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Application of HS-SPME–GC-MS-derived variables for monitoring ripening-induced changes in cheese volatile compounds

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Abstract Headspace solid-phase microextraction–gas chromatography–mass spectrometry (HS-SPME–GC-MS) and multivariate statistical analysis (MVA) are widely employed in the analyses/comparison of cheese volatiles. One shortcoming of this approach is that the SPME fiber-coat polarity has a significant effect on the adsorption of volatiles and thus, on the results of MVA. The aim of this study was to test the applicability of different sets of MVA variables, obtained from HS-SPME–GC-MS experiments using the non-polar polydimethylsiloxane (PDMS) and the polar polyacrylate (PA) fibers, in the profiling of cheese volatile compounds and tracking ripening-induced changes. The volatile profiles and ripening-induced changes of two different types of Pasta-Filata “Pirotski kačkavalj” cheese (prepared from raw ewe’s or cow’s milk) were assessed at four different stages of ripening (1, 5, 20, and 30 days). The HS-SPME–GC-MS–MVA results showed that the distribution of volatile compounds into classes gives valuable information concerning cheese type and ripening phase. The extraction with a PA fiber gives a more realistic cheese volatile profile when the relative ratio of the constituent classes is in question, whereas the PDMS fiber is better in recognizing the contribution/concentration of single dominant volatile compounds. For a better and more complete description/profiling of cheese volatiles, we propose the use of combined (transformed) variables that include the information obtained from both PA and PDMS extraction experiments.

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利用HS-SPME-GC-MS监测干酪成熟过程中挥发性化合物的变化

摘要：顶空固相微萃取-气质联用(HS-SPME-GC-MS)和多元统计分析(MVA)广泛应用于分析和比较干酪的挥发性化合物。该方法的一个缺点是SPME纤维外套的极性对挥发性化合物的吸附影响较大,因此也影响了MVA分析的结果。本论文目的是应用不同组合的MVA变量,这些变量是通过HS-SPME-GC-MS实验并采用非极性聚二甲硅氧烷(PDMS)和极性聚丙烯酸酯(PA)纤维萃取跟踪干酪成熟过程中引发的挥发性化合物变化。分析了两种不同类型的Pasta-Filata和“Pirotski kačkavalj”干酪(来自羊奶和牛奶)在四个不同成熟阶段(1,5,20,30 d)所产生挥发性化合物的变化。HS-SPME-GC-MS-MVA实验结果对干酪的类型和成熟阶段挥发性化合物的分布给出了非常重要的信息。利用PA纤维萃取对需要讨论干酪中各种挥发性化合物组分的相对比例给出了更理想的结果。然而PDMS纤维萃取对了解单个主要的挥发性化合物的分布和浓度方面更有优势。为了更好更全面的描述干酪的挥发性化合物,建议将PA和PDMS两种萃取实验的信息结合起来分析干酪成熟过程中挥发性化合物的变化。

Keywords Volatile profiles · Principal component analysis · Agglomerative hierarchical cluster analysis · Cheese ripening · Pasta-Filata “Pirotski kačkavalj” cheese

关键词 挥发性成分 · 主成分分析 · 凝聚层次聚类分析 · 干酪的成熟 · 帕斯塔费拉塔乳酪

1 Introduction

The unique flavor of different cheese varieties is due to the complex balance of the taste, aroma, and texture characteristics. Volatile compounds play a particularly important role in this respect (Arora et al. 1995; Januszkiewicz et al. 2008; Mondello et al. 2005; Peres et al. 2001; Pinho et al. 2002; Ziino et al. 2005). However, during cheese ripening, the cheese volatile/flavor profiles can change dramatically due the complex biochemical and chemical processes of lipolysis, proteolysis, and glycolysis (Fox and Wallace 1997). Due to the complexity of cheese volatile profiles (mixtures of a large number of compounds belonging to different chemical classes: alcohols, esters, acid, ketones, etc.), the monitoring of ripening-induced changes is not a straightforward task. In fact, for reasons of reliability and clarity, it is often necessary to perform multivariate statistical analyses (MVA, e.g., agglomerative hierarchical cluster analysis (AHC) and principal component analysis (PCA)) as a means of differentiating personal bias from statistical significance (Delgado et al. 2010). The variables used for MVA analyses (input data set) of cheese volatile profiles are usually obtained from gas chromatography (GC) and gas chromatography-mass spectrometry (GC-MS) experiments (Arora et al. 1995; Januszkiewicz et al. 2008; Mondello et al. 2005; Peres et al. 2001; Pinho et al. 2002, 2003; Ziino et al. 2005). Headspace solid-phase microextraction (HS-SPME) is a widely employed extraction method for GC and GC-MS analyses of cheese volatiles due to its relative simplicity and cost-effectiveness (Januszkiewicz et al. 2008; Mondello et al. 2005). Nevertheless, it has previously been demonstrated that the fiber-coat polarity has a significant effect on the adsorption of volatiles (Mondello et al. 2005; Pinho et al. 2003). Hence, one could justly formulate a question—Is it really reasonable to use data obtained from this type of sampling/analyses (HS-SPME-GC-MS) for the comparison (variables for MVA) of different types of cheese, or for monitoring ripening-induced changes in cheese volatiles? And, if it is so, does the extraction with one particular type of fiber give

better, more reliable, and more reproducible results? Furthermore, is it possible to solve this problem (fiber-dependent differences in the volatile profile of a sample) by using some transformed/combined variables for MVA, which would include information obtained from experiments based on the usage of different SPME stationary phases?

Bearing all this in mind, the main aim of this research was to test the applicability of different sets of MVA variables to profile cheese volatile compounds and monitor ripening-induced changes. The different sets of MVA variables include: the original data (relative amounts individual volatile compounds, obtained directly from HS-SPME–GC-MS experiments); transformed data (relative amounts of each class of compounds); and combined data (created by averaging the relative amounts of individual compounds obtained using both types of stationary phases).

HS-SPME–GC-MS analyses of Pasta Filata cheeses were performed using two SPME fibers of different polarity (polar: polyacrylate (PA) and non-polar: polydimethylsiloxane (PDMS)). “Pirotski kačkavalj” cheeses made from cow’s and ewe’s milk were analyzed at four stages of ripening. Results on the chemical composition of the volatile profiles of the two types of Pirotski kačkavalj cheese are reported here for the first time.

2 Materials and methods

2.1 Manufacture and sampling of cheese

Three batches of 12 0.5-kg cheeses (two different types of Pasta Filata “Pirotski kačkavalj” cheese) were manufactured in a small, certified dairy factory, Dairy School “Dr Obren Pejić”, in Pirot (Southeast Serbia) in three sequential periods within the cheese making season (i.e., November, February, and May) from raw ewe’s (EC) and cow’s milk (CC) according to traditional practices (Milosavljević et al. 2010). Three cheeses of each batch were randomly selected on the day of manufacture and after 5, 20, and 30 days of ripening. Samples of each type of cheese were taken from the inner part of the cheese and were stored at ~ -18 °C (during 1–5 weeks) prior to analysis.

2.2 Extraction of volatile compounds

Cheese volatile compounds were extracted by SPME using two types of commercially available fibers (PDMS, 100 μm film thickness and PA, 85 μm film thickness). The SPME fibers were housed in their manual holder (Supelco, Bellefonte, PA, USA).

All extractions were performed as follows—2 g of each cheese sample (eight samples in total) was placed in a 50-mL headspace glass vial sealed tightly with a crimp cap and PTFE/silicone septum. The vial was immersed at 40 mm in a thermostated waterbath at 60 °C, and the volatile compounds were extracted over 20 min using the two SPME fiber coatings maintained 10 mm above the molten samples which were continuously stirred with a magnetic stirring bar. After extraction, the samples were directly desorbed (without delay) by manual injection into the GC injection port, held at 250 °C, using a specially designed splitless 0.75 mm ID inlet liner to suit these purposes (also purchased from Supelco).

2.3 Gas chromatography-mass spectrometry (GC-MS)

Separation and identification of the volatile compounds extracted as described above were performed by GC-MS. The instruments used and the experimental conditions are identical to those described in Radulović and Blagojević (2010), except that the samples were manually injected by inserting the saturated fiber into the inlet liner for 10 min. The GC oven temperature was held at 45 °C for 5 min, then raised to 250 °C at a heating rate of 10 °C.min⁻¹, and then held isothermally for 10 min at 250 °C (carrier gas: helium at 1.0 mL.min⁻¹). The relative abundance (percent peak area) of three independent replicate samples of each compound identified by GC-MS without the use of correction factors is given in Table S1. Qualitative analysis was performed in the same way as described in Radulović and Blagojević (2010) and was based on (a) comparison of the mass spectra, (b) order of elution, and (c) co-injection of authentic standards (the corresponding pure compounds were added/spiked to cheese samples prior to HS-SPME). Standard (pure) compounds were obtained from Merck, Germany or Sigma-Aldrich, USA. All analyses were performed in triplicate, and there were no statistically significant differences between the obtained results.

2.4 Multivariate statistical analysis (MVA)

PCA and AHC were performed using the Excel program plug-in XLSTAT version 2011.3.02 and according to Radulović and Blagojević (2010). AHC was determined using Pearson dissimilarity where the aggregation criterion were simple linkage, unweighted pair-group average and complete linkage, and Euclidean distance where the aggregation criterion were weighted pair-group average, unweighted pair-group average, and Ward's method. Euclidian distance dissimilarities of the analyzed samples were within the interval [0, 2,000–12,000]. PCA of the Pearson (*n*) type was performed. Axes (F1 and F2 factors—the first and second principal components) of the resulting biplots refer to the ordination scores obtained for the samples. Both MVA methods were applied utilizing following types of variables: PA and PDMS original variables (relative amounts of individual compounds, as determined from HS-SP–GC-MS experiments with PA and PDMS fibers), PA and PDMS transformed variables (relative amounts of classes of compounds, obtained from the experiments with PA and PDMS fibers), and original-combined and transformed-combined variables (these were created by averaging (weight factor of 0.5) of the relative amounts of individual constituents/class of constituents obtained from the experiments with both types of stationary phases).

3 Results and discussion

3.1 Volatile compounds of the two types of Pirotski kačkavalj cheese

The main compounds identified in the cheeses are listed in Table 1 (all identified compounds, 73 in total, are given in the [Electronic supplementary material](#)). The major classes of volatile compounds identified in both cheese types were short- and medium-chain fatty acids, alcohols, esters, and ketones. Some of the identified

Table 1 The major compounds (relative abundance $\geq 2\%$) identified by HS-SPME–GC-MS analysis of the volatile fractions of two types of Pirotski kackavalji cheese prepared from ewes' (EC) and cows' milk (CC) at four different ripening stages

RT ^a	Method of identification	RT (min)	Compound	Relative abundance (peak area, %) ^c															
				EC1 ^b				EC2				EC3				EC4			
				PDMS	PA ^e	PDMS	PA												
Acids^f																			
600–700	MS, RI, CoI	1.73	Acetic acid	nd	tr	2.5	4.0	1.6	7.1	5.1	16.2	nd	nd	20.4	2.7	14.3	5.9	16.8	
831	MS, RI, CoI	5.78	Butanoic acid	0.6	12.0	6.3	35.5	14.3	42.0	6.2	31.2	tr	1.5	tr	2.0	4.2	17.3	2.2	13.1
1,010	MS, RI, CoI	10.64	Hexanoic acid	tr	10.4	6.2	15.6	12.2	17.6	6.9	23.2	2.5	18.0	tr	2.3	11.5	tr	6.9	
1,182	MS, RI, CoI	13.86	Octanoic acid	tr	tr	3.4	1.7	4.5	1.6	tr	3.5	tr	tr	1.7	nd	tr	nd	nd	
			Subtotal (%)	0.6	22.4	19.1	61.2	32.9	69.4	18.2	74.9	2.5	19.5	1.7	22.4	9.2	43.1	8.1	36.8
Esters																			
612	MS, RI, CoI	2.20	Ethyl acetate	3.0	nd	tr	tr	1.6	tr	7.2	nd	1.7	0.8	4.5	tr	4.9	tr	tr	3.5
801	MS, RI, CoI	5.88	Ethyl butanoate	tr	nd	tr	tr	0.5	2.2	tr	tr	nd	nd	nd	tr	nd	nd	nd	nd
950	MS, RI	9.80	Ethyl 3-oxobutanoate	nd	nd	nd	0.1	nd	nd	nd	0.3	7.0	5.7	0.6	nd	nd	0.7	nd	tr
1,002	MS, RI, CoI	10.78	Ethyl hexanoate	1.2	nd	2.8	nd	5.7	2.4	1.8	tr	1.0	0.0	tr	nd	0.7	0.3	nd	nd
1,047	MS, RI, CoI	11.83	Butyl 3-methylbutanoate	nd	nd	0.7	0.1	0.3	nd	tr	tr	nd	nd	nd	nd	nd	2.1	nd	nd
			Subtotal (%)	5.3	nd	7.3	0.7	12.8	5.3	9.0	0.6	9.7	7.6	5.1	tr	5.6	1.0	2.1	3.5
Alcohols																			
470	MS, RI, CoI	1.55	Ethanol	6.7	18.7	1.4	1.8	2.0	2.2	12.6	2.4	28.9	21.6	27.0	30.4	9.6	8.3	18.6	12.0
717	MS, RI	3.20	2-Pentanol	tr	nd	6.5	5.1	1.8	1.4	7.1	2.5	nd	nd	nd	nd	3.3	2.7	5.2	4.8
734	MS, RI, CoI	3.88	3-Methyl-1-butanol	11.0	13.4	3.3	3.0	2.2	1.6	11.7	5.2	13.3	18.1	26.3	24.6	11.6	10.2	7.2	10.3

Table 1 (continued)

RI ^a	Method of identification	RT (min)	Compound	Relative abundance (peak area, %) ^c															
				EC1 ^b		EC2		EC3		EC4		CC1		CC2		CC3		CC4	
				PDMS ^d	PA ^e	PDMS	PA	PDMS	PA	PDMS	PA	PDMS	PA	PDMS	PA	PDMS	PA	PDMS	PA
760	MS, RI, CoI	4.05	1-Pentanol	nd	nd	nd	nd	nd	nd	17.0	nd								
782	MS, RI	5.44	(±)-2,3-Butanediol	nd	nd	0.4	nd	nd	nd	3.0	9.5	nd							
807	MS, RI	6.18	<i>meso</i> -2,3-Butanediol	nd	nd	tr	2.3	nd	nd	3.8	4.2	tr	nd						
888	MS, RI	8.65	2-Heptanol	1.2	nd	2.9	2.1	1.0	0.4	nd	0.4	nd	0.4	nd	nd	nd	nd	0.6	nd
			Subtotal (%)	19.9	32.1	17.7	18.1	7.4	5.8	31.4	11.8	66.0	54.7	53.3	55.0	24.5	21.8	31.0	27.1
Ketones																			
503	MS, RI, CoI	1.64	Acetone	4.1	4.0	5.4	tr	3.8	1.5	16.2	2.9	9.5	3.8	2.7	3.6	11.7	4.4	9.9	4.8
686	MS, RI	2.99	2-Pentanone	4.6	2.4	2.9	1.1	3.4	0.9	6.9	1.2	nd	nd	nd	nd	21.5	6.7	5.1	2.2
706	MS, RI	3.45	3-Hydroxy-2-butanone	nd	nd	2.2	1.1	1.4	1.6	tr	1.1	5.9	8.2	22.1	9.8	4.0	4.3	10.1	5.6
889	MS, RI	8.34	2-Heptanone	15.0	7.7	12.4	4.1	7.8	1.7	5.0	1.1	tr	tr	tr	nd	10.4	3.9	9.1	3.3
992	MS, RI	10.62	2-Octanone	nd	nd	6.2	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd	nd
1,087	MS, RI	12.48	2-Nonanone	35.5	16.2	19.7	6.6	18.3	3.6	4.8	1.3	tr	nd	nd	nd	2.0	0.7	5.2	1.3
1,294	MS, RI	15.60	2-Undecanone	1.5	tr	0.6	0.2	2.1	0.3	nd	tr	nd							
			Subtotal (%)	61.5	30.3	50.3	13.5	37.2	9.6	32.9	7.6	15.4	12.0	24.8	13.4	49.6	20.0	39.4	17.2
Aldehydes																			
965	MS, RI, CoI	9.97	Benzaldehyde	tr	tr	0.2	0.3	tr	tr	nd	0.2	1.7	2.1	tr	tr	nd	tr	nd	nd
			Subtotal (%)	tr	tr	0.3	0.4	0.4	0.2	0.0	0.2	2.1	2.1	2.3	tr	0.0	tr	0.0	0.0

Table 1 (continued)

RI ^a	Method of identification	RT (min)	Compound	Relative abundance (peak area, %) ^c													
				EC1 ^b		EC2		EC3		EC4		CC1		CC2		CC3	
				PDMS ^d	PA ^e	PDMS	PA	PDMS	PA	PDMS	PA	PDMS	PA	PDMS	PA	PDMS	PA
Miscellaneous																	
485	MS, RI, CoI MS, RI, CoI	1.67	Diethyl ether	tr	nd	tr	3.7	tr	tr	tr	tr	tr	tr	tr	tr	tr	tr
531	MS, RI, CoI	1.76	Dichloromethane	2.8	5.0	2.1	2.0	2.2	1.3	nd	nd	0.8	1.3	2.7	4.3	4.4	3.0
600	MS, RI, CoI	2.08	Hexane	3.5	nd	0.5	nd	tr	nd	tr	nd	tr	nd	nd	nd	3.5	nd
	Subtotal (%)			6.3	5.0	5.4	5.7	2.2	1.3	tr	0.4	0.8	1.3	3.8	4.3	7.9	3.0
	Total (%)			93.6	103	93.3	60.9	56.1	99.6	91.6	87	84.7	82.4	74.4	84.3	87.0	73.1
																	90.5

nd not detected, tr trace compounds (less than 0.05% of the total peak area), MS identification based on mass spectra comparison, RI estimated retention time (index)/order of elution, CoI coinjection with an authentic standard

^a Literature values of retention indices (NIST Chemistry Web Book)

^b EC1 and CC1, day1; EC2 and CC2, day 5; EC3 and CC3, day 20; EC4 and CC4, day 30

^c Mean value of three independent measurements (both extraction and GC/MS)

^d Microextraction was conducted using polydimethylsiloxane (PDMS)

^e Microextraction was conducted using polyacryl (PA) fibers

^f Broad peaks, with a pronounced tailing effect

compounds were unique to just one sample/ripening period or reported for the first time in cheese samples. For example, methyl 2-hydroxy-4-methylpentanoate has not been previously reported in any other type of cheese prepared from cow's milk. However, this compound, (bio)synthetically related to leucine, was previously detected as a volatile metabolite in a number of different plant species (Knudsten et al. 1994). Perhaps, the origin of this ester in milk/cheese is directly connected to its presence in the fodder. The same may be true for several aliphatic and aromatic hydrocarbons detected (Viallon et al. 1999). Some of the compounds identified were characteristic for only one cheese type/ripening period and could also be products of lipid autooxidation or carotene degradation, formed during ripening (Barbieri et al. 1994; Johnson et al. 1969). Diethyl ether, dichloromethane, and chloroform were most probably contaminants introduced during the isolation procedure, during storage or from detergents used during cleaning of the plant and equipment.

3.2 Extraction of volatile compounds: influence of the stationary phase type on the resulting volatile profile

The two types of fiber stationary phases were selected due to their differentiating capacity for adsorption of volatiles (Mondello et al. 2005; Pinho et al. 2003). It has been shown that the non-polar PDMS fiber was satisfactory for compounds of low to medium polarity whereas the PA stationary phase is more suitable for the extraction of highly polar compounds of a broader volatility range (Mondello et al. 2005). The data presented in Table 1 could be used to justify and once again emphasize the necessity to use at least two fiber types, of different coating polarity, for HS-SPME extraction. The sensitivity of the PDMS phase for the extraction of octanoic acid (and other longer chain aliphatic compounds) was obviously higher than that of the PA fiber (Table 1). On the other hand, the PA fiber more effectively adsorbed ethanol and the rest of the detected acids. The results also show that both fibers had similar sensitivity for alcohols other than ethanol. When reaching conclusions on this "functional group affinity" of a particular stationary phase, one should bear in mind the changes (i.e., the decrease) of polarity of members of a (homologous) series of compounds with an increase in the number of carbon atoms in their alkyl chains. Once more, it is important to note that the use of just one type of stationary phase for HS-SPME of volatile compounds would have easily given false (over- or understating the relative contribution of certain compound(s)) results concerning the most dominant volatile compounds present in these cheese samples.

MVA analyses (see Electronic supplementary material Fig. S2) revealed that, in some cases, no significant differences were detected between the PDMS and PA cheese volatile profiles. Nevertheless, a substantial number of PA volatile profiles were recognized as statistically different from their corresponding non-polar fiber counterparts. The differences between the volatile profiles obtained through the use of different SPME phases (non-polar and polar) become even more pronounced if the sums of the volatile compound classes are used as variables. It is interesting to note that all four cheese samples made from cow's milk and the 1-day-old cheese sample made from ewe's milk had very similar volatile profiles (both fibers), whereas the volatile profiles of the three older ewe's cheese samples were very different depending on the fiber used. It seems that some more pronounced chemical changes occur

during the ripening of ewe's milk cheese in comparison to those associated with cow's cheese (this could be a consequence of the transformation of one class of compounds to another of different polarity).

3.3 Different types of variables for the comparison of volatile profiles of different cheese types and the tracking of ripening-induced changes

The results of MVA analyses (Figs. 1 and 2) clearly showed that, in general, there are significant differences in the aroma profiles of Pirotski kačkavalj made from ewe's and cow's milk. Generally speaking, all MVA analyses clearly distinguished (with few exceptions that will be discussed later) the two types of cheese, and the grouping of CC and EC samples is easily discernible from all the PCA biplots and AHC dendrograms. If considered alone, due to the “functional group affinity” of each of the SPME fiber stationary phases, the volatile profiles obtained by using a PDMS or PA fiber provide data which either over- or underestimated the relative amount of an individual volatile compound. It was our intention to check if this could be overcome by using some transformed/combined variables for MVA, which would include data

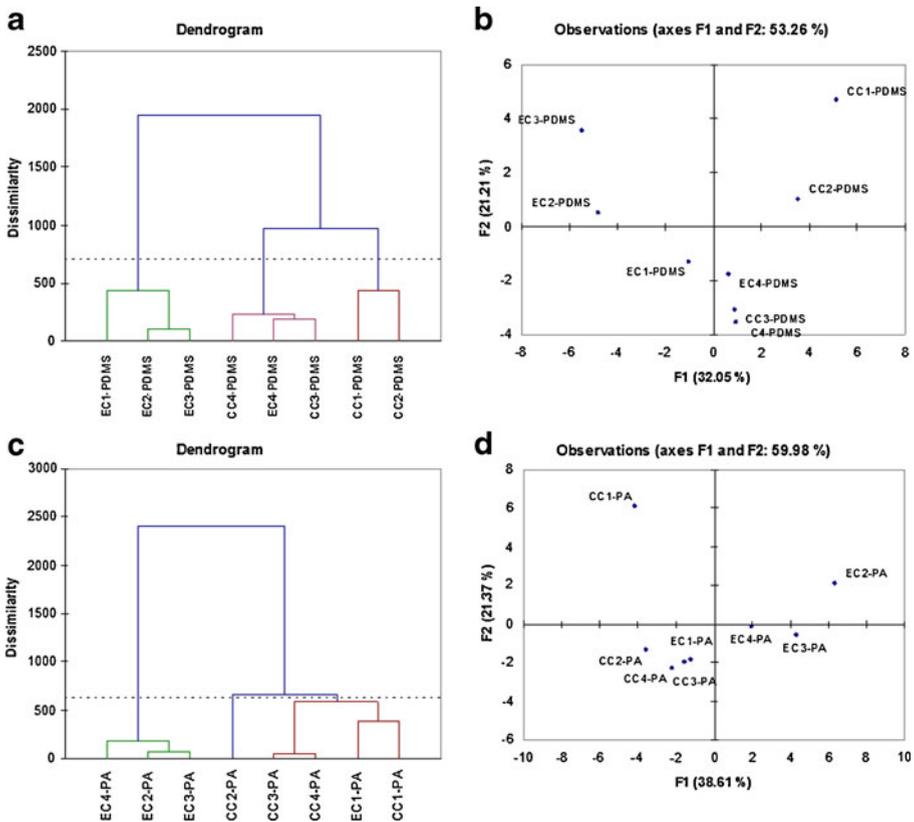


Fig. 1 Dendrograms (AHC analysis) and biplots (PCA analysis) representing chemical composition dissimilarity relationships of eight cheese samples using the original variables: **a** PDMS-AHC, **b** PDMS-PCA, **c** PA-AHC, **d** PA-PCA

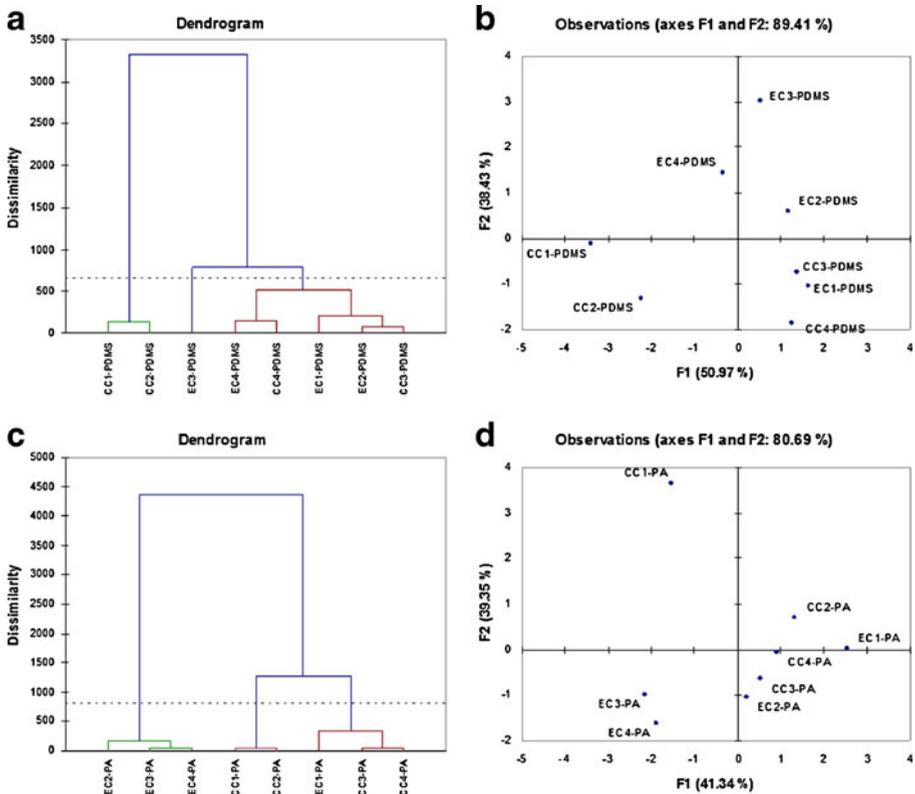


Fig. 2 Dendrograms (AHC analysis) and biplots (PCA analysis) representing chemical composition dissimilarity relationships of eight cheese samples using the transformed variables: **a** PDMS-AHC, **b** PDMS-PCA, **c** PA-AHC, **d** PA-PCA

obtained from experiments with different fiber stationary phases. In order to do that, additional variables were devised by combining (summing with the weight value equal to 0.5) relative amounts of individual compound/classes of compounds found in a cheese sample by each of the different fibers (PA and PDMS). The MVA results performed with these new variables are given in Fig. 3. Significant differences in the flavor profiles of the cheese samples prepared from different types of milk (ewe's and cow's) were still clearly highlighted.

As can be seen from Table 1 and Figs. 1, 2, and 3, significant differences in the volatile profiles of the two cheese types occurred during the ripening period. The relative amounts of some of the compounds/classes of compounds tended to increase during the cheese ripening (e.g., majority of acids), as seen in Table 1, while others decreased (e.g., ketones). Generally speaking, with ripening, a larger number of volatile compounds were identified in cheeses made using EC than in CC. It should be noted that the ripening-induced changes in the cheese volatile profiles are, most probably, the result of at least two different processes: (a) chemical/microbiological transformation of native raw milk constituents and (b) evaporation loss of the most volatile compounds. Thus, the cheese volatile profile could be considered to be the result of a complex and dynamic equilibrium.

The results of MVA analyses obtained for the EC samples, using different sets of variables, were much less coherent. It seems, according to all performed PCA analyses (differences in the corresponding score factors), that EC undergoes a more pronounced and, more or less, constant change during ripening. According to the AHC analyses, samples EC2 and EC3 were recognized as statistically mutually very similar and were always placed within the same cluster of the corresponding dendrograms. In contrast, the clustering of EC1 and EC4 was not the same if EC-PDMS instead of EC-PA profiles were used as the input datasets for AHC. This again pinpoints the issue relating to differences in the affinity of the HS-SPME stationary phases to compounds of differing polarity. Interesting results were obtained when combined variables were used (averaging of the cheese profiles was achieved by combining the profiles obtained from both of the used types of fibers, with the weight factor of 0.5). For example, AHC analysis (average of the relative amounts of individual constituents as variables) resulted in dendrograms that are essentially the same as those obtained by the use of original variables derived from PDMS profiling. However, the results of the AHC based on combined-transformed variables (averaging of the relative amounts of classes of constituents) were analogous to those of the AHC with original PA variables. Thus, it could be speculated that, perhaps, PA gives a more realistic cheese profile when classes of constituents are in question, while PDMS is better at recognizing the specific dominant volatile/flavor compounds. MVA experiments with combined variables obtained with different weighting factors (0.8/0.2, 0.6/0.4, 0.4/0.6, and 0.2/0.8) were also performed and gave expected results (in the sense of resemblance to the MVA results obtained by using the corresponding (PA or PDMS) variables).

4 Conclusion

In this paper, the applicability of different types of variables in MVA profiling of cheese volatile compounds was explored. Our results have shown that, in addition to original variables (relative amounts of detected cheese volatile compounds), the distribution of volatile compounds into classes also gives valuable information concerning cheese type and ripening stage. Additionally, the extraction with a PA fiber gives a more realistic cheese profile when the relative ratio of the classes of constituents is in question, while PDMS is better in recognizing the contribution/concentration of the dominant volatile/flavor compounds. Also, combined variables that include information obtained from the extraction experiments using different types of stationary phases (PA and PDMS fibers) could be used for a better and more complete description/profiling of the aroma of cheese.

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