



HAL
open science

Differential adaptation to multi-stressed conditions of wine fermentation revealed by variations in yeast regulatory networks

Christian C. Brion, Chloé C. Ambroset, Isabelle Sanchez, Jean-Luc Legras,
Bruno B. Blondin

► To cite this version:

Christian C. Brion, Chloé C. Ambroset, Isabelle Sanchez, Jean-Luc Legras, Bruno B. Blondin. Differential adaptation to multi-stressed conditions of wine fermentation revealed by variations in yeast regulatory networks. *BMC Genomics*, 2013, 14, pp.1-21. 10.1186/1471-2164-14-681 . hal-00939593

HAL Id: hal-00939593

<https://hal.science/hal-00939593>

Submitted on 30 Jan 2014

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

RESEARCH ARTICLE

Open Access

Differential adaptation to multi-stressed conditions of wine fermentation revealed by variations in yeast regulatory networks

Christian Brion^{1,2,3}, Chloé Ambroset^{1,2,3}, Isabelle Sanchez^{1,2,3}, Jean-Luc Legras^{1,2,3} and Bruno Blondin^{1,2,3*}

Abstract

Background: Variation of gene expression can lead to phenotypic variation and have therefore been assumed to contribute the diversity of wine yeast (*Saccharomyces cerevisiae*) properties. However, the molecular bases of this variation of gene expression are unknown. We addressed these questions by carrying out an integrated genetical-genomic study in fermentation conditions. We report here quantitative trait loci (QTL) mapping based on expression profiling in a segregating population generated by a cross between a derivative of the popular wine strain EC1118 and the laboratory strain S288c.

Results: Most of the fermentation traits studied appeared to be under multi-allelic control. We mapped five phenotypic QTLs and 1465 expression QTLs. Several expression QTLs overlapped in hotspots. Among the linkages unraveled here, several were associated with metabolic processes essential for wine fermentation such as glucose sensing or nitrogen and vitamin metabolism. Variations affecting the regulation of drug detoxification and export (*TPO1*, *PDR12* or *QDR2*) were linked to variation in four genes encoding transcription factors (*PDR8*, *WAR1*, *YRR1* and *HAP1*). We demonstrated that the allelic variation of *WAR1* and *TPO1* affected sorbic and octanoic acid resistance, respectively. Moreover, analysis of the transcription factors phylogeny suggests they evolved with a specific adaptation of the strains to wine fermentation conditions. Unexpectedly, we found that the variation of fermentation rates was associated with a partial disomy of chromosome 16. This disomy resulted from the well known 8–16 translocation.

Conclusions: This large data set made it possible to decipher the effects of genetic variation on gene expression during fermentation and certain wine fermentation properties. Our findings shed a new light on the adaptation mechanisms required by yeast to cope with the multiple stresses generated by wine fermentation. In this context, the detoxification and export systems appear to be of particular importance, probably due to nitrogen starvation. Furthermore, we show that the well characterized 8–16 translocation located in *SSU1*, which is associated with sulfite resistance, can lead to a partial chromosomal amplification in the progeny of strains that carry it, greatly improving fermentation kinetics. This amplification has been detected among other wine yeasts.

Keywords: Wine yeast, Fermentation, QTL, Transcriptome, Partial disomy, Detoxification

* Correspondence: blondin@supagro.inra.fr

¹INRA, UMR1083 Science pour l'Œnologie, 2 Place Viala, Montpellier F-34060, France

²Montpellier SupAgro, UMR1083 Science pour l'Œnologie, 2 Place Viala, Montpellier F-34060, France

Full list of author information is available at the end of the article

Background

Since the development of wine-making, wine yeasts have undergone a specific pattern of evolution and have become highly effective in the fermentation of grape juice [1]. They are able to withstand various stresses, such as low pH or high levels of ethanol. These yeasts are also resistant to inhibitors, such as sulfites [2] and medium-chain fatty acids [3], and are able to continue fermentation for long periods of nutrients starvation. In addition, commercial wine yeasts make a specific contribution to the aroma bouquet of the wine. Knowledge of the genomic bases of these specific features is a prerequisite for understanding the mechanisms underlying adaptation [1,4,5]. Such knowledge would also provide a basis for the improvement of industrial wine yeast strains [6]. Various studies have enhanced our understanding of genomic structural variation in wine yeast strains. Genome sequencing has led to the identification of genomic rearrangements and mutations specific to wine yeasts (reviewed in [7-9]). One of the best examples is the translocation between chromosomes 8 and 16 described by Pérez-Ortín *et al.* [10] leading to overexpression of the sulfite exporter gene *SSUI* and to the resistance of higher levels of sulfite. Dunn *et al.* [11,12] have shown, using comparative genomic hybridization (aCGH), that copy-number variations for several groups of genes (involved in drug responses and ion sensors) are also characteristic of wine strains. Based on an analysis of the wine yeast EC1118 sequence, Novo *et al.* [13] showed that three loci widely conserved in wine strains had been gained by horizontal transfer events from non *Saccharomyces* yeast [14]. Comparison of sequenced genomes has suggested that nucleotide polymorphisms are the major source of phenotypic diversity [15,16]. However the relationships between such genetic variations and phenotypic diversity remain unclear, particularly in the context of alcoholic fermentation.

Quantitative trait locus (QTL)-based approaches are widely used in yeast, to link allelic variations to phenotypic diversity [17-20]. This approach has been applied to wine strains to identify the origin of fermentation traits. Marullo *et al.* [21] used this approach to show that mutations of the asparaginase gene *ASPI* underlie differences in acetic acid production between two wine strains. QTL-based approaches have proved efficient for identifying the traits associated with a single mutation, but most fermentation traits are under complex polygenic control which is much more difficult to resolve. Using new approaches based on the analysis of very large populations of segregants; Extreme QTL mapping (X-QTL); Ehrenreich *et al.* [22,23] demonstrated the feasibility of identifying the genetic determinants of complex phenotypes, such as drug resistance.

Trait variation in may also results from changes in gene expression of the underlying regulatory networks.

Brem *et al.* [24] have demonstrated that regulatory variations could be linked to genomic alterations by QTL approaches. Searches for expression QTLs (eQTLs) are now widely used in yeast to decipher regulatory networks variations [25-29], reviewed in [30,31].

Such approaches can be used to elucidate the origin of regulatory network variations and their impact on fermentation traits, such as the fermentation duration, the nitrogen consumption and the metabolite production. We previously generated a segregating population by crossing the lab strain S288c and a spore derivative of the wine yeast EC1118, for eQTL linkage analysis [4]. One of the main results of this previous study was the detection of a large impact of the *ABZI* (encoding para-amino benzoate (PABA) synthase) allele on nitrogen assimilation, resulting in large variations in the fermentation rate [4]. Furthermore, a recent QTL study using this lineage implicated *ABZI* allele variation in aromatic compounds production [32] probably due to the involvement of this gene in amino-acid metabolisms.

We report here of the results of a new search for fermentation traits QTL and eQTL with an enlarged segregant population in the late stages of alcoholic fermentation, more relevant conditions for addressing the stress response. The use of this approach provided us with a broad view of expression variation in alcoholic fermentation and allowed us to identify the genetic origin of variation for several regulatory networks involved in key processes, such as detoxification and sulfate assimilation. An unexpected result of this study was the finding that fermentation rate was controlled by a partial disomy of chromosome 16, revealing a new role in wine yeasts for a well known translocation.

Results

Phenotypics and transcriptomic analysis of the 59A×S288c lineage

We phenotyped 44 segregants obtained from a cross between the laboratory strain S288c and the wine yeast derivative 59A (as described in Methods). Fermentations were performed in a synthetic medium simulating a grape must (SM425) and containing para-amino benzoate (PABA) to counteract the effect of the *ABZI* allele [4]. Our analysis were performed in more stringent conditions than that of Ambroset *et al.* [4]: the amounts of ergosterol and oleic acid used were half those generally used (the final concentrations were 7.5 mg/l for ergosterol and 2.5 µl/l for oleic acid) and fermentations were performed at 24°C. We estimated the kinetic properties regarding the lag time and the fermentation rate at 2 stages of the fermentation: Rmax (maximal fermentation rate) and R70 (fermentation rate at 70% of fermentation). The fermentation kinetics of parental strains and of several segregants are shown in Figure 1. Population

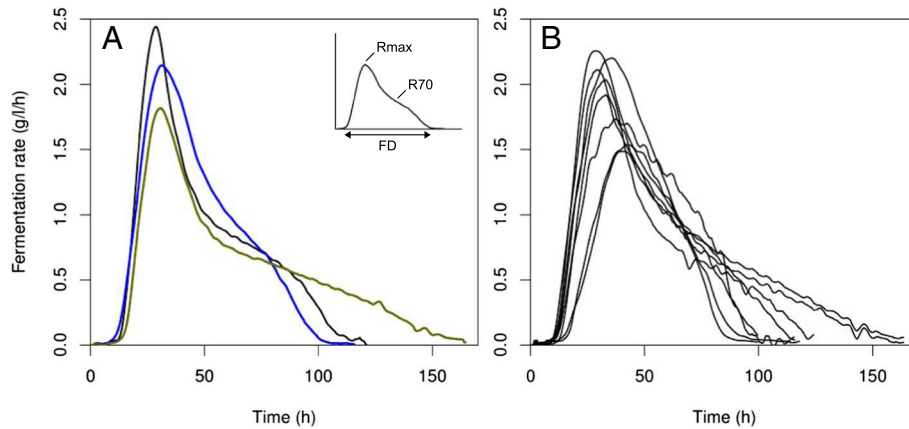


Figure 1 Fermentation profiles for parental strains and for segregants. **(A)** Fermentation profile for the two parental strains and for 59A x S288c: 59A in blue, S288c in green, 59A x S288c in black. **(B)** Fermentation profile for 8 segregants. Sugar was completely consumed for all segregants.

size and metabolite quantities were determined at the end of the fermentation.

We first checked that phenotypes affected by *ABZI* allele, such as R_{max} , were corrected by the addition of PABA and independent of the allelic form (data not shown). Most of the phenotypes had a high heritability (80% to 97%), indicating that genetic variations had a major impact on overall variations. The dry weight was the only phenotype displaying a low heritability (>50%) and was disregarded. R_{70} values followed a bimodal distribution, suggesting that the phenotype is mainly controlled by one locus. The other phenotypes, such as R_{max} and the amounts of metabolites followed continuous distributions, indicating a probable polygenic control (Additional file 1).

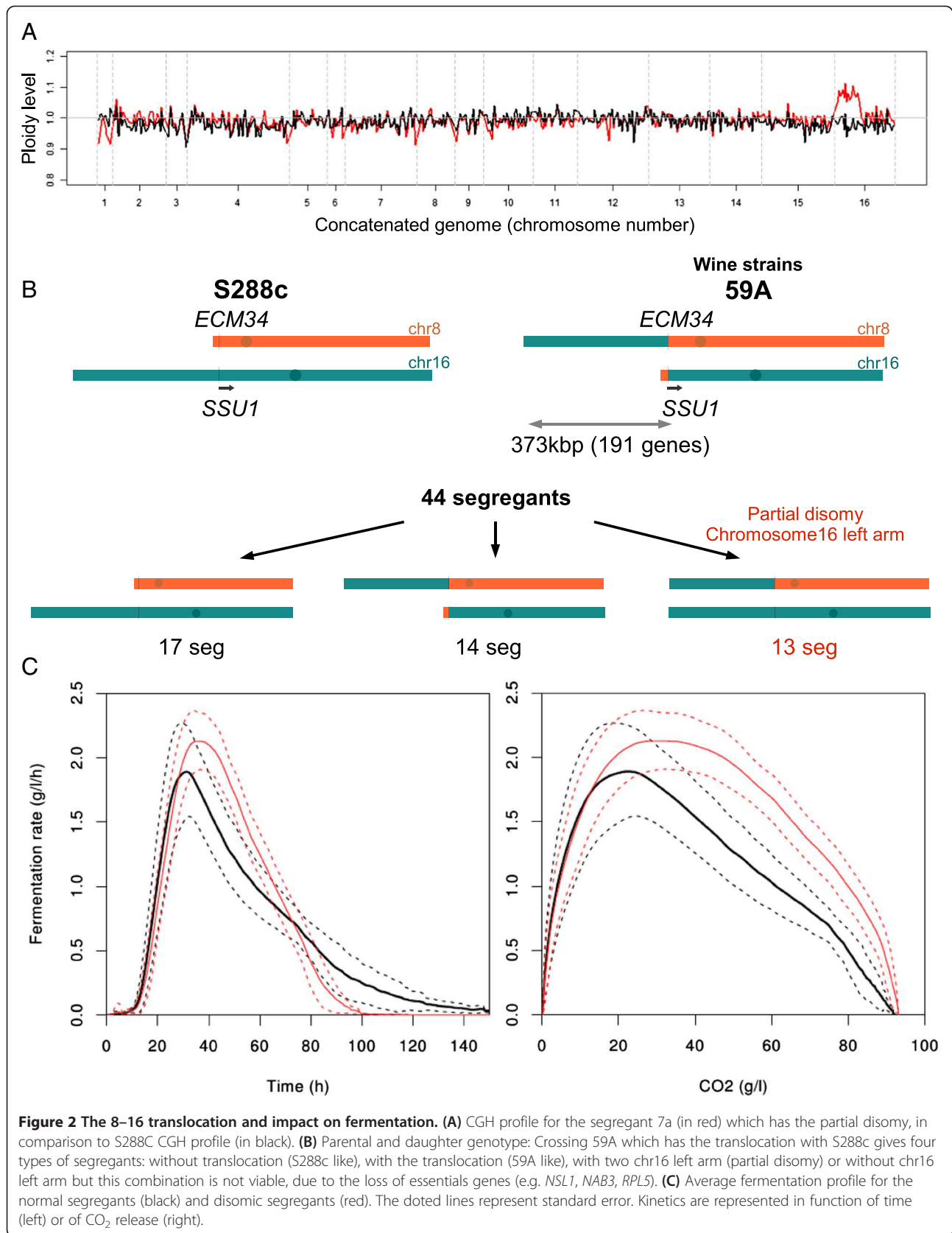
Transcriptome profiles were obtained at 70% of fermentation progress (66 g/l CO_2 released), corresponding to late stationary phase, 20 to 40 hours after the end of the growth phase (depending on the segregants). At this point in the fermentation, the yeast is subject to nutrient starvation and ethanol stress (8% v/v). We compared RNA abundance between segregants by Agilent mono-color labeling and hybridization on oligonucleotides microarrays as described in Methods. We identified 1610 genes differentially expressed between 59A and S288c with a \log_2 of fold-change (LogFC) of more than 0.7 in either direction ($\text{adj}P_v < 0.01$).

We assessed the correlation between the fermentation rate (R_{70}) and gene expression and found large sets of genes positively correlated (347 genes involved in cell trafficking and nitrogen reutilization; Funspec [33]) and negatively correlated (351 genes involved in oxidative stress and respiration; Funspec).

Clustering analysis revealed a partial disomy of chromosome 16 affecting genes expression

A clustering analysis of the whole gene expression dataset revealed significant associations (Pearson correlation coefficient ≥ 0.84) for 887 genes (Additional file 2). Most of the gene clusters obtained were enriched in functional classes, such as mating, mitochondrial translation, ribosomal proteins, ergosterol synthesis and sulfur assimilation consistent with the known coordination of expression of these genes. Some clusters also corresponded to genes correlated with phenotypes (R_{70} , R_{max}). Other clusters included physically associated genes (ASP cluster, regions A B and C [13], telomeric genes).

We identified a cluster of 37 genes displaying a higher expression in 13 segregants. These genes were physically linked and all were located in the first 373 kbp at the start of chromosome 16 (Additional file 2, cluster “chr16 left-arm genes”). A careful mining of the CGH data (from the Affymetrix chip signals) revealed that these 13 segregants carried a 373 kbp duplicated region on the left arm of chromosome 16 (Figure 2A). No other chromosomal aberration was observed. This partial disomy resulted from the translocation of an arm of chromosome 16 onto the chromosome 8 originating from the wine yeast strain, in addition to the presence of a standard chromosome 16 (Figure 2B). This 8–16 translocation is known to be responsible for higher level of sulfite resistance and was found in many wine yeasts [10]. We examined the impact of this partial disomy on transcriptome profiles by comparing the expression data for the 13 disomic segregants with those for the other 31 segregants. As expected, the genes of the duplicated region were more strongly expressed in segregants carrying the



disomic region (155 of the 191 genes in the area concerned, $\text{adj}P_v < 0.05$), consistent with the doubling of gene copy number. We also observed differences in expression for 1305 other genes ($\text{adj}P_v < 0.01$) not located in this area. The upregulated genes include a high proportion of genes involved in methionine biosynthesis (17 genes, $P_v = 10^{-9}$ -FunSpec [33]) whereas the down-regulated genes included a high proportion of genes involved in the oxidative stress response (20 genes, $P_v = 10^{-6}$ -FunSpec) and aerobic respiration (37 genes, $P_v = 10^{-14}$ -FunSpec). The pattern of functional enrichment of the deregulated genes suggests that disomic strains are less affected by stresses of alcoholic fermentation conditions. Consistent with this hypothesis, we observed that the partial disomy had a major impact on fermentation kinetics (Figure 2C). Partial disomy was associated with significantly higher fermentation rates, a shorter duration of fermentation (t -test $P_v < 10^{-6}$) and an absence of sluggish fermentation profiles. Levels of acetic acid production were also lower for the disomic segregants (data not shown, t -test $P_v < 0.01$), supporting our hypothesis of lower levels of stress in these clones.

Phenotypic and expression QTL linkages

We generated a new high-density genetic map (1.81 markers/10 kbp Additional file 3) from the Affymetrix S98 microarrays genotyping data for the 44 segregants. Linkage analysis was performed by the interval mapping method, for QTLs and eQTLs.

Three phenotypic QTLs were identified with a false discovery rate (FDR) threshold of 0.05: one QTL explaining flocculation on chromosome 1 encompassing *FLO1*, another explaining clumpiness and encompassing *AMN1* and a third, explaining fermentation rate (R70), on the left arm of the chromosome 16 (140 kbp), overlapping with the duplicated region and clearly triggered by the partial disomy. Limiting the population to the 31 segregants with no partial duplication of chromosome 16 eliminated the detection of the R70 QTL on chromosome 16. It also led to the detection of three QTLs for metabolites levels, mapping to chromosome 2 (208 kbp) for pyruvic acid and chromosome 14 (460 kbp) for succinic acid and glycerol.

Genome wide linkage analysis of gene expression led to the detection of 1063 eQTLs as summarized in Table 1. Dividing the FDR by two only slightly reduce the number of eQTLs, demonstrating the robustness of the data. A high amount (56%) of local-eQTLs (or cis-eQTLs, see Methods) were detected with higher average LOD than trans-eQTLs (Table 1). Some of the genes differentially expressed between the parents displaying cis-eQTLs corresponded to genes absent from one of the parental strains. As expected, genes from the three new regions acquired by horizontal transfer in EC1118 [13] and conserved in 59A formed

Table 1 Summary of eQTLs detected

LODscore threshold	4	4.4	4 (without absent genes)
trans-eQTL	468 (5.31*)	315	443
cis-eQTL	595 (8.78*)	508	524
Total	1063	823	967
FDR	0.10	0.05	0.10

* Average LODscore.

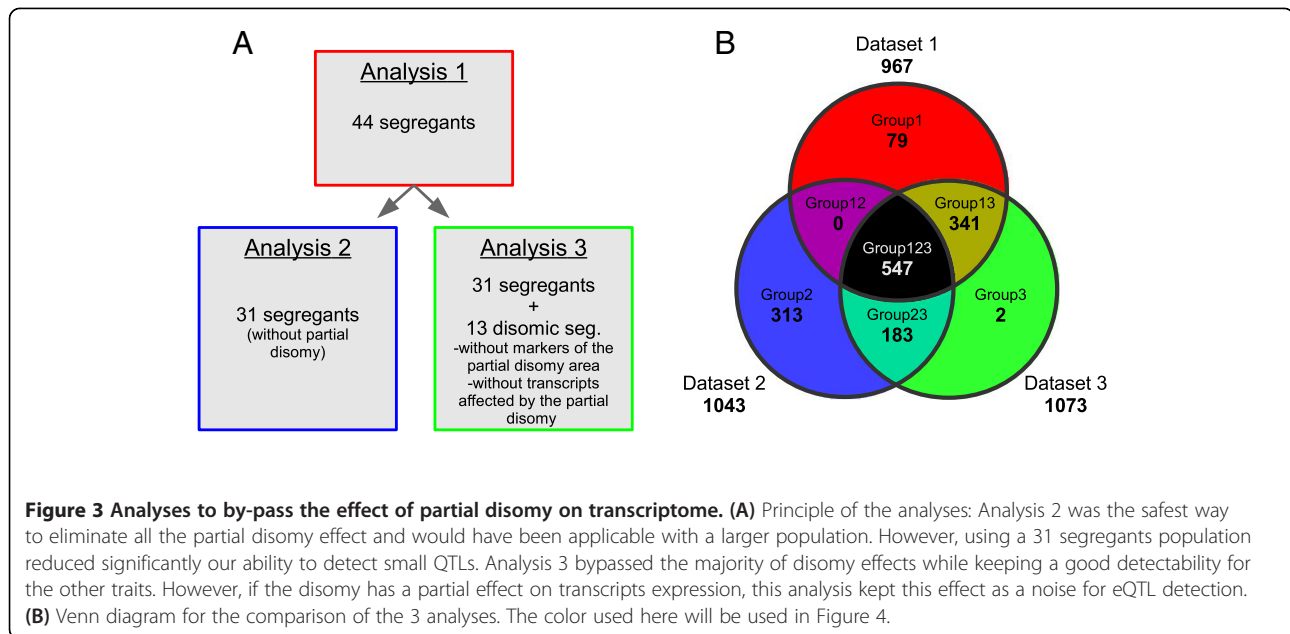
three clusters (Additional file 2), each controlled in cis-regulations. We did not consider these genes, nor the 111 genes missing from EC1118 [13] for subsequent analyses, reducing the number of eQTLs to 967.

We found that 113 of the 1460 genes affected by the partial disomy displayed eQTLs, and 48 of these eQTLs mapped to the deleted or duplicated regions on chromosomes 8 and 16. Given that the partial disomy could affect the eQTL data, we adapted the analysis (analysis 1) to take this effect into account. Two new eQTL linkage analyses were performed as described in Figure 3A: (a) a search with a population limited to 31 non disomic segregants (analysis 2) (b) a filtering of the data for the 13 disomic segregants, eliminating the markers in the duplicated region and ignoring the 1460 genes affected by the partial disomy (analysis 3).

These two new data sets were compared with the first one on a Venn diagram Figure 3B and in Additional file 4. Most of the eQTLs (547) were common to all the three data sets (group 123 on Figure 3B) and corresponded to eQTLs with high LOD-score (cis-eQTLs). In analysis 2, restricted to the 31 non disomic segregants, the detection power was lower: at lod4, the FDR was 0.13. Data set 3 overlapped strongly with data set 1, with 888 of 967 eQTLs common to these two data sets. This suggests that the partial disomy slightly decreased our ability to detect true eQTLs, but had only a limited effect on the robustness of the data. The eQTL analysis was performed for the union of the three data sets (1465 eQTLs), taking the "Venn" group for each eQTL into account. We, therefore, concluded that we could overcome the effect of the partial disomy.

Variations in expression networks are triggered by few loci resulting in the formation of hotspots

The locations of the phenotypic QTLs and eQTLs are summarized in Figure 4 and Table 2. Many eQTLs overlapped in several hotspots and most were common to the three analyses. However, the disomy was responsible for two hotspots on chromosomes 8 and 16 (hotspot 6 and hotspot 10, in red Figure 4B). Furthermore, 113 of the eQTLs discovered in the two new analyses are overlapping in two hotspots on chromosome 2, at 208 kbp and 283 kbp (hotspots 1 and 2 in blue/green Figure 4A-4B). Due to the genetic similarities between EC1118 and



RM11-1a, some phenotypic QTLs and eQTLs are identical to those previously identified for the cross between BY4716 and RM11-1a (BxR population: hotspot 3, 4, 8 and 9, [25,34,35]).

However, we also found some linkages not previously identified. Hotspot 5 on chromosome 4 was found to be enriched in genes involved in thiamine biosynthesis pathway and is probably controlled by variations of the *THI3* regulator gene located in the hotspot area. Hotspot 7 on chromosome 10 brings together 18 genes with various functions (mitochondria, glucose sensing: *SNF3*, *YDL199c*, *GRR1*) and *GRR1* seems to be the most relevant candidate gene for the control of this hotspot. It is indeed involved in glucose repression and amino acid sensing [36].

We also found variations in several networks involving fewer genes but potentially involved in the wine-making process. The high-affinity sulfate permease gene, *SUL2*, has a cis-eQTL (LOD 4.5) overlapping with trans-eQTLs for three other genes of the sulfur pathway *MET2*, *MET5* and *MET32*. *SUL2* has two SNPs in its promoter (but none in the consensus sequences for transcription factors binding sites, Yeastract [37]) and five non-synonymous SNPs in its coding sequence. Changes in sulfate transport efficiency are clearly responsible for the higher expression level of the three other genes involved in sulfur amino acid metabolism (Figure 5).

Similarly, we observed a perturbation of the nitrogen uptake system that might have a significant impact on wine yeast performance [38]. The membrane peptide transporter gene, *PTR2*, and the arginine/alanine aminopeptidase gene, *AAP1*, both have eQTLs mapping to *CUP9* (chromosome 16) which encodes the known transcriptional repressor of *PTR2*. *CUP9* has two non-synonymous

SNPs in its coding sequence and the 59A allele clearly increases the expression of *PTR2* and, probably, also that of *AAP1* (Figure 6). Byrd *et al.* [39] described the activation of *PTR2* by ubiquitin-dependent Cup9p degradation in the presence of peptide [40]. However, in the 59A strain, *PTR2* has a frame-shift in its sequence and is, therefore, probably non-functional [41]. The involvement of *CUP9* in the control of *AAP1* is consistent with a broader role of this regulator in peptide metabolism. The synthetic must used here did not contain peptides, but these molecules may have been generated by endogenous nitrogen metabolism. The mutation in 59A may act by modulating the repressive activity or degradation of Cup9p.

Finally, a flocculation QTL was found on chromosome 1, mapping to *FLO1* as previously described [24] (Figure 4C). Flocculation phenotype was found linked to *FLO1* expression level (Additional file 6). Surprisingly, multiple QTL searches on the basis of flocculation phenotype did yield a locus containing *FLO8*. However, a second QTL was found on chromosome 1, in a 45 kbp region (Additional file 6). This region contains the gene *OAF1* involved in fatty-acid and peroxisome biogenesis. It is possible that *OAF1* mutations triggered variation of cell surface hydrophobicity that could impact flocculation [42].

Variations in detoxification mechanisms are triggered by mutations in transcription factor genes

In this linkage analysis, we detected changes in the detoxification network. Several membrane transporters involved in drug export displayed eQTLs (Table 3). Some of these genes were under local regulation (cis-eQTL): *SVS1*, required for resistance to vanadate, the two polyamine exporter, *TPO1* and *TPO2*, and the ion transporters *ALR2*

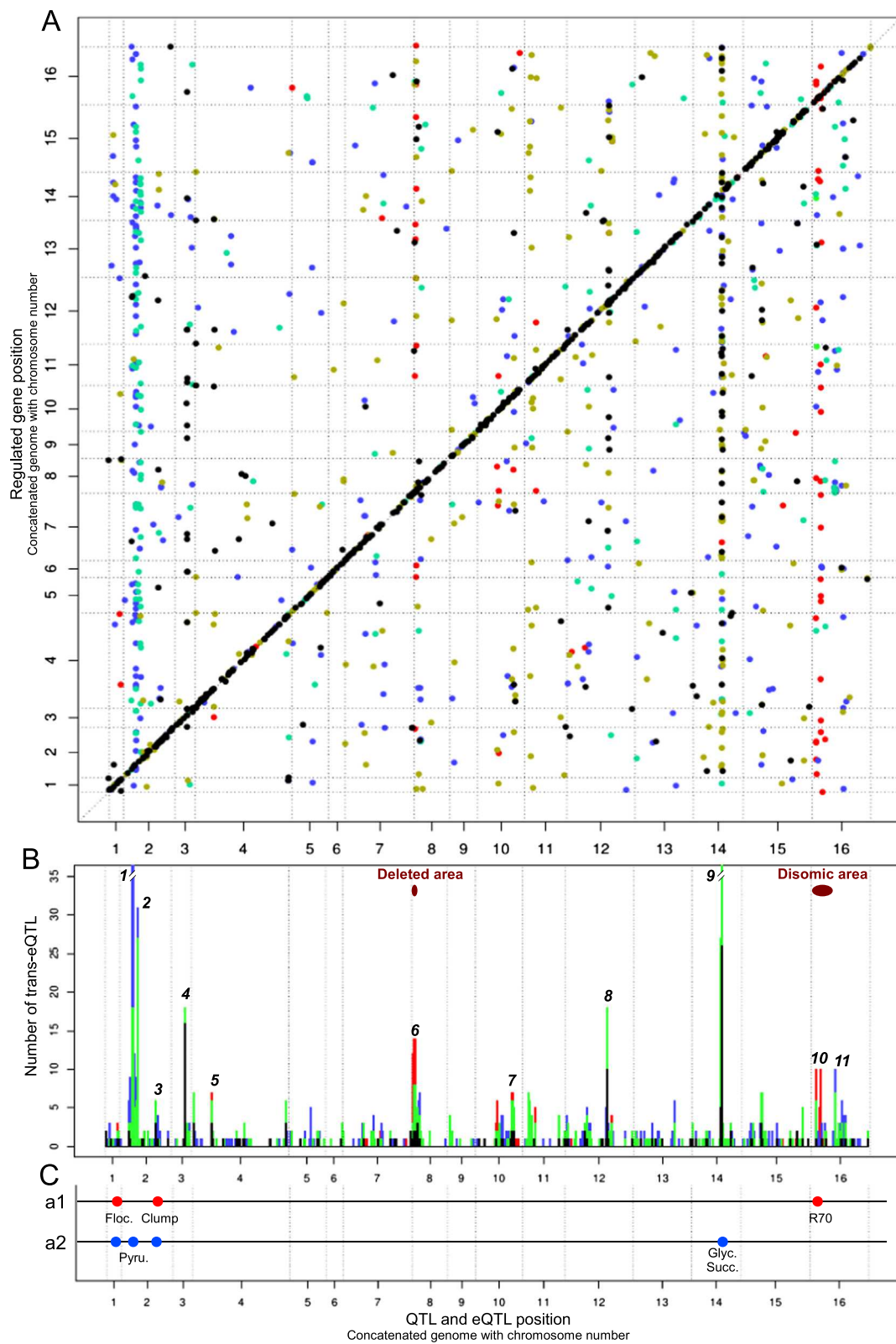


Figure 4 (See legend on next page.)

(See figure on previous page.)

Figure 4 QTL and eQTL localization. (A) Localization of the 1465 eQTLs in function of the corresponding transcript position. Colors correspond to the group in the Venn diagram Figure 3B. Spot in the first diagonal correspond to cis-eQTLs. (B) Trans-eQTL density and formation of hotspot. Descriptions of each hotspot are in Table 2. Black peaks correspond to the trans eQTL in common of the 3 analyses, Green peaks correspond to trans-eQTL detected at least in analysis 3. Blue peaks correspond trans-eQTL specific of analysis 2. Red peaks correspond trans-eQTL specific of analysis 1. (C) Phenotypic QTLs detected with 44 segregants (a1, red) and with 31 segregants (a2, blue).

and *SSU1*. The cis-eQTL of *SSU1* is due to the 8–16 translocation. Other detoxification system genes (e.g. *SNG1*, *PDR12*, *QDR2*) displayed distant control. Several eQTLs associated variations in the expression of these transporters with variations of zinc-finger transcription factor genes: *HAP1*, *PDR8*, *YRR1* and *WARI*.

The *TPO1* gene, which displays a cis-eQTL, has three SNPs in its promoter area and three non synonymous SNPs in its coding sequence. The S288c allele of *TPO1* is more strongly expressed than the wine yeast allele. As *TPO1* is involved in octanoic acid resistance [3], we investigated the effect of variations of *TPO1* expression on octanoic acid resistance. We measured population growth in the synthetic must medium (pH3.3) supplemented with octanoic acid (0.2 mM). Octanoic acid resistance dependent principally on the *TPO1* allele (*t*-test

$P < 0.005$, Figure 7) but was also weakly correlated with *TPO1* expression level (data not shown). Our data therefore suggest that the form of *TPO1* encoded by the S288c allele is more effective, conferring a higher octanoic acid resistance.

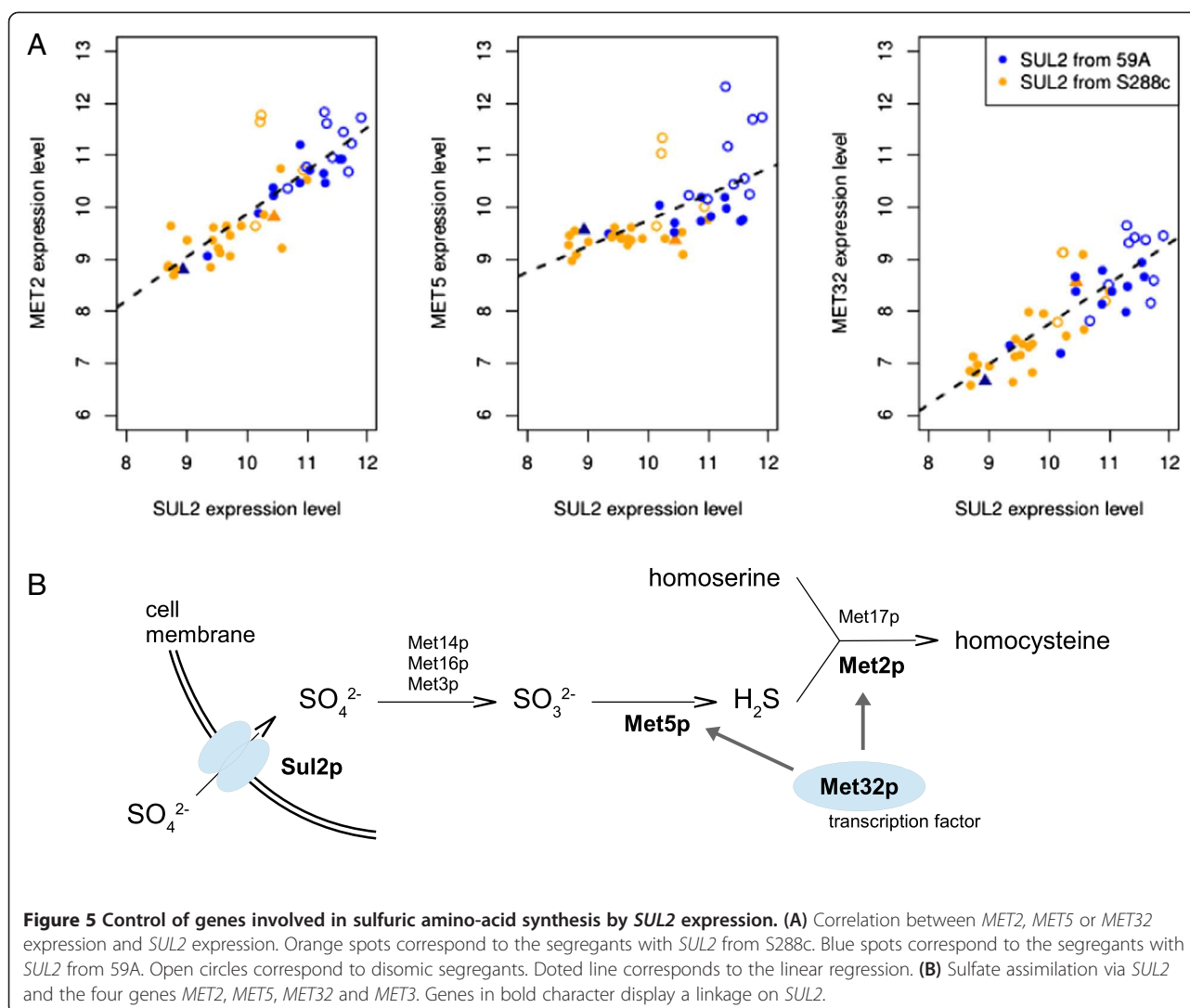
PDR11, *TPO4* and *QDR2* all had an eQTL in hotspot 8 mapping to *HAP1* (Table 3). Another gene, *PDR8*, encoding a pleiotropic drug resistance transcription factor lies in the same locus, 25 kbp away from *HAP1*, and may control some of these genes (Additional file 5). Consistent with this hypothesis, Steyer *et al.* [32], recently showed that high nerolidol production were associated with the S288c allele of *PDR8*. They demonstrated that *QDR2* regulation by *PDR8* accounted the variation of nerolidol release into the medium. We assessed the impact of *PDR8* allelic variation on gene expression by

Table 2 Hotspot description

Hotspot	Localization ^a	Number of eQTL ^b	Functional enrichment	Phenotypic QTL	Known or candidate gene regulator	Functional validation	Comments
1	chr2:207	66	No functional enrichment	Pyruvate	<i>SCT1</i>	Not realized	Hidden by the disomy, Sct1p is involved in lipid desaturation and diverges in the two strains by the size of a glutamic acid repeated sequence
2	chr2:277	47	No functional enrichment	-	Unknow	-	Hidden by the disomy
3	chr2:560	7	Daughter cell separation	Clumpness, OD, cell pop.	<i>AMN1</i>	Yvert et al. 2003 [34]	Impacting on cellular population estimation
4	chr3:198	20	Mating, pheromone	-	<i>MAT</i>	Brem et al. 2005 [35]	
5	chr4:309	9	Thiamine synthesis	-	<i>THI3</i>	Not realized	<i>THI3</i> bears two non-synonymous SNPs and a frame-shift in its coding sequence
6	chr8:20	27	Helicase	-	-	-	Disomy consequence
7	chr10:590	18	Mitochondrial protein	-	<i>GRR1</i>	Not realized	<i>GRR1</i> has a strong cis-eQTL (LOD = 13.4) and several alteration of its coding sequence
8	chr12:667	33	Fatty acid metabolism, detoxification	-	<i>HAP1/PDR8</i>	Yvert et al. 2003 [34]	Genes involved in the response to oxygen (nine targets of HAP1, Additional file 5)
9	chr14:456	101	Mitochondrial protein	Succinate, Glycerol	<i>MKT1</i>	Smith and Kruglyak 2008 [25]	Perturbation of the mitochondrial function affect the production of metabolites
10	chr16 left-arm	50	No functional enrichment	R70	-	-	Disomy consequence
11	chr16:370	17	chr8 translocated gene	-	<i>SSU1</i> translocation	Not realized	Translocation consequence

^aLocalization of the center of hotspot given in chromosome:kilo base pair.

^bNumber of total eQTL from the union of the 3 datasets.

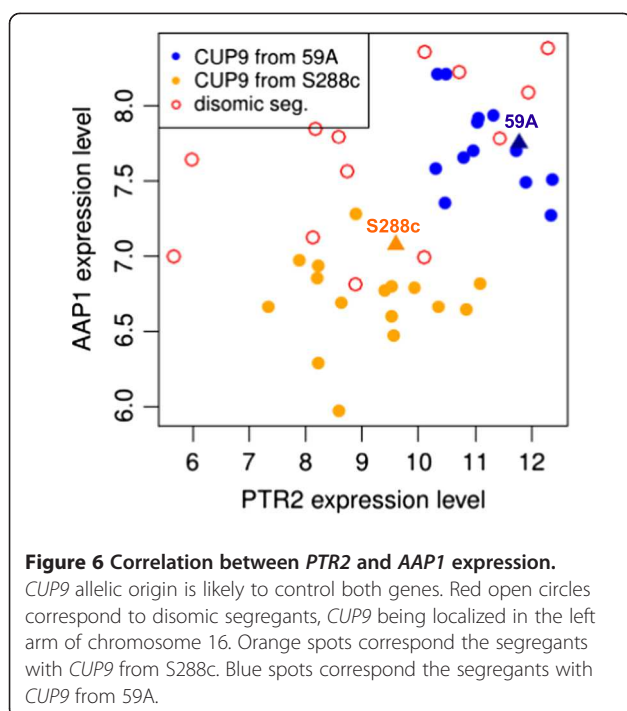


performing an allelic switch for *PDR8* and constructed 59A strain expressing the *PDR8* allele from S288c (59A *PDR8*-S288c). We compared the transcriptome of this strain with that of wild type 59A in fermentation conditions (the same conditions as for the global analysis). We detected variation of expression for very few genes ($\text{adj}P_v < 0.01$, Table 4). *QDR2* was the gene most strongly upregulated by allelic replacement, with a LogFC of 2.24, consistent with the results of Steyer *et al.* [32]. Among the other targets of *PDR8*, we only found *YLR179c* controlled by the allelic form of *PDR8*. Both *QDR2* and *YLR179c* had an eQTL mapping to *PDR8*. In addition to a slight down regulation of *PDR8* itself, we identified three other genes (*YLR149c*, *GIT1* and *QDR1*) as upregulated in the constructed strain and potentially playing a role in detoxification. These genes have not previously been described as *PDR8*-controlled. We did not detect the targets of *PDR8* reported by Hikkel *et al.* [43]. This was probably due to the specific conditions of

alcoholic stress and nutrient starvation used in our study. Furthermore, we performed an allelic switch with a smaller impact than the over-expression of *PDR8* used in other studies. Finally, only *QDR2* and *YLR179c* were controlled by *PDR8* in hotspot 8, suggesting that expressions of the other genes are triggered by *HAP1*.

The gene paralog of *PDR8*, *YRR1*, displays a cis-eQTL on chromosome 15 and has four SNPs in its promoter region and six non-synonymous SNPs in its coding sequence. *SNG1*, one of the targets of *YRR1* responsible for nitrosoguanidine resistance [44], has an eQTL mapping to the position of *YRR1*. Surprisingly, we found no linkages of this locus to other *YRR1* target genes. Moreover, the inverse patterns of parental behavior and locus segregation for *SNG1* expression indicates that other loci are involved in *SNG1* control (Figure 8).

The *PDR12* gene, encoding a plasma membrane ABC transporter responsible for organic acid efflux, had an eQTL mapping to its known transcription factor *WAR1*



(chromosome 13), an activator of weak acid stress response. The *WARI* allele of 59A has five non synonymous SNPs in its coding sequence and was associated with higher levels of *PDR12* expression level (Figure 9A). In addition, *WARI* was itself one of the genes for which expression was affected by *HAP1* variation (hotspot 8). The level of *WARI* expression had no significant impact on *PDR12* transcript abundance. Instead *PDR12* expression levels were influenced by the type of *WARI* allele (Figure 9B). *PDR12* was the only known *WARI*-dependent gene with an eQTL mapping at *WARI* (even with a LODscore below the threshold) [45]. As *PDR12* has been implicated in sorbic acid resistance [46], we investigated whether variation of the expression of this gene modulated resistance to this acid. We assessed the growth of segregants in the synthetic medium (pH3.3) supplemented with sorbic acid (0.5 mM, see Methods). Sorbic acid resistance appeared to be partly controlled by the *WARI* allele (Figure 9C, *t*-test $P_v < 0.005$) consistent with the effect of *WARI* allele on *PDR12* expression. We then performed an allelic switch for *WARI* and constructed a 59A strain expressing the *WARI* allele from S288c (59A *WARI*-S288c). The growth profiles of the strains were monitored in the synthetic must medium supplemented with various amount of sorbic acid. The lag phase was longer for 59A *WARI*-S288c than for 59A wild-type (Figure 9D). This phenotypic difference was subtler than that observed with a knockout strain, but these results nevertheless confirm that sorbic acid resistance is linked to *PDR12* expression and to *WARI* genetic variation. Variations of detoxification

networks may thus be linked to variations of several transcription factor genes. Two of these linkages were validated by allelic switch experiments. Moreover, we demonstrated that the levels of resistance to octanoic acid and sorbic acid were clearly modulated by these genetic variations.

As polymorphism may result from specific adaptation, we tried to infer the phylogeny of *War1p* and *Yrr1p* from nucleotidic sequence available for different strains in SGD. Interestingly, the two allelic versions (wine and laboratory) of these two genes diverge from their *S. paradoxus* orthologue, and are clearly divergent but apparently in a different manner. For *WARI*, S288C and 59A alleles are located on different branches, whereas for *YRR1*, the phylogeny indicates two clusters one containing palm wine and Asian alleles and the second cluster containing 59A wine, rum and bread alleles, as well as S288C alleles. However in this case S288C and 59A are also located on different branches of this cluster (Additional file 7, MEGA 5 [47]). In order to detect if these two genes have evolved neutrally, we performed a McDonald Kreitman test calculating the neutrality index (NI) [48]. This test was applied to a set of 15 *WARI* alleles from strains isolated from various substrates and NI was 2.28 ($P_v = 0.02$), indicating a significant excess of non-neutral mutations. As the cluster of Asian alleles might inflate this test, we removed the sequence and performed the test again, which indicate a similar result (NI = 2.56 and $P_v = 0.03$). We performed the similar analysis on *YRR1* alleles. We obtained NI = 2.51 ($P_v = 0.001$) for the whole set of alleles and NI = 3.93 ($P_v = 0.002$) after removal of the Asian and palm wine alleles. This suggests that *WARI* and *YRR1* do not evolve neutrally: these two genes are either subject to the accumulation of slightly deleterious mutations that are eliminated by negative selection during speciation, or alternatively they present substantial diversity that might be associated with balanced selection resulting from specific adaptation of the strain to wine fermentation conditions.

Discussion

In this study, we developed a genetical genomic approach to get insight into variation of gene regulatory networks during alcoholic fermentation and to assess the impact of this variation on wine yeast properties. We mapped 1465 eQTLs with 601 local eQTLs and 864 distant eQTLs (several distributed in 11 hotspots). As heritability exceeded 80% for 80% of the transcripts, genetic variations clearly had a major impact on global mRNA levels. The differences between the transcripts differentially expressed between the two parental strains and those displaying eQTLs confirmed that multigenetic controls are underlying most variations in transcripts abundance. These observations are consistent with the results obtained for other genetical genomic approaches

Table 3 Genes involved in detoxification having an eQTL

Name		eQTL				SGD Function
Standard	Systematic	Localization ^a	Lod score	Regulation ^b	Venn group	
Ions Detoxification						
<i>MNR2</i>	<i>YKL064W</i>	chr2:151	4,49	unknown	23	Putative magnesium transporter; has similarity to Alr1p and Alr2p which mediate influx of Mg ²⁺ and other divalent cations
<i>ALR2</i>	<i>YFL050C</i>	chr6:30	5,88	Self	123	Probable Mg(2+) transporter; overexpression confers increased tolerance to Al (3+) and Ga(3+) ions; plays a role in regulating Ty1 transposition
<i>SSU1</i>	<i>YPL092W</i>	chr16:371	5,92	Self	123	Plasma membrane sulfite pump involved in sulfite metabolism and required for efficient sulfite efflux; major facilitator superfamily protein
Drug Transport						
<i>TPO2</i>	<i>YGR138C</i>	chr7:761	17,28	Self	123	Polyamine transport protein specific for spermine; localizes to the plasma membrane; transcription of TPO2 is regulated by Haa1p; member of the major facilitator superfamily
<i>SNG1</i>	<i>YGR197C</i>	chr15:639	5,24	<i>YRR1</i>	1	Protein involved in nitrosoguanidine (MNNG) resistance; expression is regulated by transcription factors involved in multidrug resistance
<i>ERC1</i>	<i>YHR032W</i>	chr2:624	4,09	unknown	13	Member of the multi-drug and toxin extrusion (MATE) family of the multidrug/oligosaccharidyl-lipid/polysaccharide (MOP) exporter superfamily
<i>PDR11</i>	<i>YIL013C</i>	chr12:659	4,78	<i>HAP1/PDR8</i>	123	ATP-binding cassette (ABC) transporter multidrug transporter involved in MDR; mediates sterol uptake when sterol biosynthesis is compromised/regulated by Pdr1p
<i>QDR2</i>	<i>YIL121W</i>	chr12:680	10,31	<i>HAP1/PDR8</i>	123	Multidrug transporter of the major facilitator superfamily required for resistance to quinidine barban cisplatin and bleomycin; may have a role in potassium uptake
<i>GEX2</i>	<i>YKR106W</i>	chr11:658	13,66	Self	123	Proton:glutathione antiporter localized to the vacuolar and plasma membranes; almost identical to paralog Gex1p; potential role in resistance to oxidative stress
<i>TPO1</i>	<i>YLL028W</i>	chr12:86	9,89	Self	123	Polyamine transporter that recognizes spermine putrescine and spermidine; catalyzes uptake of polyamines at alkaline pH and excretion at acidic pH
<i>TPO4</i>	<i>YOR273C</i>	chr12:659	4,61	<i>HAP1/PDR8</i>	13	Polyamine transport protein recognizes spermine putrescine and spermidine; localizes to the plasma membrane; member of the major facilitator superfamily
<i>PDR12</i>	<i>YPL058C</i>	chr13:116	7,50	<i>WAR1</i>	123	Plasma membrane ATP-binding cassette (ABC) transporter weak-acid-inducible multidrug transporter required for weak organic acid resistance; regulated by War1p
Transcription factor						
<i>HAP1</i>	<i>YLR256W</i>	chr12:673	6,22	Self	123	Zinc finger transcription factor involved in the complex regulation of gene expression in response to levels of heme and oxygen
<i>PDR8</i>	<i>YLR266C</i>	chr16:138	5,00	unknown	2	Transcription factor; targets include ATP-binding cassette (ABC) transporters, major facilitator superfamily transporters, and other genes involved in the PDR phenomenon
<i>WAR1</i>	<i>YML076C</i>	chr12:673	6,57	<i>HAP1/PDR8</i>	123	Homodimeric Zn ₂ Cys ₆ zinc finger transcription factor; binds to a weak acid response element to induce transcription of PDR12 and FUN34
<i>YRR1</i>	<i>YOR162C</i>	chr15:639	5,03	Self	13	Zn ₂ -Cys ₆ zinc-finger transcription factor that activates genes involved in multidrug resistance; paralog of Yrm1p acting on an overlapping set of target genes
<i>FZF1</i>	<i>YGL254W</i>	chr7:21	4,02	Self	123	Transcription factor involved in sulfite metabolism, identified regulatory target is SSU1, overexpression suppresses sulfite-sensitivity
Other/Unknown						
<i>SVS1</i>	<i>YPL163C</i>	chr16:251	9,55	Self	123	Cell wall and vacuolar protein required for wild-type resistance to vanadate
<i>YKR104W</i>	<i>YKR104W</i>	chr11:651	4,63	Self	13	Putative transporter of the multidrug resistance-associated protein (MRP) subfamily; contains a stop codon in S288C
<i>YLR046C</i>	<i>YLR046C</i>	chr12:238	5,72	Self	123	Putative membrane protein transcription is activated by paralogous transcription factors Yrm1p and Yrr1p along with genes involved in multidrug resistance

Table 3 Genes involved in detoxification having an eQTL (Continued)

<i>YJL216C</i>	<i>YJL216C</i>	chr10:42	11,36	Self	123	Protein of unknown function similar to alpha-D-glucosidases; transcriptionally activated by both Pdr8p and Yrm1p along with other genes involved in the PDR phenomenon
<i>YLR179C</i>	<i>YLR179C</i>	chr12:689	4,08	<i>HAP1/PDR8</i>	23	Protein of unknown function transcription is activated by paralogous proteins Yrm1p and Yrr1p along with proteins involved in multidrug resistance; not essential

^aLocalization of the center of hotspot given in chromosome:kilo base pair.

^b“Self” mean the eQTL is closer than 40 kpb to the gene controlled.

in various organisms [30]. We observed several overlaps between phenotypic QTLs (glycerol, pyruvate) and eQTLs, consistent with the involvement of regulatory networks variation in yeast trait variations. Some of these linkages have been described before (clumpiness, flocculation, mating type), whereas others are probably specific to our conditions or strains. We show that various mechanisms contribute to variations of expression and highlight the role of transcription factors for toxic compound export and the contribution of large chromosomal variations, a partial chromosomal duplication in this case, to the control of fermentation rate.

The transcriptome analysis was performed during the stationary phase in starvation and alcohol stress. Therefore our approach was adapted to describe regulatory change associated to this condition. However, variations in regulatory network involved in nitrogen consumption and cell growth were not considered here. For example, expressions of only few genes were found correlated to Rmax.

Genetic control of fermentation rate

Ambroset *et al.* [4] showed that fermentation rate (Rmax) depended on the PABA biosynthesis capacity and *ABZ1* allele. By bypassing the effect of *ABZ1*, we identified another genetic control of the fermentation

rate: the improvement of fermentation kinetics by partial disomy of the left arm of chromosome 16. In the system studied here, this partial disomy was present only in the offspring, due to cross carried out. This link cannot therefore be considered a “real” QTL, but it nevertheless highlights the role of gene copy number variations in the control of a key phenotype [11].

The mechanisms underlying the improvement of fermentation rate associated with this disomy are unclear, however, and will not be easy to decipher, given the size of the chromosomal region and the large number of genes it contains (190 genes). However, candidate genes may be identified on the basis of their patterns of expression, because the fermentation phenotype is thought to be triggered by a change in expression level. One or several of the 155 genes overexpressed in the translocated area may improve tolerance to starvation or, potentially, to alcohol. We investigated the properties of the set of genes in this region of chromosome 16 and evaluated their correlation with fermentation rate (R70). We observed that levels of expression of *SAM3* and *SAM4*, involved in S-adenosyl methionine (SAdM) pool control, were correlated with R70. Interestingly, the expression levels of other genes, not located in the duplicated region, involved in SAdM synthesis (*SAM1*, *SAM2*) and involved in methyl transfers

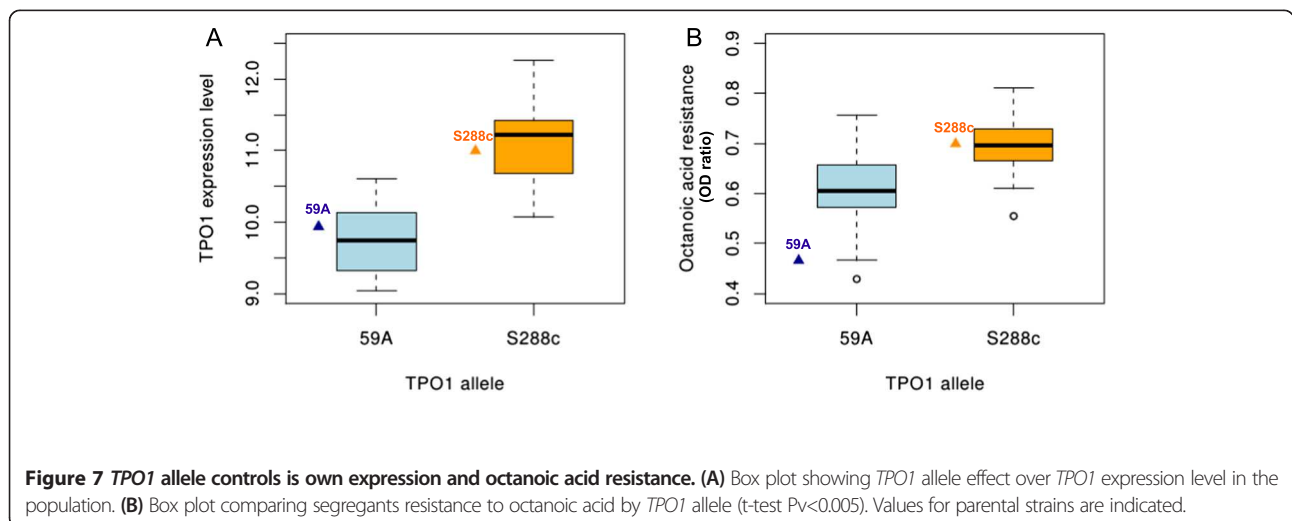


Figure 7 *TPO1* allele controls its own expression and octanoic acid resistance. (A) Box plot showing *TPO1* allele effect over *TPO1* expression level in the population. (B) Box plot comparing segregants resistance to octanoic acid by *TPO1* allele (t-test $P_v < 0.005$). Values for parental strains are indicated.

Table 4 Gene expression modified by *PDR8* allelic switch

ORF	Name	eQTL	LogFC	AdjPv	SGD function
<i>YIL121W</i>	<i>QDR2</i>	hotspot8	2,234	<10e-05	Multidrug transporter of the major facilitator superfamily required for resistance to quinidine, barban, cisplatin and bleomycin
<i>YLR179C</i>	<i>YLR179C</i>	hotspot8	0,897	0,0026	Protein of unknown function; transcription is activated by paralogous proteins Yrm1p and Yrr1p along with proteins involved in multidrug resistance
<i>YLR149C</i>	<i>YLR149C</i>	-	1,349	<10e-05	Putative protein of unknown function; overexpression causes a cell cycle delay or arrest; null mutation results in a decrease in plasma membrane electron transport
<i>YCR098C</i>	<i>GIT1</i>	-	0,548	0,005	Plasma membrane permease, mediates uptake of glycerophosphoinositol and glycerophosphocholine as sources of the nutrients inositol and phosphate
<i>YIL120W</i>	<i>QDR1</i>	-	0,472	0,0046	Multidrug transporter of the major facilitator superfamily, required for resistance to quinidine, ketoconazole, fluconazole, and barban
<i>YLR266C</i>	<i>PDR8</i>	chr16:138	-0,493	0,0032	Transcription factor; targets include ATP-binding cassette (ABC) transporters, major facilitator superfamily transporters, and other genes involved in the pleiotropic drug resistance (PDR) phenomenon

(*CHO2*, *NNT1*, *SET7*) were also correlated with R70. We suspect that *SAM3* and *SAM4* improve the fermentation rate of the disomic segregants by increasing the availability of SAdM. However, the addition of SAdM (final concentration: 0.1 μ M) to the fermentation medium did not improve the fermentation rate of the 59A strain (data not shown). We previously showed that the methyl donor synthesis and methionine biosynthesis pathways had a strong impact on fermentation rate [4], highlighting the key role of these metabolic pathways in alcoholic fermentation. Additional experiments are required to determine the mechanisms underlying this control of fermentation rate.

As this chromosomal amplification increased the fermentation capacity of wine yeasts, we checked for its

presence in wild and industrial strain. Dunn *et al.* [12] reported a aCGH analysis of 83 *S. cerevisiae* strains including 69 wine strains. We used their data to search for a partial trisomy (for diploid strains) of the left arm of chromosome 16. We detected such partial trisomy in the wine yeast strain NT45. Moreover, in their characterization of the *S. cerevisiae/S. kudriavzevii* triploid hybrid EG8, Erny *et al.* [49] detected a major amplification (four copies) of the left arm of the *S. cerevisiae* chromosome 16. The diploid *S. cerevisiae* strain EG25, which is genetically related to the *S. cerevisiae* moiety of the EG8 genome (with no *S. kudriavzevii* DNA), also displays trisomy of this region (Legras JL, personal communication). We determined the form of the chromosomes 16 and 8 (normal or translocated) present, by PCR for these

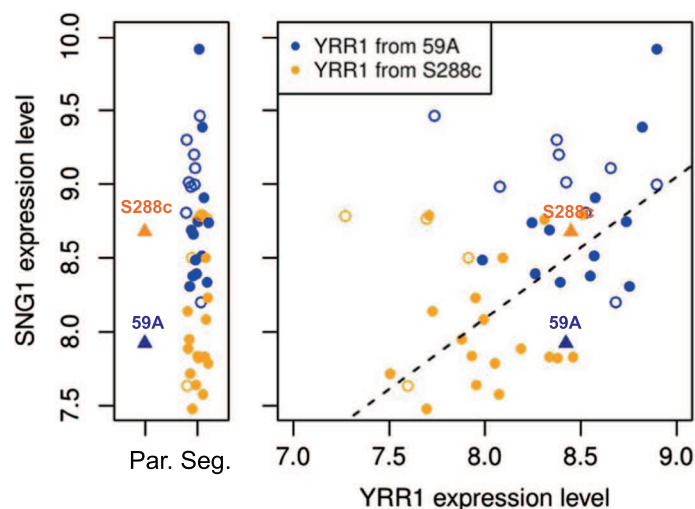
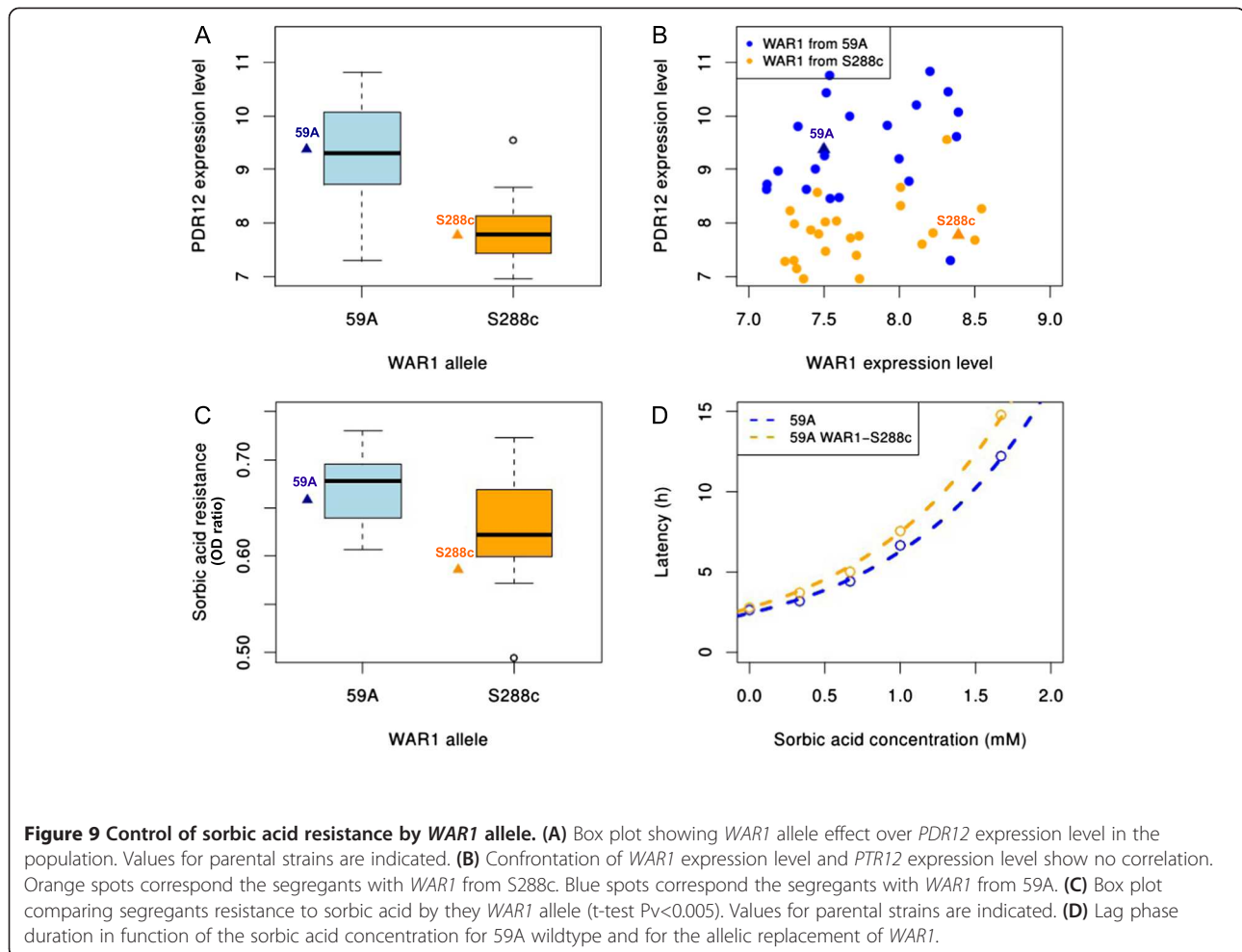


Figure 8 *YRR1* control over *SNG1* expression. Orange spots correspond the segregants with *YRR1* from S288c, blue spots correspond the segregants with *YRR1* from 59A. Open circles correspond to disomic segregants, There is a permutation between parent and the allelic effect in the segregants population, involving polygenic control.



three strains (Figure 10). The NT45 and EG25 strains carry three chromosomes 16 left arms, originating from two normal chromosomes 16 and one copy of chromosome 8 carrying the translocation from chromosome 16. The hybrid aneuploid strain EG8 carries four left arms of chromosome 16: one from a normal chromosome 16 and three from the three chromosomes 8, all carrying the translocation. This findings confirm that amplification of the left arm of chromosome 16 occurs in wild/industrial strains. The situation described here suggests a spectacular evolutionary event occurring in two independent steps: a translocation selected due to its effect on sulfite resistance followed by amplification of the chromosomal region concerned, improving fermentation properties.

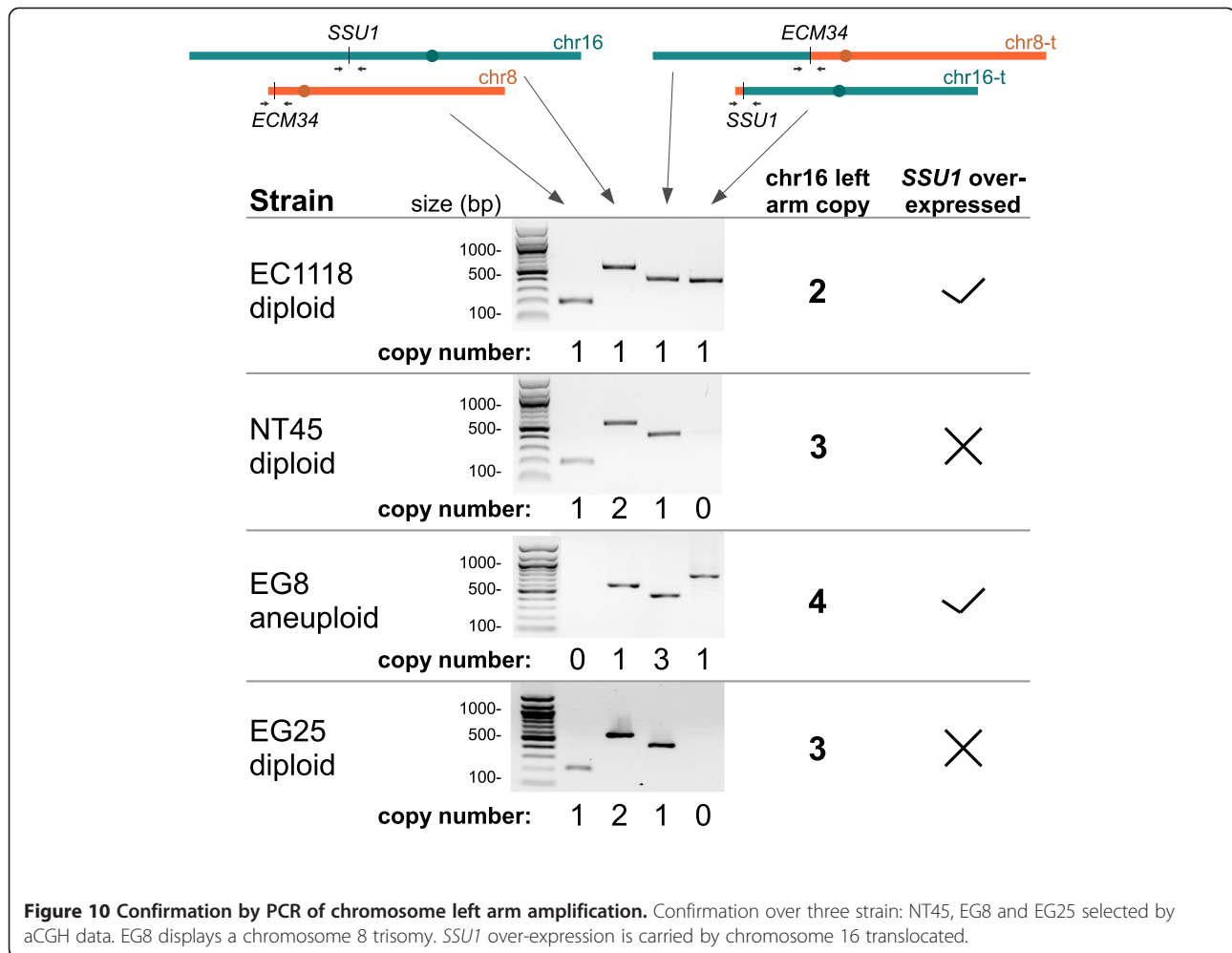
We evaluated the contribution of the disomy effect to eQTL linkage, by modifying the input dataset. We found that a large set of new eQTLs had been masked by the effect of this disomic. Most of these eQTLs (113) formed 2 hotspots on chromosome 2. Several of the genes controlled by these hotspots were found correlated with R70. Thus, these loci may partially control fermentation

rate, despite the lack of detection of a QTL for this phenotype.

Importance of variations in detoxification systems

The analysis of functional enrichment for all eQTLs (in FunSpec [33]) highlighted a set of 22 eQTLs linking genetic variation to expression level of genes involved in detoxification. Our data show the importance of variations in transcription factor genes and their impact on detoxification systems: *PDR8*, *WAR1* and *YRR1*, in addition to *HAP1* (as summarized in Figure 11).

During fermentations, yeast produces toxic compounds that may lead to stuck fermentations [50]. This is the case for the medium-chain fatty acids generated as by-products of lipid biosynthesis. *PDR12* has been shown to be involved in weak organic acid [46], and Legras *et al.* [3] have shown that *PDR12*, together with *TPO1*, is also a key transporter involved in octanoic acid and decanoic acid resistance. We were able to link genetic variations of *TPO1* to its expression and to octanoic acid resistance. *PDR12* has an eQTL in the gene encoding its known transcription

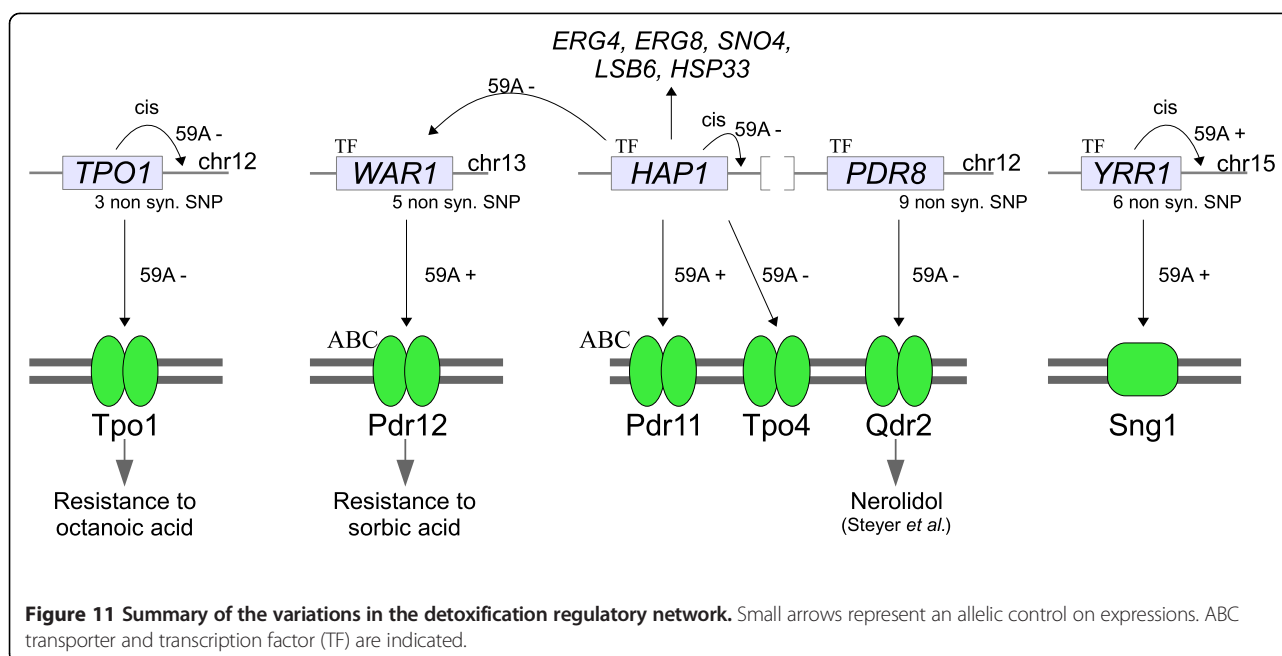


factor, *WARI*, that we could link to variations of sorbic acid resistance, but not to octanoic acid resistance, suggesting that the allelic effect of *TPO1* outweighs the control of medium-chain fatty acid resistance.

For the same population, Steyer et al. [32] shown that the allele of *PDR8* gene controls the export of nerolidol via *QDR2*. We confirm here that *PDR8* allelic variation controls the expression of *QDR2* and *YLR179c*, the function of which remain unknown. We observed that several drug exporters (*TPO1*, *TPO2*, *TPO4* and *QDR2*) were less strongly expressed by the wine form, consistent with the deletions in wine yeasts reported by Dunn et al. (*PDR3*, *SNQ1*, *QDR1* [11]). The exact physiological role of several of these transporters remains unclear. Qdr2p was initially identified as a drug efflux system but its natural targets remain a matter of debate [51,52]. It was subsequently suggested that Qdr2p might export amino acids in starvation conditions [53]. Such a function would be consistent with the lower levels of expression of this gene in wine yeast, given the major effect of nitrogen homeostasis on wine yeasts fermentation capacity.

In addition to the toxic compounds produced by the yeast during fermentation, natural grape musts contain various phenolic compounds [54], which act as key wine preservatives and contribute to aging. Stilbene compounds are clearly toxic to fungi, such as *S. cerevisiae* and *Aspergillus flavus* [55]. Pan et al. [56] studied the transcriptomic response to pterostilbene (an analog of resveratrol) and show that *YRR1* and *SNG1* were among the upregulated genes in response to the drug. We identified eQTLs for these two genes, which were upregulated by the wine allele of *YRR1* transcription factor encoding gene.

As the polymorphism observed here may result from specific adaptation to exposure to these compounds, we tried to infer to phylogeny of War1p and Yrr1p and evaluate if these genes evolve neutrally. Interestingly the McDonald Kreitman test [48] revealed a non neutral evolution for these two genes (NI calculated was greater than two): this indicates very likely that the substantial diversity observed for *WARI* and *YRR1* results from balanced selection due to specific adaptation of the strain



to wine fermentation conditions. This is consistent with the analysis of Steyer *et al.* [32], who reported also a non neutral evolution of *PDR8* according to the results of a McDonald test.

Unlike Yvert *et al.* [34], who reported that distant QTLs were not often due to variations of transcription factor genes, we found that most of the variation of detoxification gene expression was linked to transcription factors. Surprisingly, variations in these three genes (*PDR8*, *YRR1*, *WAR1*) encoding zinc-finger transcription factors modified the expression of only one or two of their known targets. This may be partly due to the different environment in which *PDR8* targets have been characterized. We performed our expression analysis in stationary phase under a stressful condition whereas cell growths in rich medium were used for the Chip on Chip assay of Hikkel *et al.* [43]. In addition, Jothi *et al.* [57] have shown that these transcription factors belong to the lower layer of the hierarchy of the yeast transcription factors network, with direct effects on a limited number of targets. This may make them more tolerant to variation than genes higher up in this hierarchy.

Regulatory network variations involved in wine fermentation

In this study, we investigated, for the first time, the expression profiles of the genes present in the three regions introgressed into EC1118 by horizontal transfer [13]. As we worked with a second parental strain devoid of these three regions, we could not identify real eQTLs for these genes. We searched for links, using the populations of segregant possessing each region (half of

segregants). On chromosome 6, at 145kbp, an eQTL was identified that trigger the expression of the two genes encoding polypeptide transporters (FOT) describes by Damon *et al.* [41]. The new fructose transporter gene, *FSY1*, [58] also had an eQTL on chromosome 13, at 446 kbp (data not shown). No clear candidate gene was identified in these loci, but our findings could lead to revealing the role of these new genes in fermentation. No phenotypic QTL or trans-eQTL were found in these three regions. More detailed analysis, with a larger set of segregants, should be carried out to assess the variations of expression associated with these regions.

The phenotypic data indicated that most of the traits considered were under multigenic control. This may reflect the large genetic distance between our two parental strains, but Marullo *et al.* [59] also found, in a study using two commercial wine yeasts and a population of 51 segregants, a preponderance of multigenic traits, such as ethanol tolerance and kinetic properties. We were nevertheless able to associate several phenotypic QTLs with eQTLs. Pyruvic acid levels, which were strongly correlated with R70, were linked to a major hotspot on chromosome 2. A candidate gene, *SCT1*, was identified. This gene is involved in lipid desaturation, suggesting that it may act by modulating the plasma membrane stability under alcoholic stress.

However, within the eQTLs set, several new linkages are probably involved in the management of the multi-stress conditions in wine fermentation. Changes were detected for genes involved in glucose sensing (*GRR1* cis-eQTL that may control *SNF3* expression) or thiamine synthesis (affected by *THI3*). Functional validations of these results

are required, together with further investigations to determine the role of such genetic variation in the adaptation of strains to the wine making process. We also showed that the sulfur amino-acid pathway (*MET32*, *MET2*, *MET5*) was partially enhanced by *SUL2* mutation, probably through effects on sulfate uptake. Such activation has not been described before and may reflect the conditions of nitrogen starvation in this study.

Conclusion

In this study, we developed an approach to deciphering the variation of transcriptional networks in yeast, during an industrial wine-making process resulting in multiple stresses. The results obtained were consistent with those of other studies. However, the wine fermentation conditions used here led to the identification of specific variations of regulatory networks not revealed previously in classical laboratory environment. We show that expression variations affect networks of genes involved in various pathways and cellular functions, through diverse mechanisms involving both local and distal regulations and copy number changes driven by chromosomal rearrangements. We focus here on variations of detoxification systems and identified and we confirm the key role of variations in transcription factor genes in modulating the expression of these systems. These results also raise questions about the role of these export systems in fermentation conditions, in which cells are starved and stressed.

The eQTL linkage analysis was generally highly consistent, but most of the phenotype changes were not linked genetic variations and were considered to be under multi-locus control. We found that fermentation rate depended on a gene copy numbers effect of a duplication of the left arm of chromosome 16. This partial disomy was also found in natural yeasts and appears to be a potential new target for yeast optimization for the wine making process.

Methods

Strains, growth conditions and fermentation medium

The two parental strains used for the segregating population are S288c (*MAT α* ; *SUC2*, *gal2*) strain and 59A (*MAT α* ; ho), which is a haploid derivative of EC1118 (HO/ho), an industrial wine strain [4]. Mutations between the two parental strains are accessible in the GenYeastTrait database previously described (<http://genome.jouy.inra.fr/genyeastrait/> [4]).

We increased the population of 30 F1 segregants described in [4] with 14 randomly selected segregants. Thereby we disposed of two complete tetrads in the analyzed population, and the other segregants were selected from 24 different asci of two or three viable spores. The phenotyping was done on fermentation in 0,9l of synthetic SM425 medium (with 425 mg/l of assimilate

nitrogen pH3.3 and previously describe as SM300 [60]) which mimics natural grape must. This medium was supplemented by 1 mg/l of p-aminobenzoic acid (PABA, sigma A-9878), 14.5 mg/l of iron (III) chloride (FeCl_3 , sigma F-7134), 10 $\mu\text{l/l}$ of antifoam silicone (rhodorsil Prolabo 27429.297). The sugar source was glucose (10%) and fructose (10%) and the quantity of anaerobic factor was reduced by $\frac{1}{2}$. After a reactivation of strain on YPED (48 h) and a preculture in shake flasks containing 30 ml of SM425 (12 h), 900 ml of medium were inoculated at 10^6 cell/ml. Fermenters were equipped with airlock to maintain anaerobiosis. Fermentation were performed at 24°C and preculture at 28°C.

Fermentation kinetics and phenotypic measures

Each parental fermentation and phenotypic measures were realized in four replicates and segregants fermentation in two replicates. The fermentation kinetics were measured by the loss of weight due to CO_2 release. The weight was record every 20 min and the data were smoothed by polynomial method. The final population size was estimated at 45 g/l of CO_2 release and 66 g CO_2 /l using electronic particle counter (Beckman Coulter) and optic density at 600 nm (OD_{600} , spectrophotometer SECOMAN-UVLine9400). Dry weights were measured at 88 g CO_2 /l by filtration using nitrocellulose filters (pore size 0.45 μm ; Millipore) and drying at 100°C (48 h).

The metabolic compounds amounts in the medium was measured after the end of the fermentation by high-pressure liquid chromatography (HPLC) using ion-exclusion column (HPX-87H BIO-RAD) with 8 mM H_2SO_4 mobile phase. Glucose, ethanol, glycerol and succinate were detected by refractometry. Acetic acid and pyruvic acid were detected by UV absorption (as described in [61]). Measures repeatability between replicate was controlled and the phenotype heritability H^2 was calculated as follow (describe in [35]):

$$H^2 = \frac{\text{Var}_{seg} - \text{Var}_{env}}{\text{Var}_{seg}} \times 100$$

where Var_{env} is the pooled variance among parental measurement and Var_{seg} is the variance among phenotype values for the segregants. Phenotypes with low H^2 are not statistically valid and were discarded.

Resistance phenotyping was performed in the same SM425 medium (pH3.3) than fermentation with a completion of the right amount of toxic compound stock solution and ethanol for a final rate of 1.7% (vol/vol). Octanoic acid stock solution: 69.3 mM in ethanol. Sorbic acid stock solution: 33.7 mM in water. The growth was taken with OD_{600} with and without toxic compound, and resistance indicator was measured by latency differences, ratio of specific growth rate and ratio of OD_{600} at 20 hours (end of

exponential stage). We here only considerate the ratio of OD₆₀₀, showing the lower experimental variation. Three replicates were done for each parental growth.

Molecular biology and strains construction

PDR8 allelic switch in 59A was obtained in three steps: 1) depletion of *PDR8* in 59A using hphMX4 cassette for hygromycin resistance (pAG32). Primers sequences for cassette amplification and verification were obtained from Euroscarf. 2) preparation of a replacement cassette containing *PDR8-loxP-kanMX4-loxP* by the insertion of loxP-kanMX4-loxP (pUG6 [62]) in the terminator of *PDR8* in S288c strain (primers in Additional file 8). 3) allelic switch by depletion of the hphMX4 cassette of 59A *PDR8Δ::hph* by the *PDR8-loxP-kanMX4-loxP* replacement cassette from S288c and selection on YEPD containing G418 (200 μg/l). The loss of hphMX4 cassette was controlled by PCR and the absence of growth on hygromycin (200 μg/l).

WARI allelic switch in 59A was obtained in two steps: 1) depletion of *WARI* in 59A using loxP-kanMX4-loxP (pUG6). Primers sequences for cassette amplification and verification were obtained from Euroscarf. 2) allelic switch by depletion of the kanMX cassette by *WARI* sequence from S288c DNA (primers in Additional file 8) and selection on YEPD containing 300mg/l sorbic acid, pH4.2. The loss of kanMX cassette was controlled by PCR and the absence of growth on YEPD supplemented by G418 (200 μg/l).

The 8–16 translocation was confirmed by PCR, using the same primers than Pérez-Ortín *et al.* [10].

Transcriptomic profiling

The transcriptome profiling was performed only one time for each segregants and in three technical replicates for each two biological replicates of the parental strains. At 66 gCO₂/l release (70% of fermentation progress), 10⁹ cells were sampled, pelleted, washed with DEPC-treated water and freezed in methanol at -80°C. This condition is corresponding to late stationary phase; cells are not in transition situation which could have increase experimental variation on transcriptome. Total RNA extractions were performed with Trizol reagent (Gibco BRL, Life Technologies), purified by isopropanol precipitation then with RNeasy kit (Qiagen). Cy3-labeled cRNA was synthesized with the One color RNA Spike-In kit (Agilent Technologies) and purified with RNeasy kit (Qiagen). Quality and quantity of RNA were controlled at each step by spectrometry (NanoDrop 1000, Thermo Scientific).

Agilent gene expression microarrays 8x15k was used for the micro array hybridization, with one-color method (Agilent Technologies, Santa Clara, CA, USA). Array design is based on ID 016322 completed with the 39 genes from the new region of EC1118 [13] and available on GEO with GPL16012 as accession number. A quantity of

600ng of labeled cRNA were hybridized for 17 h in 65°C in a rotative hybridization oven (Corning) using the Expression Hybridization kit (Agilent Technologies, 5188–5242). Plates were washed with expression wash buffer kit (Agilent Technologies, 5188–5325 5188–5326). The array pictures were analyzed on a GenePix 4000B laser Scanner (Axon Instruments) and with the GenePix PRO7 software.

Data normalization and statistical analyses were performed using R 2.13.1 software and the limma package [63–66]. Normalization was done by the quantile method considering the whole array data set (55 arrays). The normalized LOG₂ of the spot-median intensity was used as the quantitative evaluation of gene expression (5.3 corresponding to background signal, 16 to spot saturation). Biological and technical repeatability were estimated higher than 87% and 96% respectively using parental strain replicate by the intra-class coefficient of correlation [67].

Comparative transcriptomic between parental strain and between disomic/normal segregants were performed with a modified t-test using the Benjamini and Hochberg false discovery rate as multiple testing correction of the t-test p-values (adjPv) [68]. The threshold used was the adjPv lower than 0.01, and a filter with a log₂ of fold-change (logFC) greater than 0.7 or lower than -0.7 was applied for parental comparison.

Hierarchical clustering was performed using cluster v3.0 (Centered correlation and complete linkage [69]) and displayed with JavaTreeView v1.1.5r2 [70]. As previously described in Yvert *et al.* [34], we defined a statistically significant degree of correlation between genes by permutation testing (n=10) and focused further analyses to clusters in which all pairwise correlations are greater than 0.84. At this threshold, fewer than three clusters of two genes and no cluster of more than two genes are expected by chance. Functional analysis of transcriptomic output was realized using Funspec with the Bonferroni correction at p-value cutoff of 0.05 [33]. Complete array data set is available on Gene Expression Omnibus database (global analysis: GSE41025, *PDR8* allelic switch analysis: GSE41738).

Genotyping

The genotyping of the 14 new strains was performed with the same method than the first analysis [4]: Genomic DNA was isolated, fragmented, labeled, and hybridized on Affymetrix YGS98 microarrays, one time for each strain. A new marker map was designed doing the same selection than the precedent analysis described in Ambroset *et al.* 2011 [4] and inspired from Brem *et al.* 2002 [24]. This resulted in a map of 2140 markers.

To study genome duplication, comparative genomic hybridization (CGH-like) analysis was performed using Affymetrix normalized logarithm of PM signal of all the

probes. A smoothing taking the average over 200 probes was done.

We completed the 2140 markers map with 46 markers localized in area of low markers density. These markers were based on single nucleotide polymorphism detection using Illumina veracode technology. This resulted in a good genotyping cover with an average density of 1.81 markers each 10 kbp.

Linkage analysis

The linkage analysis was performed using R/qtl package [71] on the three datasets corresponding to the three analyses (Analysis1, Analysis2 and Analysis3). The normal model with Haley-Knott regression method was used resulting in logarithm of odds (LOD) score for each marker and pseudo-marker every 2.5 cM (7.5 kbp) (interval mapping method). An interval estimate of the location of each QTL or eQTL was obtained as the 1-LOD support interval: the region in which the LOD score is within 1 unit of the peak LOD score. For phenotypic QTLs, individual LOD score threshold for a false discovery rate of 0.05 was determined doing 1000 permutations. For eQTLs, the permutation was performed 20 times, and the average number of transcripts showing linkage at a specific LOD score threshold was used to calculate FDR.

The eQTLs closer than 40 kbp to the localization of the gene were considered as "local" eQTLs or cis-eQTLs. This could be due to a mutation in the gene promoter (real cis-eQTL), a mutation in the protein that control its own expression as a feedback, a mutation in an other gene close that control its expression (trans-eQTL co-localized). They also can be artifacts due to mutations in sequence that reduce the mRNA affinity with the micro-array probe.

Phylogeny and neutrality tests

To infer the evolutionary history of *WARI* and *YRR1*, we collected their sequences from genomes available at SGD (<http://www.yeastgenome.org/>). All uncompleted or frameshift-containing sequences were discarded from this set. The phylogenies were inferred with MEGA [47] by the Maximum Likelihood method based on the Kimura 2-parameter model [72]. The trees with the highest log likelihood are shown. The trees are drawn to scale, with branch lengths proportional to the number of substitutions per site.

The Neutrality Index correspond to the comparison of the ratio of non synonymous to synonymous polymorphism (intra species) to the ratio of non synonymous to synonymous divergence with the nearest species. An NI lower than one reflects a paucity of non synonymous polymorphism relative to non synonymous divergence, and is indicative of positive selection; an NI greater than one indicates negative selection of deleterious alleles driving divergence between species or balancing selection. The

significance of the NI test [48] was calculated using the <http://bioinf3.uab.cat/mkt/MKT.asp> website.

Availability of supporting data

Sequence of 59A strain is available in <http://genome.jouy.inra.fr/genyeastrait/>. Transcriptomic row data are available in Gene Expression Omnibus database (global analysis: GSE41025, *PDR8* allelic switch analysis: GSE41738). Other supporting data are included in the additional files.

Additional files

Additional file 1: Example of phenotypic value distribution among the population. Parental values are indicated in the top with open circle and black circle for 59A and S288c respectively. Heritability (H^2) is indicated. R70 exhibits a bimodal distribution while other phenotypes have a continuous distribution.

Additional file 2: Clustering analysis.

Additional file 3: Markers map. 2140 markers are from Affymetrix genotyping, 46 markers were based on single nucleotide polymorphism detection using Illumina veracode technology to cover area of low Affymetrix markers density. The global density is 1.81 markers each 10 kbp.

Additional file 4: Comparison of partial disomy correction analyses.

Additional file 5: Hotspot 8 genes triggered by *HAP1* or/and *PDR8*.

Additional file 6: Flocculation phenotype triggered by *FLO1* expression.

Additional file 7: Phylogenic trees of the protein sequence of the two transcription factor involved in drug detoxification network variation. Trees were drawn by MEGA5 software with the maximum likelihood method from the genome sequences available [SGD]. Parental strains are indicated by blue and yellow spot for 59A and S288c respectively.

Additional file 8: Primer used.

Abbreviations

QTL: Quantitative trait locus; X-QTL: Extreme QTL; eQTL: Expression QTL; cis-eQTL: eQTL closer than 40 kbp to the localization of the gene ("local" eQTL); trans-eQTL: eQTL farther than 40 kbp to the localization of the gene ("distant" eQTL); aCGH: Comparative genomic hybridization in array; logFC: Log₂ of Fold-Change; FDR: False discovery rate; LOD: Logarithm of odds; SAdM: S-adenosyl methionine; PABA: Para-amino benzoate; Rmax: Maximal fermentation rate; R70: Fermentation rate at 70% of fermentation progress; OD₆₀₀: Optic density at 600 nm; Pv: P-value; adjPv: Adjusted P-value; kbp: Kilo base-pair.

Competing interests

The authors declare that they have no competing interests.

Author's contributions

CB participated in the design of the study, performed the data acquisition and analysis, and drafted the manuscript. CA participated in the design of the study, and realized the 59AxS288c lineage. IS performed the statistical analyses, the transcriptomic data processing and the QTL and eQTL linkages. JLL performed the phylogenetic analysis and provided guidance over the data analysis and the manuscript preparation. BB conceived and coordinated the study, and provided guidance over the manuscript preparation. All authors read and approved the final manuscript.

Acknowledgment

C. Brion was supported by a grant from French Ministry for Higher Education and Research. C. Ambroset was supported by a fellowship from the INRA Département Microbiologie et Chaîne Alimentaire (MICA) and the region

Languedoc-Roussillon. This work was partly supported by Agence Nationale de la Recherche grant Genyeastrait ANR-07-BLAN-0205.

Author details

¹INRA, UMR1083 Science pour l'Œnologie, 2 Place Viala, Montpellier F-34060, France. ²Montpellier SupAgro, UMR1083 Science pour l'Œnologie, 2 Place Viala, Montpellier F-34060, France. ³Université Montpellier 1, UMR1083 Science pour l'Œnologie, 2 Place Viala, Montpellier F-34060, France.

Received: 20 June 2013 Accepted: 30 September 2013

Published: 4 October 2013

References

1. Camarasa C, Sanchez I, Brial P, Bigey F, Dequin S: **Phenotypic landscape of *Saccharomyces cerevisiae* during wine fermentation: evidence for origin-dependent metabolic traits.** *PLoS ONE* 2011, **6**:e25147.
2. Divol B, du Toit M, Duckitt E: **Surviving in the presence of sulphur dioxide: strategies developed by wine yeasts.** *Appl Microbiol Biotechnol* 2012, **95**:601–613.
3. Legras JL, Erny C, Jeune CL, Lollier M, Adolphe Y, Demuyter C, Delobel P, Blondin B, Karst F: **Activation of two different resistance mechanisms in *Saccharomyces cerevisiae* upon exposure to octanoic and decanoic acids.** *Appl Environ Microbiol* 2010, **76**:7526–7535.
4. Ambroset C, Petit M, Brion C, Sanchez I, Delobel P, Guérin C, Chiapello H, Nicolas P, Bigey F, Dequin S, Blondin B: **Deciphering the molecular basis of wine yeast fermentation traits using a combined genetic and genomic approach.** *G3: Genes, Genomes, Genetics* 2011, **1**:263–281.
5. Warringer J, Zörgö E, Cubillos FA, Zia A, Gjuvsland A, Simpson JT, Forsmark A, Durbin R, Omholt SW, Louis EJ, Liti G, Moses A, Blomberg A: **Trait variation in yeast is defined by population history.** *PLoS Genet* 2011, **7**:e1002111.
6. Marullo P, Mansour C, Dufour M, Albertin W, Sicard D, Bely M, Dubourdieu D: **Genetic improvement of thermo-tolerance in wine *Saccharomyces cerevisiae* strains by a backcross approach.** *FEMS Yeast Res* 2009, **9**:1148–1160.
7. Dequin S, Casaregola S: **The genomes of fermentative *Saccharomyces*.** *C R Biol* 2011, **334**:687–693.
8. Sicard D, Legras J-L: **Bread, beer and wine: yeast domestication in the *Saccharomyces sensu stricto* complex.** *C R Biol* 2011, **334**:229–236.
9. Bisson LF: **Geographic origin and diversity of wine strains of *Saccharomyces*.** *Am J Enol Vitic* 2012, **63**:165–176.
10. Pérez-Ortín JE, Querol A, Puig S, Barrio E: **Molecular characterization of a chromosomal rearrangement involved in the adaptive evolution of yeast strains.** *Genome Res* 2002, **12**:1533–1539.
11. Dunn B, Levine RP, Sherlock G: **Microarray karyotyping of commercial wine yeast strains reveals shared, as well as unique, genomic signatures.** *BMC Genomics* 2005, **6**:53.
12. Dunn B, Richter C, Kvitck DJ, Pugh T, Sherlock G: **Analysis of the *Saccharomyces cerevisiae* pan-genome reveals a pool of copy number variants distributed in diverse yeast strains from differing industrial environments.** *Genome Res* 2012, **22**:908–924.
13. Novo M, Bigey F, Beyne E, Galeote V, Gavory F, Mallet S, Cambon B, Legras J-L, Wincker P, Casaregola S, Dequin S: **Eukaryote-to-eukaryote gene transfer events revealed by the genome sequence of the wine yeast *Saccharomyces cerevisiae* EC1118.** *Proc Natl Acad Sci USA* 2009, **106**:16333–16338.
14. Galeote V, Bigey F, Beyne E, Novo M, Legras J-L, Casaregola S, Dequin S: **Amplification of a *Zygosaccharomyces bailii* DNA segment in wine yeast genomes by extrachromosomal circular DNA formation.** *PLoS One* 2011, **6**:e17872.
15. Liti G, Carter DM, Moses AM, Warringer J, Parts L, James SA, Davey RP, Roberts IN, Burt A, Koufopanou V, Tsai IJ, Bergman CM, Bensasson D, O'Kelly MJT, Van A O, Barton DBH, Bailes E, Nguyen AN, Jones M, Quail MA, Goodhead I, Sims S, Smith F, Blomberg A, Durbin R, Louis EJ: **Population genomics of domestic and wild yeasts.** *Nature* 2009, **458**:337–341.
16. Schacherer J, Shapiro JA, Ruderfer DM, Kruglyak L: **Comprehensive polymorphism survey elucidates population structure of *S. cerevisiae*.** *Nature* 2009, **458**:342–345.
17. Steinmetz LM, Sinha H, Richards DR, Spiegelman JJ, Oefner PJ, McCusker JH, Davis RW: **Dissecting the architecture of a quantitative trait locus in yeast.** *Nature* 2002, **416**:326–330.
18. Hu XH, Wang MH, Tan T, Li JR, Yang H, Leach L, Zhang RM, Luo ZW: **Genetic dissection of ethanol tolerance in the budding yeast *Saccharomyces cerevisiae*.** *Genetics* 2007, **175**:1479–1487.
19. Cubillos FA, Billi E, Zörgö E, Parts L, Fargier P, Omholt S, Blomberg A, Warringer J, Louis EJ, Liti G: **Assessing the complex architecture of polygenic traits in diverged yeast populations.** *Mol Ecol* 2011, **20**:1401–1413.
20. Salinas F, Cubillos FA, Soto D, Garcia V, Bergström A, Warringer J, Ganga MA, Louis EJ, Liti G, Martinez C: **The genetic basis of natural variation in oenological traits in *Saccharomyces cerevisiae*.** *PLoS ONE* 2012, **7**:e49640.
21. Marullo P, Aigle M, Bely M, Masneuf-Pomarède I, Durrens P, Dubourdieu D, Yvert G: **Single QTL mapping and nucleotide-level resolution of a physiologic trait in wine *Saccharomyces cerevisiae* strains.** *FEMS Yeast Res* 2007, **7**:941–952.
22. Ehrenreich IM, Gerke JP, Kruglyak L: **Genetic dissection of complex traits in yeast: insights from studies of gene expression and other phenotypes in the BYxRM cross.** *Cold Spring Harb Symp Quant Biol* 2009, **74**:145–153.
23. Ehrenreich IM, Torabi N, Jia Y, Kent J, Martis S, Shapiro JA, Gresham D, Caudy AA, Kruglyak L: **Dissection of genetically complex traits with extremely large pools of yeast segregants.** *Nature* 2010, **464**:1039–1042.
24. Brem RB, Yvert G, Clinton R, Kruglyak L: **Genetic dissection of transcriptional regulation in budding yeast.** *Science* 2002, **296**:752–755.
25. Smith EN, Kruglyak L: **Gene-environment interaction in yeast gene expression.** *PLoS Biol* 2008, **6**:e83.
26. Brem RB, Kruglyak L: **The landscape of genetic complexity across 5,700 gene expression traits in yeast.** *Proc Natl Acad Sci USA* 2005, **102**:1572–1577.
27. Brown KM, Landry CR, Hartl DL, Cavalieri D: **Cascading transcriptional effects of a naturally occurring frameshift mutation in *Saccharomyces cerevisiae*.** *Mol Ecol* 2008, **17**:2985–2997.
28. Zhu J, Sova P, Xu Q, Dombek KM, Xu EY, Vu H, Tu Z, Brem RB, Bumgarner RE, Schadt EE: **Stitching together multiple data dimensions reveals interacting metabolomic and transcriptomic networks that modulate cell regulation.** *PLoS Biol* 2012, **10**:e1001301.
29. Lee S-I, Dudley AM, Drubin D, Silver PA, Krogan NJ, Pe'er D, Koller D: **Learning a prior on regulatory potential from eQTL data.** *PLoS Genet* 2009, **5**:e1000358.
30. Williams RBH, Chan EKF, Cowley MJ, Little PFR: **The influence of genetic variation on gene expression.** *Genome Res* 2007, **17**:1707–1716.
31. Kliebenstein D: **Quantitative genomics: analyzing intraspecific variation using global gene expression polymorphisms or eQTLs.** *Annu Rev Plant Biol* 2009, **60**:93–114.
32. Steyer D, Ambroset C, Brion C, Claudel P, Delobel P, Sanchez I, Erny C, Blondin B, Karst F, Legras J-L: **QTL mapping of the production of wine aroma compounds by yeast.** *BMC Genomics* 2012, **13**:573.
33. Robinson MD, Grigull J, Mohammad N, Hughes TR: **FunSpec: a web-based cluster interpreter for yeast.** *BMC Bioinforma* 2002, **3**:35.
34. Yvert G, Brem RB, Whittle J, Akey JM, Foss E, Smith EN, Mackelprang R, Kruglyak L: **Trans-acting regulatory variation in *Saccharomyces cerevisiae* and the role of transcription factors.** *Nat Genet* 2003, **35**:57–64.
35. Brem RB, Storey JD, Whittle J, Kruglyak L: **Genetic interactions between polymorphisms that affect gene expression in yeast.** *Nature* 2005, **436**:701–703.
36. Iraqui I, Vissers S, Bernard F, de Craene J-O, Boles E, Urrestarazu A, André B: **Amino acid signaling in *Saccharomyces cerevisiae*: a permease-like sensor of external amino acids and f-box protein *grr1p* are required for transcriptional induction of the *agp1* gene, which encodes a broad-specificity amino acid permease.** *Mol Cell Biol* 1999, **19**:989–1001.
37. Abdulrehman D, Monteiro PT, Teixeira MC, Mira NP, Lourenco AB, dos Santos SC, Cabrito TR, Francisco AP, Madeira SC, Aires RS, Oliveira AL, Sa-Correia I, Freitas AT: **YEASTRACT: providing a programmatic access to curated transcriptional regulatory associations in *Saccharomyces cerevisiae* through a web services interface.** *Nucleic Acids Res* 2010, **39**:D136–D140.
38. Martínez-Moreno R, Morales P, Gonzalez R, Mas A, Beltran G: **Biomass production and alcoholic fermentation performance of *Saccharomyces cerevisiae* as a function of nitrogen source.** *FEMS Yeast Res* 2012, **12**:477–485.
39. Byrd C, Turner GC, Varshavsky A: **The N-end rule pathway controls the import of peptides through degradation of a transcriptional repressor.** *EMBO J* 1998, **17**:269–277.
40. Turner GC, Du F, Varshavsky A: **Peptides accelerate their uptake by activating a ubiquitin-dependent proteolytic pathway.** *Nature* 2000, **405**:579–583.
41. Damon C, Vallon L, Zimmermann S, Haider MZ, Galeote V, Dequin S, Luis P, Fraissinet-Tachet L, Marmeisse R: **A novel fungal family of oligopeptide transporters identified by functional metatranscriptomics of soil eukaryotes.** *ISME J* 2011, **5**:1871–1880.
42. Straver MH, Aar PCVD, Smit G, Kijne JW: **Determinants of flocculence of brewer's yeast during fermentation in wort.** *Yeast* 1993, **9**:527–532.

43. Hikkel I, Luciau-Danila A, Delaveau T, Marc P, Devaux F, Jacq C: **A general strategy to uncover transcription factor properties identifies a new regulator of drug resistance in yeast.** *J Biol Chem* 2003, **278**:11427–11432.
44. Grey M, Pich CT, Haase E, Brendel M: **SNG1—a new gene involved in nitrosoguanidine resistance in *Saccharomyces cerevisiae*.** *Mutat Res* 1995, **346**:207–214.
45. Schuller C: **Global phenotypic analysis and transcriptional profiling defines the weak acid stress response regulon in *saccharomyces cerevisiae*.** *Mol Biol Cell* 2003, **15**:706–720.
46. Piper P, Mahé Y, Thompson S, Pandjaitan R, Holyoak C, Egner R, Mühlbauer M, Coote P, Kuchler K: **The pdr12 ABC transporter is required for the development of weak organic acid resistance in yeast.** *EMBO J* 1998, **17**:4257–4265.
47. Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S: **MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods.** *Mol Biol Evol* 2011, **28**:2731–2739.
48. McDonald JH, Kreitman M: **Adaptive protein evolution at the Adh locus in *Drosophila*.** *Nature* 1991, **351**:652–654.
49. Erny C, Raoult P, Alais A, Butterlin G, Delobel P, Matei-Radoi F, Casaregola S, Legras JL: **Ecological success of a group of *Saccharomyces cerevisiae*/*Saccharomyces kudriavzevii* hybrids in the northern european wine-making environment.** *Appl Environ Microbiol* 2012, **78**:3256–3265.
50. Alexandre H, Charpentier C: **Biochemical aspects of stuck and sluggish fermentation in grape must.** *J Ind Microbiol Biotechnol* 1998, **20**:20–27.
51. Vargas RC, Tenreiro S, Teixeira MC, Fernandes AR, Sá-Correira I: ***Saccharomyces cerevisiae* multidrug transporter Qdr2p (Yil121wp): localization and function as a quinidine resistance determinant.** *Antimicrob Agents Chemother* 2004, **48**:2531–2537.
52. Ríos G, Cabedo M, Rull B, Yenush L, Serrano R, Mulet JM: **Role of the yeast multidrug transporter Qdr2 in cation homeostasis and the oxidative stress response.** *FEMS Yeast Res* 2012, **13**(1):97–106.
53. Vargas RC, García-Salcedo R, Tenreiro S, Teixeira MC, Fernandes AR, Ramos J, Sá-Correira I: ***Saccharomyces cerevisiae* multidrug resistance transporter Qdr2 is implicated in potassium uptake, providing a physiological advantage to quinidine-stressed Cells.** *Eukaryot Cell* 2007, **6**:134–142.
54. Waterhouse AL: **Wine Phenolics.** *Ann N Y Acad Sci* 2002, **957**:21–36.
55. Kim JH, Campbell BC, Mahoney NE, Chan KL, Molyneux RJ: **Identification of phenolics for control of *Aspergillus flavus* using *Saccharomyces cerevisiae* in a model target-gene bioassay.** *J Agric Food Chem* 2004, **52**:7814–7821.
56. Pan Z, Agarwal AK, Xu T, Feng Q, Baerson SR, Duke SO, Rimando AM: **Identification of molecular pathways affected by pterostilbene, a natural dimethylether analog of resveratrol.** *BMC Med Genomics* 2008, **1**:7.
57. Jothi R, Balaji S, Wuster A, Grochow JA, Gsponer J, Przytycka TM, Aravind L, Babu MM: **Genomic analysis reveals a tight link between transcription factor dynamics and regulatory network architecture.** *Mol Syst Biol* 2009, **5**:294.
58. Galeote V, Novo M, Salema-Oom M, Brion C, Valério E, Gonçalves P, Dequin S: **FSY1, a horizontally transferred gene in the *Saccharomyces cerevisiae* EC1118 wine yeast strain, encodes a high-affinity fructose/H⁺ symporter.** *Microbiology (Reading, Engl)* 2010, **156**:3754–3761.
59. Marullo P, Bely M, Masneuf-Pomarède I, Pons M, Aigle M, Dubourdieu D: **Breeding strategies for combining fermentative qualities and reducing off-flavor production in a wine yeast model.** *FEMS Yeast Res* 2006, **6**:268–279.
60. Bely M, Sablayrolles J-M, Barre P: **Automatic detection of assimilable nitrogen deficiencies during alcoholic fermentation in oenological conditions.** *J Ferment Bioeng* 1990, **70**:246–252.
61. Calull M, Marcé RM, Borrull F: **Determination of carboxylic acids, sugars, glycerol and ethanol in wine and grape must by ion-exchange high-performance liquid chromatography with refractive index detection.** *J Chromatogr, A* 1992, **590**:215–222.
62. Güldener U, Heck S, Fielder T, Beinhauer J, Hegemann JH: **A new efficient gene disruption cassette for repeated use in budding yeast.** *Nucleic Acids Res* 1996, **24**:2519–2524.
63. R Development Core Team: **R: A Language and Environment for Statistical Computing.** Vienna, Austria: R Foundation for Statistical Computing; 2008. URL <http://www.R-project.org>. ISBN ISBN 3-900051-07-0.
64. Smyth GK, Speed T: **Normalization of cDNA microarray data.** *Methods* 2003, **31**:265–273.
65. Smyth GK: **Linear models and empirical bayes methods for assessing differential expression in microarray experiments.** *Stat Appl Genet Mol Biol* 2004, **3**. Article3.
66. Smyth G: **limma: linear models for microarray data.** In *Bioinformatics and Computational Biology Solutions Using R and Bioconductor*. Edited by Gentleman R, Carey VJ, Huber W, Irizarry RA, Dudoit S. New York: Springer; 2005:397–420.
67. Quan H, Shih WJ: **Assessing reproducibility by the within-subject coefficient of variation with random effects models.** *Biometrics* 1996, **52**:1195–1203.
68. Benjamini Y, Drai D, Elmer G, Kafkafi N, Golani I: **Controlling the false discovery rate in behavior genetics research.** *Behav Brain Res* 2001, **125**:279–284.
69. De Hoon MJL, Imoto S, Nolan J, Miyano S: **Open source clustering software.** *Bioinformatics* 2004, **20**:1453–1454.
70. Saldanha AJ: **Java treeview—extensible visualization of microarray data.** *Bioinformatics* 2004, **20**:3246–3248.
71. Broman KW, Wu H, Sen S, Churchill GA: **R/qtl: QTL mapping in experimental crosses.** *Bioinformatics* 2003, **19**:889–890.
72. Kimura M: **A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences.** *J Mol Evol* 1980, **16**:111–120.

doi:10.1186/1471-2164-14-681

Cite this article as: Brion et al.: Differential adaptation to multi-stressed conditions of wine fermentation revealed by variations in yeast regulatory networks. *BMC Genomics* 2013 **14**:681.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

