121.20 (CH), 118.96 (CH), 115.01 (Cquat), 113.49 (Cquat), 110.67 (CH), 108.22 (CH), 103.51 (CH), 55.57 (CH₃), 46.12 (CH₂), 32.12 (CH₂). HRMS calcd for C₂₄H₁₉NNaO₃ [M + Na]⁺ 392.1257; found, 392.1248.

Compound 5i: 9-Hydroxy-5-[2-(3-methoxyphenyl)ethyl]-5H-indeno[1,2-b]indole-10-one

Red solid. Yield 50%. mp 138°C. IR (ν cm⁻¹): 3,461, 1,671, 1,604, 1,582. ¹H NMR (CDCl₃, 400 MHz) δ: 7.32 (d, 1H, *J*=6.2 Hz, Harom), 7.16–7.04 (m, 4H, Harom), 6.75–6.65

(m, 5H, Harom), 6.55 (bs, 1H, OH), 6.45 (s, 1H, Harom), 4.36 (t, 2H, $J=7.1$ Hz, CH_2N), 3.67 (s, 3H, OCH_3), 3.10 (t, 2H, $J=7.1$ Hz, CH_2Ph). ^{13}C NMR + DEPT (CDCl_3 , 100 MHz) δ : 186.18 (Cquat), 160.26 (Cquat), 157.10 (Cquat), 150.32 (Cquat), 143.66 (Cquat), 140.52 (Cquat), 138.93 (Cquat), 135.93 (Cquat), 132.40 (CH), 130.27 (CH), 129.61 (CH), 125.74 (CH), 123.55 (CH), 121.37 (CH), 118.75 (CH), 115.12 (Cquat), 115.06 (CH), 113.48 (Cquat), 112.77 (CH), 108.36 (CH), 103.43 (CH), 55.52 (CH_3), 47.84 (CH_2), 36.67 (CH_2). HRMS calcd for $\text{C}_{24}\text{H}_{19}\text{NNaO}_3$ $[\text{M} + \text{Na}]^+$ 392.1257; found, 392.1258.

Compound 5j: 9-Hydroxy-5-[2-(4-methoxyphenyl)ethyl]-5H-indeno[1,2-b]indole-10-one

Red solid. Yield 47%. mp 150°C. IR (νcm^{-1}): 3,422, 1,667, 1,601, 1,512. ^1H NMR (CDCl_3 , 400 MHz) δ : 7.31 (m, 1H, Harom), 7.10–7.03 (m, 3H, Harom), 6.96–6.93 (m, 2H, Harom), 6.76–6.66 (m, 5H, Harom), 6.45 (bs, 1H, OH), 4.31 (t, 2H, $J=7.0$ Hz, CH_2N), 3.69 (s, 3H, OCH_3), 3.07 (t, 2H, $J=7.0$ Hz, CH_2Ph). ^{13}C NMR + DEPT (CDCl_3 , 100 MHz) δ : 186.20 (Cquat), 159.20 (Cquat), 157.13 (Cquat), 150.28 (Cquat), 143.65 (Cquat), 140.56 (Cquat), 135.92 (Cquat), 132.35 (CH), 130.13 (2CH), 129.56 (CH), 129.44 (Cquat), 125.69 (CH), 123.50 (CH), 118.77 (CH), 115.06 (Cquat), 114.61 (2CH), 113.50 (Cquat), 108.32 (CH), 103.46 (CH), 55.62 (CH_3), 48.17 (CH_2), 35.81 (CH_2). HRMS calcd for $\text{C}_{24}\text{H}_{19}\text{NNaO}_3$ $[\text{M} + \text{Na}]^+$ 392.1257; found, 392.1252.

General procedure for the synthesis of compound 6

A solution containing 0.63 mmol of **5** and 0.013 g of salcomine (Co-Salen) in 13 mL of dimethylformamide was stirred under oxygen atmosphere at room temperature for 24 hours. The solution was then poured into 100 mL of ice and water and stirred for 1 hour. The resulting precipitate was filtered and washed with water and dried to give a first quantity of **6**. The filtrate was extracted with CH_2Cl_2 . The organic phase was dried over sodium sulfate and evaporated in vacuum to give a second quantity of **6**, which was purified by silica gel column chromatography with EtOAc/cyclohexane (1:2, v/v) as the eluent.

Compound 6c: 5-(2-Phenylethyl)-5H-indeno[1,2-b]indole-6,9,10-trione

Dark red solid. Yield 70%. mp 263°C. IR (νcm^{-1}): 1,709, 1,663, 1,645, 1,594, 1,526. ^1H NMR (CDCl_3 , 400 MHz) δ : 7.58 (dd, 1H, $J_1=1.0$ Hz, $J_2=7.2$ Hz, Harom), 7.34–7.16 (m, 7H, Harom), 7.04 (dd, 1H, $J_1=1.0$ Hz, $J_2=6.9$ Hz, Harom), 6.63 (AB, 2H, H-7 and H-8), 4.77 (t, 2H, $J=7.2$ Hz, CH_2N),

3.18 (t, 2H, $J=7.2$ Hz, CH_2Ph). ^{13}C NMR + DEPT (CDCl_3 , 100 MHz) δ : 186.40 (Cquat), 183.79 (Cquat), 181.95 (Cquat), 178.45 (Cquat), 155.80 (Cquat), 140.04 (Cquat), 137.47 (CH), 136.81 (Cquat), 136.70 (CH), 133.74 (Cquat), 133.58 (CH), 130.47 (CH), 129.26 (2CH), 129.23 (2CH), 127.68 (CH), 124.96 (CH), 122.96 (Cquat), 120.79 (Cquat), 119.44 (CH), 49.76 (CH_2), 37.28 (CH_2). HRMS calcd for $\text{C}_{23}\text{H}_{15}\text{NNaO}_3$ $[\text{M} + \text{Na}]^+$ 376.0944; found, 376.0936.

Compound 6d: 5-(3-Phenylpropyl)-5H-indeno[1,2-b]indole-6,9,10-trione

Red solid. Yield 30%. mp 205°C. IR (νcm^{-1}): 1,709, 1,662, 1,647, 1,589, 1,526. ^1H NMR (CDCl_3 , 400 MHz) δ : 7.55 (dd, 1H, $J_1=0.9$ Hz, $J_2=7.2$ Hz, Harom), 7.36–7.18 (m, 7H, Harom), 6.61 (AB, 2H, H-7 and H-8), 6.52 (d, 1H, $J=7.0$ Hz, Harom), 4.52 (t, 2H, $J=7.9$ Hz, CH_2N), 2.82 (t, 2H, $J=7.1$ Hz, CH_2Ph), 2.24 (m, 2H, $\text{CH}_2\text{CH}_2\text{CH}_2$). ^{13}C NMR + DEPT (CDCl_3 , 100 MHz) δ : 183.69 (Cquat), 181.84 (Cquat), 178.50 (Cquat), 155.46 (Cquat), 140.47 (Cquat), 140.11 (Cquat), 137.44 (CH), 136.65 (CH), 133.87 (Cquat), 133.74 (CH), 136.61 (Cquat), 130.43 (CH), 129.07 (2CH), 128.97 (2CH), 126.88 (CH), 124.94 (CH), 122.80 (Cquat), 120.82 (Cquat), 119.71 (CH), 47.41 (CH_2), 33.04 (CH_2), 32.21 (CH_2). HRMS calcd for $\text{C}_{24}\text{H}_{17}\text{NNaO}_3$ $[\text{M} + \text{Na}]^+$ 390.1101; found, 390.1094.

Compound 6e: 5-Benzyl-7-phenyl-5H-indeno[1,2-b]indole-6,9,10-trione

Red solid. Yield 40%. mp 255°C. IR (νcm^{-1}): 1,715, 1,646, 1,585. ^1H NMR (CDCl_3 , 400 MHz) δ : 7.61 (d, 1H, $J=6.8$ Hz, Harom), 7.43–7.27 (m, 12H, Harom), 7.16 (d, 1H, $J=7.1$ Hz, Harom), 6.73 (s, 1H, H-8), 5.88 (s, 2H, NCH_2Ph). ^{13}C NMR + DEPT (CDCl_3 , 100 MHz) δ : 183.78 (Cquat), 181.71 (Cquat), 177.86 (Cquat), 156.49 (Cquat), 146.86 (Cquat), 140.20 (Cquat), 135.14 (Cquat), 134.29 (Cquat), 133.86 (Cquat), 133.85 (CH), 133.71 (CH), 133.34 (Cquat), 130.62 (CH), 130.10 (CH), 129.81 (2CH), 129.49 (2CH), 128.65 (2CH), 128.63 (CH), 126.85 (2CH), 125.02 (CH), 123.26 (Cquat), 120.66 (Cquat), 119.96 (CH), 51.21 (CH_2). HRMS calcd for $\text{C}_{28}\text{H}_{17}\text{NNaO}_3$ $[\text{M} + \text{Na}]^+$ 438.1101; found, 438.1086.

Compound 6f: 7-Methyl-5-(2-phenylethyl)-5H-indeno[1,2-b]indole-6,9,10-trione

Red solid. Yield 80%. mp 228°C. IR (νcm^{-1}): 1,717, 1,645, 1,609, 1,532. ^1H NMR (CDCl_3 , 400 MHz) δ : 7.53 (dd, 1H, $J_1=0.7$ Hz, $J_2=7.1$ Hz, Harom), 7.32–7.15 (m, 7H, Harom), 7.01 (d, 1H, $J=7.3$ Hz, Harom), 6.48 (q, 1H, $J=1.6$ Hz, H-8), 4.74

(t, 2H, $J=7.3$ Hz, CH_2N), 3.17 (t, 2H, $J=7.3$ Hz, CH_2Ph), 2.07 (d, 3H, $J=1.6$ Hz, Me-7). ^{13}C NMR + DEPT (CDCl_3 , 100 MHz) δ : 183.81 (Cquat), 181.95 (Cquat), 178.89 (Cquat), 155.72 (Cquat), 146.31 (Cquat), 140.04 (Cquat), 136.91 (Cquat), 133.99 (Cquat), 133.85 (Cquat), 133.51 (CH), 133.45 (CH), 130.28 (CH), 129.27 (2CH), 129.17 (2CH), 127.58 (CH), 124.79 (CH), 123.30 (Cquat), 120.39 (Cquat), 119.35 (CH), 49.78 (CH_2), 37.25 (CH_2), 16.19 (CH_3). HRMS calcd for $\text{C}_{24}\text{H}_{17}\text{NNaO}_3$ [$\text{M} + \text{Na}$] $^+$ 390.1102; found, 390.1097.

Compound 6g: 7-Phenyl-5-(2-phenylethyl)-5H-indeno[1,2-b]indole-6,9,10-trione

Red solid. Yield 74%. mp 190°C. IR ($\text{v}_{\text{cm}^{-1}}$): 1,713, 1,655, 1,583. ^1H NMR (CDCl_3 , 400 MHz) δ : 7.59 (d, 1H, $J=7.3$ Hz, Harom), 7.52–7.21 (m, 12H, Harom), 7.10 (d, 1H, $J=7.1$ Hz, Harom), 6.76 (s, 1H, H-8), 4.85 (t, 2H, $J=7.3$ Hz, CH_2N), 3.25 (t, 2H, $J=7.3$ Hz, CH_2Ph). ^{13}C NMR + DEPT (CDCl_3 , 100 MHz) δ : 183.87 (Cquat), 181.83 (Cquat), 177.91 (Cquat), 156.17 (Cquat), 146.93 (Cquat), 140.11 (Cquat), 137.01 (Cquat), 134.13 (Cquat), 133.86 (Cquat), 133.75 (CH), 133.67 (CH), 133.53 (Cquat), 130.48 (CH), 130.20 (CH), 129.91 (2CH), 129.40 (2CH), 129.25 (2CH), 128.79 (2CH), 127.69 (CH), 124.91 (CH), 123.19 (Cquat), 120.34 (Cquat), 119.59 (CH), 49.94 (CH_2), 37.29 (CH_2). HRMS calcd for $\text{C}_{29}\text{H}_{20}\text{NO}_3$ [$\text{M} + \text{H}$] $^+$ 430.1438; found, 430.1426.

X-ray data

The structure of compound 5c has been established by X-ray crystallography at 170 K. Red single crystal ($0.25 \times 0.25 \times 0.03$ mm 3) of **5c** was obtained after 20 hours at 17°C from a CS_2 /chloroform (70/30) solution: monoclinic, space group P1 21/c 1, $a=10.4901(3)$ Å, $b=10.1187(3)$ Å, $c=15.8195(4)$ Å, $\alpha=90^\circ$, $\beta=91.6571(11)^\circ$, $\gamma=90^\circ$, $V=1,678.48(8)$ Å 3 , $Z=4$, $\delta(\text{calcd})=1.343$ Mg·m $^{-3}$, formula weight=339.38 for $\text{C}_{23}\text{H}_{17}\text{NO}_2$, $F(000)=712$. Crystallographic data were acquired at ICMCB (UPR 9048) on a Bruker K-CCD APEX 2. Full crystallographic results have been deposited at the Cambridge Crystallographic Data Centre (CCDC-991361), UK, as supplementary material.¹⁸ The data were corrected for Lorentz and polarization effects and for empirical absorption correction.¹⁹ The structure was solved by direct methods Shelx 2013 and refined using Shelx 2013 suite of programs²⁰ found in the integrated OLEX2 package.²¹

Biology and biochemistry

Compounds

Mitoxantrone was purchased from Sigma-Aldrich (Saint-Quentin Fallavier, France). All commercial reagents were

of the highest available purity grade. The compounds were dissolved in DMSO, and then diluted in Dulbecco's Modified Eagle's Medium (DMEM) (high-glucose DMEM). The stock solutions were stored at -20°C , and warmed to 25°C just before use.

Cell cultures

The human fibroblast human embryonic kidney 293 (HEK293) cell line was transfected with ABCG2 (HEK293-ABCG2) or its empty vector (HEK293-*pcDNA3*),²² as well as MRP1 (HEK293-*ABCC1*) and its empty vector (HEK293-*pcDNA5*). The cells were maintained in high-glucose DMEM supplemented with 10% fetal bovine serum, 1% penicillin/streptomycin at 37°C , 5% CO_2 under controlled humidity. The mouse embryonic fibroblast wild-type (NIH3T3) and Pgp-overexpressing (NIH3T3-*ABCB1*) were maintained under the same conditions. The cell culture medium was supplemented with 0.75 mg/mL G418 (HEK293ABCG2), 200 $\mu\text{g}/\text{mL}$ hygromycin B (HEK293*pcDNA5* and HEK293*ABCC1*), or 60 ng/mL colchicin (NIH3T3).

ABCG2-mediated mitoxantrone efflux and inhibition

As previously described,²² cells were seeded at a density of 1.0×10^5 cells/well into 24-well culture plates. After 72 hours incubation, the cells were exposed to 5 μM mitoxantrone for 30 minutes at 37°C , in the presence or absence of each compound, and then washed with phosphate buffer saline (PBS) and trypsinized. The intracellular fluorescence was monitored with a FACS Calibur cytometer (Becton Dickinson, Franklin Lakes, NJ, USA) equipped with a 635-nm red laser, using the FL4 channel and at least 10,000 events were collected. The percentage of inhibition was calculated by using the following equation: % inhibition = $(C-M)/(C_{\text{ev}}-M) \times 100$, where C is the intracellular fluorescence of resistant cells (HEK293-ABCG2) in the presence of compounds and mitoxantrone, M is the intracellular fluorescence of resistant cells with only mitoxantrone, and C_{ev} the intracellular fluorescence of control cells (the same HEK293-ABCG2 cells 100% inhibited with 1 μM Ko143).

Intrinsic cytotoxicity of the inhibitory compounds

Cell viability was evaluated through the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) colorimetric assay.²³ Wild-type HEK293 cells were seeded at a density of 1×10^4 cells/well, into 96-well culture plates. After overnight incubation, the cells were treated with the compounds (0–250 μM) for 72 hours. To assess the viability, the cells were exposed to 0.5 mg/mL of MTT and incubated for

4 hours at 37°C. The culture medium was discarded, and 100 µL of a DMSO/ethanol (1:1) solution was added into each well and mixed by gently shaking for 10 minutes. Absorbance was measured at 570 nm using a microplate reader at 570 nm, and the value measured at 690 nm was subtracted. Data are the mean \pm SD of at least three independent experiments.

Inhibition of Pgp- and MRPI-mediated drug efflux

NIH3T3-*ABCB1* were seeded at a density of 6×10^4 cells/well into 24-well culture plates and incubated for 48 hours at 37°C, whereas HEK293 cells transfected with *ABCC1* were seeded at 2.5×10^5 cells/well for 72 hours. The cells were respectively exposed to rhodamine 123 (0.5 µM) or calcein-AM (0.2 µM) for 30 minutes at 37°C, in the presence or absence of each compound, then washed with PBS and trypsinized. The intracellular fluorescence was monitored with a FACS Calibur cytometer (Becton Dickinson) using the FL1 channel and at least 10,000 events were collected. The percentage of inhibition was calculated relatively to 5 µM GF120918 or 35 µM verapamil, respectively, using similar equation as demonstrated to ABCG2 inhibition.

Effects on ABCG2 ATPase activity

Vanadate-sensitive ATPase activity was measured colorimetrically by determining the liberation of inorganic phosphate from ATP, the Sf9 membranes were prepared as previously and loaded with cholesterol.²⁴ The incubation was performed in 96-well plates. Sf9 insect cell membranes (1 mg/mL) were incubated in a 50 mM Tris/HCl, 50 mM NaCl buffer (pH 8.0) containing sodium azide (3.3 mM) in the absence (with or without sodium orthovanadate at 0.33 mM) or in the presence of tested compounds (2 µM). The reaction was started by the addition of ATP-Mg (3.9 mM) and the plates were incubated for 30 minutes at 37°C. The reaction was stopped with sodium dodecylsulfate (10%) and revealed with a mixture of ammonium molybdate reagent and 10% ascorbic acid (1:4). The absorbance was measured after 30-minute incubation at 880 nm using a reader plate.

Preparation of recombinant human CK2 holoenzyme and assay of inhibitors activity

The human protein kinase CK2 holoenzyme was prepared as previously described.^{25,26} In brief, human CK2 α (CSNK2A1) and CK2 β subunits (CSNK2B) were expressed separately in *Escherichia coli* BL21(DE3) cells using the pT7-7 vector. Freshly-transformed cultures were grown overnight at 37°C in lysogeny broth (LB) medium until the stationary phase was

reached. LB medium was inoculated with the starter cultures (1:100), and protein expression was induced by isopropyl β -D-1-thiogalactopyranoside addition (1 mM final concentration) when an OD₅₀₀ of 0.6 was reached. The cultures were further incubated at 30°C during 5–6 hours for the CK2 α subunit, or 3 hours for the CK2 β subunit. After harvesting the bacterial cells by centrifugation (6,000 \times g for 10 minutes at 4°C) and disruption by sonication (3 \times 30 seconds on ice), cell debris was removed by another centrifugation at 15,000 \times g (10 minutes, 4°C). Both extracts were combined and CK2 holoenzyme was purified by a three-column procedure. Fractions were analyzed by sodium dodecyl sulphate-polyacrylamide gel electrophoresis and western blot. Those containing CK2 and showing CK2 activity were pooled and stored at –80°C as aliquots, which obtained CK2 holoenzyme with a purity higher than 99%.²⁷

For testing the compounds on CK2 inhibition, a capillary electrophoresis-based CK2 activity assay²⁸ was used. Briefly, 2 µL of the inhibitor solutions in DMSO was mixed with 78 µL of kinase buffer (50 mM Tris/HCl (pH 7.5), 100 mM NaCl, 10 mM MgCl₂ and 1 mM DTT) containing 1 µg CK2, and preincubated at 37°C for 10 minutes. The CK2 reaction was initiated by the addition of preincubated assay buffer (25 mM Tris/HCl (pH 8.5), 150 mM NaCl, 5 mM MgCl₂, 1 mM DTT, 100 µM ATP and 190 µM of the CK2 substrate peptide RRRDDDSDDD), carried on for 15 minutes at 37°C and was stopped by addition of 4 µL of 0.5 M ethylenediaminetetraacetic acid. Subsequently, the reaction samples were fed to a PA800 plus capillary electrophoresis system (Beckman Coulter, Krefeld, Germany) using acetic acid (2 M, adjusted to pH 2.0 with concentrated HCl) as the background electrolyte. Detection of the separated CK2 reaction substrate and product peptide at 214 nm was accomplished by a DAD detector. As controls for 0% and 100% inhibition, samples containing pure DMSO instead of inhibitor and samples additionally lacking the CK2 holoenzyme respectively, were treated under identical conditions. Compounds that showed more than 50% inhibition in the initial test at a final inhibitor concentration of 10 µM were subjected to half-maximal inhibitory concentration (IC₅₀) determination. For this purpose, nine final inhibitor concentrations in suitable intervals ranging from 0.001 µM to 100 µM were tested. IC₅₀ values were calculated from the resulting dose–response curves using GraphPadPrism 5.02 (GraphPad Software, Inc., La Jolla, CA, USA).

Results Chemistry

The access to 5,6,7,8-tetrahydroindeno[1,2-*b*]indole-9,10-diones 4 was previously detailed.¹⁷ Their syntheses started by

the reaction of a cyclohexane-1,3-dione with the appropriate primary amine to form the enaminone derivatives, which were then condensed with ninhydrin. The resulting *vic*-dihydroxyindeno[1,2-*b*]indole-9,10-diones were deoxygenated with tetraethylthionylamide to afford compound 4.

9-Hydroxy-5*H*-indeno[1,2-*b*]indole-10-ones 5 were prepared by dehydrogenation of 4 with 10% Pd-C in refluxing diphenyl ether.²⁹ Subsequent oxidation with molecular dioxygen in the presence of salcomine³⁰ (Co-Salen), at room temperature, gave 5*H*-indeno[1,2-*b*]indole-6,9,10-triones 6 (Figure 3).

It is known that oxidation of phenols with Fremy's salt represents an excellent synthetic method for the preparation of *p*-quinones under mild conditions and usually in good yield.³¹ For this reason, we applied these conditions to oxidize compounds 5c and 5g (Figure 4), but very low yields of the desired products were obtained. This was presumably due to steric effects of R₁ and R₂, or/and electronic destabilization of the radical intermediates. Accordingly, we preferred the method of oxidation with O₂/salcomine that we applied for the synthesis of all *p*-quinones mentioned in this work.

In total, ten phenols (series 5) and seven quinones (series 6) have been synthesized, and their structures are shown in Figure 4. The 3D spatial determinations of 5c were established by X-ray crystallography, and confirmed the structure in the solid state as anticipated on the basis of IR and ¹H NMR data (Figure 5). The key bond lengths and angles of this indeno[1,2-*b*]indolone are very similar to those given in the literature for other substituted indenoindole derivatives.^{32,33} The indeno[1,2-*b*]indolone system of 5c is nearly planar with

a mean out-of-plane deviation of 0.027 Å and the largest deviation of 0.062(2) Å for atom C7. The C4=O1 double bond was noticed at 1.231(2) Å.

Biological evaluation and structure–activity relationships

The newly synthesized phenolic and *p*-quinonic indenoindoles were assayed for their capacity to modulate ABCG2 and CK2 activities, and for their cytotoxicity.

Inhibition of ABCG2-mediated mitoxantrone efflux, and cytotoxicity

The new series of compounds 5 and 6 were first screened for their ABCG2-inhibitory activities using mitoxantrone as a substrate, and the inhibitory potency was compared with the recently-characterized ketonic derivative 4c¹⁷ (Figure 6). Other reference inhibitors of mitoxantrone efflux, used here as positive controls, were chromone 6g and Ko143 with IC₅₀ values of 0.11–0.13 μM and 0.09 μM, respectively.^{22,34}

Table 1 shows that the corresponding phenolic derivative 5c, without any substituent on both D-ring and N⁵-phenethyl, was about threefold more potent (IC₅₀=0.16 μM versus 0.43 μM). In contrast to ketonic indenoindoles,¹⁷ the addition of hydrophobic methyl and phenyl substituents on D-ring did not further increase inhibition, producing either no effect or a negative contribution, as in 5f (0.16 μM) and 5g (0.51 μM) versus 5c. Replacement of N⁵-phenethyl by a shorter linker drastically decreased the inhibitory potency, as evidenced from 5g (full inhibition, with IC₅₀=0.51 μM) to 5e (only 56% inhibition at 10 μM, with IC₅₀ around 9 μM), as well as

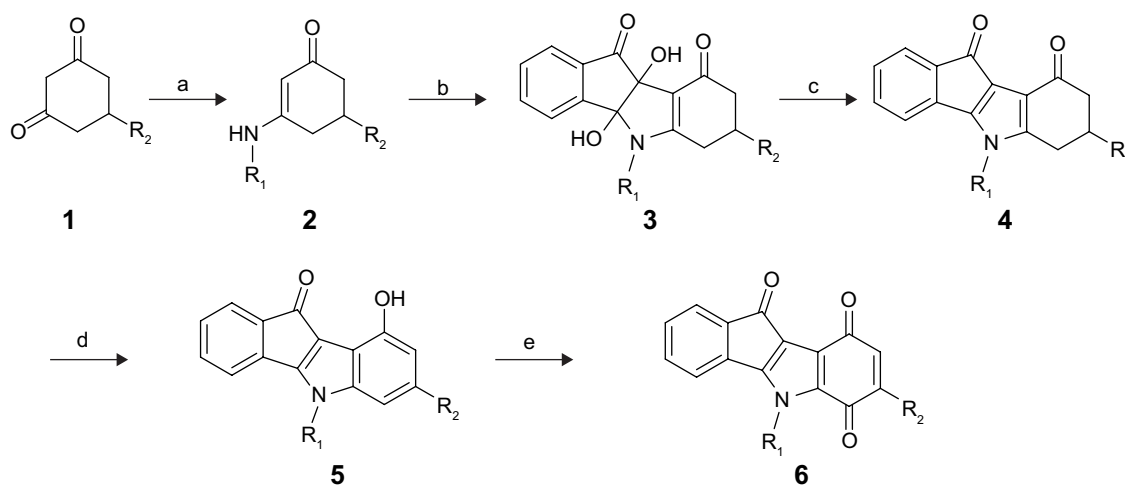
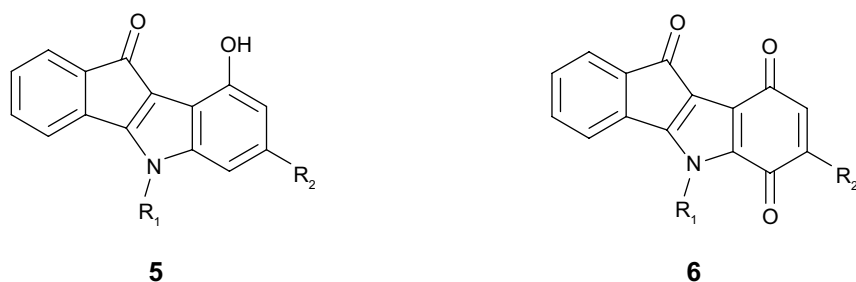


Figure 3 Reagents and conditions.

Notes: (a) R₁NH₂, Toluene, reflux; (b) ninhydrin, MeOH, rt; (c) (NEt₃)₂SO (TETA), DMF, AcOH, rt; (d) 10% Pd-C, Ph₂O, reflux, 6 hours; (e) Co-Salen, DMF, O₂, rt, 24 hours.

Abbreviations: DMF, dimethylformamide; TETA, tetraethylthionylamide; rt, room temperature.



5a⁴¹: R₁ = CH(CH₃)₂, R₂ = H

5b⁴¹: R₁ = CH₂Ph, R₂ = H

5c: R₁ = CH₂CH₂Ph, R₂ = H

5d: R₁ = CH₂CH₂CH₂Ph, R₂ = H

5e⁴¹: R₁ = CH₂Ph, R₂ = Ph

5f: R₁ = CH₂CH₂Ph, R₂ = Me

5g: R₁ = CH₂CH₂Ph, R₂ = Ph

5h: R₁ = CH₂CH₂(*ortho*-OMe)Ph, R₂ = H

5i: R₁ = CH₂CH₂(*meta*-OMe)Ph, R₂ = H

5j: R₁ = CH₂CH₂(*para*-OMe)Ph, R₂ = H

6a³⁰: R₁ = CH(CH₃)₂, R₂ = H

6b³⁰: R₁ = CH₂Ph, R₂ = H

6c: R₁ = CH₂CH₂Ph, R₂ = H

6d: R₁ = CH₂CH₂CH₂Ph, R₂ = H

6e: R₁ = CH₂Ph, R₂ = Ph

6f: R₁ = CH₂CH₂Ph, R₂ = Me

6g: R₁ = CH₂CH₂Ph, R₂ = Ph

Figure 4 Structures of the investigated indenoindole series 5 and 6.

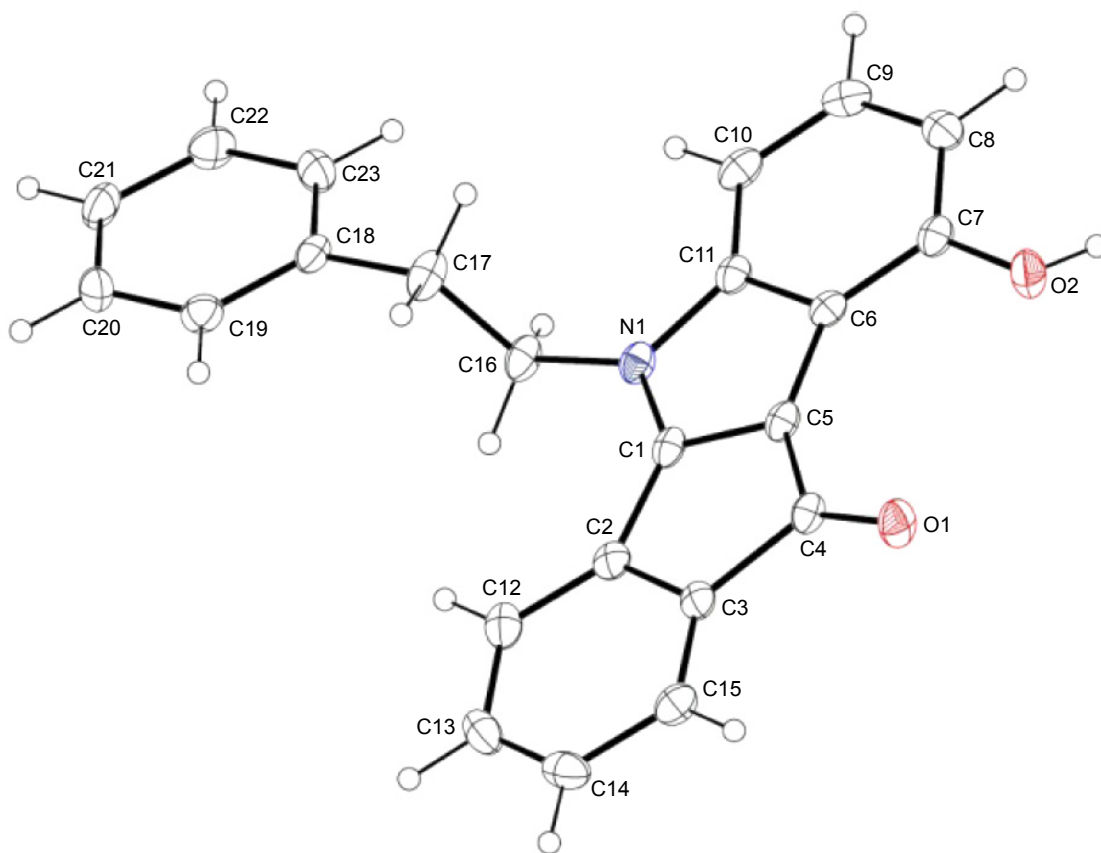
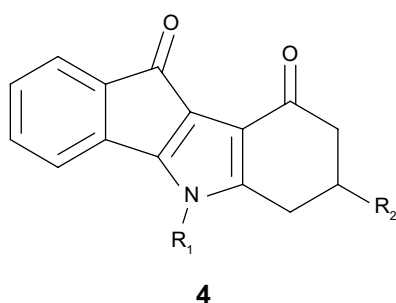


Figure 5 View of the crystal structure of 5c with our numbering scheme.

Notes: Displacement ellipsoids are drawn at the 30% probability level; the numbering used here is specific for crystallographic studies, and therefore different from that used detailed in the Materials and Methods section.



4c: R₁ = CH₂Ph, R₂ = H

4d: R₁ = CH₂CH₂CH₂Ph, R₂ = H

4h: R₁ = CH₂CH₂Ph, R₂ = Me

4j: R₁ = CH₂CH₂ (*ortho*-OMe)Ph, R₂ = H

4k: R₁ = CH₂CH₂ (*meta*-OMe)Ph, R₂ = H

4l: R₁ = CH₂CH₂ (*para*-OMe)Ph, R₂ = H

Figure 6 Structures of ketonic indenoindoles (series 4).

Note: Data from Gozzi et al.¹⁷

from 5c (0.16 μ M) to 5b (68% inhibition). However, changing the *N*⁵-phenethyl substituent by a longer linker only slightly reduced the inhibitory potency, in 5d (0.64 μ M) versus 5c (0.16 μ M). Methoxy substitution of the *N*⁵-phenethyl phenolic-indenoindole core structure (5c), at position

ortho in 5h (0.15 μ M), *para* in 5j (0.20 μ M), or *meta* in 5i (0.37 μ M) did not show substantial improvement on activity. Phenyl substitution of D-ring was not able to recover any activity for *N*⁵-benzyl derivatives (5e versus 5b). In sharp contrast, *p*-quinonic derivatives were very poor inhibitors when comparing 6c (18% inhibition) with 5c (and 4c), 6d (33% inhibition) with 5d, and 6b (10% inhibition) with 5b. However, hydrophobic substitutions of D-ring allowed a partial recovery of inhibition potency, as illustrated for methyl and phenyl additions in 6f (0.84 μ M) and 6g (0.43 μ M) versus 6c, and in 6e (41% inhibition) versus 6b.

Since the root cause of in vivo and preclinical trial failures of potent ABCG2 inhibitors are their intrinsic cytotoxic effects, one valuable complementary approach is to evaluate the in vitro cytotoxicity of new inhibitors. The ratio between cytotoxicity and inhibitory potency of the inhibitor, defined here as therapeutic ratio (TR), gives a valuable information to guide future in vivo trials. The phenolic leads, 5c, 5f, and 5h (IC₅₀=0.15–0.16 μ M), as well as 5d, 5g, and 5i (IC₅₀=0.37–0.64 μ M), displayed a low cytotoxicity, with half-maximal inhibitory growth concentrations (IG₅₀) values >42 μ M, and even >100 μ M in two cases. Such IG₅₀ values were similar to those previously observed with

Table 1 Inhibition of mitoxantrone efflux in ABCG2-transfected cells, and cytotoxicity

Indenoindoles	ABCG2 inhibition		Cytotoxicity	TR ^e
	% at 10 μM ^a	IC ₅₀ (μM)	IG ₅₀ (μM) ^d	
Ketonic reference				
4c	100±14	0.43±0.01 ^b	30.7±9.5	71
Phenolic derivatives				
5a	45	~12 ^c	>10	≥1
5b	68	~7 ^c	>4	≥1
5c	81±5	0.16±0.02 ^b	42.7±9.7	267
5d	98±25	0.64±0.23 ^b	>100	>156
5e	56	~9 ^c	4.8±0.9	<1
5f	92±22	0.16±0.01 ^b	42.3±2.8	264
5g	111±2	0.51±0.09 ^b	>100	>196
5h	85±11	0.15±0.01 ^b	54±14	360
5i	100±10	0.37±0.09 ^b	45.7±6.1	124
5j	111±14	0.20±0.01 ^b	0.8±0.1	4
<i>p</i> -Quinonic derivatives				
6a	36	~14 ^c	1.3±0.4	<1
6b	10	~50 ^c	0.53±0.15	<1
6c	18	~30 ^c	6.9±2.1	<1
6d	33	~15 ^c	6.8±0.5	<1
6e	41	~13 ^c	7.1±1.1	<1
6f	99±9	0.84±0.31 ^b	15.6±5.9	19
6g	128±6	0.43±0.01 ^b	3.7±2.2	9

Notes: ^aThe percentage inhibition of ABCG2-mediated mitoxantrone efflux was determined for each compound at a fixed concentration of 10 μ M. ^bFor the best compounds producing at least 50% inhibition at 10 μ M, a concentration range was analyzed in order to precisely determine the IC₅₀ values; the data show mean±SD. ^cFor the other, less potent compounds, a rough estimation was obtained from the experimental inhibition produced at 10 μ M. ^dThe IG₅₀ values of compounds cytotoxicity were determined after 72 hours of treatment with the MTT cell-survival method. ^eThe TR was calculated by dividing the IG₅₀ values of cytotoxicity with the corresponding IC₅₀ values of ABCG2 inhibition.

Abbreviations: IC₅₀, half-maximal inhibitory concentration; IG₅₀, half-maximal inhibitory growth concentration; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; TR, therapeutic ratio.

ketonic derivatives.¹⁷ This gave high TR values, up to 360, assumed to be quite promising for further investigations. The only exception was 5j, which displayed an unexpectedly high cytotoxicity, possibly due to strong interaction with unknown critical cellular target(s). Similarly, shortening the phenethyl substituent into either benzyl (in 5b and 5e) or isopropyl (in 5a) drastically increased cytotoxicity, leading to very low TR values (around 1). All *p*-quinonic derivatives were much more cytotoxic than phenolic compounds, except for 6e where the already high toxicity of 5e was not further increased, with IG_{50} values in the 0.5–15 μ M range, and gave very low TR values (<20, and often <1). No cross-resistance was ever observed in ABCG2-transfected cells versus the control cell line (data not shown), indicating that both phenolic and *p*-quinonic indenoindoles were apparently not transported by ABCG2.

Selectivity toward ABCG2 inhibition

The most potent indenoindole inhibitors of ABCG2 were tested against the two other major multidrug ABC transporters of cancer cells, namely Pgp/ABCB1 and MRP1/ABCC1. Figure 7 shows that none of the selected compounds, including phenolic, *p*-quinonic, and ketonic derivatives were able to induce any inhibition of Pgp-mediated rhodamine 123 efflux, as compared with the reference inhibitor GF120918/elacridar. A differential pattern was obtained with MRP1-mediated calcein efflux: while the phenolic derivatives (5c, 5f, and 5h) and *p*-quinonic derivatives (6f and 6g) produced a very limited, if any, inhibition, in contrast to the ketonic derivatives (4h, 4j, and 4k)¹⁷ (Figure 6) significantly inhibited at 2 μ M and, in two cases, as efficiently as verapamil at 10 μ M. Therefore, phenolic and *p*-quinonic, but not ketonic, inhibitors could be considered as ABCG2 selective over MRP1.

A higher selectivity of phenolic indenoindoles, relatively to the ketonic derivatives, was also observed toward the CK2 protein kinase (Table 2). Indeed, the phenolic derivatives inhibited CK2 activity with a limited potency, never

reaching a complete inhibition at 10 μ M. The high values of the ABCG2/CK2 ratio, in the range of 70–75 for 5f and 5j, were 7- to 12-fold higher than those previously obtained (in the range of 6–11) for the corresponding ketonic derivatives (4h and 4l)¹⁷ (Figure 6). A significant effect was also observed with 5d and 5i, giving a ratio value around 31, which was two- to fourfold higher than for the respective ketonic indenoindoles 4d and 4k (Figure 6). In the case of *p*-quinonic derivatives, the potency to inhibit CK2 was also lower, by about twofold, as compared with the ketonic ones, such as for 6c (IC_{50} ~13 μ M) versus 4c (7.0 μ M), and 6f (4.1 μ M) versus 4h (2.5 μ M),¹⁷ but their very low capacity to inhibit ABCG2 gave extremely low ABCG2/CK2 ratio values (<1), except for the two more active compounds, 6f and 6g, containing hydrophobic substituents on D-ring (5–27).

The interest of phenolic indenoindoles as ABCG2 inhibitors, versus ketonic and *p*-quinonic derivatives, is illustrated in Figure 8, where five phenolic leads, namely 5c, 5d, 5f, 5g, and 5i, display both a good selectivity, with a lower interaction toward CK2 than ketonic derivatives such as 4c (Figure 8A), and a low cytotoxicity by difference with all *p*-benzoquinonic derivatives (Figure 8B).

Effects on ABCG2 basal ATPase activity

The phenolic indenoindoles could also be distinguished from both ketonic and *p*-quinonic derivatives on the basis of their differential modulation of ABCG2 ATPase activity, which provided indirect information about the binding sites of the different compounds. Figure 9 shows that the vanadate-sensitive ATPase activity of insect-cell plasma membranes overexpressing human ABCG2 was strongly stimulated by the most potent phenolic inhibitors of mitoxantrone efflux: a 2- to 4.5-fold stimulation was observed with 5c, 5d, 5f, 5g, 5h, 5i, and 5j, in contrast to the very low stimulation observed with much less active 5e (Table 1). Addition of hydrophobic methoxy substitution

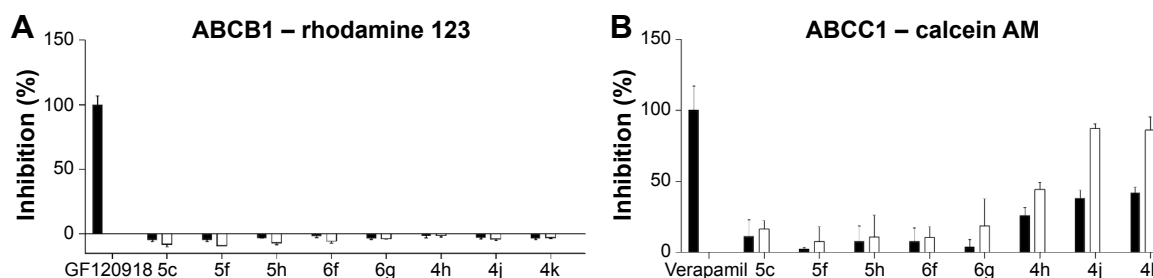


Figure 7 Ability of leads of the different series of indenoindoles to inhibit drug efflux by ABCB1 or ABCC1.

Notes: Each indenoindole derivative was assayed at either 2 μ M (black bars) or 10 μ M (white bars) for its ability to alter the efflux of either rhodamine 123 by ABCB1 (**A**) or calcein by ABCC1 (**B**) assayed by flow cytometry, as for the efflux of mitoxantrone by ABCG2 in Table 1. The inhibition produced by the two reference inhibitors, 5 μ M GF120918 and 35 μ M verapamil, was taken as 100%.

Table 2 Inhibition of human CK2 holoenzyme and comparison with ABCG2

Indenoindoles	CK2 inhibition		AGCG2/ CK2 ^d
	% at 10 μM ^a	IC ₅₀ (μM)	
Ketonic reference			
4c	59	7.0 ^b	16
Phenolic derivatives			
5a	72	2.0 ^b	<1
5b	20	~25 ^c	~4
5c	57	7.5 ^b	47
5d	27	~20 ^c	~31
5e	48	~10.5 ^c	~1
5f	42	~12 ^c	~75
5g	45	~11 ^c	~22
5h	78	1.3 ^b	9
5i	44	~11.5 ^c	~31
5j	38	~14 ^c	~70
<i>p</i> -Quinonic derivatives			
6a	60	5.5 ^b	<1
6b	70	2.2 ^b	<1
6c	40	~13 ^c	<1
6d	60	6.4 ^b	<1
6e	40	~13 ^c	<1
6f	65	4.1 ^b	5
6g	44	~11.5 ^c	~27

Notes: ^aThe percentage inhibition of CK2 activity was determined, for each compound, at 10 μ M. ^bFor the best compounds, producing at least 50% inhibition at 10 μ M, the concentration was varied in order to precisely determine the IC₅₀ values. ^cFor the other, less potent compounds, a rough estimation was obtained from the experimental inhibition produced at 10 μ M. ^dThe ABCG2/CK2 ratio, indicating the inhibitory efficiency of compounds toward ABCG2 relatively to CK2, was calculated by dividing the IC₅₀ (CK2) values with the IC₅₀ (ABCG2) values of Table 1.

Abbreviations: CK2, casein kinase II; IC₅₀, half-maximal inhibitory concentration.

on *N*⁵-phenethyl at position *ortho* did not further stimulate the ATPase activity, as in 5c versus 5h. However, shifting the methoxy substituent to position *meta* significantly increased the ATPase activity (5i), a maximal stimulation being observed for compound 5j, with a methoxy substituent at position *para*. A similar lack of positive effects was observed by lengthening the linker at *N*⁵-phenethyl when comparing 5c with 5d. However, changing the *N*⁵-phenethyl substituent into a shorter linker, from 5g to 5e, was quite detrimental to stimulation. Methyl substitution of D-ring was not able to increase the ATPase stimulation activity (5c versus 5f); however, changing the methyl into phenyl (5f versus 5g) induced a twofold increased stimulation.

In addition, the *p*-quinonic derivative 6f, as well as 6c and 6e (not shown here), did not produce any effect, while a very limited stimulation was observed for compound 6g (Figure 9). This lack of effect on ATPase activity was also observed with the four ketonic derivative leads, 4c, 4h, 4j, and 4k, which did not induce any significant stimulation, suggesting either distinct binding sites or different

induced conformational changes associated with inhibition of ABCG2-mediated mitoxantrone efflux. All indenoindoles derivatives behaved differently from the reference inhibitors, Ko143 and chromone 1, which strongly inhibited ATPase activity.

Discussion

This paper shows that phenolic indenoindoles constitute quite interesting ABCG2 modulators as potent and selective inhibitors, with low cytotoxicity. They strongly stimulate ATPase activity, by difference with ketonic derivatives and reference inhibitors.

Phenolic indenoindoles as better ABCG2 inhibitors than ketonic derivatives

The advantages of the phenolic, over the ketonic, derivatives are related to three main aspects. Firstly, a higher potency (about threefold) in inhibition of drug-efflux activity: such an increase might be due to a higher reactivity of the phenol versus the ketone group rather than to the planarity of the indenoindoles core, since the inhibition was markedly altered in planar *p*-quinonic derivatives. Secondly, a higher selectivity for ABCG2 among MDR transporters and CK2: this was monitored through the absence of interaction with MRP1, and a threefold lower interaction with CK2 giving a 3–7 higher ABCG2/CK2 ratio; this further confirms the possibility to convert CK2 inhibitors into ABCG2 inhibitors upon appropriate substitutions, as previously observed with ketonic derivatives,¹⁷ and extends the differences to high ABCG2/CK2 ratio values, of 70–75, for 5f and 5j. Thirdly, a strong modulation, up to 4.5-fold, of the basal ATPase activity: this allows considering the possibility of depleting intracellular ATP from ABCG2-overexpressing cells, with the aim of promoting selective cell death, as proposed for the Pgp-dependent collateral sensitivity observed in the presence of verapamil.^{35–37} Such a stimulation of ATPase activity contrasts with the lack of effect observed with both ketonic and *p*-quinonic derivatives, and the inhibition produced by the reference inhibitors chromone 1 and Ko143,²² as well as fumitremogin C.³⁸

Among the six best phenolic derivatives, toward both potency of ABCG2 inhibition (IC₅₀ values in the range of 0.16–0.64 μ M) and selectivity (ABCG2/CK2 ratio values in the range of 22–75), one compound, namely 5j, appeared to be cytotoxic. The five remaining leads, namely 5c, 5d, 5f, 5g, and 5i, which displayed high TR values, in the range of 124–267, constitute good candidates for additional experiments to check their suitability in future assays in animal models.

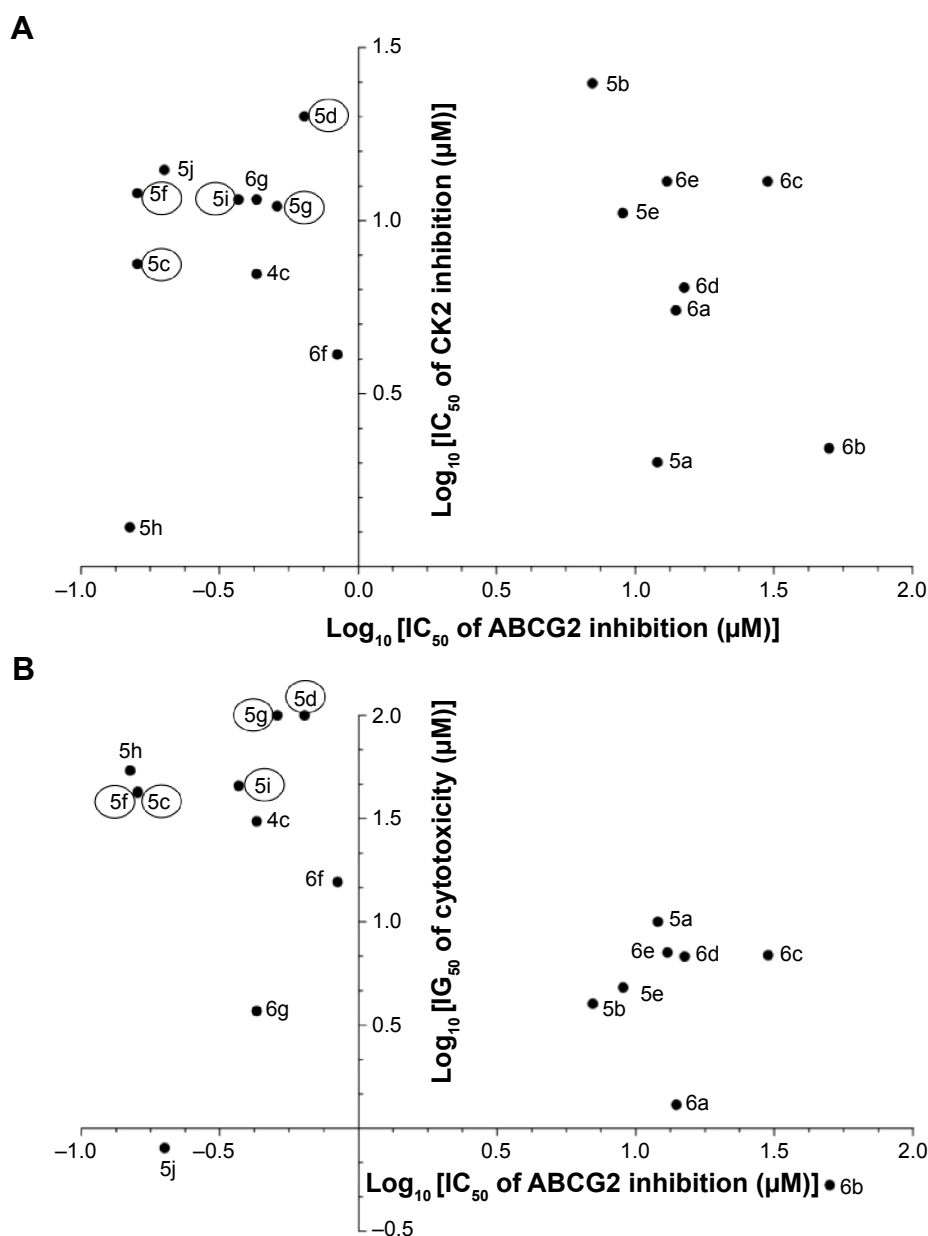


Figure 8 Phenolic indenoindoles as better ABCG2 inhibitors over ketonic and *p*-quinonic derivatives. ABCG2 inhibition versus CK2 inhibition (**A**) and versus cytotoxicity (**B**). Abbreviations: CK2, casein kinase II; IC_{50} , half-maximal inhibitory concentration.

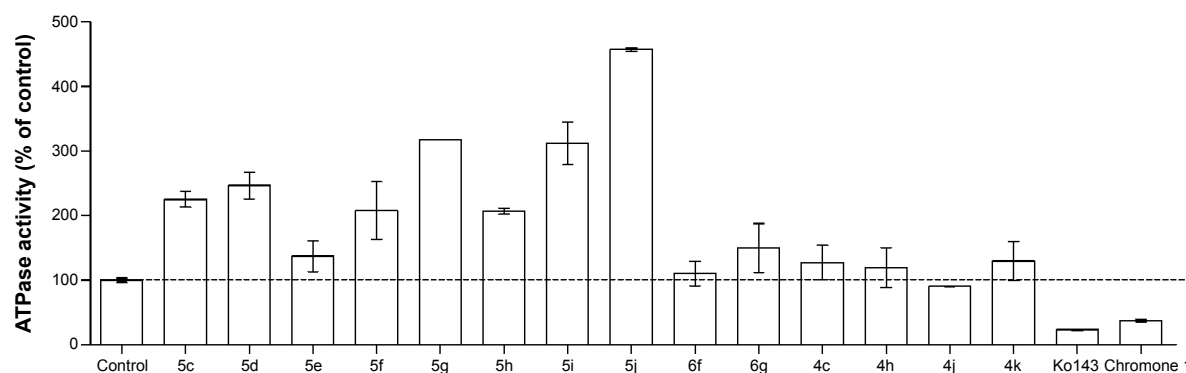


Figure 9 Modulation of basal ATPase activity by indenoindoles.

Notes: The effects of 2 μM compounds from the different series on the vanadate-sensitive ATPase activity of ABCG2 were tested on 10 μg of ABCG2-containing membrane vesicles (prepared from Sf9 cells overexpressing human ABCG2, and loaded with cholesterol). The specific basal ATPase activity of the control without inhibitor, taken as 100%, was 7.0 ± 0.4 nmol ATP hydrolyzed/min \times mg of proteins. Two reference inhibitors, Ko143¹³ and chromone 1,³⁵ were used under the same conditions for comparison.

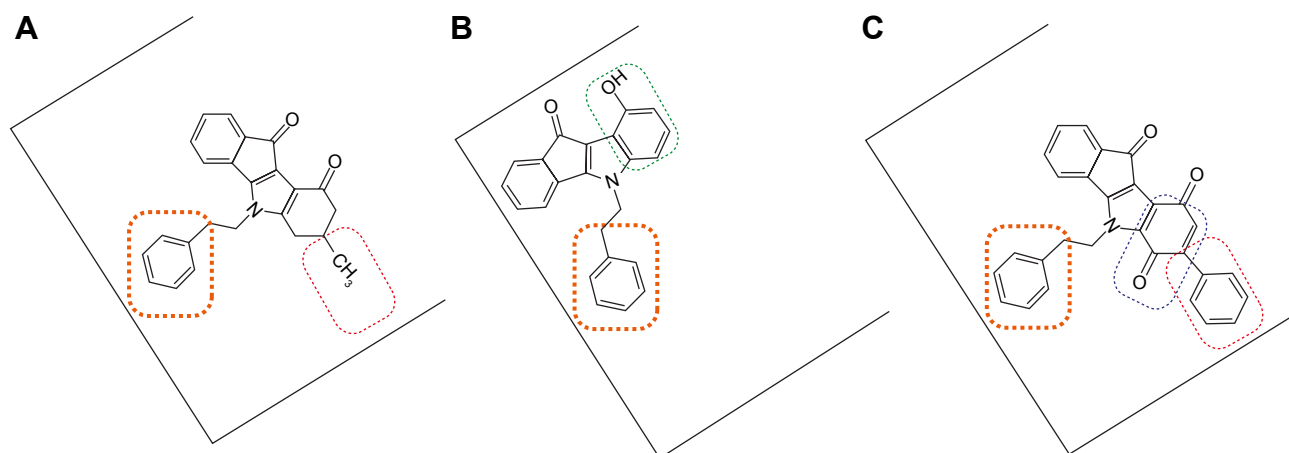


Figure 10 Tentative representation of partly-overlapping sites for indenoindoles, within ABCG2, allowing binding of, and inhibition by, the different types of derivatives. **Notes:** (A) Ketonic 4h; (B) phenolic 5c; and (C) *p*-quinonic 6g. The common phenethyl group, which is essential for ABCG2 inhibition and selectivity, is framed in orange; the other substituents positively contributing to inhibition are framed in either red (hydrophobic methyl or phenyl) or green (phenol) whereas the negatively-contributing *p*-quinone is framed in blue.

Molecular mechanism and polyspecificity

The different modulation of ABCG2 basal ATPase activity by phenolic indenoindoles versus non-stimulatory ketonic derivatives and inhibitory reference inhibitors suggests distinct binding sites. A similar situation was previously observed with methoxy *trans*-stilbenes which, in combination with GF120918, prazosin, or nilotinib, produced additive inhibitory effects on drug efflux.³⁹ Although the inhibitor binding sites are probably distant from the cytosolic nucleotide-binding domain, they are indeed able to modulate the ATPase activity; this suggests the existence of allosteric interactions, between transmembrane and cytosolic domains of the transporter, controlling the strict coupling between ATP binding/hydrolysis and drug transport.

The common *N*⁵-phenethyl substituent found in all active indenoindole derivatives, as illustrated in Figure 10, should however induce partial overlapping of the binding sites. The hydrophobic substituents shown to increase the binding affinity of ketonic indenoindoles, produced similar effects in *p*-quinonic derivatives, where they partially compensated the strongly-negative contribution of the second ketone group, at *para* position versus the first one, within the quinone moiety. In contrast, the phenol group appeared to interact with a distinct, although likely overlapping, subsite associated with ATPase-stimulation effects. A similar overlapping of binding sites was previously observed for ABCG2-selective acridones in comparison with GF120918/elacridar, a dual inhibitor strongly interacting with Pgp.⁴⁰

Finally, these diverse binding sites, located close each other and partly overlapping, constitute a new demonstration of ABCG2 polyspecificity toward inhibitors, similarly to that better known for substrates.

Acknowledgments

EW and GV were recipients of mobility doctoral fellowships from the Brazilian CNPq-CAPES (Science without Borders Program 245762/2012-4) and CAPES (Process numbers 8792127 and 2303/10-8), respectively. NDY was recipient of a postdoctoral fellowship for the Control of Cancer-CNPq (Science without Borders Program). Financial support was provided by the CNRS and Université Lyon 1 (UMR 5086), the Ligue Nationale Contre le Cancer (Equipe labellisée Ligue 2014), an international grant from French ANR and Hungarian NIH (2010-INT-1101-01). The Aquitaine Region is thanked for supporting equipment set up in CESAMO and at ICMCB. C Ozvegy-Laczka and B Sarkadi are acknowledged for providing the insect cell membranes overexpressing human ABCG2, L Luku Tchamabe for technical assistance, and Dr A Coleman for improving the English.

Author contributions

Gustavo Jabor Gozzi and Zouhair Bouaziz contributed equally to the experiments. Senior investigators, Marc Le Borgne and Attilio Di Pietro contributed equally to the work supervision. All authors contributed toward data analysis, drafting and revising the paper and agree to be accountable for all aspects of the work.

Disclosure

The authors report no conflicts of interest in this work.

References

1. Gottesman MM. Mechanisms of cancer drug resistance. *Annu Rev Med.* 2002;53:615–627.
2. Szakács G, Paterson JK, Ludwig JA, Booth-Genthe C, Gottesman MM. Targeting multidrug resistance in cancer. *Nat Rev Drug Discov.* 2006;5(3):219–234.

3. Juliano RL, Ling V. A surface glycoprotein modulating drug in chinese hamster ovary cell mutants. *Biochim Biophys Acta*. 1976; 455(1):152–162.
4. Cole SP, Bhardwaj G, Gerlach JH, et al. Overexpression of a transporter gene in a multidrug-resistant human lung cancer cell line. *Science*. 1992;258(5088):1650–1654.
5. Allikmets R, Schriml LM, Hutchinson A, Romano-Spica V, Dean M. A human placenta-specific ATP-binding cassette gene (ABCP) on chromosome 4q22 that is involved in multidrug resistance. *Cancer Res*. 1998;58(23):5337–5339.
6. Doyle LA, Yang W, Abruzzo LV, et al. A multidrug resistance transporter from human MCF-7 breast cancer cells. *Proc Natl Acad Sci U S A*. 1998;95(26):15665–15670.
7. Miyake K, Mickley L, Litman T, et al. Molecular cloning of cDNAs which are highly overexpressed in mitoxantrone-resistant cells: demonstration of homology to ABC transport genes. *Cancer Res*. 1999;59(1):8–13.
8. Cripe LD, Uno H, Paietta EM, et al. Zosuquidar, a novel modulator of P-glycoprotein does not improve the outcome of older patients with newly diagnosed acute myeloid leukemia: a randomized, placebo-controlled trial of the Eastern Cooperative Oncology Group 3999. *Blood*. 2010;116(20):4077–4085.
9. Darby RA, Callaghan R, Mc Mahon RM. P-glycoprotein inhibition: the past, the present and the future. *Curr Drug Metab*. 2011; 12(8):722–731.
10. Vlaming ML, Lagas JS, Schinkel AH. Physiological and pharmacological roles of ABCG2 (BCRP): recent findings in Abcg2 knockout mice. *Adv Drug Deliv Rev*. 2009;61(1):14–25.
11. Natarajan K, Xie Y, Baer MR, Ross DD. Role of breast cancer resistance protein (BCRP/ABCG2) in cancer drug resistance. *Biochem Pharmacol*. 2012;83(8):1084–1103.
12. Rabindran SK, He H, Singh M, et al. Reversal of a novel multidrug resistance mechanism in human colon carcinoma cells by fumitremorgin C. *Cancer Res*. 1998;58(24):5850–5858.
13. Allen JD, van Loevezijn A, Lakhai JM, et al. Potent and specific inhibition of the breast cancer resistance protein multidrug transporter in vitro and in mouse intestine by a novel analogue of fumitremorgin C. *Mol Cancer Ther*. 2002;1(6):417–425.
14. Winter E, Devantier Neuenfeldt P, Chiaradia-Delatorre LD, et al. Symmetric bis-chalcones as a new type of breast cancer resistance protein inhibitors with a different mechanism than chromones. *J Med Chem*. 2014;57(7):2930–2941.
15. Arnaud O, Boumendjel A, Geze A, et al. The acridone derivative MBLI-87 reverses breast cancer resistance protein-mediated resistance to irinotecan in xenografts. *Eur J Cancer*. 2011;47(4):640–648.
16. Honorat M, Guitton J, Gauthier C, et al. MBL-II-141, a chromone derivative, enhances irinotecan anticancer efficiency in ABCG2-positive xenografts. *Oncotarget*. 2014;5(23):11957–11970.
17. Gozzi GJ, Bouaziz Z, Winter E, et al. Converting potent indeno[1,2-*b*] indole inhibitors of protein kinase CK2 into selective inhibitors of the breast cancer resistance protein ABCG2. *J Med Chem*. 2015; 58(1):265–277.
18. Supplementary X-ray crystallographic data: Cambridge Crystallographic Data Centre, University Chemical Lab, Lensfield Road, Cambridge CB2 1EW, UK; Email: deposit@chemcrs.cam.ac.uk
19. Sheldrick GM. *SADABS*. Göttingen: University of Göttingen; 1996.
20. Sheldrick GM. A short history of SHELX. *Acta Crystallogr A*. 2008;64(Pt 1):112–122.
21. Dolomanov OV, Bourhis LJ, Gildea RJ, Howard JAK, Puschmann H. OLEX2: a complete structure solution, refinement and analysis program. *J Appl Cryst*. 2009;42:339–341.
22. Winter E, Lecerf-Schmidt F, Gozzi GJ, et al. Structure-activity relationships of chromone derivatives toward mechanism of interaction with, and inhibition of, breast cancer resistance protein ABCG2. *J Med Chem*. 2013;56(24):9849–9860.
23. Mosmann T. Rapid colorimetric assay for cellular growth and survival. Application to proliferation and cytotoxicity assays. *J Immunol Methods*. 1983;65(1–2):55–63.
24. Telbisz A, Muller M, Ozvegy-Laczka C, et al. Membrane cholesterol selectively modulates the activity of the human ABCG2 multidrug transporter. *Biochim Biophys Acta*. 2007;1768(11): 2698–2713.
25. Olgen S, Götz C, Jose J. Synthesis and biological evaluation of 3-(substituted-benzylidene)-1,3-dihydroindolin derivatives as human protein kinase CK2 and p60(c-Src) tyrosine kinase inhibitors. *Biol Pharm Bull*. 2007;30(4):715–718.
26. Grankowski N, Boldyreff B, Issinger OG. Isolation and characterization of recombinant human casein kinase II subunits α and β from bacteria. *Eur J Biochem*. 1991;198(1):25–30.
27. Guillon J, Le Borgne M, Rimbault C, et al. Synthesis and biological evaluation of novel substituted pyrrolo[1,2-*a*]quinoxaline derivatives as inhibitors of the human protein kinase CK2. *Eur J Med Chem*. 2013;65:205–222.
28. Gratz A, Götz C, Jose J. A CE-based assay for human protein kinase CK2 activity measurement and inhibitor screening. *Electrophoresis*. 2010;31(4):634–640.
29. Poumaroux A, Bouaziz Z, Domard M, Fillion H. Regiospecific hetero Diels-Alder synthesis of pyrido[2,3-*b*]- and [3,2-*b*]carbazole-5,11-diones. *Heterocycles*. 1997;45:585–596.
30. Hundsdörfer C, Hemmerling HJ, Hamberger J, et al. Novel indeno[1,2-*b*]indoloquinones as inhibitors of the human protein kinase CK2 with antiproliferative activity towards a broad panel of cancer cell lines. *Biochem Biophys Res Commun*. 2012;424(1): 71–75.
31. Zimmer H, Lankin DC, Horgan SW. Oxidations with potassium nitrosodisulfonate (Fremy's radical). Teuber reaction. *Chem Rev*. 1971;71(2):229–246.
32. Janreddy D, Kavala V, Bosco JWW, Kuo C-W, Yao C-F. An easy access to carbazolones and 2,3-disubstituted indoles. *Eur J Org Chem*. 2011;12:2360–2365.
33. Bill K, Black GG, Falshaw CP, Sainsbury M. The coupling reactions of 3-acylindoles and proof of structure of the palladium (II) acetate mediated cyclisation reaction product of 3-benzoyl-1-methylindole. *Heterocycles*. 1983;20:2433–2436.
34. Valdameri G, Genoux-Bastide E, Peres B, et al. Substituted chromones as highly potent nontoxic inhibitors, specific for the breast cancer resistance protein. *J Med Chem*. 2012;55(2):966–970.
35. Karwatsky J, Lincoln MC, Georges E. A mechanism for P-glycoprotein-mediated apoptosis as revealed by verapamil hypersensitivity. *Biochemistry*. 2003;42(42):12163–12173.
36. Hall MD, Handley MD, Gottesman MM. Is resistance useless? Multidrug resistance and collateral sensitivity. *Trends Pharmacol Sci*. 2009;30(10):546–556.
37. Szakacs G, Hall M, Gottesman MM, et al. Targeting the Achilles heel of multidrug-resistant cancer by exploiting the fitness cost of resistance. *Chem Rev*. 2014;114(11):5753–5774.
38. Ozvegy C, Litman T, Szakacs G, et al. Functional characterization of the human multidrug transporter, ABCG2, expressed in insect cells. *Biochem Biophys Res Commun*. 2001;285(1):111–117.
39. Valdameri G, Pereira Rangel L, Spatafora C, et al. Methoxy stilbenes as potent, specific, untransported, and nontoxic inhibitors of breast cancer resistance protein. *ACS Chem Biol*. 2012;7(2):322–330.
40. Boumendjel A, Macalou S, Valdameri G, et al. Targeting the multidrug ABCG2 transporter with flavonoidic inhibitors: in vitro optimization and in vivo validation. *Curr Med Chem*. 2011;18(22):3387–3401.
41. Hundsdörfer C, Hemmerling HJ, Götz C, et al. Indeno[1,2-*b*]indole derivatives as novel class of potent human protein kinase CK2 inhibitors. *Bioorg Med Chem*. 2012;20(7):2282–2289.

Drug Design, Development and Therapy

Dovepress

Publish your work in this journal

Drug Design, Development and Therapy is an international, peer-reviewed open-access journal that spans the spectrum of drug design and development through to clinical applications. Clinical outcomes, patient safety, and programs for the development and effective, safe, and sustained use of medicines are a feature of the journal, which

has also been accepted for indexing on PubMed Central. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <http://www.dovepress.com/drug-design-development-and-therapy-journal>