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► To cite this version:

Julie Suzanne Grange, Hervé Rybarczyk, Aline Tribollet. The three steps of the carbonate biogenic dissolution process by microborers in coral reefs (New Caledonia). *Environmental Science and Pollution Research*, 2015, *Microbial Ecology of the Continental and Coastal Environments*, 22 (18), pp.13625-13637. 10.1007/s11356-014-4069-z . hal-01107579

HAL Id: hal-01107579

<https://hal.science/hal-01107579v1>

Submitted on 21 Jan 2015

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1 **The three steps of the carbonate biogenic dissolution process by microborers in coral reefs**
2 **(New Caledonia)**

3
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13
14 **Keywords** Microbioerosion dynamics; biogenic dissolution of calcium carbonate; euendoliths; succession of
15 microboring communities; coral reefs; New Caledonia

16
17 **Abstract**

18 Biogenic dissolution of carbonates by microborers is one of the main destructive forces in coral reefs and is
19 predicted to be enhanced by eutrophication and ocean acidification by 2100. The chlorophyte *Ostreobium* sp.,
20 the main agent of this process, has been reported to be one of the most responsive of all microboring species to
21 those environmental factors. However, very little is known about its recruitment, how it develops over
22 successions of microboring communities and how that influences rates of biogenic dissolution. Thus, an
23 experiment with dead coral blocks exposed to colonization by microborers was carried out on a reef in New
24 Caledonia over a year period. Each month, a few blocks were collected to study microboring communities and
25 the associated rates of biogenic dissolution. Our results showed a drastic shift in community species composition
26 between the 4th and 5th months of exposure, i.e. pioneer communities dominated by large chlorophytes such as
27 *Phaeophila* sp. were replaced by mature communities dominated by *Ostreobium* sp.. Prior the 4th month of
28 exposure, large chlorophytes were responsible of low rates of biogenic dissolution while during the community
29 shift, rates increased exponentially (x10). After 6 months of exposure, rates slowed down and reached a 'plateau'
30 with a mean of 0.93 kg of CaCO₃ dissolved per m² of reef after 12 months of exposure. Here we show that (a)
31 *Ostreobium* sp. settled down in new dead substrates as soon as the 3rd month of exposure but dominated
32 communities only after 5 months of exposure, and (b) microbioerosion dynamics comprise 3 distinct steps which
33 fully depend on community development stage and grazing pressure.

41 Introduction

42

43 Coral reefs are highly diverse and complex marine ecosystems, providing many services including commercial
44 fishing, tourism and coastal protection against storms (Knowlton et al. 2010). Their maintenance results from the
45 delicate balance between constructive forces, mainly calcification of corals and encrusting coralline algae, and
46 destructive forces, mostly bioerosion processes (Scoffin et al. 1980, Tribollet & Golubic 2011). Unfortunately,
47 this equilibrium is increasingly threatened by anthropogenic and environmental factors such as eutrophication,
48 overfishing, rising sea surface temperature and ocean acidification (Hoegh-Guldberg et al. 2007, Pandolfi et al.
49 2011, Huand 2012). To date, 20% of coral reefs have already disappeared and 20% show signs of imminent
50 extinction (Wilkinson 2008). Although both constructive and destructive forces are affected by anthropogenic
51 and climatic stresses (Anthony et al. 2008, Tribollet et al. 2009, Chauvin et al. 2011, Carreiro-Silva et al. 2012,
52 Andersson & Gledhill 2013, Jessen et al. 2014, Wisshak et al. 2014), much less attention has been given to
53 bioerosion than reef accretion.

54

55 Reef bioerosion results from the mechanical and/or chemical activity of living organisms (Neumann 1966)
56 comprising grazers (e.g. urchins and parrotfishes), macroborers (e.g. bivalves and sponges) and microborers or
57 euendoliths (cyanobacteria, algae and fungi) (Glynn 1997, Tribollet & Golubic 2011). Grazers abrade substrate
58 surfaces to feed on epilithic and euendolithic algae (Pari et al. 1998, Tribollet & Golubic 2005) while macro- and
59 micro-borers penetrate actively by mechanical and/or chemical means into substrates to find a shelter (Lazar &
60 Loya 1991, Zundevich et al. 2007, Garcia-Pichel et al. 2010, Tribollet & Golubic 2011). Previous studies
61 showed that microbioerosion in synergy with grazing is the main process driving reef bioerosion of dead
62 carbonates when exposed several months to three years to bioeroding agents (Chazottes et al. 1995, Tribollet &
63 Golubic 2005). Despite their microscopic size, euendoliths are probably the main agents of reef bioerosion as
64 they are able to colonize each available micrometer of carbonate surface area, -from carbonate sand to skeletons
65 of dead and live calcifiers such as corals, mollusks and crustose coralline algae-, and are able to dissolve as much
66 as 1.1 kg of CaCO₃ per linear m² of reef per year, which converts to 30 mmol CaCO₃.m².d⁻¹ (value obtained
67 from the highest rate of microbioerosion quantified in coral reefs and recalculated using coral microdensity and
68 porosity; see Tribollet & Golubic 2005). Chazottes et al. (1995) and Tribollet (2008a) highlighted that the
69 highest rates of carbonate dissolution result mostly from the activity of the chlorophytes of the genus
70 *Ostreobium*. Recently, experiments realized under controlled conditions or semi-controlled conditions over short
71 periods of time showed that ocean acidification and eutrophication enhance biogenic dissolution by stimulating
72 the activity of chlorophytes such as *Ostreobium* sp., suggesting an acceleration of reef framework degradation in
73 the near future (Chazottes et al. 2002, Tribollet et al. 2009, Carreiro-Silva et al. 2012, Reyes-Nivia et al. 2013,
74 Tribollet et al. 2014). These experiments strongly suggested the dependence of the response of biogenic
75 dissolution on species composition of microboring communities (see also Tribollet 2008a). To better understand
76 dynamics of the biogenic dissolution process under various environmental conditions, it is therefore essential to
77 determine simultaneously species composition of microboring communities, their successions over time and
78 their associated rates of biogenic dissolution under natural conditions. Interestingly these aspects have been
79 rarely studied limiting possibilities of modeling dynamics of the biogenic dissolution process for better
80 prediction of coral reef future.

81
82 Very few studies have indeed investigated changes in euendolithic assemblages at different time scales and the
83 influence of such changes on rates of carbonate biogenic dissolution. Le Campion-Alsumard (1975) was first to
84 report successions of euendolithic communities in exposed calcite spar in the Mediterranean Sea over a period of
85 one month. She showed that colonization of calcite by euendoliths starts after 8 days of exposure and that the
86 early colonizers are cyanobacteria such as *Mastigocoleus testarum*. Unfortunately, she did not quantify rates of
87 carbonate dissolution. Later, Kiene et al. (1995), Gektidis (1999) and Vogel et al. (2000) reported successions of
88 euendolithic communities in various reef carbonate substrates after 1, 3, 6, 12, 24 months and very scarce
89 information on rates of biogenic dissolution associated with those communities. They confirmed that pioneer
90 euendolithic communities, also called ‘immature communities’, comprise short lived cyanobacteria such as *M.*
91 *testarum* and the chlorophyte *Phaeophila* sp., and that the long-lived euendolithic chlorophyte *Ostreobium* sp.
92 appears later to dominate communities after 1 or 2 years of exposure (‘mature communities’). Chazottes et al.
93 (1995), Tribollet et al. (2006) and Tribollet (2008a) specified later that euendolithic communities in coral reefs
94 become mature at least after 6-12 months of exposure. However, the exact period of settlement of *Ostreobium*
95 and when it becomes dominant remain unknown. Among all those studies, only Chazottes et al. (1995) and
96 Tribollet (2008a) studied and reported simultaneously species composition of euendolithic communities and
97 rates of reef biogenic dissolution over time, i.e. after 2, 6, 12, 24 months and 12 and 36 months, respectively.
98 They reported that biogenic dissolution does not vary linearly nor proportionally over time, however those
99 authors did not present the full dynamics of the process. Thus, to better understand dynamics of biogenic
100 dissolution of carbonates by euendoliths, we carried out an experiment with a high temporal resolution on a reef
101 in New Caledonia. Experimental coral blocks were exposed to colonization by euendoliths and were recovered
102 every month over a year period. The experiment did not last longer to avoid colonization by macroboring
103 organisms (see review by Tribollet & Golubic 2011) but was long enough to report interactions between
104 euendoliths and grazers. The specific goals of the present study were to determine (1) when *Ostreobium* sp.
105 settles on dead carbonate substrates newly available, (2) when exactly it dominates communities, (3) how
106 euendolithic communities succeed to each other over a year period with a monthly resolution, and (4) how does
107 that influence biogenic dissolution. In addition we quantified grazing pressure and how it influenced biogenic
108 dissolution. Then, the first model of the biogenic dissolution process dynamics was developed.

109
110

111 **Material and Methods**

112

113 Site and experimental design description

114 This study was carried out on the coral reef of the Island ‘Ile aux Canards’ in New Caledonia (lat. 22°31’S, long.
115 166°43’E, Fig. 1a, b) located in the lagoon, a few hundred meters away from Nouméa city (South province of the
116 Island Grande Terre). This site was selected because of its easy access by boat from the laboratory (Center of the
117 Institut de Recherche pour le Développement), it presents a well developed reef which is relatively healthy
118 although it is impacted by anthropogenic activities such as tourism. At this site, the tide amplitude is low (~1.5
119 m) and seawater is weakly turbid over the year due to the hydrodynamism occurring around the island. Local
120 currents resuspend regularly carbonate sediments and organic matter. The mean seawater temperature during the

121 hot season is 26 ± 2 C° (November-April) and 23 ± 1 C° during the cold season (May-October). Seawater
122 salinity varies between 35.1 and 35.8 over a year period (monthly measurements with CTD sensors on the reef
123 during the experiment).

124 To determine successions of euendolithic communities and to quantify associated biogenic dissolution and
125 grazing rates, 60 experimental coral blocks were fixed on two steel grids (1 m x 1 m) affixed on the reef at 3 m
126 depth on the 15th of December 2010, i.e. during the austral summer season (Fig. 1c). Experimental coral blocks
127 (~1.5 cm x 2 cm x 2 cm) were cut from the interior of a live colony of the massive coral *Porites* sp. using a band
128 saw. *Porites* skeletons have been used in several previous bioerosion studies thus allowing for comparison of
129 results (Kiene & Hutchings 1994, Chazottes et al. 1995, Pari et al. 1998, Edinger et al. 2000, Schonberg 2001,
130 Chazottes et al. 2002, Tribollet & Golubic 2005, Tribollet 2008a). Micro- and bulk density of the coral skeleton
131 were measured using the buoyant weight technique as described by Bucher et al. (1998). Between January and
132 December 2011, 3 coral blocks were collected every month and were preserved in a buffered 4% solution of
133 formaldehyde in seawater.

134
135 Sample analyses

136 Colonized blocks were cut in half perpendicular to the upper side. The first halves were used to estimate the
137 bioeroded surface area by euendoliths (in cm²) on the upper side, by taking pictures under a Scanning Electronic
138 Microscope (SEM, Zeiss EVO LS15, Alizés Platform, IRD, Bondy, France). Twenty pictures were taken per
139 coral block ($n = 60$ per month) at panoramic magnification x1500, and were analyzed using the software ImageJ.
140 The second halves of blocks were used to prepare thin sections according to the technique described by Golubic
141 et al. (1970), and Tribollet et al. (2002). These thin sections were observed under a light microscope (Nikon
142 Eclipse LV100) to allow the determination of (1) the species composition of euendolithic communities, (2) the
143 relative abundance (RA) of living filaments inside substrates ($n > 120$ measurements per month) expressed in %
144 (\pm the confidence interval), and (3) the mean depth of penetration (P_{80}) in cm at which 80%-90% of the living
145 euendolithic filaments could be observed inside coral blocks ($n > 120$ measurements per month; see technique
146 description in Tribollet et al. 2009). To complete the identification of euendoliths, small pieces of sample were
147 decalcified (HCl < 10%) allowing the observations of a few euendolithic filaments at a time. Identification was
148 based on morphological criteria with the help of Stjepko Golubic, and according to Bornet and Flahault (1889)
149 and Wisshak et al. (2011). Only species with relative abundance higher than 5% are presented in this paper.
150 The quantity of carbonate dissolved per month, called here ‘biogenic dissolution’ (BD), was calculated using the
151 following equation (adapted from Tribollet et al. 2009):

$$152 \quad BD = \frac{\text{Depth of penetration} \times \text{Bioeroded surface area} \times \text{Density}}{\text{Surface area of blocks}}$$

153
154 where ‘BD’ is expressed in g of CaCO₃ dissolved per linear square meter of reef after a certain time of exposure
155 (e.g. one month of exposure for blocks colonized by euendoliths during one month), ‘depth of penetration’ is P_{80}
156 in cm, ‘bioeroded surface area’ is the bioeroded surface area of block surfaces exposed to colonization by
157 euendoliths multiplied by (100 - (% of porosity of coral skeleton) in cm² (here we considered only 5 sides per
158 block as the bottom side was exposed to a different light regime and porosity was 53%), ‘density’ is the coral
159 skeleton microdensity (2.58 ± 0.06 g.cm⁻³) as filaments of euendoliths grow into the microstructure of the

160 skeleton, not in its void spaces, and ‘surface area of block’ is the surface area of the 5 sides of each block
161 exposed to colonization (the bottom side was discarded). We assumed here that the measured bioeroded surface
162 area and P_{80} on the upper sides of blocks were representative of the 5 sides as blocks were small enough to be
163 exposed to the same light intensity.

164
165 By knowing the initial dimensions of coral blocks, it was also possible to estimate the percentage of carbonate
166 removed by grazers on each sample, by analyzing pictures of thin sections taken by a high resolution Nikon
167 camera (Nikon Camera Digital D5100 with AF-S Micro Nikkor 60mm objective) using the software imageJ ($n >$
168 12 measurements per month). Grazing rates were then calculated using the following equation:

$$\text{Grazing} = \frac{\text{Percent of grazing} \times \text{Volume of blocks} \times \text{Density}}{\text{Surface area of blocks}}$$

170
171 where ‘grazing’ is expressed in g of CaCO_3 eroded per linear square meter of reef after a certain time of
172 exposure (i.e. xx months), ‘volume of block’ is expressed in cm^3 (initial length x height x width), ‘density’ is the
173 bulk density of the coral skeleton in g.cm^{-3} ($1.86 \pm 0.1 \text{ g.cm}^{-3}$) as grazers abrade simultaneously voids and
174 skeleton, and ‘surface area of block’ is similar to that used for BD, in cm^2 .

175
176 Statistical analysis

177 Statistical analyzes were performed under the R statistical framework (R. Development Core Team 2014). All
178 data sets were analyzed with non-parametric tests because homogeneity of variance and normality distribution
179 were not met (Bartlett test and Shapiro test, respectively) even after data transformation (Hollander et al. 2014).
180 The Kruskal-Wallis test (with $\alpha = 5 \%$) was used to determine if measured variables (i.e. relative abundance and
181 depth of penetration of euendolithic filaments, biogenic dissolution, and grazing) varied significantly among the
182 different durations of exposure (from 1 month to 12 months). When the Kruskal-Wallis test was significant a
183 pairwise post-hoc analysis of Mann and Whitney was realized using a correction of Bonferroni (Hollander et al.
184 2014). To fit biogenic dissolution values and to obtain a rate of the biogenic dissolution after 12 months of
185 exposure, we used a logistic growth model in R statistical framework with the nls2 library (Grothendieck 2013):

$$\frac{A}{(1 + e^{X-t})/S}$$

187
188 where ‘A’ is the maximum asymptotic value, ‘X’ is the time where A/2 is observed, ‘S’ is the maximum slope
189 and ‘t’ is the time of exposure. To see possible correlation between grazing and biogenic dissolution, the
190 correlation of Spearman was used (Hollander et al. 2014). Finally to determine the main variables driving
191 biogenic dissolution dynamics over a year period, a hierarchical cluster analysis was performed based on
192 dissimilarity matrix using Bray-Curtis distances (Murtagh & Legendre 2014). A non-metric multidimensional
193 scaling analysis (nMDS) was performed using this matrix as well as a hierarchical cluster analysis to graphically
194 represent results (Faith et al. 1987).

195
196

197 **Results**

198

199 Euendolithic species

200 Euendoliths observed in the experimental coral blocks comprised (i) chlorophytes including *Phaeophila* sp. (Fig.
201 2a), *Eugomontia* sp., (Fig. 2b) and *Ostreobium* sp. (Fig. 2c), (ii) two cyanobacteria comprising *Mastigocoleus*
202 *testarum* (Fig. 2d) and *Plectonema terebrans* (Fig. 2e), and (iii) unidentified fungi (Fig. 2f). Due to relative
203 morphological similarities between *Eugomontia* sp. and *Phaeophila* sp. and difficulties sometimes to distinguish
204 these two species from unidentified large chlorophytes, all these euendoliths were grouped in one category
205 named ‘unidentified chlorophytes’. *Ostreobium* sp. was easily identified as this chlorophyte presents siphonous
206 and polymorphic filaments. *Plectonema terebrans* and fungi were also grouped in one category named ‘thin
207 filaments’ as the diameter of their filaments did not exceed 1-2 μm and cells in some filaments of *P. terebrans*
208 were difficult to distinguish. Moreover perpendicular branches and reproductive organs of fungi were usually
209 absent rendering difficult their differentiation from *P. terebrans* filaments.

210

211 Successions of communities: abundance and distribution of euendoliths

212 After one month of exposure, euendolithic communities were mostly composed of unidentified chlorophytes
213 even though thin filaments and the cyanobacterium *M. testarum* were observed (mean RA of 83% for
214 unidentified chlorophytes, Fig. 3a). Filaments of *M. testarum* and *P. terebrans* as well as fungi were observed in
215 all blocks during the overall duration of the experiment but their mean relative abundance never exceeded 20%
216 (Fig. 3c,d). The cyanobacterium *Hyella* sp. was also observed in one month-old blocks but was very rare (RA <
217 0.5%). In 2 and 3 months-old blocks, the unidentified chlorophytes remained dominant with a mean RA of 52%
218 (Fig. 3a). About 2% can be attributable to *M. testarum*, 0.5% to thin filaments and the rest to *O. quekettii*. The
219 unidentified chlorophytes never penetrated deeper than 0.5 mm into coral skeletons (Fig. 3a). Similarly filaments
220 of *M. testarum* were abundant beneath the surface of substrates although a few filaments could be observed
221 down to 2 mm (their relative abundance at this depth was less than 2%; Fig. 3d). Between 4 and 6 months, the
222 relative abundance of unidentified chlorophytes and *O. quekettii* was inversely correlated (Spearman test, $r = -$
223 0.81, $p < 0.01$), with RA of unidentified chlorophytes reaching less than 1% after 6 months of exposure (Fig. 3a)
224 and RA of *O. quekettii* 42% (Fig. 3b). After more than 6 months of exposure, *O. quekettii* dominated
225 euendolithic communities. Its relative abundance was maximal (> 70%) after 11 months of exposure (Fig. 3b).
226 The rest of the communities comprised thin filaments (13%) and rare filaments of *M. testarum* (4%). *O. quekettii*
227 and thin filaments could penetrate deep into coral skeletons reaching sometimes more than 4 mm (Fig. 3b, c). *O.*
228 *quekettii* was the most abundant species between 0.5 mm and 4 mm below the substrate surface with a mean RA
229 of 43% (Fig. 3b).

230

231 Surface area bioeroded and depth of penetration (P_{80})

232 After one month of exposure, the mean surface area bioeroded by euendoliths was about $17.6\% \pm 2.1\%$. During
233 the first four months of exposure, the surface area bioeroded increased significantly (Mann and Whitney, $p < 0.01$;
234 Table 1) and reached a mean value of 40%. After 4 months of exposure, the surface area bioeroded was 38% on
235 average, except between 7 and 9 months of exposure (lowest mean quantified, 28%).

236 The mean depth of penetration of living euendolithic filaments (P_{80}) increased slowly during the first four
237 months of exposure (Table 1). Between the 4th and 7th months of exposure, P_{80} increased exponentially and
238 reached a maximum mean value of 3.19 ± 0.19 mm after 7 months of exposure. Finally, after 8 months of
239 exposure, depths of penetration tend to stabilize with a mean value of 2.55 ± 0.20 mm.

240

241 Biogenic dissolution rates

242 The quantity of dissolved calcium carbonate by euendoliths increased significantly over time (Mann and
243 Whitney, $p < 0.001$) but non-linearly (Fig. 4a). The dynamics of the biogenic dissolution process presented indeed
244 3 steps. First, between 1 and 3 months of exposure, biogenic dissolution increased slowly reaching 66 ± 0.8 g of
245 dissolved per square meter of reef after 3 months of exposure (Fig. 4a). Within this period of 2 months, the mean
246 rate of dissolved CaCO_3 was $30 \text{ g.m}^{-2}.\text{mo}^{-1}$. Second, between 4 and 5 months of exposure, biogenic dissolution
247 increased exponentially reaching a mean of $739 \pm 11 \text{ g.m}^{-2}$ after 5 months of exposure. During this short period
248 of time, the mean rate of biogenic dissolution was $564 \text{ g.m}^{-2}.\text{mo}^{-1}$. Finally after 6 months of exposure, the
249 process slowed down dramatically. Biogenic dissolution reached a 'plateau' with a mean value varying between
250 742 and 1134 g.m^{-2} after 8 months and 11 months, respectively (Fig. 4a); the mean rate of biogenic dissolution
251 was $23 \text{ g.m}^{-2}.\text{mo}^{-1}$ for the period comprised between 7 and 12 months of exposure. This process, as described
252 above, is typically a logistic growth model in three steps. A preliminary model was then developed to highlight
253 the process kinetics and to estimate the mean rate of biogenic dissolution for a 12 months period (Fig. 5). The
254 expression of a logistic model growth was used in R to fit our biogenic dissolution dataset. The asymptotic value
255 or the maximum value of biogenic dissolution was $0.91 \pm 0.04 \text{ kg.m}^{-2}$; half of the biogenic dissolution was
256 realized after 4.5 ± 0.2 months and the maximum of slope was $0.37 \pm 0.15 \text{ kg.m}^{-2}.\text{mo}^{-1}$. The 3 steps-process was
257 confirmed by the non-metric multidimensional scaling (nMDS) with the hierarchical cluster analysis (Fig. 6).
258 The low value of stress indicates that the representation is reliable: the distance between studied months (circle)
259 correlate well with the dissimilarity of the three clusters (ellipses). In addition, the nMDS showed how the
260 different measured biological parameters influenced each step. The early step of biogenic dissolution appeared to
261 be driven by the abundance of unidentified chlorophytes while the second step seemed to be related to the shift
262 in community species composition. In contrast, the final step was clearly driven by the abundance of the
263 chlorophyte *Ostreobium quekettii* and the intensity of grazing.

264

265 Grazing pressure

266 Based on observations of block surfaces under a binocular microscope, grazing traces appeared to be due mostly
267 to parrotfishes, confirming a few *in situ* observations during block collection. Between 1 and 7 months of
268 exposure, grazing increased almost linearly (from 35 ± 11 to 365 ± 51 g of CaCO_3 eroded per square meter of
269 reef, respectively; Fig. 4b). Except for the two peaks at 8 and 10 months of exposure (grazing rate $> 600 \text{ g.m}^{-2}$),
270 after 7 months of exposure grazing rate values oscillated around the mean obtained after 12 months of exposure,
271 i.e. $339 \pm 81 \text{ g.m}^{-2}$ (Fig. 4b). Biogenic dissolution and grazing were strongly negatively correlated after 7 months
272 of exposure (Spearman correlation, $r = -0.92$, $p < 0.01$).

273

274

275 Discussion

276

277 The present study of euendolithic community successions with a monthly resolution reveals the three main steps
278 in the colonization of newly available dead carbonate substrates: (i) an early community development stage
279 between 0 and 3 months where large chlorophytes such as *Phaeophila* sp. are dominant, (ii) an intermediate
280 stage between 3 and 6 months where the chlorophyte *Ostreobium* sp. the main agent of biogenic dissolution,
281 starts to settle down and to dominate communities, and (iii) a final stage, after more than 6 months of exposure,
282 where *Ostreobium* sp. dominates largely microboring communities. The present work confirms that colonization
283 by euendoliths of newly available carbonate substrates is rapid (a few days or weeks) and is dominated by large
284 pioneer chlorophytes, especially the green alga *Phaeophila* sp., and cyanobacteria such as *Mastigocoleus*
285 *testarum* (see Chazottes et al. 1995, Kiene et al. 1995, Le Campion-Alsumard et al. 1995, Gektidis 1999, Vogel
286 et al. 2000, Tribollet et al. 2006, Tribollet 2008b). Those communities are characteristic of ‘immature
287 communities’ (Gektidis 1999). Here, the relative abundance of large unidentified chlorophytes, comprising
288 mostly *Phaeophila* filaments, was higher than 60% during the first 4 months of exposure which is similar to the
289 findings of Kiene et al. (1995) and Vogel et al. (2000) in limestone in Bahamian and Australian reefs. Although
290 Chazottes et al. (1995) also found *Phaeophila* in abundance in one-month old coral blocks in French Polynesia,
291 the relative abundance of this species in communities decreased rapidly after the second month of exposure. This
292 could result from the intense grazing quantified on blocks in French Polynesia which certainly limited the
293 development of epiliths and *Phaeophila*, and postponed *Ostreobium* settlement (seen after 6 months of
294 exposure). In the present study, significant traces of grazing were only observed on blocks after a few months of
295 exposure suggesting that epilithic and euendolithic communities developed slowly on the studied reef in New
296 Caledonia compared to those growing in French Polynesian reefs. Grazers are indeed not attracted by substrates
297 until epilithic (mostly turfs) and euendolithic communities on which they feed, start to be well developed
298 (Bruggemann et al. 1994). Interestingly in the present study, *Ostreobium* sp. started settling on new dead coral
299 skeletons as soon as the 3rd month of exposure (but represented less than 5% of euendolithic communities). In
300 mollusc shells Carreiro-Silva et al. (2005) also observed *Ostreobium* filaments after 49 days. To the contrary of
301 what was suggested in previous studies, this green euendolithic alga appears thus as an early colonist of dead
302 carbonate substrates in shallow reefs. Most studies have indeed reported the presence of *Ostreobium* sp. in dead
303 substrates comprising limestone, calcite spar, mollusc shells and coral skeletons, only after more than 6-12
304 months of exposure (Chazottes et al. 1995, Kiene et al. 1995, Le Campion-Alsumard et al. 1995, Gektidis 1999,
305 Vogel et al. 2000, Tribollet et al. 2006, Tribollet 2008a).

306

307 All pioneer euendolithic species were located within the first few hundred micrometers beneath the substrate
308 surface, with the exception of a few filaments of *M. testarum* seen down to 2 mm below the surface. These
309 species are indeed known as photophile euendoliths which do not penetrate deep into substrates as they require
310 high light intensities to insure photosynthesis (see review by Tribollet 2008b). In contrast, *Ostreobium* is a
311 sciaphile or oligophotic euendolith, i.e. that it is adapted to low light intensities (Fork & Larkum 1989, Tribollet
312 et al. 2006), and thus, can penetrate deep inside substrates (max depth > 4 mm after 6 months of exposure) and
313 presents a large bathymetric distribution (found down to 300 m depth; Lukas (1978); Kiene et al. (1995); Radtke
314 et al. 1996). It can also be easily photoinhibited by sudden important light intensities like in the case of a rapid
315 bleaching event in live corals (Fine et al. 2005). Its light regime requirements may thus explain why it was not

316 observed on one month-old blocks. Like suggested by Vogel et al. (2000), we strongly emphasize that
317 *Ostreobium* can only settle on substrates once they are colonized by epilithic organisms (biofilms, and then turfs
318 at the early stages) and pioneer euendoliths, which provide shade (see also Radtke et al. 1996). As soon as those
319 conditions were met in the present experiment, *Ostreobium* started rapidly to dominate communities. The shift in
320 euendolithic community species composition in favour of *Ostreobium* occurred between the 4th and 5th months of
321 exposure. Although such shift was suggested previously by Chazottes et al. (1995) and Kiene et al. (1995), its
322 period of occurrence was not highlighted due to the temporal resolution selected in each experiment (2-3 months
323 and then, 6 months or more). Here, after 6 months of exposure, the relative abundance of *Ostreobium* reached
324 more than 45% while that of large unidentified chlorophytes (mostly *Phaeophila*) dropped down to 1%. The
325 outcompetition between *Ostreobium* and *Phaeophila* was first reported by Le Campion-Alsumard et al. (1995) in
326 shallow waters where light intensity is high. The other species observed in the present study after 6 months of
327 exposure were essentially the cyanobacterium *P. terebrans* and fungi. The presence of these species in addition
328 to *Ostreobium* characterizes ‘mature communities’ (Chazottes et al. 1995, Kiene et al. 1995, Gektidis 1999,
329 Vogel et al. 2000, Tribollet 2008a). Such community species composition seems to be maintained after 12
330 months of exposure (Chazottes et al. 1995, Kiene et al. 1995, Tribollet 2008a) unless a major event occurs. An
331 extreme grazing pressure for instance, can modify dramatically community assemblages by removing most of
332 settled euendolithic filaments in substrates allowing more light to penetrate inside the carbonate structure. The
333 remaining filaments can then grow inside substrates until they reach their new depth of compensation
334 (photosynthesis = respiration) while new pioneer euendoliths colonize substrate surfaces (Schneider & Torunski
335 1983, Chazottes et al. 1995). In general, grazers do not remove all euendolithic filaments inside coral skeleton
336 allowing *Ostreobium* to remain the dominant species in substrates exposed more than 6 months to colonization,
337 but are sufficient to offer new surfaces available to colonization by cyanobacteria such as *Hyella* sp. and *M.*
338 *testarum*. This is why these pioneer species are often observed within mature communities, but in relatively low
339 abundance (less than 0.5% for *Hyella* sp. and 20% for *M. Testarum* here; see also Chazottes et al. 1995, Tribollet
340 et al. 2006, and Tribollet 2008a).

341
342 With a monthly resolution, we highlight how much the dynamics of euendolithic communities and grazing
343 pressure controlled the process of biogenic dissolution over a year period. First of all, pioneer communities
344 within the first 3 months of exposure drove a slow process of biogenic dissolution with a mean rate of 30 g.m⁻²
345 .mo⁻¹ although the amount of dissolved CaCO₃ after 3 months of exposure was 66 g.m⁻², which is similar to
346 what found Chazottes et al. (1995), and Peyrot-Clausade et al. (1995) in 2 months-old blocks in French
347 Polynesia (~73 g.m⁻² after recalculation with a microdensity for *Porites* coral skeleton of 2.58 g.cm⁻³ and a
348 porosity of 53%). This was due to the low depth of penetration of euendolithic filaments at this early stage of
349 community development (< 400 μm), and not to the colonization of surfaces which was already important (>
350 30% like in Chazottes et al. 1995) and the large diameter of pioneer euendolithic filaments (> 10 μm). Carbonate
351 biogenic dissolution became a very efficient process during the community shift with a mean rate of 564 g.m⁻²
352 .mo⁻¹ (rate obtained the 4th and 5th month of exposure). The rate of the biogenic dissolution increased
353 exponentially during this short period of time (one month) and was multiplied by a factor 10. This resulted from
354 the 3D network development of *Ostreobium* filaments within substrates (increase of filament abundance and
355 depth of penetration) as described above. Once *Ostreobium*’s population became stable after 6 months of

356 exposure, -i.e. *Ostreobium* filaments reached their depth of compensation (about 4 mm which is similar to that
357 found by Tribollet 2008a)-, the mean rate of biogenic dissolution slowed down and reached a 'plateau' ($23 \text{ g.m}^{-2}.\text{mo}^{-1}$).
358 Interestingly the estimated average rate of biogenic dissolution provided by the model after 12 months of
359 exposure ($0.91 \text{ kg m}^{-2}.\text{y}^{-1}$) was of the same order to rates estimated on offshore reefs on the Great Barrier Reef
360 after one year of exposure, although those reefs were more influenced by oceanic conditions than the present
361 studied site (0.5 to $1.1 \text{ kg m}^{-2}.\text{y}^{-1}$ after recalculation with microdensity and porosity of *Porites* skeletons, Tribollet
362 & Golubic 2005). Those rates are comparable as the same techniques were used for their quantification (SEM
363 picture analysis and thin sections; see Tribollet 2008a). Tribollet (2008a)'s methodology, although it prevents
364 from confounding initial microbioerosion like in Chazottes et al. (1995) and avoids subjective estimations like in
365 Vogel et al. (2000) and Carreiro-Silva et al. (2005), allows only the quantification of the amount of CaCO_3
366 dissolved by euendoliths alive at the time of collection. It is like an instantaneous 'snap shot' of a community
367 activity at a given time. It cannot provide the cumulative effects of euendolithic communities on carbonate
368 substrates over a given time period and does not reflect the potential long term impact of abiotic factors. The
369 survey of biogenic dissolution rates at different periods of time like in the present study allows however,
370 determining the possible impacts of biotic and abiotic factors on the process. Here, biogenic dissolution variation
371 within the last 7 months of the experiment resulted from grazing activity as a strong inverse correlation between
372 the two processes was found during this period. Grazing was especially important after 8 and 10 months of
373 exposure reducing biogenic dissolution by about 30% (the 'residual biogenic dissolution' was thus measured; see
374 definition in Chazottes et al. 1995). This synergy between grazers and euendoliths has first been well described
375 by Schneider and Torunski (1983) in the Adriatic Sea and later by Chazottes et al. (1995) in coral reefs. It has
376 been reported as the main bioerosion process in dead substrates after more than 12 months by Tribollet and
377 Golubic (2005). Here, it also appears as the main bioerosion process on dead carbonates exposed 6 to 12 months
378 to colonization.

379
380 In conclusion, the present experiment revealed the 3 steps in the process of biogenic dissolution over a year
381 period confirming that it is a neither linear, nor proportional process over time. It also suggested that rates
382 estimated after 12 months of exposure should be considered as conservative rates (probably minimum values).
383 This is the first time that such pattern is highlighted under natural conditions with a moderate grazing pressure.
384 Due to the experimental design, temporal resolution, and biotic conditions (intense grazing), Chazottes et al.
385 (1995) could only report part of the kinetics of the process. The present study also highlights that the biogenic
386 dissolution process is particularly efficient during the community shift between the 4th and the 5th month of
387 exposure and after a grazing episode (similar slopes). It took indeed about one month for the remaining
388 *Ostreobium* filaments to reach again their depth of compensation as the amount of calcium carbonate dissolved
389 per square meter after a high grazing episode (e.g. 8 months) was of the same order to that estimated prior to
390 grazing (Fig. 7). This fast growth in depth inside substrates may be the result of the avoidance of an increased
391 light intensity by *Ostreobium* filaments as epiliths and euendoliths near the surface have been scrapped off by
392 grazers. This reinforces the idea that the chlorophyte *Ostreobium* is a very efficient bioeroding microorganism in
393 coral reef systems. It should be noticed here however, that euendolithic community dynamics over 12 months
394 were studied starting in the summer season (December). Biofilms and epilithic covers may develop slowly in
395 winter due to reduce light intensity and seawater temperature, and thus may slow down euendolith settlement

396 (especially that of *Ostreobium*), successions and therefore, biogenic dissolution. This should be investigated over
397 several years to highlight possible seasonal and inter-annual variations. Such data would definitely improve our
398 capacity to better predict the response of biogenic dissolution to climatic and anthropogenic factors, and
399 therefore to better evaluate coral reef future by 2100.

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402 **Acknowledgements**

403 We would like to deeply thank John Butscher and the diving team of IRD located in New Caledonia (Center of
404 IRD, Nouméa), especially Armelle Renaud and Bertrand Bourgeois, for helping us on the field. We thank Cécile
405 Dupouy for *in situ* CTD measurements. We thank Sandrine Caquineau for helping using the Scanning Electronic
406 Microscope on the ALIZES facility (IRD-UPMC), which was funded by the Région Ile-de-France. We also
407 thank Stjepko Golubic for discussions and help with euendolith identification. Finally, we thank the Institut
408 National des Sciences de l'Univers (INSU) – EC2CO (Microbien and PNEC programs), the Institut de
409 Recherche pour le Développement, and the Grand Observatoire de l'Environnement et de la Diversité Terrestre
410 et Marine du Pacifique Sud (GOPS) for funding this project.

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413 **References**

- 414 Andersson AJ, Gledhill D (2013): Ocean acidification and coral reefs: Effects on breakdown, dissolution, and
415 net ecosystem calcification. *Annual Review of Marine Science* 5, 321-348
- 416 Anthony KRN, Kline DI, Diaz-Pulido G, Dove S, Hoegh-Guldberg O (2008): Ocean acidification causes
417 bleaching and productivity loss in coral reef builders. *Proceedings of the National Academy of Sciences*
418 of the United States of America 105, 17442-17446
- 419 Bornet E, Flahault C (1889): Sur quelques plantes vivant dans le test calcaire des mollusques. *Bulletin de la*
420 *Société Botanique de France* 36, CXLVII-CLXXVI
- 421 Bruggemann JH, Vanoppen MJH, Breeman AM (1994): Foraging by the stoplight Parrotfish *Sparisoma viride*.
422 1. Food selection in different, socially determined habitats. *Marine Ecology Progress Series* 106, 41-55
- 423 Bucher DJ, Harriott VJ, Roberts LG (1998): Skeletal micro-density, porosity and bulk density of acroporid
424 corals. *Journal of Experimental Marine Biology and Ecology* 228, 117-136
- 425 Carreiro-Silva M, McClanahan TR, Kiene WE (2005): The role of inorganic nutrients and herbivory in
426 controlling microbioerosion of carbonate substratum. *Coral Reefs* 24, 214-221
- 427 Carreiro-Silva M, Kiene WE, Golubic S, McClanahan TR (2012): Phosphorus and nitrogen effects on microbial
428 euendolithic communities and their bioerosion rates. *Marine Pollution Bulletin* 64, 602-613
- 429 Chauvin A, Denis V, Cuet P (2011): Is the response of coral calcification to seawater acidification related to
430 nutrient loading? *Coral Reefs* 30, 911-923
- 431 Chazottes V, Le Campion-Alsumard T, Peyrot-Clausade M (1995): Bioerosion rates on coral reefs: interactions
432 between macroborers, microborers and grazers (Moorea, French Polynesia). *Palaeogeography,*
433 *Palaeoclimatology, Palaeoecology* 113, 189-198
- 434 Chazottes V, Le Campion-Alsumard T, Peyrot-Clausade M, Cuet P (2002): The effects of eutrophication-related
435 alterations to coral reef communities on agents and rates of bioerosion (Reunion Island, Indian Ocean).
436 *Coral Reefs* 21, 375-390
- 437 Edinger EN, Limmon GV, Jompa J, Widjatmoko W, Heikoop JM, Risk MJ (2000): Normal coral growth rates
438 on dying reefs: Are coral growth rates good indicators of reef health? *Marine Pollution Bulletin* 40,
439 404-425
- 440 Faith DP, Minchin PR, Belbin L (1987): Compositional dissimilarity as a robust measure of ecological distance.
441 *Vegetatio* 69, 57-68
- 442 Fine M, Meroz-Fine E, Hoegh-Guldberg O (2005): Tolerance of endolithic algae to elevated temperature and
443 light in the coral *Montipora monasteriata* from the southern Great Barrier Reef. *Journal of*
444 *Experimental Biology* 208, 75-81
- 445 Fork DC, Larkum AWD (1989): Light harvesting in green-alga *Ostreobium* sp., a coral symbiont adapted to
446 extreme shade. *Marine Biology* 103, 381-385

447 Garcia-Pichel F, Ramirez-Reinat E, Gao QJ (2010): Microbial excavation of solid carbonates powered by P-type
448 ATPase-mediated transcellular Ca²⁺ transport. Proceedings of the National Academy of Sciences of the
449 United States of America 107, 21749-21754

450 Gektidis M (1999): Development of microbial euendolithic communities: The influence of light and time.
451 Bulletin of the Geological Society of Denmark 45, 147-150

452 Glynn PW (1997): Bioerosion and coral reef growth: a dynamic balance. In: Birkeland C (Editor), Life and
453 Death Of Coral Reefs. Chapman and Hall, USA, pp. 68-98

454 Golubic S, Brent G, Le Campion-Alsumard T (1970): Scanning electron microscopy of endolithic algae and
455 fungi using a multipurpose casting-embedding technique. Lethaia 3, 203-209

456 Grothendieck G (2013): nls2: Non-linear regression with brute force

457 Hoegh-Guldberg O, Mumby PJ, Hooten AJ, Steneck RS, Greenfield P, Gomez E, Harvell CD, Sale PF, Edwards
458 AJ, Caldeira K, Knowlton N, Eakin CM, Iglesias-Prieto R, Muthiga N, Bradbury RH, Dubi A,
459 Hatzitolos ME (2007): Coral reefs under rapid climate change and ocean acidification. Science 318,
460 1737-1742

461 Hollander M, Douglas AV, Chicken E (2014): Nonparametric statistical methods, Hoboken, New Jersey

462 Huand D (2012): Threatened reef corals of the World. Plos One 7

463 Jessen C, Voolstra CR, Wild C (2014): *In situ* effects of simulated overfishing and eutrophication on settlement
464 of benthic coral reef invertebrates in the Central Red Sea. PeerJ 2:e339

465 Kiene W, Radtke G, Gektidis M, Golubic S, Vogel K (1995): Factors controlling the distribution of microborers
466 in Bahamian Reef environments. In: Schuhmacher H, Kiene W , Dullo WC (Editors), Factors
467 controlling Holocene reef growth: An interdisciplinary approach. Facies, pp. 174-188

468 Kiene WE, Hutchings PA (1994): Bioerosion experiments at Lizard-Island, Great Barrier Reef. Coral Reefs 13,
469 91-98

470 Knowlton N, Brainard RE, Fisher R, Moews M, Plaisance L, Caley MJ (2010): Coral reef biodiversity. In:
471 McIntyre A (Editor), Life in the World's Oceans: Diversity, Distribution, and Abundance. Wiley-
472 Blackwell, Oxford, pp. 65-77

473 Lazar B, Loya Y (1991): Bioerosion of coral reefs - A chemical approach. Limnol. Oceanogr. 36, 377-383

474 Le Campion-Alsumard T (1975): Experimental study of the colonization of calcite fragments by marine
475 endolithic Cyanophyceae. Cahiers de Biologie Marine 16, 177-185

476 Le Campion-Alsumard T, Golubic S, Hutchings P (1995): Microbial endoliths in skeletons of live and dead
477 corals - porites lobata (Moorea, French Polynesia). Marine Ecology Progress Series 117, 149-157

478 Lukas KJ (1978): Depth distribution and form among common microboring algae from the Florida continental
479 shelf. Geological Society of America - Abstracts with Programs 10, 1 - 448

480 Murtagh F, Legendre P (2014): Ward's hierarchical agglomerative clustering method: which algorithms
481 implement Ward's criterion? Journal of Classification 31, 274-295

482 Neumann AC (1966): Observations on coastal erosion in Bermuda and measurements of boring rate of sponge
483 *Cliona lampa*. Limnol. Oceanogr. 11, 92-108

484 Pandolfi JM, Connolly SR, Marshall DJ, Cohen AL (2011): Projecting coral reef futures under global warming
485 and ocean acidification. Science 333, 418-422

486 Pari N, Peyrot-Clausade M, Le Campion-Alsumard T, Hutchings P, Chazottes V, Golubic S, Le Campion J,
487 Fontaine MF (1998): Bioerosion of experimental substrates on high islands and on atoll lagoons
488 (French Polynesia) after two years of exposure. Marine Ecology Progress Series 166, 119-130

489 Peyrot-Clausade M, Le Campion-Alsumard T, Hutchings P, Le Campion J, Payri C, Fontaine MC (1995): Initial
490 bioerosion and bioaccretion on experimental substrates in high island and atoll lagoons (French
491 Polynesia). Oceanologica Acta 18, 531-541

492 R. Development Core Team (2014): R: A language and environment for statistical computing, R Foundation for
493 Statistical Computing, Vienna, Austria

494 Radtke G, le Campion-Alsumard T, Golubic S (1996): Microbial assemblages of the bioerosional "notch" along
495 tropical limestone coasts. Algological Studies 83, 469-482

496 Reyes-Nivia C, Diaz-Pulido G, Kline D, Guldberg O-H, Dove S (2013): Ocean acidification and warming
497 scenarios increase microbioerosion of coral skeletons. Global Change Biology 19, 1919-1929

498 Schneider J, Torunski H (1983): Biokarst on limestone coasts, morphogenesis and sediments production. Marine
499 Ecology 4, 45-63

500 Schonberg CHL (2001): Small-scale distribution of great barrier reef bioeroding sponges in shallow water.
501 Ophelia 55, 39-54

502 Scoffin P, Alexandersson E, Bowes G, Clokie J, Farrow G, Milliman J (1980): Recent, temperate, sub-photic,
503 carbonate sedimentation: Rockall bank, Northeast Atlantic. Journal of Sedimentary Research 50, 331-
504 355

505 Tribollet A, Decherf G, Hutchings PA, Peyrot-Clausade M (2002): Large-scale spatial variability in bioerosion
506 of experimental coral substrates on the Great Barrier Reef (Australia): importance of microborers. *Coral*
507 *Reefs* 21, 424-432

508 Tribollet A, Golubic S (2005): Cross-shelf differences in the pattern and pace of bioerosion of experimental
509 carbonate substrates exposed for 3 years on the northern Great Barrier Reef, Australia. *Coral Reefs* 24,
510 422-434

511 Tribollet A, Langdon C, Golubic S, Atkinson M (2006): Endolithic microflora are major primary producers in
512 dead carbonate substrates of Hawaiian coral reefs. *Journal of Phycology* 42, 292-303

513 Tribollet A (2008a): Dissolution of dead corals by euendolithic microorganisms across the northern Great
514 Barrier Reef (Australia). *Microbial ecology* 55, 569-80

515 Tribollet A (2008b): The boring microflora in modern coral reef ecosystems: a review of its roles. *Current*
516 *Developments in Bioerosion*, 67-94

517 Tribollet A, Godinot C, Atkinson M, Langdon C (2009): Effects of elevated pCO_2 on dissolution of coral
518 carbonates by microbial euendoliths. *Global Biogeochemical Cycles* 23

519 Tribollet A, Golubic S (2011): Reef bioerosion : agents and processes. In: Dubinsky Z , Stambler N (Editors),
520 *Coral reefs: an ecosystem in transition*. Springer, Berlin Heidelberg, pp. 435-450

521 Tribollet A, Atkinson M, Cuet P, Chauvin A (2014): Production of seawater alkalinity by bioeroding microflora
522 increases with ocean acidification, *Ocean Sciences Meeting*, Honolulu, Hawaii, USA

523 Vogel K, Gektidis M, Golubic S, Kiene W, Radtke G (2000): Experimental studies on microbial bioerosion at
524 Lee Stocking Island, Bahamas and One Tree Island, Great Barrier Reef, Australia: implications for
525 paleoecological reconstructions. *Lethaia* 33, 190-204

526 Wilkinson C (2008): Status of coral reefs of the world: 2008, *Coral Reef Monitoring Network and Reef and*
527 *Rainforest Research Centre*, Townsville, Australia

528 Wisshak M, Tribollet A, Golubic S, Jakobsen J, Freiwald A (2011): Temperate bioerosion: ichnodiversity and
529 biodiversity from intertidal to bathyal depths (Azores). *Geobiology* 9, 492-520

530 Wisshak M, Schoenberg CHL, Form A, Freiwald A (2014): Sponge bioerosion accelerated by ocean
531 acidification across species and latitudes? *Helgoland Marine Research* 68, 253-262

532 Zundeleovich A, Lazar B, Ilan M (2007): Chemical versus mechanical bioerosion of coral reefs by boring sponges
533 - lessons from *Pione cf. vastifica*. *Journal of Experimental Biology* 210, 91-96

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TABLES

Table 1 Mean bioeroded surface area and mean depth of penetration P_{80} by euendoliths in dead coral blocks over a 12 months period.

| Time of exposure <i>(Months)</i> | Bioeroded surface area <i>(% ± SEM)</i> | Depth of penetration <i>(mm ± SEM)</i> |
|--|---|--|
| 1 | 17.62 ± 2.10 | 0.08 ± 0.02 |
| 2 | 28.64 ± 2.60 | 0.10 ± 0.01 |
| 3 | 32.22 ± 2.40 | 0.19 ± 0.02 |
| 4 | 40.55 ± 2.40 | 0.42 ± 0.04 |
| 5 | 39.44 ± 3.80 | 1.63 ± 0.12 |
| 6 | 33.76 ± 2.40 | 2.12 ± 0.15 |
| 7 | 27.91 ± 2.00 | 3.19 ± 0.19 |
| 8 | 25.61 ± 2.10 | 2.66 ± 0.22 |
| 9 | 29.20 ± 2.20 | 2.77 ± 0.22 |
| 10 | 36.32 ± 2.80 | 2.35 ± 0.22 |
| 11 | 38.43 ± 2.60 | 2.69 ± 0.18 |
| 12 | 39.70 ± 2.60 | 2.29 ± 0.24 |

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FIGURES

Fig. 1 The area of study. a - Position of New Caledonia in the Southwest Pacific Ocean; b - Study site at “Ile aux Canards” in New Caledonia; c - Two experimental exposure grids with 60 experimental coral blocks fixed on the reef at 3 m-depth

Fig. 2 Euendolithic filaments or borings observed on colored thin sections or after decalcification under light microscopy, or under SEM. A – *Phaeophila* sp. filament (Chlorophyta, on thin section); b – *Eugomontia* sp. filament with a few cells (Chlorophyta, after decalcification); c – *Ostreobium* sp. filaments (Chlorophyta, on thin section) with the white arrow indicating the typical shape of this alga, and the black arrow highlighting filaments of fungi; d – *Mastigocoleus testarum* filaments (Cyanobacteria, after decalcification) with heterocystous cells (black arrow); e – *Plectonema terebrans* filaments (Cyanobacteria, after decalcification); f – Borings of unidentified euendolithic fungi (under SEM). Scale bare = 20 μ m

Fig. 3 Mean relative abundance (RA) of euendolithic species as a function of time (in months) and their depth of penetration in substrates (in mm). a - Unidentified chlorophytes (i.e. *Phaeophila* sp. + *Eugomontia* sp. + unidentified species); b - The chlorophyte *Ostreobium* sp., c; Thin filaments (i.e. *Plectonema terebrans* + fungi); d - The cyanobacterium *Mastigocoleus testarum*

Fig. 4 Bioerosion by euendoliths (a) and grazers (b) as a function of time. Rates are expressed in kg of CaCO_3 dissolved per linear m^2 of reef (Mean \pm standard error)

Fig. 5 Preliminary logistic growth model of the process of biogenic dissolution in kg of CaCO_3 dissolved per linear m^2 of reef as a function of time. Solid line relates to fitted values, and dotted lines show the confidence interval of fitted values

Fig. 6 Non-metric multidimensional scaling (nMDS) realized after clustering the relative abundance of euendolithic species, biogenic dissolution (BD), and grazing over the duration of the experiment

Fig. 7 Graphic representation of temporal dynamics of the process of biogenic dissolution of carbonates by euendoliths and interaction with grazers













