

X-tox: a new family of putative antimicrobial protein specific to Lepidoptera

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Species-level phylogenetic hypotheses can be used to explore patterns of divergence and speciation. Warningly-colored mimetic butterflies interesting models, because their wing patterns are under strong selection, and might be involved in speciation. Here we present an almost complete phylogenetic hypothesis for neotropical mimetic butterfly genus Napeogenes (20 species), based on mitochondrial genes (entire cytochrome oxydase I and II) and nuclear genes (tektin, 715bp; and elongation factor 1a, 1028bp). Whenever possible, we included several subspecies of the same species (with different wing patterns). The results showed good congruence between the different genetic regions. In most cases individuals of the same species clustered together, supporting the current taxonomy. However, N. larina otaxes and N. larina aethra appeared to be two sister species rather than two subspecies. A combined evidence topology is presented based on a Bayesian analysis of all the genes regions. Using the phylogentic hypothesis we investigate whether changes in wing pattern were associated with speciation, and whether speciation was dominantly sympatric or allopatric. Using the phylogenetic hypothesis we explore patterns of diversification, and we investigate whether changes in altitudinal range and in wing pattern were associated with speciation.

X-tox: a new family of putative antimicrobial protein specific to Lepidoptera

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A new family of putative antimicrobial proteins, characterized by imperfectly conserved tandem repeats (up to 11) of cystein-stabilized alpha beta motifs (CS- $\alpha\beta$), a structural scaffold characteristic of invertebrate defensins and scorpion toxins, was identified through functional genomic approaches lepidopteran species Spodoptera frugiperda. Orthologs were also found in ESTs or genomic databases from other lepidopteran species (Bombyx mori, Galleria mellonella, Papillo dardanus), but not in other insect orders other invertebrate or metazoans, thus suggesting that this protein family is specific to the lepidopteran insects. In S. frugiperda as well as in B. mori, more than one transcript can be related to this protein family. In silico analysis of B. mori genome (the sole lepidopteran genome available to date) suggests that the three mRNA encoding X-tox proteins that were characterized in this insect are generated through a mechanism of alternative splicing. This mechanism thus allows the production of different X-tox proteins containing five or six CS-αβ motifs. S. frugiperda X-tox genomic organization is currently under investigation. The role of these proteins in lepidopteran immune response was investigated by transcriptional analyses with one of the S. frugiperda cDNAs. Results indicate that the corresponding gene, spod-11-tox, is expressed mainly in larval fat body and hemocytes and that transcription is enhanced in both immune tissues upon bacterial challenge. Phylogenetic analyses show that CS-αβ motifs-containing proteins are divided into three clusters. The first, considered as "ancestral" group, comprises molecules from different invertebrate taxa (odonate insect, arachnids and mollusks) and even a fungus molecule whereas the second group includes defensins isolated from neopteran insects. Finally, the third cluster contains all the CS- $\alpha\beta$ motifs stem from lepidopteran X-tox proteins. According to this phylogenetic analysis and to the position of lepidoptera in insect evolution (one of the latest order in insect speciation), we propose that X-tox proteins evolved from insect defensins to generate a new family of antimicrobial proteins. The main feature of this family is the multi-domain organisation who leads to molecular diversity generated through a mechanism of alternative splicing. The full characterization of this new protein family (gene organization and expression, protein structure and mechanism of action) in several lepidopteran species should contribute to solve the puzzle of the evolutionary history of arthropod defensins.

The role of serineprotease homolgue 3 (SPH-3) in *Manduca* sexta shown by RNA interference

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Photorhabdus luminescens lives in symbiosis with nematodes that invade insects. Following entry into the insect, the bacteria are released from the nematode gut into the open blood system of the insect. Here they secrete factors, which kill the host and also convert the host tissues into food for the replicating bacteria and nematodes. One of the secreted proteins is an rtx-like metalloprotease PrtA. Using a proteomic approach to determine the natural substrates of PrtA in host haemolymph, we identified a number of Manduca sexta haemolymph proteins that are selectively cleaved when plasma was incubated with purified PrtA. One of the PrtA protein targets in M. sexta haemolymph included the Serine Protease Homologue Protein 3 (SPH-3). We found that SPH-3 is not transcribed in the fat-body or haemocytes of naive M. sexta, but only when insects are exposed to bacteria, including Photorhabdus. We used RNA interference (RNAi)-mediated inhibition of expression through injection of double-stranded RNA of SPH-3 into M. sexta caterpillars and we achieved reduced transcription of SPH-3 in both fat-body and haemocyte tissues. Knock-down of SPH-3 dramatically reduced the ability of insects to resist infection to a standard dose of *Photorhabdus*, as measured by the rate at which infected insects die. RNAi of SPH-3 was also associated with decreased levels of phenoloxidase (PO) activity in haemolymph cell-free plasma from insects infected with either a non-pathogenic strain of E. coli or Photorhabdus. The reduction in PO activity was also reflected by a reduction in the number of melanotic nodules present in tissues of dissected larvae. Finally, we found that RNAi of SPH-3 did not affect the transcription of pattern

recognition protein (Hemolin, genes **Peptidoglycan** Immulectin-2, recognition protein, Pattern recognition Serine Proteinase, recognition protein-1, β -glucan β -glucan recognition protein-2), but down-regulated the transcription of certain antibacterial effector genes (Attacin, Cecropin, Moricin). Our findings strongly suggest that SPH-3 serves specific immune functions and plays a distinctive role in M. sexta immune pathways.

Host plant adaptation and specialization in the Pieridae family

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A metabolic diversion of a plant chemical defense caused by an enzyme (Nitrile Specifier Protein, NSP) enables a majority of Pieridae species to feed on Brassicaceous plants. The NSP is a newly recruited detoxifying protein in this family. It shows a specific repeat structure and has no sequence homology to any protein with a known function. We are trying to unravel the molecular mechanisms leading to the evolution of this protein and the subsequent adaptation to a plant defense system. The nucleotide sequence is known for two model species. By generating cDNA libraries of related species and amplifying intron and upstream regions, species and allelic differences are being located. Although NSP shows no sequence homology to any protein of known function, single domains show similarities to the so-called major allergen proteins, found in the gut lumen of many insect species. By establishing heterologous expression systems for both proteins we are trying to investigate the mode of action of NSP and the function of MA.

The effect of microorganisms in the food on the growth rate and immune system of cabbage looper (*Trichoplusia ni*)

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