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# Ultrasonic emissions reveal individual cavitation bubbles in water-stressed wood

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## Summary

Under drought conditions, the xylem of trees that conducts ascending sap produces ultrasonic emissions whose exact origin is not clear. We introduce a new method to record simultaneously both acoustical events and optical observation of the xylem conduits within slices of wood that were embedded in a transparent material setting a hydric stress. In this article, we resolved the fast development of all cavitation bubbles and demonstrated that each ultrasound emission was linked to the nucleation of one single bubble, whose acoustic energy is an increasing function of the size of the conduit where nucleation occurred and also of the hydric stress. We modeled these observations by the fact that water columns in conduits store elastic energy and release it into acoustic waves when they are broken by cavitation bubbles. Water columns are thus elastic, and not rigid, "wires of water" set under tension by hydric stresses. Cavitation bubbles are at the origin of an embolism, whose development was followed in our experiments. Such an embolism of sap circulation can evolve in a fatal condition for living trees. These findings provide new insight for the non-destructive monitoring of embolism within trees, and suggest a new approach to study porous media under hydric stress.

**Keywords:** bubble, nucleation, xylem, acoustic emissions

## 1 Introduction

Sap ascends in trees through the xylem tissues (wood), a porous structure containing a network of parallel and in-

terconnected conduits that are a few dozen micrometers in diameter, using a seemingly hazardous mechanism relying on water under tension [1, 2]. This mechanism has for long generated controversy because tensed water is metastable: water columns may be seen as "wires of water" [3] as opposed to the "ropes of sand" proposed by early opponents of the cohesion tension theory [4, 5]. Physicists have since established that pure liquid water does sustain extremely negative pressures, negative by tens of MPa [6]. Beyond this pressure, water vaporizes by cavitation. Because of the large contact area between the wood cell walls and the water column that greatly favors the bubble nucleation, the risk of heterogeneous cavitation in trees is very high. Cavitation occurs when the sap pressure is abnormally negative, in case of drought for instance, generating an embolism of the sap circulation which can prove to be fatal for the trees [7]. Most trees have a narrow safety margin away from this hydraulic rupture [8]. If cavitation is beneficial in other context such as the discharge of spores in ferns [9] (or harmless in human cracking joints [10]), it is now seen as a major player in the determination of tree mortality by drought [11].

However, the physical mechanism of cavitation development in trees remains unknown, the two main hypotheses being "nucleation" and "air seeding". Experimental investigations are still largely constrained by methodological issues. Most methods are invasive and subject to artifacts [12], while non-invasive observation tools such as magnetic resonance imaging [13] or X-ray microtomography [14] do not provide fast high resolution images.

An indirect but very attractive approach is to record acoustic emissions from intact wood, as for non-destructive testing of engineering structures [15]. Milburn and Johnson were the first to record "clicks" [16] in the audible acoustic range, while Tyree and Dixon [17] evidenced ultrasonic emissions. It is established that ultrasonic emissions are more frequent under water stress conditions [7] and are correlated with embolism patterns [18, 19, 20] suggesting that ultrasounds may be linked to cavitation. But there is still

no direct evidence of the exact origin of ultrasounds, which represents a major limitation for its use in cavitation studies.

Here we investigated the question of the origin of ultrasounds emissions. We developed a new method to record simultaneously both acoustical emission and optical observations of thin wood slices molded in a transparent hydrogel reproducing the live conditions in a tree. This method could also be useful to study any porous media under hydric stress [21].

## 2 Experiment

To monitor functional xylem conduits, we cut 50  $\mu\text{m}$  thick slices from fresh xylem samples (Scots pine, *Pinus sylvestris* L.) cut along the longitudinal axis of the conduits, called tracheids in gymnosperms. This thickness ensured that the sample became transparent to visible light while it still contained several intact conduits. We chose Scots pine species whose wood is made of short tracheids. The intact tracheids lengths we measured in our samples ranged from 20  $\mu\text{m}$  to 1200  $\mu\text{m}$ . The samples were around 5 mm long. This ensured that the samples contained intact tracheids.

This technique using slice samples was used in several studies [22, 23] but has the drawback of exposing the sectioned slice to air entry. In order to generate the hydric stress in wood and to avoid air infiltration, we molded the wood slices in polyhydroxyethylmethacrylate (pHEMA), a wet and stiff hydrogel formulated according to Wheeler & Stroock [24]. The hydrogel is porous to water, allowing mass transfer of water from the wood to the hydrogel surface, while preventing air invasion, since the mesh size of the hydrogel is nanometric [24]. The molding in between two half-cured hydrogel slabs is described in Fig. 1 and detailed in the Appendix A. Materials and Methods. Note that the final UV curing did not result in a shrinking of the gel, therefore the gel closely matched the shape of wood. It resulted in a close bounding of the hydrogel with the wood slice, without visible void space in between them where bubbles could propagate (see Fig. 1b).

We placed the sample in a controlled subsaturated atmosphere: evaporation at the surface of the porous hydrogel induced a hydric stress within the sample (Fig. 2a). The level of hydric stress is quantified by the water potential  $\Psi$ . At equilibrium,  $\Psi$  is directly set by the humidity level: it is 0 for an atmosphere saturated in humidity, and negative for a non-saturated atmosphere. The building of a hydric stress was similarly to what occurs naturally within the leaf of a tree. The atmospheric potential of water vapor equi-

librated through the leaf/hydrogel (acting as a membrane) with the liquid contained in the conduits: xylem conduits or artificial conduits as in [24]. This hydric stress was thus transmitted to the liquid contained in the wood inclusion, which resulted in the lowering of the liquid pressure  $p$  compared to the atmospheric pressure  $p_0$  (since  $\Psi = p - p_0$  in a pure liquid), until cavitation occurred in the wood conduits, at a water pressure potential expected to be around -3.6 MPa for Scots pine [25]. Note that the water potential slowly built within the hydrogel sample, because it took time for water to diffuse out of the gel [26]. The hydrogel was sufficiently stiff to sustain this very large pressure and did not cavitate (rupture) itself. Under evaporation both wood and hydrogel shrunk, and we did not observe the apparition of voids due to detachment of hydrogel from wood in our experiments reported.

During this drought experiment we monitored the inclusion both (i) optically with a camera mounted on a microscope (Fig. 2b) and (ii) acoustically with high sensitivity microphones (Fig. 2c).

## 3 Results

During the water stress, air bubbles suddenly appeared in conduits (defining an "optical event", that was detected by image analysis). Then, one air bubble usually took a characteristic time of  $4 \pm 3$  s before it filled the entire conduit. We distinguished two types of optical events: on the one hand, the "nucleation" events, starting in a fully wet area and, on the other hand, the "air seeding" events, here defined as the appearance of bubbles near an already gas filled conduit. The first type of events showed that pre-existing bubbles were not necessary for the start of the cavitation process (Fig. 3ab). The second phenomenon resulted in extending patches of gas (Fig. 3c). This proved that the embolism development in our system was not the gradual propagation of a gas front, but the abrupt appearance of bubbles, and their intermittent development from several nuclei.

A second major finding was obtained from the detection of all "acoustic events" (signals whose amplitude exceeded a defined threshold value) in the ultrasonic range. We investigated what was the precise moment of these acoustic events, zooming on a few conduits and recording with a high-speed camera. Recordings showed that a sound was always synchronized with a bubble appearance at the millisecond time scale, see Fig. 3a. This result clearly indicated that the ultrasound emission was correlated in time with bubble nucleation.

Monitoring entire samples (at 3.75 frames per second)

we found that nearly all acoustic events (118 over a total of 118 and 84 over a total of 87 for samples 1 and 2 respectively) were correlated in time with one specific bubble nucleation with a delay that was less than the time interval between images. This therefore discards other possible origins of sounds, such as wood failures or conduit collapses [17] at least for our specific case of a conifer species whose conduits are only tracheids.

From the correlation of acoustic events with optical events we obtained the spatiotemporal distribution of emissions, i.e. we could plot the position of the conduits where single ultrasound emissions were detected, obtaining the analog of a seismic map, see Fig. 4A, providing the point of origin of each sound. Plotting the time of filling for each conduit, we clearly observed on Fig. 4B the extending patches of gas propagating by "air seeding".

A striking result was that sound energies were widely scattered (size of circles in Fig. 4A), and that some bubbles do not even produce any recordable ultrasound. For a typical experiment with a millimetric sample the counts were 84 acoustic events and 463 optical events (sample 2). The acoustic recordings synchronized with optical events clearly suggested that the energy of each acoustic event increased with the volume of the corresponding conduit, for two samples (Fig. 5a for sample 1 and Fig. 5c for sample 2).

## 4 Discussion

We now interpret the acoustic results, considering the elastic energy that is stored in the conduits. Under the large hydric pressures developed by evaporation, the volumic elasticity of water comes into play. As water is tensed, it is elastically stretched with respect to its initial volume at rest: the larger the water conduit, the higher the storage of total elastic energy. Assuming that always the same fraction of elastic energy could be converted in acoustic energy, the variations during the experiment were thus explained by the variability of the conduit sizes and the degree of water tension at cavitation.

The stored elastic energy can be predicted quantitatively. It depends on the hydric negative pressure  $p$  of the liquid. The nucleation of a bubble suddenly relaxes this pressure to a near vanishing value  $p_{\text{relax}} \simeq 0$ . Indeed, after nucleation the liquid pressure is in equilibrium with the gas pressure (neglecting capillary effects), the latter being close to vacuum in a freshly cavitated bubble, before water vapor and then air dissolved in surrounding liquid fills the bubble [26].

The initial negative pressure  $p$  is stretching both (i) the

volume of water that is contained in a conduit of volume  $V_c$  and (ii) the conduit walls. The volume  $\Delta V$  of the bubble just after nucleation ( $< 1\mu s$ ) is therefore the sum of the volumes released by the relaxation of these two stretched volumes. It is such as  $\Delta V/V_c = p/K$ , where  $K$  is an effective elastic bulk modulus. Because water and wood volumes are stretched in series by the same pressure, the effective modulus writes  $K = 1/(1/K_{\text{water}} + 1/K_c)$  [26, 27], with the compression modulus of water  $K_{\text{water}}$  (2.2 GPa), and a volumetric elastic modulus  $K_c \sim G$  linked to the deformation of the material surrounding the cavity ( $G$  being the elastic shear modulus of the solid material surrounding the conduit). Here we followed [28] and we used the value of  $K_c = 0.75$  GPa, typical of conifers. This results in an effective modulus of  $K = 0.5$  GPa. With this new parameter we can now evaluate the density of elastic energy prior cavitation to be  $1/2 \times K(\Delta V/V_c)^2 = 1/2 \times p^2/K$ .

The total elastic energy stored in the cavity volume therefore writes:

$$E_{\text{elastic}} = \frac{1}{2} \frac{p^2}{K} V_c \quad (1)$$

The energy stored is a function of the stretching tension  $p$ , and the cavity volume  $V_c$ . Upon bubble nucleation, this elastic energy was partly released into acoustic radiation [26]. The microphone captured a part of this energy; the remaining energy being emitted in air or absorbed in the different materials (wood, hydrogel and structure). The same experiments were performed on artificial wood with a simpler geometry in order to validate this prediction. This mimetic material consisted of a hydrogel containing spheroidal cavities that were initially filled with water, and then set under hydric stress according to the Wheeler et al. method [24], providing nucleation at  $-20$  MPa. Thanks to this calibration experiment, we showed that over a wide range of cavities volumes the acoustic energy of events was proportional to the water volume, see Fig. 6. Our results thus provide a first mechanistic demonstration of previous observations of Mayr et al. [29], who suggested that the energy of acoustic emissions may correlate with the average tracheid dimensions.

The ultrasound energy therefore mainly reflected the conduit volume, so that smaller volumes produced a sound level that was below the detection threshold value of our microphones. The threshold value was fixed in order to remove the noise. Variations in tension at cavitation (because of different conduit anatomies) or variations in distances to the microphone accounted for the distribution of acoustic energies at a given volume.

In addition, we noted a difference between the cavitation bubbles occurring near intact conduits ("nucleation"), and cavitation bubbles occurring near already cavitating conduits ("air seeding", extending the gaseous patches). A minority of bubbles originate from nucleations (15 %) at earlier times, but on average they were much more likely to produce a recordable emission (41 % against 21%). Our interpretation is that the level of liquid pressure to be specified in equation (1) is less negative for "air seeding" bubbles. We expect such bubbles to be triggered when the pressure difference between the filled conduit and the cavitating conduit,  $\Delta p = p - p_{\text{cavitating}} \simeq p$ , overcomes a threshold value. This threshold value may be less negative on average than the "nucleation" pressure (else we would observe isolated nucleations only). This results in a lower released elastic energy according to equation (1), decreasing the likelihood of detecting such acoustic events. Note that two hypotheses remain for the origin of "air seeding" bubbles: (a) air propagates through a pit pore (a few dozens of nanometers [30]) or (b) the pit ruptures and breaks open irreversibly [31]. Note also that we cannot here detect optically the propagation of air, and a nucleation occurring near a cavitating conduit would be classified as an air seeding event. Last, we must specify that the acoustic signals were similar for both types of bubbles, but nucleations occurred only at the beginning of the experiments.

In the course of time, the average acoustic energy of each bubble clearly increased and then decreased after a maximum value (Fig. 5b for sample 1 and Fig. 5d for sample 2). We can also interpret this climax with the prediction. The stretching tension is expected to initially increase because of the progression of dehydration, which accounts for the energy increase, but the volume of cavitating conduits also decreased drastically (insert Fig. 5bd), which accounts for the final decrease.

## 5 Conclusion

In summary, we proposed a technique to monitor the development of embolism in a slice of xylem wood under hydric stress. The propagation of embolism is found to be the consequence of two abrupt phenomena: nucleation and discontinuous propagation from conduit to conduit (air-seeding). The original combination of acoustic and fast optical microscopy brought insights in the induction of cavitation. Remarkably, we demonstrated that every ultrasound emission in xylem was triggered by a bubble nucleation. This discards other possible origins of sounds, such as wood failure or conduit collapse at least for our specific case of a

conifer wood whose conduits are tracheids. Moreover, we showed that the energy we recorded with the microphones was linked to the mechanical elastic energy that was released during the cavitation event. In a living tree, water stressed bark also emits ultrasounds but they are not linked to xylem cavitation [32]: further experiments are needed to filter out those emissions. The origin of sound in xylem is the consequence of the sudden relaxation of tension by a bubble. The vibration causing the acoustic emission remains to be resolved. One relevant hypothesis is a bubble vibration: indeed within artificial wood in hydrogel it was found that nucleated bubbles oscillate in volume for a few periods, at a MHz frequency linked to the compression modulus of water and the surrounding matrix [27].

These findings strengthen the ultrasonic approach to monitor cavitation. The results enlightened that the energy of the acoustic signals was linked to the volumes of the cavitating conduits. A promising application opened by this study is that the energy of acoustic events goes through a maximum in the middle of the cavitation process, which proves useful to interpret the cavitation state of a living tree. With a simple and non-invasive ultrasonic sensor, there is now a great potential to follow *in situ* the effect of severe droughts associated with climate changes on the survival of forests.

## Appendix. Materials and Methods

To prepare wood inclusions, 50  $\mu\text{m}$  thick xylem slices were cut with a microtome, tangentially to the surface of the tree (radial plane) allowing most pits connecting conduits to be observed from the side. The fresh and still humid xylem was molded in pHEMA hydrogel, see Fig. 1. The pHEMA hydrogel was built by polymerization of a monomer solution of HEMA. The solution of HEMA was directly inspired from Wheeler et al. formulation [24]: 2-hydroxyethyl methacrylate (65 vol%), ethyleneglycol (6 vol%), methacrylic acid (1 vol%), de-ionized water (28 vol%) with an additional 1 vol% of photoinitiator solution (600 mg 2,2-dimethoxy-2-phenylacetophenone dissolved in 1 ml of n-vinyl pyrrolidone). All chemicals were from Sigma Chemicals. To include the xylem slice in hydrogel, we prepared two half-cured (1 minute in UV light, within a UV chamber, CL 508 from Uvitec Cambridge) hydrogel slabs 400  $\mu\text{m}$  thick, prepared in between two glass slides spaced by a rectangular jig. Then we bound the three layers of gel/xylem slice/gel as shown on Fig. 1 and terminated the polymerization of the hydrogel with UV light in the UV chamber during 3 minutes. The assembly was done with watered slabs, in or-

der to avoid trapping bubbles and keep the system always fully wet. Applying a slight finger pressure before curing was required in order to ensure a good binding and to avoid detachment during drying (as observed with unsuccessfully bound samples). Artificial wood was prepared as described in [24], by bubbling Argon gas in lightly cured monomer solution, resulting in spheroidal water-filled cavities with a wide range of sizes. Both artificial wood and wood inclusion were soaked for at least 24h in deionized and degassed water in order to dissolve potential bubbles.

The samples were held directly in between two high-frequency microphones (PICO-HF 1.2, bandwidth 0.5-1.85 MHz, from Physical Acoustic Corporation) facing each other, using a thin layer of coupling elastomer (Tensive gel, from Parker laboratories) to insure a good acoustic coupling between the gels and the microphones. The clamping configuration of the microphones avoided any contact with other surfaces and thus avoided frictional noises. The sample and microphones were placed in a closed chamber made of two Petri dishes and a home-made centimetric spacer, all being wrapped with parafilm. The hydric stress was imposed by a saturated KCl solution in the bottom dish, ensuring a relative humidity of 85 %, corresponding to a water potential of around -22 MPa at equilibrium [33]. This imposed hydric stress was well below the one causing 50 % reduction in hydraulic conductance of Scots pine (-3.6 MPa [25]), and was thus never reached in the conduits because of cavitation.

The samples were optically monitored under a zoom stereo-microscope with a camera (AVT Marlin F-421B, imaging at 3.75 Hz). Image analysis helped to detect appearance of bubble, by looking at the differences between

images separated by around 1 second. The volume of conduits was estimated from the conduit length as measured on images, multiplied by the cross-section area. To follow the fast nucleation of bubbles, we used a fast camera (Miro 4, Vision Research, operating at 1000 frames per second).

An acquisition card (USB AE Node, Physical Acoustics) recorded the microphone signals during 700  $\mu$ s when the acoustic pressure exceeded a threshold value of 4 mPa. The acoustic energy received by a microphone is given by  $E_{\text{mic}} = S_{\text{mic}} \int \frac{p(t)^2}{\rho_p c} dt$ , with  $S_{\text{mic}}$  the microphone area in contact with the hydrogel,  $p(t)$  the acoustic pressure received by the microphone (using the calibration chart to convert voltages into Pascals),  $\rho_p = 1274 \text{ kg/m}^3$  and  $c = 2000 \text{ m/s}$  the density and the speed of sound in the hydrogel. The speed of sound within the hydrogel was measured with pure hydrogel samples of thicknesses  $e$  ranging from 0.3 to 1.8 mm, and placed inside a water tank and probed in echo mode with ultrasound pulses generated by a piezo-transducer connected to a pulser-receiver (model 5073PR from Panametrics).

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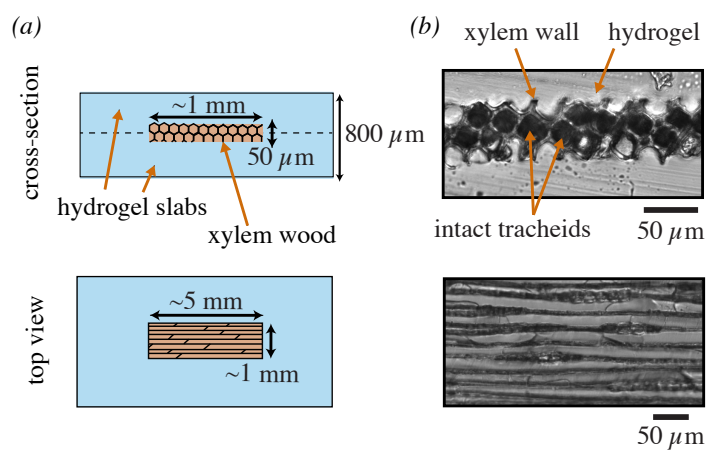


Fig. 1: Inclusion of wood samples in a stiff hydrogel matrix: the slices of xylem are inserted underwater in between two half-cured hydrogel slabs. (a) schematic views of the inclusion after assembly and total curing of hydrogel slabs (dashes indicated the assembly). (b) Optical observations. Above, a cross-section cut performed with a scalpel clearly shows that the hydrogel is intimately bound to the wood. Intact channels are still in contact side by side. Below: top view of the wood inclusions.



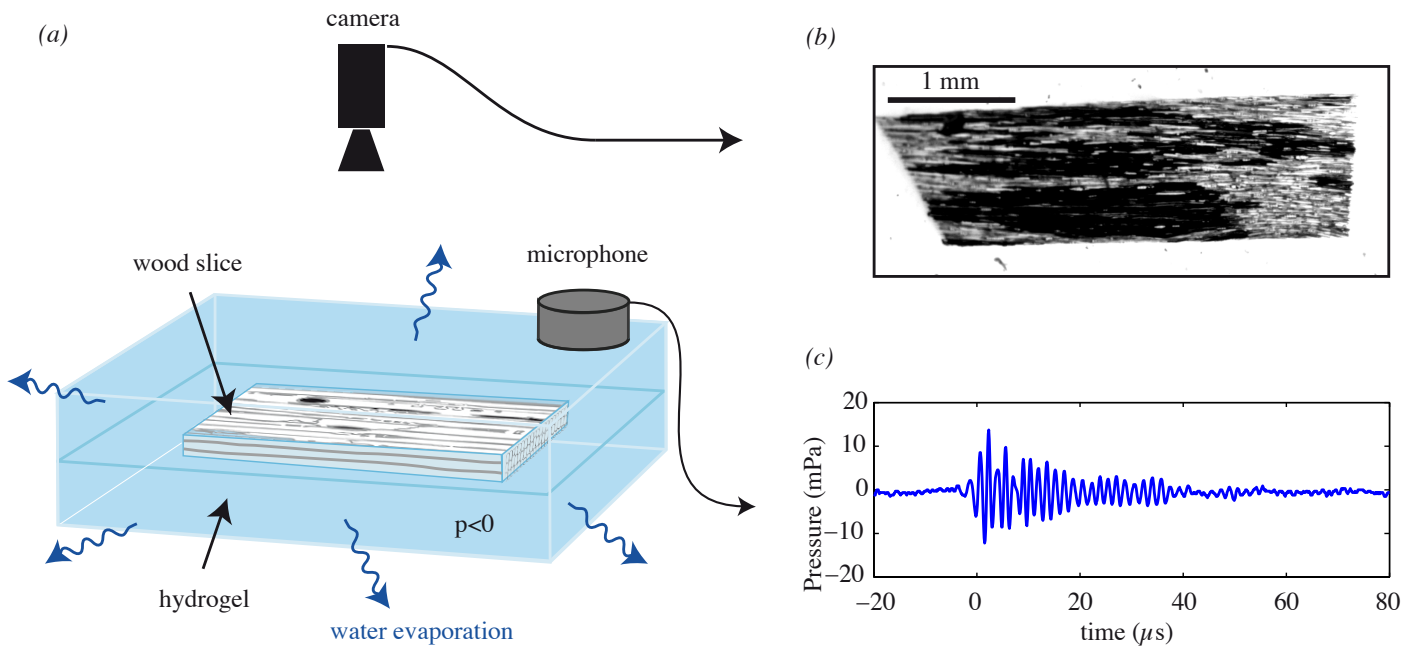


Fig. 2: Experimental set-up: (a) the water contained in the wood inclusion in hydrogel underwent negative pressure as water evaporates from the hydrogel. The camera installed on an optical microscope with magnification between 1.25x and 20x recorded images (b) of bubbles development (black conduits on image) in the water columns (grey on image). (c) Simultaneously the microphone monitored the ultrasonic signals.

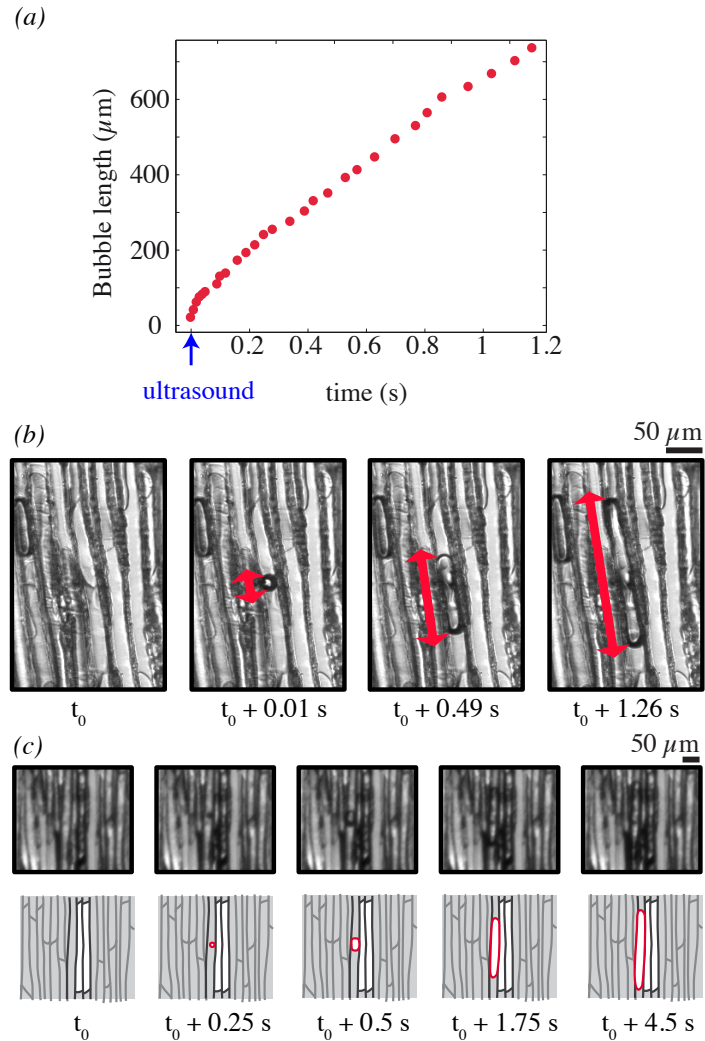


Fig. 3: (a) Bubble length as a function of time (red symbols, measured on some selected of images). The timing of the detected sound is indicated with the blue arrow. (b) Nucleation of a bubble in a conduit, recorded with a high-speed camera (1000 frames per second). The red line indicates the bubble length. (c) Air-seeding event, when a bubble nucleates near an already cavitated conduit, recorded with a 4 Hz camera. The evolution is sketched below each images, to increase the readability.

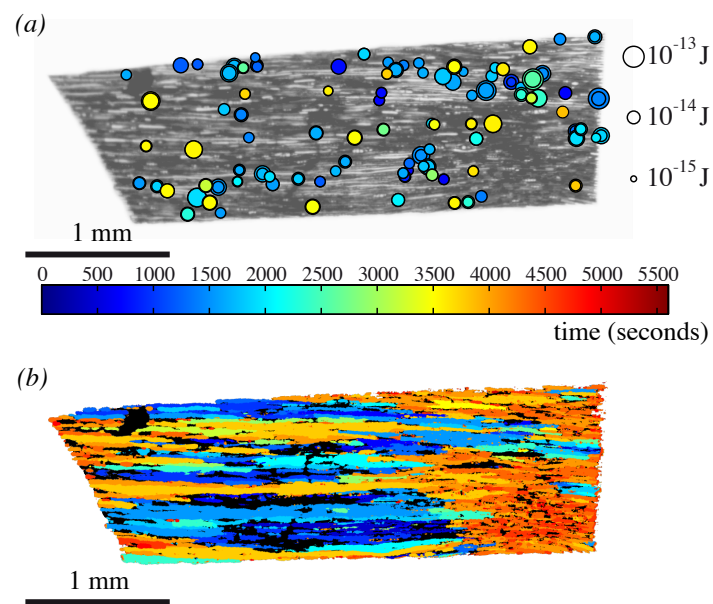


Fig. 4: (a) "Seismic" map, showing the location of bubbles appearance. The circle size indicates the magnitude of the acoustic energy, while color indicates apparition time. For clarity, the events after 3600 s were not shown. See also Supplementary movie 1. (b) Global map of air invasion by cavitation: color indicates the time of the filling of conduits with gas. See also Supplementary movie 2.

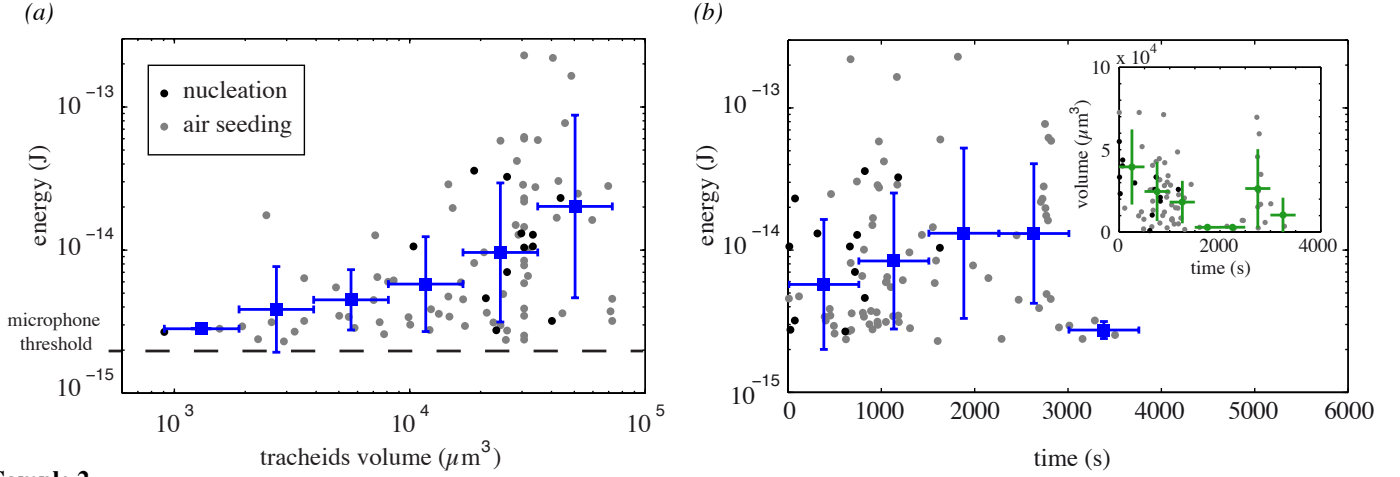
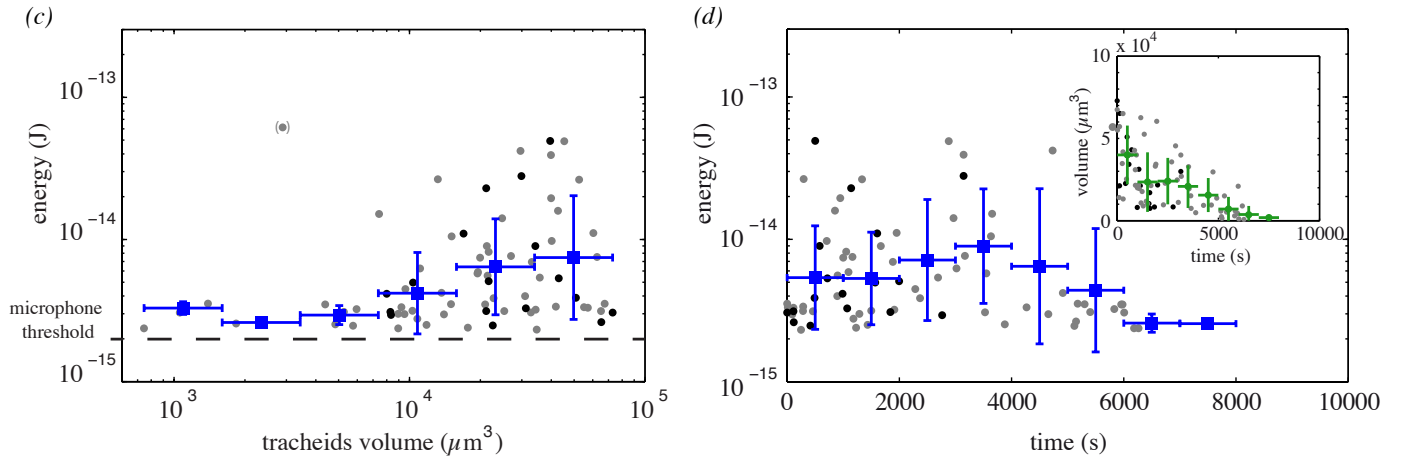
**Sample 1****Sample 2**

Fig. 5: (a) Variation of the acoustic energy correlated with bubble appearance as a function of the conduit volume, in xylem. The blue squares indicate the logarithmic average in a bin whose size is specified by the horizontal blue line. The vertical blue line represents the standard deviation of the points in the bin. The grey symbols show individual events. "Nucleations" are highlighted by dark symbols. The dashed line is a guide to the eye showing the microphone threshold. (b) Energy in the course of time. Insert: volume of each cavitating conduit in the course of time. For the figure and the insert,  $t = 0$  s corresponds to the first acoustic event and the error bars are determined in the same way as in (a). The sample for (a) and (b) is the one presented in Fig.2 and Fig.4. (c) and (d): same graphs for a second sample.

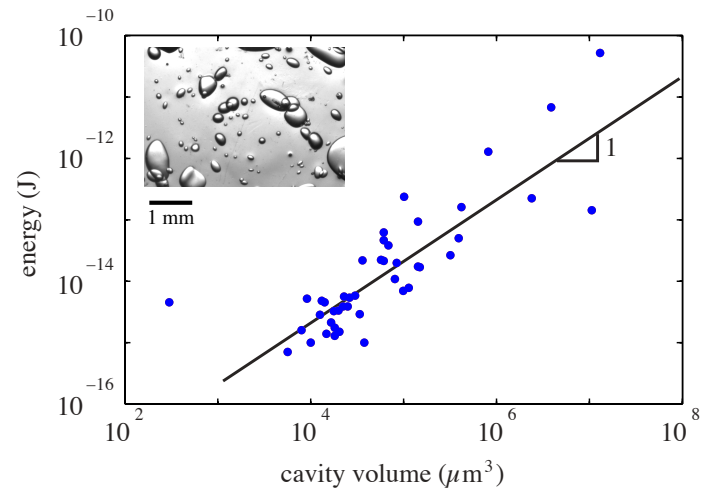


Fig. 6: Synthetic wood consisting in spheroidal cavities in hydrogel: acoustic energy as a function of the volume of artificial cavities. A picture of the experiment is shown in insert. To calculate cavities volumes, we considered them to be spherical. The radius is deduced from the pictures where we fit an ellipse to the observed cross section of the cavities and take the mean of the two ellipse axes as the radius.

Fig. 7: Supplementary material, Movie 1. Animated version of figure 4 (a) played a hundred times faster, with cavitation sounds. The original acoustic emissions (in the ultrasound range) are slowed down (1000 x) to become audible.

Fig. 8: Supplementary material, Movie 2. Animated version of figure 4 (b), with cavitation sounds, same parameters as Movie 1.