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Physical characteristics and Chemical compositions of Leaves extracts of *Sorindeia grandifolia* Engl. (Anacardiaceae) harvested at Kato, Benin

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Abstract

The results brought back at the end of this work concern various chemical constituents of *S. grandifolia* leaves collected in Benin. The major compounds ($\geq 4\%$) of essential oils obtained after hydrodistillation and analysis by coupling gas chromatography with spectrometry mass are constituted of : limonene (20.2%), (*E*)- β -ocimene (7.6-17.8%), (*Z*)- β -ocimene (11.4%), γ -cadinene (7.0%), selin-11-en-4- α -ol (4.5-5.7%), palmitic acid (4.3-6.2%), β -pinene (5.5%), α -selinene (5.5%), α -phellandrene (5.2%), β -selinene (4.9%), β -elemene (4.5%). The lipidic fractions realized from petroleum ether extracts are marked by important rates of arachidic acid (22.5%), palmitic acid (20.3%) and of linoleic acid (15.9%). The phytochemical analysis showed relatively a large content of coumarins, gallic tannins, flavones, leucoanthocyanins and saponins in opposition to the other secondary metabolites.

Keywords: limonene, arachidic acid, metabolites, coumarins

Introduction

Since ancient times, plant extracts, in particular aromatic, are sought for their bioactive properties. These plants are frequently used in the traditional pharmacopoeia in Benin for the therapeutic purposes in particular to relieve certain cutaneous affections, respiratory disorders, gastrointestinal diseases and cardiovascular etc. It is also attributed to the aromatic plants extracts likely biological properties of potential applications in modern medicine, food industry, perfumery and cosmetics: antiradical and anti-inflammatory powers¹.

Sorindeia grandifolia is an aromatic species of anacardiaceae family met in wooded savannas, dry forests, dense forests and humid regions of Western and Central Africa, situated in the North of the equator. This shrub, lianescent, bears leaves and supporting leaflets oblong and lanceolate, a pale green calyx, petal pink, white or pink flowers and fruits of ellipsoidal shape at maturity². In Benin, the decoction of *S. grandifolia* leaves, consumed in the fresh state, treats the cough while its juice possesses antipyretic properties³. To date, the literature has not reported at least one bioactive chemical compounds in this plant whose therapeutic performance in traditional medicine has always proven. This work has for objective the valorization of this species (*Sorindeia grandifolia* Engl., Anacardiaceae) from Benin by the extraction, the identification and the characterization of bioactive compounds it contains. The purpose of this study is to determine, through appropriate analytical methods, the physicochemical characteristics of this plant leaf extracts.

Material and Methods

Plant material and distillation of the volatile constituents: *Sorindeia grandifolia* leaves were collected in June and July 2007 in the classified forest of Lama (Benin). They were authenticated at the National Herbarium of the University of Abomey-Calavi. In the laboratory, these leaves are kept between 18 and 20°C in the shade of sunlight throughout the study period. Essential oil is extracted from leaves (450g) by the technique of hydrodistillation during five hours on Clevenger, according to the method used in british pharmacopoeia⁴. They was dried over anhydrous sodium sulfate and analyzed by GC/FID and GC/MS. For the determination of non-volatile compounds, powders were obtained from the leaves, dried in the dark for one month, by grinding with a knives machine Ika Werke MF 10 basic type. Vegetable powders collected are then sieved in the size grading 0.425.

Physical properties: Physical parameters of the essential oil of *S. grandifolia* leaves were determined using the methods described by AFNOR^{5,6}.

Density at 20°C: The density measure was carried out using a micro-pycnometer and a precision balance.

Refractive index at 20°C: The refractive index was determined by means of the refractometer CARL ZEISS JENA 234678.

Rotatory power at 20°C: The measurement was made by Carl Zeiss polarimeter 128291.

Phytochemical analysis: Analysis of the volatile constituents:

GC/FID: The extracts were analysed on a Hewlett-Packard gas chromatograph Model 6890, equipped with a DB5 MS column (30 m x 0.25 mm, 0.25 μ m), programming from 50°C (5 min) to 300°C at 5°C/min, 5 min hold. Hydrogen as carrier gas (1.0 mL/min); injection in split mode (1:60); injector and detector temperature: 280 and 300°C respectively. Each extract is diluted in hexane: 1/30.

GC/MS: The extracts compositions were analysed on a Hewlett-Packard gas chromatograph Model 5890, coupled to a Hewlett-Packard MS model 5871, equipped with a DB5 MS column (30 m x 0.25 mm, 0.25 μ m), programming from 50°C (5 min) to 300°C at 5°C/min, 5 min hold. Helium as carrier gas (1.0 mL/min); injection in split mode (1:30); injector and detector temperature, 250 and 280°C respectively. The MS working in electron impact mode at 70 eV; electron multiplier: 2500 eV; ion source temperature: 180°C; mass spectra data were acquired in the scan mode in *m/z* range 33-450.

The compounds assayed by GC in the different essential oils were identified by comparing their retention indices with those of reference compounds in the literature and confirmed by GC-MS by comparison of their mass spectra with those of reference substances^{7,8,9}.

Analysis of non-volatile constituents of leaves of *S. Grandifolia*:

The phytochemical screening was made according to the standard techniques described by Paris and Moysé¹⁰, Bouquet¹¹ and Debray *et al.*¹².

Coumarins: Infused at a 10% made from the plant powder was examined under the UV light (365 nm). The appearance of a bluish fluorescence indicated a positive reaction.

Tannins gallic: An aqueous infusion was prepared from 5 g of plant powder and 100 mL of boiling distilled water. After 15 min, the mixture was filtered. The residue was rinsed with hot water to bring the volume of the filtrate to 100 mL. 20 mL of the filtrate are saturated with sprayed sodium sulfate and then, it was added dropwise 1 mL of ferric chloride (1%). The development of a blue-black tint corresponded to the presence of gallic tannins, not precipitated by Stiasny's reagent.

Flavones: It was introduced in a test tube, 5 mL of infused. In this content, 5 mL of hydrochloric alcohol constituted by equal volumes of ethanol at 95°, distilled water, concentrated hydrochloric acid (37%) and 1 mL of isoamylic alcohol were added. In the presence of shavings magnesium, it emerged at the supernatant layer (layer isoamyl alcohol) a pink-orangy color indicating the presence of genins of flavonoids.

Leucoanthocyan: They were identified by introducing into a test tube 5 mL of infused (5%) and 5 mL of hydrochloric (ethanol 95° + distilled water + hydrochloric acid 37% of equal volumes). The mixture was compete with 1 mL of isoamylic

alcohol and then, heated to 90° through a water bath. After fifteen minutes, it had developed a red-cerise tint (or purple) indicating the presence of leucoanthocyan.

Saponins: A decoction was prepared during 30 min from two grams of plant powder and 100 mL of distilled water. After filtered the obtained mixture, the filtrate was divided into 10 different volumes (1 mL, 2 mL, 3 mL, 10 mL) in 10 calibrated tubes (internal diameter :1.3 cm). The content of each tube was adjusted to 10 mL with distilled water. After shaking each tube in a horizontal position for 15 seconds, followed by a rest of 15 min in an upright position, the height of the foam supernatant was measured in cm. When this height is close to 1 cm in the Xth tube, the foam index (I) is calculated by the following formula: $I = \text{Foam height (in cm) in the X}^{\text{th}} \text{ tube} \times 5/0.0X$. The presence of saponins in the plant is confirmed when the value of the foam index is greater than 100.

Fatty acids (FA) and unsaponifiable (Un): Extraction of lipids: 15.0 g of the vegetable material powder were twice extracted successively by 100 mL of petroleum ether (40-60°C) with magnetic stirring at room temperature. After filtration and evaporation of the solvent under reduced pressure, the extracts were dried and weighed. The yields were established in calculating the average of three extractions.

Saponification and fatty acids obtention: The saponification was conducted by refluxing, during thirty minutes past one o'clock, 0.5 g of plant extract and 25 mL of an ethanolic and potassium hydroxide solution (2N). After cooling, there was added 50 mL of water and the unsaponifiable matter are extracted by 3 x 50 mL of cyclohexane. The soap solution produced was then acidified to precipitate the FA ($5 \leq \text{pH} \leq 6$). The FA released were yet extracted by 3 x 50 mL of diethyl ether^{13,14}.

Methylation of FA: Fatty acids were converted to their methylic esters by addition a methanolic solution (10%) of boron trifluoride (BF₃) and the methylic esters were extracted with cyclohexane.

The analysis of FA and Un collected was made through GC/FID and GC/MS¹⁵.

Results and Discussion

The measurement of density, refractive index and rotatory power of the essential oils samples of *S. grandifolia* leaves collected at different dates to Kato (the Lama botanical forest reserve of Benin) gave the values included in the table 1.

Table-1
Physical characteristics of the essential oils of *Sorindeia grandifolia*

	Density (20°C)	Refractive index (20°C)	rotatory power (20°C)
L ₁	0.917	1.4935	-7.996
L ₂	0.921	1.4932	5.000
L ₁ = Leaves (07-06-07) ; L ₂ = Leaves (07-07-07)			

The values determined for the density and the rotatory power conversely varies to those of the refractive index after a month of harvest interval of L₁ and L₂. According to these values, there is a slight difference between the densities of essential oils extracted from *S. grandifolia* leaves. Between the values of refractive indexes, this difference is more weak. By cons, the difference between the values of the rotatory power is very pronounced. These differences noted could be bound to several factors, in particular, edaphic, climatic as well as the cultural practices^{16,17,18}. It is important to appreciate in the future such homogeneity over several months.

S. grandifolia is a little fragrant species. The yields in essential oil of the leaves of this plant are low (1.98x10⁻²% to 2.17x10⁻²%). The values of these yields are seventeen for eighteen times lower than the value of the yield in essential oil of *Diplolophium africanum* stem-foliage reported by Koudoro *et al.* in 2011¹⁹. On the other hand, the *S. grandifolia* leaves seem richer in volatile extracts contrary to those of *Anona senegalensis* (1.4x10⁻²%) investigated by Noudogbessi *et al.* in 2011²⁰. The volatile compounds identified in the essential oils collected by hydrodistillation of leaves are presented in table 2.

GC-FID and GC- MS analysis of *S. grandifolia* essential oil revealed 40 compounds in the sample L₁ and 46 in that of L₂ representing respectively 96.9 and 94.1% of the total weight of the essential oil (table 2). The main families volatile of compounds which marking L₁ and L₂ are the hydrogenated monoterpenes (31.3 to 57.9%), hydrogenated sesquiterpenes (15.3% to 41.5%) and oxygenated sesquiterpenes (11.7 to 13.6%). The proportions by oxygenated monoterpenes no neighborhood that 3.5% in L₁ and 2.3% in L₂. The main constituents (≥ 4%) susceptible to generate a bioactive character in L₁ are (Z)-β-ocimene (11.4%), (E)-β-ocimene (7.6%), γ-cadinene (7.0%), selin-11-en-4-α-ol (5.7%), palmitic acid (6.2%), α-selinene (5.5%), α-phellandrene (5.2%), β-selinene (4.9%) and β-elemene (4.5%). The major compounds (≥ 4%) of L₂ volatile extract were constituted of limonene (20.2%), (E)-β-ocimene (17.8%), β-pinene (5.5%), selin-11-en-4-α-ol (4.5%) and palmitic acid (4.3%). Further phytochemical investigations on various extracts realized with appropriate solvents allowed indicate the presence in *S. grandifolia* leaves of other metabolites, proven pharmacological properties²¹. This is coumarin, gallic tannins, flavones, leucoanthocyanins and saponins (table 3). The presence of saponins, assessed by calculating the foam index (83.33), would explain the plant high aphrodisiac power. It was also noticed traces of carotenoids, catechic tannins and free quinone.

The lipid content (specifically FA) obtained by quantitation after extraction with petroleum ether was estimated at 0.7% compared to the mass of plant material used.

FA identified by gas chromatography coupled with spectrometry mass are predominantly saturated (44.2%). As for the unsaturated FA, they represent only 15.9%. The major FA

identified in *S. grandifolia* leaves are arachidic acid (22.5%), palmitic acid (20.3%), linoleic acid (15.9%), followed by traces of myristic acid (1.4%). The investigations led on the unsaponifiables did not reveal an interesting chemical composition. Indeed the compounds constituting the footprints of Un collected could not be identified by GC/MS.

Conclusion

The chromatographic studies of *S. grandifolia* leaves extracts showed the presence of high proportions of monoterpenes and sesquiterpenes hydrogenated in essential oils and FA often found in the vegetable kingdom. The phytochemical screening, performed on the basis of specific tests allowed to characterize in *S. grandifolia* leaves: coumarins, gallic tannins, flavones and leucoanthocyanins. The phytochemical heritage of the *S. grandifolia* plant, omen, because it is referenced by traditional medicine in Benin, a useful contribution in the development of the vegetable biotechnology. These preliminary results will be used to support a rigorous exploration of pharmacological properties of the endemic species *S. grandifolia* of Benin.

References

1. Alitonou G.A., Huiles essentielles extraites de plantes aromatiques acclimatées au Bénin : étude chimique, évaluation biologique et applications potentielles, Thèse de Doctorat en cotutelle, UAC et Univ. Montpellier II, (2006)
2. Akoegninou A., Van Der Burg W.J., Van Der Maesen L.J.G. *Flore Analytique du Bénin*, (2006)
3. Adjanohoun E.J., Médecine traditionnelle et Pharmacopée, Contribution aux études ethnobotaniques et floristiques en République Populaire du Bénin, ACCT, Paris (1989)
4. British Pharmacopoeia, 11. P. A. HMSO: London, (1980)
5. AFNOR, NFT 75-006, Les huiles essentielles-vocabulaire-1ere liste, (1998)
6. AFNOR, Recueil de normes : les huiles essentielles, Tome 2, Monographies relatives aux huiles essentielles, Paris, 661-663 (2000)
7. Rösch P., Popp J., Kiefer W., Raman and SERS Investigations on Lamiaceae, *J. Mol. Struct.*, **121**, 480-481 (1999)
8. Adams R.P., Identification of essential oils by ion trap mass spectrometry, *Academy Press, Inc, New-York*, (1989)
9. Swigar A.A., Silverstein R.M., Monoterpènes, Infrared, Mass, NMR Spectra and Kovats Indices, *Aldrich Chem. Co. Milwaukee, WI, USA*, (1981)
10. Paris R., Moysse H., Précis de matière médicinale, Masson, Paris, (1969)
11. Bouquet M., Travaux et documents de l'Orstom, Paris, 13, (1971)

Table-2
Chemical compositions of essential oils from the leaves of *S. grandifolia*

Compounds	KI _{exp}	KI _{th}	(%)	
			L ₁	L ₂
<i>trans</i> -hex-2-enal	848	846	0.4	1.1
<i>trans</i> -hex-2-en-1-ol	858	854	-	0.6
hexan-1-ol	867	863	-	0.3
tricyclene	925	921	2.5	-
α -pinene	936	932	-	2.5
β-pinene	978	974	3.3	5.5
myrcene	991	988	0.5	1.2
meta-mentha-1(7),8-diene	1002	1000	-	0.3
α-phellandrene	1004	1002	5.2	2.7
p-cymene	1022	1020	-	3.2
o-cymene	1026	1022	-	0.5
limonene	1029	1024	0.4	20.2
(Z)-β-ocimene	1037	1032	11.4	0.4
(E)-β-ocimene	1046	1044	7.6	17.8
terpinolene	1090	1086	-	3.1
p-cymenene	1092	1089	-	0.5
linalool	1098	1095	1.7	0.4
p-cymen-8-ol	1184	1179	-	0.8
α -terpineol	1191	1186	1.4	1.1
geraniol	1254	1249	0.4	-
<i>trans</i> -verbenyl acetate	1295	1291	-	0.3
α -longipinene	1354	1350	1.3	-
α -ylangene	1377	1373	1.3	-
α -copaene	1379	1974	1.1	0.5
β-elemene	1394	1389	4.5	2.2
β -longipinene	1405	1400	1.8	0.2
β -caryophyllene	1421	1417	3.2	1.7
<i>cis</i> -thujopsene	1432	1429	1.3	0.3
β -copaene	1433	1430	-	0.2
selina-4(15),6-diene	1447	-	1.6	-
α -humulene	1454	1452	1.4	t
selina-4,11-diene	1471	-	2.0	1.1
γ -muurolene	1480	1478	0.7	0.5
β-selinene	1492	1489	4.9	3.6
α-selinene	1501	1498	5.5	2.7
α -bulnesene	1512	1509	1.2	0.5
γ-cadinene	1515	1513	7.0	1.1
δ -cadinene	1524	1522	1.5	0.7
<i>trans</i> -calamenene	1530	1528	1.2	-
spathulenol	1579	1577	0.5	0.5
caryophyllene oxide	1584	1582	1.2	1.1
globulol	1593	1590	-	0.3
humulene epoxide II	1611	1608	0.7	0.6
1,10-di-epi-cubenol	1622	1618	0.5	0.2
citronellyl pentanoate	1628	1624	-	0.3
1-epi-cubenol	1632	1627	-	0.2
epi- α -cubenol	1640	-	-	1.1
epi- α -muurolol	1645	1640	1.1	-

α -muurolol	1649	1644	0.5	-
cubenol	1651	1645	-	0.4
α -cadinol	1658	1652	1.9	1.7
selin-11-en-4-α-ol	1664	1658	5.7	4.5
β -sinensal	1704	1699	-	0.6
(2Z, 6E)-farnesol	1718	1722	1.0	0.5
iso-longifolol	1723	1728	0.5	-
palmitic acid	1967	1959	6.2	4.3
oleic acid	2148	2141	0.8	-
Monoterpene hydrocarbons			31.3	57.9
Oxygenated monoterpenes			3.5	2.3
Sesquiterpene hydrocarbons			41.5	15.3
Oxygenated sesquiterpenes			13.6	11.7
Total			96.9	94.1
L₁ = Leaves (07-06-07) ; L₂ = Leaves (07-07-07)				

Table-3
Chemical families identified in the leaves of *Sorindeia grandifolia*

Coumarin	Gallic tannins	Flavones	Leucoant hocyanes	Saponins (foam index)
+++	+++	+++	+++	++ (83.33)
+++ : Abundant ; ++ : Average				

12. Debray M., Jacquemin H., Razafindrambo R., Travaux et documents de l'Orstom, Paris, 8, (1971)
13. Joulain D., Konig W., the Atlas of Spectral Data of sesquiterpene hydrocarbons, E.B. – Verlag, Hamburg, Germany, (1998)
14. Wolff J.P., Manuel d'Analyse des Corps Gras, Azoulay, Paris, (1968)
15. Metcalfe L.D., Schmitz A.A., Rapid preparation of fatty acid esters for gas chromatography analysis, *Anal. Chem.*, **33**, 363-364 (1961)
16. Demarne F.E., Le géranium rosat, Parfums, Cosmétiques et Arômes, n°62 (1985)
17. Gilly G., Les plantes à parfum et huiles essentielles à Grasse, L'Harmattan, Paris 11-19 (1997)
18. Lis-Balchin M., Geranium and pelargonium: the genera *Geranium* and *Pelargonium*, CRC Press, Taylor & Francis, London, 116-131, 147-165, 184-217, (2002)
19. Koudoro Y.A., Alitonou G.A., Sossou D.J., Yehouenou B., Avlessi F., Menut C., Sohounhloué D., Chemical Composition and biological Activities of Essential Oil from Benin *Diplolophium africanum* Turcz stem leaves, *J. Soc. Ouest-Afr. Chim.*, 032; 1-8 (2011)
20. Noudogbessi J.P., Natta A.K., Avlessi F., Sohounhloué D.C.K., Figueredo G., Chalchat J.C., Chemical Composition of the Essential Oils Extracted from Two Annonaceae Required in Beninese Pharmacopeia, *Australian Journal of Basic and Applied Sciences*, **5(2)**, 34-40 (2011)
21. Bruneton J., Pharmacognosie, Phytochimie, Plantes médicinales, éd. 3, Techniques et Documentation, Lavoisier, Paris, (1999)