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Systematic molecular and cytogenetic screening of 100 patients with marfanoid syndromes and intellectual disability

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The association of marfanoid habitus (MH) and intellectual disability (ID) has been reported in the literature, with overlapping presentations and genetic heterogeneity. A hundred patients (71 males and 29 females) with a MH and ID were recruited. Custom-designed 244K array-CGH (Agilent®; Agilent Technologies Inc., Santa Clara, CA) and *MED12*, *ZDHHC9*, *UPF3B*, *FBNI*, *TGFBR1* and *TGFBR2* sequencing analyses were performed. Eighty patients could be classified as isolated MH and ID: 12 chromosomal imbalances, 1 *FBNI* mutation and 1 possibly pathogenic *MED12* mutation were found (17%). Twenty patients could be classified as ID with other extra-skeletal features of the Marfan syndrome (MFS) spectrum: 4 pathogenic *FBNI* mutations and 4 chromosomal imbalances were found (2 patients with both *FBNI* mutation and chromosomal rearrangement) (29%). These results suggest either that there are more loci with genes yet to be discovered or that MH can also be a relatively non-specific feature of patients with ID. The search for aortic complications is mandatory even if MH is associated with ID since *FBNI* mutations or rearrangements were found in some patients. The excess of males is in favour of the involvement of other X-linked genes. Although it was impossible to make a diagnosis in 80% of patients, these results will improve genetic counselling in families.

Conflict of interest

The authors declare no conflict of interest.

P Callier^{a,b,†}, B Aral^{b,c,†}, N Hanna^{d,†}, S Lambert^{b,c}, H Dindy^d, C Ragon^{a,b}, M Payet^{a,b}, G Collod-Beroud^e, V Carmignac^b, MA Delrue^f, C Goizet^f, N Philip^g, T Busa^g, Y Dulac^h, I Missotteⁱ, Y Sznajer^j, A Toutain^k, C Francannet^l, A Megarbane^m, S Juliaⁿ, T Edouard^o, P Sarda^p, J Amiel^q, S Lyonnet^q, V Cormier-Daire^q, B Gilbert^r, A Jacqueline^s, D Heron^s, P Collignon^t, D Lacombe^f, F Morice-Picard^f, PS Jouk^u, V Cusin^v, M Willems^q, E Sarrazin^w, K Amarof^w, C Coubes^p, MC Addor^x, H Journal^y, E Colin^z, P Khau Van Kien^{aa}, C Baumann^{ab}, B Leheup^{ac}, D Martin-Coignard^{ad}, M Doco-Fenzy^{ae}, A Goldenberg^{af}, G Plessis^{ag}, J Thevenon^{ah}, L Pasquier^{ai}, S Odent^{ai}, P Vabres^b, F Huet^b, N Marle^{a,b}, AL Mosca-Boidron^{a,b}, F Mugneret^a, S Gauthier^{ah}, C Biquet^{aj}, C Thauvin-Robinet^{b,ah}, G Jondeau^v, C Boileau^d and L Faivre^{b,ah}

^aService de Cytogénétique, Plateau technique de Biologie, CHU, Dijon, France, ^bEquipe GAD, EA 4271, Université de Bourgogne, Dijon, France, ^cLaboratoire de Biologie Moléculaire, Plateau technique de Biologie, CHU, Dijon, France, ^dAPHP, Laboratoire de Biochimie et de Génétique moléculaire, Hôpital Ambroise Paré, Boulogne, France, ^eINSERM, UMR_S 910, Marseille, F-13000, France, ^fDépartement de Génétique, CHU Bordeaux et Université

de Bordeaux, Bordeaux, France,
⁹Département de Génétique, Assistance
Publique Hôpitaux de Marseille, Marseille,
France, ^hService de Cardiologie, CHU,
Toulouse, France, ⁱService de Pédiatrie,
Centre Hospitalier Territorial, Nouvelle
Calédonie, France, ^lService de
Génétique, CHU, Louvain, Belgique,
France, ^kService de Génétique, CHU,
Tours, France, ^lService de Génétique,
CHU, Clermont-Ferrand, France, ^mUnité
de Génétique Médicale, Faculté de
Médecine, Université Saint Joseph,
Beirut, Lebanon, ⁿService de Génétique,
CHU, Toulouse, France, ^oService de
Pédiatrie, CHU, Toulouse, France,
^pService de Génétique, CHU, Montpellier,
France, ^qDépartement de Génétique,
Hôpital Necker-Enfants Malades, Paris,
France, ^rGénétique Médicale, CHU,
Poitiers, France, ^sDépartement de
Génétique, Hôpital Pitié Salpêtrière, Paris,
France, ^tService de Génétique Médicale,
CH Font Pré, Toulon, France, ^uService de
Génétique Médicale, CHU, Grenoble,
France, ^vCentre de Référence Maladie de
Marfan, Hôpital Bichat, Paris, France,
^wCentre de Référence Caraibéen des
Maladies Rares Neurologiques et
Neuromusculaires, CHU de Fort de
France, Hôpital Pierre Zobda-Quitman,
La Martinique, France, ^xService de
Génétique Médicale, Centre Hospitalier
Vaudois, Lausanne, Switzerland, ^yService
de Génétique Médicale, CH, Vannes,
France, ^zDépartement de Génétique,
CHU, Angers, France, ^{aa}Service de
Biologie Moléculaire, CHU, Montpellier,
France, ^{ab}Service de Génétique
Médicale, Hôpital Robert Debré, Paris,
France, ^{ac}Département de Génétique,
CHU, Nancy, France, ^{ad}Service de
génétique Médicale, CH, Le Mans,
France, ^{ae}Département de Génétique et
Centre de Référence Maladies Rares
'Anomalies du Développement et
Syndromes Malformatifs' de la région Est,
CHU Reims et EA3801 SFR Cap-Santé,
Reims, France, ^{af}Département de
Génétique, CHU, Rouen, France,
^{ag}Service de Génétique, CHU, Caen,
France, ^{ah}Centre de Génétique et Centre
de Référence Anomalies du
Développement et Syndromes
Malformatifs, Hôpital d'Enfants, Dijon,
France, ^{ai}Service de Génétique Médicale
et Centre de Référence Anomalies du
Développement de l'Ouest, CHU,
Rennes, France, and ^{aj}Centre
d'Investigation Clinique – Epidémiologie
Clinique, CHU, Dijon, France

†These authors equally contributed to the
work.

Key words: *FBNI* gene – intellectual disability – Lujan-Fryns – marfanoid syndromes – *MED12* gene – submicroscopic chromosomal rearrangements

Corresponding author: Laurence Faivre, MD, PhD, Centre de Génétique, Hôpital d'Enfants, 10 bd Maréchal de Lattre de Tassigny, 21034 Dijon Cedex, France.

Tel.: 00 33 380 293 300;

fax: 00 33 380 293 266;

e-mail: laurence.faivre@chu-dijon.fr

Marfan syndrome (MFS) is a multisystem genetic disease that can affect the cardiovascular, skeletal, ophthalmic, and integumentary systems and the dura mater. The diagnosis is based on an international classification (1–3). The severity of the condition lies in the risk of dilation and subsequent dissection of the ascending aorta. The *FBNI* gene is the major gene in patients with MFS, and variable expression of the disease has been largely identified within and between families (4, 5). Patients with atypical presentations not fulfilling the criteria involving only one organ system exist but are rare (6). Patients with MFS do not usually present with intellectual disability (ID) (7). Patients with MFS can also display mutations in the *TGFBR1* and *TGFBR2* genes (8).

The term marfanoid habitus (MH) is used to describe patients with skeletal signs suggestive of MFS but who do not meet the international criteria (5, 7). The association of a MH and ID has been reported in the literature in syndromes with overlapping features. ID is defined, according to the American Association of Intellectual and Developmental Disabilities as significant limitations both in intellectual functioning (referring to general mental capacity such as learning, reasoning, and problem solving) and in adaptive behaviour (comprising conceptual, social and practical skills), and originates before the age of 18. Its incidence is 1.26/100 (9). The Lujan-Fryns syndrome (LJS) was first described in male patients with a family history suggesting an X-linked mode of inheritance (10, 11). The term LJS is now recognized to describe patients with MH (long, hyperlax fingers and toes, tall stature, dolichostenomelia), facial dysmorphism (prominent forehead contrasting with a long, narrow face, maxillary hypoplasia, small mandible, long nose with a high narrow nasal root); mild to moderate ID with behavioural abnormalities, that could include emotional instability, shyness, or even psychotic disturbances, generalized hypotonia, nasal speech, normal development of sexual features (12, 13). This syndrome has also been reported occasionally in females (14, 15). Thoracic aortic aneurysm (TAA) is usually

absent, but has been reported in two families, including one patient who required aortic surgery (16, 17). In 2007, the systematic screening of 737 genes annotated in the Vertebrate Genome Annotation database on the human chromosome X in 250 families with syndromic or non-syndromic X-linked ID led to the identification of hemizygous mutations in the *MED12*, *ZDHHC9*, and *UPF3B* genes in a very small number of familial cases (18–20). The *MED12* gene (MIM 300188, also called *HOPA* or *TRAP230*), located in Xq13, encodes for a subunit of the macromolecular complex known as Mediator, which is required for thyroid hormone-dependent activation and repression of transcription by RNA polymerase II (21). Med12-deficient zebra fish embryos show defects in the brain and neural crest and do not survive beyond 1 week after fertilization (22). *MED12* mutation have been found in only one large family with LJS, which was the index family reported by Lujan, and carrier females displayed no bias in chromosome X inactivation (11, 19). Mutations in the *ZDHHC9* gene (MIM 300646, zinc finger, and DHHC-type containing 9), located in Xq26.1, have been found in three families with LJS (18). It encodes a palmitoyl transferase, highly expressed in the brain, which catalyzes the post-translational modification of rat sarcoma viral oncogene homolog (RAS). The mechanism by which loss of function mutations of *ZDHHC9* lead to ID is unclear, but it may be through alteration of the relative proportion of the RAS proteins within the different compartments of nerve cells (23). Finally, mutations of the *UPF3B* gene (MIM 200298), located on Xq24, have been found in two families with LJS (20). The UPF3B protein is an important component of the nonsense-mediated mRNA decay surveillance machinery (24) and the mechanism by which loss of function mutations of *UPF3B* lead to ID is also unknown. Mutations of these three genes have also been found in patients with Opitz–Kaveggia (FG) syndrome (18, 20, 25) and in non-syndromic ID for *UPF3B* (20). These results have not been replicated to date in LJS.

ID and MH can also be found in Shprintzen–Goldberg syndrome (SGS) associated with

craniosynostosis (26, 27) and rare *de novo* mutations have been described in the *FBN1* or *TGFBR2* genes. These results are still a subject of debate in the literature (28–30). ID is only reported very occasionally in Loey-Dietz syndrome (LDS), secondary to mutations in the *TGFBR1* and *TGFBR2* genes (31). Finally, chromosomal imbalances have also been reported in sporadic observations, including one 15q21.1q21.3 deletion involving the *FBN1* gene (32) and two cases of terminal deletion of chromosome 5p (33, 34).

Therefore, the genetic aetiology of the association of a MH and ID represents a true challenge for individual diagnosis, follow-up and genetic counselling, particularly for sporadic presentations. Besides the clinical overlap between these different entities, the implication of each gene and its clinical spectrum in such a phenotype is unknown and little is known about the risk of aortic involvement. The aim of this research project was to conduct a clinical, cytogenetic and molecular study of a large cohort of 100 probands presenting with a skeletal marfanoid phenotype and ID, in order to provide answers to these questions, and help in the management of these patients.

Materials and methods

Patients

This study was prospectively designed. Patients were included by a clinical geneticist over an 18 months period. Inclusion criteria were the presence of ID and certain skeletal features of the Marfan spectrum, including at least two of the following clinical signs: height greater than the 97th centile in the absence of tall stature in parents, long, thin habitus, long, thin fingers, pectus abnormalities, and dolichostenomelia. A cardiac ultrasound and anterior eye chamber examination were required for all patients. The presence of heart and eye manifestations of the Marfan spectrum was not an exclusion criterion. Neuropsychological examination in order to evaluate non-subjectively the degree of ID could be performed in 15% of the patients only. In the other cases, ID was graduated according to the geneticist experience. Patients with a family history of MFS without ID were excluded from the study because the segregation was in favour of a different cause for MH and ID, and homocystinuria also needed to be ruled out. Inclusion was validated also after standard karyotype and *FMRI* gene study (Fragile-X) did not show any anomaly, although patients with apparently balanced chromosomal rearrangements were not excluded. Patients with the MH and ID were mainly recruited through the network of reference centres for rare diseases in France. Informed consent was obtained for all patients and the study was approved by the local ethics committee.

A detailed clinical evaluation was performed for every patient during a specialized consultation, and a detailed clinical file specifically designed for this project was completed by the referring geneticist, describing the family history, clinical features of

MFS and information regarding developmental data. Patients were divided into two groups, depending on the absence (group 1) or presence (group 2) of associated features characteristic of MFS (TAA, mitral valve prolapse or ectopia lentis). The median systemic score was calculated in the total cohort and among groups, according to the 2010 new Ghent criteria for the diagnosis of MFS and related disorders. A systemic score above 7 indicated systemic involvement (3).

Molecular and cytogenetic analyses

For etiological purposes, all patients were screened for mutations of the *ZDHHC9*, *UPF3B*, *MED12*, *TGFBR1*, *TGFBR2*, and *FBN1* genes and for chromosomal micro-rearrangements using 244 K custom microarray. Female patients were also screened for X-inactivation bias.

Direct sequencing of ZDHHC9, UPF3B, MED12, FBN1, TGFBR1 and TGFBR2 genes

Genomic DNA was extracted from blood samples of the patients and parents when available. Exons and exon–intron boundaries sequencing analyses of the *MED12*, *ZDHHC9*, *UPF3B*, *FBN1*, *TGFBR1* and *TGFBR2* genes were performed according to previously reported methods (8, 18, 20, 35, 36). Corresponding reference of the genomic DNA sequences were downloaded using Ensembl Genome Browser, www.ensembl.org. Sequence analyses were searched for between consensus and reference sequences using SEQSCAPE® software v2.5 package (Applied Biosystems, Foster City, CA). Mutation nomenclature numbering was based on the current Ensembl transcript (<http://www.hgvs.org>). The pathogenic nature of a mutation was determined according to a database search, bioinformatic predictions and segregation of the mutation in the family. The Universal Mutation Database (UMD) was used for *FBN1* (37) and *TGFBR2* (38) requests. When a mutation was suspected to lead to the creation of a splice site, additional mRNA studies were performed. All patients were screened for *MED12*, *ZDHHC9*, *UPF3B*, *FBN1* and *TGFBR2* gene mutations. Only patients with TAA were screened for *TGFBR1* mutations.

X-inactivation assay in female patients

This assay was performed as previously reported (39) with some slight modifications, notably the use of fluorescent primers and the detection mode. A control was used in the analyses. X-inactivation assay in female patients was considered as biased when it was superior or equal to 85%, or even 95%.

Array-CGH experiments

A custom-designed array was designed with approximately 240,000 oligonucleotides manufactured by Diagenode™ (Division Imaxio, Biopôle Saint-Beauzire, France). This array contains oligonucleotides

selected from the Agilent online library (earray; <https://earray.chem.agilent.com/earray>) and has been further empirically optimized. In addition, we increased probe density within seven selected genes (12,307, Table S1). The entire genome was covered with an average resolution of 20 kb (NCBI, hg 18). The procedures of array-CGH were performed according to the Agilent instructions with minor modifications (40). Slides were scanned using the Agilent G2565 Microarray scanner and images quantified using AGILENT FEATURE EXTRACTION (v9.0) and a graphical overview was obtained using GENOMIC WORKBENCH software (v5.0). Mapping data were analysed on the human genome sequence using Ensembl (www.ensembl.org; hg18). Copy Number Variations were assessed in the Database of Genomic Variants (<http://projects.tcag.ca/variation/>). When a chromosomal imbalance was detected, quantitative polymerase chain reaction (qPCR) or fluorescence in situ hybridization (FISH) studies with probes derived from bacterial artificial chromosomes were performed in order to confirm the chromosomal imbalances, as well as the family segregation. When a chromosomal imbalance was suspected to be pathogenic, the gene content was determined in order to find candidate genes for the phenotype, with a particular interest for genes expressed in the central nervous system and connective tissues (<http://www.ensembl.org/biomart/>). We also checked if a known 'microdeletion/microduplication syndrome' was described through the decipher database (<http://sanger.ac.uk>) and the available literature, and if additional patients with a similar phenotype had already been reported/gathered.

Results

Patients

Among the 100 patients with marfanoid phenotype and ID, 71 were males and 29 females (sex ratio: 2.45), originating from 98 families. They were aged from 2 to 45 years with a mean of 19.1 ± 8.6 years. Fifty percent of patients were adults. A family history of ID was found in 23/98 families, including 48% in a first-degree relative (91% in a sibling and 9% in a parent), 17% in a second-degree relative (including 50% compatible with an X-linked inheritance) and 35% in other degrees (including 12% compatible with an X-linked inheritance). MH was associated with ID in only four familial cases. Two patients had apparently balanced *de novo* reciprocal translocations: t(12;19)(q13.3;p12), and t(2;22)(q33;q11.2).

Detailed description of the overall cohort is given in Table 1. A minority was screened for dural ectasia and protrusio acetabulae. Other miscellaneous features included nasal speech ($n = 18$), abnormal genitalia ($n = 15$), ptosis ($n = 5$), vertebral instability ($n = 4$), deafness ($n = 3$), hypertrophic cardiomyopathy ($n = 2$), pneumothorax ($n = 1$), atrial septal defect ($n = 1$), aneurysm of the interventricular septum ($n = 1$), cleft lip ($n = 1$), absent uvula ($n = 1$), optic nerve atrophy

($n = 1$), horseshoe kidney ($n = 1$) and supernumerary mammary ($n = 1$).

General clinical, molecular and cytogenetic characteristics of patients belonging to group 1 and 2 are presented in Table 1. Clinical details of patients with a molecular or cytogenetic abnormality are presented in Tables 2–4. The systemic score for patients with a positive cytogenetic or molecular result was not significantly different from patients with normal results (7.6 ± 3.2 vs 6.6 ± 2.9).

Molecular and cytogenetics results

FBN1 sequencing analysis

Five pathogenic *FBN1* mutations were found, one splicing and four missense mutations (Table 2). Their pathogenicity was verified according to the UMD-predictor tool (41) and except the c.2534G>A mutation (p.Cys845Tyr), they had already been reported at least once in the literature. Interestingly, the c.4270C>G (p.Pro1424Ala) mutation was found in two unrelated patients with no aortic manifestation at 18 and 36 years. The segregation of the mutation could not be verified in either family. The mutation had already been reported in six other instances in the UMD-*FBN1* database (8, 42–46), and no ID was noticed when clinical data were available. The pathogenic nature of the *FBN1* c.8176C>T (p.Arg2726Trp) mutation in exon 64 was demonstrated by *in vitro* studies (47) and reported in a few instances in patients with isolated skeletal features and even incomplete penetrance (48). Within the UMD-*FBN1* database, we found 11 families with this mutation, and aortic dilatation was reported in two patients (37, 49). The mutation was also found in the father and sister of the proband. When re-examined, they displayed some very mild skeletal features of the Marfan spectrum, including dolichostenomelia, arachnodactyly, high arched palate, dental crowding and myopia in the father, as well as dolichostenomelia, mild scoliosis, significant striae and high arched palate in the sister. The presence of ID in the proband was finally explained by the co-occurrence of a *FBN1* mutation and a 17q21.31 microdeletion.

Four variants were considered as polymorphic or probably non-pathogenic, according to UMD-*FBN1* predictor and segregation analyses (Table S2). Of note, although considered a polymorphism in this study, the c.6700G>A (p.Val2234Met) variant was considered pathogenic in various publications (45, 50). Additional investigations were necessary to draw conclusions regarding the pathogenicity of the synonymous variant c.5097 C>T (p.Tyr1699Tyr), based on the prediction of a potential exonic splicing enhancers (ESE) disruption by the Human Splicing Finder tool (51). This hypothesis was ruled out as mRNA studies on fibroblasts were not in favour of abnormal splicing. The median systemic score of patients with *FBN1* mutations or deletions was above 7, whereas the median systemic score of patients without *FBN1* mutations or deletions was below 7, but the results were not significantly different (8.1 ± 4.1 vs 6.7 ± 2.9).

Table 1. Description of the population and results obtained in the total cohort, and depending on the classification of patients

	Group 1 Isolated MH and ID <i>n</i> = 80	Group 2 MH, ID and another MFS feature ^a <i>n</i> = 20	Overall series <i>n</i> = 100
Clinical characteristics			
Sex ratio (<i>n</i> = 100)	2.8	1.5	2.4
Mean age ± SD (<i>n</i> = 100)	18.4 ± 7.8	21.8 ± 11.1	19.1 ± 8.6
<i>Cardiovascular</i>			
TAA (<i>n</i> = 94)	0/74 (0%)	10/20 (50%)	11/94 (10%)
Mitral valve prolapse (<i>n</i> = 94)	0/74 (0%)	11/20 (55%)	12/94 (12%)
Other valvular abnormalities (<i>n</i> = 94)	6/74 (8%)	12/20 (60%)	19/94 (19%)
<i>Eye</i>			
Ectopia lentis (<i>n</i> = 92)	0/73 (0%)	2/19 (10%)	2/92 (2%)
Myopia (<i>n</i> = 92)	20/73 (27%)	3/19 (16%)	23/92 (25%)
Other eye abnormality (strabismus, astigmatism, hypermetropia, and nystagmus) (<i>n</i> = 79)	46/62 (74%)	8/17 (47%)	54/79 (68%)
<i>Skeleton</i>			
Long and thin habitus (<i>n</i> = 99)	76/80 (95%)	16/19 (84%)	92/99 (93%)
Arachnodactyly (<i>n</i> = 98)	61/79 (77%)	13/19 (68%)	74/98 (75%)
Dolichostenomelia (<i>n</i> = 96)	51/78 (65%)	14/18 (78%)	65/96 (68%)
Scoliosis (<i>n</i> = 99)	44/80 (55%)	12/19 (63%)	56/99 (56%)
Pectus abnormalities (<i>n</i> = 100)	38/80 (48%)	15/20 (75%)	53/100 (53%)
Joint laxity (<i>n</i> = 99)	39/80 (49%)	8/19 (42%)	47/99 (47%)
Flat feet (<i>n</i> = 98)	35/79 (44%)	8/19 (42%)	43/98 (44%)
Limitation of extension of the elbow (<i>n</i> = 97)	16/78 (20%)	1/19 (5%)	17/97 (17%)
Camptodactyly (<i>n</i> = 91)	10/74 (13%)	3/17 (18%)	13/91 (14%)
<i>Others</i>			
Striae (<i>n</i> = 98)	13/79 (16%)	5/19 (26%)	18/98 (18%)
Translucent skin (<i>n</i> = 88)	2/70 (3%)	2/18 (11%)	4/88 (4%)
<i>Median systemic score</i>			
According to 2010 Ghent criteria ± SD (<i>n</i> = 100)	6.6 ± 2.9	7.3 ± 3.1	6.8 ± 3.0
<i>Cerebral and cognitive</i>			
Degree of ID (<i>n</i> = 95) ^b			
• Mild ID	13/76 (17%)	4/19 (21%)	17/95 (18%)
• Moderate ID	43/76 (56%)	13/19 (68%)	56/95 (59%)
• Severe ID	20/76 (26%)	2/19 (10%)	22/95 (23%)
Behavioural abnormalities (including anxiety, hyperactivity, psychotic troubles, and ASD) (<i>n</i> = 98)	56/79 (71%)	13/19 (68%)	69/98 (70%)
Epilepsy (<i>n</i> = 95)	28/77 (36%)	5/18 (28%)	33/95 (35%)
Abnormal MRI (<i>n</i> = 45) ^c	10/40 (25%)	3/6 (50%)	13/45 (29%)
Molecular and cytogenetic results (<i>n</i> = 100)			
Submicroscopic chromosome rearrangements	11/80 (14%) ^d	5/20 (25%) ^d	16/100 (16%)
<i>FBN1</i>	2/80 (2%) ^d	3/20 (15%) ^d	5/100 (5%)
<i>TGFBR1</i> , <i>TGFBR2</i>	0/80 (0%)	0/20 (0%)	0/100 (0%)
<i>MED12</i>	1/80 (1%)	0/20 (0.0%)	1/100 (1%)
<i>UPF3B</i> , <i>ZDHHHC9</i>	0/80 (0%)	0/20 (0%)	0/100 (0%)
X-inactivation bias in females			
• >85% skewing	28/80 (35%)	0/20 (0%)	28/100 (28%)
• >95% skewing	1/80 (1.2%) ^e	0/20 (0%)	1/100 (1%)
Total	14/80 (18%)	6/20 (30%) ^d	20/100 (20%)

ASD, autism spectrum disorder; ID, intellectual deficiency; MFS, Marfan syndrome; MH, marfanoid habitus; MRI, magnetic resonance imaging; TAA, thoracic aortic aneurysm.

^aIncluding TAA, ectopia lentis, sinuous aorta and/or mitral valve prolapse.

^bTwenty-three percent of patients had neuropsychological evaluation.

^cIncluding absent corpus callosum, thin corpus callosum, corpus callosum dysgenesis, vermis hypoplasia, Arnold Chiari malformation, pituitary stalk interruption syndrome, asymmetric ventricles, cortical atrophy, hydrocephaly, arachnoidian cyst, enlarged Virchow Robin spaces, and enlarged cisterna major.

^dOne patient had both a *FBN1* mutation and a chromosomal rearrangement.

^ePatient P20 not carrying any cytogenetic nor molecular abnormality.

Table 2. Pathogenic *FBN1* mutations obtained from direct sequencing in the cohort of interest

Identifier	Nomenclature	Segregation	Number of reports in UMD-FBN1 database	Clinical features
Group 1 86	c.8176C>T (p.Arg2726Trp) exon 64 EVS: 14T/10744T	Inherited from pauci symp- tomatic father	11	<ul style="list-style-type: none"> • Male, 21 years, second- and third-degree relatives with ID on both sides • Skeletal features: 168 cm, dolichostenomelia, scoliosis, joint laxity, arachnodactyly, hyperlaxity with recurrent sprains, crowded teeth with irregular shape • Other MFS features: myopia • Moderate ID, epilepsy, abnormal pronunciation, attention deficit disorder, abnormal co-ordination, abnormal behaviour (automutilation and shyness), corpus callosum dysgenesis • Other features: VSD, recurrent ENT infections, anal fistula, right pyelocaliceal dilatation, dysmorphism, cryptorchidism, hyperelastic skin, pescaucus, fractures, gingival enlargement • Male, 18 years, no FH • Skeletal features: 188 cm, asymmetric pectus excavatum, dolichostenomelia, arachnodactyly, joint laxity, severe thoraco-lumbar cyphosis, protrusio acetabulae • Other MFS features: none • Moderate ID • Other features: nasal speech, perceptive deafness
97	c.4270C>G (p.Pro1424Ala) exon 34 EVS: 3G/10755C	Parents not available	6	<ul style="list-style-type: none"> • Female, 21 years, no FH • Skeletal features: 188 cm, long and thin habitus, dolichostenomelia, arachnodactyly, pectus excavatum, scoliosis, hyperlaxity, flat feet, typical dysmorphism, high arched palate. Surgery for metatarsus varus at age 2 • Other MFS features: TAA diagnosed at age 5 years, mitral valve prolapse with mitral insufficiency that required surgery at age 4 years, ectopialentis, striae, duralectasia • Moderate ID with Chiari malformation • Male, 54 years, father unknown • Skeletal features: 180 cm, scoliosis, pectus excavatum, arachnodactyly, high-arched palate with dental malposition • Other MFS features: lens dislocation that required surgery, mitral valve prolapse • Mild ID • Other features: hypertrophic cardiomyopathy • Male, 36 years, no FH • Skeletal features: long and thin habitus, pectus excavatum, dolichostenomelia, arachnodactyly • Other MFS features: severe mitral valve prolapse with massive mitral insufficiency requiring surgery at age 36 years, myopia • Moderate ID, epilepsy • Other features: ASD, vascular embolization of two intracranial sylvian aneurysms, dolichocephaly, plagiocephaly, down slanting palpebral fissures, small and round ears
Group 2 56	c.247+1G>A exon 2_3	<i>De novo</i>	13	<ul style="list-style-type: none"> • Female, 21 years, no FH • Skeletal features: 188 cm, long and thin habitus, dolichostenomelia, arachnodactyly, pectus excavatum, scoliosis, hyperlaxity, flat feet, typical dysmorphism, high arched palate. Surgery for metatarsus varus at age 2 • Other MFS features: TAA diagnosed at age 5 years, mitral valve prolapse with mitral insufficiency that required surgery at age 4 years, ectopialentis, striae, duralectasia • Moderate ID with Chiari malformation • Male, 54 years, father unknown • Skeletal features: 180 cm, scoliosis, pectus excavatum, arachnodactyly, high-arched palate with dental malposition • Other MFS features: lens dislocation that required surgery, mitral valve prolapse • Mild ID • Other features: hypertrophic cardiomyopathy • Male, 36 years, no FH • Skeletal features: long and thin habitus, pectus excavatum, dolichostenomelia, arachnodactyly • Other MFS features: severe mitral valve prolapse with massive mitral insufficiency requiring surgery at age 36 years, myopia • Moderate ID, epilepsy • Other features: ASD, vascular embolization of two intracranial sylvian aneurysms, dolichocephaly, plagiocephaly, down slanting palpebral fissures, small and round ears
83 ^a	c.2534A>G (p.Cys845Tyr) exon 20 EVS: not reported	Father not available	0	<ul style="list-style-type: none"> • Female, 21 years, no FH • Skeletal features: 188 cm, long and thin habitus, dolichostenomelia, arachnodactyly, pectus excavatum, scoliosis, hyperlaxity, flat feet, typical dysmorphism, high arched palate. Surgery for metatarsus varus at age 2 • Other MFS features: TAA diagnosed at age 5 years, mitral valve prolapse with mitral insufficiency that required surgery at age 4 years, ectopialentis, striae, duralectasia • Moderate ID with Chiari malformation • Male, 54 years, father unknown • Skeletal features: 180 cm, scoliosis, pectus excavatum, arachnodactyly, high-arched palate with dental malposition • Other MFS features: lens dislocation that required surgery, mitral valve prolapse • Mild ID • Other features: hypertrophic cardiomyopathy • Male, 36 years, no FH • Skeletal features: long and thin habitus, pectus excavatum, dolichostenomelia, arachnodactyly • Other MFS features: severe mitral valve prolapse with massive mitral insufficiency requiring surgery at age 36 years, myopia • Moderate ID, epilepsy • Other features: ASD, vascular embolization of two intracranial sylvian aneurysms, dolichocephaly, plagiocephaly, down slanting palpebral fissures, small and round ears
96	c.4270C>G (p.Pro1424Ala) exon 34 EVS: 3G/10755C	Parents not available	6	<ul style="list-style-type: none"> • Female, 21 years, no FH • Skeletal features: 188 cm, long and thin habitus, dolichostenomelia, arachnodactyly, pectus excavatum, scoliosis, hyperlaxity, flat feet, typical dysmorphism, high arched palate. Surgery for metatarsus varus at age 2 • Other MFS features: TAA diagnosed at age 5 years, mitral valve prolapse with mitral insufficiency that required surgery at age 4 years, ectopialentis, striae, duralectasia • Moderate ID with Chiari malformation • Male, 54 years, father unknown • Skeletal features: 180 cm, scoliosis, pectus excavatum, arachnodactyly, high-arched palate with dental malposition • Other MFS features: lens dislocation that required surgery, mitral valve prolapse • Mild ID • Other features: hypertrophic cardiomyopathy • Male, 36 years, no FH • Skeletal features: long and thin habitus, pectus excavatum, dolichostenomelia, arachnodactyly • Other MFS features: severe mitral valve prolapse with massive mitral insufficiency requiring surgery at age 36 years, myopia • Moderate ID, epilepsy • Other features: ASD, vascular embolization of two intracranial sylvian aneurysms, dolichocephaly, plagiocephaly, down slanting palpebral fissures, small and round ears

ASD, ventricular septal defect; ID, intellectual disability; ENT, ear, nose and throat; FH, family history; MFS, Marfan syndrome; TAA, thoracic aortic aneurysm; VSD, ventricular septal defect.

^aThe patients also carries a submicroscopic chromosomal rearrangement that was already reported in the literature for patient 86 (61) EVS, exome variant server: <http://evs.gs.washington.edu/EVS/>.

Table 3. Possibly pathogenic *MED12* mutation obtained from direct sequencing in the cohort of interest

Identifier	Nomenclature	Segregation	Clinical features
68	c.3884G>A (p.Arg1295His)* exon 28	Inherited from his mother with mild ID, and not found in the two asymptomatic brothers	<ul style="list-style-type: none"> • Male, 18 years, mild phenotype in her mother (skeletal features, mitral valve prolapse) • Skeletal features: 187 cm, long and thin habitus, pectus excavatum, joint laxity, malar hypoplasia, down slanting palpebral fissures, teeth malposition • Other MFS features : none • Mild ID • Other features: small and low-set ears, nasal speech, bilateral ptosis

ID, intellectual disability; MFS, Marfan syndrome.

*Not reported in EVS (<http://evs.gs.washington.edu/EVS/>).

TGFBR1 and TGFBR2 sequencing analysis

Neither pathogenic mutations nor known variants were found in the *TGFBR1* gene. One missense and three synonymous variants were found in the coding sequence of the *TGFBR2* gene. For three of them, their presence in an asymptomatic parent combined with database predictions were in favour of a polymorphism (Table S2). The c.1039C>T (p.Leu347Leu) variant was found to be *de novo*. On the basis of the prediction of a potential ESE disruption by the Human Splicing Finder tool (51), mRNA studies on lymphocytes were performed but the results were not in favour of abnormal splicing.

MED12, UPF3B, and ZDHHC9 sequencing analysis

One possibly pathogenic *MED12* mutation was found (c.3884G>A, p.Arg1295His) in patient 68 (Table 3). The use of different databases gave opposite results, databases against the pathogenicity, taking into account the fact that a histidine is present at this position in another gene of the MED family. However, this amino acid is highly conserved across species (Fig. S1), and was not found in 200 male controls tested in the laboratory, or in the 1000 Genomes Project Consortium or in the EVS database (<http://evs.gs.washington.edu/EVS/>). Segregation in the family showed that the variant was inherited from the mother, who had a milder phenotype, which could be compatible with an X-linked inheritance, and was absent in the two healthy brothers. Additional functional studies should be performed in order to arrive at a definitive conclusion.

Table S2 summarizes the likely non-pathogenic variants found in this cohort. The c.3692-22A>C *MED12* variant was predicted to break the potential branch point (tcctcAt, -22 position in the intron) according to the *in silico* splice site analysis program Human Splicing Finder (HSF) (51), but this hypothesis could not be confirmed by normal mRNA studies.

X-inactivation studies

An X-inactivation bias >85% was found in seven females of group 1 (7/20, 35%), but absent in both other groups. When considering X-inactivation bias >95%, only one female was concerned.

Targeted array-CGH

Among the 100 patients, 84 displayed normal results, with no copy number variants (CNVs) or only the presence of benign CNVs. For 16 patients, genomic imbalances were detected ranging from 500 kb to 11.6 Mb, (i) ten deletions, including one deletion 15q21.1q21.3 encompassing the *FBNI* gene; (ii) five duplications; and (iii) one patient with an unbalanced translocation (Table 4). The overviews of copy number changes of all patients along the whole genome are shown in Fig. 1. Two patients had both chromosomal rearrangements and a pathogenic *FBNI* mutation. Five patients had a known microdeletional syndrome (3q29, 15q13.2q13.3 and 17q12 microdeletions, 15q11.2 and 16p11.2 microduplications).

Discussion

The systematic screening of genes that may be involved in MH and ID in this study provides useful information regarding the work-up necessary in such patients. From a study of 100 patients, we showed that: (i) submicroscopic rearrangements are the most prevalent abnormalities, (ii) the presence of ID should not rule out the possibility of an *FBNI* mutation; (iii) the association of two pathogenic abnormalities was possible, which means that extreme caution must be exercised when considering relating the overall phenotype to a given anomaly, unless additional data become available in the literature; (iv) the X-linked *MED12*, *ZDHHC9* and *UPF3B* genes are not major genes for MH and ID, although a skewed sex ratio in favour of males was found.

Submicroscopic rearrangements were identified in 16/100 patients. This percentage is in the range of the overall detection rate of genomic imbalances by array-CGH in patients with ID and/or developmental delay (52). Retrospectively, the use of targeted custom oligonucleotide array-CGH did not permit the identification of intragenic rearrangement in the genes of interest. The chromosomal imbalances identified were all different, strengthening the hypothesis that high genetic heterogeneity exists in this phenotype. For each abnormality, we tried to determine if there was evidence that the rearrangement might explain the MH, and

Table 4. Submicroscopic chromosomal rearrangements detected by array-CGH in the cohort of patients with marfanoid habitus (MH) and mental retardation according to the group concerned (in grey, rearrangement that has already been described in the literature in association with a MH)

Identifier	Copy number change Chromosomal location	Size of rearrangement (Mb) Sequences co-ordinates (hg18)	Inheritance	Clinical features	References
Group 1 31	Loss 2q24.1q24.2	6.44 155397362–161841920	<i>De novo</i>	<ul style="list-style-type: none"> • Male, 15 years, FH of ID in maternal uncle • Skeletal features: 167 cm, long and thin habitus, arachnodactyly, dolichostenomelia, hyperlaxity, high arched palate, coxavalga • Other features of MFS: none • Severe ID, hyperactivity • Other features: facial dysmorphism, strabismus 	(62, 63)
103	Loss 3q23.3	1.5 137357633–138147814	<i>De novo</i>	<ul style="list-style-type: none"> • Male, 15 years, no FH • Skeletal features: size –1 SD, long and thin habitus, pectus excavatum, scoliosis, flat feet • Other features of MFS: none • Mild ID (IQ 66), autistic features • Other features: mild myopia 	
52	Loss 3q29.1 ^a	1.42 197216153–198643540	<i>De novo</i>	<ul style="list-style-type: none"> • Male, 16 years, no FH • Skeletal features: 172 cm, long and thin habitus, dolichostenomelia, arachnodactyly, pectus excavatum, scoliosis, flat feet, high arched palate • Other features of MFS: pneumothorax • Mild ID (IQ 54), anxiety, psychiatric manifestations with auditive hallucinations, dysarthria 	(53,54)
101	Loss 5p15.31p15.2 ^a	1.5 8824218–10286748	<i>De novo</i>	<ul style="list-style-type: none"> • Other features: mild camptodactyly • Female, 18 years, no FH • Skeletal features: 180 cm, long and thin habitus, dolichostenomelia, arachnodactyly, incomplete extension of the elbows, facial dysmorphism • Other features of MFS: striae • Mild ID, severe epilepsy • Other features: none 	(33,34)
13	Loss 6q27	2.42 168323802–170748862	Not known (father dead)	<ul style="list-style-type: none"> • Male, 14 years, no FH • Skeletal features: 167.5 cm, long and thin habitus, dolichostenomelia, arachnodactyly, pectus excavatum, scoliosis, hyperlaxity, facial dysmorphism • Other features of MFS: none • Moderate ID (IQ 46), hyperactivity, anxiety, autistic traits, mild ventricular dilatation • Other features: microcephaly, apparent blood vessels 	(64)

Table 4. Continued

Identifier	Copy number change Chromosomal location	Size of rearrangement (Mb) Sequences co-ordinates (hg18)	Inheritance	Clinical features	References
102 ^b	Gain 9q33.3q34.3 ^a	11.66 128531258–140193874	Not verified	<ul style="list-style-type: none"> • Male, 20 years, younger brother with the same appearance at birth who died of a cardiopathy at 2 years of age • Skeletal features: 169 cm, long and thin habitus, dolichostenomelia, limited extension of the elbows, scoliosis, high arched palate • Other features of MFS: none • Severe ID (IQ 35), sociable, jovial, very affectionate, nervous • Other features: mitral regurgitation, hypernasal voice, malrotation of the right kidney, epilepsy, delayed puberty, dysmorphism, hypodontia, hamershaped of the second toe, previously by Mégarbané et Chammas, (1997) • Male, 18 years, no FH • Skeletal features: 190.5 cm, long and thin habitus, dolichostenomelia, arachnodactyly, abnormal pectus, scoliosis, hyperlaxity, flat feet, high arched palate • Other features of MFS: mild myopia • Mild ID, hyperactivity, epilepsy • Other features: mild camptodactyly, mild dysmorphism, nasal speech, stammering and dysarthria 	(66)
47	Loss 11q23.1q23.2	5.18 110122508–115306206	<i>De novo</i>	<ul style="list-style-type: none"> • Male, 14 years, no FH • Skeletal features: >97th centile, long and thin habitus, arachnodactyly, asymmetric pectus excavatum, scoliosis, high-arched palate with crowded teeth, facial dysmorphism • Other features of MFS: surgery for inguinal herniae, ascending aorta upper the normal range at the level of the sinus of Valsalva, redundant mitral valve with mitral insufficiency • Mild ID, schizophrenia • Other features: nasal speech • Male, 16 years, no FH • Skeletal features: 181 cm, long and thin habitus, dolichostenomelia, arachnodactyly, scoliosis • Other features of MFS: none • Mild ID, schizophrenia • Other features: hypodontia 	(58)
3	Gain 12q13.3q14.4 (at the breakpoint of a <i>de novo</i> translocation 46,XY,t(12;19) (q13.3;p12)	0.701 56177083–56878427	<i>De novo</i>	<ul style="list-style-type: none"> • Male, 14 years, no FH • Skeletal features: 181 cm, long and thin habitus, dolichostenomelia, arachnodactyly, scoliosis • Other features of MFS: surgery for inguinal herniae, ascending aorta upper the normal range at the level of the sinus of Valsalva, redundant mitral valve with mitral insufficiency • Mild ID, schizophrenia • Other features: nasal speech • Male, 16 years, no FH • Skeletal features: 181 cm, long and thin habitus, dolichostenomelia, arachnodactyly, scoliosis • Other features of MFS: none • Mild ID, schizophrenia • Other features: hypodontia 	(66)
99	Gain 15q11.2 ^a	5.2 20811696–26138064	<i>De novo</i>	<ul style="list-style-type: none"> • Male, 14 years, no FH • Skeletal features: 181 cm, long and thin habitus, dolichostenomelia, arachnodactyly, scoliosis • Other features of MFS: surgery for inguinal herniae, ascending aorta upper the normal range at the level of the sinus of Valsalva, redundant mitral valve with mitral insufficiency • Mild ID, schizophrenia • Other features: nasal speech • Male, 16 years, no FH • Skeletal features: 181 cm, long and thin habitus, dolichostenomelia, arachnodactyly, scoliosis • Other features of MFS: none • Mild ID, schizophrenia • Other features: hypodontia 	(58)

Table 4. Continued

Identifier	Copy number change Chromosomal location	Size of rearrangement (Mb) Sequences co-ordinates (hg18)	Inheritance	Clinical features	References
6	Gain 16p11.2 ^a	1.70 28399426–30107008	<i>De novo</i>	<ul style="list-style-type: none"> • Male, 23 years, FH of 2 maternal cousins with autistic behaviour • Skeletal features: 193 cm, long and thin habitus, dolichostenomelia, arachnodactyly, scoliosis, asymmetric pectus excavatum, flat feet, and high arched palate • Other features of MFS: none • Mild ID, obsessive compulsive behaviour, anxiety, autistic features and psychosis episodes • Other features: mild dysmorphic features 	(56)
100	Loss 17q12 ^a	1.3 31891335–33242358	Not verified	<ul style="list-style-type: none"> • Female, 25 years, FH of ID in her mother and maternal uncle • Skeletal features: 193 cm, long and thin habitus, arachnodactyly, multiple surgery for severe scoliosis • Other features of MFS: none • Mild ID, psychotic behaviour • Other features: interventricular septal aneurysm with abnormal cardiac rhythm, recurrent fractures of the fingers with osteoporosis, facial dysmorphism (round face with hypertelorism), strabismus, pescavum, thenar and hypothenar hypoplasia 	(60)
86 ^c	Loss 17q21.31 ^a	0.504 41062469–41566740	<i>De novo</i>	Described in Table 2	(61)
69	Gain 19p13.3	1.66 4899227–6563324	<i>De novo</i>	<ul style="list-style-type: none"> • Male, 20 years, no FH • Skeletal features: 187 cm, dolichostenomelia, hyperlaxity, limited elbow extension, cyphosis, high arched palate and facial dysmorphism • Other features of MFS: none • Mild ID, abnormal behaviour including anxiety, hypersomnia and obsessive compulsive behaviour necessitating treatment • Other features: chiari I malformation and supra tentorial white matter hypersignals 	(67)
Group 2 83 ^b	Gain 2q37.3 ^a Loss 4q35.1q35.2	4.51 238189799–242701179 7.03 183993225–191028016	Unknown (father not available)	Described in Table 2	
39	Loss 15q13.2q13.3 ^a	1.57 28718936–30298296	<i>De novo</i>	<ul style="list-style-type: none"> • Male, 23 years, brother with mild school difficulties • Skeletal features: 194 cm, long and thin habitus, scoliosis, hyperlaxity, facial dysmorphism • Other features of MFS: TAA and bicuspid aortic valve • Mild ID, sleep disturbances with anxiety • Other features: hypermetropia 	(40, 59)

Table 4. Continued

Identifier	Copy number change Chromosomal location	Size of rearrangement (Mb) Sequences co-ordinates (hg18)	Inheritance	Clinical features	References
36	Loss 15q21.1q21.3	9.69 45866359–55560424	<i>De novo</i>	<ul style="list-style-type: none"> Female, 14 years, brother with mild school difficulties Skeletal features: 160 cm, long and thin habitus, dolichostenomelia, arachnodactyly, pectuscarinatum, scoliosis necessitating surgery (60°), flat feet, typical dysmorphism and high arched palate. Other features of MFS: mitral valve prolapse with mitral insufficiency, mild myopia Severe ID, anxious and coleric, epilepsy, incontinence, normal cerebral MRI Other features: delayed puberty that required hormonal treatment 	(55)

ID, intellectual disability; MFS, Marfan syndrome; MRI, magnetic resonance imaging; TAA, thoracic aortic aneurysm.

^aKnown decipher syndrome.

^bClinical description of this patient has been described previously (65).

^cThis patient also carries a pathogenic *FBNI* mutation.

if the results could provide some information regarding the genes responsible for MH. In our series, the presence of a marfanoid phenotype was recurrent in 4/16 rearrangements, including the 3q29 microdeletion (53, 54), the 5p15.31p15.2 microdeletion (33, 34), the 15q21.1q21.3 microdeletion (55) and the 16p11.2 microduplication (56) (Table 4). Except for the *FBNI* gene located in the 15q21.1q21.3 microdeletion, none of the other rearrangements comprised a good candidate gene for the MH. The recurrent *de novo* 3q29 microdeletion had a commonly deleted region of 1.6 Mb, most likely secondary to non-allelic homologous recombination. Besides mild/moderate ID and a long, narrow face, chest-wall deformity has been described in 3/15 patients, long, tapering fingers in 3/15 patients, and ligamentous hyperlaxity in 1/15 patient (53, 54). Of note, our patient had a more generalized MH and pneumothorax, which can be part of the MFS spectrum. Some patients with the 5pter microdeletion and with various marfanoid features in adolescence have already been described (33, 34). Unfortunately, previous patients were not studied using high resolution techniques. It was therefore not possible to determine the smallest region of overlap. Finally, it is highly probable that the phenotype found in the patient with chromosome 16p11.2 duplication could be fully explained by the cytogenetic abnormality as it has been recently shown that it was the countertype of patients presenting with chromosome 16p11.2 microdeletion with obesity and a wide range of behavioural abnormalities (57).

Conversely, we have data to conclude that the rearrangements in five patients were certainly responsible for ID but cannot explain the MH. Four of them are well-known frequent genomic rearrangements, comprising the 15q11.2 microduplication including *UBE3A/SNRPN* (58), the 15q13.3 microdeletion including *CHRNA7* (59), the 17q12 microdeletion (60) and finally the 17q21.31 microdeletion including *MAPT* (61), and manifestations of MFS have never been described in large series. The best argument against the causality of the 17q21.31 microdeletion in MH is that we identified in the same patient a concomitant pathogenic *FBNI* mutation. Similar conclusions can be drawn for the patient with an unbalanced translocation responsible for a 2q37.3 duplication and a 4q35.1q35.2 deletion, since the patient also carried a pathogenic *FBNI* mutation. These observations especially emphasize the importance of a systematic screening approach for accurate genetic counselling and clinical follow-up. Unfortunately, no conclusion can be drawn for the other rearrangements.

The second take home message from this study is that the presence of ID should not rule out the diagnosis of MFS with its aortic risk, in a patient with MH. Indeed, 6% of our patients had a *FBNI* mutation or a rearrangement involving *FBNI*, i.e. 35% of the abnormalities found in the study. This conclusion is of importance as approximately a third of the patients were not screened by echocardiography prior to the study. When an *FBNI* mutation was found, we have data showing that ID could be attributable to a different

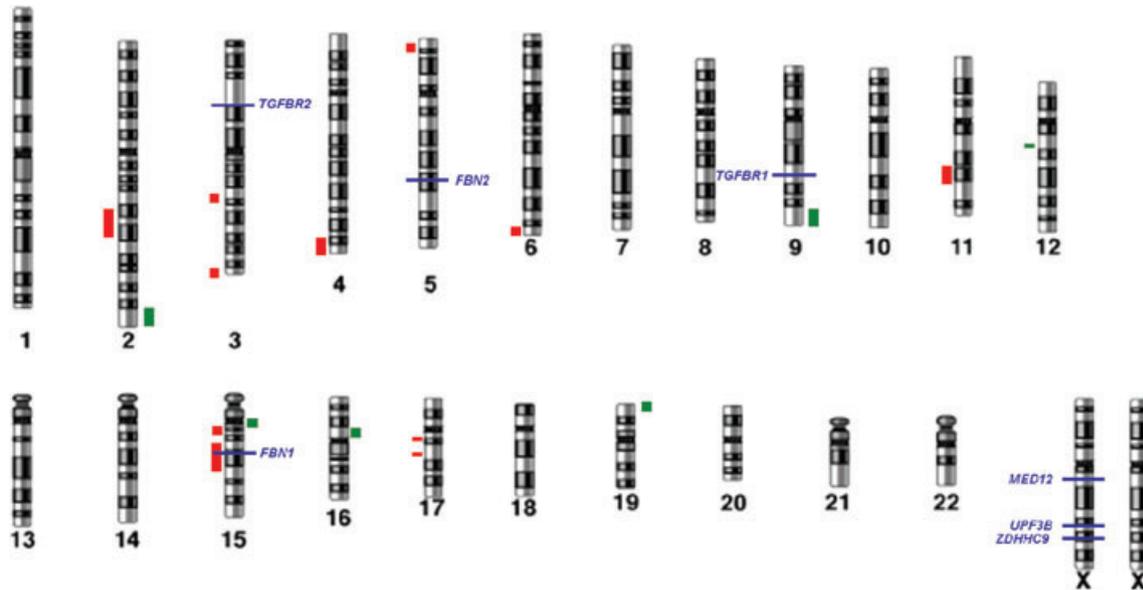


Fig. 1. Whole genome of the 16 abnormalities detected by custom array-CGH and map of targeted genes. DNA copy number losses and gains are highlighted in red and green, respectively, at the left and right, respectively, along the chromosome location.

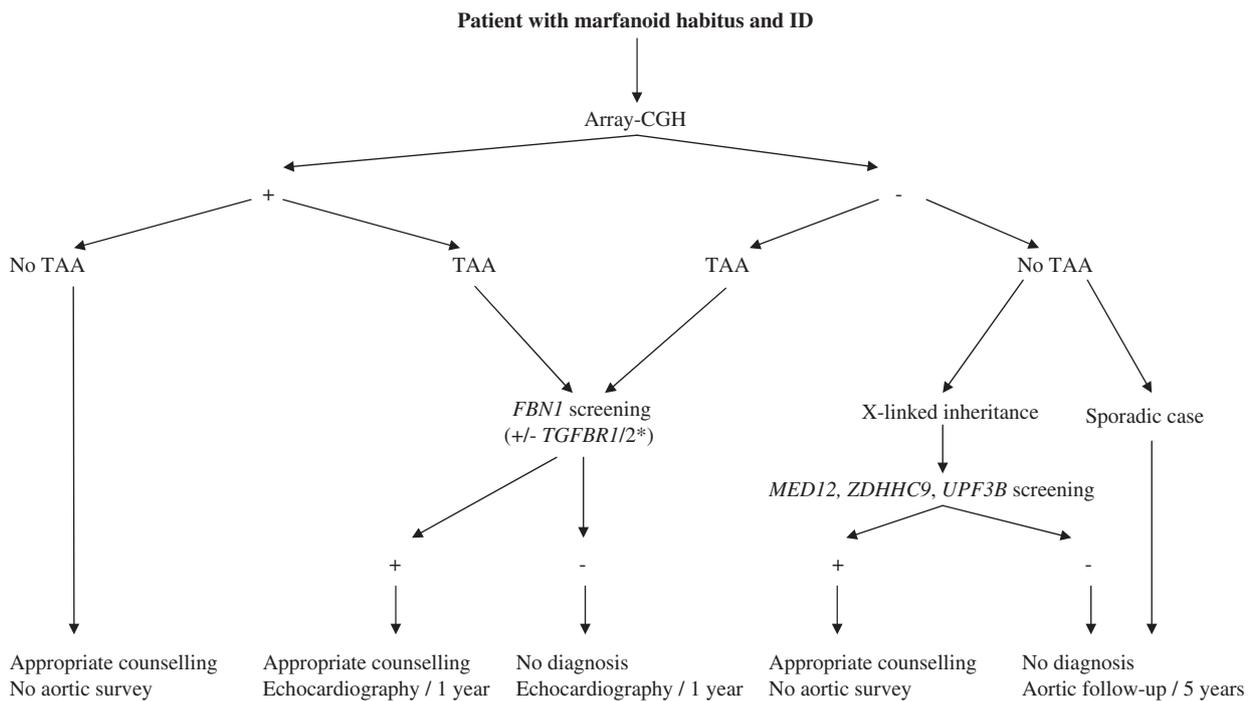


Fig. 2. Flow chart for the diagnosis of patients with MH and ID. TAA, thoracic aortic aneurysm. *The list of genes to be tested will lengthen as new syndromic TAA genes are identified.

cause, since two of the patients also had a chromosomal micro-rearrangement that explained the ID. Also, a large study of patients with MFS did not reveal an increased risk of ID (7). Given the very high allelic heterogeneity of the *FBNI* gene, it was surprising to find the same mutation (c.4270C>G, p.Pro1424Ala) in two unrelated patients of the study. However, six additional patients with the same mutation and with no ID were reported in the UMD-*FBNI* database. The

majority of *FBNI* mutations/rearrangements was found in patients with TAA, ectopia lentis and/or mitral valve prolapse (four out of five were from group 2), but only skeletal manifestations were diagnosed at 22 years of age (patient 97, Table 2), confirming the high clinical variability of patients with *FBNI* mutations, who can present with isolated skeletal features (47).

By combining all these data it was possible to create a flow chart (Fig. 2) with the following recommendation:

echocardiography and eye examination should be systematically performed in patients with MH and ID. If TAA or ectopia lentis is found, *FBNI* screening is warranted for genetic counselling and follow-up. The systemic score in patients with an *FBNI* mutation did not appear to be significantly different from that in other patients of the cohort. If the echocardiography and eye examination are normal, they should be performed at least every 5 years, to take into account the features that evolve with time.

The third answer from this study is that the three X-linked genes found in association with LFS are not major genes in this clinical population, at least in sporadic patients. Indeed, only one probably pathogenic *MED12* mutation (c.3884G>A, p.Arg1295His) was identified in our cohort of 100 patients. However, very few familial cases were included, and we therefore cannot draw any conclusions about the frequency of *MED12* involvement in cases with X-linked inheritance. Nevertheless, the role of other X-linked genes appears probable because there was an obvious skewed sex ratio in favour of males in our cohort of patients.

Finally, no molecular or cytogenetic abnormalities were found in 80% of the patients (82% in group 1 and 70% in group 2). Chromosomal micro-rearrangements found in this study could point to new candidate regions to be studied to identify new genes responsible for the phenotype. Further studies with next-generation sequencing technology such as the exome approach using a trio strategy will hopefully help in the diagnosis of such patients. In particular, a distinct entity which associates TAA and intellectual deficiency may exist because no *FBNI* mutation was found in the majority of the patients. Besides to the hypothesis that more loci with genes are yet to be discovered, an alternative explanation would be that MH is a relatively non-specific feature of patient with ID. Future studies on the subject would be useful to know if a diagnostician should give importance or not on the marfanoid findings in association to ID, given that it can be a common genetic problem in the population.

In conclusion, this collaborative work suggests a practical diagnostic pathway resulting in a better clinical differentiation, and providing a basis for more effective management and appropriate genetic counselling for families with MH and ID.

Supporting Information

The following Supporting information is available for this article:

Fig S1. Interspecies conservation of the amino acid concerned by the *MED12* missense mutation in patient 68.

Table S1. Candidate genes with numbers of clones added to the custom array-CGH design.

Table S2. Non-pathogenic variant obtained from direct sequencing of the *FBNI*, *TGFBR1*, *TGFBR2*, *MED12*, *UPF3B* and *ZDHC9* in the cohort of interest.

Additional Supporting information may be found in the online version of this article.

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References

1. De Paepe A, Devereux RB, Dietz HC, Hennekam RC, Pyeritz RE. Revised diagnostic criteria for the Marfan syndrome. *Am J Med Genet* 1996; 62: 417–426.
2. Faivre L, Collod-Beroud G, Ades L et al. The new Ghent criteria for Marfan syndrome: what do they change? *Clin Genet* 2012; 81: 433–442.
3. Loeys BL, Dietz HC, Braverman AC et al. The revised Ghent nosology for the Marfan syndrome. *J Med Genet* 2010; 47: 476–485.
4. Faivre L, Collod-Beroud G, Loeys BL et al. Effect of mutation type and location on clinical outcome in 1,013 probands with Marfan syndrome or related phenotypes and *FBNI* mutations: an international study. *Am J Hum Genet* 2007; 81: 454–466.
5. Faivre L, Collod-Beroud G, Child A et al. Contribution of molecular analyses in diagnosing Marfan syndrome and type I fibrillinopathies: an international study of 1009 probands. *J Med Genet* 2008; 45: 384–390.
6. Faivre L, Collod-Beroud G, Callewaert B et al. Pathogenic *FBNI* mutations in 146 adults not meeting clinical diagnostic criteria for Marfan syndrome: further delineation of type I fibrillinopathies and focus on patients with an isolated major criterion. *Am J Med Genet A* 2009; 149A: 854–860.
7. Faivre L, Masurel-Paulet A, Collod-Beroud G et al. Clinical and molecular study of 320 children with Marfan syndrome and related type I fibrillinopathies in a series of 1009 probands with pathogenic *FBNI* mutations. *Pediatrics* 2009; 123: 391–398.
8. Stheneur C, Collod-Beroud G, Faivre L et al. Identification of 23 *TGFBR2* and 6 *TGFBR1* gene mutations and genotype-phenotype investigations in 457 patients with Marfan syndrome type I and II, Loeys-Dietz syndrome and related disorders. *Hum Mutat* 2008; 29: E284–E295.
9. Heikura U, Taanila A, Olsen P et al. Temporal changes in incidence and prevalence of intellectual disability between two birth cohorts in Northern Finland. *Am J Ment Retard* 2003; 108: 19–31.
10. Fryns JP, Buttiens M. X-linked mental retardation with marfanoid habitus. *Am J Med Genet* 1987; 28: 267–274.
11. Lujan JE, Carlin ME, Lubs HA. A form of X-linked mental retardation with marfanoid habitus. *Am J Med Genet* 1984; 17: 311–322.
12. Fryns JP, Van Den Berghe H. X-linked mental retardation with Marfanoid habitus: a changing phenotype with age? *Genet Couns* 1991; 2: 241–244.
13. Van Buggenhout G, Fryns JP. Lujan-Fryns syndrome (mental retardation, X-linked, marfanoid habitus). *Orphanet J Rare Dis* 2006; 1: 26.
14. Fryns JP. X-linked mental retardation with marfanoid habitus. *Am J Med Genet* 1991; 38: 233.
15. Gurrieri F, Neri G. A girl with the Lujan-Fryns syndrome. *Am J Med Genet* 1991; 38: 290–291.
16. Lerma-Carrillo I, Molina JD, Cuevas-Duran T et al. Psychopathology in the Lujan-Fryns syndrome: report of two patients and review. *Am J Med Genet A* 2006; 140: 2807–2811.
17. Wittine LM, Josephson KD, Williams MS. Aortic root dilation in apparent Lujan-Fryns syndrome. *Am J Med Genet* 1999; 86: 405–409.
18. Raymond FL, Tarpey PS, Edkins S et al. Mutations in *ZDHC9*, which encodes a palmitoyltransferase of *NRAS* and *HRAS*, cause X-linked mental retardation associated with a Marfanoid habitus. *Am J Hum Genet* 2007; 80: 982–987.
19. Schwartz CE, Tarpey PS, Lubs HA et al. The original Lujan syndrome family has a novel missense mutation (p.N1007S) in the *MED12* gene. *J Med Genet* 2007; 44: 472–477.
20. Tarpey PS, Raymond FL, Nguyen LS et al. Mutations in *UPF3B*, a member of the nonsense-mediated mRNA decay complex, cause syndromic and nonsyndromic mental retardation. *Nat Genet* 2007; 39: 1127–1133.
21. Blazek E, Mittler G, Meisterernst M. The mediator of RNA polymerase II. *Chromosoma* 2005; 113: 399–408.

22. Hong SK, Haldin CE, Lawson ND et al. The zebrafish *kohtalo/trap230* gene is required for the development of the brain, neural crest, and pronephric kidney. *Proc Natl Acad Sci U S A* 2005; 102: 18473–18478.
23. Swarthout JT, Lobo S, Farh L et al. DHHC9 and GCP16 constitute a human protein fatty acyltransferase with specificity for H- and N-Ras. *J Biol Chem* 2005; 280: 31141–31148.
24. Conti E, Izaurralde E. Nonsense-mediated mRNA decay: molecular insights and mechanistic variations across species. *Curr Opin Cell Biol* 2005; 17: 316–325.
25. Risheg H, Graham JM Jr, Clark RD et al. A recurrent mutation in MED12 leading to R961W causes Opitz-Kaveggia syndrome. *Nat Genet* 2007; 39: 451–453.
26. Greally MT, Carey JC, Milewicz DM et al. Shprintzen-Goldberg syndrome: a clinical analysis. *Am J Med Genet* 1998; 76: 202–212.
27. Robinson PN, Neumann LM, Demuth S et al. Shprintzen-Goldberg syndrome: fourteen new patients and a clinical analysis. *Am J Med Genet A* 2005; 135: 251–262.
28. Kosaki K, Takahashi D, Udaka T et al. Molecular pathology of Shprintzen-Goldberg syndrome. *Am J Med Genet A* 2006; 140: 104–108 author reply 109–110.
29. Sood S, Eldadah ZA, Krause WL, McIntosh I, Dietz HC. Mutation in fibrillin-1 and the Marfanoid-craniosynostosis (Shprintzen-Goldberg) syndrome. *Nat Genet* 1996; 12: 209–211.
30. van Steensel MA, van Geel M, Parren LJ, Schrandt-Stumpel CT, Marcus-Soekarman D. Shprintzen-Goldberg syndrome associated with a novel missense mutation in TGFBR2. *Exp Dermatol* 2008; 17: 362–365.
31. Loeys BL, Chen J, Neptune ER et al. A syndrome of altered cardiovascular, craniofacial, neurocognitive and skeletal development caused by mutations in TGFBR1 or TGFBR2. *Nat Genet* 2005; 37: 275–281.
32. Ades LC, Sullivan K, Biggin A et al. FBN1, TGFBR1, and the Marfan-craniosynostosis/mental retardation disorders revisited. *Am J Med Genet A* 2006; 140: 1047–1058.
33. Stathopulu E, Ogilvie CM, Flinter FA. Terminal deletion of chromosome 5p in a patient with phenotypical features of Lujan-Fryns syndrome. *Am J Med Genet A* 2003; 119A: 363–366.
34. Zhang SZ, Tang YC, Dai FP, Niebuhr E. 5p:12q translocation with manifestations of cri du chat syndrome and Marfanoid arachnodactyly. *Clin Genet* 1990; 37: 153–157.
35. Sandhu HK, Sarkar M, Turner BM, Wassink TH, Philibert RA. Polymorphism analysis of HOPA: a candidate gene for schizophrenia. *Am J Med Genet B Neuropsychiatr Genet* 2003; 123B: 33–38.
36. Stheneur C, Oberkamp B, Chevallier B. Marfan syndrome: diagnostic criteria and molecular biology contribution. *Arch Pediatr* 2008; 15: 564–567.
37. Colod-Beroud G, Le Bourdelles S, Ades L et al. Update of the UMD-FBN1 mutation database and creation of an FBN1 polymorphism database. *Hum Mutat* 2003; 22: 199–208.
38. Frederic MY, Hamroun D, Faivre L et al. A new locus-specific database (LSDB) for mutations in the TGFBR2 gene: UMD-TGFBR2. *Hum Mutat* 2008; 29: 33–38.
39. Allen RC, Zoghbi HY, Moseley AB, Rosenblatt HM, Belmont JW. Methylation of HpaII and HhaI sites near the polymorphic CAG repeat in the human androgen-receptor gene correlates with X chromosome inactivation. *Am J Hum Genet* 1992; 51: 1229–1239.
40. Masurel-Paulet A, Andrieux J, Callier P et al. Delineation of 15q13.3 microdeletions. *Clin Genet* 2010; 78: 149–161.
41. Frederic MY, Lalonde M, Boileau C et al. UMD-predictor, a new prediction tool for nucleotide substitution pathogenicity -- application to four genes: FBN1, FBN2, TGFBR1, and TGFBR2. *Hum Mutat* 2009; 30: 952–959.
42. Arbustini E, Grasso M, Ansaldo S et al. Identification of sixty-two novel and twelve known FBN1 mutations in eighty-one unrelated probands with Marfan syndrome and other fibrillinopathies. *Hum Mutat* 2005; 26: 494.
43. Biggin A, Holman K, Brett M, Bennetts B, Ades L. Detection of thirty novel FBN1 mutations in patients with Marfan syndrome or a related fibrillinopathy. *Hum Mutat* 2004; 23: 99.
44. Comeglio P, Evans AL, Brice GW, Child AH. Detection of six novel FBN1 mutations in British patients affected by Marfan syndrome. *Hum Mutat* 2001; 18: 251.
45. Comeglio P, Johnson P, Arno G et al. The importance of mutation detection in Marfan syndrome and Marfan-related disorders: report of 193 FBN1 mutations. *Hum Mutat* 2007; 28: 928.
46. Howarth R, Yearwood C, Harvey JF. Application of dHPLC for mutation detection of the fibrillin-1 gene for the diagnosis of Marfan syndrome in a National Health Service Laboratory. *Genet Test* 2007; 11: 146–152.
47. Milewicz DM, Grossfield J, Cao SN et al. A mutation in FBN1 disrupts profibrillin processing and results in isolated skeletal features of the Marfan syndrome. *J Clin Invest* 1995; 95: 2373–2378.
48. Buoni S, Zannolli R, Macucci F et al. The FBN1 (R2726W) mutation is not fully penetrant. *Ann Hum Genet* 2004; 68: 633–638.
49. Van Dijk FS, Hamel BC, Hilhorst-Hofstee Y et al. Compound-heterozygous Marfan syndrome. *Eur J Med Genet* 2009; 52: 1–5.
50. Tjeldhorn L, Rand-Hendriksen S, Gervin K et al. Rapid and efficient FBN1 mutation detection using automated sample preparation and direct sequencing as the primary strategy. *Genet Test* 2006; 10: 258–264.
51. Desmet FO, Hamroun D, Lalonde M et al. Human Splicing Finder: an online bioinformatics tool to predict splicing signals. *Nucleic Acids Res* 2009; 37: e67.
52. de Vries BB, Pfundt R, Leisink M et al. Diagnostic genome profiling in mental retardation. *Am J Hum Genet* 2005; 77: 606–616.
53. Ballif BC, Theisen A, Coppinger J et al. Expanding the clinical phenotype of the 3q29 microdeletion syndrome and characterization of the reciprocal microduplication. *Mol Cytogenet* 2008; 1: 8.
54. Willatt L, Cox J, Barber J et al. 3q29 microdeletion syndrome: clinical and molecular characterization of a new syndrome. *Am J Hum Genet* 2005; 77: 154–160.
55. Faivre L, Khau Van Kien P, Callier P et al. De novo 15q21.1q21.2 deletion identified through FBN1 MLPA and refined by 244K array-CGH in a female teenager with incomplete Marfan syndrome. *Eur J Med Genet* 2010; 53: 208–212.
56. Jacquemont S, Reymond A, Zufferey F et al. Mirror extreme BMI phenotypes associated with gene dosage at the chromosome 16p11.2 locus. *Nature* 2011; 478: 97–102.
57. Walters RG, Jacquemont S, Valsesia A et al. A new highly penetrant form of obesity due to deletions on chromosome 16p11.2. *Nature* 2010; 463: 671–675.
58. Kitsiou-Tzeli S, Tzetis M, Sofocleous C et al. De novo interstitial duplication of the 15q11.2-q14 PWS/AS region of maternal origin: clinical description, array CGH analysis, and review of the literature. *Am J Med Genet A* 2010; 152A: 1925–1932.
59. Sharp AJ, Mefford HC, Li K et al. A recurrent 15q13.3 microdeletion syndrome associated with mental retardation and seizures. *Nat Genet* 2008; 40: 322–328.
60. Nagamani SC, Erez A, Shen J et al. Clinical spectrum associated with recurrent genomic rearrangements in chromosome 17q12. *Eur J Hum Genet* 2010; 18: 278–284.
61. Dubourg C, Sanlaville D, Doco-Fenzy M et al. Clinical and molecular characterization of 17q21.31 microdeletion syndrome in 14 French patients with mental retardation. *Eur J Med Genet* 2011; 54: 144–151.
62. Magri C, Piovani G, Pilotta A et al. De novo deletion of chromosome 2q24.2 region in a mentally retarded boy with muscular hypotonia. *Eur J Med Genet* 2011; 54: 361–364.
63. Palumbo O, Palumbo P, Palladino T et al. A novel deletion in 2q24.1q24.2 in a girl with mental retardation and generalized hypotonia: a case report. *Mol Cytogenet* 2012; 5: 1.
64. Mosca AL, Callier P, Masurel-Paulet A et al. Cytogenetic and array-CGH characterization of a 6q27 deletion in a patient with developmental delay and features of Ehlers-Danlos syndrome. *Am J Med Genet A* 2010; 152A: 1314–1317.
65. Megarbane A, Chammas C. Severe mental retardation with marfanoid habitus in a young Lebanese male. A diagnostic challenge. *Genet Couns* 1997; 8: 195–200.
66. Vermeesch JR, Syrrou M, Salden I et al. Mosaicism for duplication 12q (12q13-->12q21.2) accompanied by a pericentric inversion in a dysmorphic female infant. *J Med Genet* 2002; 39: e72.
67. Sigberg L, Olsen P, Nanto-Salonen K, Knuutila S. 19p13.3 aberrations are associated with dysmorphic features and deviant psychomotor development. *Cytogenet Genome Res* 2011; 132: 8–15.