



**HAL**  
open science

## **Pan genome of the phytoplankton *Emiliana underpins* its global distribution**

Betsy A Read, Jessica Kegel, Mary J Klute, Alan Kuo, Stephane C Lefebvre,  
Florian Maumus, Christoph Mayer, John Miller, Adam Monier, Asaf Salamov,  
et al.

► **To cite this version:**

Betsy A Read, Jessica Kegel, Mary J Klute, Alan Kuo, Stephane C Lefebvre, et al.. Pan genome of the phytoplankton *Emiliana underpins* its global distribution. *Nature*, 2013, 499 (7457), pp.209-213. 10.1038/nature12221 . hal-01074841

**HAL Id: hal-01074841**

**<https://hal.science/hal-01074841>**

Submitted on 15 Oct 2014

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial - ShareAlike 4.0 International License

# Pan genome of the phytoplankton *Emiliana* underpins its global distribution

Betsy A. Read<sup>1</sup>, Jessica Kegel<sup>2</sup>, Mary J. Klute<sup>3</sup>, Alan Kuo<sup>4</sup>, Stephane C. Lefebvre<sup>5</sup>, Florian Maumus<sup>6</sup>, Christoph Mayer<sup>7,8</sup>, John Miller<sup>9</sup>, Adam Monier<sup>10</sup>, Asaf Salamov<sup>4</sup>, Jeremy Young<sup>11</sup>, Maria Aguilar<sup>3</sup>, Jean-Michel Claverie<sup>12</sup>, Stephan Frickenhaus<sup>2,13</sup>, Karina Gonzalez<sup>14</sup>, Emily K. Herman<sup>3</sup>, Yao-Cheng Lin<sup>15</sup>, Jonathan Napier<sup>16</sup>, Hiroyuki Ogata<sup>12</sup>, Analissa F. Sarno<sup>1</sup>, Jeremy Shmutz<sup>4,17</sup>, Declan Schroeder<sup>18</sup>, Colomban de Vargas<sup>19</sup>, Frederic Verret<sup>20</sup>, Peter von Dassow<sup>21</sup>, Klaus Valentin<sup>2</sup>, Yves Van de Peer<sup>15</sup>, Glen Wheeler<sup>18,22</sup>, *Emiliana huxleyi* Annotation Consortium†, Joel B. Dacks<sup>3\*</sup>, Charles F. Delwiche<sup>9\*</sup>, Sonya T. Dyhrman<sup>23,24\*</sup>, Gernot Glöckner<sup>25\*</sup>, Uwe John<sup>2\*</sup>, Thomas Richards<sup>26\*</sup>, Alexandra Z. Worden<sup>10\*</sup>, Xiaoyu Zhang<sup>27\*</sup> & Igor V. Grigoriev<sup>4</sup>

**Coccolithophores have influenced the global climate for over 200 million years<sup>1</sup>. These marine phytoplankton can account for 20 per cent of total carbon fixation in some systems<sup>2</sup>. They form blooms that can occupy hundreds of thousands of square kilometres and are distinguished by their elegantly sculpted calcium carbonate exoskeletons (coccoliths), rendering them visible from space<sup>3</sup>. Although coccolithophores export carbon in the form of organic matter and calcite to the sea floor, they also release CO<sub>2</sub> in the calcification process. Hence, they have a complex influence on the carbon cycle, driving either CO<sub>2</sub> production or uptake, sequestration and export to the deep ocean<sup>4</sup>. Here we report the first haptophyte reference genome, from the coccolithophore *Emiliana huxleyi* strain CCMP1516, and sequences from 13 additional isolates. Our analyses reveal a pan genome (core genes plus genes distributed variably between strains) probably supported by an atypical complement of repetitive sequence in the genome. Comparisons across strains demonstrate that *E. huxleyi*, which has long been considered a single species, harbours extensive genome variability reflected in different metabolic repertoires. Genome variability within this species complex seems to underpin its capacity both to thrive in habitats ranging from the equator to the subarctic and to form large-scale episodic blooms under a wide variety of environmental conditions.**

Fundamental uncertainties exist regarding the physiology and ecology of *E. huxleyi*, and the relationships between different morphotypes (Fig. 1a). To investigate its gene repertoire and physiological capacity, we sequenced the diploid genome of CCMP1516 using the Sanger shotgun approach. The haploid genome is estimated to be 141.7 megabases (Mb) and 97% complete on the basis of conserved eukaryotic single-copy genes<sup>5,6</sup> (Supplementary Table 1, Supplementary Data 7 and Supplementary Information 1.1–1.4). It is dominated by repetitive elements, constituting >64% of the sequence, much greater than seen for sequenced diatoms (Fig. 2 and Supplementary Information 2.10). Of the 30,569 protein-coding genes predicted—93% of which have transcriptomic support (expressed sequence tag or RNA-seq)

(Supplementary Information 1.5–1.7, 2.1–2.2 and Supplementary Data 1–3)—we identified expansions in gene families specific to iron/macromolecular transport, post-translational modification, cytoskeletal development and signal transduction relative to other sequenced eukaryotic algae (Supplementary Information 2.3).

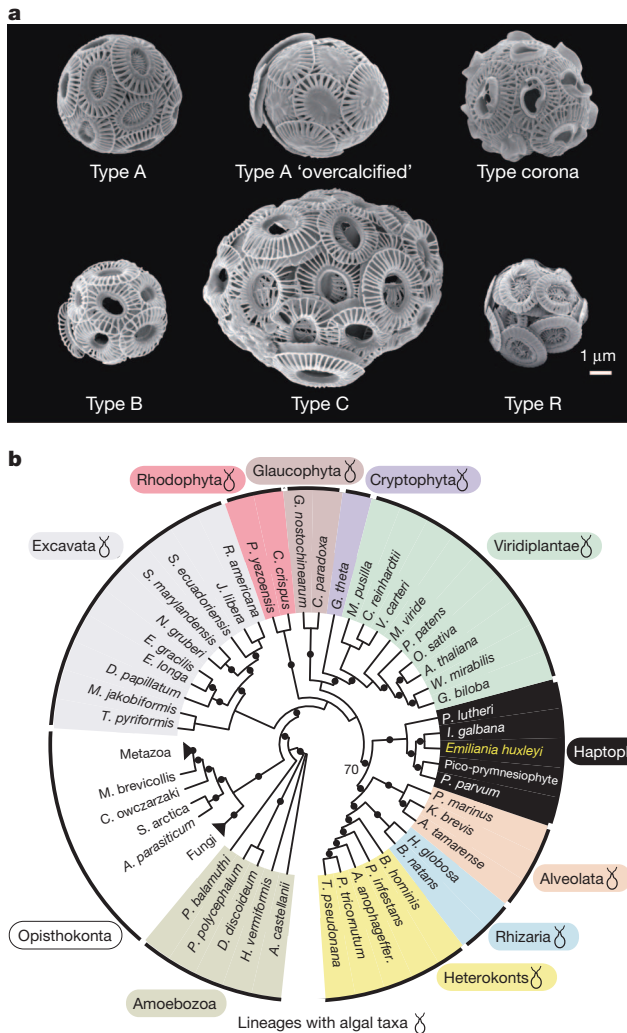
The *E. huxleyi* genome provides a crucial reference point for evolutionary, cellular and physiological studies because haptophytes represent a distinct branch on the eukaryotic tree of life (Fig. 1b). Consistent with other published analyses<sup>7</sup>, conserved marker genes demonstrate the haptophytes branch as a sister clade to heterokonts, alveolates and rhizarians. However, as a lineage possessing secondary plastids, the evolutionary history of haptophyte genomes may be more complex<sup>8</sup> than that suggested by a single concatenated analysis. Thus, individual gene phylogenies were constructed using clusters of orthologous proteins (1,563) identified by comparative analysis of *E. huxleyi* and at least 9 of 48 taxa sampled from across eukaryotes (Supplementary Information 2.4). *E. huxleyi* was monophyletic, with heterokonts in 28–33% of the resolved trees and the green lineage (green algae and plants) in 11–14%. Less frequent relationships were also observed, presumably reflecting a mosaic genome<sup>8</sup> with contributions from the host lineage, the eukaryotic endosymbiont, and possibly horizontal gene transfer (Supplementary Fig. 1 and Supplementary Data 4).

Coccolithophores produce the anti-stress osmolyte dimethylsulphoniopropionate (DMSP), which can be demethylated to produce methylmercaptopropionate and/or cleaved by some organisms, such as *E. huxleyi*, to produce the predominant natural source of atmospheric sulphur, dimethylsulphide. Although the gene encoding the DmdA protein, which catalyses the initial demethylation of DMSP, was not detected in the genome, genes that produce sulphur and carbon intermediates and function in later stages of DMSP degradation were identified<sup>9</sup>. Also present is an intron-containing, but otherwise bacterial *dddD*-like, gene encoding an acetyl-coenzyme A (acetyl-CoA) transferase proposed to add CoA to DMSP before cleavage<sup>9</sup> (Supplementary Table 2). These data will facilitate molecular approaches

<sup>1</sup>Department of Biological Sciences, California State University San Marcos, San Marcos, California 92096, USA. <sup>2</sup>Alfred Wegener Institute Helmholtz Center for Polar and Marine Research (AWI), 27570 Bremerhaven, Germany. <sup>3</sup>Department of Cell Biology, University of Alberta, Edmonton, Alberta T6G 2H7, Canada. <sup>4</sup>US Department of Energy Joint Genome Institute, Walnut Creek, California 94598, USA. <sup>5</sup>J. Craig Venter Institute, San Diego, California 92121, USA. <sup>6</sup>Institut National de la Recherche Agronomique, Unité de Recherche en Génomique-Info, Versailles 78026, France. <sup>7</sup>Forschungsmuseum Alexander Koenig, 53113 Bonn, Germany. <sup>8</sup>Department of Animal Ecology, Evolution and Biodiversity, Ruhr-University, D-44801 Bochum, Germany. <sup>9</sup>Cell Biology and Molecular Genetics and the Maryland Agricultural Experiment Station, University of Maryland, College Park, Maryland 20742, USA. <sup>10</sup>Monterey Bay Aquarium Research Institute, Moss Landing, California 95039, USA. <sup>11</sup>Department of Earth Sciences, University College London, Gower Street, London WC1E 6BT, UK. <sup>12</sup>Structural and Genomic Information Laboratory, CNRS, Aix-Marseille University, Mediterranean Institute of Microbiology, Marseille FR3479, France. <sup>13</sup>Biotechnology, Hochschule Bremerhaven, An der Karlstadt 8, 27568 Bremerhaven, Germany. <sup>14</sup>Department of Cell Biology, Harvard Medical School, Boston, Massachusetts 02115, USA. <sup>15</sup>Department of Plant Systems Biology, VIB, Ghent University, 9052 Ghent, Belgium. <sup>16</sup>Department of Biological Chemistry, Rothamsted Research, Harpenden AL5 2JQ, UK. <sup>17</sup>HudsonAlpha Genome Sequencing Center, Huntsville, Alabama 35806, USA. <sup>18</sup>Marine Biological Association of the UK, Plymouth PL12PB, UK. <sup>19</sup>CNRS UMR 7144 and Université Pierre et Marie Curie, EPEP team, Station Biologique de Roscoff, 29682 Roscoff Cedex, France. <sup>20</sup>School of Biological Sciences, University of Essex, Colchester CO4 3SQ, UK. <sup>21</sup>Departamento de Ecología, Facultad de Ciencias Biológicas, Pontificia Universidad Católica de Chile, Santiago, Chile. <sup>22</sup>Plymouth Marine Laboratory, Prospect Place, The Hoe, Plymouth PL1 3DH, UK. <sup>23</sup>Biology Department, Woods Hole Oceanographic Institution, Woods Hole, Massachusetts 02543, USA. <sup>24</sup>Department of Earth and Environmental Sciences and Lamont-Doherty Earth Observatory, Columbia University, Palisades, New York 10964, USA. <sup>25</sup>Institute for Biochemistry I, Medical Faculty, University of Cologne, D-50931, Germany and Leibniz-Institute of Freshwater Ecology and Inland Fisheries, D-12587 Berlin, Germany. <sup>26</sup>Department of Zoology, Natural History Museum, London SW7 5BD, UK. <sup>27</sup>Department of Computer Science and Information Systems, California State University San Marcos, California 92096, USA.

†A list of participants and their affiliations appears at the end of the paper.

\*These authors contributed equally to this work.

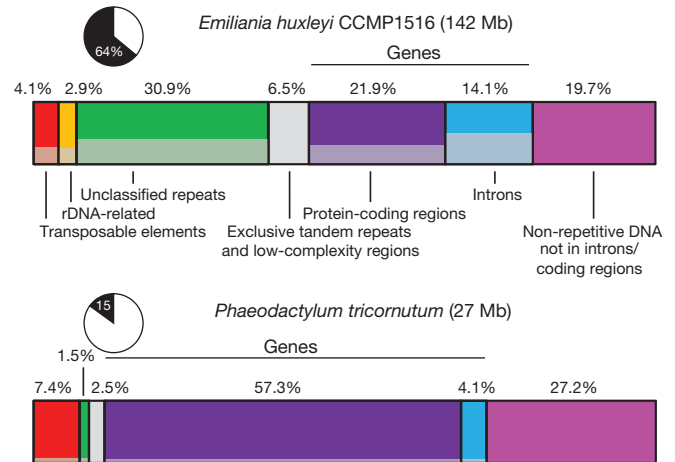


**Figure 1 | *Emiliana huxleyi* and its position in the eukaryotic tree of life.** **a.** *E. huxleyi* has five well-characterized calcification morphotypes and an overcalcified state<sup>1</sup>. **b.** Cladogram showing the distinct branch occupied by the haptophyte lineage on the basis of RAXML analysis of concatenated, nuclear-encoded proteins after addition of homologues from CCMP1516 and a pico-prymnesiophyte-targeted metagenome<sup>8</sup>. Lineages with algal taxa are indicated (symbol). Filled circles represent nodes with  $\geq 70\%$  bootstrap support. The tree is rooted for display purposes only.

for probing DMSP biogeochemistry and the environmental importance of sulphur production and biotransformations.

*E. huxleyi* synthesizes unusual lipids that are used as nutritional/feedstock supplements, polymer precursors and petrochemical replacements. Two functionally redundant pathways for the synthesis of omega-3 polyunsaturated eicosapentaenoic and docosahexaenoic fatty acids were partially characterized<sup>10</sup> (Supplementary Table 3). Pathway analysis indicates that *E. huxleyi* sphingolipids are primarily glucosylceramides, often with an unusual C9 methyl branch (Supplementary Table 3) found only in fungi and some animals<sup>11</sup>. Genes for two zinc-containing quinone reductases, involved in reduction of alkenone  $\alpha, \beta$ -double bonds used in paleotemperature reconstructions and proposed biofuels, were also identified<sup>12,13</sup>.

Coccoliths have precise nanoscale architecture and unique light-scattering properties of interest to material and optoelectronic scientists. Carbonic anhydrase is associated with biomineralization in other organisms<sup>14</sup> and accelerates bicarbonate formation. The 15 *E. huxleyi* carbonic anhydrase isozymes and genes involved in calcium and carbon transport,  $H^+$  efflux, cytoskeleton organization and polysaccharide modulation (Supplementary Table 4) represent targets for resolving



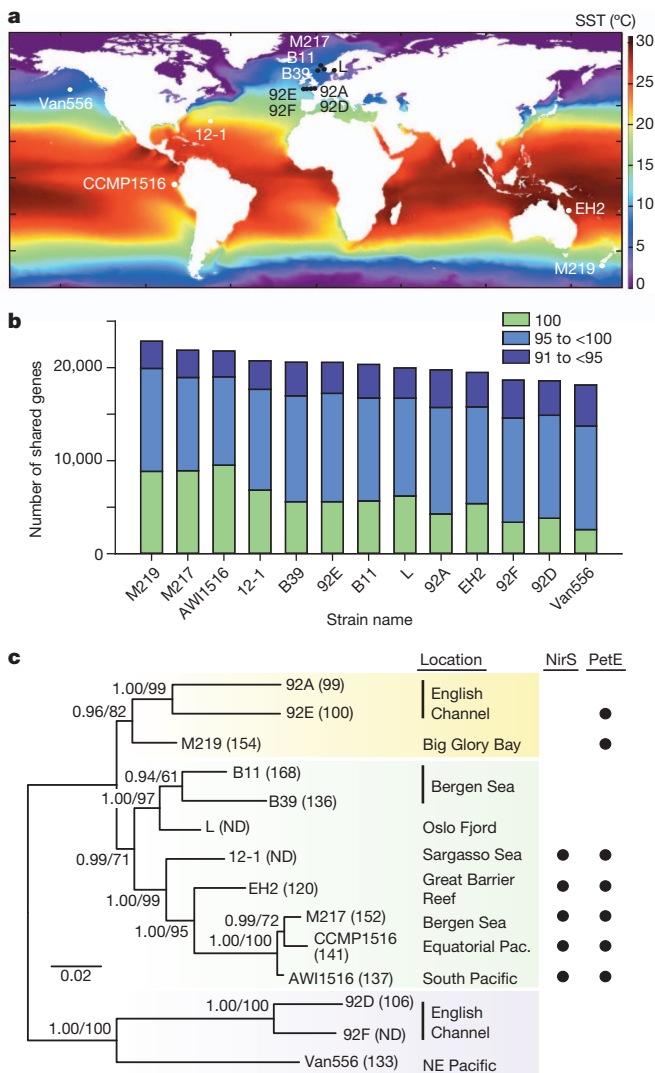
**Figure 2 | Relative composition of the *E. huxleyi* genome.** Structural composition of genomes from CCMP1516 and the diatom *P. tricornutum*. Grey-shaded regions of each class depict proportions of tandem repeats and low-complexity regions. The grey vertical box contains only tandem repeats and low-complexity sequence. Pie charts indicate the proportion of non-repeated (white) and repeated or low-complexity (black) sequences in each haploid genome.

molecular mechanisms governing coccolith formation, and will aid in predicting response patterns to anthropogenic  $CO_2$  increases and ocean acidification.

The global distribution of *E. huxleyi* (for example, Fig. 3a, c) and its capacity for bloom formation under different physiochemical parameters are puzzling. To investigate the potential influence of genome variation in this ecological dynamic, three *E. huxleyi* isolates (92A, EH2 and Van556) from different oceanic regions were deeply sequenced (265–352-fold coverage) (Fig. 3a, c, Supplementary Tables 5–7 and Supplementary Information 2.6). Two approaches were used to compare genomes. First, sequence reads were assembled and contigs aligned to the CCMP1516 reference genome using Standard Nucleotide BLAST (BLASTn; Supplementary Information 2.6.1). Although these isolates show  $>98\%$  18S ribosomal RNA (rRNA) identity, only 54–77% of their contigs showed similarity to CCMP1516. 71 Mb of the remaining contigs were shared between at least two deeply sequenced strains. 8–40 Mb appeared to be isolate specific, as did 27 Mb of CCMP1516. Flow cytometric genome-size estimates also showed heterogeneity across isolates, with haploid genome sizes ranging from 99 to 133 Mb (Supplementary Information 2.5, 2.6.1 and Supplementary Table 5). These findings indicated considerable intraspecific variation.

To examine potential variations in gene content further, sequence reads were directly mapped to the CCMP1516 genome. Of the 30,569 predicted genes in CCMP1516, between 1,373 and 2,012 different genes were not found in 92A, Van556 and EH2 (cumulatively 5,218, or 17% of CCMP1516 genes), and 364 appeared to be missing from all three. These findings cannot be explained by poor coverage or sequencing bias alone. Of 458 highly conserved eukaryotic genes from the CEGMA set<sup>5</sup>, 95–97% were identified in the isolates, indicating nearly complete genome sequences (Supplementary Data 7). Together, *de novo* assemblies and direct mapping to CCMP1516 indicate that the pan genome of *E. huxleyi* represents a rapidly changing repository of genetic information with genomic fluidity estimated to be  $\geq 10\%$ <sup>15</sup> (on the basis of CCMP1516 gene content).

*E. huxleyi* isolate differences were assessed further by Illumina sequencing of ten additional strains. Although sequenced at lower coverage, these strains were estimated to be 91–95% complete (Supplementary Tables 6, 7 and Supplementary Data 7). Direct mapping of reads from the 13 strains to CCMP1516 revealed a ‘core genome’ containing about two-thirds of the genes predicted in the reference genome (Supplementary Information 2.6.2 and Supplementary Data 5), a core



**Figure 3 | Predicted proteome comparisons and concatenated phylogeny of *E. huxleyi* strains.** **a**, Isolation locations shown over the averaged Reynolds monthly sea-surface temperature (SST) climatology (1985–2007). **b**, tBLASTn homology search results using predicted CCMP1516 proteins against assemblies from other strains. Bars are coloured according to the number of gene products and nucleotide per cent identity. **c**, Best Bayesian topology, where node values indicate posterior probability/maximum-likelihood bootstrap support. Haploid genome sizes (in Mb) are provided in brackets (with ND indicating not determined), and shaded boxes denote robust clades of geographically dispersed strains. The variable distribution of nitrite reductase (NirS) and plastocyanin (PetE) is shown.

independently confirmed by comparative DNA microarrays (Supplementary Information 2.7, Supplementary Data 6 and Supplementary Fig. 2). Nearly 25% of CCMP1516 genes were not found in at least three other strains, indicating that *E. huxleyi* represents a species complex with a genetic repertoire much greater than that of any one strain (Supplementary Figs 3, 4). Although the most extensive gene-sequence divergence was observed between CCMP1516 and deeply sequenced isolates Van556, 92A and EH2, concatenated phylogenies define three well-supported clades that are not necessarily reflective of geographic distributions (Fig. 3b, c and Supplementary Information 2.61, 2.8).

We searched the CCMP1516 genome for evidence of molecular mechanisms contributing to genome plasticity. There was limited evidence for horizontal gene transfers (Supplementary Information 2.9 and Supplementary Table 8), and although diverse, the complement of transposable elements was also small (Fig. 2 and Supplementary Information 2.10.2). However, *E. huxleyi* has a high density of unclassified

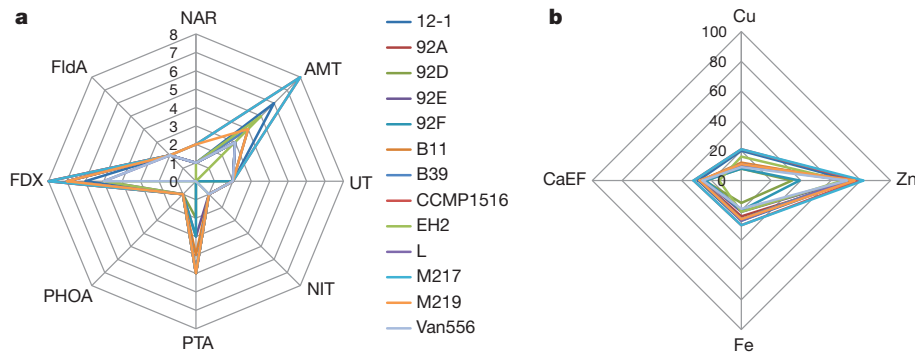
repeats (~31%) and tandem repeats/low-complexity regions (~34%) with tandem-repeat/low-complexity density highest in introns (Fig. 2, Supplementary Information 2.10.1 and Supplementary Table 9). Most protein-coding genes contain multiple introns, often with noncanonical GC donor sites (Supplementary Fig. 5). The preference for 10–11-base-pair repeats in introns and their strong strandedness (meaning that on the sense and antisense strand either the motif or its reverse complement is highly favoured) raises the possibility that intronic tandem repeats have a functional role in exon swapping (Supplementary Information 2.10.3–2.10.5 and Supplementary Table 9).

*E. huxleyi* blooms under many different oceanographic regimes. We explored how the core genome and variable components in different ecotypes might influence success (Supplementary Information 2.11 and Supplementary Fig. 6). The remarkable capacity of *E. huxleyi* to withstand photoinhibition<sup>16</sup> lies in the core genome, which encodes a variety of photoreceptors; proteins that function in the assembly and repair of photosystem II, such as D1-specific proteases and FtsH enzymes; and proteins that have a role in non-photochemical quenching (NPQ) or synthesis of NPQ compounds (Supplementary Table 10). Genes encoding reactive oxygen species (ROS) scavenging antioxidants, enzymes for synthesis of vitamin B<sub>6</sub> constituents used during photo-oxidative stress in plants<sup>17</sup> (Supplementary Tables 10, 15) and many light-harvesting complex (LHC) proteins are also in the core. Of the 68 LHCs, 17 belong to LI818 or LHCZ classes with photoprotective capabilities<sup>18</sup> (Supplementary Table 11 and Supplementary Information 3.1). The complex repertoire of photoprotectors facilitates tolerance to high light by minimizing ROS accumulation and preventing oxidative damage.

Phosphorus and nitrogen are key determinants of oceanic primary production. A suite of core genes allows *E. huxleyi* to thrive in low phosphorus conditions. This includes six inorganic phosphate transporters (Fig. 4), a high-efficiency alkaline phosphatase (Fig. 4)<sup>19</sup>, purple acid phosphatases and other enzymes used to hydrolyse and acquire organic phosphorus compounds<sup>20</sup>. Genes for the synthesis of betaine and sulpholipids used as replacements for cellular phospholipids<sup>21</sup> are also present (Supplementary Table 12). Numbers of phosphate transporters and alkaline phosphatases, (Fig. 4) however, vary considerably from strain to strain, supporting previous observations of differences in phosphorus uptake and hydrolysis kinetics<sup>22</sup>.

Genes for inorganic nitrogen uptake and assimilation (nitrate, nitrite and ammonium) and for acquisition and degradation of nitrogen-rich compounds (for example, urea) (Fig. 4 and Supplementary Table 13) are present in the core genome and may explain the broad range of nitrogen concentrations in which *E. huxleyi* blooms<sup>23</sup>. Although present in multiple copies, the number of genes encoding nitrite (4), nitrate (8) and urea (3) transporters was relatively small compared to ammonium transporters (20). This enrichment, and the varied distribution across strains (Fig. 4), may be indicative of strain-specific ammonium preference, or the need for tightly regulated transporters to mediate high-affinity ammonium/ammonia uptake while offering ammonium-toxicity protection. Surprisingly, core iron-containing (*nirK*) versus clade-restricted copper-containing (*nirS*) nitrite reductases were identified (Fig. 3), although iron is often more limiting than copper in oceanic environments.

*E. huxleyi* grows well in surface waters where iron levels are generally low (0.02–1 nM)<sup>24</sup>. The core genome indicates that iron is acquired using the natural resistance-associated macrophage protein (NRAMP) class of metal transporters, multicopper oxidases, surface-bound ferric reductases, and possibly, membrane-bound siderophores (Supplementary Data 8). Genes involved in mechanisms limiting iron requirements are also in the core, including manganese and copper/zinc superoxide dismutases, both zinc and iron alcohol dehydrogenases and rubredoxins, and copper- and haem- plastocyanins (PetE) and ascorbate oxidases. Selective recruitment of these enzymes as well as flavodoxin, a functional analogue of ferredoxin, may reduce iron demands<sup>25</sup>. *E. huxleyi* encodes many iron-binding proteins,



**Figure 4 | Distribution of genes in the variable genome reflecting niche specificity.** **a**, Key genes (gene numbers on axes) involved in nutrient acquisition and metabolism, including ammonium transporters (AMT), urea transporters (UT), nitrilase (NIT), phosphate transporters (PTA), alkaline

phosphatase (PHOA), ferredoxin (FDX), flavodoxin (FldA) and nitrate reductase (NAR) (Supplementary Information 3.2). **b**, Genes encoding calcium EF hand (CaEF) proteins and others that bind metals such as copper, zinc and iron (Supplementary Information 3.2).

80 in the core and 30 linked to the variable genome (Fig. 4). Iron limitation is linked to reduced calcification and photosynthesis<sup>26</sup>, and our analysis suggests cellular demands and mechanisms to alleviate iron deprivation differ between strains and are probably important factors shaping *E. huxleyi* ecological dynamics.

The *E. huxleyi* pan genome encodes nearly 700 proteins whose structure and function is dependent upon metal binding (Supplementary Data 8). Selenium is essential for growth<sup>27</sup> and potentially incorporated into at least 49 proteins (20 gene families) present in nearly all strains (Supplementary Table 14). Zinc affects growth and nitrogen usage<sup>26</sup>, and is a cofactor of more than 400 proteins, many present in the variable genome (Fig. 4). Heterogeneity in zinc-binding proteins across strains may explain variations in zinc quotas between cultured isolates<sup>26,28</sup>.

In addition to metals, *E. huxleyi* relies on a range of vitamins. Genes for *de novo* synthesis of antioxidants such as pro-vitamin A, vitamins C, E, B<sub>6</sub> and B<sub>9</sub> and the ultraviolet-light-absorbing vitamin D are uniformly present across strains. *E. huxleyi*, however, is ostensibly unable to inhabit ocean regions where vitamins B<sub>1</sub> and B<sub>12</sub> are inaccessible. ThiC, a key B<sub>1</sub> biosynthesis enzyme, was not found in the genome, and despite relying exclusively on a vitamin-B<sub>12</sub>-dependent methionine synthase, genes for a B<sub>12</sub> transporter and several enzymes required for B<sub>12</sub> synthesis are also absent (Supplementary Table 15).

*E. huxleyi* is the dominant bloom-forming coccolithophore and can be abundant in oligotrophic oceans, directly influencing global carbon cycling. Distributions in modern oceans and those dating back to the Pleistocene era demonstrate its tremendous capacity for adaptation. Until now, the underlying mechanisms for the physiological and morphological variations between isolates have been elusive. Evidence presented here indicates that this capacity can be explained, in part, by its pan genome, the first of its kind reported for what was thought to be a single microbial eukaryotic algal species. Variations in gene complements (Fig. 4) within this species complex may drive phenotypic variation, ecological dynamics and the physiological heterogeneity observed in past studies. The high level of diversity indicates that a single strain is unlikely to be typical—or representative—of all strains. Future sequencing of phytoplankton isolates will reveal whether this discovery is a unique or more common feature in microalgae. Together, the physiological capacity and genomic plasticity of *E. huxleyi* make it a powerful model for the study of speciation and adaptations to global climate change.

## METHODS SUMMARY

The diploid genome of CCMP1516 (isolated from the Equatorial Pacific (02.6667S 82.7167W)) was Sanger sequenced and assembled using the Arachne assembler. Gene models were predicted and validated using computational tools, experimental data (including transcriptomics; Sanger and Illumina sequenced) and NimbleGen tiling array experiments. Thirteen additional strains were sequenced

using Illumina and mapped to the reference genome. A detailed description of materials and methods is in Supplementary Information.

Received 18 June 2012; accepted 25 April 2013.

Published online 12 June 2013.

- Paasche, E. A review of the coccolithophorid *Emiliania huxleyi* (Prymnesiophyceae), with particular reference to growth, coccolith formation, and calcification-photosynthesis interactions. *Phycologia* **40**, 503–529 (2001).
- Poulton, A. J., Adey, T. R., Balch, W. M. & Holligan, P. M. Relating coccolithophore calcification rates to phytoplankton community dynamics: regional differences and implications for carbon export. *Deep-Sea Res.* **II** **54**, 538–557 (2007).
- Holligan, P. M., Viollier, M., Harbour, D. S., Campus, P. & Champagne-Philippe, M. Satellite and ship studies of coccolithophore production along a continental shelf edge. *Nature* **304**, 339–342 (1983).
- Rost, B. & Riebesell, U. in *Coccolithophores: From Molecular Processes to Global Impact* (eds Thierstein, H. R. & Young, J. R.) 99–125 (Springer, 2004).
- Parra, G., Bradnam, K., Ning, Z., Keane, T. & Korf, I. Assessing the gene space in draft genomes. *Nucleic Acids Res.* **37**, 289–297 (2009).
- Colbourne, J. K. *et al.* The ecoresponsive genome of *Daphnia pulex*. *Science* **331**, 555–561 (2011).
- Burki, F., Okamoto, N., Pombert, J. F. & Keeling, P. J. The evolutionary history of haptophytes and cryptophytes: phylogenomic evidence for separate origins. *Proc. R. Soc. B* **279**, 2246–2254 (2012).
- Cuvelier, M. L. *et al.* Targeted metagenomics and ecology of globally important uncultured eukaryotic phytoplankton. *Proc. Natl Acad. Sci. USA* **107**, 14679–14684 (2010).
- Todd, J. D. *et al.* Structural and regulatory genes required to make the gas dimethyl sulfide in bacteria. *Science* **315**, 666–669 (2007).
- Sayanova, O. *et al.* Identification and functional characterisation of genes encoding the omega-3 polyunsaturated biosynthetic pathway from the coccolithophore *Emiliania huxleyi*. *Phytochemistry* **72**, 594–600 (2011).
- Oura, T. & Kajiwara, S. *Candida albicans* sphingolipid C9-methyltransferase involved in hyphal elongation. *Microbiology* **156**, 1234–1243 (2010).
- Conte, M. N., Eglinton, G. & Madureira, L. A. S. Long-chain alkenones and alkyl alkenoates as palaeotemperature indicators: their production, flux, and early sedimentary diagenesis in the Eastern North Atlantic. *Advances in Organic Chemistry* **19**, 287–298 (1992).
- Wu, Q., Shiraiwa, Y., Takeda, H., Sheng, G. & Fu, J. Liquid-saturated hydrocarbons resulting from pyrolysis of the marine Coccolithophores *Emiliania huxleyi* and *Gephyrocapsa oceanica*. *Mar. Biotechnol.* **1**, 346–352 (1999).
- Väänänen, H. K. & Parvinen, E. K. in *The Carbonic Anhydrases* (eds Tashian, R. E., Dodgson, S. J., Gros, G. & Carter, N. D.) Ch. 32, 351–356 (Springer, 1991).
- Kislyuk, A. O., Haegeman, B., Bergman, H. & Weitz, J. S. Genomic fluidity: an integrative view of gene diversity within microbial populations. *BMC Genomics* **12**, 32 (2011).
- Nanninga, H. J. & Tyrrell, T. Importance of light for the formation of algal blooms by *Emiliania huxleyi*. *Mar. Ecol. Prog. Ser.* **136**, 195–203 (1996).
- Havaux, M. *et al.* Vitamin B6 deficient plants display increased sensitivity to high light and photo-oxidative stress. *BMC Plant Biol.* **9**, 130 (2009).
- Zhu, S. H. & Green, B. R. Photoprotection in the diatom *Thalassiosira pseudonana*: role of L1818-like proteins in response to high light stress. *Biochim. Biophys. Acta* **1797**, 1449–1457 (2010).
- Xu, Y., Wahlund, T. M., Feng, L., Shaked, Y. & Morel, F. M. M. A novel alkaline phosphatase in the coccolithophore *Emiliania huxleyi* (Prymnesiophyceae) and its regulation by phosphorus. *J. Phycol.* **42**, 835–844 (2006).
- Karl, D. M. & Björkman, K. M. *Dynamics of DOP in Biogeochemistry of Marine Dissolved Organic Matter* (eds Hansell, D. A. & Carlson, C. A.) Ch. 6, 249–348 (Elsevier Science, 2002).

21. Van Mooy, B. A. *et al.* Phytoplankton in the ocean use non-phosphorus lipids in response to phosphorus scarcity. *Nature* **458**, 69–72 (2009).
22. Reid, E. L. *et al.* Coccolithophores: functional biodiversity, enzymes and bioprospecting. *Mar. Drugs* **9**, 586–602 (2011).
23. Lessard, E. J., Merico, A. & Tyrrell, T. Nitrate:phosphate ratios and *Emiliania huxleyi* blooms. *Limnol. Oceanogr.* **50**, 1020–1024 (2005).
24. Turner, D. R., Hunter, K. A. & de Baar, H. J. W. *The Biogeochemistry of Iron in Seawater* Vol. 7, Ch. 1, 1–7 (John Wiley & Sons, 2001).
25. Erdner, D. L. & Anderson, D. M. Ferredoxin and flavodoxin as biochemical indicators of iron limitation during open-ocean iron enrichment. *Limnol. Oceanogr.* **44**, 1609–1615 (1999).
26. Schulz, K. G. *et al.* Effect of trace metal availability on coccolithophorid calcification. *Nature* **430**, 673–676 (2004).
27. Danbara, A. & Shiraiwa, Y. The requirement of selenium for the growth of marine coccolithophorids, *Emiliania huxleyi*, *Gephyrocapsa oceanica* and *Helladosphaera* sp. (Prymnesiophyceae). *Plant Cell Physiol.* **40**, 762–766 (1999).
28. Sunda, W. G. & Huntsman, S. A. Feedback interactions between zinc and phytoplankton in seawater. *Limnol. Oceanogr.* **37**, 25–40 (1992).

**Supplementary Information** is available in the online version of the paper.

**Acknowledgements** Joint Genome Institute (JGI) contributions were supported by the Office of Science of the US Department of Energy (DOE) under contract no. 7DE-AC02-05CH11231. We thank A. Gough for assistance with figures, C. Gentemann for Fig. 3 ocean colour analysis and P. Keeling for discussions.

**Author Contributions** Genome sequencing was performed by the US DOE JGI. B.A.R. coordinated the project and I.V.G. coordinated JGI sequencing/analysis; J.S. performed assemblies; A.K. and A.S. conducted automated annotation and analysis; U.J. at the AWI performed Illumina sequencing of 13 additional strains; A.K., X.Z., U.J., G.G., F.M., C.d.V., S.F., C.M., H.O., F.V., D.S., S.C.L., A.M., J.-M.C., Y.-C.L., Y.V.d.P., J.K., K.V., K.G., A.F.S., J.N., P.v.D. and G.W. performed genome and transcriptome analyses; U.J. and G.G. provided Illumina genomic sequence data, F.V. and D.S., tiling array data, and J. K., microarray data; J.Y. provided SEM images; phylogenetic analyses was contributed by A.M., and A.Z.W. (Fig. 1b); E.K.H., M.J.K. and J.B.D. (Fig. 3c); J.M., C.F.D., M.A. U.J., and J.B.D. (Supplementary Fig. 1); B.A.R. wrote the manuscript in collaboration with J.B.D., C.F.D., S.T.D., G.G., U.J., T.R., A.Z.W., X.Z. and I.V.G. (co-second senior authors). Authors in the first alphabetical list of the paper are equally contributing second authors who made substantial contributions to the paper. The remaining authors are members of the *E. huxleyi* Annotation Consortium who contributed additional analyses and/or annotations.

**Author Information** This paper is distributed under the terms of the Creative Commons Attribution-Non-Commercial-Share Alike licence, and the online version of this paper is freely available to all readers. Assembly and annotation data for *E. huxleyi* strain 1516 are available through JGI Genome Portal at <http://jgi.doe.gov/Ehux> and at DDBJ/EMBL/GenBank under accession number AHAL00000000. The version described in this paper is the first version, AHAL01000000. Sequence information for other strains can be found at the Sequence Read Archive (<http://www.ncbi.nlm.nih.gov/sra>) under the accession number SRA048733.2. Reprints and permissions information is available at [www.nature.com/reprints](http://www.nature.com/reprints). The authors declare no competing financial interests. Readers are welcome to comment on the online version of the paper. Correspondence and requests for materials should be addressed to B.A.R. ([bread@csusm.edu](mailto:bread@csusm.edu)).



This work is licensed under a Creative Commons Attribution-NonCommercial-Share Alike 3.0 Unported licence. To view a copy of this licence, visit <http://creativecommons.org/licenses/by-nc-sa/3.0>

### *Emiliania huxleyi* Annotation Consortium

Andrew E. Allen<sup>1</sup>, Kay Bidle<sup>2</sup>, Mark Borodovsky<sup>3,4</sup>, Chris Bowler<sup>5</sup>, Colin Brownlee<sup>6</sup>, J. Mark Cock<sup>7,8</sup>, Marek Elias<sup>9</sup>, Vadim N. Gladyshev<sup>10</sup>, Marco Groth<sup>11</sup>, Chittibabu Guda<sup>12</sup>, Ahmad Hadaegh<sup>13</sup>, Maria Debora Iglesias-Rodriguez<sup>14,15</sup>, Jerry Jenkins<sup>16</sup>, Bethan M. Jones<sup>15,17</sup>, Tracy Lawson<sup>18</sup>, Florian Leese<sup>19</sup>, Erika Lindquist<sup>20</sup>, Alexei Lobanov<sup>1</sup>, Alexandre Lomsadze<sup>3</sup>, Shehre-Banoo Malik<sup>21</sup>, Mary E. Marsh<sup>22</sup>, Luke Mackinder<sup>6</sup>, Thomas Mock<sup>23</sup>, Bernd Mueller-Roeber<sup>24</sup>, António Pagarete<sup>25</sup>, Micaela Parker<sup>26</sup>, Ian Probert<sup>27</sup>, Hadi Quesneville<sup>28</sup>, Christine Raines<sup>18</sup>, Stefan A. Rensing<sup>29,30</sup>, Diego Mauricio Riaño-Pachón<sup>31</sup>, Sophie Richier<sup>15,32,33</sup>, Sebastian Rokitta<sup>34</sup>, Yoshihiro Shiraiwa<sup>35</sup>, Darren M. Soanes<sup>36</sup>, Mark van der Giezen<sup>36</sup>, Thomas M. Wahlund<sup>37</sup>, Bryony Williams<sup>36</sup>, Willie Wilson<sup>38</sup>, Gordon Wolfe<sup>39</sup> & Louie L. Wurch<sup>40,41</sup>

<sup>1</sup>J. Craig Venter Institute, San Diego, California 92121, USA. <sup>2</sup>Environmental Biophysics and Molecular Ecology Group, Institute of Marine and Coastal Sciences, Rutgers University, New Brunswick, New Jersey 08901, USA. <sup>3</sup>Joint Georgia Tech and Emory Department of Biomedical Engineering, School of Computational Science and Engineering, Georgia Tech, Atlanta, Georgia 30322, USA. <sup>4</sup>Department of Bioinformatics, Moscow Institute for Physics and Technology, Moscow 117303, Russia. <sup>5</sup>Environmental and Evolutionary Genomics Section, Institut de Biologie de l'Ecole Normale Supérieure, Centre National de la Recherche Scientifique, Unité Mixte de Recherche 8197, Institut National de la Santé et de la Recherche Médicale U1024, Ecole Normale Supérieure, 75230 Paris Cedex 05, France. <sup>6</sup>Marine Biological Association of the UK, Plymouth PL12PB, UK. <sup>7</sup>CNRS, UMR 7139, Laboratoire International Associé Dispersal and Adaptation in Marine Species, Station Biologique de Roscoff, Place Georges Teissier, BP74, 29682 Roscoff Cedex, France. <sup>8</sup>UPMC Université Paris 06, The Marine Plants and Biomolecules Laboratory, UMR 7139, Station Biologique de Roscoff, Place Georges Teissier, BP74, 29682 Roscoff Cedex, France. <sup>9</sup>University of Ostrava, Faculty of Science, Department of Biology and Ecology, Life Science Research Centre, 710 00 Ostrava, Czech Republic. <sup>10</sup>Division of Genetics, Department of Medicine, Brigham and Women's Hospital and Harvard Medical School, Boston, Massachusetts 02115, USA. <sup>11</sup>Leibniz Institute for Age Research - Fritz Lipmann Institute, Beutenbergstraße 11, 07745 Jena, Germany. <sup>12</sup>Department of Genetics, Cell Biology & Anatomy, Bioinformatics and Systems Biology Core, University of Nebraska Medical Center, Omaha, Nebraska 68198, USA. <sup>13</sup>Department of Computer Science and Information Systems, California State University San Marcos, San Marcos, California 92096, USA. <sup>14</sup>Department of Ecology, Evolution and Marine Biology, University of California Santa Barbara, Santa Barbara, California 93106, USA. <sup>15</sup>Ocean and Earth Science, National Oceanography Centre Southampton, University of Southampton, Southampton SO17 1BJ, UK. <sup>16</sup>HudsonAlpha Genome Sequencing Center, Huntsville, Alabama 35806, USA. <sup>17</sup>Department of Microbiology, Oregon State University, Corvallis, Oregon 97331, USA. <sup>18</sup>School of Biological Sciences, University of Essex, Colchester CO4 3SQ, UK. <sup>19</sup>Department of Animal Ecology, Evolution and Biodiversity, Ruhr-University D-44801 Bochum, Germany. <sup>20</sup>US Department of Energy Joint Genome Institute, Walnut Creek, California 94598, USA. <sup>21</sup>Canadian Institute for Advanced Research Program in Integrated Microbial Biodiversity, Dalhousie University, Halifax, Nova Scotia B3H 4R2, Canada. <sup>22</sup>University of Texas-Houston Medical School, Houston, Texas 77030, USA. <sup>23</sup>School of Environmental Sciences, University of East Anglia, Norwich Research Park, Norwich NR47TJ, UK. <sup>24</sup>University of Potsdam, Institute of Biochemistry and Biology, Karl-Liebknecht-Straße 24-25, Haus 20, 14476 Potsdam-Golm, Germany. <sup>25</sup>Department of Biology, University of Bergen, Thormøhlensgate 53 A & B, N-5006 Bergen, Norway. <sup>26</sup>Center for Environmental Genomics, PNW Center for Human Health and Ocean Studies, University of Washington, Seattle, Washington 98195-7940, USA. <sup>27</sup>CNRS UMR 7144 and Université Pierre et Marie Curie, EPEP team, Station Biologique de Roscoff, 29682 Roscoff Cedex, France. <sup>28</sup>Institut National de la Recherche Agronomique, Unité de Recherche en Génomique-Info, Versailles 78026, France. <sup>29</sup>Faculty of Biology and BIOS Centre for Biological Signalling Studies, University of Freiburg, Friedrichstrasse 39, 79098 Freiburg, Germany. <sup>30</sup>Faculty of Biology, University of Marburg, Karl-von-Frisch-Strasse 8, 35043 Marburg, Germany. <sup>31</sup>Departamento de Ciencias Biológicas, Universidad de los Andes, Bogotá Distrito Capital, 111711, Colombia. <sup>32</sup>INSU CNRS, Lab Oceanography Villefranche, UMR7093, F-06234 Villefranche Sur Mer, France. <sup>33</sup>Université Paris 06, Observatoire Océanologique Villefranche, F-06230 Villefranche Sur Mer, France. <sup>34</sup>Alfred Wegener Institute Helmholtz Center for Polar and Marine Research (AWI), 27570 Bremerhaven, Germany. <sup>35</sup>Faculty of Life and Environmental Sciences, University of Tsukuba, 1-1-1 Tennodai, Tsukuba Ibaraki Prefecture 305-8572, Japan. <sup>36</sup>Biosciences, College of Life & Environmental Sciences, University of Exeter, Stocker Road, Exeter EX4 4QD, UK. <sup>37</sup>Department of Biological Sciences, California State University San Marcos, San Marcos, California 92096, USA. <sup>38</sup>Provasoli-Guillard National Center for Marine Algae and Microbiota, Bigelow Laboratory for Ocean Sciences, 60 Bigelow Way, East Boothbay, Maine 04544, USA. <sup>39</sup>Department of Biological Sciences, California State University Chico, 1205 West 7th Street, Chico, California 95929-0515, USA. <sup>40</sup>Biology Department, Woods Hole Oceanographic Institution, Woods Hole, Massachusetts 02543, USA. <sup>41</sup>Oak Ridge National Laboratory, Oak Ridge, Tennessee 37831, USA.