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# Five decades of N<sub>2</sub> fixation research in the North Atlantic Ocean

Mar Benavides<sup>1\*</sup> and Maren Voss<sup>2</sup>

<sup>1</sup> Centre National de la Recherche Scientifique/Institut National des Sciences de l'univers, Institut de Recherche pour le Développement, Mediterranean Institute of Oceanography, Aix Marseille Université, Université de Toulon, UM 110, Nouméa, New Caledonia, <sup>2</sup> Department of Biological Oceanography, Leibniz Institute for Baltic Sea Research, Rostock, Germany

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### \*Correspondence:

Mar Benavides,  
Centre National de la Recherche  
Scientifique/Institut National des  
Sciences de l'univers, Institut de  
Recherche pour le Développement,  
Mediterranean Institute of  
Oceanography, Aix Marseille  
Université, Université de Toulon, 101  
Promenade Roger Laroque, BP A5,  
98848 Nouméa  
cedex, New Caledonia  
mar.benavides@ird.fr

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Dinitrogen (N<sub>2</sub>) fixation (the reduction of atmospheric N<sub>2</sub> to ammonium by specialized prokaryotic microbes), represents an important input of fixed nitrogen and contributes significantly to primary productivity in the oceans. Marine N<sub>2</sub> fixation was discovered in the North Atlantic Ocean (NA) in the 1960s. Ever since, the NA has been subject to numerous studies that have looked into the diversity and abundance of N<sub>2</sub>-fixing microbes (diazotrophs), the spatial and temporal variability of N<sub>2</sub> fixation rates, and the range of physical and chemical variables that control them. The NA provides 10–25% of the globally fixed N<sub>2</sub>, ranking as the third basin with the largest N<sub>2</sub> fixation inputs in the world's oceans. This basin suffers a chronic depletion in phosphorus availability, more aeolian dust deposition than any other basin in the world's oceans, and significant nutrient inputs from important rivers like the Amazon and the Congo. These characteristics make it unique in comparison with other oceanic basins. After five decades of intensive research, here we present a comprehensive review of our current understanding of diazotrophic activity in the NA from both a geochemical and biological perspective. We discuss the advantages and disadvantages of current methods, future perspectives, and questions which remain to be answered.

**Keywords:** Nitrogen, *nifH*, tropical, upwelling, Amazon River plume, phosphorus, N\*

## Introduction

Dinitrogen (N<sub>2</sub>) fixation—the reduction of atmospheric nitrogen to ammonia— is an energy intensive process performed by prokaryotes from a variety of habitats, including rice fields, lacustrine waters, and the ocean (Karl et al., 2002). In the latter, N<sub>2</sub> fixation provides an input of fixed nitrogen that may become the primary source of this nutrient when other inputs like coastal runoff, atmospheric deposition, or subsurface nutrient upwelling and diffusion are minor or insufficient to meet the photosynthetic phytoplankton demand. This can be the case in some of the remote and chronically stratified oligotrophic subtropical gyres, which make up approximately half of the world's ocean surfaces. In that sense, N<sub>2</sub> fixation plays a key role in sustaining primary production on a global basis, powering the biological carbon pump and eventually conferring the oceans the ability to sequester atmospheric carbon dioxide (CO<sub>2</sub>). The availability of fixed nitrogen ultimately depends on the difference between “gains” and “losses” through N<sub>2</sub> fixation vs. denitrification and anammox (Falkowski, 1997; Gruber and Galloway, 2008), and so does the magnitude and distribution of diazotrophic activity in the oceans (Weber and Deutsch, 2014). As a result, the complexity of the many reactions involved between the different nitrogen chemical forms and the diversity of the microorganisms responsible for them have encouraged a wide body of research in the last decades

(Zehr and Kudela, 2011). A better understanding of the nitrogen cycle is needed in order to predict the behavior of our oceans in the light of climate change and other anthropogenic perturbations.

Probably due to the proximity to the wealthier surrounding countries of North America and Europe, the North Atlantic Ocean (NA) has been subject to the vast majority of the N<sub>2</sub> fixation related studies conducted so far. Marine nitrogen fixation was first observed in the NA in the early 1900s. After Brandt (1899) introduced the concept of limiting nutrients in controlling phytoplankton production in the ocean, the German scientists Joseph Keutner and Wilhelm Benecke firstly reported that diazotrophic bacteria common in terrestrial environments like *Clostridium* and *Azobacter* were also present in the sea, and hence their role in providing these nutrients could be important (Mills, 2012). Modern marine N<sub>2</sub> fixation research in the NA was pioneered by the research of Richard Dugdale, John Goering, and colleagues in the tropical and subtropical Northwest Atlantic (NWA) (Dugdale et al., 1961, 1964; Goering et al., 1966). These authors found considerable <sup>15</sup>N atom % excess values (up to 0.9) after incubating isolated *Trichodesmium* colonies in the presence of <sup>15</sup>N<sub>2</sub>. Intrigued by the recurrent observation of *Trichodesmium* in offshore nutrient-poor waters of the NWA, Goering et al. (1966) conducted further research in an area affected by the Amazon River plume. In subsequent years, the drawdown of dissolved inorganic carbon (DIC) at BATS in the absence of sufficient nitrate was tentatively attributed to N<sub>2</sub> fixation, possibly in combination with atmospheric nitrogen deposition and deviations from Redfield stoichiometry (Michaels et al., 1994). Overcoming the temporal and spatial scale constraints of *in situ* <sup>15</sup>N<sub>2</sub> uptake measurements, the calculation of the N\* parameter—a quasi-conservative tracer which indicates the excess of nitrate as compared to phosphate considering the Redfield ratio of nitrate:phosphate ratio (N:P) = 16:1 (Michaels et al., 1996; Gruber and Sarmiento, 1997)—allowed basin-scale estimations of nitrate concentrations in excess of that predicted by Redfield stoichiometry. Michaels et al. (1996) calculated a nitrogen excess generation rate of 51.8–89.6 Tg N y<sup>-1</sup> in the top 1000 m of the Sargasso Sea, followed by Gruber and Sarmiento (1997) who estimated a fixed nitrogen excess rate of 28 Tg N y<sup>-1</sup> for the tropical and subtropical NA. The estimated nitrogen excess was attributed to N<sub>2</sub> fixation, predominantly by *Trichodesmium* (Gruber and Sarmiento, 1997). *Trichodesmium* is strongly limited by the availability of iron, which is a cofactor of its nitrogenase enzyme and makes an important part of its biomass (Berman-Frank et al., 2001). Early on, the importance of the input of Saharan dust with relatively high iron content to the NA was related to N<sub>2</sub> fixation activity by *Trichodesmium* and the subsequent formation of a nitrogen excess signal (Duce et al., 1991; Michaels et al., 1996; Gruber and Sarmiento, 1997 the role of iron in shaping diazotrophic activity and diversity is further discussed in Section Responses to Atmospheric Dust Inputs below). At that time, the continued observation of dense surface blooms of *Trichodesmium* in the NWA and in the Caribbean Sea (e.g., Carpenter and Price, 1977), together with the associated high N<sub>2</sub> fixation rates observed (Carpenter and Romans, 1991), pointed

toward a prominent role for this conspicuous cyanobacterium in N<sub>2</sub> fixation activity in the NA. Research was not strictly limited to *Trichodesmium*, as other diazotrophs could be studied with the microscopy-based techniques of the time, notably diatom-diazotroph associations (DDAs). These DDAs are commonly formed by the cyanobacteria *Richelia intracellularis* and *Calothrix rhizosoleniae* which form symbiotic associations with several diatom genera like *Rhizosolenia*, *Hemiaulus*, and *Chaetoceros* (e.g., Carpenter et al., 1999).

During the late 1990s and 2000s, the especial interest in *Trichodesmium* as a key species made way for a great deal of research initiatives focused on the independent contribution of this cyanobacterium to basin or global-scale N<sub>2</sub> fixation (e.g., estimated at 22.4 Tg N y<sup>-1</sup> for the NA by Capone et al., 2005). These added to a variety of studies on its physiology, autoecology, and distribution (recently reviewed by Bergman et al., 2012). Later studies found that *Trichodesmium* is mostly restricted to the tropical and subtropical NWA waters due to their growth temperature optimum (24–30°C; Breitbart et al., 2007), and their preference for low dissolved inorganic nitrogen and calm stratified waters (Bergman et al., 2012).

The revolutionary application of polymerase chain reaction (PCR) techniques to amplify the *nifH* gene (Zehr et al., 1998)—the gene encoding the iron subunit of the nitrogenase enzyme—revolutionized the way diazotrophy was scrutinized in the ocean. Molecular studies unveiled the ubiquity of unicellular diazotrophic cyanobacteria (UCYN) which can equal or even exceed N<sub>2</sub> fixation rates by *Trichodesmium* (Falcón et al., 2004; Montoya et al., 2004), as well as inhabit colder and more nutrient rich areas of the NA. Studies targeting the *nifH* gene in the tropical and subtropical NA have found a predominance of *Trichodesmium*, followed by UCYN-A, with the latter present in deeper and colder waters than the former (Langlois et al., 2005, 2008; Goebel et al., 2010). The distribution of these two phylotypes seems to follow a “shifting” pattern, so that *Trichodesmium* may dominate at the surface while UCYN-A dominates in deeper levels of the photic zone (Langlois et al., 2005). Additionally, current evidence points toward the dominance of *Trichodesmium* in the NWA while UCYN-A dominates in the NEA (Agawin et al., 2014). Other UCYN like *Crocospaera* (UCYN-B) and UCYN-C phylotypes have also been recurrently detected in the NA, however in much lower abundances (e.g., Langlois et al., 2008).

Five decades have passed since the work of Dugdale et al. (1964) in the Sargasso Sea. Numerous studies on the ecological, biogeochemical and molecular aspects of diazotrophy have arisen since then, and work in the NA now represents the greatest number of N<sub>2</sub> fixation and diazotrophic diversity studies of the world's oceans (Luo et al., 2012). Despite this, other facets of diazotrophy are not well-constrained yet. The discovery of new organisms with surprising metabolisms like the unicellular diazotrophic cyanobacterium UCYN-A (*Candidatus* Atelocyanobacterium thalassa; Zehr et al., 2008; Thompson et al., 2012), the measurement of significant N<sub>2</sub> fixation rates in unexpected habitats (e.g., Bonnet et al., 2013), and the possibility that global N<sub>2</sub> fixation rates have been systematically underestimated in the past (Großkopf et al., 2012) has left room

for plenty of new questions still to be answered, including: how much do heterotrophic diazotrophs and UCYN contribute to total N<sub>2</sub> fixation?, how much does N<sub>2</sub> fixation contribute to new production and what is its role in the biological carbon pump?, can the methodological underestimation of available N<sub>2</sub> fixation rates explain the current differences between denitrification and nitrogen fixation at a global scale?, to what extent published N<sub>2</sub> fixation rates are affected by <sup>15</sup>N<sub>2</sub> gas contamination?, and, how will diazotrophs respond to climate change? The advent of new molecular biology methods, high resolution mass spectrometry techniques and single-cell techniques, together with more powerful modeling approaches should provide new insights and answers to these questions.

It is the purpose of this review to synthesize the knowledge gained on the distribution, magnitude and controls of diazotrophic activity and diversity in the pelagic NA through the years, providing an outlook toward better constraints of basin-scale N<sub>2</sub> fixation rates and identification of the various diazotroph species including previously unexplored sites with diverging environmental conditions. Benthic -and coral-associated N<sub>2</sub> fixation studies are out of the scope of this review. The reader is referred to comprehensive reviews on those topics (see Capone et al., 2008).

## Physical and Chemical Controls of Diazotrophic Activity, Diversity, and Distribution

Diazotrophic activity and diversity is known to be regulated by a variety factors, broadly divided into physical (light, temperature, salinity, water column stability, turbulence, mesoscale variability, density fronts), and chemical factors (N:P ratios and iron availability, among other trace metals) (Luo et al., 2014). Both groups of controlling factors are inextricably connected as physical structures induce changes in nutrient availability patterns (e.g., diffusion through fronts, upwelling in the core of eddies, etc.). The NA spans a suite of regional environments where the physical and/or chemical constraints can be substantially different. For example, we can compare the turbulent, cold and nutrient rich waters of the Northwest African upwelling to the stratified and oligotrophic Sargasso Sea, the steep gradients of the Gulf Stream to the Amazon River plume, and the highest desert dust aeolian inputs in the world (Jickells, 2005). In this section we explore how these different factors shape the distribution and activity of diazotrophs in the NA.

### Impact of Physical Factors on Diazotrophy

Hydrographic structures such as filaments, eddies or fronts can have profound effects on biological activity. Besides the changes in nutrient availability that these structures may cause, the spatial distribution of microorganisms can also change in response to physical forcing phenomena. For example, phytoplankton activity may be reduced when cells are transported to dimmer waters in an anticyclonic eddy, or cells can accumulate along a density front between riverine and open ocean waters.

In the case of diazotrophs, *Trichodesmium* is an obvious candidate to experience physical-driven changes in its spatial

distribution given the buoyancy conferred by intracellular gas vesicles (Villareal and Carpenter, 2003). Local accumulations of *Trichodesmium* have been observed in anticyclonic eddies across the NA (Davis and McGillicuddy, 2006), with an increasing abundance toward the West. In the NEA however, *Trichodesmium* is too scarce and usually present as free trichomes instead of colonies (Moore et al., 2009; Fernández et al., 2010; Benavides et al., 2011). This low abundance probably prevents any structuring effects from mesoscale activity (Agawin et al., 2013). However, relative peaks of *Trichodesmium* abundance have been observed in typical NEA hydrographic structures such as the Azores Front or the upwelling filament off Cape Ghir (in the Northwest Africa coastal upwelling; Benavides et al., 2011).

In terms of N<sub>2</sub> fixation activity, diazotrophs may respond to changes in nutrient availability caused by hydrographic phenomena. To date, research on this aspect of marine diazotrophy has been sparse, especially in the Atlantic as relative to the Pacific. The few studies available do not reveal a clear trend between hydrographic features and N<sub>2</sub> fixation activity trends. For example, Agawin et al. (2013) could not find a relationship between anticyclonic/cyclonic eddies and N<sub>2</sub> fixation rates have been observed in the eddy field downwind of the Canary Islands (Agawin et al., 2013). Ultimately, the study of how physical forcing influences N<sub>2</sub> fixation requires research strategies capable of resolving meso- and submesoscale features.

Another important factor controlling the activity of nitrogen fixers is light. Light provides energy for N<sub>2</sub> fixation and determines the circadian rhythm of autotrophic diazotrophs. In heterocystous diazotrophs, carbon and N<sub>2</sub> fixation can take place concurrently, however non-heterocystous diazotrophs are challenged to protect nitrogenase from photosynthesis-derived oxygen. *Trichodesmium* possesses a photosynthetic apparatus adapted to a high-light regimes and uses a combined approach of high respiration rates and intracellular spatial segregation which allows it to fix both carbon and N<sub>2</sub> during daylight hours (Capone, 1997; Berman-Frank et al., 2007), while UCYN (Zehr et al., 2008) species must perform carbon fixation during daylight, while using the obtained carbohydrates to sustain N<sub>2</sub> fixation at night. As such, light controls the physiology of photosynthetic diazotrophs, while heterotrophic bacterial and archaeal diazotrophs may not be affected and have been shown to express the *nifH* gene independently of light availability (Church et al., 2005). However, the potential circadian rhythms of these diazotrophs is not yet well-understood.

On a greater scale, light levels structure how diazotrophs are distributed in the water column. While *Trichodesmium* usually prefers the top sunlit 75 m, most UCYN tend to appear at deeper depths and down to ~150 m (e.g., Langlois et al., 2008). The lower availability of light at these depths presumably results in a less efficient photosynthesis and hence less evolution of oxygen, thus making it easier for UCYN species to protect their nitrogenase (Karl et al., 2002). Alternatively, UCYN-A presents a more complex situation as this group is not capable of carrying out the oxygen-evolving steps of photosynthesis and because it is thought to be an extracellular symbiont of an eukaryotic algae that does require light for photosynthesis (Zehr et al., 2008; Thompson et al., 2012). While UCYN-A or *Crocospaera* can



appear at deeper levels of the water column than *Trichodesmium*, some studies have shown that their abundance still tends to be greater at the surface (Luo et al., 2012). Finally, the statistical analysis of global N<sub>2</sub> fixation data and diazotroph abundance data indicates that solar radiation is the most influencing factor determining diazotrophic activity, and explains why the highest N<sub>2</sub> fixation rates and abundances of diazotrophs have been reported for the tropical and subtropical bands of the oceans, which also receive the greatest amounts of light worldwide (Luo et al., 2014). However, it must be borne in mind that different diazotroph phylotypes have different responses to daily fluctuations in irradiance (e.g., Church et al., 2005). Finally, the presence and activity of heterotrophic diazotrophs in aphotic waters (e.g., Hewson et al., 2007; Bonnet et al., 2013; Rahav et al., 2013) challenges this strict control and defines physiological differences among diazotrophic species.

### Nutrient and Trace Metal Controls

Fertilization of the photic layer of the oceans depends principally on upwelling, Ekman pumping, mixing, fronts, convection (in polar and temperate regions mainly) and vertical diffusion of deeper waters rich in remineralized nutrients, the remineralization efficiency on those deeper waters, and finally on how the aforementioned physical processes are constrained by the formation and transport of mode waters (Palter et al., 2005). Other external sources include atmospheric deposition, and internal sources include *in situ* regeneration processes like photic layer nitrification or dissolved organic matter (DOM) breakdown into bioavailable molecules through remineralization and photobleaching processes (e.g., Hedges, 2002; Yool et al., 2007). The NA is largely composed of a subtropical gyre where nitrate and phosphate concentrations are low (Figures 1A,B). This gyre is flanked by more nutrient-rich areas such as the Northwest African upwelling system, the Equatorial upwelling region, the Amazon River plume, the Gulf Stream, and the subpolar gyre. These nutrient-rich flanks fertilize the oligotrophic central gyre via advection and epipycnal (along isopycnals) mixing (Pelegri et al., 2006). The NA subtropical gyre differs from others in that it receives the greatest quantity of desert dust worldwide, which in principle alleviates iron limitation in diazotrophs (Jickells, 2005). The extent to which nutrients and trace metals control N<sub>2</sub> fixation vary depending

on the diazotroph phylotype, their physiology and nutrient requirements (reviewed in Sohm et al., 2011c). These are trends that at present are only fairly clear for *Trichodesmium* (Bergman et al., 2012). In this section we explore how the distribution of nutrients and the input of aeolian trace metals controls N<sub>2</sub> fixation and shapes the distribution of different diazotrophic phylotypes in the NA.

### Inorganic and Organic Nutrients Driving the Distribution of Diazotrophic Activity

N<sub>2</sub> fixation provides a source of nitrogen that is unaccompanied by a concomitant input of phosphorus, eventually resulting in increased N:P ratios (with respect to the canonical Redfield ratio of N:P = 16:1; Redfield et al., 1963). This imbalance creates a fixed nitrogen excess signal in the NA (Redfield et al., 1963; Gruber and Sarmiento, 1997), which has been attributed to enhanced N<sub>2</sub> fixation activity driven by the high aeolian iron supply (Michaels et al., 1996, 2001; Gruber and Sarmiento, 1997). As aforementioned, iron plays a key role in controlling diazotrophic activity and diversity in the ocean (see also Section Responses to Atmospheric Dust Inputs). This N:P imbalance results in a severe phosphorus deficiency in the NA basin, a feature which is remarkably different from the other basins of the world's oceans (Wu, 2000; Ammerman et al., 2003). Conversely, high concentrations of dissolved inorganic nitrogen forms (mainly nitrate and ammonium) are usually considered to inhibit diazotrophic activity, given that N<sub>2</sub> fixation is energetically expensive relative to the assimilation of more readily available dissolved inorganic nitrogen forms (Holl and Montoya, 2005).

The comparison of nitrate and phosphate concentrations (Figures 1A,B), with the distribution of N:P ratios (Figure 2A), and that of integrated N<sub>2</sub> fixation rates (Figure 2B) reveals a greater diazotrophic activity in relatively phosphate rich waters (0.1–0.3 μM; Figure 1B) where nitrate is low (<0.5 μM; Figure 1A), leading to low N:P ratios. Indeed, the relationship between phosphate supply or phosphate concentrations and N<sub>2</sub> fixation rates has been documented for different locations of the NA (e.g., Fernández et al., 2012; Benavides et al., 2013d), although this does not hold true in all cases (Turk et al., 2011), because the response of diazotrophs to phosphate limitation may vary geographically and with each diazotroph phylotype

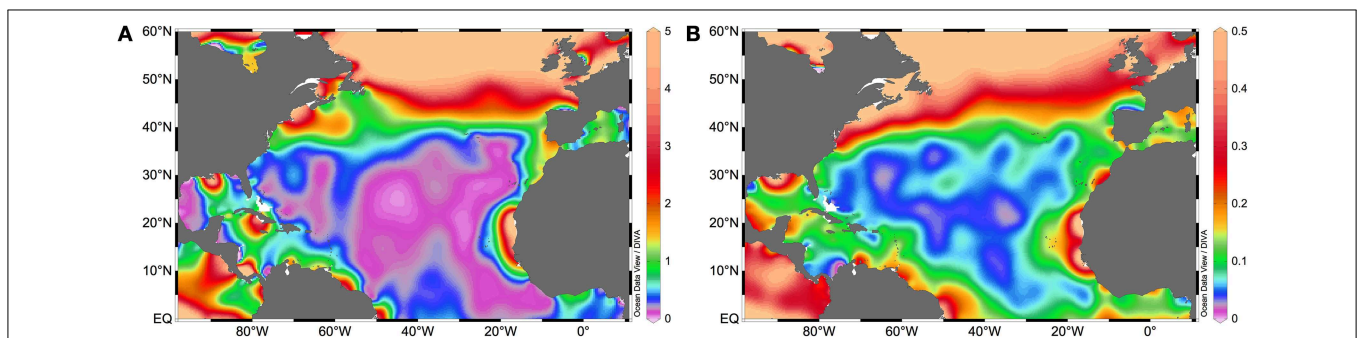
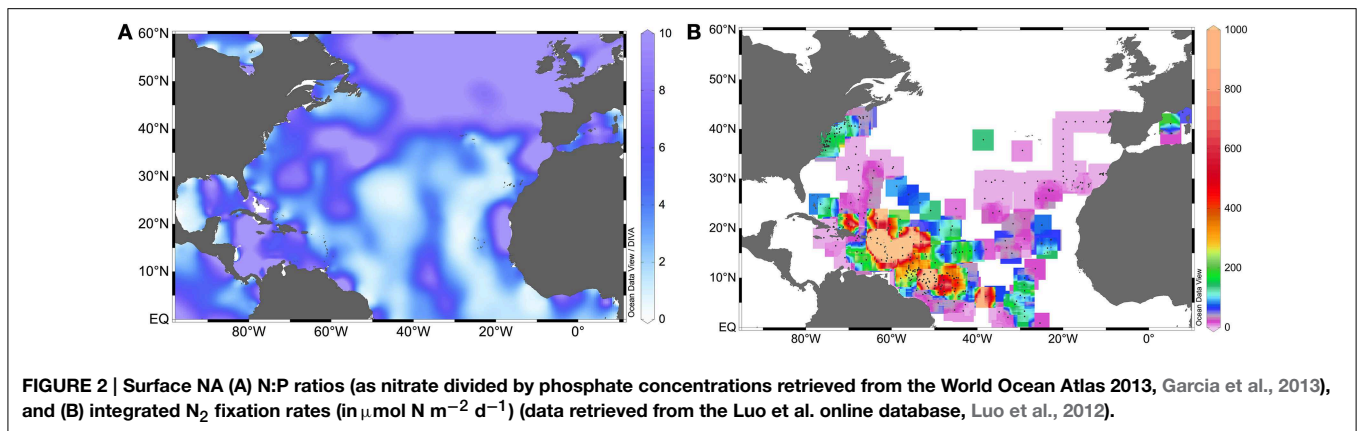


FIGURE 1 | NA surface (A) nitrate and (B) phosphate concentrations (in μM). Data retrieved from the World Ocean Atlas 2013 Garcia et al., 2013.

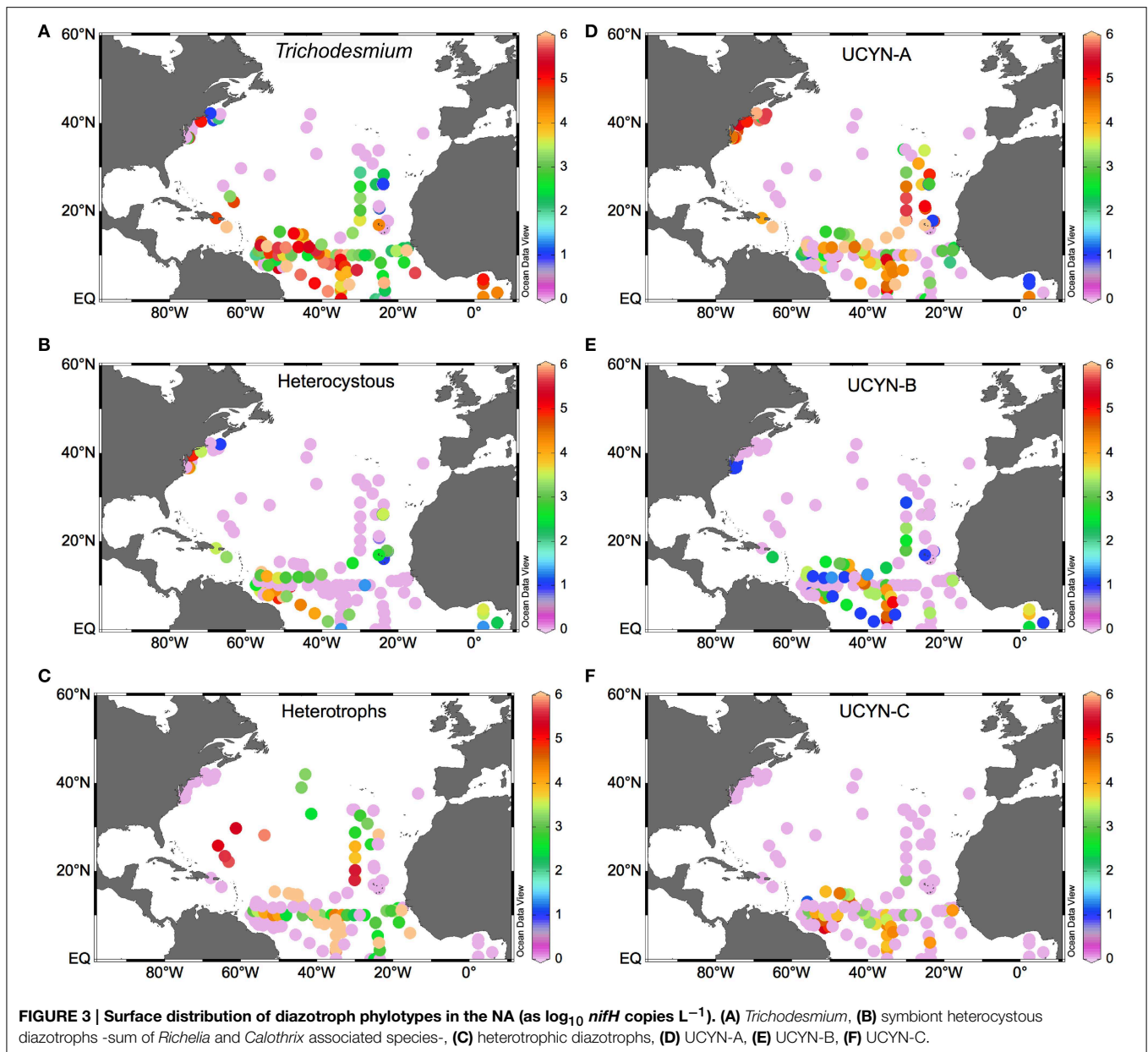


(Turk-Kubo, 2012). In sites of nitrate and concomitant phosphate loading such as the Amazon River plume in the NWA or the Equatorial upwelling on the NEA, N<sub>2</sub> fixation is thought to be fueled by excess phosphate remaining after the consumption of nitrate by non-N<sub>2</sub> fixing phytoplankton (Subramaniam et al., 2008, 2013). As such, diazotroph abundances peak where nitrate is exhausted but there remains enough phosphate to fuel N<sub>2</sub> fixation, such as in the mesohaline waters (salinities 30–35) of the Amazon River plume (Subramaniam et al., 2008). Integrated N<sub>2</sub> fixation rates associated with the plume and East of the Caribbean Sea easily reach values  $>1000 \mu\text{mol N m}^{-2} \text{d}^{-1}$  (Figure 2B). Within the plume, DDAs dominate (mainly *Richelia-Rhizosolenia* symbioses which benefit from excess silica also provided by the river), closely followed by *Trichodesmium*. UCYN phylotypes are found where the plume meets the saltier waters of the open ocean (Foster et al., 2007). Similarly, in the NEA where the Congo River plume meets the Equatorial upwelling -note that the mouth of the Congo is in the southern hemisphere, but its plume affects the NA- and phosphate concentrations are relatively high ( $0.18 \mu\text{M}$ ; Subramaniam et al., 2013), the abundances of DDAs and *Trichodesmium* are greatest within the plume and at the edge of the upwelling, while UCYN-A abounds within the area influenced by the upwelling (Foster et al., 2009). In general, high abundances of *Trichodesmium* are observed down to 75 m depth (Luo et al., 2012), and their geographical extent ranges from the waters adjacent to the Amazon plume to the eastern NA coast along the Equator.

Away from relatively phosphate-rich waters, active diazotroph assemblages in the inner oligotrophic subtropical gyre are likely sustained by phosphate transferred vertically (diapycnal mixing), transported along isopycnals (epiypcnal mixing), or advected from the surrounding gyre flanks (Pelegri et al., 2006), such as excess phosphate from the Gulf Stream (Palter et al., 2011). Alternatively, dissolved organic phosphorus transported to the interior of the gyre from the productive NEA coastal and Equatorial upwelling sites (Mather et al., 2008; Reynolds et al., 2014) may play an important role in supporting N<sub>2</sub> fixation (Mahaffey et al., 2014). The ability of the NEA coastal upwelling sites to partly sustain autotrophic and heterotrophic production within the subtropical gyre has been widely recognized. For example, the Canary Current

upwelling exports  $\sim 60\%$  of production as dissolved organic matter (DOM) (Álvarez-Salgado et al., 2007), while dissolved organic nitrogen and dissolved organic phosphorus are estimated to support 40–70% of export production within the gyre (Torres-Valdés et al., 2009; Reynolds et al., 2014). The role of DOM in sustaining N<sub>2</sub> fixation is much less known. In the tropical and subtropical NA phosphate concentrations are too low to sustain *Trichodesmium* growth using this compound as the solely source of phosphorus (Sohm and Capone, 2006). Indeed, molecular biology approaches have demonstrated that *Trichodesmium* is capable of taking up dissolved organic phosphorus in the form of phosphomonoesters via alkaline phosphatase activity (Dyhrman et al., 2002), or as phosphonates via a carbon-phosphorus bond (C-P) lyase pathway (Dyhrman et al., 2006). The only other marine diazotroph that has been studied in terms of their dissolved organic phosphorus acquisition capability is *Crocospaera watsonii*, which does not possess the genes to utilize phosphonates, but it does have the genes to hydrolyze phosphomonoesters, as well as a suite of genes for inorganic phosphorus transport (high-affinity and low-affinity phosphate transport genes) (Dyhrman et al., 2006). Despite these differences in dissolved organic phosphorus uptake capability, whether the differential distribution of *Trichodesmium* vs. *Crocospaera* responds to phosphomonoesters and phosphonates distributions in the NA is unknown, a fact further worsened by the dearth of knowledge regarding the composition of the dissolved organic phosphorus pool in the ocean (Karl and Björkman, 2002; Kononova and Nesmeyanova, 2002).

Within the NA subtropical gyre, UCYN-A and *Trichodesmium* are the most abundant phylotypes (Figure 3), although they appear to be spatially segregated (Montoya et al., 2007; Agawin et al., 2014; Ratten et al., 2014). While UCYN occupy a wide latitudinal and longitudinal range (Figures 3C,D), *Trichodesmium* is constrained to  $<30^\circ\text{N}$  in the NWA, with the exception of the data reported by Mulholland et al. on the US East Coast (Mulholland et al., 2012, Figure 3A). Indeed, most of the N<sub>2</sub> fixation in the NWA is supported by *Trichodesmium*, and a greater contribution by UCYN phylotypes is found in the NEA (Montoya et al., 2007). UCYN-C is less abundant than the other UCYN and appears on a narrower latitudinal range ( $<20^\circ\text{N}$ ),



mostly over the NWA (Figure 3F). Nevertheless, it must be borne in mind that matching N<sub>2</sub> fixation activity to the presence on diazotrophs based solely on *nifH* abundance as determined by qPCR may lead to misleading results, and *nifH* expression can be high in low abundance phylotypes (Turk-Kubo, 2012).

This spatial segregation may not be straightforward given that the low nitrate-high phosphate (low N:P ratio) conditions responsible for them occur in both the Amazon River plume and in the upwelling-affected areas of the NEA. However, upwelling-affected waters are colder, which likely explains the lower abundance of diazotrophic organisms in the NEA as compared to the NWA (Figure 3), and the lower N<sub>2</sub> fixation rates (generally <1 nmol N L<sup>-1</sup> d<sup>-1</sup>; Moore et al., 2009; Fernández et al., 2010; Benavides et al., 2011). The low N<sub>2</sub> fixation rates

of the NEA do not likely represent an important source of new production in this area (Benavides et al., 2013b). Thus, it appears that the fixed nitrogen requirements of autotrophic planktonic communities in the NEA are met by a combination of atmospheric fixed nitrogen deposition (as this area is located downwind of European industrialized countries), *in situ* remineralization of advected DOM and, to a minor extent, *in situ* photic nitrate regeneration (nitrification) and N<sub>2</sub> fixation. However, the presence of N<sub>2</sub> fixation activity and/or diazotrophic microorganisms in the cold and nutrient rich waters of the NA such as upwelling sites (Voss et al., 2004; Foster et al., 2009; Benavides et al., 2011; Subramaniam et al., 2013; Agawin et al., 2014) or mesopelagic waters (Hewson et al., 2007) has been observed, though their ecological role in these environments is

unclear and warrants further study. High dissolved inorganic nitrogen environments have been usually regarded as inhibitory for diazotrophic activity (Holl and Montoya, 2005). However, this effect is not clear. Recent culture and field studies indicate that diazotrophy can continue upon exposure to dissolved inorganic nitrogen (nitrate and ammonium) as long as phosphate is available (reviewed in Knapp, 2012). Furthermore, active N<sub>2</sub> fixation in dissolved inorganic nitrogen-rich environments suggests that restricting diazotrophy studies to the upper ocean layer of the tropics and subtropics can result in large underestimations of N<sub>2</sub> fixation rates (Raimbault and Garcia, 2008; Benavides et al., 2011; Sohm et al., 2011b; Bonnet et al., 2013). Foster et al. (2009) found UCYN-A over the Equatorial upwelling region in the Gulf of Guinea. Although these authors did not find a high level of *nifH* expression in UCYN-A, in a later study Subramaniam et al. (2013) reported N<sub>2</sub> fixation rates between 15 and 424 μmol N m<sup>-2</sup> d<sup>-1</sup> for the same site, with a ~50% contribution of the <10 μm fraction presumably including UCYN-A. Similarly, in the upwelling sites of Cape Silleiro (Northwest Iberian peninsula) and Cape Ghir (Northwest Africa), the <10 μm fraction contributed 60–90% of total N<sub>2</sub> fixation, although the rates were much lower (Benavides et al., 2011). Finally, this group was found in high abundances along the East Coast of the US where it occurred at high dissolved inorganic nitrogen concentrations and comparatively low temperatures (Mulholland et al., 2012). Relatively high abundances of DDAs have been also reported off the eastern US coast, which could be due to actual *in situ* growth or as a result of transport from Amazon River plume affected waters along the Gulf Stream (Ratten et al., 2014).

In summary, N:P ratios seem to shape the distribution of diazotrophs and their associated N<sub>2</sub> fixation rates, while the phosphorus requirements of diazotrophs could be met by direct uptake of dissolved organic phosphorus or hydrolysis of these compounds into inorganic phosphorus (however note that UCYN-A does not possess the genes needed to use dissolved organic phosphorus; Tripp et al., 2010). *Trichodesmium* needs low N:P ratios, UCYN can stand higher dissolved inorganic nitrogen concentrations, DDAs need low N:P accompanied by sufficient silica, and heterotrophic diazotrophs seem to be more flexible and better able to exploit the DOM accumulated in the inner subtropical gyre.

When dividing the NA into its western (80–40°W) and eastern (40–0°W) sides, and considering three latitudinal bands (Tropic 0–23.5°N, Subtropic 23.5–40°N, and Temperate >40°N), the distribution of *nifH* copies per liter of different diazotroph phylotypes show interesting patterns (based on the database of Luo et al., 2012; **Table 1**—note that the number of data points is strikingly different between latitude/longitude ranges-). The dominance of *Trichodesmium* in the tropical and subtropical NWA becomes evident, with one order of magnitude higher abundances than its eastern counterpart. Heterocystous diazotrophs are also clearly dominant in the NWA due to the influence of the Amazon River plume. The distribution of heterotrophic diazotrophs is more disparate, with two orders of magnitude higher abundance in the tropical NEA than in the NWA, but three orders of magnitude higher

**TABLE 1 | Average ± standard deviation *nifH* copies L<sup>-1</sup> of different diazotroph phylotypes as determined by qPCR.**

	West (80–40°W)					East (40–0°W)						
	<i>Trichodesmium</i>	DDAs	Heterotrophs	UCYN-A	UCYN-B	UCYN-C	<i>Trichodesmium</i>	DDAs	Heterotrophs	UCYN-A	UCYN-B	UCYN-C
Tropic (0–23.5°N)	1.4 ± 3.4 × 10 <sup>5</sup>	4.3 ± 21 × 10 <sup>5</sup>	81 ± 320	1.1 ± 4.7 × 10 <sup>7</sup>	2.6 ± 16 × 10 <sup>4</sup>	6.9 ± 24 × 10 <sup>3</sup>	7.1 ± 23 × 10 <sup>4</sup>	84 ± 330	9.9 ± 41 × 10 <sup>2</sup>	4.4 ± 13 × 10 <sup>4</sup>	4.3 ± 21 × 10 <sup>3</sup>	1.8 ± 4.6 × 10 <sup>3</sup>
Subtropic (23.5–40°N)	4.4 ± 23 × 10 <sup>3</sup>	1.1 ± 3.2 × 10 <sup>4</sup>	1.0 ± 2.5 × 10 <sup>3</sup>	2.1 ± 7.7 × 10 <sup>6</sup>	47 ± 51	0	1.3 ± 3.2 × 10 <sup>2</sup>	2.0 ± 6.7 × 10 <sup>2</sup>	18 ± 97	1.8 ± 7.2 × 10 <sup>4</sup>	49 ± 260	0
Temperate (>40°N)	1.0 ± 3.0 × 10 <sup>4</sup>	3.2 ± 9.7 × 10 <sup>2</sup>	6.9 ± 11 × 10 <sup>2</sup>	2.7 ± 3.3 × 10 <sup>5</sup>	0	0	n/a	n/a	n/a	n/a	n/a	n/a

Data compiled from the Luo et al. database (Luo et al., 2012).  
n/a indicates no data available.



abundance in the subtropical NWA than in the NEA (Table 1). It is perhaps remarkable that heterotrophic diazotrophs in temperate waters are only one order of magnitude lower than in subtropical waters of the NWA. Nevertheless, the abundance of heterotrophic diazotrophs needs to be interpreted with great caution as their diversity is less explored than that of UCYN and contamination problems with PCR reagents are well known (Zehr et al., 2003). UCYN-A is also more abundant in the NWA as compared to the NEA. Their average abundances decrease by one order of magnitude by latitudinal band in the NWA, and surpass the average abundances of the NEA by more than two orders of magnitude. UCYN-B seem to be more homogeneously distributed, with similar abundances in the tropical and subtropical NEA and NWA, and practically absent at temperate latitudes. This seems also to be the case of UCYN-C, which is however constrained to tropical latitudes. It should be noted that the NWA has been more extensively researched in terms of diazotrophic activity and diversity than the NEA, and therefore these averages could be biased by undersampling. This is clearly reflected by the generally higher standard deviation values of the NEA average *nifH* abundances, as well as by the number of data included for these analyses (NWA = 136 data points, NEA = 30 data points).

### Responses to Atmospheric Dust Inputs

The nitrogenase enzyme consists of an iron subunit (dinitrogenase reductase), and an iron-molybdenum subunit (dinitrogenase) (Postgate, 1982), which makes iron an essential metal for diazotrophs. Due to the low solubility of iron in the oxidized and slightly basic contemporary ocean (Liu and Millero, 2002), planktonic microorganisms have developed specialized iron scavenging mechanisms to provide them with the iron needed for the functioning of many photosynthetic and respiratory redox enzymes. For example, conspicuous pelagic diazotrophs like *Trichodesmium* and *Crocospaera* express the IdiA protein in response to iron stress (Webb et al., 2001). Pioneering studies showed that the addition of iron enhanced growth and N<sub>2</sub> fixation rates in cultured and field populations of *Trichodesmium* (Rueter, 1988; Paerl et al., 1994). Later, the cellular iron quotas of *Trichodesmium* were reported to be very high (Berman-Frank et al., 2001), with iron requirements during diazotrophic growth found to be as much as five-fold greater than when grown on ammonium (Kustka et al., 2003). When grown under moderate iron deficiency, cultured *Crocospaera* responds by reducing its cellular size to overcome the limitation of growth and N<sub>2</sub> fixation, while severe iron deficiency leads to disruption of the diazotrophic activity (Jacq et al., 2014). The release of extracellular polysaccharides has been hypothesized as an iron binding strategy in *Crocospaera* (Sohm et al., 2011a), and in unidentified pico- and nano-sized diazotrophs of the NEA (Benavides et al., 2013c). UCYN-A increases *nifH* expression upon the addition of Fe or Saharan dust (Turk-Kubo, 2012; Krupke et al., 2014), suggesting that its N<sub>2</sub> fixation activity may be partially controlled by dust deposition events, the chemical processes by which dust-derived Fe is dissolved into seawater, as well as any Fe-binding strategies of these microbes may have (Benavides et al., 2013c).

The biochemical mechanisms by which diazotrophs obtain iron *in situ* have yet to be fully characterized, but several experiments have confirmed that *in situ* N<sub>2</sub> fixation activity responds to iron inputs. For example, Mills et al. (2004) reported a two- to three-fold increase of N<sub>2</sub> fixation rates upon iron and phosphorus enrichment of seawater samples collected in the tropical NA. Langlois et al. (2012) observed an equivalent enhancement of N<sub>2</sub> fixation rates in response to Saharan dust additions, which was also accompanied by an increase in the abundance of *nifH* phylotypes. In accordance with these artificially fertilized bioassays, natural peak inputs of Saharan dust to the NEA have been observed to increase N<sub>2</sub> fixation rates and UCYN abundance by ~90–~60%, respectively (Benavides et al., 2013c). Dust inputs may also affect diazotrophic communities in areas distant from the NEA. For example, sporadic peaks in *Trichodesmium* abundance are often observed off Florida as a response to dust deposition (Lenes et al., 2001). This phenomenon produces enough dissolved organic nitrogen to promote toxic phytoplankton blooms (Lenes and Heil, 2010). These results confirm that N<sub>2</sub> fixation reacts to dust and/or iron additions in time scales of hours to days.

The NA receives more desert dust than any other basin in the world, providing an estimated iron deposition of 6 Tg Fe y<sup>-1</sup> (Prospero et al., 1996). Despite this great supply, the potential bioavailability of the iron deposited with dust is not well-understood (Mahowald et al., 2009). Significant correlations between dissolved iron concentrations, dust deposition rates (or dust deposition proxies like aerosol optical depth at 550 nm) and N<sub>2</sub> fixation rates or diazotroph phylotypes abundance have been recurrently reported in the NA (e.g., Moore et al., 2009; Fernández et al., 2010; Benavides et al., 2013d; Ratten et al., 2014). In addition, it is now clear that the shifts in the geographical extent of coverage of the intertropical convergence zone (ITCZ) controls iron inputs and hence N<sub>2</sub> fixation activity in the tropical and subtropical Atlantic (Schlosser et al., 2013). This “iron-rich cloud” acts as a frontier between the phosphorus-rich/iron-poor South Atlantic, and the phosphorus-poor/iron-rich NA (Capone, 2014).

Despite the evident control of iron on N<sub>2</sub> fixation in the NA, it is unclear why the North Pacific has comparable N<sub>2</sub> fixation rates given the negligible aeolian iron inputs to this basin (Mahowald et al., 2009). The predominance of different diazotroph phylotypes in the North Pacific and the North Atlantic has been hypothesized as an explanation for these differences (Deutsch et al., 2007), although the *nifH* sequences of different phylotypes isolated from the NA seem to be highly similar to those from the North Pacific (Langlois et al., 2005). As Luo et al. suggested (2014), other explanations include the saturation of N<sub>2</sub> fixation by present iron supply, the adaptation of diazotrophs to low iron environments, and/or the biological unavailability of the iron deposited to the ocean. These authors also reported a poor relationship between N<sub>2</sub> fixation and dust deposition from a global database (Luo et al., 2012), suggesting that iron is not the ultimate controlling factor of marine diazotrophy on a global basis (Luo et al., 2014). This implies that while empirical evidence suggests dust explains short-term diazotrophic responses, it cannot explain N<sub>2</sub> fixation rates on a

basin-scale or over geological time-scales in the NA (Straub et al., 2013).

## N<sub>2</sub> Fixation Contribution to New Production and Nitrogen Excess in the North Atlantic

Diazotrophic N<sub>2</sub> fixation has been classically regarded as an important contribution to new production in chronically stratified oligotrophic areas of the oceans where the upwelling of deep nitrate rich waters is inhibited (Karl et al., 2002). Several studies in the NA have compared the upward diffusive flux of nitrate to <sup>15</sup>N-labeled nitrate uptake rates. In most cases, nitrate uptake is greater than nitrate diffusion (e.g., Planas et al., 1999 see **Table 2**), a difference attributed to external fixed nitrogen inputs via atmospheric deposition, *in situ* remineralization of advected DOM, photic zone nitrification and/or N<sub>2</sub> fixation (Capone et al., 2005; Landolfi et al., 2008; Mouriño-Carballido et al., 2011; Benavides et al., 2013b). The comparison of nitrate diffusive fluxes to *in situ* N<sub>2</sub> fixation rates has resulted in a wide range of results (**Tables 2, 3**).

Capone et al. (2005) calculated that N<sub>2</sub> fixation rates ranged from 50 to 180% of the parallel measured nitrate diapycnal

fluxes in the tropical NWA, suggesting the major influence of diazotrophy in sustaining new production over this area. In the central subtropical and equatorial NA (~30 °W), Mouriño-Carballido et al. (2011) reported a contribution of N<sub>2</sub> fixation to total new nitrogen inputs to the mixed layer of 22% (12°S to 16°N) and 2% (16 to 9°N). More recently, Painter et al. (2013) reported a contribution of 62% in an area north of that studied by Mouriño-Carballido et al. (26.5°N–29.5°W) 2011.

Differences between these estimates may arise from a variety of factors, including the geographical variability of hydrographic factors. The transport of nitrate into the surface layer occurs in two different ways, by advection, upwelling, or turbulent diffusion. Upwelling is caused by a vertically upward velocity (which is driven in general by the curl of the wind stress; Yoshida, 1967), or horizontal deformation fields in velocity (e.g., divergence or stretching deformation; Dippner et al., 2007). Turbulent diffusion depends on the gradient of the nitrate concentration and the characteristics of the turbulence field such as the vertical shear of the velocity field and the stability of the water column given by the vertical density gradient. The intensity of the turbulence and the depth of mixed layer are driven by the wind speed (Mellor and Yamada, 1982). Thus, nitrate uptake by phytoplankton is not directly coupled to N<sub>2</sub> fixation, but the

**TABLE 2 | Compilation of nitrate diapycnal flux data in the NA.**

Location	$\mu\text{mol N m}^{-2} \text{d}^{-1}$	Tg N y <sup>-1</sup>	References
Subtropical NA ("Beta-Triangle": 26.5°N × 38.5°W, 32.5°N × 30.0°W, 22.5°N × 28.5°W)	1644	329.3	Jenkins, 1982
Whole NA	100	20.0	Michaels et al., 1996
NA (anticyclonic eddy at 60°N)	1250	250.4	Law et al., 2001
Central Atlantic Ocean (35°S–28°N)	380–838	76.1–167.9	Planas et al., 1999
Equatorial NA	137	27.4	Oschlies, 2002
Subtropical and tropical NWA	46–736	9.2–147.4	Capone et al., 2005
Subtropical NA (Canary Current-North Equatorial Current)	270	54.1	Dietze et al., 2004
Subtropical NA (WOCE section A05; 24.5°N)	860	172.3	Bahamón et al., 2003
Subtropical NEA (Eddy field southern Canary Islands)	1710	342.5	González-Dávila et al., 2006
Subtropical NEA (Azores Current-Subtropical Front)	10	2.0	Mouriño et al., 2004
Central Atlantic Ocean (31°S–16°N)	840	168.3	Mouriño-Carballido et al., 2011
Subtropical NEA (Northwest African upwelling)	20–180	4–36.1	Arcos-Pulido et al., 2014

**TABLE 3 | List of integrated N<sub>2</sub> fixation rates from the NA, compiled in Luo et al. Luo et al., 2012.**

Location	$\mu\text{mol N m}^{-2} \text{d}^{-1}$	Tg N y <sup>-1</sup>	References
Subtropical NEA (Canary Current and Northwest African upwelling)	0.001–0.09	0–0.02	Benavides et al., 2011
Subtropical NEA (Sargasso and Caribbean Seas)	0.05–540	0–108	Carpenter and Price, 1977
Subtropical and tropical NWA	0.03–19235	0.01–3853	Capone et al., 2005
Tropical NWA	73–90	15–18	Falcón et al., 2004
Tropical and subtropical NA	1.2–298	0.2–60	Fernández et al., 2010
Tropical NWA	4.5–68.1	0.9–14	Goering et al., 1966
NWA	0.1–0.9	0.02–0.2	McCarthy and Goldman, 1979
Temperate, tropical and subtropical NA	1.8–182	0.4–36.4	Moore et al., 2009
Northeast American coast (Cape Hatteras–Georges Bank)	2.4–532	0.5–107	Mulholland et al., 2012
Tropical NEA	28–142	8–29	Turk et al., 2011
Tropical NEA	56–60	11–12	Turk-Kubo, 2012

enhanced stability of the water column in the NWA as compared to the NEA translates into a lessened nitrate diffusion, which eventually makes the contribution of N<sub>2</sub> fixation to total fixed nitrogen inputs greater than that derived from nitrate diffusion (Capone et al., 2005; Montoya et al., 2007).

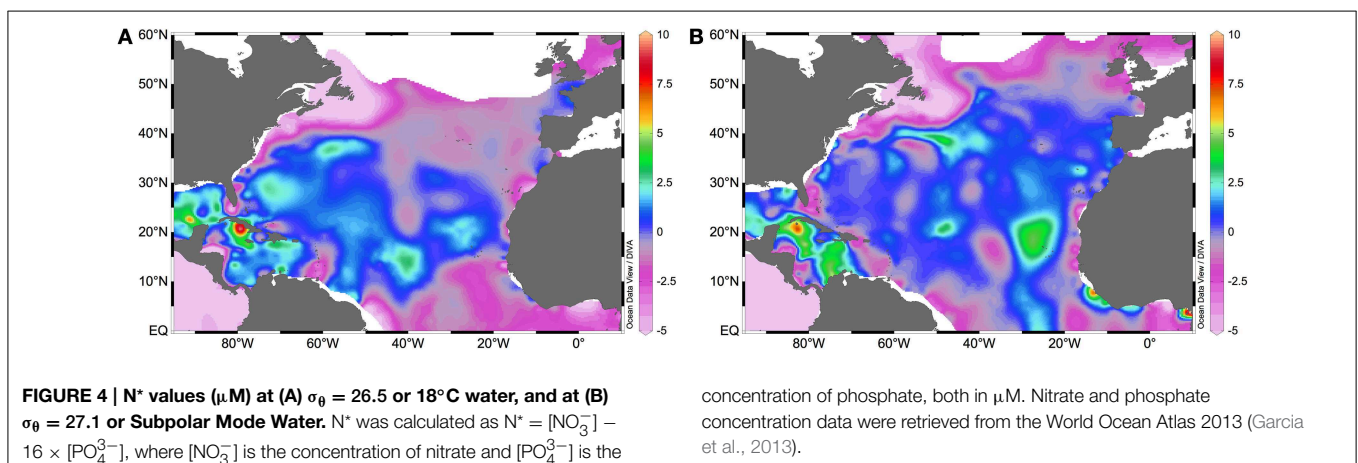
In addition, the inherent seasonal variability of the NA, and the intermittent nature of mesoscale and submesoscale features may result in a highly variable contribution of N<sub>2</sub> fixation to new production (Mouriño-Carballido et al., 2011; Painter et al., 2013). Finally, other factors such as the application of theoretical vs. empirical estimated turbulent diffusion coefficients ( $K_z$ ) (Mouriño-Carballido et al., 2011; Arcos-Pulido et al., 2014), biases derived from measuring N<sub>2</sub> fixation on sporadic *Trichodesmium* blooms (which might be not be representative of the long-term situation; Capone et al., 2005), <sup>15</sup>N<sub>2</sub> method underestimations (Mohr et al., 2010), or overestimations (Dabundo et al., 2014 see discussion below), and substantial nitrate regeneration in the photic zone (Yool et al., 2007), may also play an important role.

The N<sub>2</sub> fixation rates reported in these studies are strikingly different. Capone et al. (2005) reported *Trichodesmium*-associated integrated N<sub>2</sub> fixation rates averaging 239 μmol N m<sup>-2</sup> d<sup>-1</sup> (but up to 19235 μmol N m<sup>-2</sup> d<sup>-1</sup>, see **Table 3**), while Mouriño-Carballido et al. (2011) and Painter et al. (2013) reported bulk seawater N<sub>2</sub> fixation rates ranging from 1.3 to 154.5 μmol N m<sup>-2</sup> d<sup>-1</sup> and 15.4 to 95.6 μmol N m<sup>-2</sup> d<sup>-1</sup>, respectively. Thus, it is clear that the contribution of N<sub>2</sub> fixation to new production is greater in *Trichodesmium*-dominated waters of the NWA (Montoya et al., 2007), whereas minimal in the UCYN-dominated waters of the NEA (Benavides et al., 2013b; Arcos-Pulido et al., 2014).

Although N<sub>2</sub> fixation has traditionally been regarded as an important source of new production, little is known about the fate of this fixed N<sub>2</sub> or to what extent it fuels the biological carbon pump (Mulholland, 2007). The efficiency of fixed N<sub>2</sub> transfer between trophic compartments and the mechanisms by which it is governed remain unclear (Mulholland, 2007), and whether the process is more efficient in *Trichodesmium* vs. UCYN or heterotrophic diazotrophs is yet to be determined. In principle, the transfer of fixed N<sub>2</sub> should be highly efficient

because the product of this reaction is ammonium, which is readily available to photoautotrophic microbes. *Trichodesmium* also releases large amounts of fixed N<sub>2</sub> as labile dissolved organic nitrogen (Capone et al., 1994; Sipler et al., 2013), and the role of UCYN in providing these organic compounds could also be important (Benavides et al., 2013d). Modern high-resolution mass spectrometry techniques such as nano-scale secondary ion mass spectrometry (nanoSIMS) have recently been used to yield new insights into the efficiency of this transfer and the partitioning of fixed N<sub>2</sub> between planktonic groups in different trophic levels. Symbiotic diazotrophs represent clear examples of fixed N<sub>2</sub> transfer, like UCYN-A, which has been found to provide fixed N<sub>2</sub> to its prymnesiophyte host from whom it receives carbon compounds (Thompson et al., 2012). This transfer has also been demonstrated in DDAs (Foster et al., 2011). However, the fate of N<sub>2</sub> fixed by other non-symbiotic diazotrophs is unknown.

From a geochemical perspective, the recognition of biological N<sub>2</sub> fixation as a significant input of fixed nitrogen to the NA has also been sustained by the recurrent observation of an excess fixed nitrogen signal over the intermediate waters of this basin (Michaels et al., 1996; Gruber and Sarmiento, 1997). The role of this excess in counterbalancing fixed nitrogen losses in oxygen minimum zones (OMZs) has been repeatedly discussed in the literature (e.g., Codispoti, 2007), and has even prompted improvements of the <sup>15</sup>N<sub>2</sub> fixation method that can potentially reconcile geochemical and biological estimates of N<sub>2</sub> fixation in the ocean (Großkopf et al., 2012). Low phosphate availability can be driven by either diazotrophic phosphate-drawdown boosted by high iron availability in mid to low latitudes (Wu, 2000; Mather et al., 2008; Sohm and Capone, 2010) or by low non-Redfieldian (N:P <16) fast-growing phytoplankton nutrient uptake ratios at higher latitudes (Mills and Arrigo, 2010). Both N<sub>2</sub> fixation and non-Redfieldian nutrient uptake patterns leave a fixed nitrogen excess signal (depicted by the N\* parameter) which is maximal in mode waters (isopycnal surface  $\sigma_\theta = 26.5$  or 18°C water, and  $\sigma_\theta = 27.1$  or Subpolar Mode Water (SPMW) (e.g., Gruber and Sarmiento, 1997; Hansell et al., 2004), as shown in **Figure 4**. This excess nitrogen signal has been translated into basin-wide N<sub>2</sub> fixation rates ranging from ~5



**TABLE 4 | Estimates of N<sub>2</sub> fixation using a geochemical approach (N\*) for different areas of the North Atlantic Ocean.**

Method	Area considered (10 <sup>6</sup> km <sup>2</sup> )	Tg N y <sup>-1</sup>	References
$N^* = [\text{NO}_3^-] - 16 \times [\text{PO}_4^{3-}] + 2.72$	7–19	51–89.6	Michaels et al., 1996
$N^* = ([\text{NO}_3^-] - 16 \times [\text{PO}_4^{3-}] + 2.9) \times 0.87$	28	28	Gruber and Sarmiento, 1997
$\text{DIN}_{\text{XS}} = [\text{NO}_3^-] - 16 \times [\text{PO}_4^{3-}]$	6.83	4.3	Hansell et al., 2004
$\text{DIN}_{\text{XS}} = [\text{NO}_3^-] - 16 \times [\text{PO}_4^{3-}]$	16	8–59.2	Bates and Hansell, 2004
$\text{DIN}_{\text{XS}} = [\text{NO}_3^-] - 16 \times [\text{PO}_4^{3-}]$	16	5.6	Hansell et al., 2007
$N^* = [\text{NO}_3^-] - 14.63 \times [\text{PO}_4^{3-}]$	27.8	16.8	Singh et al., 2013

to 90 Tg N y<sup>-1</sup> (Table 4) for the NA tropical and subtropical waters.

The rates estimated by Michaels et al. (1996) are four- to seven-fold higher than the most recent direct estimate rate (12.8 Tg N y<sup>-1</sup>; see the Supplementary Information of Großkopf et al., 2012), while the estimates of Hansell et al. (2004, 2007) are more conservative. However, it must be noted that these estimations rely on the extrapolation of nitrate excess formation rates over different areas of the NA (Table 4). In a recent paper, Singh et al. (2013) revisited the magnitude and distribution of N\* in the NA taking into account other factors causing positive N\* values. They calculated a basin-wide nitrate excess rate formation of 16.8 Tg N y<sup>-1</sup>, which is reasonably close to the average direct N<sub>2</sub> fixation rate calculated by Großkopf et al. (2012). N<sub>2</sub> fixation rates derived from N\* estimates are potentially overestimated by non-diazotrophic positive N\* signatures such as atmospheric deposition and of high N:P ratio compounds, and lateral advection of high N:P DOM (Landolfi et al., 2008; Letscher et al., 2013; Singh et al., 2013). Also, the high N:P uptake quotas typical of slow-growing picocyanobacteria which dominate subtropical gyres (i.e. *Prochlorococcus*) may reduce fixed nitrogen availability promoting diazotrophy which eventually produces a nitrogen excess signal in the thermocline of the NA subtropical gyre via (Mills and Arrigo, 2010). As Singh et al. (2013) stated, the N\* parameter has to be interpreted with caution, because positive N\* values are caused by a variety of processes other than N<sub>2</sub> fixation. Predominantly, the temporal and spatial scales taken into account when deriving N<sub>2</sub> fixation rates from N\* and from direct <sup>15</sup>N<sub>2</sub> incubations are strikingly different. According to the timescales of thermocline ventilation, present N\* observations likely reflect the N<sub>2</sub> fixation activity that occurred a decade before (Gruber and Sarmiento, 1997). Moreover, positive N\* signals travel downstream from the formation site along isopycnal surfaces (Hansell et al., 2004). As a result, there is a considerable spatial and temporal lag between the process and its signature. The N\* signature is undoubtedly affected by atmospheric nitrogen deposition (which usually has high N:P; Duce et al., 1991), and by non-Redfieldian fluxes and remineralization of organic matter (Zamora et al., 2010; Singh et al., 2013). In turn, N<sub>2</sub> fixation rates derived from <sup>15</sup>N<sub>2</sub> incubations assess the uptake of N<sub>2</sub> in a closed volume of water (usually ranging from hundreds of mL to <5 L), along a limited incubation time (hours to days), in a specific geographical location and water column depth. In summary, while N\* offers an easy and large-scale alternative to evaluate N<sub>2</sub> fixation, it must be

interpreted with great care, taking into account all other possible sources of positive N\*, which might be difficult to constrain.

In general, the rates obtained with geochemical approaches exceed those obtained with biological approaches two to seven-fold (Tables 3, 4), although both types of estimates are highly variable and arguably have considerable uncertainties. This could either mean that geochemical-approach methods overestimate N<sub>2</sub> fixation rates, or alternatively biological-approach methods underestimate (Mohr et al., 2010; Großkopf et al., 2012) or overestimate them (Dabundo et al., 2014). While the underestimation of N<sub>2</sub> fixation rates by incomplete dissolution of the <sup>15</sup>N<sub>2</sub> bubble during short duration incubations generally translates into two to six-fold increases of the rates (Mohr et al., 2010; Großkopf et al., 2012; Wilson et al., 2012; Benavides et al., 2013d), the use of contaminated <sup>15</sup>N<sub>2</sub> gas stocks with other nitrogen compounds besides N<sub>2</sub> (notably ammonium) may result in N<sub>2</sub> fixation rates of 0.01 to 530 nmol N L<sup>-1</sup> d<sup>-1</sup> (as estimated by Dabundo et al., 2014). The upper limit of this broad range is of serious concern, as these rates surpass N<sub>2</sub> fixation rates reported in many marine systems. However, the degree of contamination depends on the brand of <sup>15</sup>N<sub>2</sub> used (Dabundo et al., 2014), and thus not all published studies can be considered to be equally overestimated. Since current geochemical methods are not able to discern N<sub>2</sub> fixation from other processes resulting in nitrogen excess signals, these methods are prone to result in overestimations of N<sub>2</sub> fixation rates. Improvements in the large scale estimation of N\* as well as in the empirical measurement of <sup>15</sup>N<sub>2</sub> fixation rates should bring denitrification and N<sub>2</sub> fixation rates closer in the near future, eventually elucidating a contemporary balanced oceanic nitrogen cycle.

## Crossing Boundaries: Heterotrophic and Aphotic N<sub>2</sub> Fixation

Oceanic N<sub>2</sub> fixation was once attributed solely to the filamentous cyanobacterium *Trichodesmium* (e.g., Capone, 1997). As discussed earlier, the advent of molecular techniques identified the previously unrecognized prevalence of UCYN (Zehr et al., 1998). These methodological approaches have also enabled the detection of non-cyanobacterial diazotrophs (bacteria and archaea), although their role in global marine N<sub>2</sub> fixation has been largely overlooked (Farnelid and Riemann, 2008). Heterotrophic diazotroph *nifH* sequences group into four major clusters, namely Cluster I (including molybdenum nitrogenases



from cyanobacteria,  $\alpha$ -,  $\beta$ -, and  $\gamma$ -proteobacteria, and vanadium nitrogenases from  $\gamma$ -proteobacteria), Cluster II (composed of archaeal nitrogenases and bacterial iron nitrogenases, also including methanogens), Cluster III (comprising anaerobic bacteria and sulfate-reducing  $\delta$ -proteobacteria), and Cluster IV (which includes non-functional archaeal *nifH* homologs). Present estimates indicate that heterotrophic *nifH* sequences account for >80% of the total number of *nifH* clones obtained from marine samples in a range of studies (Farnelid and Riemann, 2008). This high proportion highlights the need to understand the quantitative role of heterotrophic N<sub>2</sub> fixation, although the usual nested PCR approach taken to obtain these clones (Zehr et al., 1998; Zehr and Turner, 2001) is not quantitative and hence the numerical majority of heterotrophic *nifH* derived from these methods is arguable (Riemann et al., 2010; Farnelid et al., 2011; Turk-Kubo et al., 2014). However, inconsistencies between the presence and the expression of their *nifH* genes has led to skeptical points of view concerning their importance and contribution to total N<sub>2</sub> fixation (Turk-Kubo et al., 2014).

The ecological role and the energy sources that maintain heterotrophic N<sub>2</sub> fixation are uncertain. The differences between day and night heterotrophic *nifH* expression are not significant (Church et al., 2005; Moisander et al., 2014), suggesting the activity of these organisms could be independent of light. This opens the possibility of active N<sub>2</sub> fixation below the euphotic zone, challenging the paradigm that N<sub>2</sub> fixation is constrained to sunlit oligotrophic epipelagic waters. On the contrary, the predominance of non-cyanobacterial nitrogenase gene amplicons from surface oceanic waters collected around the world (Farnelid et al., 2011), suggests that heterotrophic diazotrophs may also obtain their energy through photoheterotrophic processes (e. g., Riemann et al., 2010; Moisander et al., 2014). This possibility is also supported by the significant relationship between solar light radiation and euphotic zone N<sub>2</sub> fixation rates obtained worldwide (Luo et al., 2014), as previously hypothesized for the North Pacific by Karl et al. (2008). In any case, photic and aphotic heterotrophic diazotrophs probably belong to different phylotypes and obtain energy through different metabolic strategies.

Heterotrophic metabolisms imply that a source of bioavailable organic matter is needed for growth. At depth, fixed nitrogen concentrations are high and the lability of organic matter is low (Hedges, 2002), which inevitably leads to questions about the ecological benefit of aphotic heterotrophic N<sub>2</sub> fixation. However, aphotic N<sub>2</sub> fixation could well-occur in association with particles. Organic materials leaking from particles as a result of bacterial extracellular enzyme activity may be an important source of sustenance for heterotrophic N<sub>2</sub> fixation (Riemann et al., 2010). In addition, particles may provide unique oxygen-deficient loci where the nitrogenase enzyme would be protected from destruction (Riemann et al., 2010). The pioneering work of Hans Paerl on particle-attached N<sub>2</sub> fixation (Paerl, 1985; Paerl and Prufert, 1987) has, unfortunately, not been continued in modern microbial oceanography research, although some authors are currently working on the nitrogen metabolism of diazotrophic filamentous cyanobacteria aggregates (Klawonn et al., 2015).

Heterotrophic N<sub>2</sub> fixation activity has been confirmed in the coastal OMZ off Peru (Bonnet et al., 2013; Loescher et al., 2014), and in the oxygen-deficient waters of the Baltic Sea chemocline (Farnelid et al., 2013). However, matching rates to diazotroph phylotypes has been difficult (e.g., Turk-Kubo et al., 2014). Moreover, the mechanisms by which these organisms obtain energy are unknown. It has been hypothesized that in such environments, heterotrophic N<sub>2</sub> fixation could be sustained by organic matter oxidation processes like denitrification or sulfate reduction, through the use of electron acceptors such as nitrate, nitrite or sulfate (Bonnet et al., 2013; Loescher et al., 2014).

In comparison with other oceanic basins, the NA OMZ is less pronounced than that of the Southeast Pacific (Stramma et al., 2008a). In principle, this would indicate a minor role for heterotrophic N<sub>2</sub> fixation in the NA OMZ as compared to the Southeast Pacific, unless these heterotrophs rely on oxygen-poor microzones provided by particle-attached modes of living. In the NA, Baltar et al. (2009) observed an equatorward increase in bacterial abundance and activity in the mesopelagic and bathypelagic zones, related to a higher abundance of suspended particles. The potential role of these microhabitats in sustaining pelagic heterotrophic N<sub>2</sub> fixation in the North Atlantic is an unsolved mystery. In the Sargasso Sea, Hewson et al. (2007) detected heterotrophic *nifH* genes in mesopelagic to abyssopelagic waters. Although these authors did not provide parallel N<sub>2</sub> fixation rates, the presence of these phylotypes suggests that nitrogenase activity should be significant there. To our knowledge, there are no other aphotic diazotrophy studies in the NA. In this basin, the dependence of aphotic prokaryotes on particulate organic matter (Baltar et al., 2009) and on nutrient availability controlled by water mass mixing (Reinthal et al., 2013) suggests that heterotrophic aphotic N<sub>2</sub> fixation may be affected by these factors too. In the future, new investigations coupling organic matter characterization, *nifH* quantification and N<sub>2</sub> fixation rate measurement in aphotic waters should help reveal the real importance of aphotic diazotrophy. Considering N<sub>2</sub> fixation rates in aphotic waters could potentially increase global estimates enough to reconcile present differences between N<sub>2</sub> fixation and denitrification (Bonnet et al., 2013), which has important consequences in the ability of our oceans to maintain uptake of CO<sub>2</sub> (Codispoti, 2007).

## What is there Left to be Known?

Ever since the pioneering work of Dugdale et al. (1961), the North Atlantic has been subject to a great amount of diazotrophy-related studies, with a clear NWA bias in their geographical distribution. To date, we have amassed considerable good knowledge of the distribution and magnitude of N<sub>2</sub> fixation rates, the abundance and distribution of common qPCR target groups (most commonly UCYN-A, -B, -C, *Trichodesmium* and Het-1), and the importance of phosphorus and aeolian dust-related iron in controlling diazotrophic activity and diversity in this basin. After five decades of N<sub>2</sub> fixation research in the North Atlantic, there are still open questions to be answered, which are also open questions for N<sub>2</sub> fixation research in other basins:

1. How do different groups contribute to N<sub>2</sub> fixation rates?: Many authors have used size-fractionation to ascribe rates to different groups of diazotrophs (e.g., Moore et al., 2009; Benavides et al., 2011). Such manipulations are prone to alter the behavior of the cells and to enhance dissolved organics release, among a variety of other problems (Mulholland, 2007; Benavides et al., 2013d). The comparison of *in situ* N<sub>2</sub> fixation rates with parallel *nifH* copy abundance has been used extensively to indicate which groups are responsible for the measured activity (most studies are compiled in Luo et al., 2012), but the presence of *nifH* genes does not guarantee a measurable N<sub>2</sub> fixation activity (Turk-Kubo, 2012). The tropical and subtropical NA has been long thought to be dominated by *Trichodesmium*, but despite they abound in the western NA, on a general basis UCYN phylotypes are more abundant (especially UCYN-A; Luo et al., 2012), and *nifH* expression by UCYN can be more important than by *Trichodesmium* locally, suggesting that in fact UCYN may be more representative of bulk N<sub>2</sub> fixation rates than *Trichodesmium*, and changing classic paradigms of *Trichodesmium* dominance in the NA (Turk-Kubo, 2012). Still, the quantification of *nifH* transcripts does not indicate how to divide the measured N<sub>2</sub> fixation activity among the active phylotypes found (Church et al., 2005). The identity-activity matching problem is worsened by the lack of specificity of currently published qPCR primer-probe sets, a problem which is being solved in part for some phylotypes like UCYN-A (Thompson et al., 2014). The development of single-cell approaches like nanoSIMS combined with halogen *in situ* hybridization (HISH-SIMS) has allowed the quantification of specific N<sub>2</sub> fixation rates (Thompson et al., 2012; Krupke et al., 2013, 2014) although the HISH procedure has been shown to dilute the <sup>15</sup>N signal, resulting in inaccurate rates (Musat et al., 2014). Other procedures like the combined use of catalyzed reporter deposition fluorescence *in situ* hybridization (CARD-FISH) with flow cytometry sorting holds great promise, but also comes with difficulties (McInnes et al., 2014). In summary, the combination of new methodologies and the development of single-cell approaches is significantly increasing our capability to quantify compound fluxes within and between planktonic cells, but there is much work to be done before straight-forward methodologies are developed and all aforementioned drawbacks are overcome. All in all, the days of compound “black box” studies are soon to be over.
2. Where does the fixed N<sub>2</sub> go?: N<sub>2</sub> fixation is considered an input of “new nitrogen” (sensu Dugdale and Goering, 1967), as it comes from outside the system and not from *in situ* recycled production. As a new nitrogen source, diazotrophy-derived fixed nitrogen theoretically fuels export production (Capone et al., 2005). In practice however, current research has not resolved whether fixed N<sub>2</sub> is preferentially transferred to higher trophic levels, recycled via the microbial loop, or if it fuels export production and the biological pump (Mulholland, 2007), nor have any of these fluxes been accurately quantified, largely due to the lack of appropriate methods. DDAs seem to clearly contribute to carbon sequestration and supply

fixed nitrogen for remineralization at depth, as it has been seen in the Amazon River plume and at station ALOHA (Subramaniam et al., 2008; Karl et al., 2012), which is logical given the sinking character of these symbioses. The role of slow sinking diazotrophs like UCYN or floating colonies like those of *Trichodesmium* is relatively unknown. Over the past two decades several authors have quantified the release of recently fixed N<sub>2</sub> in the form of ammonium and/or dissolved organic nitrogen, mainly from *Trichodesmium* (Glibert and Bronk, 1994; Mulholland et al., 2004), but also from smaller diazotrophs (Benavides et al., 2013a,d). The metabolism behind this behavior is unknown, but this activity is recurrently found in different marine systems and culture studies like those referenced above. The lability of dissolved organic nitrogen and its potential to fuel autotrophic production is not well-understood due to constraints in compositional analysis (Bronk et al., 2007). Recent advances in DOM characterization are currently starting to answer some of these questions. For example, Sipler et al. (2013) have shown that *Trichodesmium* produces labile dissolved organic nitrogen compounds capable of triggering toxic microalgae blooms. Integrative approaches including stable isotope tracing experiments in combination with state-of-the-art DOM characterization techniques hold great promise for new discoveries.

The other path of fixed nitrogen transfer is direct consumption of diazotrophs by higher trophic levels. Cyanobacteria had been traditionally considered being less palatable than other phytoplankton due to their low nutritional value and the toxin production in the cells (Sellner, 1997). There are two pathways that transport fixed nitrogen to upper trophic levels. One is the direct ingestion of cyanobacteria (e.g., for *Trichodesmium* filaments by a harpacticoid copepod *Macrosetella gracilis*; O’Neil and Roman, 1994), or the uptake of released fixed nitrogen (as described above) by heterotrophic bacteria, entering the microbial food web. Subsequently, microzooplankton sustaining their growth on fixed nitrogen can be consumed by mesozooplankton (Sellner et al., 1994; Montoya et al., 2002; Wannicke et al., 2013). Up the chain, this transport of fixed nitrogen can even stimulate fish production (Rolf et al., 2007). Montoya et al. (2002) estimated that the contribution of diazotrophy to the transfer of bulk surface particulate nitrogen can be as high as 30–40% in a bloom of *Trichodesmium*. However, when the abundance of cyanobacterial cells is low, the transfer may be also low and hence their contribution to mesozooplankton nutrition negligible (Wannicke et al., 2010). A mesocosm study revealed a strong dependency of grazing processes on the development of heterocysts (Chan et al., 2006). Successful growth could be interrupted by a high grazing activity of zooplankton, which may reduce the filament length and heterocyst frequency in cyanobacteria, as it occurs with *Anabaena* (Chan et al., 2006). In summary, cyanobacteria play a significant but mainly indirect role for the nitrogen transfer to higher trophic levels. The role of large colony forming species vs. the role of unicellular species cannot be currently assessed.

3. Are heterotrophic diazotrophs important?: Recently, there has been increased interest on diazotroph phylotypes other than cyanobacteria. Some authors have shown that non-cyanobacterial diazotrophs are widespread in the oceans and potentially very diverse (Riemann et al., 2010; Farnelid et al., 2011), as well as present in previously unrecognized sites like the dark pelagic realm (Hewson et al., 2007; Bonnet et al., 2013; Rahav et al., 2013). It has yet to be determined how much these phylotypes contribute to measured N<sub>2</sub> fixation rates, given current difficulties in matching activity to identity, as discussed above. The ubiquity of heterotrophic diazotrophs in the world's oceans and their predominance in certain systems like the South Pacific Gyre (Halm et al., 2011) suggests that, although their specific activity might be lower than that of other diazotrophs like *Trichodesmium*, their N<sub>2</sub> fixation rates are far from negligible and deserve further study (Falcón et al., 2004). There is an urgent need to develop methodologies that allow us to discern autotrophic from heterotrophic N<sub>2</sub> fixation, as well as to further investigate the diversity of this group, which spreads among the four recognized *nifH* clusters (Riemann et al., 2010). Being heterotrophic, the N<sub>2</sub> fixation activity of this group of diazotrophs may significantly differ from that of autotrophic cyanobacterial diazotrophs, posing a major challenge for microbial oceanographers and opening a variety of new questions which require novel and interdisciplinary approaches.
3. How will diazotrophy react to climate change?: Current CO<sub>2</sub> partial pressure (*p*CO<sub>2</sub>) levels are expected to double by the end of this century (IPCC, 2007). The main consequences of this increase which will affect planktonic organisms are (a) seawater acidification, (b) increase of seawater temperature, (c) water column stratification, (d) expansion of OMZs, and (e) desertification of terrestrial systems and associated increase of desert dust inputs to the ocean (Boyd and Doney, 2002; Jickells, 2005; Stramma et al., 2008b). While seawater acidification can either enhance photosynthetic activity in some species, or be harmful for others (Malakoff, 2012), seawater warming and stratification will likely enhance the proliferation of diazotrophic cyanobacteria such as *Trichodesmium* (Lomas et al., 2012), although their response to increased *p*CO<sub>2</sub> levels may depend on a variety of environmental factors (Shi et al., 2012). Indeed, culture experiments have revealed that the length of *Trichodesmium* filaments and their N<sub>2</sub> fixation rates increase when exposed to increasing levels of CO<sub>2</sub> (Levitan et al., 2007). However, experiments including several different diazotrophic strains suggest that the response to increased *p*CO<sub>2</sub> is strain-specific and depends on the CO<sub>2</sub> uptake kinetics of each diazotroph type (Hutchins et al., 2013). Similarly, field studies have found divergent responses to increasing *p*CO<sub>2</sub> levels, indicating that different phylotypes are unequally affected by acidification, which results in a diluted effect on bulk N<sub>2</sub> fixation field measurements (e.g., Gradoville et al., 2014). Heterotrophic N<sub>2</sub> fixation in hypoxic zones (Hamersley et al., 2011), suggests that the predicted expansion of OMZs (Stramma et al., 2008b) will likely enhance global N<sub>2</sub> fixation rates too, either by *in situ* heterotrophic N<sub>2</sub> fixation, or by the

enhancement of diazotrophy in other oceanic areas in order to balance increased denitrification occurring in the OMZs (Deutsch et al., 2007). N<sub>2</sub> fixation by *Trichodesmium* and other diazotrophs would be further enhanced by an increased availability of iron through desert dust deposition in the oceans (Jickells, 2005). Michaels et al. (2001) proposed a theoretical climate-based N<sub>2</sub> fixation feedback cycle. In this conceptual model, desertification causes a global increase of N<sub>2</sub> fixation rates as a result of iron limitation alleviation through increased desert dust deposition, which in turn lowers atmospheric CO<sub>2</sub> levels and decreases global temperatures. Global cooling may in turn decrease desert dust inputs diminishing N<sub>2</sub> fixation and raising CO<sub>2</sub> levels, which would again raise global temperatures. However, increased inputs of iron to the ocean will likely affect N<sub>2</sub> fixation activity as a function of *in situ* phosphorus availability, suggesting that diazotrophic activity in the phosphorus-deficient NA may not be positively affected by increased dust inputs in the future (Garcia et al., 2014). In conclusion, diazotrophic responses to atmospheric CO<sub>2</sub> levels will likely have an important role in carbon and nitrogen cycling and subsequent productivity in the future oceans, although their impact will depend on a variety of factors which are currently poorly constrained.

## Summary and Concluding Remarks

The role of the NA in comparison with other basins: In **Table 5** we compile the most recent basin-wide N<sub>2</sub> fixation estimates for the main oceanic basins. Different basin-wide averaging methods yield different results, which are also affected by the latitudinal -and hence areal- range considered. All methods indicate that the NA ranks as the third most important oceanic basin in contributing to global N<sub>2</sub> fixation (**Table 5**). Depending on the method used (geometric and arithmetic mean areal rates by Luo et al., 2012, or station-weighted mean areal rates by Großkopf et al., 2012), the percentage contribution of NA N<sub>2</sub> fixation to the global sum ranges from 2.74 to 23.36%. Considering that the global arithmetic mean areal N<sub>2</sub> fixation rate of Luo et al. (2012), and the global weighted mean of Großkopf et al. (2012) are closer to current geochemical global estimates of N<sub>2</sub> fixation (100–200 Tg N y<sup>-1</sup> Deutsch et al., 2007; Gruber and Galloway, 2008), we consider these estimates more appropriate and thus conclude that the NA most probably provides 10–25% of global fixed N<sub>2</sub>. The estimates presented here are somewhat uncertain given the current undersampling in other basins as compared to the NA (e.g., the South Atlantic, South Pacific, and especially the Indian Ocean), as well as the potential importance of diazotrophic activity at higher latitudes (>50°N or 50°S), and in the aphotic zone (>200 m).

The biogeochemical and physical features of the NA influence the distribution and activity of diazotrophs. In the NWA, the low N:P ratio conditions and stratified water column appears to select for *Trichodesmium*. Although the NEA receives more iron-rich dust inputs than the NWA, the colder waters and more turbulent conditions of the NEA (influence of Northwest African upwelling) seems to constrain the proliferation of

**TABLE 5 | Basin-wide N<sub>2</sub> fixation rates compiled from the literature (Modified from Großkopf et al., 2012; Luo et al., 2012).**

Ocean basin	Luo et al., 2012				Ocean basin	Großkopf et al., 2012		
	Latitudinal range (°N)	Area (10 <sup>6</sup> km <sup>2</sup> )	Geometric mean areal rate (Tg N y <sup>-1</sup> )	Arithmetic mean areal (Tg N y <sup>-1</sup> )		Latitudinal range (°N)	Area (10 <sup>6</sup> km <sup>2</sup> )	Station-weighted mean areal rate (Tg N y <sup>-1</sup> )
North Atlantic	0/55	36	1.7	32	North Atlantic	50/−5	39.2	12.7
South Atlantic	0/−40	27	1.1	1.8	South Atlantic	−5/−30	15.9	1.5
North Pacific	0/55	89	35	56	North Pacific	0/30	56.3	33.0
South Pacific	0/−40	72	24	46	South Pacific	0/−35	56.1	29.5
Indian	25/−40	62	n/q	n/q	North Indian	25/0	15.8	9.3
					South Indian	0/−35	31.8	16.7
Global			62*	137*	Global			102.7
% N <sub>2</sub> fixation provided by the North Atlantic			2.74	23.36				12.37

\*Note that these global estimates do not take into account the Indian Ocean.

n/q means not quantifiable

*Trichodesmium*. These colder and dissolved inorganic nitrogen-rich waters are preferentially inhabited by UCYN phylotypes. UCYN-A is the most widespread of the three, while UCYN-B and UCYN-C are constrained to tropical latitudes, with the latter more associated with coastal regions. The Amazon and Congo River plumes are characterized by high dissolved inorganic nitrogen and phosphate inputs, but the rapid consumption of dissolved inorganic nitrogen by non-diazotrophic phytoplankton leaves a low N:P signal which, combined with high silica inputs, sustains the proliferation of DDAs and UCYN (the latter closer to saltier open-ocean waters). Heterotrophic diazotrophs seem to be widespread and more flexible, capable of adapting to contrasting conditions and presumably able to exploit *in situ* DOM.

The wide distribution of small diazotrophs (UCYN and heterotrophs) predicts that their contribution to total basin-scale fixed nitrogen inputs is important. However, linking rates to species has been hampered by several difficulties (Zehr et al., 2007; Turk-Kubo et al., 2014) and so far UCYN-associated N<sub>2</sub> fixation rates have been mostly calculated from size-fractionation studies (usually discerning between >10 and <10 μm fractions (e.g., Benavides et al., 2011), although new single-cell approaches are currently able to give specific N<sub>2</sub> fixation rates for some phylotypes (Foster et al., 2011; Krupke et al., 2013). Still, N<sub>2</sub> fixation rates in UCYN-dominated areas of the NEA are lower than those measured in the NWA, suggesting that the N<sub>2</sub> fixation capacity of UCYN is lower than that of *Trichodesmium* and DDAs. Thus, it seems clear that the contribution of N<sub>2</sub> fixation to new production is greater in *Trichodesmium*-dominated waters of the NWA (Montoya et al., 2007), but minimal in the UCYN-dominated waters of the NEA. The contribution of N<sub>2</sub> fixation to new production is thus greater in *Trichodesmium*-dominated

waters of the NWA. The low contribution of diazotrophic N<sub>2</sub> fixation to new production in the NEA suggests that the fixed nitrogen requirements of autotrophic planktonic communities in the subtropical NEA and inner subtropical gyre are met by a combination of atmospheric fixed nitrogen deposition (as this area is located downwind of European industrialized countries), *in situ* remineralization of advected DOM and, to a minor extent, *in situ* photic nitrate regeneration (nitrification) and N<sub>2</sub> fixation.

N<sub>2</sub> fixation rates obtained with geochemical approaches exceed those obtained with biological approaches two to seven-fold. The differences between both groups of methods stem from nitrogen excess signals derived from processes other than N<sub>2</sub> fixation (such as non-Redfieldian uptake ratios and atmospheric deposition of nitrogenous compounds), but also from methodological underestimations (and overestimations) of the <sup>15</sup>N<sub>2</sub> method. Future methodological developments should allow overcoming current drawbacks in the estimation of N<sub>2</sub> fixation rates (Mohr et al., 2010; Dabundo et al., 2014). Nevertheless, our ability to reconcile present imbalances between global denitrification and N<sub>2</sub> fixation rates will eventually depend on the accuracy of denitrification estimates and the quantification of other sources of fixed nitrogen to the ocean.

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**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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