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Mercury(II) trace detection by a gold nanoparticle-modified glassy carbon electrode using square-wave anodic stripping voltammetry including a chloride desorption step

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ABSTRACT

Gold nanoparticles (AuNPs) were deposited on a glassy carbon (GC) substrate by constant potential electrolysis and characterized by cyclic voltammetry in H₂SO₄ and field emission gun scanning electron microscopy (FEG-SEM). The modified AuNPs-GC electrode was used for low Hg(II) concentration detection using a Square Wave Anodic Stripping Voltammetry (SWASV) procedure which included a chloride desorption step. The comparison of the obtained results with our previous work in which no desorption step was used showed that this latter step significantly improved the analytical performances, providing a three time higher sensitivity and a limit of detection of 80 pM for 300 s preconcentration, as well as a lower average standard deviation. The influence of chloride concentration on the AuNPs-GC electrode response to Hg(II) trace amounts was also studied and its optimal value confirmed to be in the 10⁻² M range. Finally, the AuNPs-GC electrode was used for the determination of Hg(II) in a natural groundwater sample from south of France. By using a preconcentration time of 3000 s, a Hg(II) concentration of 19 ± 3 pM was found, which compared well with the result obtained by cold vapor atomic fluorescence spectroscopy (22 ± 2 pM).

Keywords:

Hg(II) sensor
Gold nanoparticles
Stripping voltammetry
Chloride desorption step
Natural water

1. Introduction

Like many other micropollutants, heavy metals have emerged as a major concern both in terms of human health [1,2] and environmental risks [3,4]. They are considered as a major source of ecological issues because of their widespread occurrence in natural media [5]. This is particularly true for methylmercury (MeHg) which bioaccumulates along the trophic chain [6,7] and has adverse health effects on mammals including humans [8,9]. As it is commonly

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suggested now that MeHg production relies on the (bio-)availability of inorganic Hg(II) [10], it appears of critical importance to thoroughly understand Hg(II) dynamics that is, daily variations of inorganic Hg(II) concentration and speciation. Hg(II) may be dangerous even at very low concentration [7,11,12] so that the World Health Organization has delivered a guideline value of 1 µg L⁻¹ (ca. 5 nM) for drinking water [13]. As a consequence, environmental monitoring of Hg(II) is of critical importance and requires in situ, real-time and highly-sensitive sensors [14].

Hg(II) trace analysis is mainly performed by spectroscopic techniques [15] such as cold-vapor atomic fluorescence spectrometry (CV-AFS) [16–18] or inductively coupled plasma mass spectrometry (ICPMS) generally associated with gas chromatography [19–22]. All these techniques offer good selectivity and sensitivity [23], reaching 0.2 pM for total Hg(II) [15,18,21]. However, they suffer important limitations, i.e. expensive material and complex and time-consuming procedures, thus limiting any in situ or on line and *operando* analysis. Moreover, rigorous

sampling and storage conditions are necessary to avoid any external Hg(II) contamination or speciation changes [24,25].

In this context, tremendous attention has been paid to electrochemical sensors due to their manifold advantages over other detection methods: they are usually cheap, user-friendly, require little energy and simple procedures and are suitable for on-site analysis. Detection limits in the pM range are frequently reached by using sensitive methods like differential pulse (DPASV) or square wave (SWASV) anodic stripping voltammetry [26,27] either on platinum [28], carbon-based materials [29–33] or gold (Au) electrode. Because of its high affinity for Hg, Au is the most commonly used electrode material in various configurations: bulk [34], film [35,36], microwire [37,38], microdisk [39] or microdisk array electrode [40]. Another strategy is the use of chemically functionalized electrodes [27,41]. In recent years, Hg(II) detection by means of gold nanoparticles (AuNPs)-modified electrodes has been the subject of many reports [42–45]. This is due to the specific physicochemical properties of these nanomaterials [46]: enhanced diffusion of electroactive species, high effective surface area, improved selectivity, catalytic activity, and higher signal-to-noise ratio [47]. AuNPs may be prepared either by chemical [44] or electrochemical route [42,43,45]. Recently, we reported on an AuNPs-modified glassy carbon (AuNPs-GC) sensor dedicated to Hg(II) trace determination [48,49]. The amperometric response towards Hg(II) was found to be dependent on AuNPs size and density [49] and was hampered by a broad baseline due to chloride anions contribution. We proved that this was rather due to Cl⁻ adsorption onto the Au surface than to calomel formation [50], contrary to what has been reported previously in the literature [51]. Considering that the use of Cl⁻ is a necessary condition to improve Hg(II) trace detection [52], Salaün and van den Berg added a desorption step to the ASV procedure in order to minimize the broad baseline [38]. The detection of Hg(II) and Cu(II) on a Au microwire electrode was improved using this supplementary step. However, no comparative feature was provided with respect to analytical performances whatever the desorption step was used or not. Surprisingly, this up-and-coming strategy did not give rise to a large number of reports. We found only one work from Abollino's group which took advantage of applying a negative potential of -0.8 V between the deposition and the stripping step of the ASV procedure to minimize the adsorption of iodide ions present in Sea lettuce sample [53]. However, few details have been provided since no experimental data were given to support the improvement brought by this additional step into the analytical procedure.

In this study, we report Hg(II) trace determination results obtained with a AuNPs-GC modified electrode using a SWASV procedure which included a chloride desorption step. The improvement brought was evidenced by comparison of the analytical performances of the sensor with and without this latter step. Different calibration curves were recorded using varying Cl⁻ concentrations and the AuNPs-GC modified electrode was successfully tested for Hg(II) determination in natural water.

2. Experimental section

2.1. Reagents

All solutions were prepared in ultrapure water (Milli-Q, Millipore, 18.2 M Ω cm). NaNO₃, 65% HNO₃, 30% HCl, NaCl and NaOH were purchased from Merck (suprapur grade). Pb(NO₃)₂ (ACS grade) was also provided by Merck. 95% H₂SO₄ (normapur grade) was obtained from VWR Prolabo. H₂AuCl₄·3H₂O pro-analysis grade was supplied by Acros Organics. Hg(II) solutions were prepared by dilution of 1001 ± 2 mg L⁻¹ Hg(NO₃)₂ NIST standard solution (certiPUR grade, Merck) in 0.01 M HNO₃ (pH 2) and then used as it.

2.2. Apparatus

All the electrochemical experiments were performed at the Laboratoire de Génie Chimique (LGC) (Toulouse, France) at room temperature in a Teflon PFA three-electrode cell (Metrohm) using a Metrohm Autolab PGSTAT 128N potentiostat interfaced to a computer and controlled with the NOVA 1.10 software package. A Metrohm Ag/AgCl/KCl 3 M electrode separated from the electrochemical cell by a Teflon PTFE capillary containing 0.1 M NaNO₃ solution and ended by a ceramic diaphragm (D type), and a Metrohm glassy carbon wire were used as reference and counter electrodes, respectively. All the potentials are referred to this reference electrode. The working electrode was a glassy carbon (GC) rotating disk electrode from Radiometer (3 mm diameter, A=7.07 mm²) modified by gold nanoparticles (AuNPs). The electrochemical cell was maintained in a Faraday cage in order to minimize electrical interferences. The solution was deaerated with a high purity N₂ stream during 10 min and maintained under N₂ atmosphere during each experiment.

The AuNPs-GC surface was characterized by field emission gun scanning electron microscopy (FEG-SEM) using a Quanta 250 FEG FEI equipment with an accelerating voltage of 5 kV and a working distance 7 mm depending on the sample. Image analysis was carried out using a LGC homemade program for particle counting (density estimation) and average diameter measurement developed using MatLab image processing toolbox software [48]. The density and average size of AuNPs were evaluated from a 7.2 μ m² GC surface analysis counting 1165 or 2806 particles (depending on the micrograph was recorded before or after a series of electrochemical measurements). For each deposit, the error was calculated from the analysis of three different SEM images using the adequate magnification.

Total Hg analyses were performed at the Géosciences Environnement Toulouse laboratory (France) with cold-vapor atomic fluorescence spectrometry (CV-AFS) following a method from the Environmental Protection Agency (EPA-1631) [16,54] modified by Cossa et al. [55]. Briefly, total Hg concentration analysis was achieved by chemical reduction of dissolved Hg(II) to vapor phase Hg(0) using SnCl₂ in acidic media after BrCl preoxidation. Hg(0) vapor was then amalgamated on a gold trap and released by flash heating into a Brooks Rand Model III AFS. The accuracy was verified daily with the ORMS-5 certified reference material of riverine water from the National Research Council of Canada.

2.3. Water sampling and conditioning

Natural water dedicated to Hg(II) trace detection was sampled from Font Estramar (42° 51' 31.73"N 2° 57' 30.776"E), which is a submarine groundwater discharge located at Salses-le-Château (Pyrénées Mountains) at the edge of Corbières Mountains (France). An unfiltered sample was taken and stored into a 2 L FEP Teflon bottle. Chloride concentration in this water was 34 mM and dissolved organic carbon concentration was 1.2 mg L⁻¹. The sample was acidified with 0.01 M HNO₃ overnight before SWASV analysis.

2.4. Working electrode preparation

GC surface was carefully polished successively by 9 μ m (5 min), 3 μ m (2 min), 1 μ m (1 min) and 0.3 μ m (1 min) mecaprex diamond compounds (Presi) on a cloth polishing pad. The GC surface was cleaned in an ultrasonic bath in ethanol for 5 min between each polishing in order to remove impurities. AuNPs were electrochemically deposited at 20 °C in a standard three-electrode water-jacketed cell from a 0.1 M NaNO₃ deaerated solution containing 0.25 mM H₂AuCl₄ by potentiostatic electrolysis at 0 V for 15 s, according to our previous work [49]. Then the electrode was

activated by recording 10 voltammograms between 0 and 1.4 V at a scan rate of 100 mV s^{-1} in $0.5 \text{ M H}_2\text{SO}_4$.

2.5. Hg(II) detection

Hg(II) detection on AuNPs–GC electrode was achieved in deaerated 0.01 M HCl or 0.01 M HNO_3 solution containing NaCl (from 0.2 mM to 500 mM). Hg(II) was analyzed by SWASV under the following conditions: constant cleaning potential between 0.5 and 0.8 V , depending on NaCl concentration, for 15 s ; preconcentration cathodic potential at 0 V for 300 s ; desorption cathodic potential at -0.8 V for 60 s ; pulse amplitude = 25 mV , step amplitude = 5 mV , frequency = 200 Hz ; anodic scan from 0 to 0.5 – 0.8 V depending on NaCl concentration. The working electrode rotation frequency was 2000 rpm during the preconcentration step. A second scan was recorded immediately after the first one using the same conditions except that the preconcentration and desorption cathodic times were reduced to 30 s and 6 s , respectively (10% of first measurement) and considered as a blank [48,49]. Hg(0) reoxidation peak heights (Δi_p) were calculated after subtraction of this blank. This procedure, called “subtractive ASV method”, has been previously reported in the literature [56]. It allows the analytical results to be released from background vagaries. It has to be noticed that the subtractive anodic signals recorded with low Hg(II) concentrations were noisy; hence we needed to use a Savitzky-Golay smoothing function.

3. Results and discussion

3.1. AuNPs electrodeposition and characterization

AuNPs were electrochemically deposited onto the GC electrode by applying a constant electrode potential of 0 V for 15 s to a 0.1 M NaNO_3 solution containing 0.25 mM HAuCl_4 . This potential being nearly 500 mV more cathodic than the peak potential corresponding to Au(III) reduction, AuNPs deposition onto the electrode surface occurred according to Reaction (1):



These conditions were chosen in accordance with our previous work in order to get a dense deposit of small NPs, which ensures the best analytical performances with respect to Hg(II) trace determination [49]. The obtained deposit was characterized by cyclic voltammetry in $0.5 \text{ M H}_2\text{SO}_4$ and FEG-SEM. Fig. 1 shows the typical CV obtained in acidic media for electrodeposited AuNPs (solid line). The voltammogram exhibited on the forward scan the anodic signals between 1.1 and 1.35 V which correspond to the formation of several kinds of Au oxides, the major one being AuO according to Reaction (2):



On the backward scan, a single reduction peak at 0.88 V with a post-peak shoulder was observed, which corresponded to the reduction of the oxides formed during the forward scan. Fig. 2A shows a FEG-SEM micrograph recorded on a GC electrode freshly functionalized by electrodeposited AuNPs. The different features corresponding to the deposit extracted from Figs. 1 and 2, namely the amount of charge corresponding to Au oxides reduction (Q_{oxides}) and AuNPs average size and density are summarized in Table 1. As expected, the FEG-SEM micrograph exhibited a dense array of AuNPs, homogeneously distributed over the GC surface. Both AuNPs average size and density compared favorably with our previous work (ca. 17 nm and $332 \text{ particles } \mu\text{m}^{-2}$, respectively)

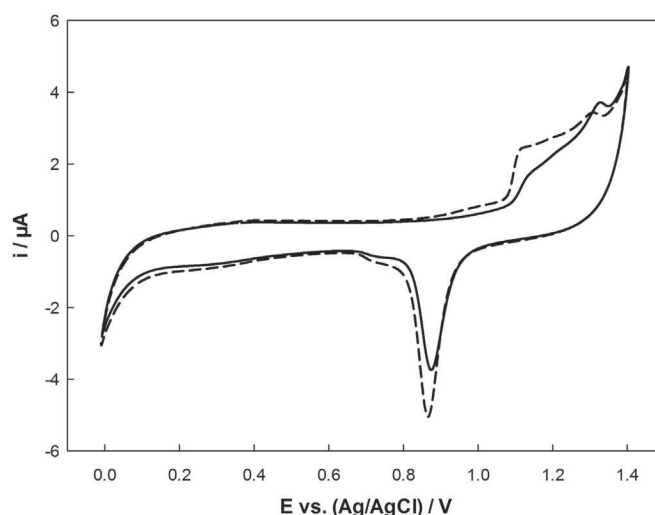


Fig. 1. CVs recorded in a $0.5 \text{ M H}_2\text{SO}_4$ solution on a AuNPs–GC electrode before (solid line) and after (dashed line) a series of Hg(II) measurements using SWASV. Scan rate: 100 mV s^{-1} . AuNPs–GC electrode was prepared from a deaerated 0.1 M NaNO_3 solution containing 0.25 mM HAuCl_4 by constant potential electrolysis at 0 V for 15 s .

[49], thus demonstrating good reproducibility of the deposition method.

3.2. Influence of chloride desorption potential on analytical performances

The influence of the insertion of a chloride desorption potential in the analytical procedure was checked by studying the SWASV response of the AuNPs–GC electrode to a 0.01 M HCl solution containing 4 nM Hg(II) . This latter condition was chosen in order to allow direct comparison to be made with our previous work [49] in which no desorption potential was used (see Table 2 for comparative features). Fig. 3 presents the SWASVs obtained depending on whether a desorption step was included or not in the analytical procedure. When using this latter step, the peak around 0.6 V which corresponded to Hg(0) reoxidation was sharpened and the peak current was increased. Indeed, once the analytical blank subtraction operated (see Experimental Section for details), the resulting peak current (Δi_p) was nearly twice when using the chloride desorption step, suggesting an improved sensitivity of the AuNPs toward Hg(II). Applying a negative desorption potential also induced simultaneously a slight decrease in the broad amperometric signal observed in the potential range 0.2 – 0.4 V , which was proved to result from Cl^- adsorption [50]. Consequently, this additional desorption step clearly modified the overall shape of the voltammogram by reducing the broad baseline and by improving the amperometric response with respect to Hg(II) detection.

The analytical performances of the AuNPs–GC electrode were checked by adding successive amounts of Hg(II) to a 0.01 M HCl solution and recording the corresponding SWASVs using a chloride desorption potential. The amperometric response exhibited a clear linear trend up to 6 nM Hg(II) (Fig. 3, inset). For higher Hg(II) concentrations, the Δi_p values obtained were lower than that expected from the linear regression, suggesting a saturation effect of the electrode surface. As can be seen from Table 2, the linear range was wider when no desorption step was applied, since Δi_p values obtained for Hg(II) concentrations higher than 6 nM fitted to the linear regression curve. These data confirmed that when no negative potential desorption step was applied between the deposition and the stripping steps of the ASV procedure, a bigger amount of chloride ions remained adsorbed at the electrode

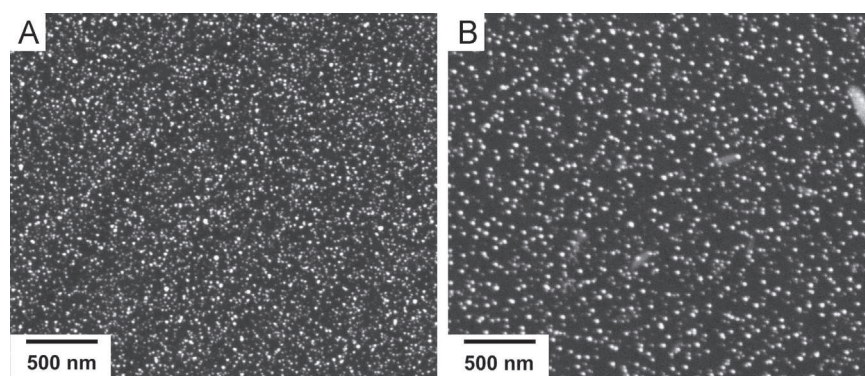


Fig. 2. FEG-SEM micrographs of a AuNPs-GC electrode prepared from a deaerated 0.1 M NaNO₃ solution containing 0.25 mM HAuCl₄ by constant potential electrolysis at 0 V for 15 s before (A) and after (B) a series of Hg(II) measurements.

Table 1
Characteristics of the AuNPs deposit on GC electrode obtained by CV and FEG-SEM.

	Q_{oxides} (μC) ^a	NPs density (μm^{-2}) ^b	Average diameter (nm) ^b
Before Hg(II) analysis	3.2	391 ± 15 (2806)	19 ± 8
After Hg(II) analysis	4.8	162 ± 19 (1165)	27 ± 9

^a Q_{oxides} is the charge corresponding to the reduction of Au oxides in 0.5 M H₂SO₄ obtained by integration of the peak at 0.88 V (Fig. 1, solid line).

^b See Section 2 for details on NPs density and average diameter estimation. The values in brackets correspond to the average number of particles considered for the calculation (Fig. 2).

surface, which constituted an additional diffusion barrier with respect to Hg(II) reduction. On the contrary in the presence of a desorption step, the electrode surface is more reachable for Hg(II) thus making the sensor more sensitive. This is consistent with the significant increase in sensitivity which may be noticed between the procedure with ($0.60 \mu\text{A nM}_{\text{Hg}}^{-1} \text{min}^{-1}$) and without ($0.23 \mu\text{A nM}_{\text{Hg}}^{-1} \text{min}^{-1}$) chloride desorption step. The limit of detection (LOD) was also significantly improved by the addition of this latter step since Hg(II) concentrations down to 80 pM may be detected and quantified vs. 400 pM in our previous work, using a same preconcentration time of 300 s. It has to be noticed that the 80 pM value was calculated for a signal-to-noise ratio of 3 [57] and confirmed experimentally by adding successive 10 pM Hg(II) amounts to a Hg-free solution. This lowered LOD is in accordance with the results reported by Salaün and van den Berg who proposed a flatter baseline to be at the origin of this improvement [38], however, without providing any demonstration. Our results also agreed well with this latter work since we found that the best desorption duration was 20% of the deposition time, ca. 1 min. Thus, the AuNPs-GC electrode seemed to behave in a similar way to a Au microwire since its analytical performances were significantly enhanced by the use of a chloride desorption step, considering the actual goal of the system to be Hg(II) trace determination. Finally, an improvement of the accuracy of the sensor was also noticed, since the average relative standard deviation (RSD) was more than twice lower when operating with the chloride desorption step. The analytical performances of the sensor also compared well with literature data. In particular, the normalized sensitivity was found to be one of the best, being closed to that reported by Abollino et al. [43]. Only Gong et al. reported a value 1.6 times higher [58]. However the functionalized interface was a little bit complicated in this latter case, since previously prepared bimetallic Au-Pt NPs were electrodeposited onto organic nanofibers on a GC electrode. The average RSD of the AuNPs-GC sensor was also found to be one of the lowest. The

widest linear ranges were observed on systems which used organic compounds or polymers combined to NPs [42,59], except that reported by Abollino et al. [43], but in this case the metal deposit looked like more a 3D porous nanostructured Au film than a AuNPs array. Finally, it has to be noticed that, as far as we know, this work is the only one using a chloride desorption step which reports clear analytical performances features.

3.3. Influence of chloride concentration on analytical performances

The influence of chloride concentration on the analytical performances was studied by performing calibrations curves in the range 0.3–10 nM Hg(II) using an analytical procedure which included a chloride desorption step. The Cl⁻ concentrations were chosen in order to mimic the most common conditions that may be encountered in natural waters, 0.2 mM and 505 mM corresponding to the Cl⁻ concentrations frequently encountered in fresh and sea water, respectively. The results obtained are summarized in Table 3. Clearly, the best responses were obtained for Cl⁻ concentrations around 10⁻² M, in accordance with literature data [34,38,60]. Linear range, sensitivity and LOD reached their optimum values under these conditions, the best ones corresponding to 60 mM Cl⁻. Thus, from this point of view, the AuNPs-GC electrode behave rather like thick film graphite electrodes modified with a Au(III) inorganic complex [60] than like a Au disk [34] or microwire [38]. For Cl⁻ concentration corresponding to sea water, the sensitivity remained quite good but the LOD was 400 nM, this latter value being the same as that obtained in 0.01 M Cl⁻ without chloride desorption step. The linear range was narrowed both from lower and higher concentrations, being only 0.9–4.0 nM. In this case Cl⁻ anions seemed to hamper Hg(II) trace detection, probably because of their very strong interactions with the Au surface. When operating at 0.2 mM Cl⁻, the AuNPs-GC electrode exhibited sensitivity more than 4 times lower than under the best conditions. It has to be noticed that the LOD was better in this latter case than for 505 mM Cl⁻, leading to the observation that a better sensitivity did not give rise to lower LOD. The reason for that is not clear but may lie in the Cl⁻ interactions with the Au surface.

3.4. Stability of the AuNPs deposit toward measurements

The stability of the AuNPs-GC electrode analytical response was checked vs. time and measurements. It was found that the amperometric signal corresponding to a given Hg(II) concentration decreased very fast over few days (not shown). An alteration of the signal was also noticed when increasing the number of successive measurements, although a cleaning step was included in the whole analytical procedure. To get information on this phenomenon which is poorly documented in the literature, the AuNPs

Table 2

Comparison of the analytical performances of a AuNPs–GC electrode in 0.01 M HCl with respect to Hg(II) trace determination.

Cl ⁻ desorption step	Electrode modification procedure	Linear range (nM)	Normalized sensitivity ^a ($\mu\text{A nM}^{-1} \text{min}^{-1}$)	LOD (pM)	Average RSD (%)	Réf.
No	AuNPs	0.64–4.0	0.274	420	–	[48]
No	AuNPs	0.8–9.9	0.23	400	4.0	[49]
No	MBT–Au nanopores arrays ^b	0.05–10	0.37	20	2.1	[65]
No	Imprinted MPMBT/GCE ^c	1.0–160.0	0.053	100	2.5	[59]
No	Imprinted PMBT/AuNPs/SWCNTs ^d	0.4–96.0	0.062	80	2.6	[45]
No	MES/AuNPs ^e	5–500	0.009	660	–	[42]
No	AuNPs	50–250	0.65	7.5	2.7	[43]
No	Au–PtNPs/NFs ^f	Up to 50	0.962	40	1.2	[58]
Yes	AuNPs	–	–	–	3.5	[53]
Yes	AuNPs	0.4–6.0	0.60	80	1.5	This work

^a The value of the normalized sensitivity was calculated by dividing the slope of the linear response by the electrochemical or chemical preconcentration time. All electrochemical measurements were performed by SWASV excepted Ref. [45] which used DPASV.

^b 2-Mercaptobenzothiazole adsorbed on gold micro-/nanopore arrays.

^c Hg(II) imprinting microporous poly(2-mercaptobenzothiazole) film electropolymerized on GC electrode.

^d Hg(II) imprinting poly(2-mercaptobenzothiazole) film electropolymerized on the surface of gold nanoparticles/single-walled carbon nanotube nanohybrids modified glassy carbon electrode.

^e Mercaptoethanesulfonate monolayer-modified AuNPs.

^f Bimetallic Au–Pt nanoparticles/organic nanofibers.

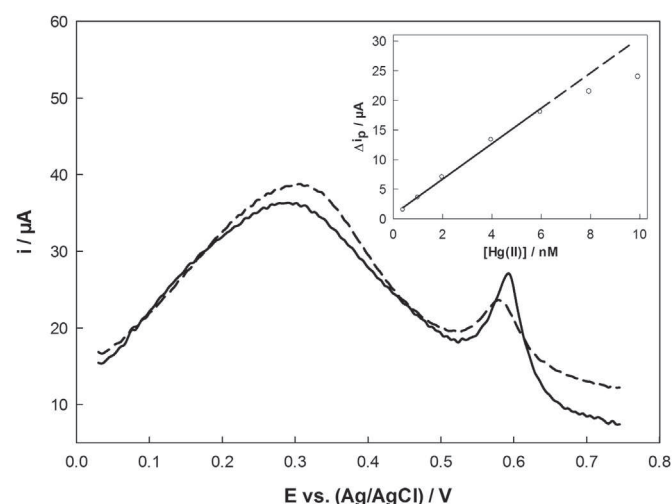


Fig. 3. SWASVs recorded on a AuNPs–GC electrode in a deaerated 0.01 M HCl solution containing 4 nM Hg(II) using a procedure including (solid line) or not including (dashed line) a chloride desorption step. Inset: Calibration curve obtained by SWASV on a AuNPs–GC electrode in a deaerated 0.01 M HCl solution with increasing amounts of Hg(II). Each point was the average value obtained for 3 successive measurements. Error bars were not provided in the graph because they were too much tiny (between 0.04 and 0.17 μA) to be seen.

Table 3Influence of chloride concentration on the AuNPs–GC electrode analytical performances in 0.01 M HNO₃.

[Cl ⁻] (mM) ^a	Linear range (nM Hg(II))	Normalized sensitivity ($\mu\text{A nM}^{-1} \text{min}^{-1}$)	LOD (pM)
0.2	0.9–4.0	0.153	200
34	0.4–6.0	0.656	80
60	0.4–6.0	0.711	60
505	0.9–4.0	0.520	400

^a Added as NaCl salt.

deposit was characterized by CV and SEG-FEM after a series of Hg(II) measurements. The comparison of the two CVs recorded in 0.5 M H₂SO₄ before and after Hg(II) measurements (Fig. 1) clearly indicates a modification of the surface structure of the AuNPs, as illustrated by the very different morphologies observed for the part of the CVs corresponding to Au oxides formation. Moreover, on the backward scan, the cathodic peak of Au oxides reduction

was larger after Hg(II) measurements than before, suggesting the active surface area of the whole deposit to be increased by successive analytical procedures. This observation indicates that during the preconcentration step, Hg does not only form a monolayer on the AuNPs but amalgamates, thus modifying the structure of the electrode surface and probably increasing its roughness. However, complex phenomena may occur during Hg preconcentration since the active surface increase did actually lead to a decrease in the amperometric response corresponding to Hg(0) reoxidation. Fig. 2B shows the SEM-FEG micrograph obtained for the AuNPs–GC electrode after a series of Hg(II) measurements. A less dense deposit of larger AuNPs was observed, compared to Fig. 2A (see Table 1 for quantitative features), which confirms that the deposit morphology strongly evolves during trace Hg(II) measurements. The almost 50% increase in average diameter of the AuNPs suggests that coalescence phenomena occur but it could be also possible that very small AuNPs redissolve into the solution during the reoxidation step due to their poor stability [61,62], only the biggest AuNPs remaining on the electrode surface. The fact that both NPs average diameter increases and their density decreases implies that the geometric surface of the whole deposit decreases. However, the integration of the Au oxides reduction peak at 0.88 V (Fig. 1) indicated that the active surface area increased, which is consistent with the hypothesis of a higher roughness.

3.5. Analysis of a natural water sample

To check the applicability of the AuNPs–GC sensor combined to the electroanalytical procedure including a chloride desorption step toward real sample analysis, Hg(II) measurements were conducted on natural water from Font Estramar. After addition of 0.01 M HNO₃ and stirring overnight, the sample was analyzed using the established procedure. This latter did not allow any peak in the area corresponding to Hg(0) reoxidation to be detected so that an optimization became necessary. Tests were first performed using lower preconcentration potentials until a reoxidation peak was observed. –0.1, –0.2, –0.3 and –0.4 V were successively tested and only the latter potential allowed a Hg(0) reoxidation peak to be observed (not shown). However the corresponding peak current was too low to be used for quantification so that further optimization was performed by increasing the duration of the preconcentration step. A satisfactory peak current value (ca. 0.35 μA) was obtained for 3000 s preconcentration. Using the new parameters, four successive

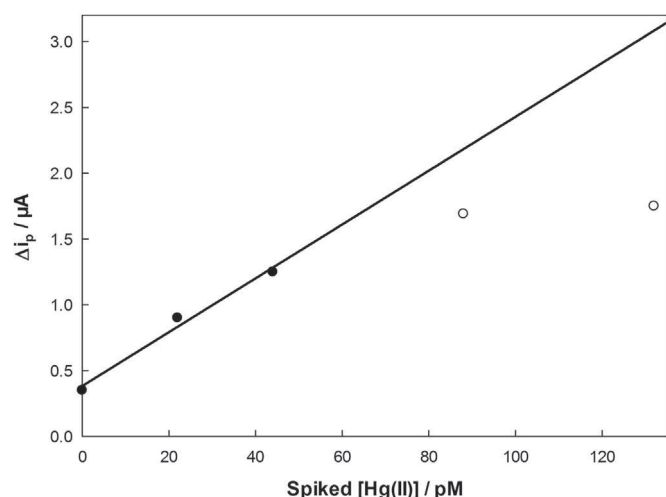


Fig. 4. Calibration curve obtained by SWASV on a AuNPs-GC electrode in an acidified (0.01 M HNO₃) sample of Font Estramar sample. The measurements were performed using 3000 s preconcentration at -0.4 V. White circles corresponds to data which were not included in the regression.

amounts of Hg(II) were spiked into the Font Estramar sample. The corresponding analytical responses are depicted in Fig. 4. Only the first two spiked concentrations allowed a linear regression curve to be calculated, the two higher ones inducing a saturation phenomenon in a similar trend to that reported above, mainly because of the significantly longer preconcentration time. On the basis of the regression equation ($\Delta I_p = 0.6881 \pm 2.9963 [\text{Hg(II)}]$; $r^2 = 0.9940$), the Hg(II) concentration in the Font Estramar sample was estimated to be 19 ± 3 pM. This value was compared to reference technique analysis, namely CV-AFS, which yielded $[\text{Hg(II)}] = 22 \pm 2$ pM. Thus, both the reference method and the AuNPs-GC electrode provided comparable values for the Hg(II) content at the 95% confidence level. This last result makes us confident concerning the selectivity of the sensor and rules out major interferences and especially Cu(II). The concentration of this latter metal cation in submarine groundwater discharge such as Font Estramar has been reported to be around 20 nM [63]. Also, Cu(II) average concentration in the Rhône River (closed to Font Estramar) was 32.7 ± 12.6 nM [64], which confirmed the value reported by Beck et al. Thus, the favorable comparison of the results obtained by AFS and the AuNPs-GC sensor suggests that Hg(II) detection using this latter is not affected by Cu(II) concentration in a 1/1000 Hg to Cu ratio. In a similar way, it may be assumed that organic matter did not undergo interferences when present at concentrations up to 1.2 mg L^{-1} , which corresponded to the dissolved organic carbon concentration found in Font Estramar water. These results proved that our system was reliable and potentially suitable for in situ analysis of natural water samples. Work is currently in progress in our laboratory in this sense.

4. Conclusions

In this work, we showed that the addition of a chloride desorption step in the analytical procedure aiming at Hg(II) trace determination allows to significantly improve the analytical performances of a AuNPs-GC electrode. In particular, the sensitivity was three times better and the LOD was as low as 80 pM for 300 s preconcentration. The influence of chloride amount on the analytical performances was also studied and it was proved that AuNPs behave in a similar trend to a Au microwire from this point of view. Finally, the AuNPs-GC electrode succeeded in the determination of Hg(II) trace in a natural groundwater sample, allowing 19 pM Hg(II) to be detected for 3000 s preconcentration. Further

works are in progress to improve the AuNPs deposits stability over time and the number of measurements and to better understand their physico-chemical evolution. Efforts will also focus on the decrease of the preconcentration time in natural water and on a more complete study of potential interfering species.

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References

- [1] J. Musarrat, A. Zaidi, M.S. Khan, M.A. Siddiqui, A.A. Al-Khedhairi, *Environ. Pollut.* 20 (2011) 323–342.
- [2] S. Prabhakar, A.K. Singh, D.S. Pooni, *Indian J. Anim. Sci.* 82 (2012) 244–255.
- [3] E. Callender, *Treatise Geochem.* 9 (2004) 67–105.
- [4] S.P. Roy, *Ecoscan* 4 (2010) 235–240.
- [5] G.N. Mhatre, *Encycl. Environ. Control Technol.* 8 (1995) 427–454.
- [6] W.F. Fitzgerald, C.H. Lamborg, *Geochemistry of Mercury in the Environment*, Oxford, Pergamon, 2007.
- [7] F.M.M. Morel, A.M.L. Kraepiel, M. Amyot, *Annu. Rev. Ecol. Syst.* 29 (1998) 543–566.
- [8] D. Mergler, H.A. Anderson, L.H.M. Chan, K.R. Mahaffey, M. Murray, M. Sakamoto, A.H. Stern, *Ambio* 36 (2007) 3–11.
- [9] M. Sakamoto, K. Murata, A. Kakita, M. Sasaki, in: Y. Cai, G. Liu, N. O'Driscoll (Eds.), *Advances in Environmental Chemistry and Toxicology of Mercury*, Wiley-VCH, Weinheim, 2012, pp. 501–516, Chapter 15.
- [10] D. Cossa, C. Garnier, R. Buscail, F. Elbaz-Poulichet, N. Mikac, N. Patel-Sorrentino, E. Tessier, S. Rigaud, V. Lenoble, C. Gobeil, *Biogeochemistry* 119 (2014) 35–43.
- [11] R. Eisler, *Environ. Geochem. Health* 25 (2003) 325–345.
- [12] Q. Wang, D. Kim, D.D. Dionysiou, G.A. Sorial, D. Timberlake, *Environ. Pollut.* 131 (2004) 323–336.
- [13] *Guidelines for Drinking-water Quality*, Vol. 1 third ed., World Health Organization, Geneva, 2008. (http://www.who.int/water_sanitation_health/dwq/gdwq3rev/en/) (accessed 16.06.11).
- [14] S.L. Suib, *Front. Chem.* 1 (2013) 1–2.
- [15] K. Leopold, M. Foulkes, P. Worsfold, *Anal. Chim. Acta* 663 (2010) 127–138.
- [16] N. Bloom, *Can. J. Fish. Aquat. Sci.* 46 (1989) 1131–1140.
- [17] N.S. Bloom, M. Horvat, C.J. Watras, *Water Air Soil Pollut.* 80 (1995) 1257–1268.
- [18] D. Cossa, L.-E. Heimbürger, D. Lannuzel, S.R. Rintoul, E.C.V. Butler, A.R. Bowie, B. Averty, R.J. Watson, T. Remenyi, *Geochim. Cosmochim. Acta* 75 (2011) 4037–4052.
- [19] K.L. Bowman, C.R. Hammerschmidt, *Limnol. Oceanogr.—Methods* (2011) 121–128.
- [20] C.-M. Tseng, C.R. Hammerschmidt, W.F. Fitzgerald, *Anal. Chem.* 76 (2004) 7131–7136.
- [21] M. Monperrus, E. Tessier, S. Veschambre, D. Amouroux, O. Donard, *Anal. Bioanal. Chem.* 381 (2005) 854–862.
- [22] D. Point, W.C. Davis, J.I. Garcia Alonso, M. Monperrus, S.J. Christopher, O.F.X. Donard, P.R. Becker, S.A. Wise, *Anal. Bioanal. Chem.* 389 (2007) 787–798.
- [23] S.K. Pandey, K.-H. Kim, R.J.C. Brown, *Trends Anal. Chem.* 30 (2011) 899–917.
- [24] C.H. Lamborg, C.R. Hammerschmidt, G.A. Gill, R.P. Mason, S. Gichuki, *Limnol. Oceanogr.—Methods* 10 (2012) 90–100.
- [25] C.R. Hammerschmidt, K.L. Bowman, M.D. Tabatchnick, C.H. Lamborg, *Limnol. Oceanogr.—Methods* 9 (2011) 426–431.
- [26] J. Buffle, M.-L. Tercier-Waeber, *Trends Anal. Chem.* 24 (2005) 172–191.
- [27] C. Gao, X.-J. Huang, *Trends Anal. Chem.* 51 (2013) 1–12.
- [28] J.M. Pinilla, L. Hernandez, A.J. Conesa, *Anal. Chim. Acta* 319 (1996) 25–30.
- [29] S. Meyer, F. Scholz, R. Trittler, *Fresenius' J. Anal. Chem.* 356 (1996) 247–252.
- [30] G.G. Muntyanu, *J. Anal. Chem.* 56 (2001) 546–551.
- [31] H. Yi, *Anal. Bioanal. Chem.* 377 (2003) 770–774.
- [32] M. Shamsipur, J. Tashkhourian, B. Hemmateejad, H. Sharghi, *Talanta* 64 (2004) 590–596.
- [33] J. Svarc-Gajic, Z. Stojanovic, Z. Suturovic, N. Marjanovic, S. Kravic, *Desalination* 249 (2009) 253–259.
- [34] Y. Bonfil, M. Brand, E. Kirowa-Eisner, *Anal. Chim. Acta* 424 (2000) 65–76.
- [35] E.A. Viltchinskaia, L.L. Zeigman, D.M. Garcia, P.F. Santos, *Electroanalysis* 9 (1997) 633–640.
- [36] L.A. Khustenko, L.N. Larina, B.F. Nazarov, *J. Anal. Chem.* 58 (2003) 262–267.
- [37] S. Daniele, C. Bragato, M.A. Baldo, J. Wang, J. Lu, *Analyst* 125 (2000) 731–735.
- [38] P. Salaün, C.M.G. van den Berg, *Anal. Chem.* 78 (2006) 5052–5060.
- [39] C. Garnier, L. Lesven, G. Billon, A. Magnier, O. Mikkelsen, I. Pizeta, *Anal. Bioanal. Chem.* 386 (2006) 313–323.

- [40] O. Ordeig, C.E. Banks, Jd Campo, F.X. Munoz, R.G. Compton, *Electroanalysis* 18 (2006) 573–578.
- [41] L. Pujol, D. Evrard, K. Groenen-Serrano, M. Freyssinier, A. Ruffien-Cizsak, P. Gros, *Front. Chem.* 2 (2014) 19.
- [42] X.H. Gao, W.Z. Wei, L. Yang, T.J. Yin, Y. Wang, *Anal. Lett.* 38 (2005) 2327–2343.
- [43] O. Abollino, A. Giacomino, M. Malandrino, G. Piscionieri, E. Mentasti, *Electroanalysis* 20 (2008) 75–83.
- [44] A. Safavi, E. Farjami, *Anal. Chim. Acta* 688 (2011) 43–48.
- [45] X.-C. Fu, J. Wu, L. Nie, C.-G. Xie, J.-H. Liu, X.-J. Huang, *Anal. Chim. Acta* 720 (2012) 29–37.
- [46] Y. Liu, G. Su, B. Zhang, G. Jiang, B. Yan, *Analyst* 136 (2011) 872–877.
- [47] C.M. Welch, R.G. Compton, *Anal. Bioanal. Chem.* 384 (2006) 601–619.
- [48] T. Hezard, K. Fajerweg, D. Evrard, V. Colliere, P. Behra, P. Gros, *J. Electroanal. Chem.* 664 (2012) 46–52.
- [49] T. Hezard, K. Fajerweg, D. Evrard, V. Colliere, P. Behra, P. Gros, *Electrochim. Acta* 73 (2012) 15–22.
- [50] T. Hezard, L. Laffont, P. Gros, P. Behra, D. Evrard, *J. Electroanal. Chem.* 697 (2013) 28–31.
- [51] A. Giacomino, O. Abollino, M. Malandrino, E. Mentasti, *Talanta* 75 (2008) 266–273.
- [52] E. Pinilla Gil, P. Ostapczuk, *Anal. Chim. Acta* 293 (1994) 55–65.
- [53] O. Abollino, A. Giacomino, M. Ginepro, M. Malandrino, I. Zelano, *Electroanalysis* 24 (2012) 727–734.
- [54] USEPA, Method 1631, Revision E: Mercury in Water by Oxidation, Purge and Trap, and Cold Vapor Atomic Fluorescence Spectrometry, 2002.
- [55] D. Cossa, B. Averty, J. Bretaudeau, A.S. Senard, *Speciation du Mercure Dissous Dans les Eaux Marines: Analytical Methods for the Marine Environments*, Ifremer and French Ministry for Ecology and Sustainable Development Publication MA0303, Paris, France, 2003.
- [56] L. Sipos, H.W. Nürnberg, P. Valenta, M. Branica, *Anal. Chim. Acta* 115 (1980) 25–42.
- [57] Analytical Methods Committee, *Analyst* 112 (1987) 199–204.
- [58] J. Gong, T. Zhou, D. Song, L. Zhang, X. Hu, *Anal. Chem.* 82 (2010) 567–573.
- [59] X.-C. Fu, X. Chen, Z. Guo, C.-G. Xie, L.-T. Kong, J.-H. Liu, X.-J. Huang, *Anal. Chim. Acta* 685 (2011) 21–28.
- [60] C. Faller, N.Y. Stojko, G. Henze, K.Z. Brainina, *Anal. Chim. Acta* 396 (1999) 195–202.
- [61] G. Sandmann, H. Dietz, W. Plieth, *J. Electroanal. Chem.* 491 (2000) 78–86.
- [62] M. Ueda, H. Dietz, A. Anders, H. Knepe, A. Meixner, W. Plieth, *Electrochim. Acta* 48 (2002) 377–386.
- [63] A.J. Beck, Y. Tsukamoto, A. Tovar-Sanchez, M. Huerta-Diaz, H.J. Bokuniewicz, S.A. Sañudo-Wilhelmy, *Appl. Geochem.* 22 (2007) 477–490.
- [64] P. Ollivier, O. Radakovitch, B. Hamelin, *Chem. Geol.* 285 (2011) 15–31.
- [65] X.-C. Fu, X. Chen, Z. Guo, L.-T. Kong, J. Wang, J.-H. Liu, X.-J. Huang, *Electrochim. Acta* 56 (2010) 463–469.