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Synthesis and evaluation of substituted indolizines as peptidomimetics of RGD tripeptide sequence

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The synthesis of seven peptidomimetics of RGD is presented. The indolizine building block was obtained by condensation of allylglycine with dimethoxydihydrofuran followed by an intermolecular cyclisation. The bicyclic ring was functionalised with a carboxylic acid and a guanidinium appendage. The seven peptidomimetics were evaluated by cell-adhesion assays.

Introduction

Integrins are transmembrane receptors which regulate cell attachment and response to the extracellular matrix. They are involved in many physiopathological processes: coagulation of blood, tumor-induced angiogenesis, osteoporosis, restenosis, acute renal failure, ocular diseases, metastasis formation and sickle cell anemia, and represent an interesting target for therapeutic.¹

Among the various known integrins the $\alpha_v\beta_3$ and $\alpha_v\beta_5$ integrin receptors are of great interest since they are involved in angiogenesis, a target for cancer therapy by inhibiting vascularisation of tumors.²

The tripeptide Arg-Gly-Asp (RGD) is the motif of recognition of most integrins on several matrix proteins (vitronectin (VN), fibronectin (FN))... The specificity of each integrin depends on the conformation of the triad induced by the other residues.³

To date, several inhibitors have been imagined and prepared.⁴ A cyclic pentapeptide (c(-RGDf[MMe]V-)) Cilengitide was found quite active while a lot of studies have emerged in order to access peptidomimetics.⁵ In these series of compounds, azabicycloalkanes have been developed as versatile scaffolds to mimic peptide conformation.⁶ Their bicyclic ring backbone gives geometry with fixed dihedral angles, on which the acid (Asp) and basic (Arg) side chains can be placed at an optimal distance of 13 Å allowing the recognition.

We have been interested in developing the synthesis of several new structures as integrin inhibitors since we recently developed an efficient synthesis of indolizin-3-one.⁷ We envisaged it as a scaffold which could be converted into a potent integrin ligand by introducing a carboxylic and a guanidine appendage (Fig 1).

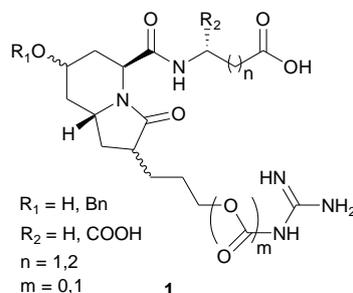


figure 1

Results and discussion

In this paper, we report the synthesis and the biological activity of RGD peptidomimetics **1**. On this bicyclic compound, the length and the relative geometry of the two side chains bearing the acid and basic functions can be adapted to optimize the affinity with the receptor. The peptidomimetics were evaluated in vitro by cell-adhesion assays.

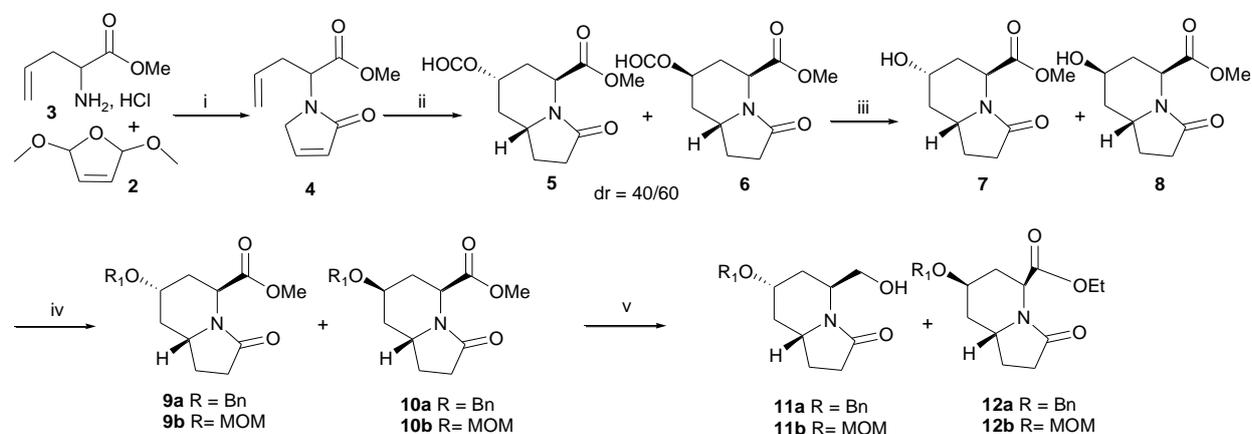
Synthesis of the indolizine building block

The first step was the condensation in acidic medium of dimethoxydihydrofuran⁸ (**2**) with racemic allylglycine (**3**) (scheme 1).

This condensation led to lactam **4** which was directly cyclised in refluxing formic acid to give only two diastereoisomers at the C7 position **5** and **6** (60/40) which can be separated through column chromatography. Practically, the crude reaction mixture was used. The formiate was hydrolysed by KHCO_3 (1.1 equivalent) in MeOH to

yield alcohols **7** and **8** in 86% yield. The hydroxyl group was protected as a benzyl or a MOM ether in good yield to give **9a**, **10a** (76%) and **9b**, **10b** (87%) respectively.

Their reaction with NaBH₄ in refluxing ethanol allowed an easy separation of the two diastereoisomers. Indeed isomer **10a** and **10b** which exhibited a *cis* group were not reduced but just transesterified by ethanol. **12a**, **12b**, and **11a**, **11b** were isolated in 84% and 83% overall yield. NMR studies made it possible to establish a *cis*-diaxial relationship between the hydroxyl and ester groups in compound **10** (or **12**) and could explain the chemoselectivity of the NaBH₄ reduction.

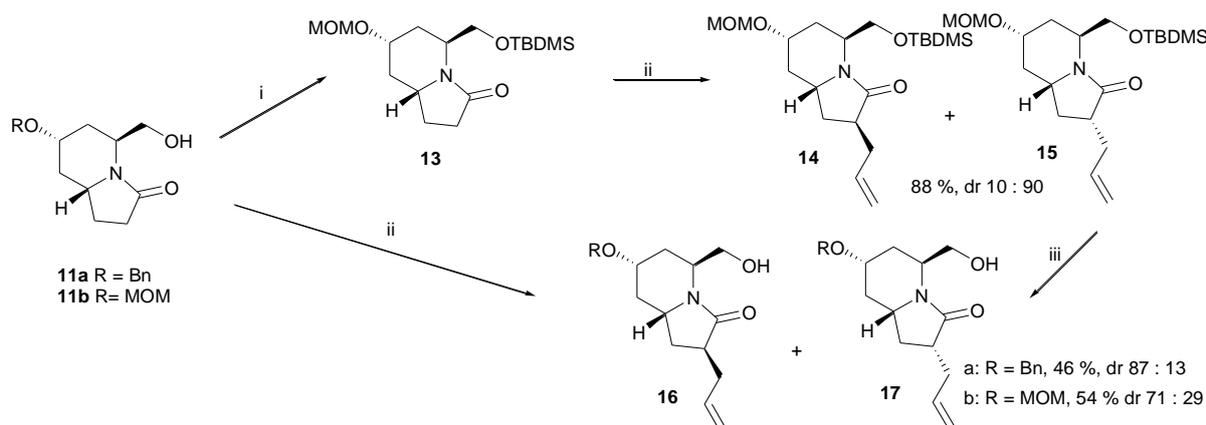


Scheme 1 synthesis of the indolizine building block i) HCl 1 M; ii) HCOOH, reflux, 60% (2 steps); iii) KHCO₃ (1.1 eq), MeOH, 0 °C, 86%; iv) R₁ = Bn: trichloroacetimidate (3 equiv), TfOH (0.25 equiv), C₆H₁₂/CH₂Cl₂, 22 h, 76%; R₁ = MOM: *i*Pr₃NEt (4.5 equiv), MOMCl (4.5 equiv), THF, rt, 2 days, 87%; v) NaBH₄, EtOH, reflux, R₁ = Bn, 84%, R₁ = MOM, 83%.

Functionalisation of the indolizine building block

Having prepared the bicyclic skeleton, we had to introduce the two side chains with an appropriate relative configuration. An allyl group was first introduced (α to the carbonyl of the lactam) to be transformed into the guanidine. This allylation was performed after deprotonation using LDA in tetrahydrofuran and allyl bromide (scheme 2). Lactams **11a** and **11b** bearing the ether and hydroxyl functions in a *trans* relationship were deprotonated with 2 equivalents of LDA to furnish, after treatment with allyl bromide, **16a** + **17a** (87:13, 46% yield) and **16b** + **17b** (71:29, 54%) respectively. When the allylation was conducted with the diprotected derivative **13** (MOM and TBDMS ether), **14** and **15** were isolated in improved yield and diastereoselectivity (10:90, 88%). It is noteworthy that the diastereoselectivity was inverted compared to the free hydroxyl series. Thus both isomer series can be easily attained by using protected or unprotected alcohol. In all cases, the relative configuration of the allyl group was assigned by NOE experiments.

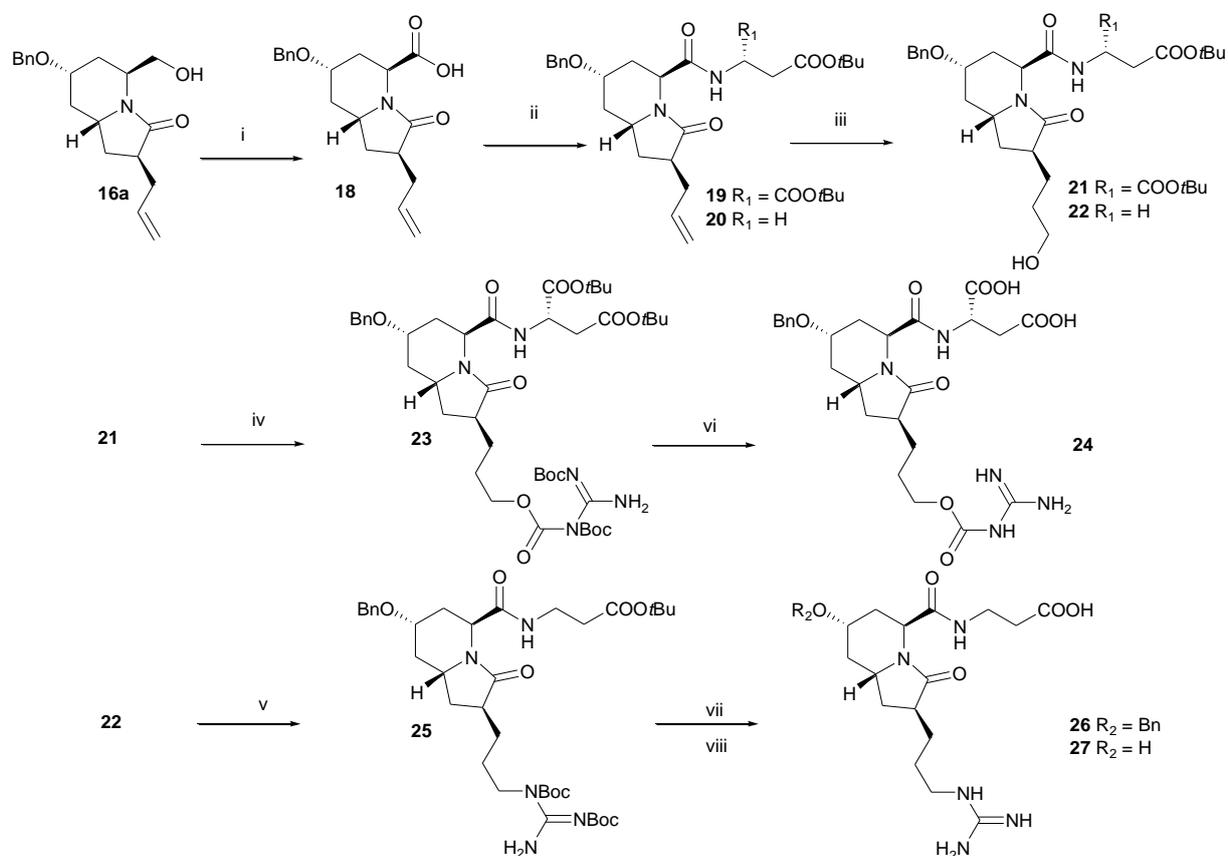
The next task was to perform the functional group interchanges to obtain the acidic and basic side chains.



Scheme 2 allylation of the indolizine i) Et₃N, TBDMSOTf, CH₂Cl₂, -10 °C, 88%; ii) LDA, -78 °C, allylbromide, THF; iii) Bu₄NF, THF, 97%.

We anticipated transforming the allyl group into guanidine. Furthermore, different acid side chains could be obtained from the same intermediates. This transformation was attempted with benzyl ether **16a**. Actually, this synthetic way was not useful: the yield was low (oxidation of the primary alcohol into acid) and purification of the guanidine compound

was difficult (several chromatographies were needed). Consequently, we decided to begin with the acid side chain. In this sequence, we started working on the acid side chain (scheme 3). Alcohol **16a** was oxidized by Jones reagent to give the acid **18** in good yield (67%). Coupling with β -Alanine *ter*-butylester and L-Aspartic acid di-*ter*-butylester furnished **19** (83%) and **20** (88%), respectively.

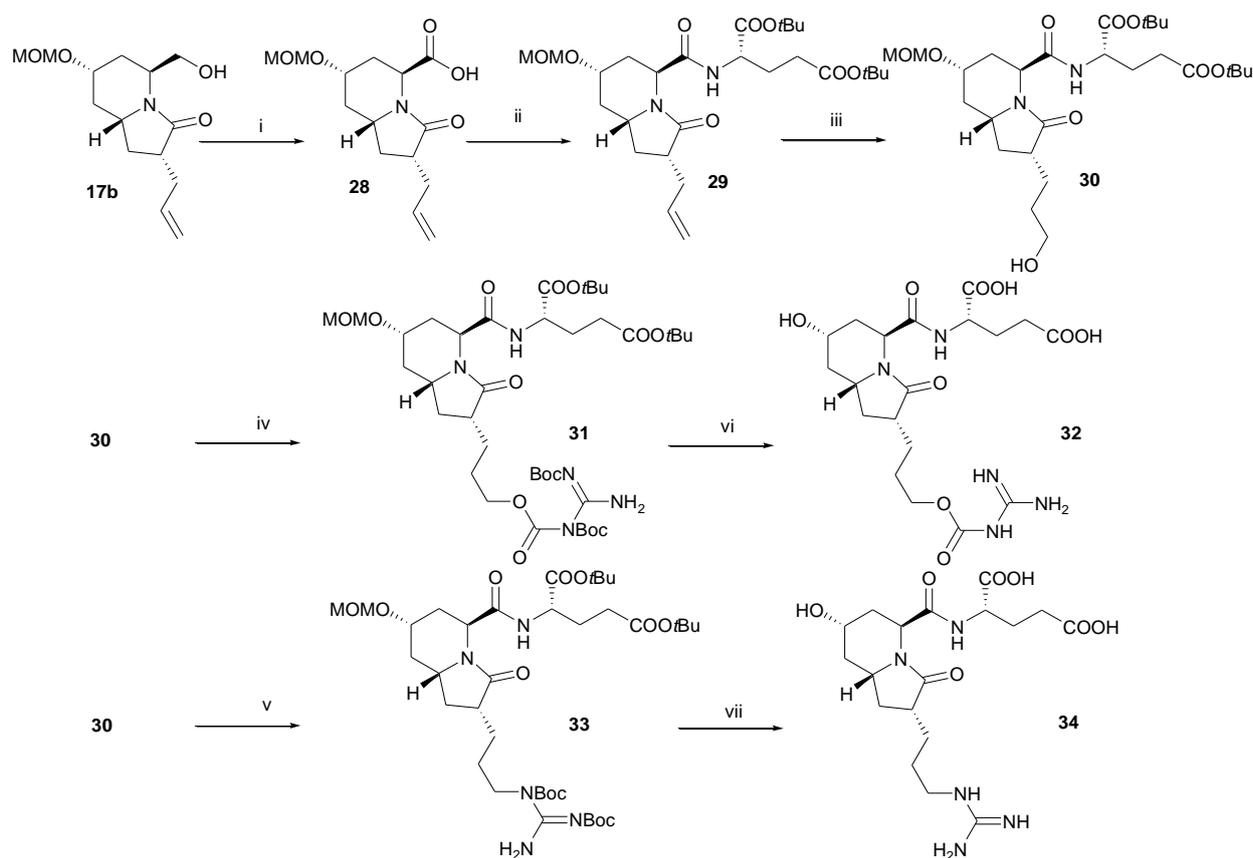


Scheme 3 functionalisation of the *cis* diastereoisomer: i) CrO₃, H₂SO₄, acetone, 0 °C, 40 min, 67%; ii) NMM, ClCOOtBu, CH₂Cl₂, R₁ = H, 83%, R₁ = COOtBu, 88%; iii) a) 9-BBN, b) H₂O₂, AcONa, R₁ = COOtBu, 68%, R₁ = H, 71%; iv) ADDP, PPh₃, di-Boc-guanidine, toluene, reflux, 2h, 60%; v) DEAD, PPh₃, di-Boc-guanidine, THF, rt, 64h, 92%; vi) TFA, CH₂Cl₂, 100%; vii) R₂ = Bn: TFA, CH₂Cl₂, 100%; viii) R₂ = H: a) H₂ Pd/C 10%, 72 % b) TFA, CH₂Cl₂, 100%.

Hydroboration of **19** or **20** gave the alcohols **21** (71%) or **22** (68%). Contrary to several reports, substitution of the hydroxyl group of **21** by a guanidine through a Mitsunobu reaction with DIAD/PPh₃ and di-Boc- or di-Cbz-guanidine was proved to be unsuccessful.⁹ We thus investigated the more reactive reagents dipiperidin-azodicarboxylate (ADDP), tributylphosphine (Bu₃P).¹⁰ Surprisingly, use of these reagents led to an unexpected compound: the guanidine carbamate **23**.¹¹

Eventually, starting from compound **22**, use of DEAD/PPh₃⁴ instead DIAD/PPh₃ permitted to access to the guanidilated product **25**. Boc deprotection with trifluoroacetic acid gave two new mimics **24** and **26**. Hydrogenation on Pd/C of **25** followed by trifluoroacetic acid treatment also gave another analogue **27**.

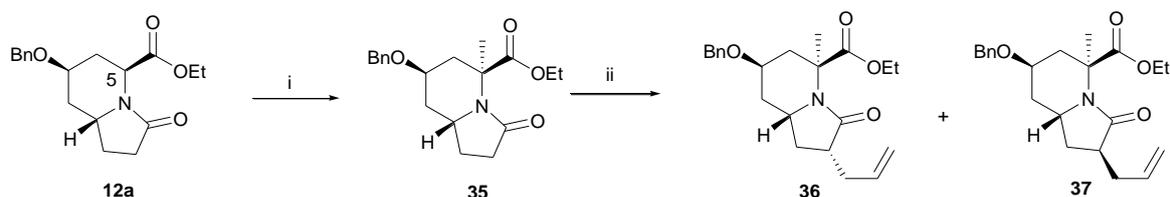
The same sequence was applied to the *trans* diastereoisomer **17b** (scheme 4).



Scheme 4 functionalisation of the *trans* diastereoisomer: i) CrO₃, H₂SO₄, acetone, 0 °C, 40 min, 54%; ii) NMM, ClCOO*t*Bu, CH₂Cl₂, 90%; iii) a) 9-BBN, b) H₂O₂, AcONa, 65%; iv) ADDP, PPh₃, di-Boc-guanidine, THF, reflux, 2h, 43% v) DEAD, PPh₃, di-Boc-guanidine, THF, rt, 64h, 60%; vi) TFA, CH₂Cl₂, 100%; vii) TFA, CH₂Cl₂, 100%.

Oxidation of alcohol **17b** gave acid **28** in moderate yield (54%), which was coupled with L-glutamic acid di-*tert*-butylester (90%). Then after hydroboration of **29** (65%), subsequent reaction of thus obtained **30** with di-Boc-guanidine led to **31** (carbamate) and **33** (guanidinium). TFA-cleavage of the tert-butoxy and tert-butoxycarbonyl groups of **31** and **33** afforded **32** and **34**, respectively.

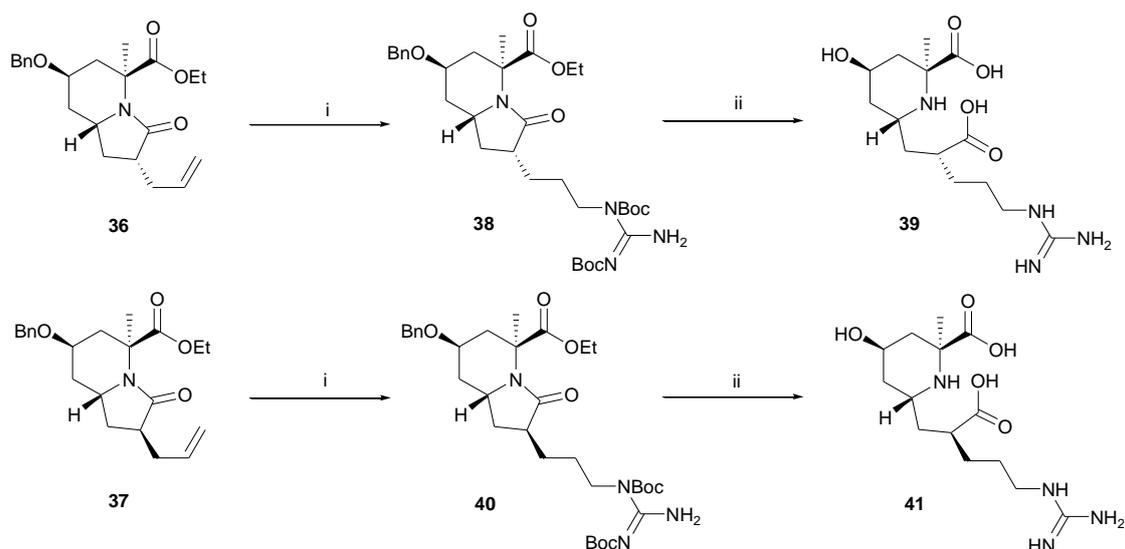
We investigated the transformation of compounds **12a**. When these esters reacted with strong bases, a carbanion was formed at carbon 5. First, this center was methylated by using KHMDS and methyl iodide in good yield to obtain single diastereoisomer **35** (scheme 5).



Scheme 5 alkylation of the *trans* diaxial diastereoisomer: i) KHMDS, MeI, THF, 99%; ii) KHMDS, allylbromide, 55% dr 60:40.

Then alkylation was performed with the same organic base and allylbromide, the diastereoselectivity was low (de 20%) and the yield was moderate (55%) (scheme 5).

The double bond of **36** and **37** was converted into guanidinium group with DEAD/PPh₃ in good overall yield (54% and 87%, respectively, for 3 steps). Saponification of ethyl ester was very difficult because of the steric hindrance. We checked several bases: NaOH, KOH, KO*t*Bu, NaSMe, LiSMe, leading to no reaction or degradation of the starting material. Indeed, acidic conditions were needed: hydrolysis in aqueous ethanol with 3 M hydrochloric acid at reflux for 20 h gave compounds **39** and **41** of which the amide bond, the benzyl ether were also hydrolyzed (scheme 6). These compounds **39** and **41** had been also considered as potential peptidomimetics and they have been tested in adhesion assays.



Scheme 6 functionalisation and hydrolysis of the *trans* diaxial diastereoisomer i) a) 9-BBN, b) H₂O₂, AcONa c) DEAD, PPh₃, di-Boc-guanidine, THF, rt (**38**, 54%; **40**, 87%); ii) EtOH/H₂O, HCl 3M, 100%.

We synthesised 7 potential peptidomimetics of RGD in racemic form, in 10 or 11 steps and in 0.6% to 3.4% total yield from racemic allylglycine.

Biological test of the 7 peptidomimetics

The compounds **24**, **26**, **27**, **32**, **34**, **39** and **41** were evaluated *in vitro* for their ability to inhibit cell adhesion. Their activity was compared to that of c(RGDfV) and GRGES that were used as positive and negative competitors for cell adhesion. We used two different cell lines, mouse sarcoma S180 cells and human vascular endothelial Eahy926 cells. These cells express $\alpha 5\beta 1$ and $\alpha v\beta 3$ integrins.¹² The c(RGDfV) peptide strongly inhibited S180 cell adhesion to VN (Fig. 2A, black bars) with 19% and 5% of cell adhesion in the presence of 10 μ M and 200 μ M of its competitor, respectively. This cyclic peptide inhibited less efficiently S180 cell adhesion to FN-coated surface (Fig. 2B, black bars) with 95% and 69% of adhesion in the presence of 10 μ M and 500 μ M c(RGDfV), respectively. In contrast, the control peptide GRGES has no inhibitory effect on the two types of coated surfaces (Fig. 2, white bars). Among the peptidomimetics tested, only the **26** and **27** compounds showed inhibitory effect on S180 cell adhesion to VN but they are less efficient compared to the c(RGDfV). We measured 65% and 74% of adhesion in the presence of the **26** or **27** compound at 200 μ M, respectively (Fig. 2A, dashed bars). They did not interfere with S180 cell adhesion to FN-coated surfaces.

The c(RGDfV) peptide also interfered with Eahy926 adhesion to VN but to a lesser extent compared to S180 cell adhesion, with 74.8 \pm 7.0 and 9.2 \pm 4.4% of adhesion measured at 10 μ M and 200 μ M c(RGDfV). All compounds tested showed no inhibitory effect on Eahy926 adhesion to VN nor FN-coated surfaces.

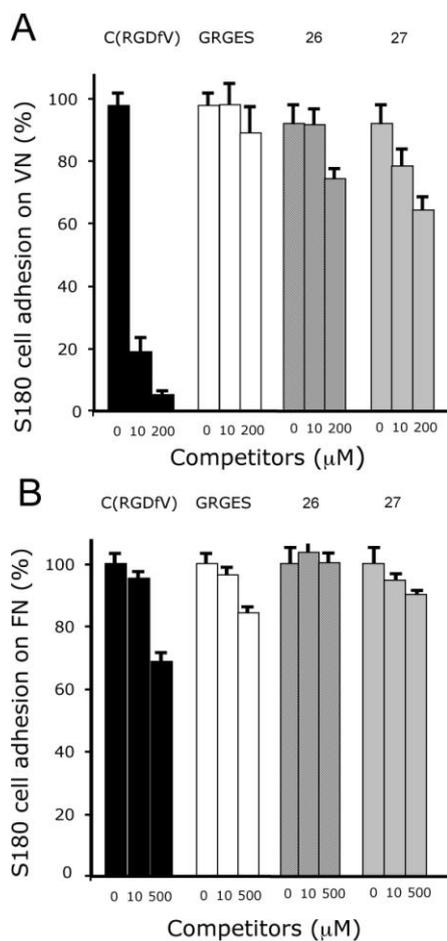


figure 2

Conclusions

The scaffold of indolizin-3-one was used to prepare seven structures as potential mimics of the RGD tripeptide sequence. These compounds were evaluated in cell adhesion test. Two of them showed some activities: they possess the acid and guanidine side chains in a *cis* relationship. This configuration corresponds to the required geometry of a β -turn (type II). Optimisation of the length of the side chains and use of optically active allyl glycine as starting material are under investigation.

Experimental section

General experimental methods

All commercially available reagents were used without further purification unless otherwise noted. Tetrahydrofuran was freshly distilled from benzophenone ketyl radical under argon prior to use. Column chromatography was performed with silica gel (35-70 mesh). Melting points are reported uncorrected.

NMR spectra were recorded on 300 and 400 MHz spectrometers Bruker Avance. Chemical shifts are reported in ppm. Coupling constants (J values) are reported in Hertz. ^{13}C and ^1H peak assignments were made based on DEPT, HMQC, HSQC, HSBC and NOESY data and IR spectra were recorded on a FT-IR spectrometer Perkin Elmer 1600.

MS experiments were performed on a Q-ToF Micromass-1 spectrometer.

(5*S,7*S**,8*aR**)-7-hydroxy-3-oxo-octahydro-indolizin-5-carboxylic acid methyl ester 7 and (5*S**,7*R**,8*aR**)-7-hydroxy-3-oxo-octahydro-indolizin-5-carboxylic acid methyl ester 8**

To a solution of formiates **5** and **6** (10.91 g, 45.2 mmol, 1.0 equiv) in 210 mL of anhydrous methanol was added solid KHCO_3 (4.54 g, 45.4 mmol, 1.0 equiv) at 0 °C and the reaction was stirred 40 min at 0 °C. KHCO_3 was filtered and the filtrate was concentrated in vacuo. Purification by flash column chromatography on silica gel (EtOAc-MeOH = 95:5 then 90:10) gave **7** and **8** as a yellow solid (8.3 g, 86%).

A sample of each diastereoisomer was obtained

8: $R_f = 0,3$ (AcOEt/MeOH 90/10) mp: 127 °C; $^1\text{H NMR}$ (CDCl_3 , 300 MHz) δ_{H} 1.30 (1H, ddd, $J = 13.8, 12.0, 2.2$ Hz, $\text{H}_{8\text{ax}}$); 1.55 (1H, dddd $J = 12.4, 9.8, 9.8, 8.5$ Hz, H_1); 1.76 (1H, ddd, $J = 14.3, 7.0, 2.2$ Hz, $\text{H}_{6\text{ax}}$); 2.01 (1H, dm, $J = 13.4$ Hz, $\text{H}_{8\text{eq}}$); 2.23 (1H, dddd, $J = 12.3, 9.3, 7.1, 3.0$ Hz, H_1); 2.3-2.5 (3H, m, 2H_2 H_6); 3.67 (3H, s, OMe); 4.08 (1H, dddd, $J = 11.8, 8.0, 8.0, 3.5$ Hz, $\text{H}_{8\text{a}}$); 4.19 (1H, m, $\text{H}_{7\text{eq}}$); 4.68 (1H, d(l), $J = 6.0$ Hz, H_5). $^{13}\text{C NMR}$ (75 MHz, CDCl_3) δ_{C} 25.8 (C_1); 30.2 (C_2); 32.7 (C_6); 38.8 (C_8); 48.1 (C_5); 49.2 ($\text{C}_{8\text{a}}$); 52.4 (C_{OMe}); 63.7 (C_7); 171.9 175.3 (CO C_3). IR (NaCl disc, cm^{-1}): $\nu_{\text{max}} = 3388, 2962, 1749, 1665, 1441, 1313, 1215, 1090, 1071$. MS (ES^+): $m/z = 236$ ($[\text{MNa}]^+$). HRMS (ES^+ , $[\text{MNa}]^+$): Calcd for $\text{C}_{10}\text{H}_{15}\text{NO}_4$ 236.0899 Found 236.0893.

7: $R_f = 0,3$ (AcOEt/MeOH 90/10) mp: 94 °C. $^1\text{H NMR}$ (CDCl_3 , 300 MHz): δ_{H} 1.1 (1H, ddd, $J = 11.7, 11.7, 11.7$ Hz, $\text{H}_{8\text{ax}}$); 1.48 (1H, m, H_6); 1.6 (1H, m, H_1); 2.09 (1H, bd, $J = 11.8$ Hz, $\text{H}_{8\text{eq}}$); 2.2 (1H, m, H_1); 2.38 (3H, m, 2H_2 H_6); 3.66 (4H, bs, OMe H_7); 3.75 (1H, dddd, $J = 7.5, 7.5, 7.5, 7.5$ Hz, $\text{H}_{8\text{a}}$); 4.8 (1H, d(l), $J = 5.7$ Hz, H_5). $^{13}\text{C NMR}$ (75 MHz, CDCl_3) δ_{C} 25.5 (C_1); 30.4 (C_2); 34.7 (C_6); 41.8 (C_8); 50.2 (C_5); 52.6 (OMe); 53.9 ($\text{C}_{8\text{a}}$); 65.2 (C_7); 170.9 174.8 (CO C_3). IR (NaCl disc, cm^{-1}): $\nu_{\text{max}} = 3432, 2952, 1736, 1666, 1432, 1213$. HRMS (ES^+ , $[\text{MNa}]^+$): Calcd for $\text{C}_{10}\text{H}_{15}\text{NO}_4$ 236.0899 Found 236.0909.

(5S*,7S*,8aR*)-7-benzyloxy-3-oxo-octahydro-indolizin-5-carboxylic acid methyl ester 9a and (5S*,7R*,8aR*)-7-benzyloxy-3-oxo-octahydro-indolizin-5-carboxylic acid methyl ester 10a

Alcohols **7** and **8** (10.7g, 50.4 mmol) were dissolved in 40 mL of dichloromethane under argon, 80 mL of cyclohexane were added. At rt, 19 mL (102 mmol, 2 equiv) of benzyl-trichloroacetimidate were added, followed by 440 μL (5 mmol, 0.1 equiv) of triflic acid. The mixture was refluxed for 16 h, 3 mL of benzyl-trichloroacetimidate was further added and the reflux was continued for 20 h. The mixture was filtered on silica gel and washed with dichloromethane. The filtrate was evaporated and purification by flash column chromatography on silica gel (EtOAc-cyclohexane-MeOH = 80: 15: 5) gave **9a** and **10a** as a colourless oil (13.6 g, 89%).

9a: $R_f = 0.49$ (AcOEt/cyclohexane/MeOH 80/15/5). $^1\text{H NMR}$ (CDCl_3 , 300 MHz) δ_{H} 1.16 (1H, ddd, $J = 11.7, 11.7, 11.7$ Hz, $\text{H}_{8\text{ax}}$); 1.58 (2H, m, H_1 $\text{H}_{6\text{ax}}$); 2.2 (2H, m, H_1 $\text{H}_{8\text{eq}}$); 2.38 (2H, m, H_2); 2.55 (1H, dm, $J = 7, 5$ Hz, $\text{H}_{6\text{eq}}$); 3.44 (1H, dddd, $J = 11.3, 11.3, 3.8, 3.8$ Hz, $\text{H}_{7\text{ax}}$); 3.66 (3H, s, OMe); 3.73 (1H, m, $\text{H}_{8\text{a}}$); 4.51 (2H, s, CH_2Ph); 4.89 (1H, d, $J = 5.6$ Hz, H_5); 7.26 (5H, m, Ar). $^{13}\text{C NMR}$ (75 MHz, CDCl_3) δ_{C} 25.2 (C_1); 29.9 (C_2); 31.7 (C_6); 38.8 (C_8); 49.8 (C_5); 52.2 (OMe); 53.3 ($\text{C}_{8\text{a}}$); 69.9 (CH_2Ph); 72.0 (C_7); 127.4 (3C_{Ar}); 128.1 (C_{Ar}); 137.8 (C_{Arquat}); 170.9 174.8 (CO C_3). IR (NaCl disc, cm^{-1}): $\nu_{\text{max}} = 3352, 2926, 1732, 1682, 1454, 1366, 1291, 1208, 1110$. HRMS (ES^+ , $[\text{MNa}]^+$): Calcd for $\text{C}_{17}\text{H}_{21}\text{NO}_4$ 326.1368 Found 326.1365.

10a: $R_f = 0.43$ (AcOEt/cyclohexane/MeOH: 80/15/5) $^1\text{H NMR}$ (CDCl_3 , 300 MHz): δ_{H} 1.29 (1H, ddd, $J = 13.7, 12.0, 2.2$ Hz, $\text{H}_{8\text{ax}}$); 1.57 (1H, dddd, $J = 12.4, 10.5, 10.5, 8.7$ Hz, H_1); 1.71 (1H, ddd, $J = 14.3, 7.0, 2.1$ Hz, $\text{H}_{6\text{ax}}$); 2.19 (1H, dddd, $J = 13.4, 13.4, 13.4, 13.4$ Hz, $\text{H}_{8\text{eq}}$); 2.28 (1H, m, H_1); 2.40 (2H, m, 2H_2); 2.71 (1H, dddd, $J = 14.2, 3.3, 1.7, 1.7$ Hz, $\text{H}_{6\text{eq}}$); 3.51 (3H, s, OMe); 3.84 (1H, bs, $\text{H}_{7\text{eq}}$); 4.11 (1H, dddd, $J = 11.8, 8.4, 8.4, 3.6$ Hz, $\text{H}_{8\text{a}}$); 4.41 (1H, d, $J = 11.8$ Hz, CH_2Ph); 4.51 (1H, d, $J = 11.8$ Hz, CH_2Ph); 4.74 (1H, d, $J = 6.0$ Hz, H_5); 7.25 (5H, m, H_{Ar}). $^{13}\text{C NMR}$ (75 MHz, CDCl_3) δ_{C} 25.9 (C_1); 29.3 (C_6); 30.1 (C_2); 37.1 (C_8); 48.2 (C_5); 49.4 ($\text{C}_{8\text{a}}$); 52.0 (OMe); 70.1 (CH_2Ph); 70.7 (C_7); 127.0 (2C_{Ar}); 127.4 (C_{Ar}); 128.2 (C_{Ar}); 138.2 (C_{Arquat}); 171.0 174.6 (CO C_3). IR (NaCl disc, cm^{-1}): $\nu_{\text{max}} = 3448, 2951, 1747, 1696, 1417, 1359, 1293, 1209, 1093$. MS (ES^+): m/z 326 ($[\text{MNa}]^+$).

(5S*,7S*,8aR*)-7-methoxymethoxy-3-oxo-octahydro-indolizin-5-carboxylic acid methyl ester 9b and (5S*,7R*,8aR*)-7-methoxymethoxy-3-oxo-octahydro-indolizin-5-carboxylic acid methyl ester 10b

To alcohols **7** and **8** (371 mg, 1.74 mmol) dissolved in 7 mL of anhydrous THF under argon and cooled to 0 °C, was successively added diisopropylethylamine (860 μL , 5.2 mmol, 3 equiv) and chloromethoxymethane (395 μL , 5.2 mmol, 3 equiv). The mixture was stirred for 20 h, 1.5 equiv of each reagent was added again, stirring was continued for 16 additional hours. After addition of a saturated solution of NH_4Cl , the ester was extracted by CH_2Cl_2 , the organic layers were dried (MgSO_4) and concentrated under vacuum. Purification by flash column chromatography on silica gel gave 388 mg (87 %) of **9b** and **10b**.

10b: $R_f = 0.39$ (AcOEt/cyclohexane/MeOH: 80/15/5). $^1\text{H NMR}$ (CDCl_3 , 300 MHz): δ_{H} 1.25 (1H, ddd, $J = 13.6, 11.7, 1.9$ Hz, $\text{H}_{8\text{ax}}$); 1.52 (1H, m, H_1); 1.66 (1H, ddd, $J = 14.4, 7.0, 2.0$ Hz, H_6); 2.03 (1H, dm, $J = 13.4$ Hz, $\text{H}_{8\text{eq}}$); 2.21 (1H, m, H_1); 2.26-2.44 (2H, m, 2H_2); 2.48 (1H, bd, $J = 14.4$ Hz, $\text{H}_{6\text{eq}}$); 3.26 (3H, s, COOMe); 3.63 (3H, s, OMe); 4.0 (2H, m, H_7 $\text{H}_{8\text{a}}$); 4.41 (1H, d, $J = 6.8$ Hz, OCH_2O); 4.55 (1H, d, $J = 6.8$ Hz, OCH_2O); 4.66 (1H, d, $J = 6.6$ Hz, H_5). $^{13}\text{C NMR}$ (75 MHz, CDCl_3) δ_{C} 25.9 (C_1); 29.9 (C_6); 30.2 (C_2); 37.2 (C_8); 48.2 (C_5); 49.5 ($\text{C}_{8\text{a}}$); 52.2 (CH_2OMe); 55.5 (OMe); 67.7 (C_7); 94.0 (OCH_2O); 171.2 174.9 (CO C_3). IR (NaCl disc, cm^{-1}): $\nu_{\text{max}} = 3478, 2950, 1747, 1682, 1417, 1209, 1038, 919$. MS (ES^+): m/z 280 ($[\text{MNa}]^+$). HRMS (ES^+ , $[\text{MH}]^+$): Calcd for $\text{C}_{12}\text{H}_{19}\text{NO}_5$ 258.1341 Found 258.1351.

9b: $R_f = 0.46$ (AcOEt/cyclohexane/MeOH 80/15/5). $^1\text{H NMR}$ (CDCl_3 , 300 MHz) δ_{H} 1.19 (1H, ddd, $J = 11.7, 11.7, 11.7$ Hz, $\text{H}_{8\text{ax}}$); 1.61 (2H, m, H_1 H_6); 2.24 (2H, m, H_1 H_8); 2.3-2.5 (3H, m, H_2 H_6); 3.34 (3H, s, OMe); 3.63 (1H, m, H_7); 3.72 (3H, s, OMe); 3.80 (1H, m, $\text{H}_{8\text{a}}$); 4.65 (2H, s, OCH_2O); 4.9 (1H, d, $J = 5.7$ Hz, H_5). $^{13}\text{C NMR}$ (75 MHz, CDCl_3) δ_{C} 25.5 (C_1); 30.2 (C_2); 32.5 (C_6); 39.7 (C_8); 50.1 (C_5); 52.2 (OMe); 53.5 ($\text{C}_{8\text{a}}$); 70.9 (C_7); 95.0 (OCH_2O); 170.9 174.2

(CO C₃).

(5S*,7R*,8aR*)-7-benzyloxy-3-oxo-octahydro-indolizin-5-carboxylic acid ethyl ester 12a

To a mixture of **9a** and **10a** (4.7 g, 15.5 mmol, cis/trans: 20/80) dissolved in 250 mL of absolute alcohol, sodium borohydride (6g, 159 mmol, 10 equiv) was added. The mixture was refluxed for 3 h. After cooling, the solution was concentrated under vacuum, borane precipitates were filtrated and washed with CH₂Cl₂. The filtrate was concentrated under reduced pressure and the residue was diluted with CH₂Cl₂ and H₂O, extracted twice by CH₂Cl₂. The organic phase was dried with MgSO₄ and evaporated to dryness, whereafter the residue was purified by flash chromatography on silica gel (EtOAc/MeOH, 99/1 to 90/10) to afford 780 mg of **11a** as a light yellow oil (16%, 80% from **10a**) and 2.9 g of **12a** as a white solid 68%, 85% from **9a**).

R_f = 0.28 (AcOEt); ¹H NMR (CDCl₃, 300 MHz): δ_H 1.11 (3H, t, *J* = 7.2, CH₃); 1.28 (1H, ddd, *J* = 13.8, 12.0, 2.3 Hz, H_{8ax}); 1.56 (1H, dddd, *J* = 12.4, 9.6, 9.6, 9.1 Hz, H₁); 1.70 (1H, ddd, *J* = 14.4, 7.1, 2.2 Hz, H_{6ax}); 2.10 (1H, ddd, *J* = 13.4, 13.4, 13.4 Hz, H_{8eq}); 2.25 (1H, dddd, *J* = 12.2, 9.4, 6.8, 2.7 Hz, H₁); 2.42 (2H, m, 2H₂); 2.71 (1H, dddd, *J* = 14.3, 3.2, 1.6, 1.6 Hz, H_{6eq}); 3.83 (1H, m, H₇); 3.92 (1H, dq, *J* = 10.7, 7.2, OCH₂); 4.02 (1H, dq, *J* = 10.7, 7.2, OCH₂); 4.11 (1H, dddd, *J* = 11.8, 8.5, 8.5, 3.4 Hz, H_{8a}); 4.41 (1H, d, *J* = 11.9 Hz, CH₂Ph); 4.52 (1H, d, *J* = 11.9 Hz, CH₂Ph); 4.72 (1H, bd, *J* = 5.9 Hz, H₅); 7.27 (5H, m, Ar). ¹³C NMR (75 MHz, CDCl₃) δ_C 14.0 (CH₃); 26.1 (C₁); 29.3 (C₆); 30.4 (C₂); 37.1 (C₈); 48.4 (C₅); 49.6 (C_{8a}); 61.3 (CH₂O); 70.2 (CH₂Ph); 70.6 (C₇); 94.0 (OCH₂O); 127.1 (2C_{Ar}); 127.5 (C_{Ar}); 128.3 (2C_{Ar}); 138.4 (C_{Arquat}); 170.7 174.9 (CO C₃). IR (NaCl disc, cm⁻¹): ν_{max} = 3468, 2935, 1742, 1694, 1417, 1292, 1198, 1093. MS (ES⁺): *m/z* 340 ([MNa]⁺). HRMS (ES⁺, [MNa]⁺): Calcd for C₁₈H₂₃NO₄ 340.1525 Found 340.1534.

(5S*,7S*,8aR*)-7-benzyloxy-5-hydroxymethyl-hexahydro-indolizin-3-one 11a

R_f = 0.39 (AcOEt/MeOH: 90/10); mp: 95 °C. ¹H NMR (CDCl₃, 300 MHz): δ_H 1.17 (1H, ddd, *J* = 11.8, 11.8, 11.8 Hz, H_{8ax}); 1.47 (1H, ddd, *J* = 12.9, 11.6, 6.6 Hz, H_{6ax}); 1.59 (1H, dddd, *J* = 12.5, 9.6, 9.6, 7.3 Hz, H₁); 2.2 (3H, m, H₆ H_{8eq} H₁); 2.38 (2H, m, H₂); 3.56 (2H, d, *J* = 7.0 Hz, CH₂O); 3.7 (2H, m, H₇ H_{8a}); 4.35 (1H, ddd, *J* = 6.7, 6.7, 6.7 Hz, H₅); 4.54 (2H, s, OCH₂Ph); 7.30 (5H, m, Ar). ¹³C NMR (75 MHz, CDCl₃) δ_C 19.7 (C₁); 29.2 (C₂); 29.9 (C₆); 38.4 (C₈); 48.5 (C₅); 52.0 (C_{8a}); 60.5 (CH₂O); 68.7 (CH₂Ph); 70.8 (C₇); 126.2 (3C_{Ar}); 127.2 (2C_{Ar}); 137.6 (C_{Arquat}); 174.9 (C₃). IR (KBr disc, cm⁻¹): ν_{max} = 3319, 2943, 1742, 1664, 1458, 1290, 1100, 1074, 740. HRMS (ES⁺, [MH]⁺): Calcd for C₁₆H₂₁NO₃ 276.1600 Found 276.1602. Anal. Calcd. for: C₁₆H₂₁NO₃ C, 69.79; H, 7.69; N, 5.09. Found: C, 69.33; H, 7.74; N 4.99%.

(5S*,7R*,8aR*)-7-methoxymethoxy-3-oxo-octahydro-indolizin-5-carboxylic acid ethyl ester 12b

To a mixture of **9b** and **10b** 128 mg (0.5 mmol, cis/trans: 74/26) dissolved in 10 mL of absolute alcohol, sodium borohydride (189 mg, 5 mmol, 10 equiv) was added. The solution was refluxed for 3 h. After cooling, the solution was concentrated under vacuum, borane precipitates were filtrated and washed with CH₂Cl₂. The filtrate was concentrated under reduced pressure; the residue was diluted with CH₂Cl₂ and 10 mL of H₂O, extracted twice by CH₂Cl₂. The organic phase was dried with MgSO₄ and evaporated to dryness, whereafter the residue was purified by flash chromatography on silica gel (EtOAc/cyclohexane 95/5, 99/1 to EtOAc/MeOH 95/5) to afford 86 mg of **12b** as a colorless oil (63%, 86% from **10b**) and 23 mg of **11b** as a colorless oil 20%, 77% from **9b**).

R_f = 0.38 (AcOEt/cyclohexane/MeOH: 80/15/5). ¹H NMR (CDCl₃, 300 MHz): δ_H 1.10 (3H, t, *J* = 7.2 Hz, CH₂CH₃); 1.25 (1H, ddd, *J* = 13.8, 12.1, 2.2 Hz, H_{8ax}); 1.43 (1H, dddd, *J* = 12.4, 9.8, 9.8, 8.9 Hz, H₁); 1.58 (1H, ddd, *J* = 14.4, 7.1, 2.3 Hz, H₆); 1.95 (1H, ddd, *J* = 13.4, 13.4, 13.4 Hz, H_{8eq}); 2.12 (1H, dddd, *J* = 12.3, 8.3, 5.4, 2.6 Hz, H₁); 2.22 (1H, ddd, *J* = 17.0, 9.7, 2.6 Hz, H₂); 2.29 (1H, m, H₂); 2.40 (1H, dddd, *J* = 14.4, 3.3, 1.7, 1.7 Hz, H_{6eq}); 3.19 (3H, s, OMe); 3.89 (2H, m, H₇ H_{8a}); 3.97 (1H, dq, *J* = 10.8, 7.1 Hz, CH₂O); 4.03 (1H, dq, *J* = 10.8, 7.2 Hz, CH₂O); 4.27 (1H, d, *J* = 6.8 Hz, OCH₂O); 4.43 (1H, d, *J* = 6.7 Hz, OCH₂O); 4.48 (1H, dd, *J* = 6.6, 1.1 Hz, H₅). ¹³C NMR (75 MHz, CDCl₃) δ_C 14.1 (CH₃CH₂O); 26.0 (C₁); 29.3 (C₆); 30.3 (C₂); 37.1 (C₈); 48.3 (C₅); 49.5 (C_{8a}); 55.4 (CH₂OMe); 61.1 (OCH₂CH₃); 67.7 (C₇); 93.9 (OCH₂O); 170.4 174.8 (CO C₃). IR (NaCl disc, cm⁻¹): ν_{max} = 3468, 2938, 1744, 1694, 1417, 1292, 1199, 1038. MS (ES⁺): *m/z* 294 ([MNa]⁺). HRMS (ES⁺, [MH]⁺): Calcd for C₁₃H₂₁NO₅ 272.1498 Found 272.1494.

(5S*,7S*,8aR*)-5-hydroxymethyl-7-methoxymethoxy-hexahydro-indolizin-3-one 11b

R_f = 0.09 (AcOEt/cyclohexane/MeOH: 80/15/5). ¹H NMR (CDCl₃, 300 MHz): δ_H 1.13 (1H, ddd, *J* = 11.7, 11.7, 11.7 Hz, H_{8ax}); 1.44 (1H, ddd, *J* = 13.2, 13.2, 6.6 Hz, H₆); 1.58 (1H, m, H₁); 2.05 (1H, bd, *J* = 13.2 Hz, H₆); 2.18 (2H, m, H₁ H_{8eq}); 2.35 (2H, m, 2 H₂); 3.30 (3H, s, OMe); 3.57 (2H, bd, *J* = 6.7 Hz, CH₂OH); 3.67 (1H, m, H_{8a}); 3.83 (1H, m, H₇); 4.30 (1H, ddd, *J* = 6.6, 6.6, 6.6 Hz, H₅); 4.62 (2H, bs, OCH₂O). ¹³C NMR (75 MHz, CDCl₃) δ_C 25.4 (C₁); 30.5 (C₂); 31.6 (C₆); 40.3 (C₈); 49.7 (C₅); 53.2 (C_{8a}); 55.4 (CH₂OMe); 62.5 (CH₂OH); 70.4 (C₇); 94.9 (OCH₂O); 175.0 (C₃). IR (NaCl disc, cm⁻¹): ν_{max} = 2943, 1672, 1656, 1421, 1287, 1111, 1064. HRMS (ES⁺, [MH]⁺): Calcd for C₁₁H₁₉NO₄ 252.1212 Found 272.1205.

(2S*,5S*,7S*,8aR*)-2-Allyl-7-benzyloxy-5-hydroxymethyl-hexahydro-indolizin-3-one 16a and (2R*,5S*,7S*,8aR*)-2-Allyl-7-benzyloxy-5-hydroxymethyl-hexahydro-indolizin-3-one 17a

A 15 mL LDA solution in THF (from diisopropylamine (22.1 mmol; 3.1 mL; 2.1 equiv) and BuLi 1.94 M in hexane (22.1 mmol; 11.4 mL) cooled to -78 °C was cannulated to amide **11a** (10.5 mmol; 2.9 g) in THF (35 mL) maintained at

-78 °C under argon. The solution was stirred at the same temperature for 20 min and allyl bromide (11.6 mmol; 1 mL; 1.1 equiv). was slowly added. The mixture was stirred at -78 °C for 2h and then treated with an aqueous saturated NH₄Cl solution (20 mL). The aqueous phase was extracted several times with CH₂Cl₂. The organic phase was then dried with MgSO₄ and the solvent removed in vacuo. The residue was purified by flash chromatography (AcOEt/MeOH: 99.5/0.5 to 95/5) to give the two diastereomeric compounds **16a** and **17a** in a 87/13 ratio and a 46% overall yield. (827 mg of pure **16a** as a pale yellow oil, 97 mg of pure **17a** as a pale yellow oil and 778 mg of a mixture were isolated).

16a: R_f = 0.42 (AcOEt/MeOH: 98/2). ¹H NMR (CDCl₃, 400 MHz) δ_H (ppm): 1.17 (1H, ddd, *J* = 11.6, 11.6, 11.6 Hz, H_{8ax}); 1.51 (1H, m, H₆); 1.79 (1H, m, H₁); 2.01 (1H, m, H₁); 2.13-2.28 (3H, m, H₆ H₈ CH₂CH=CH₂); 2.44 (1H, m, CH₂CH=CH₂); 2.59 (1H, m, H₂); 3.1 (1H, bs, H-O); 3.5-3.8 (4H, m, CH₂OH H_{8a} H₇); 4.38 (1H, ddbd, *J* = 6.8, 6.8, 6.8 Hz, H₅); 4.55 (2H, s, CH₂Ph); 5.06 (1H, bd, *J* = 9.1 Hz, CH₂=CH); 5.09 (1H, bd, *J* = 9.1 Hz, CH₂=CH); 5.77 (1H, dddd, *J* = 16.0, 9.0, 7.0, 7.0 Hz, CH₂=CH); 7.27 (5H, m, Ar). ¹³C NMR (75 MHz, CDCl₃) δ_C 30.3 (C₁); 30.8 (C₆); 35.8 (CH₂CH=CH₂); 39.4 (C₈); 40.9 (C₂); 49.7 (C₅); 51.4 (C_{8a}); 62.4 (CH₂OH); 70.1 (CH₂Ph); 71.8 (C₇); 117.3 (CH₂=CH); 127.5; 127.6; 128.3 (2C_{Ar} C_{Ar} 2C_{Ar}); 135.0 (CH₂=CH); 138.1 (C_{Arquat}); 176.4 (C₃). IR (NaCl disc, cm⁻¹): ν_{max} 3391, 2942, 1681, 1650, 1454, 1367, 1280, 1094, 917, 739. MS (ES⁺): *m/z* 338 ([MNa]⁺). HRMS (ES⁺, [MNa]⁺), Calcd for C₁₉H₂₅NO₃ 338.1732 Found 338.1717.

17a: R_f = 0.5 (AcOEt/MeOH : 98/2). ¹H (CDCl₃, 300 MHz): δ_H 1.11 (1H, ddd, *J* = 11.7, 11.7, 11.7 Hz, H_{8ax}); 1.3 (1H, ddd, *J* = 11.3, 11.3, 11.3 Hz, H₁); 1.49 (1H, ddd, *J* = 13.1, 11.6, 6.6 Hz, H₆); 2.1 (2H, m, H₆ CH₂CH=CH₂); 2.3 (2H, m, H₁ H₈); 2.57 (2H, m, H₂ CH₂CH=CH₂); 3.1 (1H, bs, H-O); 3.57 (3H, m, H_{8a} CH₂OH); 3.68 (1H, dddd, *J* = 11.2, 11.2, 4.0, 4.0 Hz, H₇); 4.37 (1H, ddd, *J* = 6.7, 6.7, 6.7, Hz, H₅); 4.52 (2H, s, CH₂Ph); 5.02 (1H, dd, *J* = 10.1, 1.6 Hz, H₁₄ CH_{2cis}=CH); 5.05 (1H, dd, *J* = 17.1, 1.6 Hz CH_{2trans}=CH); 5.71 (1H, dddd, *J* = 16.9, 10.1, 6.8, 6.8 Hz, CH₂=CH); 7.3 (5H, m, Ar). ¹³C NMR (100 MHz, CDCl₃) δ_C 31.1 (C₆); 32.4 (C₁); 35.4 (CH₂CH=CH₂); 40.0 (C₈); 41.3 (C₂); 49.9 (C₅); 51.5 (C_{8a}); 62.5 (CH₂OH); 70.4 (CH₂Ph); 72.0 (C₇); 117.0 (CH₂=CH); 127.7 (2C_{Ar}); 127.8 (C_{Ar}); 128.5 (2C_{Ar}); 135.5 (CH₂=CH); 138.4 (C_{Arquat}); 176.0 (C₃). IR (NaCl disc, cm⁻¹): ν_{max} 3396, 2927, 2867, 1666, 1454, 1367. MS (ES⁺): *m/z* 316 ([MH]⁺); 338 ([MNa]⁺); 354 ([MK]⁺). HRMS (ES⁺, [MNa]⁺), Calcd for C₁₉H₂₅NO₃ 338.1732 found 338.1732.

(2S*,5S*,7S*,8aR*)-2-Allyl-5-hydroxymethyl-7-methoxymethoxy-hexahydro-indolizin-3-one 16b and (2R*,5S*,7S*,8aR*)-2-Allyl-5-hydroxymethyl-7-methoxymethoxy-hexahydro-indolizin-3-one 17b

The same experimental procedure as described for allylation was applied to lactam **11b** (100 mg; 0.43 mmol) using LDA prepared from diisopropylamine (152 μL; 1.07 mmol; 2.5 equiv) and butyllithium 2.28 M (471 μL; 1.07 mmol; 2.5 equiv). Stirring was maintained 3h after addition of allyl bromide (41 μL; 0.47 mmol; 1.1 equiv). Flash chromatography on silica gel (AcOEt/MeOH: 95/5) allowed the isolation of **16b** and **17b** as a 71/29 ratio in a 54% yield. (**17b**: 8 mg as a pale yellow oil, **16b**: 22 mg as a pale yellow oil and 32 mg of a mixture).

16b: R_f = 0.38 (AcOEt/MeOH: 90/10); ¹H NMR (CDCl₃, 300 MHz) δ_H 1.15 (1H, ddd, *J* = 11.7, 11.7, 11.7, Hz, H_{8ax}); 1.33 (1H, m, H₁); 1.55 (1H, ddd, *J* = 13.2, 11.7, 6.7 Hz, H_{6ax}); 2.11 (2H, m, H_{6eq} CH₂CH=CH₂); 2.28 (1H, bd, *J* = 12.1 Hz, H_{8eq}); 2.37 (1H, ddd, *J* = 12.6, 8.5, 6.7 Hz, H₁); 2.61 (2H, m, H₂ CH₂CH=CH₂); 3.39 (3H, s, CH₂OMe); 3.62 (1H, m, H_{8a}); 3.66 (2H, d, *J* = 6.9 Hz, CH₂OH); 3.87 (1H, dddd, *J* = 11.4, 11.4, 4.3, 4.3 Hz, H_{7ax}); 4.4 (1H, ddd, *J* = 6.7, 6.7, 6.7 Hz, H_{5eq}); 4.7 (2H, s, OCH₂O); 5.06 (1H, dd, *J* = 10.1, 1.6 Hz, CH₂=CH); 5.11 (1H, dd, *J* = 17.1, 1.6 Hz, CH₂=CH); 5.27 (1H, dddd, *J* = 17.0, 10.1, 6.8, 6.8 Hz, CH₂=CH). ¹³C NMR (CDCl₃, 100 MHz) δ_C 31.6 (C₆); 32.6 (C₁); 35.4 (CH₂CH=CH₂); 40.4 (C₈); 41.3 (C₂); 49.8 (C₅); 51.3 (C_{8a}); 55.5 (CH₂OMe); 63.5 (CH₂OH); 70.5 (C₇); 95.0 (OCH₂O); 126.9 (CH₂=CH); 135.5 (CH₂=CH); 176.3 (C₃). IR (NaCl disc, cm⁻¹): ν_{max} = 3403, 2931, 1667, 1436, 1276, 1151, 1105, 1039, 916. MS (ES⁺): *m/z* 270 ([MH]⁺) 292 ([MNa]⁺). HRMS (ES⁺, [MNa]⁺): Calcd for C₁₄H₂₃NO₄ 292.1525 Found :292.1534.

From **15** : to silyl compound **15** (35 mg; 0.87 mmol) dissolved in 14 mL of anhydrous THF was added slowly a solution 1 M of tetrabutylammonium fluoride in THF (2.6 mL; 2.6 mmol; 3 equiv) at 0 °C. Stirring was continued 30 min after removal of the ice bath. At 0 °C 7 mL of water was added, the mixture was extracted 3 times with dichloromethane. The organic phase was dried with MgSO₄ and the solvent removed in vacuo. The residue was purified by flash chromatography (AcOEt/MeOH: 97/3) to give **17b** as a pale yellow oil (226 mg, 97%).

17b: R_f = 0.44 (AcOEt/MeOH : 90/10). ¹H NMR (CDCl₃, 300 MHz): δ_H 1.16 (1H, ddd, *J* = 11.6, 11.6, 11.6 Hz, H_{8ax}); 1.48 (1H, ddd, *J* = 13.1, 11.7, 6.7 Hz, H_{6ax}); 1.77 (1H, ddd, *J* = 13.1, 9.5, 6.3 Hz, H₁); 2.0 (1H, m, H₁); 2.07 (1H, ddd, *J* = 13.3, 13.3, 13.3 Hz, H_{6eq}); 2.2 (2H, m, H_{8eq}, CH₂CH=CH₂); 2.45 (1H, m, CH₂CH=CH₂); 2.57 (1H, dddd, *J* = 4.3, 4.3, 8.8, 8.8 Hz, H₂); 3.34 (3H, s, CH₂OMe); 3.61 (2H, d, *J* = 7.2 Hz, CH₂OH); 3.65 (1H, m, H_{8a}); 3.85 (1H, dddd, *J* = 11.3, 11.3, 4.2, 4.2 Hz, H_{7ax}); 4.38 (1H, ddd, *J* = 6.7, 6.7, 6.7 Hz, H_{5eq}); 4.65 (2H, d, *J* = 1.2 Hz, OCH₂O); 5.06 (2H, m, CH₂=CH); 5.76 (1H, dddd, *J* = 17.0, 10.1, 7.0, 7.0 Hz, CH₂=CH). ¹³C NMR (75 MHz, CDCl₃) δ_C 30.3 (C₁); 31.4 (C₆); 35.8 (CH₂-CH=CH₂); 40.0 (C₈); 40.9 (C₂); 49.7 (C₅); 51.4 (C_{8a}); 55.4 (CH₂OMe); 62.5 (CH₂OH); 70.4 (C₇); 94.8 (OCH₂O); 117.3 (CH₂=CH); 135.1 (CH₂=CH); 176.4 (C₃). IR (NaCl disc, cm⁻¹): ν_{max} = 3397, 2943, 1684, 1441, 1279, 1151, 1105, 1034, 917. MS (ES⁺): *m/z* 270 ([MH]⁺). HRMS (ES⁺, [MNa]⁺): Calcd for C₁₄H₂₃NO₄ 292.1525 Found:

292.1520.

(5S*,7S*,8aR*)-5-(tert-butyl-dimethyl-silyloxymethyl)-7-methoxymethoxy-hexahydro-indolizin-3-one 13

Alcohol **11b** (343 mg; 1.5 mmol) in CH₂Cl₂ (10 mL) was cooled to -10 °C and triethylamine (525 µL; 3.75 mmol; 2.5 equiv) and *tert*-butyldimethylsilyl triflate (444 µL, 1.95 mmol; 1.3 equiv) were added. The solution was stirred between -5 et -10 °C for 30 min. and an aqueous saturated NaHCO₃ solution (3 mL) was added. The aqueous solution was extracted with CH₂Cl₂. The organic phase was washed with water (2 mL) and then NaCl solution, dried and the solvent removed in vacuo. The residue was purified by flash-chromatography on silica gel (Ether/MeOH: 99.5/0.5). Silyl ether **13** was obtained as a pale yellow oil in 88% yield (453 mg).

R_f = 0.45 (Ether/MeOH: 99/1). ¹H NMR (CDCl₃, 300 MHz): δ_H 0.04 (3H, s, SiMe); 0.06 (3H, s, SiMe); 0.89 (9H, s, *Sitert*Bu); 1.15 (1H, ddd, *J* = 11.8, 11.8, 11.8 Hz, H_{8ax}); 1.49 (1H, ddd, *J* = 13.2, 11.8, 6.7 Hz, H_{6ax}); 1.61 (1H, dddd, *J* = 12.4, 9.5, 9.5, 7.5 Hz, H₁); 2.20 (3H, m, H_{6eq} H_{8eq} H₁); 2.37 (2H, m, 2H₂); 3.36 (3H, s, OMe); 3.61 (1H, dd, *J* = 10.0, 4.7 Hz, CH₂O); 3.68 (1H, dd, *J* = 10.0, 6.0 Hz, CH₂OH); 3.76 (1H, dddd, *J* = 11.5, 7.3, 7.3, 3.1 Hz, H_{8a}); 4.06 (1H, dddd, *J* = 11.3, 11.3, 4.2, 4.2 Hz, H_{7ax}); 4.30 (1H, ddd, *J* = 5.8, 5.8, 5.8 Hz, H_{5eq}); 4.68 (2H, s, OCH₂O). ¹³C NMR (CDCl₃, 100 MHz) δ_C -5.49, -5.53 (2 SiMe); 18.1 (SiC(CH₃)₃); 25.7 (C₁); 25.9 (3 SiC(CH₃)₃); 30.5 (C₂); 32.1 (C₆); 40.1 (C₈); 49.0 (C₅); 54.2 (C_{8a}); 55.3 (OMe); 64.0 (CH₂O); 70.5 (C₇); 94.8 (OCH₂O); 173.8 (C₃). IR (NaCl disc, cm⁻¹): ν_{max} = 2886, 1689, 1416, 1257, 1154, 1103, 1041, 837, 777. MS (ES⁺): *m/z* 366 ([MNa]⁺). HRMS (ES⁺, [MNa]⁺): Calcd for C₁₇H₃₃NO₄Si 366.2077 Found 366.2072.

(2S*,5S*,7S*,8aR*)-2-Allyl-5-(tert-butyl-dimethyl-silyloxymethyl)-7-methoxymethoxy-hexahydro-indolizin-3-one 14 and (2R*,5S*,7S*,8aR*)-2-Allyl-5-(tert-butyl-dimethyl-silyloxymethyl)-7-methoxymethoxy-hexahydro-indolizin-3-one 15

The same experimental procedure as described for allylation was applied to lactame **13** (120 mg; 0.35 mmol) using LDA prepared from diisopropylamine (63 µL; 0.45 mmol; 1.3 equiv) and butyllithium 2.3 M (195 µL; 0.45 mmol; 1.3 equiv). Stirring was maintained 3h after addition of allyl bromide (33 µL; 0.39 mmol; 1.1 equiv). Flash chromatography on silica gel (ether) allowed the isolation of **15** and **14** as a 90/10 ratio in a 88% yield. (**14**: 12 mg as a yellow oil, **15**: 106 mg as yellow oil.

(2R*,5S*,7R*,8aR*)-**15**: R_f = 0.66 (AcOEt). ¹H NMR (CDCl₃, 300 MHz): δ_H 0.04 (3H, s, SiMe); 0.05 (3H, s, SiMe); 0.87 (9H, s, *Sitert*Bu); 1.1 (1H, ddd, *J* = 11.8 Hz, H_{8ax}); 1.47 (1H, ddd, *J* = 13.6, 11.6, 6.9 Hz, H_{6ax}); 1.73 (1H, ddd, *J* = 13.0, 11.6, 6.9 Hz, H₁); 1.96 (1H, ddd, *J* = 12.9, 7.6, 4.1 Hz, H₁); 2.17 (3H, m, H₆ CH₂ CH=CH₂ H₈); 2.48 (1H, m, CH₂ CH=CH₂); 2.52 (1H, m, H₂); 3.32 (3H, s, OMe); 3.60 (1H, dd, *J* = 10.0, 4.7 Hz, CH₂O); 3.65 (1H, dd, *J* = 10.0, 5.8 Hz, CH₂O); 3.70 (1H, m, H_{8a}); 4.05 (1H, dddd, *J* = 11.3, 11.3, 4.3, 4.3 Hz, H_{7ax}); 4.29 (1H, ddd, *J* = 5.6, 5.6, 5.6 Hz, H_{5ax}); 4.64 (2H, d, *J* = 0.6 Hz, OCH₂O); 5.06 (2H, m, CH₂=CH); 5.74 (1H, dddd, *J* = 16.9, 10.1, 6.8, 6.8 Hz, CH₂=CH). ¹³C NMR (75 MHz, CDCl₃) δ_C -5.6 (2SiMe); 18.1 (SiC(CH₃)₃); 25.8 (SiC(CH₃)₃); 30.5 (C₁); 32.1 (C₆); 35.9 (CH₂ CH=CH₂); 39.8 (C₈); 40.7 (C₂); 49.0 (C₅); 52.5 (C_{8a}); 55.6 (OMe); 64.4 (CH₂O); 70.6 (C₇); 94.6 (OCH₂O); 117.1 (CH₂=CH); 135.2 (CH₂=CH); 175.1 (C₃). IR (NaCl disc, cm⁻¹): ν_{max} = 2953, 2856, 1694, 1422, 1257, 1151, 1106, 1037, 916, 837, 777. MS (ES⁺): *m/z* 406 ([MNa]⁺). HRMS (ES⁺, [MNa]⁺): Calcd for C₂₀H₃₇NO₄Si 384.2570 Found 384.2575.

(2S*,5S*,7R*,8aR*)-**14**: R_f = 0.86 (AcOEt). ¹H NMR (CDCl₃, 300 MHz): δ_H 0.04 (3H, s, SiMe); 0.06 (3H, s, SiMe); 0.89 (9H, s, *Sitert*Bu); 1.1 (1H, ddd, *J* = 11.7, 11.7, 11.7 Hz, H_{8ax}); 1.27 (1H, m, H₁); 1.5 (1H, ddd, *J* = 11.8, 11.8, 6.9 Hz, H_{6ax}); 2.07-2.28 (3H, m, CH₂ CH=CH₂ H_{8(eq)} H_{6(eq)}); 2.33 (1H, m, H₁); 2.5 (1H, dddd, *J* = 9.9, 9.9, 9.9, 3.9 Hz, H₂); 2.65 (1H, ddd, *J* = 14.1, 5.3, 5.3 Hz, CH₂ CH=CH₂); 3.36 (3H, s, OMe); 3.64 (3H, m, H_{8a} CH₂ CH=CH₂); 4.08 (1H, dddd, *J* = 11.2, 11.2, 4.0, 4.0 Hz, H_{7ax}); 4.28 (1H, ddd, *J* = 5.8, 5.8, 5.8 Hz, H_{5eq}); 4.68 (2H, bs, OCH₂O); 5.06 (2H, m, CH₂=CH); 5.77 (1H, dddd, *J* = 17.0, 10.1, 6.9, 6.9 Hz, CH₂=CH). ¹³C NMR (CDCl₃, 100 MHz) δ_C -5.6 (2SiMe); 18.1 (SiC(CH₃)₃); 25.8 (SiC(CH₃)₃); 32.1 (C₆); 32.9 (C₁); 35.4 (CH₂ CH=CH₂); 40.2 (C₈); 41.3 (C₂); 49.1 (C₅); 52.4 (C_{8a}); 55.2 (OMe); 63.9 (CH₂O); 70.6 (C₇); 94.8 (OCH₂O); 116.7 (CH₂=CH); 135.7 (CH₂=CH); 174.5 (C₃). IR (NaCl disc, cm⁻¹): ν_{max} = 2930, 1691, 1419, 1255, 1104, 1042, 840. MS (ES⁺): *m/z* = 384 ([MH]⁺), 406 ([MNa]⁺). HRMS (ES⁺, [MNa]⁺): Calcd for C₂₀H₃₇NO₄Si 406.2390 Found 406.2386.

(2S*,5S*,7S*,8aR*)-2-allyl-7-benzyloxy-3-oxo-octahydro-indolizin-5-carboxylic acid 18

To alcohol **16a** (757 mg; 2.4 mmol) dissolved in 22 mL in acetone was added dropwise Jones reagent 2.67 M (1.1 mL; 2.9 mmol; 1.2 equiv) at 0 °C. Stirring was continued for 40 min. 10 mL of isopropanol was added, the mixture was stirred for 20 min at 0 °C. After addition of 0.5 mL of water and 5 mL of 1 M HCl, the mixture was extracted twice with 100 mL of dichloromethane. The organic phase was dried (MgSO₄) and the solvent removed in vacuo. The residue was purified by flash-chromatography on silica gel (AcOEt/MeOH/AcOH: 96/2/2) and the carboxylic acid **23** was isolated as a beige solid (67%; 530 mg).

R_f = 0.27 (AcOEt/MeOH/AcOH: 96/2/2). mp: 146 °C. ¹H NMR (CDCl₃, 400 MHz) δ_H (ppm): 1.18 (1H, ddd, *J* = 11.7, 11.7, 11.7 Hz, H_{8ax}); 1.36 (1H, m, H₁); 1.63 (1H, ddd, *J* = 12.1, 12.1, 6.7 Hz, H₆); 2.18 (1H, m, CH₂CH=CH₂); 2.27-2.4 (2H, m, H₁ H₈); 2.58-2.69 (3H, m, H₆, H₂, CH₂CH=CH₂); 3.57 (1H, m, H₇); 3.72 (1H, m, H_{8a}); 4.54 (1H, d, *J* = 9.1 Hz, CH₂Ph); 4.59 (1H, d, *J* = 9.1 Hz, CH₂Ph); 4.91 (1H, bd, *J* = 6.2 Hz, H₅); 5.03-5.13 (2H, m, CH₂=CH); 5.75 (1H,

dddd, $J = 16.9, 10.1, 6.8, 6.8$ Hz, $\text{CH}_2=\text{CH}$); 7.31 (5H, m, Ar). ^{13}C NMR (75 MHz, CDCl_3) δ_{C} 31.9 (C_6); 32.4 (C_1); 35.0 ($\text{CH}_2\text{CH}=\text{CH}_2$); 39.2 (C_8); 41.4 (C_2); 50.5 (C_5); 52.4 (C_{8a}); 70.6 (CH_2Ph); 72.7 (C_7); 117.3 ($\text{CH}_2=\text{CH}$); 127.7; 127.9; 128.6 (2C_{Ar} C_{Ar} 2C_{Ar}); 135.2 ($\text{CH}_2=\text{CH}$); 138.1 (C_{Arquat}); 173.1 (COOH); 176.4 (C_3). MS (ES^+): m/z 352 ($[\text{MNa}]^+$).

(2S)-2-[(2S*,5S*,7S*,8aR*)-(2-allyl-7-benzyloxy-3-oxo-octahydro-indolizin-5-carbonyl)-amino]succinic acid di-tert-butyl ester 19

To carboxylic acid **18** (55 mg; 0.167 mmol) dissolved in 1.5 mL of anhydrous dichloromethane was added N-methylmorpholine (NMM) (20 μL , 0.18 mmol, 1.1equiv) at -10 °C under argon, the mixture was stirred for 30 min at the same temperature. Isobutyl chloroformiate (23 μL , 0.18 mmol, 1.1 equiv) was added dropwise and stirring was continued for 1 h at -10 °C. The amino acid (51 mg, 0.18 mmol, 1.1 equiv) was dissolved in 1.5 mL of anhydrous dichloromethane under argon, NMM (40 μL , 0.36 mmol, 2.2 equiv) was added at 0 °C. This solution was stirred for 30 min and added via a syringe to the mixed anhydride solution. Stirring was continued for 20 h at rt. The residue was taken off with 10 mL of ethyl acetate, washed with 5 mL of 10% Na_2CO_3 solution, 5 mL of 10% KHSO_4 solution and 5 mL of brine. The organic phase was dried over Na_2SO_4 and the solvent removed in vacuo. The residue was purified by flash-chromatography on silica gel (AcOEt/cyclohexane: 25/75 to 70/30) and the compound was isolated as a white foam (88%, 65 mg).

2 diastereoisomers $R_f = 0.53$ (AcOEt/cyclohexane: 40/60). ^1H NMR (CDCl_3 , 400 MHz) δ_{H} (ppm): 1.16/1.17 (1H, ddd, $J = 11.8, 11.8, 11.8$ Hz, H_{8ax}); 1.23-1.52 (20H, m, H_1 , H_6 , $2\text{C}(\text{CH}_3)_3$); 2.11-2.5 (3H, m, H_1 , $\text{CH}_2\text{CH}=\text{CH}_2$, H_8); 2.51-2.9 (5H, m, $\text{CH}_2\text{CH}=\text{CH}_2$, CH_2CO , H_2 , H_6); 3.58-3.89 (2H, m, H_7 H_{8a}); 4.51-4.67 (3H, m, CH_2Ph , NHCHCO); 4.85 (1H, bd, $J = 6.2$ Hz, H_5); 5.02-5.14 (2H, m, $\text{CH}_2=\text{CH}$); 5.67-5.84 (1H, m, $\text{CH}_2=\text{CH}$); 6.96/7.20 (1H, d/d, $J = 8.0/8.6$ Hz, NH); 7.23-7.37 (5H, m, Ar). ^{13}C NMR (75 MHz, CDCl_3) δ_{C} 27.9 and 28.0 ($\text{C}(\text{CH}_3)_3$); 30.8/31.2 32.2/32.4 35.1 37.2/37.3 (C_6 , C_1 , $\text{CH}_2\text{CH}=\text{CH}_2$, CH_2CO); 39.6/39.7 (C_8); 40.9/41.0 (C_2); 49.1/49.3 50.9 52.52.1 (C_5 , C_{8a} , NHCHCO); 70.5 (CH_2Ph); 72.4 (C_7); 81.6/81.9 and 82.2/82.4 ($\text{C}(\text{CH}_3)_3$); 117 ($\text{CH}_2=\text{CH}$); 127.6; 128.3 (2C_{Ar} C_{Ar} 2C_{Ar}); 135.3 ($\text{CH}_2=\text{CH}$); 138.1 (C_{Arquat}); 169.2/169.3 169.5/169.5 169.9/170 175.6/175.7 (3CO, C_3). HRMS (ES^+ , $[\text{MNa}]^+$): Calcd for $\text{C}_{31}\text{H}_{44}\text{N}_2\text{O}_7$ 579.3046 Found 579.3019.

3-[(2S*,5S*,7S*,8aR*)-(2-allyl-7-benzyloxy-3-oxo-octahydro-indolizin-5-carbonyl)-amino]propionic acid tert-butyl ester 20

The same experimental procedure as described for peptide coupling was applied to carboxylic acid **18** (136 mg; 0.41 mmol) using NMM (50 μL , 0.45 mmol, 1.1equiv), isobutyl chloroformiate (60 μL , 0.45 mmol, 1.1 equiv). Stirring was maintained 20 h after addition of amino acid (82 mg, 0.45 mmol, 1.1 equiv) and NMM (100 μL , 0.9 mmol, 2.2 equiv). Flash chromatography on silica gel (AcOEt/cyclohexane: 50/50 to 70/30) allowed the isolation of a colorless oil (83%, 154 mg).

$R_f = 0.45$ (AcOEt/cyclohexane: 60/40). ^1H NMR (CDCl_3 , 400 MHz) δ_{H} (ppm): 1.17 (1H, ddd, $J = 11.8, 11.8, 11.8$ Hz, H_{8ax}); 1.2-1.5 (11H, m, H_1 , H_6 , $2\text{C}(\text{CH}_3)_3$); 2.18 (1H, m, H_1); 2.29 (1H, bd, $J = 12$ Hz, H_8); 2.35 (1H, m, $\text{CH}_2\text{CH}=\text{CH}_2$); 2.41 (2H, t, $J = 6$ Hz, $\text{NHCH}_2\text{CH}_2\text{CO}$); 2.52-2.7 (3H, m, H_2 , H_6 , $\text{CH}_2\text{CH}=\text{CH}_2$); 3.32-3.52 (2H, m, $\text{NHCH}_2\text{CH}_2\text{CO}$); 3.57 (1H, m, H_{8a}); 3.9 (1H, dddd, $J = 11, 11, 4.5, 4.5$ Hz, H_7); 4.57 (1H, d, $J = 11.6$ Hz, CH_2Ph); 4.63 (1H, d, $J = 11.6$ Hz, CH_2Ph); 4.78 (1H, d, $J = 5.9$ Hz, H_5); 5.02-5.12 (2H, m, $\text{CH}_2=\text{CH}$); 5.77 (1H, dddd, $J = 15, 12, 5.9, 5.9$ Hz, $\text{CH}_2=\text{CH}$); 6.65 (1H, bs, NH); 7.25-7.4 (5H, m, Ar). ^{13}C NMR (75 MHz, CDCl_3) δ_{C} 28.1 ($\text{C}(\text{CH}_3)_3$); 30.9 (C_6); 32.3 (C_1); 35.1 ($\text{CH}_2\text{CH}=\text{CH}_2$, $\text{NHCH}_2\text{CH}_2\text{CO}$); 39.5 (C_8); 41.1 (C_2); 50.9 (C_5); 52.1 (C_{8a}); 70.5 (CH_2Ph); 72.4 (C_7); 81.1 ($\text{C}(\text{CH}_3)_3$); 117.1 ($\text{CH}_2=\text{CH}$); 127.6; 128.4 (2C_{Ar} C_{Ar} 2C_{Ar}); 135.2 ($\text{CH}_2=\text{CH}$); 138.5 (C_{Arquat}); 169.9 171.4 176.4 (2CO C_3). IR (NaCl disc, cm^{-1}): $\nu_{\text{max}} = 3307, 2929, 1727, 1682, 1427, 1278, 1156, 737$. MS (ES^+): m/z 479 ($[\text{MNa}]^+$).

(S)-2-[(2S*,5S*,7S*,8aR*)-(7-benzyloxy-2-(3-hydroxy-propyl)-3-oxo-octahydro-indolizin-5-carbonyl)-amino] succinic acid di-tert-butyl ester 21

The same experimental procedure as described for hydroboration was applied to alkene **19** (81 mg; 0.14 mmol) using a 0.5 M solution of 9-BBN (1.5 mL; 0.73 mmol; 5 equiv). Stirring was maintained 15 h after addition of sodium acetate 5 M (570 μL , 2.8 mmol, 20 equiv) and a 30 % H_2O_2 solution (0.160 mL; 22 equiv). Flash chromatography on silica gel (AcOEt/cyclohexane: 50/50 to 90/10) allowed the isolation of a yellow oil (68%, 57 mg).

2 diastereoisomers $R_f = 0.16$ (AcOEt/cyclohexane: 90/10). ^1H NMR (CDCl_3 , 400 MHz) δ_{H} (ppm): 1.13/1.15 (1H, ddd/ddd, $J = 11.7, 11.7, 11.7$ Hz, H_8); 1.3-1.98 (23H, m, H_6 , $2\text{C}(\text{CH}_3)_3$, $\text{CH}_2\text{CH}_2\text{CH}_2\text{OH}$); 2.27 (1H, m, H_8); 2.32-2.87 (5H, m, H_1 , CH_2CO , H_2 , H_6); 3.6-3.66 (3H, m, CH_2OH , H_{8a}); 3.72-3.82 (1H, m, H_7); 4.5-4.64 (3H, m, CH_2Ph , NHCHCO); 4.79/4.85 (1H, m, H_5); 6.74/6.82 (1H, d/d, $J = 8.0/8.6$ Hz, NH); 7.25-7.35 (5H, m, Ar). ^{13}C NMR (75 MHz, CDCl_3) δ_{C} 20.2/27.1 (CH_2); 28.0 and 28.1 ($\text{C}(\text{CH}_3)_3$); 30.3 31.0 31.4 36.4 37.3 (CH_2); 39.6/39.7 (C_8); 41.2/41.3 (C_2); 49.1/49.4 (NHCHCO); 51.0 (C_5); 52.1/52.3 (C_{8a}); 62.2 (CH_2OH); 70.6 (CH_2Ph); 72.4/72.5 (C_7); 81.7 82.0 and 82.3/82.5 ($\text{C}(\text{CH}_3)_3$); 127.6; 128.5 (2C_{Ar} C_{Ar} 2C_{Ar}); 138.1 (C_{Arquat}); 169.4/169.6 170.1 170.2 176.7/176.8 (3CO, C_3).

3-[(2S*,5S*,7S*,8aR*)-(7-benzyloxy-2-(3-hydroxy-propyl)-3-oxo-octahydro-indolizin-5-carbonyl)-amino]propionic acid tert-butyl ester 22

The same experimental procedure as described for hydroboration was applied to alkene **20** (106 mg; 0.23 mmol) using a 0.4 M solution of 9-BBN in hexane (1.5 mL; 0.58 mmol; 2.5 equiv). Stirring was maintained 16 h after addition of sodium acetate 5 M (550 μ L, 2.76 mmol, 12 equiv) and a 30 % H₂O₂ solution (0.280 mL; 12 equiv). Flash chromatography on silica gel (AcOEt/MeOH: 100 to 99/1) allowed the isolation of a yellow oil (71%, 74 mg).

R_f = 0.3 (AcOEt/MeOH: 95/5). ¹H NMR (CDCl₃, 300 MHz) δ _H (ppm): 1.14 (1H, ddd, *J* = 11.8, 11.8, 11.8 Hz, H_{8ax}); 1.22-1.52 (12H, m, H₁, H₆, CH₂CH₂CH₂OH, 2C(CH₃)₃); 1.6 (2H, q, *J* = 11.8 Hz, CH₂CH₂CH₂OH); 1.93 (1H, m, CH₂CH₂-CH₂OH); 2.27 (1H, bd, *J* = 12 Hz, H₈); 2.39 (2H, t, *J* = 6 Hz, NHCH₂CH₂CO); 2.41-2.54 (2H, m, H₂, H₁); 2.6 (1H, bd, *J* = 12 Hz, H₆); 3.04 (1H, s, OH); 3.2-3.5 (2H, m, NHCH₂CH₂CO); 3.63 (1H, m, H_{8a}); 3.63 (2H, t, *J* = 6.1 Hz, CH₂OH); 3.88 (1H, dddd, *J* = 12, 12, 4.5, 4.5 Hz, H₇); 4.55 (1H, d, *J* = 11.7 Hz, CH₂Ph); 4.62 (1H, d, *J* = 11.7 Hz, CH₂Ph); 4.74 (1H, d, *J* = 5.9 Hz, H₅); 6.79 (1H, t, *J* = 6 Hz, NH); 7.2-7.45 (5H, m, Ar). ¹³C NMR (75 MHz, CDCl₃) δ _C: 27.2 (CH₂CH₂CH₂OH); 28.1 (C(CH₃)₃); 29.7 (CH₂CH₂CH₂OH); 31.1 (C₆); 33.3 (C₁); 35.1 (NHCH₂CH₂CO); 39.5 (C₈); 41.3 (C₂); 50.9 (C₅); 52.3 (C_{8a}); 62.1 (CH₂OH); 70.5 (CH₂Ph); 72.3 (C₇); 81.1 (C(CH₃)₃); 127.6; 128.4 (2C_{Ar} C_{Ar} 2C_{Ar}); 138.4 (C_{Arquat}); 169.9 171.4 176.4 (2CO C₃).

guanidine carbamate **23**

Alcohol **21** (60 mg, 0.1 mmol) and 1,3-bis(*tert*-butoxycarbonyl) guanidine (53 mg, 0.2 mmol, 2 equiv) was dried under vacuo, dissolved in 2.5 mL of anhydrous toluene under argon. Tributylphosphine (50 μ L, 0.2 mmol, 2 equiv) was added to the solution at rt, then dipiperidinazodicarboxylate (ADDP) (52 mg, 0.2 mmol, 2 equiv) at 0 °C. The mixture was refluxed for 4 h. After removal of the toluene under vacuo, the residue was purified by flash chromatography on silica gel (Et₂O/cyclohexane/MeOH: 85/14/1) allowed the isolation of **23** as a colorless oil (46 mg, 60%).

2 diastereoisomers R_f = 0.14 (Et₂O/cyclohexane/MeOH: 85/14/1). ¹H NMR (CDCl₃, 400 MHz) δ _H (ppm): 1.01-1.4 (31H, m, H₈, H₆, 3C(CH₃)₃, CH₂CH₂CH₂OH); 1.5-1.7 (2H, m, CH₂CH₂CH₂OH); 1.5-1.7 (1H, m, CH₂CH₂CH₂OH); 2.1-2.4 (2H, m, H₁, H₈); 2.4-2.88 (4H, m, CH₂CO, H₂, H₆); 3.48-3.62 (1H, m, H_{8a}); 3.62-3.78 (1H, m, H₇); 4.03 (2H, bs, CH₂OH); 4.43-4.59 (3H, m, CH₂Ph, NHCHCO); 4.76 (1H, d, *J* = 11.7 Hz, H₅); 7.21/7.26 (1H, d/d, *J* = 8.1/8.6 Hz, NH); 7.15-7.29 (5H, m, Ar). ¹³C NMR (75 MHz, CDCl₃) δ _C: 26.7 27.6/27.7 (CH₂CH₂CH₂OH); 28.0 and 28.1 (3C(CH₃)₃); 29.8/30.4 30.9/31.3 33.3/33.4 (C₆, C₁, CH₂CO); 39.6/39.8 (C₈); 41.2/41.3 (C₂); 49.1/49.4 50.9/51.0 52.0/52.2 (C₅, NHCHCO, C_{8a}); 65.5 (CH₂OH); 70.9 (CH₂Ph); 72.4/72.5 (C₇); 81.7 81.9 82.2 and 82.5 (C(CH₃)₃); 127.7 127.8 128.5 (2C_{Ar} C_{Ar} 2C_{Ar}); 138.6 (C_{Arquat}); 158.5 169.4 169.7 169.8 170 170.1 170.2 176/176.2 (C=N 5CO C₃). MS (ES⁺): *m/z* 760 ([MH]⁺). HRMS (ES⁺, [MNa]⁺): Calcd for C₃₈H₅₇N₅O₁₁ 782.3952 Found 782.3969.

peptidomimetic **24**

TFA (0.5 mL; 6.5 mmol) was added to a cooled (0 °C) solution of compound **23** (46 mg; 0.06 mmol) dissolved in CH₂Cl₂ (2 mL) with a droplet of water. The mixture was stirred at rt for 4h. The mixture was concentrated in vacuo after addition of toluene (2 mL). Two more additions and evaporations of toluene were done and finally the residue was taken in ether (1 mL) and extracted with water (3 times). The aqueous phase was lyophilysed to leave **24** as a foam in a quantitative yield (33 mg).

2 diastereoisomers mp: 60 °C. ¹H NMR (D₂O, 400 MHz) δ _H (ppm): 1.07 (1H, ddd, *J* = 11.7, 11.7, 11.7 Hz, H₈); 1.32 (2H, m, CH₂CH₂CH₂O, H₁); 1.52 (1H, m, H₆); 1.65 (2H, m, CH₂CH₂CH₂O); 1.75 (1H, m, CH₂CH₂CH₂O); 2.1 (1H, bd, *J* = 10.3 Hz, H₈); 2.24 (1H, m, H₆); 2.4 (1H, m, H₁); 2.59 (1H, m, H₂); 2.7-2.88 (2H, m, CH₂COOH); 3.65-3.8 (2H, m, H₇ H_{8a}); 4.12 (2H, t, *J* = 6.0 Hz, CH₂OCO); 4.48-4.70 (3H, m, NHCHCO, CH₂Ph); 4.70 (1H, m, H₅); 7.29 (5H, m, Ar). ¹³C NMR (100 MHz, MeOD) δ _C: 25.6 (CH₂CH₂CH₂OH); 26.6 (CH₂CH₂CH₂OH); 32.6 et 32.7 (C₁); 34.4 et 34.5 (C₆); 35.4 (CH₂COOH); 41.0 (C₈); 41.3 (C₂); 48.9 (NHCHCO); 51 (C₅); 52.7 et 52.9 (C_{8a}); 64.2 (C₇); 67.0 (CH₂OCO); 70.3 (CH₂Ph); 127.5; 127.7; 128.1 and 128.3 (2C_{Ar} C_{Ar} 2C_{Ar}); 138.1 (C_{Arquat}); 153.0 and 155.4 (C=N OCONH); 170.7; 170.8; 172.6; 173; 173.1 and 177.2 (2NHCO 2 COOH C₃). [MH]⁺. HRMS (ES⁺, [MH]⁺): Calcd for C₂₅H₃₃N₅O₉ 548.2357 Found 548.2339.

3-[[[(2*S**,5*S**,7*S**,8*R**)-7-benzyloxy-2-(3-bis(*tert*-butyloxycarbonyl)guanidino-propyl)-3-oxo-octahydro-indolizin-5-carbonyl]-amino]-propionic acid *tert*butyl ester **25**

Alcohol **22** (76 mg, 0.16 mmol) and 1,3-bis(*tert*-butoxycarbonyl) guanidine (70 mg, 0.27 mmol, 2.1 equiv) was dried under vacuo, dissolved in 5 mL of anhydrous THF under argon. Triphenylphosphine (87 mg, 0.33 mmol, 2 equiv) was added to the solution at rt, then diethylazodicarboxylate (DEAD) (52 μ L, 0.33 mmol, 2 equiv) at 0 °C. The mixture was stirred for 60 h. at rt. After addition of dichloromethane, the solvent was removed in vacuo. The residue was purified by flash chromatography on silica gel (Et₂O/cyclohexane: 90/10) to give a white foam (105 mg, 92%).

R_f = 0.22 (Et₂O/cyclohexane: 90/10). ¹H NMR (CDCl₃, 300 MHz) δ _H (ppm): 1.15 (1H, ddd, *J* = 11.7, 11.7, 11.7 Hz, H₈); 1.18-1.4 (3H, m, H₁, H₆, CH₂CH₂CH₂N); 1.44 (9H, s, C(CH₃)₃); 1.49 (9H, s, C(CH₃)₃); 1.55 (9H, s, C(CH₃)₃); 1.65 (2H, m, CH₂CH₂CH₂N); 1.94 (1H, m, CH₂CH₂CH₂N); 2.28 (1H, bd, *J* = 11.6 Hz, H₈); 2.39 (2H, t, *J* = 6 Hz, NHCH₂CH₂CO); 2.43-2.49 (2H, m, H₂, H₁); 2.63 (1H, bd, *J* = 12.7 Hz, H₆); 3.34-3.49 (2H, m, NHCH₂CH₂CO); 3.56 (1H, m, H_{8a}); 3.7-4.05 (3H, m, H₇, CH₂N); 4.55 (1H, d, *J* = 11.8 Hz, CH₂Ph); 4.63 (1H, d, *J* = 11.8 Hz, CH₂Ph); 4.74 (1H, d, *J* = 5.6 Hz, H₅); 6.68 (1H, t, *J* = 5.9 Hz, NH); 7.3 (5H, m, Ar); 9.1-9.4 (2H, bs, NH₂). ¹³C NMR (75 MHz, CDCl₃) δ _C: 26.2 (CH₂CH₂CH₂N); 28.1 and 28.3 (3C(CH₃)₃, CH₂CH₂CH₂N); 30.8 (C₆); 33.3 (C₁); 35.2 (NHCH₂CH₂CO); 39.6 (C₈); 41.1 (C₂); 44.3 (CH₂CH₂CH₂N); 50.9 (C₅); 52.3 (C_{8a}); 70.6 (CH₂Ph); 72.4 (C₇); 78.6 81.1

and 83.7 (3C(CH₃)₃); 127.6; 128.3 (2C_{Ar} C_{Ar} 2C_{Ar}); 138.5 (C_{Arquat}); 155 160.5 and 163.8 (C=N CON); 169.9 171.3 176.4 (2CO C₃). IR (NaCl disc, cm⁻¹): ν_{max} = 3382, 2977, 1715, 1608, 1514, 1368, 1251, 1152, 737. MS (ES⁺): *m/z* 739([MNa]⁺).

3-[(2S*,5S*,7S*,8aR*)-7-benzyloxy-2-(3-guanidino-propyl)-3-oxo-octahydro-indolizin-5-carbonyl]-amino]-propionic acid **26**

The same protocol for deprotection with TFA was applied to compound **25** (43 mg; 0.06 mmol). Guanidyl compound was obtained as foam in a quantitative yield (28 mg).

¹H NMR (D₂O, 300 MHz) δ_H (ppm): 1.04 (1H, ddd, *J* = 11.7, 11.7, 11.7 Hz, H₈); 1.28 (2H, m, H₁, CH₂CH₂CH₂NH); 1.48 (3H, m, CH₂CH₂CH₂NH H₆); 1.7 (1H, m, CH₂CH₂CH₂NH); 2.21 (1H, bd, *J* = 11.7 Hz, H₈); 2.27-2.48 (4H, m, NHCH₂CH₂CO H₆ H₁); 2.55 (1H, m, H₂); 3.06 (2H, t, *J* = 6.0 Hz, CH₂CH₂CH₂NH); 3.31 (2H, t, *J* = 5.7 Hz, NHCH₂CH₂CO); 3.51 (1H, m, H₇); 3.65 (1H, m, H_{8a}); 4.47 (1H, s, CH₂Ph); 4.62 (1H, d, *J* = 5.6 Hz, H₅); 7.28 (5H, m, Ar). ¹³C NMR (75 MHz, D₂O) δ_C 25.2 (CH₂CH₂CH₂NH); 27.2 (CH₂CH₂CH₂NH); 31.6 (C₆); 31.9 (C₁); 33.4 (NHCH₂CH₂CO); 35.2 (NHCH₂CH₂CO); 38.2 (C₈); 40.8 (CH₂CH₂CH₂NH); 41.0 (C₂); 51.6 (C_{8a}); 53.3 (C₅); 70.2 (CH₂Ph); 71.8 (C₇); 128.3; 128.5 and 128.7 (2C_{Ar} C_{Ar} 2C_{Ar}); 137.1 (C_{Arquat}); 156.6 (C=N); 171.4 175.8 178.6 (2CO C₃). MS (ES⁺): *m/z* 460([MH]⁺). HRMS (ES⁺, [MH]⁺): Calcd for C₂₃H₃₃N₅O₅ 460.2560 Found 460.2552.

3-[(2S*,5S*,7S*,8aR*)-2-(3-guanidino-propyl)-7-hydroxy-3-oxo-octahydro-indolizin-5-carbonyl]-amino]-propionic acid **27**

To benzyl ether **25** (45 mg, 0.063 mmol) dissolved in 7 mL of methanol was added 35 mg of palladium on activated carbon (10% Pd). The mixture was put under hydrogen, stirred 24 h. After filtration on celite, washing with ethyl acetate, the solvents were removed under vacuo. The residue was purified by flash chromatography on silica gel (Ethyl acetate) to give a colorless oil (28 mg, 71%).

R_f = 0.36 (AcOEt/MeOH: 99/1). ¹H NMR (CDCl₃, 300 MHz) δ_H (ppm): 1.14 (1H, ddd, *J* = 11.7, 11.7, 11.7 Hz, H₈); 1.23-1.38 (2H, m, H₁ CH₂CH₂CH₂N); 1.38-1.45 (28H, s, H₆ 3C(CH₃)₃); 1.59-1.73 (2H, m, CH₂CH₂CH₂N); 1.95 (1H, m, CH₂CH₂CH₂N); 2.21 (1H, dm, *J* = 12 Hz, H₈); 2.4 (2H, t, *J* = 6.2 Hz, NHCH₂CH₂CO); 2.43-2.63 (3H, m, H₂ H₁ H₆); 3.42 (2H, m, NHCH₂CH₂CO); 3.59 (1H, m, H_{8a}); 3.8 (1H, m, CH₂N); 4.01 (1H, m, CH₂N); 4.13 (1H, m, H₇); 4.73 (1H, d, *J* = 5.7 Hz, H₅); 6.74 (1H, t, *J* = 4.5 Hz, NH); 9.26 (2H, bs, NH₂). ¹³C NMR (75 MHz, CDCl₃) δ_C 26.2 (CH₂CH₂CH₂N); 28.1 and 28.3 (3C(CH₃)₃); 33.3 33.9 35.1 (NHCH₂CH₂CO, C₈, C₆, C₁); 41.1 (C₂); 41.7 44.3 (NHCH₂CH₂CO CH₂CH₂CH₂N); 50.9 52.3 (C₅ C_{8a}); 65.0 (C₇); 78.6 81.1 and 83.7 (3C(CH₃)₃); 127.6; 128.3 (2C_{Ar} C_{Ar} 2C_{Ar}); 138.5 (C_{Arquat}); 155 (C=N); 160.5 and 163.8 (C=N, CON); 169.9 171.4 176.5 (2CO C₃). IR (NaCl disc, cm⁻¹): ν_{max} = 3383, 2978, 1715, 1608, 1515, 1251, 1151, 1101, 736. MS (ES⁺): *m/z* 626 ([MH]⁺).

The same protocol for deprotection with TFA was applied to compound described above (28 mg; 0.045 mmol). Guanidyl compound **27** was obtained as an oil in a quantitative yield (17 mg).

¹H NMR (D₂O, 400 MHz) δ_H (ppm): 1.14 (1H, ddd, *J* = 11.7, 11.7, 11.7 Hz, H₈); 1.24-1.35 (2H, m, H₁ CH₂CH₂CH₂N); 1.45-1.55 (2H, m, H₆ CH₂CH₂CH₂N); 1.69 (1H, m, CH₂CH₂CH₂N); 2.12 (1H, dm, *J* = 12.1 Hz, H₈); 2.21 (1H, dm, *J* = 13.2 Hz, H₆); 2.39 (1H, ddd, *J* = 12.6, 8.9, 7.1 Hz, H₁); 2.47 (1H, t, *J* = 6.4 Hz, NHCH₂CH₂CO); 2.58 (1H, dddd, *J* = 9.4, 9.4, 9.4, 4.2 Hz, H₂); 3.08 (2H, t, *J* = 6.8 Hz, CH₂CH₂CH₂N); 3.35 (2H, t, *J* = 6.4 Hz, NHCH₂CH₂CO); 3.65-3.75 (2H, m, H₇ H_{8a}); 4.62 (1H, d, *J* = 5.6 Hz, H₅). ¹³C NMR (75 MHz, D₂O) δ_C 25.2 (CH₂CH₂CH₂N); 27.3 (CH₂CH₂CH₂N); 31.5 (C₁); 33.5 (NHCH₂CH₂CO); 34.1 (C₆); 35.3 (NHCH₂CH₂CO); 40.5 (C₈); 40.8 (CH₂CH₂CH₂N); 41.0 (C₂); 51.2 (C₅); 53.3 C_{8a}); 64.3 (C₇); 156.7 (C=N); 171.7 176.1 178.8 (2CO C₃). MS (ES⁺): *m/z* 370 ([MH]⁺) 392 ([MNa]⁺). HRMS (ES⁺, [MNa]⁺): Calcd for C₁₆H₂₇N₅O₅ 370.2090 Found 370.2078.

(2R*,5S*,7S*,8aR*)-2-allyl-7-methoxymethoxy-3-oxo-octahydro-indolizin-5-carboxylic acid **28**

To alcohol **17b** (105 mg; 0.39 mmol) dissolved in 3.5 mL of acetone was added dropwise Jones reagent 2.67 M (220 μL; 0.58 mmol; 1.5 equiv) at 0 °C. Stirring was continued for 30 min. 3 mL of isopropanol was added, the mixture was stirred for 10 min at 0 °C. After addition of 1 mL of water and 1 mL of a saturated solution of KHSO₄, the mixture was extracted twice with 100 mL of dichloromethane. The organic phase was dried (MgSO₄) and the solvent removed in vacuo. The residue was purified by flash-chromatography on silica gel (AcOEt/MeOH/AcOH: 97/1/2) and the carboxylic acid **28** was isolated as a white foam (54%; 60 mg).

R_f = 0.35 (AcOEt/MeOH/AcOH: 97/1/2). ¹H (CDCl₃, 300 MHz): δ_H 1.19 (1H, ddd, *J* = 11.8, 11.8, 11.8 Hz, H_{8ax}); 1.58 (1H, ddd, *J* = 12.9, 12.9, 6.8 Hz, H₆); 1.79 (1H, ddd, *J* = 13.0, 9.2, 7.7 Hz, H₁); 2.05 (1H, m, H₁); 2.25 (2H, m, H₈ CH₂CH=CH₂); 2.55 (3H, m, H₂ H₆ CH₂CH=CH₂); 3.38 (3H, s, CH₂OMe); 3.70 (1H, dddd, *J*=11.4, 11.4, 3.9, 3.9 Hz, H_{7ax}); 3.82 (1H, dddd, *J*=11.6, 8.0, 8.0, 3.8 Hz, H_{8a}); 4.69 (2H, s, OCH₂O); 4.95 (1H, d, *J* = 5.4 Hz, H_{5eq}); 5.06 (1H, bd, *J* = 17.1 Hz CH₂=CH); 5.09 (1H, dd, *J* = 7.0, 1.3 Hz, CH₂=CH); 5.8 (1H, dddd, *J* = 17.1, 10.1, 7.0, 7.0 Hz, CH₂=CH). ¹³C NMR (75 MHz, CDCl₃) δ_C 30.4 (C₁); 32.3 (C₆); 35.4 (CH₂-CH=CH₂); 39.4 (C₈); 41.0 (C₂); 50.0 (C₅); 52.3 (C_{8a}); 55.4 (CH₂OMe); 70.8 (C₇); 94.7 (OCH₂O); 117.4 (CH₂=CH); 135.3 (CH₂=CH); 172.3 (COOH); 176.4 (C₃). IR (NaCl disc, cm⁻¹): ν_{max} = 2936, 2368, 1735, 1651, 1451, 1211, 1112, 1039. MS (ES⁺): *m/z* 306 ([MNa]⁺). HRMS (ES⁺, [MNa]⁺): Calcd for C₁₄H₂₁NO₅ 306.1317 Found: 306.1306.

(2S)-2-[(2R*,5S*,7S*,8aR*)-(2-allyl-7-methoxymethoxy-3-oxo-octahydro-indolizin-5-carbonyl)-amino] pentanedioic acid *tert*butyl ester **29**

The same experimental procedure as described for peptide coupling was applied to carboxylic acid **28** (88 mg; 0.31 mmol) using NMM (37 μ L, 0.34 mmol, 1.1 equiv), isobutyl chloroformate (44 μ L, 0.34 mmol, 1.1 equiv). Stirring was maintained 20 h after addition of amino acid (100 mg, 0.34 mmol, 1.1 equiv) and NMM (74 μ L, 0.68 mmol, 2.2 equiv). Flash chromatography on silica gel (AcOEt/cyclohexane: 80/20) allowed the isolation of a colorless oil (90%, 146 mg).

2 diastereoisomers. Rf = 0.6 (AcOEt/cyclohexane: 90/10). ^1H (CDCl₃, 300 MHz): δ_{H} 1.12 (1H, ddd, J = 10.8, 10.8, 10.8 Hz, H_{8ax}); 1.39 (19H, m, 2C(CH₃)₃, H₆); 1.72-1.87 (2H, m, NHCHCH₂CH₂CO, H₁); 1.98-2.07 (1H, m, NHCHCH₂CH₂CO, H₁); 2.1-2.3 (4H, m, NHCHCH₂CH₂CO H₈ CH₂CH=CH₂); 2.4-2.6 (3H, m, H₂ H₆ CH₂CH=CH₂); 3.29 (3H, s, CH₂OMe); 3.66 (1H, m, H_{8a}); 3.82/3.92 (1H, dddd/dddd, J = 11.3, 11.3, 3.9, 3.9 /11.2, 11.2, 3.9, 3.9 Hz, H_{7ax}); 4.33 (1H, m, NHCHCH₂CH₂CO); 4.63 (2H, m, OCH₂O); 4.77/4.80 (1H, d/d., J = 5.7 /6.2 Hz, H_{5eq}); 5.06 (1H, bd, J = 17.1 Hz CH₂=CH); 5.04 (1H, m, CH₂=CH); 5.73 (1H, m, CH₂=CH); 6.66/6.73 (1H, d/d., J = 7.7 /8.0 Hz, NH). ^{13}C NMR (75 MHz, CDCl₃) δ_{C} 27.3/27.4 (2C(CH₃)₃); 30.2 (C₁ NHCHCH₂CH₂CO); 31.2 (C₆); 31.5 (NHCHCH₂CH₂CO); 35.4 (CH₂-CH=CH₂); 39.3 (C₈); 40.4 (C₂); 50.0 50.9 52.2 (C₅, C_{8a}, NHCHCH₂CH₂CO); 55.3 (CH₂OMe); 70.8 (C₇); 80.65 (C(CH₃)₃); 82.1/82.2 (C(CH₃)₃); 95.2 (OCH₂O); 117.3/117.5 (CH₂=CH); 135.0/135.3 (CH₂=CH); 169.6/169.7 170.4/170.6 171.8 (3CO); 176.4/176.4 (C₃). MS (ES⁺): m/z 547 ([MNa]⁺).

(2S)-2-[(2R*,5S*,7S*,8aR*)-2-(3-hydroxy-propyl)-7-methoxymethoxy-3-oxo-octahydro-indolizin-5-carbonyl]-amino] pentanedioic acid *tert*butyl ester **30**

The same experimental procedure as described for hydoboration was applied to **29** (139 mg; 0.27 mmol) using a 0.5 M solution of 9-BBN (1.6 mL; 0.81 mmol; 3 equiv). Stirring was maintained 15 h after addition of sodium acetate 5 M (0.57 mL, 2.84 mmol, 10.5 equiv) and a 30 % H₂O₂ solution (0.33 mL; 12 equiv). Flash chromatography on silica gel (AcOEt/MeOH: 98/2 to 95/5) allowed the isolation of a colorless oil (65%, 93 mg).

Rf = 0.15 (AcOEt/MeOH: 98/2). ^1H (CDCl₃, 300 MHz): δ_{H} 1.18 (1H, ddd, J = 11.2, 11.2, 11.2 Hz, H₈); 1.3-1.72 (23H, m, 2C(CH₃)₃ H₆ CH₂CH₂CH₂OH); 1.78-1.99 (3H, m, NHCHCH₂CH₂CO, H₁); 1.99-2.12 (1H, m, H₁); 2.12-2.3 (3H, m, NHCHCH₂CH₂CO H₈); 2.45-2.64 (2H, m, H₂, H₆); 3.35 (3H, s, CH₂OMe); 3.62-3.78 (3H, m, CH₂OH, H_{8a}); 3.89/3.99 (1H, dddd/dddd, J = 11.2, 11.2, 3.9, 3.9/11.2, 11.2, 4, 4 Hz, H₇); 4.33-4.45 (1H, m, NHCHCH₂CH₂CO); 4.67/4.68 (1H, d/d., J = 6.8/6.8, OCH₂O); 4.72/4.73 (1H, d/d., J = 6.8/6.8, OCH₂O); 4.83/4.86 (1H, d/d., J = 6 /5.3 Hz, H₅); 6.74/6.82 (1H, d/d., J = 8.1 /7.7 Hz, NH). ^{13}C NMR (75 MHz, CDCl₃) δ_{C} 25.7 27.4 27.7 (CH₂); 28.0 (2C(CH₃)₃); 30.1 30.2 31.5 (CH₂); 31.1 (C₆); 39.3/39.5 (C₈); 40.5/40.6 (C₂); 50.9 (C₅); 52.3 (C_{8a} NHCHCH₂CH₂CO); 55.4 (CH₂OMe); 62.3/62.5 (CH₂OH); 70.9 (C₇); 80.8/81.1 82.5 (C(CH₃)₃); 95.2:95.3 (OCH₂O); 169.6/169.7 170.5/171 172/172.2 (3CO); 177.1/177.3 (C₃). IR (NaCl disc, cm⁻¹): ν_{max} = 3313, 2934, 1730, 1536, 1453, 1369, 1257, 1154. MS (ES⁺): m/z 565 ([MNa]⁺). HRMS (ES⁺, [MNa]⁺): Calcd. for C₂₇H₄₆N₂O₉ 565.3101 Found 565.3084.

guanidine carbamate **31**

Alcohol **30** (67 mg, 0.127 mmol) and 1,3-bis(*tert*-butoxycarbonyl) guanidine (70 mg, 0.27 mmol, 2.1 equiv) was dried under vacuo, dissolved in 2.5 mL of anhydrous THF under argon. Tributylphosphine (63 μ L, 0.25 mmol, 2 equiv) was added to the solution at rt, then dipiperidinazodicarboxylate (ADDP) (63 mg, 0.25 mmol, 2 equiv) at 0 °C. The mixture was refluxed 2 h. After addition of water and dichloromethane, the mixture was extracted. The organic phase was dried (MgSO₄) and the solvent removed in vacuo. The residue was purified by flash chromatography on silica gel (Et₂O/MeOH: 97/3) to give a white solid (40 mg, 43%).

Rf = 0.1 (Et₂O/MeOH: 98/2). ^1H (CDCl₃, 400 MHz): δ_{H} 1.18 (1H, ddd, J = 11.7, 11.7, 11.7 Hz, H₈); 1.27-1.48 (28H, m, 3C(CH₃)₃ H₆); 1.5-1.63 (1H, m, CH₂CH₂CH₂OH); 1.73-2.28 (10H, m, H₈, NHCHCH₂CH₂CO CH₂CH₂CH₂OH H₁); 2.43-2.59 (2H, m, H₂ H₆); 3.33 (3H, s, CH₂OMe); 3.64-3.76 (1H, m, H_{8a}); 3.85/3.93 (1H, dddd/dddd, J = 11.2, 11.2, 4.0, 4.0/11.2, 11.2, 4.0, 4.0 Hz, H₇); 4.05-4.15 (2H, m, CH₂OH); 4.32-4.43 (1H, m, NHCHCH₂CH₂CO); 4.65/4.66 (1H, d/d., J = 6.8/6.8 Hz, OCH₂O); 4.69/4.695 (1H, d/d., J = 6.8/6.8, OCH₂O); 4.81/4.84 (1H, d/d., J = 6 /5.8 Hz, H₅); 6.74/6.82 (1H, d/d., J = 8.0 /7.6 Hz, NH). ^{13}C NMR (75 MHz, CDCl₃) δ_{C} 26.5 26.6 27.3 27.4 28.0 (3C(CH₃)₃); 29.6 31.2 31.5 (CH₂); 39.0/39.3 (C₈); 40.3 (C₂); 50.9 (C₅); 52/52.1 52.2/52.4 (C_{8a} NHCHCH₂CH₂CO); 55.3 (CH₂OMe); 64.9 (CH₂OH); 70.9 (C₇); 80.8 82.2 82.3 82.7 (C(CH₃)₃); 95.2 (OCH₂O); 158.7 169.7 169.8 170.7 172.0 (5CO C=N); 176.4/176.8 (C₃). IR (NaCl disc, cm⁻¹): ν_{max} = 3403, 2979, 1730, 1554, 1370, 1256, 1156, 1042, 733. MS (ES⁺): m/z 750 ([MNa]⁺). HRMS (ES⁺, [MNa]⁺): Calcd. for C₃₄H₅₇N₅O₁₂ 750.3901 Found 750.3885.

peptidomimetic **32**

The same protocol for deprotection with TFA was applied to compound **31** (29 mg; 0.04 mmol). Guanidyl compound was obtained as a foam in a quantitative yield (19 mg).

^1H NMR (D₂O, 400 MHz) δ_{H} (ppm): 1.11 (1H, ddd, J = 11.7, 11.7, 11.7 Hz, H₈); 1.4-1.58 (2H, m, H₆, CH₂CH₂CH₂O); 1.58-1.7 (3H, m, CH₂CH₂CH₂O); 1.72-1.9 (2H, m, NHCHCH₂CH₂COOH, H₁); 1.9-2.17 (5H, m, NHCHCH₂CH₂COOH H₈ H₁); 2.27 (1H, m, H₆); 2.53 (1H, m, H₂); 3.78-3.9 (2H, m, H₇ H_{8a}); 4.03 (1H, m, NHCHCH₂CH₂COOH); 4.11 (2H, bs, CH₂CH₂CH₂O); 4.68 (1H, m, H₄). HRMS (ES⁺, [MNa]⁺): Calcd for C₁₉H₂₉N₅O₉ 494.1863 Found 494.1860.

(2S)-2-[[[(2R*,5S*,7S*,8aR*)-2-(3-bis(*tert*-butyloxycarbonyl)-guanidino-propyl)-7-methoxymethoxy-3-oxo-octahydro-indolizin-5-carbonyl]-amino] pentanedioic acid *tert*butyl ester] 33

The same experimental procedure as described for **25** above was applied to **30** (33 mg; 0.062 mmol) using and 1,3-bis(*tert*-butoxycarbonyl) guanidine (39 mg, 0.15 mmol, 2 equiv), triphenylphosphine (40 mg, 0.15 mmol, 2 equiv) and diethylazodicarboxylate (DEAD) (24 μ L, 0.15 mmol, 2 equiv). Stirring was continued for 3 days. Flash chromatography on silica gel (Et₂O/cyclohexane: 70/30 to 100/0) allowed the isolation of **33** as a colorless oil (29 mg, 60%) (2 diastereoisomers).

R_f = 0.45 (Et₂O). ¹H (CDCl₃, 400 MHz): δ_{H} 1.21 (1H, ddd, *J* = 11.7, 11.7, 11.7 Hz, H₈); 1.35-1.6 (37H, m, 4C(CH₃)₃ H₆); 1.62-2.12 (8H, m, CH₂CH₂CH₂NH H₈ NHCHCH₂CH₂CO H₁); 2.15-2.3 (3H, m, NHCHCH₂CH₂CO H₈); 2.47-2.63 (2H, m, H₆ H₂); 3.36 (3H, s, CH₂OMe); 3.7-4.06 (4H, m, H_{8a} CH₂CH₂CH₂NH H₇); 4.09 (1H, m, NHCHCH₂CH₂CO); 4.67 (1H, d, *J* = 6.8 Hz, OCH₂O); 4.71 (1H, d, *J* = 6.8, OCH₂O); 4.74 (1H, d, *J* = 5.4 Hz, H₅); 6.72 (1H, d, *J* = 7.6 Hz, NH); 9.26 (2H, bs, NH₂). ¹³C NMR (75 MHz, CDCl₃) δ_{C} 26.3 26.5 27.4 (CH₂CH₂CH₂NH NHCHCH₂CH₂CO); 28.1 28.3 (4C(CH₃)₃); 30.8 31.5 31.6 (CH₂CH₂CH₂NH NHCHCH₂CH₂CO C₆ C₁); 38.9/39.2 (C₈); 40.1 (C₂); 44.2 (CH₂CH₂CH₂NH); 51 52 52.1 52.3 55.4 (C_{8a} C₅ NHCHCH₂CH₂CO CH₂OMe); 64; 70.9 71.0(C₇); 78.6 80.8 82.3 83.7 (4C(CH₃)₃); 95.2 (OCH₂O); 155.0 160.5 and 163.8 (C=N N-CO); 169.8 170.5 170.7 172.0 176.9 177.2 (4CO C₃). IR (NaCl disc, cm⁻¹): ν_{max} = 3382, 2977, 1715, 1609, 1513, 1368, 1251, 1152, 737. MS (ES⁺): *m/z* 784 ([MH]⁺).

(2S)-2-[[[(2R*,5S*,7S*,8aR*)-2-(3-guanidino-propyl)-7-hydroxy-3-oxo-octahydro-indolizin-5-carbonyl]-amino] pentanedioic acid 34

The same protocol for deprotection with TFA was applied to compound **33** (46 mg; 0.06 mmol). Guanidyl compound was obtained as a white solid in a quantitative yield (33 mg).

mp: 60 °C. ¹H (D₂O, 400 MHz): δ_{H} 1.1 (1H, m, H₈); 1.35-1.53 (5H, m, CH₂CH₂CH₂NH H₆); 1.77-1.98 (3H, m, NHCHCH₂CH₂CO H₈); 1.98-2.15 (2H, m, H₈); 2.21 (1H, m, H₆); 2.31 (2H, bs, NHCHCH₂CH₂CO); 2.52 (1H, bs, H₂); 3.06 (2H, t, *J* = 5.9 Hz, CH₂CH₂CH₂NH); 3.71 (1H, m, H₇); 3.79 (1H, m, H_{8a}); 4.25 (1H, m, NHCHCH₂CH₂CO); 4.63 (1H, m, H₅). ¹³C NMR (100 MHz, D₂O) δ_{C} 25.5 25.6 (CH₂CH₂CH₂NH NHCHCH₂CH₂CO); 27.7 (CH₂CH₂CH₂NH); 30.1 30.3 (NHCHCH₂CH₂CO, C₁); 34.0 (C₆); 39.8 (C₈); 40.8 (C₂); 40.9 (CH₂CH₂CH₂NH); 51.0/51.1 (C₅); 52.4 (NHCHCH₂CH₂CO); 53.7/53.9 (C_{8a}); 64.6 (C₇); 156.7 (C=N); 171.9 172.1 (CONH); 175.1 (NHCHCOOH) 177.1 (COOH); 179.3 (C₃). MS (ES⁺): *m/z* 428 ([MH]⁺). HRMS (ES⁺, [MH]⁺): Calcd for C₁₈H₂₉N₅O₇ 428.2145 Found 428.2146.

(5S*,7R*,8aR*)-7-benzyloxy-5-methyl-3-oxo-octahydro-indolizin-5-carboxylic acid ethyl ester 35

To **12a** (430 mg; 1.42 mmol) dissolved in 90 mL of anhydrous THF under argon at -78 °C, was added 23.3 mL (11.7 mmol, 1.1 equiv) of KHMDS 0.5 M in toluene. After 30 min, methyl iodide (filtered on neutral aluminium oxide; 725 μ L, 11.65 mmol, 1.1 equiv) was added. Stirring was continued for 3h at -78°C, reaction was quenched with a saturated solution of NH₄Cl. The product was extracted twice with CH₂Cl₂. The organic phase was dried (MgSO₄) and the solvent removed in vacuo. Flash chromatography on silica gel (AcOEt/cyclohexane: 50/50) allowed the isolation of **35** as a pale yellow oil (3.5 mg, 99%).

R_f = 0.37 (AcOEt/cyclohexane: 70/30) ¹H NMR (CDCl₃, 300 MHz): δ_{H} 1.12 (3H, t, *J* = 7.1 Hz; CH₃CH₂); 1.36 (1H, ddd, *J* = 13.5, 12.2, 2.6 Hz, H_{8ax}); 1.50 (1H, dddd, *J* = 10.4, 10.4, 10.4, 10.4 Hz, H₁); 1.56 (1H, dd, *J* = 14.4, 2.5 Hz, H_{6ax}); 1.89 (3H, s, Me); 2.12 (2H, m, H₁ H_{8eq}), 2.33 (2H, m, 2H₂); 2.65 (1H, ddd, *J* = 14.4, 3.5, 2.2 Hz, H_{6eq}); 3.77 (1H, dddd, *J* = 2.9, 2.9, 2.9, 2.9 Hz, H_{7eq}); 3.94 (1H, dq, *J* = 10.7, 7.1 Hz, OCH₂CH₃); 3.94 (1H, m, H_{8a}); 4.04 (1H, dq, *J* = 10.7, 7.1 Hz, OCH₂CH₃); 4.40 (1H, d, *J* = 11.9 Hz, CH₂Ph); 4.51 (1H, d, *J* = 11.9 Hz, CH₂Ph); 7.29 (5H, m, Ar). ¹³C NMR (75 MHz, CDCl₃) δ_{C} 14.0 (CH₃CH₂); 26.0 (Me); 26.6 (C₁); 31.9 (C₂); 36.7 (C₈); 40.4 (C₆); 52.1 (C_{8a}); 59.2 (C₅); 61.3 (OCH₂CH₃); 70.1 (CH₂Ph); 70.7 (C₇); 127.2 (2C_{Ar}); 127.5 (C_{Ar}); 128.3 (C_{Ar}); 138.5 (C_{Arquat}); 173.1 177.7 (CO C₃). IR (NaCl disc, cm⁻¹): ν_{max} = 3460, 2938, 1682, 1455, 1271, 1027, 731. HRMS (ES⁺, [MNa]⁺): Calcd for C₁₉H₂₅NO₄ 354.1681 Found 354.1665.

(2R*,5S*,7R*,8aR*)-2-allyl-7-benzyloxy-5-methyl-3-oxo-octahydro-indolizin-5-carboxylic acid ethyl ester 36 and (2S*,5S*,7R*,8aR*)-2-allyl-7-benzyloxy-5-methyl-3-oxo-octahydro-indolizin-5-carboxylic acid ethyl ester 37

The same experimental procedure as described for allylation was applied to **35** (1.5 g; 4.53 mmol) using KHMDS 0.5 M (9.5 mL; 4.75 mmol, 1.01 equiv) in 40 mL of THF. Stirring was maintained 20 min, after cooling to -110 °C, addition of distilled allyl bromide (470 μ L; 5.4 mmol; 1.2 equiv) stirring was continued 1h, then 3h30 at -78 °C. Flash chromatography on silica gel (AcOEt/cyclohexane: 50/50) allowed the isolation of **36** as a pale yellow oil (561 g, 33%) and **37** as a pale yellow oil (366 mg, 22%). Starting material was also recovered (35%).

36: R_f = 0.33 (Et₂O/cyclohexane: 50/50) ¹H NMR (CDCl₃, 300 MHz): δ_{H} 1.13 (3H, t, *J* = 7.1 Hz; CH₃CH₂); 1.20 (1H, q, *J* = 10.9 Hz, H₁); 1.32 (1H, ddd, *J* = 14.6, 12.2, 2.4 Hz, H_{8ax}); 1.56 (1H, dd, *J* = 14.4, 2.4 Hz, H_{6ax}); 1.90 (3H, s, Me); 2.18 (3H, m, CH₂CH=CH₂ H₁ H_{8eq}); 2.48 (1H, m, H₂); 2.66 (1H, ddd, *J* = 14.4, 3.4, 1.8 Hz, H_{6eq}); 2.68 (1H, m, CH₂CH=CH₂); 3.77 (1H, bs, H_{7eq}); 3.88 (1H, m, H_{8a}); 3.93 (1H, dq, *J* = 10.7, 7.1 Hz, OCH₂CH₃); 4.04 (1H, dq, *J* = 10.7, 7.1 Hz, OCH₂CH₃); 4.40 (1H, d, *J* = 11.9 Hz, CH₂Ph); 4.52 (1H, d, *J* = 11.9 Hz, CH₂Ph); 5.04 (1H, bd, *J* = 17 Hz, CH₂CH=CH₂); 5.09 (1H, bd, *J* = 17 Hz, CH₂CH=CH₂); 5.80 (1H, dddd, *J* = 17, 10.1, 6.9, 6.9 Hz, CH₂CH=CH₂);

7.28 (5H, m, Ar). ^{13}C NMR (75 MHz, CDCl_3) δ_{C} 13.9 (CH_3CH_2); 26.2 (Me); 32.8 (C_1); 35.0 ($\text{CH}_2\text{CH}=\text{CH}_2$); 36.5 (C_8); 40.2 (C_6); 42.1 (C_2); 49.9 (C_{8a}); 59.1 (C_5); 61.3 (OCH_2CH_3); 70.0 (CH_2Ph); 70.6 (C_7); 116.4 ($\text{CH}_2\text{CH}=\text{CH}_2$); 127.1 (2C_{Ar}); 127.4 (C_{Ar}); 128.2 (2C_{Ar}); 136.0 ($\text{CH}_2\text{CH}=\text{CH}_2$); 138.5 (C_{Arquat}); 173.1 178.5 (CO C_3). IR (NaCl disc, cm^{-1}): $\nu_{\text{max}} = 2925, 1737, 1698, 1454, 1396, 1307, 1274, 1207, 1130, 1028, 913, 736$. MS (ES^+): m/z 394 ($[\text{MNa}]^+$). Anal. Calcd. for: $\text{C}_{22}\text{H}_{29}\text{NO}_4$ C, 71.13; H, 7.87; N, 3.77. Found: C, 71.52; H, 7.96; N 3.77.

37: $R_f = 0.2$ ($\text{Et}_2\text{O}/\text{cyclohexane}$: 50/50) ^1H NMR (CDCl_3 , 300 MHz): δ_{H} 1.14 (3H, t, $J = 7.2$ Hz; CH_3CH_2); 1.34 (1H, ddd, $J = 13.3, 12.3, 2.5$ Hz, H_{8ax}); 1.54 (1H, dd, $J = 14.4, 2.5$ Hz, H_{6ax}); 1.60 (1H, m, H_1); 1.89 (3H, s, Me); 1.97 (1H, dd, $J = 12.7, 6.0$ Hz, H_1); 2.10 (1H, ddd, $J = 13.4, 13.4, 13.4$ Hz, H_{8eq}); 2.28 (1H, m, $\text{CH}_2\text{CH}=\text{CH}_2$); 2.45 (1H, m, H_2); 2.50 (1H, m, $\text{CH}_2\text{CH}=\text{CH}_2$); 2.64 (1H, ddd, $J = 14.4, 3.3, 2.2$ Hz, H_{6eq}); 3.76 (1H, q, $J = 2.7$ Hz, H_7); 3.95 (1H, dq, $J = 10.8, 7.1$ Hz, OCH_2CH_3); 3.98 (1H, m, H_{8a}); 4.04 (1H, dq, $J = 10.7, 7.1$ Hz, OCH_2CH_3); 4.38 (1H, d, $J = 11.9$ Hz, CH_2Ph); 4.52 (1H, d, $J = 11.9$ Hz, CH_2Ph); 5.06 (1H, bd, $J = 10.1$ Hz, $\text{CH}_2\text{CH}=\text{CH}_2$); 5.09 (1H, dd, $J = 17.1, 1.4$ Hz, $\text{CH}_2\text{CH}=\text{CH}_2$); 5.85 (1H, dddd, $J = 17, 10.1, 7.8, 6.0$ Hz, $\text{CH}_2\text{CH}=\text{CH}_2$); 7.28 (5H, m, Ar). ^{13}C NMR (75 MHz, CDCl_3) δ_{C} 14.0 (CH_3CH_2); 25.9 (Me); 30.7 (C_1); 34.7 ($\text{CH}_2\text{CH}=\text{CH}_2$); 36.8 (C_8); 40.2 (C_6); 41.9 (C_2); 50.0 (C_{8a}); 59.0 (C_5); 61.3 (OCH_2CH_3); 70.1 (CH_2Ph); 70.6 (C_7); 116.7 ($\text{CH}_2\text{CH}=\text{CH}_2$); 127.2 (2C_{Ar}); 127.5 (C_{Ar}); 128.3 (2C_{Ar}); 136.4 ($\text{CH}_2\text{CH}=\text{CH}_2$); 138.3 (C_{Arquat}); 173.0 178.8 (CO C_3). IR (NaCl disc, cm^{-1}): $\nu_{\text{max}} = 2937, 1736, 1694, 1446, 1396, 1276, 1208, 1132, 1067, 912, 729$. MS (ES^+): m/z 394 ($[\text{MNa}]^+$).

(2R*,5S*,7R*,8aR*)-7-benzyloxy-2-(3-bis(tert-butyloxycarbonyl)guanidino-propyl)-5-methyl-3-oxo-octahydro-indolizin-5-carboxylic acid ethyl ester 38

-*trans* intermediate alcohol:

The same experimental procedure as described for hydoboration was applied to alkene **36** (309 mg; 0.83 mmol) using a 0.5 M solution of 9-BBN in THF (5 mL; 2.49 mmol; 3 equiv). Stirring was maintained 16 h after addition of sodium acetate 5 M (2 mL, 9.9 mmol, 12 equiv) and a 30 % H_2O_2 solution (1 mL; 12 equiv). Flash chromatography on silica gel ($\text{AcOEt}/\text{cyclohexane}$: 90/10) allowed the isolation of a yellow oil (60%, 193 mg).

$R_f = 0.24$ ($\text{AcOEt}/\text{cyclohexane}$: 95/5) ^1H NMR (CDCl_3 , 300 MHz): δ_{H} 1.09 (3H, t, $J = 7.1$ Hz; CH_3CH_2); 1.17 (1H, ddd, $J = 11.4, 11.4, 11.4$ Hz, H_1); 1.31 (1H, ddd, $J = 13.9, 11.4, 2.4$ Hz, H_{8ax}); 1.44 (1H, m, $\text{CH}_2\text{CH}_2\text{CH}_2\text{OH}$); 1.54 (1H, dd, $J = 14.3, 2.3$ Hz, H_6); 1.6 (2H, m, $\text{CH}_2\text{CH}_2\text{CH}_2\text{OH}$); 1.78-1.95 (4H, s, Me $\text{CH}_2\text{CH}_2\text{CH}_2\text{OH}$); 2.14 (1H, dm, $J = 11.4$ Hz, H_8); 2.25 (1H, ddd, $J = 11.5, 7.9, 5.4$ Hz, H_1); 2.38 (1H, m, H_2); 2.62 (1H, bd, $J = 14.4$ Hz, H_6); 3.6 (2H, t, $J = 6.3$ Hz, $\text{CH}_2\text{CH}_2\text{CH}_2\text{OH}$); 3.75 (1H, m, H_7); 3.85 (1H, m, H_{8a}); 3.93 (1H, dq, $J = 10.7, 7.1$ Hz, OCH_2CH_3); 3.97 (1H, dq, $J = 10.7, 7.1$ Hz, OCH_2CH_3); 4.38 (1H, d, $J = 11.9$ Hz, CH_2Ph); 4.49 (1H, d, $J = 11.9$ Hz, CH_2Ph); 7.29 (5H, m, Ar). ^{13}C NMR (75 MHz, CDCl_3) δ_{C} 13.9 (CH_3CH_2); 26.1 (Me); 26.6 ($\text{CH}_2\text{CH}_2\text{CH}_2\text{OH}$); 30.1 ($\text{CH}_2\text{CH}_2\text{CH}_2\text{OH}$); 33.3 (C_1); 36.4 (C_8); 40.2 (C_6); 42.2 (C_2); 50.0 (C_{8a}); 59.1 (C_5); 61.3 (OCH_2CH_3); 62.1 ($\text{CH}_2\text{CH}_2\text{CH}_2\text{OH}$); 70.0 (CH_2Ph); 70.5 (C_7); 127.1 (2C_{Ar}); 127.4 (C_{Ar}); 128.2 (2C_{Ar}); 138.3 (C_{Arquat}); 173.0 179.7 (CO C_3). IR (NaCl disc, cm^{-1}): $\nu_{\text{max}} = 3412, 2928, 1734, 1681, 1399, 1307, 1273, 1210, 1131, 735$. MS (ES^+): m/z 412 ($[\text{MNa}]^+$).

The same experimental procedure as described for **25** was applied to intermediate *trans* (300 mg; 0.77 mmol) using and 1,3-bis(tert-butoxycarbonyl) guanidine (311 mg, 1.2 mmol, 1.5 equiv), triphenylphosphine (315 mg, 1.2 mmol, 1.5 equiv) and diethylazodicarboxylate (DEAD) (190 μL , 1.2 mmol, 1.5 equiv). Stirring was continued for 24 h. Flash chromatography on silica gel ($\text{Et}_2\text{O}/\text{cyclohexane}$: 50/50) allowed the isolation of **38** as a white solid (436 mg, 90%).

$R_f = 0.27$ ($\text{Et}_2\text{O}/\text{cyclohexane}$: 50/50). mp: 132 $^\circ\text{C}$ ^1H NMR (CDCl_3 , 300 MHz): δ_{H} 1.04 (3H, t, $J = 7.1$ Hz; CH_3CH_2); 1.07 (1H, m, H_1); 1.25 (2H, m, $\text{CH}_2\text{CH}_2\text{CH}_2\text{N}$ H_8); 1.4-1.55 (19H, m, $2\text{C}(\text{CH}_3)_3$ H_6); 1.58 (2H, m, $\text{CH}_2\text{CH}_2\text{CH}_2\text{N}$); 1.81 (3H, s, Me); 1.85 (1H, m, $\text{CH}_2\text{CH}_2\text{CH}_2\text{N}$); 2.08 (1H, bd, $J = 12.5$ Hz, H_8); 2.34 (2H, m, H_2 , H_1); 2.59 (1H, bd, $J = 14.2$ Hz, H_6); 3.62-3.75 (2H, m, H_7 , $\text{CH}_2\text{CH}_2\text{CH}_2\text{N}$); 3.75-3.89 (2H, H_{8a} , OCH_2CH_3); 3.89-4.0 (2H, m, $\text{CH}_2\text{CH}_2\text{CH}_2\text{N}$ OCH_2CH_3); 4.32 (1H, d, $J = 11.9$ Hz, CH_2Ph); 4.45 (1H, d, $J = 11.9$ Hz, CH_2Ph); 7.21 (5H, m, Ar); 9.22 (2H, m, NH_2). ^{13}C NMR (75 MHz, CDCl_3) δ_{C} 13.9 (CH_3CH_2); 26.1 (Me); 27.6 ($\text{CH}_2\text{CH}_2\text{CH}_2\text{N}$); 28.0 and 28.3 ($\text{CH}_2\text{CH}_2\text{CH}_2\text{N}$, $2\text{C}(\text{CH}_3)_3$); 33.1 (C_1); 36.5 (C_8); 40.3 (C_6); 42.0 (C_2); 44.3 ($\text{CH}_2\text{CH}_2\text{CH}_2\text{N}$); 49.9 (C_{8a}); 59.0 (C_5); 61.2 (OCH_2CH_3); 69.9 (CH_2Ph); 70.5 (C_7); 78.4 ($\text{C}(\text{CH}_3)_3$); 83.6 ($\text{C}(\text{CH}_3)_3$); 127.0 (2C_{Ar}); 127.3 (C_{Ar}); 128.2 (2C_{Ar}); 138.3 (C_{Arquat}); 155.0; 160.5; 163.8 (C=N, NCOO); 173.0 179.1 (CO C_3). IR (NaCl disc, cm^{-1}): $\nu_{\text{max}} = 3390, 2881, 1714, 1608, 1271, 1150$. MS (ES^+): m/z 653 ($[\text{MNa}]^+$).

(2S*,4R*,6R*)-6-[(2R*)-2-carboxy-5-guanidino-pentyl]-4-hydroxy-2-methyl-piperidin-2-carboxylic acid 39

Compound **38** (70 mg, 0.11 mmol) was dissolved in 5 mL of HCl 3 M. The mixture was refluxed for 20 h. The solvent was evaporated under vacuo. To the residue were added 1 mL of diethyl ether and 2 mL of water, insoluble solids were filtered and the product was extracted 3 times with 2 mL of water. The aqueous phase was lyophilysed to leave an orange oil in a 100% yield (38 mg).

^1H NMR (D_2O , 400 MHz): δ_{H} 1.3-1.65 (8H, m, CH_2CHCOOH H_5 Me, $\text{CH}_2\text{CH}_2\text{CH}_2\text{N}$); 1.67 (1H, bd, $J = 15.0$ Hz, H_3); 1.75-1.95 (3H, m, $\text{CH}_2\text{CH}_2\text{CH}_2\text{N}$ CH_2CHCOOH H_5); 2.3-2.5 (2H, m, H_3 CH_2CHCOOH); 3.0 (2H, bs, $\text{CH}_2\text{CH}_2\text{CH}_2\text{N}$); 3.67 (1H, m, H_6); 4.05 (1H, bs, H_4). ^{13}C NMR (75 MHz, D_2O) δ_{C} 24.4 (Me); 25.4 ($\text{CH}_2\text{CH}_2\text{CH}_2\text{N}$); 28.7 ($\text{CH}_2\text{CH}_2\text{CH}_2\text{N}$); 33.2 (C_5); 34.4 (CH_2CHCOOH); 37.8 (C_3); 40.7 (CH_2CHCOOH $\text{CH}_2\text{CH}_2\text{CH}_2\text{N}$); 47.5 (C_6); 59.2 (C_2); 61.7 (C_4); 156.6 (C=N); 173.8 178.6 (CO C_3). MS (ES^+): m/z 313 ($[\text{M}-\text{H}_2\text{O}]^+$), 331 ($[\text{M}-\text{H}]^+$). HRMS (ES^+ , $[\text{MH}]^+$): Calcd for $\text{C}_{14}\text{H}_{26}\text{N}_4\text{O}_5$ 331.1981 Found 331.1965.

(2S*,5S*,7R*,8aR*)-7-benzyloxy-2-(3-bis(*tert*-butyloxycarbonyl)guanidino-propyl-5-methyl- -3-oxo-octahydro-indolizin-5-carboxylic acid ethyl ester 40

-*cis* intermediate alcohol:

The same experimental procedure as described for hydroboration was applied to alkene **37** (178 mg; 0.48 mmol) using a 0.5 M solution of 9-BBN in THF (2 mL; 1 mmol; 2.1 equiv). Stirring was maintained 16 h after addition of sodium acetate 5 M (1.15 mL, 5.76 mmol, 12 equiv) and a 30 % H₂O₂ solution (0.6 mL; 5.76 mmol, 12 equiv). Flash chromatography on silica gel (AcOEt/Et₂O: 90/10) allowed the isolation of a yellow oil (87%, 162 mg).

R_f = 0.18 (AcOEt/Et₂O: 90/10) ¹H NMR (CDCl₃, 300 MHz): δ_H 1.15 (3H, t, *J* = 7.1 Hz, CH₃CH₂); 1.34 (1H, ddd, *J* = 13.6, 11.2, 2.4 Hz, H₈); 1.57 (1H, dd, *J* = 14.4, 2.5 Hz, H₆); 1.61-1.96 (9H, m, Me CH₂CH₂CH₂OH H₁); 2.13 (1H, dm, *J* = 13.5 Hz, H₈); 2.28 (1H, bs, OH); 2.46 (1H, m, H₂); 2.67 (1H, ddd, *J* = 14.4, 3.4, 2.1 Hz, H₆); 3.7 (2H, m, CH₂CH₂CH₂OH); 3.78 (1H, m, H₇); 3.93-4.09 (3H, m, H_{8a} OCH₂CH₃); 4.41 (1H, d, *J* = 11.9 Hz, CH₂Ph); 4.53 (1H, d, *J* = 11.9 Hz, CH₂Ph); 7.3 (5H, m, Ar). ¹³C NMR (100 MHz, CDCl₃) δ_C 14.1 (CH₃CH₂); 26.0 (Me); 27.2 (CH₂CH₂CH₂OH); 30.5 (CH₂CH₂CH₂OH); 32.4 (C₁); 34.7 (CH₂CH=CH₂); 36.9 (C₈); 40.3 (C₆); 41.9 (C₂); 50.3 (C_{8a}); 59.1 (C₅); 61.5 (OCH₂CH₃); 62.6 (CH₂CH₂CH₂OH); 70.2 (CH₂Ph); 70.6 (C₇); 127.3 (2C_{Ar}); 127.6 (C_{Ar}); 128.4 (2C_{Ar}); 138.4 (C_{Arquat}); 173.1 179.6 (CO C₃). IR (NaCl disc, cm⁻¹): ν_{max} = 3389, 2952, 1651, 1454, 1286, 1066, 910, 728. MS (ES⁺): *m/z* 412 ([MNa]⁺).

The same experimental procedure as described for **25** was applied to intermediate *cis* (24 mg; 0.062 mmol) using and 1,3-bis(*tert*-butoxycarbonyl) guanidine (26 mg, 0.1 mmol, 1.6 equiv), triphenylphosphine (26 mg, 0.1 mmol, 1.6 equiv) and diethylazodicarboxylate (DEAD) (16 μL, 0.1 mmol, 1.6 equiv). Stirring was continued for 44 h. Flash chromatography on silica gel (Et₂O/cyclohexane: 70/30) allowed the isolation of **40** as a white solid (40 mg, 100%).

R_f = 0.31 (AcOEt/Et₂O: 70/30). mp > 60 °C ¹H NMR (CDCl₃, 300 MHz): δ_H 1.13 (3H, t, *J* = 7.2 Hz; CH₃CH₂); 1.31 (1H, m, H₈); 1.41-1.62 (20H, m, CH₂CH₂CH₂N 2C(CH₃)₃ H₆); 1.63-1.62 (4H, m, CH₂CH₂CH₂N H₁); 1.95 (4H, m, Me, H₁); 2.12 (1H, bd, *J* = 13.4 Hz, H₈); 2.4 (1H, m, H₂); 2.67 (1H, bd, H₆); 3.78 (1H, bs, H₇); 3.82-4.09 (5H, m, CH₂CH₂CH₂N H_{8a} OCH₂CH₃); 4.39 (1H, d, *J* = 12 Hz, CH₂Ph); 4.52 (1H, d, *J* = 12 Hz, CH₂Ph); 7.3 (5H, m, Ar); 9.23 (2H, bs, NH₂). ¹³C NMR (75 MHz, CDCl₃) δ_C 14.0 (CH₃CH₂); 25.9 (Me); 26.8 and 27.3 (CH₂CH₂CH₂N, CH₂CH₂CH₂N); 28.0 and 28.3 (2C(CH₃)₃); 31.6 (C₁); 36.9 (C₈); 40.2 (C₆); 42.0 (C₂); 44.5 (CH₂CH₂CH₂N); 50.1 (C_{8a}); 59.0 (C₅); 61.3 (OCH₂CH₃); 70.0 (CH₂Ph); 70.6 (C₇); 78.6 (C(CH₃)₃); 83.7 (C(CH₃)₃); 127.2 (2C_{Ar}); 127.5 (C_{Ar}); 128.3 (2C_{Ar}); 138.3 (C_{Arquat}); 155.1, 156.8, 163.9 (C=N, NCOO); 173.0 179.2 (CO C₃). IR (NaCl disc, cm⁻¹): ν_{max} = 3382, 2930, 1714, 1614, 1454, 1359, 1255, 1152. MS (ES⁺): *m/z* 631 ([MNa]⁺).

(2S*,4R*,6R*)-6-[(2S*)-2-carboxy-5-guanidino-pentyl]-4-hydroxy-2-methyl-piperidin-2-carboxylic acid 41

The same experimental procedure as described for **39** was applied to **40** (67 mg; 0.1 mmol) in 5 mL of HCl 3 M. The aqueous phase was lyophilised to leave brown oil in a 100% yield (44 mg).

¹H NMR (D₂O, 400 MHz): δ_H; 1.45 (3H, s, Me); 1.46-1.6 (5H, m, CH₂CHCOOH H₅ CH₂CH₂CH₂N); 1.75 (1H, bd, *J* = 14.4 Hz, H₃); 1.78-1.93 (3H, m, CH₂CH₂CH₂N CH₂CHCOOH H₅); 2.48 (1H, bd, , *J* = 15.1 Hz, H₃); 2.58 (1H, q, *J* = 6.4 Hz, CH₂CHCOOH); 3.09 (2H, t, *J* = 6.5 Hz, CH₂CH₂CH₂N); 3.75 (1H, m, H₆); 4.09 (1H, bs, H₄). ¹³C NMR (75 MHz, D₂O) δ_C 24.5 (Me); 25.2 (CH₂CH₂CH₂N); 28.2 (CH₂CH₂CH₂N); 33.5 (C₅); 34.1 (CH₂CHCOOH); 37.7 (C₃); 40.7 (CH₂CH₂CH₂N); 40.9 ((CH₂CHCOOH)); 47.8 (C₆); 59.4 (C₂); 61.8 (C₄); 156.6 (C=N); 174.1 179.1 (CO C₃). MS (ES⁺): *m/z* 331 ([M-H]⁺). HRMS (ES⁺, [MH]⁺): Calcd for C₁₄H₂₆N₄O₅ 331.1981 Found 331.1965.

Cell culture and cell adhesion assay

S180 and Eahy926 cells were cultured in DMEM+10% FCS and DMEM+20%FCS+HAT, respectively. For adhesion assays, cells were harvested with cell dissociation non-enzymatic solution (SIGMA). Cells were pelleted by centrifugation and resuspended at a density of 1x10⁵ cells/ml in PBS/CaMg. Assays of cell adhesion to coated substrates were performed on non-tissue culture treated 96-well plates,. The wells were incubated with FN or VN at 10 μg.ml⁻¹ in PBS overnight at 4 °C followed by a 30-min incubation with 3 mg.ml⁻¹ bovine serum albumin in PBS (previously heat-inactivated for 3 min at 80 °C). The substrates were thoroughly washed and maintained in PBS until use. The peptidomimetics solutions were prepared either at 20 μM, 400 μM or 1 mM in PBSCaMg. 50 μl of peptidomimetic solution were added to the well with 50 μl of the cell suspension reaching the final peptidomimetic solution to 10 μM, 200 μM or 500 μM), for the assay. Each test was done in triplicate. The plates were incubated at 37 °C for 1 h, washed with PBS to remove the non adherent cells. Then, the wells were fixed in 5% glutaraldehyde in PBS, washed, stained with 1% crystal violet in 200 mM MES and washed in 200 mM MES buffer, pH 6. The crystal violet fixed to the cells was dissolved in 100 μL of 10% acetic acid and OD was read with a microplate reader at 570 nm. Data are the mean±S.D of triplicates and are expressed as the % of cell adhesion obtained on FN in the absence of competitor. The positive and negative controls for the inhibition of cell adhesion were done using the cyclic c(RGDfV) and GRGES peptides, respectively. Two independent experiments were done for each competitor and for each cell type.

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References and notes

1. (a) Hynesw, R. O *Cell*, **1992**, *69*, 11-25; Gottschalk, K. E.; Kessler, H. *Angew. Chem. Int. Ed.* **2002**, *41*, 3767-3774; (b) Benneth, J. S. *J. Clin. Invest.* **2005**, *115*, 3363-3369.
2. (a) Brooks, P.C. *Eur. J. Cancer* **1996**, *32A*, 2423-2429; (b) Friedlander, M.; Brooks, P. C.; Shaffer, R. W.; Kincaid, C. M.; Varner, J. A.; Cheresch, D. A. *Science* **1995**, *270*, 1500-1502; (c) Brooks, P.C.; Montgomery, A.M.P.; Rosenfeld, M.; Reisfeld, R.A.; Hu, T.; Klier, G.; Cherech, D.A. *Cell* **1994**, *79*, 1157-1164; (d) Enns, A.; Korb, T.; Schlüter, K.; Gassmann, P.; Spiegel, H.-U.; Senninger, N.; Mitjans, F.; Haier, J. *Eur. J. Cancer* **2005**, *41*, 1065-1072.
3. (a) Ruoslahti, E. *Annu. Rev. Cell. Dev. Biol.* **1996**, *12*, 697-715; (b) Marinelli, L.; Lavecchia, A.; Gottschalk, K.-E.; Novellino, E.; Kessler, H. *J. Med. Chem.* **2003**, *46*, 4393-4404; (c) Moitessier, N.; Henry, C.; Maigret, B.; Chapleur, Y. *J. Med. Chem.* **2004**, *47*, 4178-4187.
4. Kumar, S; Wang, Q.; Sasaki, A. *Tetrahedron* **2007**, *63*, 2084-2090 and references cited herein.
5. (a) Dechantstreiter, M.A.; Planker, E.; Mathä, B.; Lohof, E.; Hölzemann, G.; Jonczyk, A.; Goodman, S.L.; Kessler, H. *J. Med. Chem.* **1999**, *42*, 3033-3040; (b) Uman, S.; Gaus, K.; Yang, Y.; Strijowski, U.; Sewald, N.; De Pol, S.; Reiser, O. *Angew. Chem. Int. Ed.* **2007**, *46*, 3976-3978; (c) Macdonald, T.; Stewart, C. F.; Kocak, M.; Goldman, S.; Ellenbogen, R. G.; Phillips, P.; Lafond, D.; Poussaint, T. Y.; Kieran, M. W.; Boyet, J. M.; Kun, L. E. *J. Clin. Oncol.* **2008**, *26*, 919-924; (d) Zanardi, F.; Burreddu, P.; Rassu, P.; Auzzas, G.; Battistini, L.; Curti, L.; Sartori, C.; Nicastro, A.; Menchi, G.; Cini, G.; Bottonocetti, N.; Raspanti, A.; Casiraghi, G. *J. Med. Chem.* **2008**, *51*, 1771-1782.
6. (a) Belvisi, L.; Colombo, L.; Manzoni, L.; Potenza, D.; Scolastico, C. *Synlett* **2004**, 1449-1471; (b) Maison, W.; Prenzel, A. H. G. P. *Synthesis* **2005**, 1031-1048; (c) Manzoni, L.; Bassanini, M.; Belvisi, L.; Motto, I.; Scolastico, C.; Castorina, M.; Pisano, C. *Eur. J. Org. Chem.* **2007**, 1309-1317.
7. Halie, D.; Pérard-Viret, J.; Royer, J. *Heterocycles* **2006**, *68*, 2471-2476.
8. Baussanne, I.; Dudot, B.; Perard-Viret, J.; Planas, L.; Royer, J. *ARKIVOC* **2006**, 57-66.
9. (a) Osterkamp, F.; Ziemer, B.; Koert, U.; Wiesner, M.; Raddatz, P.; Goodman, S.L. *Chem. Eur. J.* **2000**, *6*, 666-683; (b) Chen, Z.; Kende, A.S.; Colson, A.O.; Mendezandino, J.L.; Ebetino, F.H.; Bush, R.D.; Hu, X.E. *Synth. Commun.* **2006**, *36*, 473-479; (c) Osterkamp, F.; Wehlan, H.; Koert, U.; Wiesner, M.; Raddatz, P.; Goodman, S.L. *Tetrahedron* **1999**, *55*, 10713-10734.
10. Tsunoda, T.; Yamamiya, Y.; Itô, S. *Tetrahedron Lett.* **1993**, *34*, 1639-1642.
11. Robinson, D.E.; Seth, P.P.; Jefferson, E.A. *Synth. Commun.* **2004**, *34*, 2743-2749. This paper described the same reaction. But, we do not agree with the proposed mechanism. We suggest a simple transesterification type reaction since we obtained the same product in the absence of ADDP and Bu₃P.
12. Beauvais, A.; Erickson, A.; Goins, T.; Craig, S.E.; Suzan, E.; Humphries, J. P.; Dufour, S. *J. Cell. Biol.* **1995**, *128*, 699-713.