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Enhancing Transcriptomic Data Mining with Semantic Ranking: Towards a new Functional Spectral Representation.

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Abstract. In biomedical domains, high throughput technologies produce large amount of transcriptomic data used for studying comportment of genes. The analysis and the interpretation of such data require important databases and efficient mining methods, in order to extract specific biological functions belonging to a group of genes of an expression profile. To this aim, we propose here a new approach for mining transcriptomic data combining domain knowledge and classification methods. Firstly, we propose the definition of Fuzzy Differential Gene Expression Profiles (FD-GEP) based on fuzzy classification and a differential definition between the considered biological situations. Secondly, we will use our previously defined efficient semantic similarity measure (called IntelliGO), that is applied on Gene Ontology (GO) annotation terms, for computing semantic and functional similarities between genes of the resulting FD-GEP and well known genetic markers involved in the development of cancers. After that, the similarity matrices will be used to introduce a novel Functional Spectral Representation (FSR) calculated through a semantic ranking of genes regarding their similarities with the tumoral markers. The FSR representation should help expert to interpret by a new way transcriptomic data and infer new genes having similar biological functions regarding well known diseases.

Availability: The semantic similarity measure and the ranking method are available at <http://plateforme-mbi.loria.fr/intelligo/ranking.php>.

1 Introduction

Nowadays, the pharmaceutical industries require large volume of biological data and need a sophisticated computational methods to manage and extract relevant knowledge. Recently, DNA microarrays were used for measuring the expression levels of thousands of genes under various biological conditions. Hence, gene expression data analysis proceeds in two steps: Firstly, expression profiles are produced by grouping genes displaying similar expression levels under biological situations [1]. Secondly, a functional analysis, based on functional annotations, is applied on genes sharing the same expression profile, in order to identify their relevant biological functions [2]. In fact, the main goal of this functional analysis

is to identify and characterize genes that can serve as diagnostic signatures or prognostic markers for different stages of a disease. One of the interesting source of functional annotations in the biological domain is the Gene Ontology (GO) [3]. The interpretation of transcriptomic data requires efficient mining methods for extracting specific biological functions belonging to genes of an expression profile. In this context, we introduce here, by an innovative way, a cascade of methods for mining such data combining domain knowledge and classification methods. Firstly, we propose the definition of Fuzzy Differential Gene Expression Profiles (FD-GEP) based on fuzzy classification and a differential definition of the expression between the considered biological situations. This classification approach affects genes through fuzzy sets when there are differential physiological relations between the studied biological situations. Secondly, we will use our previously defined efficient semantic similarity measure (called IntelliGO), that is applied on Gene Ontology (GO) annotation terms, for computing semantic and functional similarities between genes of resulting FD-GEP and set of well known genetic markers involved in the development of cancers. After that, the similarity matrices will be used to introduce a novel Functional Spectral Representation (FSR) calculated through a semantic ranking of genes regarding their similarities with the tumoral markers. This new paradigm for visualizing expression data displays as a bar code, genes of a given FD-GEP w.r.t. disease markers. Genes that are at the top of the sorted list are functionally similar to these markers and can hence be explored by biologists to verify experimental hypothesis.

This paper is organized as follows. The next section, outlines the presentation of the used dataset and introduce the new proposed method for profiles extraction. The next subsections present the new gene functional analysis based IntelliGO similarity and semantic Ranking. Finally, we discuss in the last section the relevance of the obtained results of the proposed methods.

2 Material and methods

2.1 DNA microarrays dataset

In this work, we will use a list (L) of 222 differentially expressed genes of colorectal cancers. An Affymetrix HGU133+¹ microarray was used for experiments. In this dataset, we dispose of three biological situations in the gene expression matrix (M) that correspond to three biological samples: (i) healthy tissue (normal); (ii): tumor tissue (cancer); (iii) cell line. We name these situations: S_1 , S_2 , S_3 respectively. Each situation represents the average of multiple replicates and multiple specimens in each type of tissue during experiences [4]. An example of the expression data in the matrix M is illustrated in Table 1. The expression value for a given gene g from a set of genes G in a situation S_i is given by ν_{si} . The selected 222 genes represent a significant fold change observed between S_2 and S_1 . Thus biologists are interested by genes for which the expression varies between these two situations, i.e., found deregulated in cancer tissues.

¹ www.affymetrix.com/products_services/arrays/specific/hgu133plus.affx

Gene	Healthy : S_1	Cancer: S_2	Cell line: S_3
KIAA1199	33,6	827,87	735,75
FOXQ1	65,36	1240,21	2631,71
PSAT1	89,03	1019,0	3025,66
CLDN1	12,15	119,9	78,5
SLC6A6	56,6	551,1	568,6
...	...	...	...
Gene g	ν_{s1}	ν_{s2}	ν_{s3}
...	...	...	...
PSAT1	113,1	407,1	1258,0

Table 1. Example of the expression matrix M of the 222 genes relative to colorectal cancer, used in this study. This matrix is used for extracting gene expression profiles.

2.2 Definition of the Fuzzy Differential Genes Expression Profiles: FD-GEP

In the state-of-the-art methods for extracting expression profiles [1], there is no prior exploitable knowledge between the nature of biological situations. However, the interpretation of groups of genes in an expression profile is usually guided by hypotheses related to the objectives of the study. This is the case in particular when relations exist between the biological situations. Such kind of relations could be temporal when expression levels are measured at different time stamp, or kinetic when a kind of a tissue is considered but in different physiological states (e.g. stages of tumor). In this last case, the differences existing between pairs of situations may interest biologists, and it would be interesting to regroup together genes with the same variations between two situations regardless of the level of expression they have each in one situation. This observation conducts us to introduce here the notion of a priori definition of *Differential Genes Expression Profiles* (D-GEP). It is the first contribution of this paper. Constructed from pairs of situations chosen by the user, these profiles can be considered as combinations, for each pair of situations, of the *Differential Expression Sets* (DES), to regroup genes having similar variations (over-expression, or even under-expression) between two situations. The membership of a gene to a given profile begins by studying its membership to a particular DES for each pair of situations. Here we use a fuzzy modeling of the expression level variations between two situations to consider noise in the data and provide an opportunity for a gene to belong to more than one profile. With our data set, and having three situations, we have 3 possible pairs of situations (S_3, S_1) (S_2, S_1) and (S_3, S_2) , respectively. For n situations, we can have $n(n-1)/2$ pairs of situations. Identifying a D-GEP for a gene g leads us to affect this gene, for each pair of situation (S_i, S_j) , either in the set of genes over-expressed in S_i with respect to S_j (which we denote $\text{Over}_{i,j}$), or in the set of genes under-expressed in S_i compared to S_j (which we denote $\text{Under}_{i,j}$), or rather in set of genes with similar expression in S_i w.r.t. S_j (which we denote $\text{Iso}_{i,j}$). By extension, the sets $\text{Over}_{i,j}$, $\text{Under}_{i,j}$, $\text{Iso}_{i,j}$ are called *Differential Expression Sets* (DES), and allow to represent the differential expression of genes between two situations of interest S_i and S_j . Consequently, the formal definition of a D-GEP is a K-uplet of DES, where K is the number of pairs of situations. For example, in our case, the profile $(\text{Over}_{3,1}, \text{Over}_{2,1}, \text{Under}_{3,2})$, is a D-GEP in which genes are: over expressed in Cell Line

(S_3) w.r.t. Normal tissue (S_1), over expressed in cancer (S_2) w.r.t. Normal tissue (S_1), and under expressed in the Cell Line (S_3) w.r.t. Cancer (S_2). As we said before, the affectation of genes in the different DES is performed with fuzzy logic. Firstly, we need to define a value of variation of expression noted $\xi_{i,j}$ to represent the difference of the expression of a gene g in two situations (S_i, S_j): $\xi_{i,j}(g) = \frac{\nu_{s_i} - \nu_{s_j}}{\min_{k=1..n} \nu_{s_k}}$ for all i, j in k situations. The proposed fuzzy model conducts to define a threshold value σ for the expression variation value $\xi_{i,j}(g)$, and for each pair of situations (S_i, S_j). Three membership functions of three fuzzy sets allow to define the affectation of genes to DES depending on the value of $\xi_{i,j}(g)$. These membership functions are defined as:

$$Over_{i,j} : G \longrightarrow [0, 1] \quad Over_{i,j}(g) = \begin{cases} 1 & \text{if } \xi_{i,j}(g) \geq \sigma \\ \frac{1}{\sigma} \times \xi_{i,j}(g) & \text{if } 0 < \xi_{i,j}(g) < \sigma \\ 0 & \text{otherwise} \end{cases}$$

$$Under_{i,j} : G \longrightarrow [0, 1] \quad Under_{i,j}(g) = \begin{cases} 1 & \text{if } \xi_{i,j}(g) \leq -\sigma \\ \frac{-1}{\sigma} \times \xi_{i,j}(g) & \text{if } -\sigma < \xi_{i,j}(g) < 0 \\ 0 & \text{otherwise} \end{cases}$$

$$Iso_{i,j} : G \longrightarrow [0, 1] \quad Iso_{i,j}(g) = \begin{cases} 0 & \text{if } \xi_{i,j}(g) < -\sigma \text{ or } \xi_{i,j}(g) > \sigma \\ \frac{1}{\sigma} \times \xi_{i,j}(g) + 1 & \text{if } -\sigma < \xi_{i,j}(g) < 0 \\ \frac{-1}{\sigma} \times \xi_{i,j}(g) + 1 & \text{if } 0 < \xi_{i,j}(g) < \sigma \end{cases}$$

The defuzzification process allows the final classification of a gene g to one or more DES in accordance to a given threshold value μ . This is done with the following constraint: if the value taken by function $Over_{i,j}(g)$, $Under_{i,j}(g)$ or $Iso_{i,j}(g)$ is $\geq \mu$, thus the gene is classified in the corresponding DES. The figure (1) represents the three membership functions $Over_{i,j}$, $Under_{i,j}$, $Iso_{i,j}$. For example, for the gene g , we have $Over_{i,j}(g) = 0.25$, $Under_{i,j}(g) = 0$, $Iso_{i,j}(g) = 0.75$. We observe that for $\mu = 0.5$, this gene is classified in the DES $Iso_{i,j}$ only, otherwise for $\mu = 0.25$ it will be affected both in $Iso_{i,j}$ with membership probability of 0.75 and $Over_{i,j}$ with a complementary membership value of 0.25. We can remark that with this modelization, the bi-classification criteria of a gene in two DES is possible for $\mu \leq 0.5$. Hence, a D-GEP is a combination of DES, and the fuzzy modelization introduced here allows to genes to be affected to more than one D-GEP. By consequence a Fuzzy differential genes expression profile (FD-GEP) is a D-GEP where the affectation of genes in DES is performed with fuzzy logic. Note that, with 3 situations, and with 3 DES, we can obtain 27 possible FD-GEP. As we can observe, there are two parameters that can be taken into account during the classification algorithm, namely: σ and μ . The different values affected to these variables depend on the used datasets. We performed an exploratory analysis by testing different values for $\mu = \{0.1, 0.2, 0.3, 0.4, 0.5\}$ and $\sigma = \{0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1\}$. Each pair of values of the two parameters were tested for extracting the

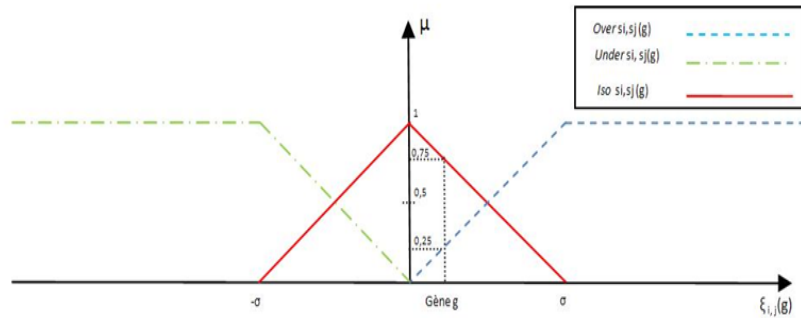


Fig. 1. Representation of the membership functions defining the affectation of a gene to the differential expression sets $Over_{i,j}$, $Under_{i,j}$, $Iso_{i,j}$ used for defining the FD-GEP profiles.

possible FD-GEP profiles. Results for the different combination are not shown here. Although, we used some prior knowledge to eliminate certain values that generate outliers FD-GEP profiles. For example, for $\sigma \geq 0.6$ we observed that some genes were classified in the DES $Iso_{2,1}$ (same expression in cancer and normal tissues), even if in our dataset the genes are differentially expressed between the two situations (S2) and (S1). We decided after multiple testing, to keep the combination $(\mu, \sigma) = (0.3, 0.4)$ which expects a reasonable number of genes in the obtained profiles. With this pair of values, and from the list of 222 colorectal cancer genes, we extracted the FD-GEP expression profiles presented in Table 2. The number of genes in each profile is displayed in the diagonal of the matrix, while overlaps (due to fuzzy classification) are observed between some profiles in terms of number of shared genes are displayed in the rest of cells of the matrix.

Name of the FD-GEP	Definition of the FD-GEP	Profile 1	Profile 2	Profile 3	Profile 4	Profile 11	Profile 13	Profile 14	Profile 15	Profile 20	Profile 21
Profile 1	$Over_{2,1}, Over_{3,1}, Over_{3,2}$	51		2							
Profile 2	$Over_{2,1}, Over_{3,1}, Under_{3,2}$		108	17						24	1
Profile 3	$Over_{2,1}, Over_{3,1}, Iso_{3,2}$			30						1	1
Profile 4	$Over_{2,1}, Under_{3,1}, Over_{3,2}$				1						
Profile 11	$Under_{2,1}, Over_{3,1}, Under_{3,2}$					1					
Profile 13	$Under_{2,1}, Under_{3,1}, Over_{3,2}$						9		1		
Profile 14	$Under_{2,1}, Under_{3,1}, Under_{3,2}$							7	1		
Profile 15	$Under_{2,1}, Under_{3,1}, Iso_{3,2}$								5		
Profile 20	$Iso_{2,1}, Over_{3,1}, Under_{3,2}$									56	
Profile 21	$Iso_{2,1}, Over_{3,1}, Iso_{3,2}$										1

Table 2. Distribution of genes in the obtained expression profiles. Note that if a cell is empty then the two corresponding FD-GEP profiles do not share any gene. The diagonal represents the number of genes in each FD-GEP.

2.3 Gene Expression Functional Analysis

Calculating semantic and functional similarity between genes: A lot of functional analysis methods have been proposed using either statistical enrichment [5] or gene functional clustering [6]. In this last case, a semantic similarity is used for grouping genes regarding Gene Ontology (GO) functions [3]. We admit that the performance of the used similarity will have an impact on the clustering results. Recently, we proposed an hybrid functional analysis method combining both fuzzy clustering and statistical enrichment analysis [7–9], using our *IntelliGO* measure [10]. Our second contribution in this paper (after having defined FD-GEP), is to propose a complementary visualization method that should help biologists to investigate interesting genes during the transcriptomic data study. The principle of the technique is detailed below.

In the rest of the experiments, we will use the same list of 222 colorectal cancer genes, for computing the IntelliGO similarity matrices. As a first step, and in order to have a global overview of the distribution of the biological annotations of the used list of genes, we produced a Heatmap with a two way hierarchical clustering. The results are shown in figure (2). Despite the change in color intensity in different regions of the heatmap, several clusters can be distinguished in its diagonal. Homogeneous cluster in the upper left of the heatmap and with very little cross similarity with other genes in the data set has been studied from the point of view of its gene content. The strong functional similarity between these genes can be explained by recurrent Biological Process annotations on transport processes. Transport processes are important in the physiology of the digestive system and cluster genes are already known to be deregulated in colorectal cancer. This is the case of gene AQP8 aquaporin 8 whose expression is no longer detectable in colorectal tumors [11]. In fact AQP8 gene is found in the FD-GEP Profile_14 corresponding to genes under-expressed in tumor vs the normal situation. Another gene (ATP11A) is also present in the functional cluster analyzed here, with an annotation transport of phospholipids, and was recently described as a new predictive marker of metachronous metastasis of colorectal cancer [12]. Associated in our study to FD-GEP Profile_1, it actually appears as over-expressed in tumor position relative to the healthy situation. These two genes can be considered as positive controls, confirming the validity of classification results.

Functional Spectral Representation: A Genes functional ranking approach: As reported above, to enhance the analysis of the transcriptomic data, we propose a new functional spectral representation of genes. The main idea of the proposition is summarized in the Figure (3). Firstly, we proceed by ranking with descending order (from most to less similar) an input list of genes regarding their IntelliGO functional semantic similarity with given well known biological markers. After the ranking step, we identify for a given FD-GEP profile, the position of its genes in the ranked list. The position of each gene of this profile is marked with a red line. Blue line indicates the 0.5 functional similarity threshold. The main objective here is to verify if the genes belonging to the profile

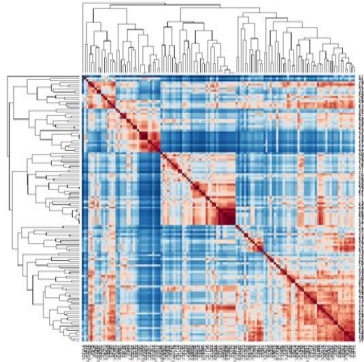


Fig. 2. Generated Heatmap with 222 colorectal cancer.

(marked with red lines) are top ranked in the list or not, i.e., the number of red lines over the blue line is important or not. If it is the first case, these genes are judged to be functionally similar to the well-known biological markers and therefore they could be suggested to biologists for further analysis.

In preliminary experiences, we have selected 6 genes involved in target WNT signaling pathway² known as transcriptionally repressed in colorectal cancer [13]. Genes of this pathway are known as contributors in mutations in degenerative diseases and cancers. Namely we have chosen: AXIN2 (The Axin-related protein), CD44 (Cell-surface glycoprotein), MET (MNNG HOS Transforming gene), MYC (v-myc myelocytomatosis viral oncogene homolog), SOX9 (SRY (sex determining region Y)-box 9) and ASCL2.

Results for FD-GEP profile1 are presented in Figure (4). We can observe that with AXIN2, MET, MYC, SOX9 markers, the blue line (50% of similarity) is positioned in low levels of the ranked list. It means that genes of this profile are very functionally similar to these markers, in particular with MET and MYC ($\geq 60\%$ of genes of the profile) as it is shown in figure 5. We recall that these two biomedical markers are confirmed to be catalyst in the colorectal cancer, and we know that the studied data set is relative to this kind of cancer [14]. Thus, these results confirm the pertinence of the proposed approach. For deepening the analysis, we processed the same analysis for FD-GEP 2, 3, 14, 20 since genes in these profiles are dysregulated in the cancer situation vs normal (either Over or Under in S_2 vs S_1). We calculated for each profile its FSR and also the average percentage of its genes having functional similarity w.r.t. the used cancer biomedical markers over than 0.50. Results are shown in figure 5.

We can observe that all FD-GEPs except Profile 14 have similar functional background regarding the used markers, and it appears very clearly that genes of the used data set share biological processes that are very similar to MET and

² <http://www.genome.jp/kegg/pathway/hsa/hsa04310.html>

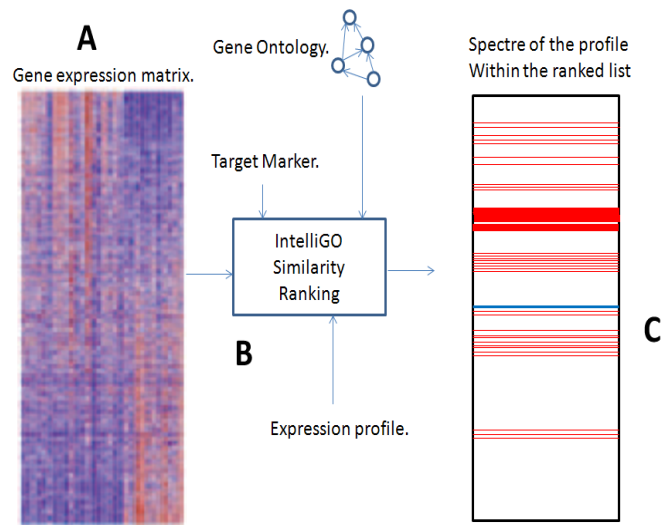


Fig. 3. A Functional Spectral Representation (FSR) overview illustrating the method. A: An expression data matrix is used as input. B: The IntelliGO functional similarity is calculated within all genes of the input list and a given bio-marker. C: The ranked list is displayed by highlighting with red lines genes of a studied expression profiles. A blue line is used to identify the 0.5 functional similarity threshold.

MYC cancer genes (more than 40 %). Hence, we can suggest to biologists the top positioned genes in the produced ranked lists of the FSRs of each profile, in order to verify their sequence similarity with the used markers. It could be then possible to extend the WNT signaling pathway with those genes, and verify other hypothesis of their involvement in the colorectal cancers.

3 Conclusion and perspectives

In this paper, we proposed data mining methods combining classification and visualization for analyzing transcriptomic data by an innovative way. In a first step, we propose a new definition of the Fuzzy Differential Genes Expression Profiles FD-GEP, for classifying genes through fuzzy sets when there are differential physiological relations between biological situations. After the step of profiles extraction, a gene functional analysis is essential to give sense to the affected genes in the profiles. We propose a new functional analysis approach, with a new paradigm for visualizing expression data through a process of semantic ranking regarding interesting disease markers. Indeed, the proposed FSR displays as a bar code, genes of a given FD-GEP w.r.t. disease markers. Genes that are at the top of the sorted list are functionally similar to these markers and can hence be explored by biologists to verify experimental hypothesis.

Further, we would consider the idea of performing the same study on genes be-

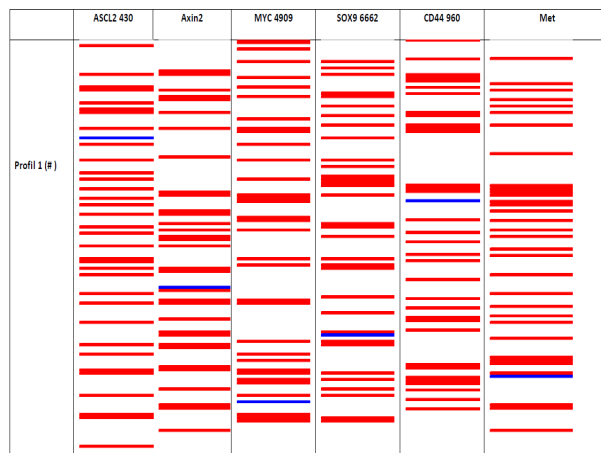


Fig. 4. Functional Spectral Representation (FRS) using genes of FD-GEP Profile 1.

longing to other disease processes. Indeed, the OMIM database (Online Mendelian Inheritance in Man)³ provides markers for different types of diseases that are listed in this database. One can test these markers with different lists of gene expression data.

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³ <http://www.omim.org/>

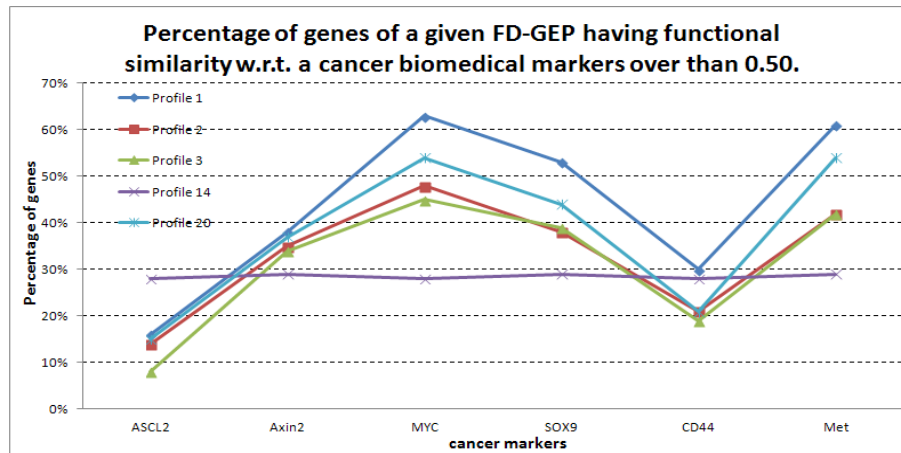


Fig. 5. Average percentage of genes of an FD-GEP having functional similarity w.r.t. a cancer biomedical markers over than 0.50. Values are varying from a marker to another for the considered FD-GEPs. We can observe that majority of the FD-GEP have similar biological behavior regarding their functional similarities with biological functions of the markers.

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