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Bioactivation of zearalenone by porcine hepatic biotransformation

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Abstract – Zearalenone (ZEA) is a resorcylic acid lactone derivative produced by various *Fusarium* species that are widely found in food and feeds. The structure of zearalenone is flexible enough to allow a conformation able to bind to mammalian oestrogen receptors, where it acts as an agonist. Using oestrogen-dependent Human Breast Cancer (MCF-7) cells, the oestrogenic activity of zearalenone and its derivatives were compared using 17 β -oestradiol as a positive control. The results obtained demonstrate that the oestrogenic potency of ZEA derivatives could be ranked in the following order: α -zearalenol > α -zearalanol > zearalenone > β -zearalenol. Since pigs have been reported to be among the most sensitive animal species, biotransformation studies with pig liver subcellular fractions were conducted. These studies indicated that α -zearalenol is the main hepatic metabolite of zearalenone in pigs, and it is assumed that 3 α - and 3 β -hydroxysteroid dehydrogenases are involved in the hepatic biotransformation, since the formation of α -zearalenol and β -zearalenol could be inhibited by prototypic substrates for either enzyme. The bioactivation of ZEA into the more active α -zearalenol seems to provide a possible explanation for the observed high sensitivity of pigs towards feedingstuffs contaminated with the mycotoxin.

zearalenone / MCF-7 cells / oestrogenic effects / biotransformation / pigs

1. INTRODUCTION

Human and animal diets may contain several non-steroidal weakly oestrogenic compounds that are either produced by plants (phyto-oestrogens) or by fungi (myco-oestrogens). The most important myco-oestrogen is zearalenone (ZEA), [6-(10-hydroxy-6-oxo-trans-1-undecenyl)- β -resorcylic acid], a non-steroidal compound mainly produced by the fungal species *Fusarium graminearum* and *Fusarium culmorum*. It is one of the most frequently

occurring contaminants of corn and corn-derived plants and it has also been detected in cereals such as barley, rice, oats, rye, sorghum, soy beans and wheat [8, 28].

Following ingestion with feed, zearalenone is rapidly absorbed from the gastro-intestinal tract, and oral availability is estimated to reach 80–85% in monogastric species. In the liver, zearalenone is reduced to the isomeric α - and β -zearalenols (α -ZOL and β -ZOL), a reaction that competes with glucuronidation of the parent molecule. Previous data indicated that the ratio

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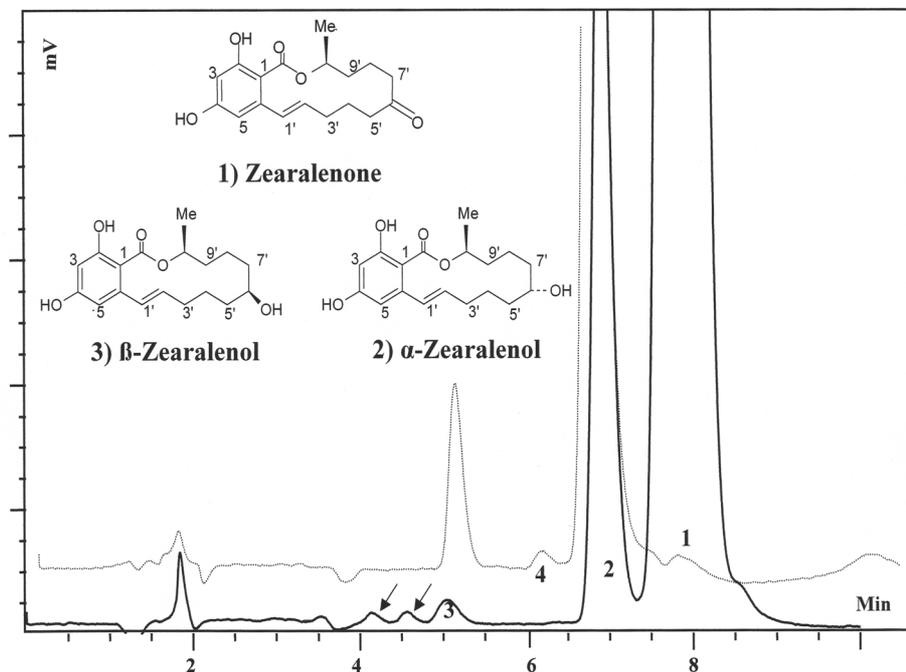


Figure 1. Chemical structures of zearalenone and its metabolites, and an HPLC profile with ZEA (1), α -ZOL (2), β -ZOL (3), and (4) α -ZAL. The grey chromatogram depicts the mixture of the standards, the black chromatogram is a representative chromatogram of samples (pig liver microsomes incubated with zearalenone (for details see Materials and methods section)).

between α -ZOL and β -ZOL varies considerably among species, as for example pigs seem to form predominantly α -ZOL, whereas in cattle predominantly β -ZOL formation is observed. The chemical structures of ZEA and its major metabolites are shown in Figure 1. In some animal species, including cattle, goats, sheep and horses, a further reduction of α -ZOL into the corresponding α -zearalanol (zeranol) and β -zearalanol (taleranol) takes place in vivo. Whether or not hepatic metabolism contributes to this second methylation step remains to be elucidated [27]. The possible biotransformation of zearalenone into zeranol is of particular interest, since in non-European countries zeranol might be used as a growth promoter in cattle, whereas it is strictly inhibited in Europe (Council Directive 96/22/EC).

ZEA as well as many of its metabolites act as oestrogen receptor (ER) agonists. They can bind competitively to oestrogen receptors located in the uterus, mammary gland, liver and hypothalamus, as demonstrated in various species [21, 24], and the consumption of contaminated feed has been associated with clinical signs of hyperoestrogenism in pigs and other animal species [8, 13, 25]. Pigs appear to be especially sensitive to the oestrogenic effects of ZEA and adverse affects in sexually mature pigs include decreased fertility, ovarian atrophy, prolonged oestrus, oedematous swelling and reddening of the vulvae, prolapsed uterus and rectum. In piglets, stillbirth, reddening and swelling of the vulva, nipples and the prepuce as well as necrosis of the tail have been described [7, 8, 41, 42].

In the present study, the oestrogenic activity of ZEA and its metabolites is compared to that of 17- β -oestradiol in a bioassay. In this assay, proliferation of oestrogen-dependent human breast cancer cells (MCF-7 cells) is used as a marker to assess oestrogenic potency. Since pigs are known to be very susceptible to the adverse effects of ZEA, the biotransformation of this mycotoxin was investigated *in vitro* using subcellular fractions of the pig liver to assess the ratio of the individual metabolites.

2. MATERIALS AND METHODS

The test compounds 17- β -oestradiol (E_2), zearalenone (ZEA), α -zearalenol (α -ZOL), β -zearalenol (β -ZOL), zeranol (α -ZAL), activated charcoal, and MTT [(3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide)] were purchased from Sigma Chemical Co. (St. Louis, MO, USA). DMEM medium was obtained from GIBCO (Aickland, United Kingdom), and T77 dextran was purchased from Amersham Pharmacia Biotech (Uppsala, Sweden). MCF-7 cells were obtained from ECACC (European Collection of Cell Cultures). All test compounds were dissolved in absolute ethanol in order to prepare 100 μ M and 10 μ M stock solutions, respectively. All solutions were stored at 4 °C prior to use.

2.1. Bioassay for oestrogenic activity of ZEA and its metabolites

2.1.1. Cell cultures

Human breast cancer cells (MCF-7) were cultured in DMEM without phenol red, supplemented with 10% bovine calf serum (BCS), 1% penicillin (100 units/mL), streptomycin (100 μ g/mL), 1% L-glutamine, 1% sodium pyruvate (100 mM), and 0.1% bovine insulin (1 mM), subsequently denoted as a complete DMEM medium. The cells were maintained as monolayer cultures in

standard culture flasks, kept at 37 °C in a humidified atmosphere of 5% CO₂ in air. Every 4–5 days, the subcultures were obtained using trypsin.

2.1.2. Preparation of charcoal-dextran-treated bovine calf serum (CD-BCS)

Charcoal-Dextran (CD) was used to remove natural oestrogens from the BCS according to Ju et al. [20]. Briefly, activated charcoal (5 g, Norit-Sigma) was mixed with 0.5 g T77 dextran (Pharmacia, Uppsala, Sweden), and the mixture was suspended in 100 mL demineralised water. After 3 times washing by centrifugation (2 500 rpm, 20 °C, and 3 min), the supernatant was removed and 5 mL BCS was added to 5 mL of the CD suspension. Following mixing on a roller-plate for 1 h, the suspension was centrifuged (2500 rpm for 20 min) and the supernatant was filter-sterilised (Nalgene SFCA membrane, 0.22 μ m). Aliquots of the CD-BCS obtained were stored at –20 °C in glass tubes until use.

2.1.3. Oestrogen-dependent cell proliferation

MCF-7 cells were harvested by trypsinisation of stock cultures and were re-cultured in 75-mm flask and incubated in 37 °C and 5% CO₂. After 72 h the medium was removed and replaced by complete DMEM without phenol red that contained 10% DC-BCS. In the first set of the experiments, MCF-7 cells were then seeded in 96-well cell culture plates with different densities (2 500, 5 000, 10 000, and 20 000 cells/well) and incubated with 17- β oestradiol for three different time periods (96, 120, and 144 h) in order to assess oestrogen dependent cell proliferation as compared to solvent treated controls by means of the MTT assay. Each experiment was conducted in triplicate, and the data represent means \pm SD from three independent experiments.

2.1.4. Assessment of mycoestrogen potency by the MCF-7 cell proliferation assay

As described above, MCF-7 cells were seeded in 96 well-culture plates (10 000 cells/well in 200 μ L medium). After 24 h, the medium was removed and replaced with supplemented DMEM containing 10% CD-FCS and different concentrations of the test chemicals including E_2 (10^{-3} – 10^4 pM), ZEA and its derivatives (10^{-3} – 10^7 pM); α -ZAL was included as second positive reference due to its known oestrogenic activity [22, 23, 27]. After 6 days of incubation at 37 °C and 5% CO_2 , the MTT-assay was conducted and data are compared with the corresponding solvent-treated controls. The data represent the results of three independent experiments each conducted in triplicate. EC_{50} values were calculated using non-linear regression analysis (Win Non-line 4.1., Pharsight, North Carolina, USA).

2.1.5. MTT Test

Cell proliferation was quantified by the colorimetric MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide] assay. This assay measures the reduction of dimethylthiazol diphenyl tetrazolium bromide (MTT; stock solution 3 mg/mL dissolved in PBS) to formazan by the mitochondrial enzyme succinate dehydrogenase. This reduction capacity reflects the number of viable cells, hence the MTT assay is used for both the assessment of cytotoxicity and the demonstration of cell proliferation. Following the described incubation period, the medium was discarded and 0.2 mL MTT solution was added to the cells. After 3 h incubation at 37 °C the MTT solution was discarded and the intracellular purple insoluble formazan was solubilised by adding 100 μ L/well of lysis buffer (0.5% sodium dodecyl sulphate, 36 mM HCl, and isopropanol acid). Following careful mixing, the optical density (OD) was measured at 594 nm using a micro plate reader (BIORAD Model 3550). Cell proliferation rate

was expressed as the percentage of non-treated controls as follows: A_{595} treated cells/ A_{595} of control) \times 100.

2.2. Biotransformation of ZEA by subcellular fractions derived from porcine livers

2.2.1. Preparation of liver subcellular fractions

Immediately after slaughter, liver specimens from four male pigs (Great Yorkshire \times Dutch Landrace), aged approximately 12 weeks, and weighing between 30 and 35 kg, were collected, cut into small pieces, and rinsed three times with ice-cold saline to remove excess blood. The subcellular fractions were prepared according to Rutten et al. [34]. Briefly, two volumes of KCL (1.15%) and EDTA (0.1mM) solution were added to a tissue sample and the mixture was homogenised in a Potter-Elvehjem apparatus with a Teflon pestle. The homogenate was then centrifuged at 9 000 g at 4 °C for 30 min and an aliquot of the supernatant was collected as the post-mitochondrial (S9) fraction. The remaining sample was centrifuged at 100 000 g at 4 °C for 90 min in order to obtain the microsomal fraction. After centrifugation, the supernatant was discarded and the pellet was carefully dissolved in the same volume of phosphate buffer (0.05 M; pH 7.4) with glycerol (20%), and then homogenised with an ultra-turrax. The individual fractions of the four pigs were pooled and stored at -70 °C until use. Protein concentrations of the microsomal and the S9-fractions were determined according to Lowry et al. [26].

2.2.2. Incubations with subcellular fractions

In order to measure the biotransformation products, different concentrations of ZEA were added to the reaction mixtures of 250 μ L, containing either pooled liver microsomes (0.375 mg protein per mL) or

the post-mitochondrial fraction of the liver (1.25 mg protein/mL) and supplemented with NADPH (0.5 mM) in a final volume of 250 μ L phosphate buffer (50 mM, pH 7.4). The samples were incubated in a shaking water bath at 37 °C for 30 min. The reaction was stopped by transferring the samples to an ice-cold environment followed by extraction with chloroform (1.25 mL). One millilitre of the organic phase was collected and evaporated to dryness under a stream of N₂. The residue was redissolved in the mobile phase. Analysis of the enzyme kinetics revealed under these conditions an enzyme velocity (V_{\max}) of 953 ± 75 pmol.mg/min and 173 ± 30 pmol.mg/min for the α -ZOL and β -ZOL formation in liver microsomes, and 401 ± 38 pmol/mg/min and 80 ± 7.7 pmol.mg/min for the formation of α -ZOL and β -ZOL in the post-mitochondrial fraction, respectively.

2.2.3. Inhibition studies

In order to assess the involvement of 3α - and 3β -hydroxysteroid dehydrogenases (3α and 3β -HSD) in the biotransformation of ZEA, 5α -androstenedione (5α -AND) as the endogenous substrate of 3α -HSD, and pregnenolone as the endogenous substrate for 3β -HSD were added to the incubation mixture at increasing concentrations (50, 100, 250, 500 and 1 000 μ M), whereas the concentration of ZEA (1 000 μ M) remained constant. These incubations were conducted with porcine liver microsomes in the presence of NADPH as the co-factor and at pH 7.4 for 30 min. Data were compared to control incubation conducted with ZEA only.

2.2.4. HPLC determination and confirmation of ZEA metabolites

ZEA and its metabolites were determined by high-performance liquid chromatography (HPLC) according to Trenholm et al. [39]. The chromatographic system consisted of an auto sampler (Gynkotek Autosampler Model GINA 50, Separations

Analytical Instruments BV, Hendrik Ido Ambacht, The Netherlands) and dual pumps (Gynkotek High Precision Pump Model 300, Separations Analytical Instruments BV, Hendrik Ido Ambacht, The Netherlands). Fifty microlitres of the extracted sample was injected onto a LUNA 5 μ C18 (150 \times 4.60 mm, Phenomenex) column. The mobile phase consisted of a mixture of methanol-water (70:30, v/v) eluted at a flow-rate of 0.8 mL/min. Detection occurred by means of a fluorescence detector (Jasco Fluorescence Detector Model FP-920, Separations Analytical Instruments BV, Hendrik Ido Ambacht, The Netherlands), set at an excitation and emission wavelength of 236 and 418 nm, respectively. ZEA and its metabolites were quantified by measuring peak areas, and comparing them to the relevant calibration curves.

2.3. Statistical analysis

All results are presented as means \pm SD. Differences in the cell proliferation between the control and treatment groups and differences between the control and treated groups in ZEA biotransformation in the presence of different competitors were evaluated with one-way ANOVA. Differences between the amounts of produced metabolites by either the microsomal or the post-mitochondrial fractions were analysed by a student *t*-test. $P < 0.05$ was considered as significant.

3. RESULTS

3.1. Bioassays with MCF-7 cells

Standardisation of the bioassay with MCF-7 cells has to address a number of variables including the cell number, incubation time and non-specific responses exerted by oestrogenic compounds present in commercial cell culture medium supplements. Initial studies indicated that the optimal density of MCF-7 cells in a 96-well tissue culture plate is 1×10^4 cells/well (Fig. 2),

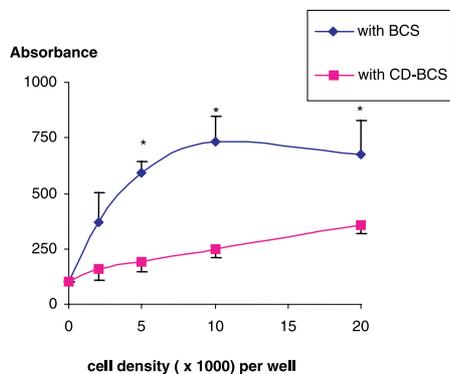


Figure 2. Effect of serum composition on MCF-7 cell proliferation: The cells were left to grow for 6 days and proliferation was measured by an MTT test. Optical density of control incubations was set to 100, and data are expressed as the percentage of this control. Individual data points represent the mean \pm SD of three independent experiments conducted in triplicate.

since at this concentration a measurable but not-significant cell proliferation could be measured in the control samples.

Also in this study, the difference between native BCS and charcoal-stripped CD-BCS could be seen, since there was a significant higher proliferation rate of cells incubated with BCS as compared to the cells incubated with CD-BCS. This difference is considered to be attributable to residual amounts of natural oestrogens present in BCS. In the presence of CD-BCS serum cell proliferation was minimal ($P < 0.001$; Fig. 2). These findings implied that all assays have to be conducted with stripped, oestrogen-depleted serum.

A second set of experiments was conducted to assess the optimal incubation time. Data shown in Figure 3 indicate that the late exponential phase of cell proliferation in response to the exposure to graded concentrations of the positive control 17β -oestradiol is reached after 144 h. This time period was subsequently used in all further experiments. Moreover, it could be demonstrated that 17β -oestradiol (see also Fig. 4A) induced a maximal proliferation rate at a

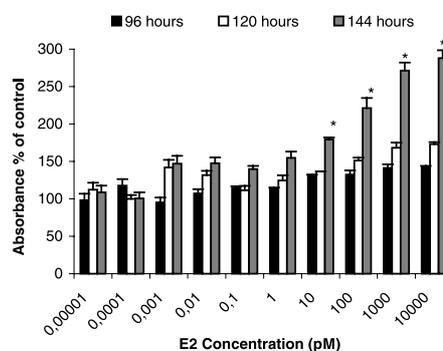


Figure 3. Dose-response curve of 17β -oestradiol at various treatment times: 10^4 cells/well were treated with various concentrations of 17β -oestradiol and incubated for different time intervals. Cell proliferation was assessed by the MTT test. Data points represent the mean \pm SD of three independent experiments conducted in triplicate.

concentration of 10^3 pM. The calculated EC_{50} value corresponded to 2.8 pM.

3.2. Effect of ZEA and its derivatives on cell proliferation

Proliferation of the MCF-7 cells was analysed in the presence of ZEA and its metabolites. The data in Figure 4B show that ZEA and its reduced metabolites significantly enhance the proliferation of MCF-7 cells. The following EC_{50} values were calculated: 49.9, 105.6, 1638.7, and 20007.9 pM for α -ZOL, α -ZAL, ZEA, and β -ZOL, respectively, implying that the most potent zearalenone derivative, α -ZOL is about 20 times less potent than 17β -oestradiol.

3.3. ZEA metabolism by liver subcellular fractions

Following the incubation of either porcine microsomes or post-mitochondrial fractions with different concentrations of ZEA, the chloroform-extracted samples were subjected to HPLC analysis and ZEA

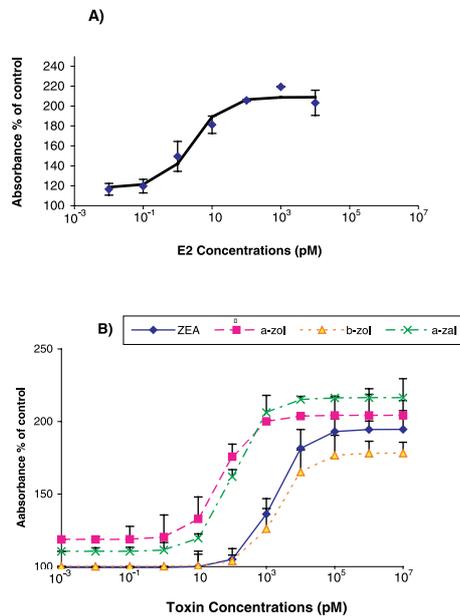


Figure 4. Dose-response curves of (A) 17 β -estradiol, and (B) ZEA, α -ZOL, β -ZOL, and α -ZAL in the MCF-7 bioassay. Data points represent the mean \pm SD of three independent experiments conducted in triplicate.

(peak 1), as well as α -ZOL (peak 2) and β -ZOL (peak 3) were quantified according to the respective standard curves. Two other unknown minor metabolites were detected as presented in Figure 1, but could not be identified.

As shown in Tables I and II, the major metabolite of ZEA formed by pig liver microsomes and the liver post-mitochondrial fraction was α -ZOL. Only at low concentrations of ZEA (< 100 μ M) did the amount of β -ZOL exceed that of α -ZOL. A comparison of the biotransformation capacity in the microsomal fraction with that of the post-mitochondrial fraction indicated a higher biotransformation capacity in the microsomal fraction. The ratio of α -ZOL/ β -ZOL, however, was higher in the incuba-

tion with the post-mitochondrial fraction as compared to incubations with the microsomal fraction. The absolute amount of metabolites formed by either the microsomal or the post-mitochondrial fraction differed significantly ($P < 0.001$).

3.4. Inhibition of the biotransformation of Zearalenone by endogenous substrates for 3 α - and 3 β -HSD

In order to demonstrate that 3 α - and 3 β -HSD are indeed involved in the hepatic biotransformation of ZEA and subsequently in the formation of α - and β -ZOL, porcine liver microsomes were incubated with ZEA (1000 μ M) in the presence of endogenous substrates for either 3 α -HSD or 3 β -HSD. The results summarised in Figure 5A show that the NADPH-dependent biotransformation of ZEA into α -ZOL was significantly inhibited by 5 α -AND and PGN. This inhibition was concentration-dependent. In contrast, biotransformation of ZEA into β -ZOL by porcine liver microsomes was only inhibited in the presence of PGN at concentrations higher than 250 μ M. Incubations with increasing concentrations of 5 α -AND did not significantly inhibit β -ZOL formation (Fig. 5B).

4. DISCUSSION

One of the most reliable bioassays to assess the oestrogenic activity of natural, semi-synthetic and synthetic chemical compounds measures the cell proliferation of the human breast cancer cells MCF-7 [3, 9]. MCF-7 cells are known to express both types of oestrogen receptors, and cell proliferation depends on the presence of agonists to these receptors. This assay, also denoted as the E-screen, was applied with minor modifications in the current study, in order to compare the effects of ZEA and its hepatic metabolites with those induced by the synthetic growth promoter α -zearalanol or the natural agonist 17 β -oestradiol.

Table I. Metabolism of ZEA by pig liver microsomes and ratio of α -ZOL/ β -ZOL. Individual values represent the means \pm SD of three independent experiments.

ZEA Concentration (μ M)	α -zol (pmol/min.mg protein)	β -zol (pmol/min.mg protein)	Ratio α -zol: β -zol
10	11.88 \pm 1.5	64.74 \pm 38.79	0.18
25	37.02 \pm 5.7	79.56 \pm 31.81	0.46
50	71.33 \pm 1.5	86.36 \pm 13.19	0.82
100	170.97 \pm 7.4	84.06 \pm 7.14	2.03
250	370.60 \pm 11.1	189.31 \pm 41.13	1.95
500	586.69 \pm 111.4	287.63 \pm 47.96	2.03
750	510.41 \pm 143.9	251.43 \pm 53.55	2.03
1000	598.53 \pm 204.2	362.92 \pm 35.24	1.64
1250	643.64 \pm 196.6	426.56 \pm 15.98	1.50
1500	776.21 \pm 230.2	521.06 \pm 17.38	1.48
1750	977.26 \pm 349.9	658.99 \pm 162.19	1.48
2000	891.15 \pm 1.6	697.86 \pm 111.58	1.27

Table II. Metabolism of ZEA by pig liver post-mitochondrial fraction and ratio of α -ZOL/ β -ZOL. Individual values represent the means \pm SD of three independent experiments.

ZEA Concentration (μ M)	α -zol (pmol/min.mg protein)	β -zol (pmol/min.mg protein)	Ratio α -zol: β -zol
10	4.8 \pm 2.8	8.9 \pm 2.5	0.53
25	4.1 \pm 0.4	23.4 \pm 3.7	0.60
50	18.5 \pm 14.5	22.9 \pm 4.2	0.80
100	55.1 \pm 26.9	38.2 \pm 7.0	1.44
250	128.5 \pm 60.5	56.7 \pm 6.4	2.26
500	239.9 \pm 49.1	87.3 \pm 17.5	2.74
750	337.0 \pm 43.1	108.0 \pm 4.0	3.12
1000	292.1 \pm 43.4	110.4 \pm 20.8	2.64
1250	435.7 \pm 8.7	154.2 \pm 11.1	2.82
1500	423.1 \pm 88.1	152.2 \pm 21.4	2.77
1750	477.7 \pm 6.4	197.7 \pm 42.8	2.41
2000	421.3 \pm 48.6	170.9 \pm 51.1	2.46

A specific prerequisite for any assay designed to measure oestrogenic activity, is the avoidance of all oestrogenic compounds in the cell culture medium. Since natural BCS contains residual amounts of physiological oestrogens, demonstrated by the significant differences on cell proliferation between natural BCS and CD-BCS (see Fig. 2), the serum has to be stripped carefully prior to use by washing with activated charcoal and dextran. Moreover, in any cell proliferation

assay, the optimal density of cells at seeding has to be estimated. To this end, a series of initial experiments was conducted with 17β -oestradiol showing that in 96 well plates MCF-7 cells reach the stationary phase of growth after 144 h, when seeded at a density of 10^4 cells per well. Although our results are in accordance with previously published data [1, 3, 11], the exact assay conditions have to be established prior to the testing of new chemicals, since

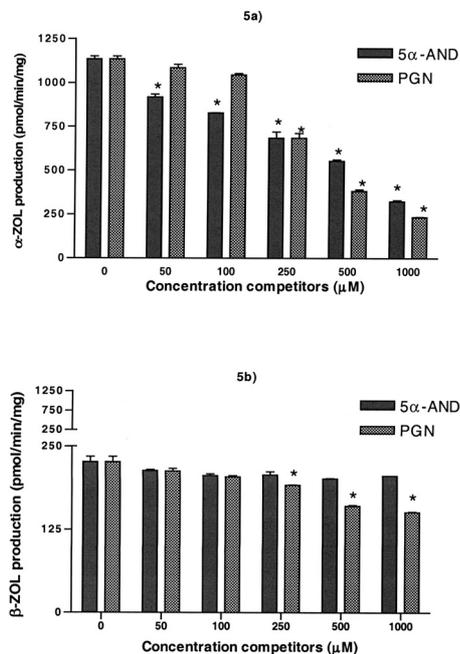


Figure 5. ZEA biotransformation by pig liver microsomes in the presence of endogenous substrates for 3 α - and 3 β -HSD: (a) α -ZOL production and (b) β -ZOL formation in the presence of 5 α -androstane-3-one (5 α -AND), and pregnenolone (PGN).

the number of cell passages prior to the actual assay might influence cell proliferation rates as seen in SaOS cells [14], in MC3T3-E1 pre-neoblastic cells [18], in C-6 glioma cells [17], in LNCap prostatic adenocarcinoma cells [12], in Caco-2 cells [6], and in rat mesangial cells [16].

Dose-dependent cell proliferation induced by oestrogenic compounds can be used to compare their relative oestrogenic activity. Based on the dose-response curves and the calculated EC₅₀ values, the oestrogenic potency of ZEA and its derivatives can be ranked as follows: α -ZOL > α -ZAL > ZEA > β -ZOL. The higher potency of α -ZOL and α -ZAL as compared to parent ZEA and β -ZOL, reflect their higher affinity to the oestrogen receptors in MCF-7 cells. These findings were in accordance

with previous results, demonstrating that in a conventional eutopic assay, which assesses the increase of the organ weight of the juvenile uterus in mice or rats, the same ranking of the oestrogenic effects of ZEA and its metabolites was observed [35, 40].

The major advantages of an in vitro assay are its higher specificity and accuracy. In the eutopic assay, the uterus weight might be increased due to different factors, including among others inflammatory processes. Moreover, in vitro assays are independent of laboratory animals and allow the analysis of a wide range of concentrations of diverse contaminants. Hence, the MCF-7 cell proliferation assay has been widely used for the assessment of diverse oestrogenic compounds and is considered to be very specific [3, 10, 36, 37].

The agonistic property of ZEA to oestrogen receptors, inducing cell proliferation in MCF-7 cells, has been shown previously. Dees et al. [9] demonstrated that ZEA stimulates MCF-7 cells to enter the cell cycle by increasing the cyclin D1 protein levels, and activating cyclin-dependent kinase (Cdk)-2 resulting in the hyperphosphorylation of the retinoblastoma gene product [9]. These actions, in turn, lead to an increased rate of G1 to S phase transitions and mitosis. By inducing cell-cycle progression in MCF-7 cells, ZEA has anti-apoptotic properties [1].

In the intact animal, there are two main biotransformation pathways for ZEA. The reduction of ZEA by hydroxysteroid dehydrogenases (HSD) yields α - and β -zearalenol. In addition, both ZEA and the reduced metabolites are known to be conjugated to glucuronic acid in the presence of UDPG-glucuronyl transferase, facilitating biliary and renal excretion [4, 5].

The reduction of ZEA resembles that of steroid metabolism catalysed by steroid dehydrogenases [29]. There are two different 3-HSD known so far to catalyse the formation of either 3 α -hydroxysteroids, or 3 β -hydroxysteroids. The 3 α -HSD isoform is expressed predominantly in the brain, prostate, liver, skin, lung, kidney, and gonads.

Physiologically, this enzyme acts as an oxidoreductase (reductase-dehydrogenase) catalysing the reduction of the 3-ketogroup on steroids into a 3 α -hydroxygroup and vice versa. This enzyme is also able to metabolise many non-oestrogenic compounds thereby acting mainly as a dehydrogenase [30, 31, 33]. 3 β -hydroxysteroid dehydrogenase occurs in humans in two distinct forms, 3 β -HSD-1 and 3 β -HSD-2. 3 β -HSD-1 is found in the placenta, skin, mammary gland, prostate, and endometrium, as well as in breast cancer cells, whereas 3 β -HSD-2 prevails in the gonads and adrenals [15]. Human 3 β -HSD-isomerases belong to the SDR (short-chain dehydrogenase/reductase) family, catalysing a dehydrogenase activity involving NADH as the co-factor [38]. PCR analysis demonstrated a moderate amount of 3 β -HSD-1 RNA in human foetal liver [32], but reports on these enzymes in pig tissues are lacking.

Incubations with porcine liver microsomal or post-mitochondrial fractions indicate that α -ZOL is the main metabolite of ZEA in pigs. However, at low concentrations of ZEA (< 100 μ M), the ratio shifted towards higher amounts of β -ZOL, indicating the involvement of both isoenzymes in the hepatic biotransformation of ZEA. This observation was further substantiated by the observation that higher amounts of reduced metabolites of ZEA were produced by the microsomal fraction as compared to the post-mitochondrial fraction (at equal amounts of cellular protein). When calculating the α -ZOL/ β -ZOL ratio in either fraction (see Tabs. I and II), it became obvious that at almost all tested concentrations of ZEA, the amount of both metabolites produced by microsomes was higher than the amount produced by the post-mitochondrial fraction, and that the amount of α -ZOL exceeded that of β -ZOL. These findings suggest that the enzymatic activity of 3 α -HSD and probably 3 β -HSD varies in different subcellular fractions, i.e. occurring as microsomal and cytosolic forms [2], depending on the organ or part of the organ [27], and the animal species investigated. Indeed,

it has been shown for example that cytosolic 3 α -HSD activity is greater than the microsomal one in the liver and skin of rats, while in humans and dogs microsomal 3 α -HSD activity is about one order of magnitude lower than the cytosolic activity in the liver and prostate [19, 33].

Further evidence for the involvement of 3 α -HSD in the conversion of ZEA in α -ZOL is provided by competition studies with porcine hepatic microsomes. In the presence of 5 α -androstanedione, a keto-steroid and prototypical substrate for 3 α -HSD, the α -ZOL production was significantly inhibited. Also in the presence of pregnenolone, the apparent formation of α -ZOL was inhibited, corresponding to the finding that pregnenolone can be a substrate for human Type 3 3 α -HSD [32]. Hence, it can be concluded that in the pig liver the reduction of ZEA towards α -ZOL is catalysed predominantly by microsomal 3 α -reductases. In contrast to the α -ZOL production, the formation of β -ZOL by pig liver microsomes was only inhibited in the presence of pregnenolone, a typical endogenous substrate for 3 β -HSD. The concentration-dependent inhibition of α - and β -ZOL production confirms that this inhibition is competitive. Further investigations are necessary to quantify enzyme expression in pig livers and to compare enzyme velocity in pig subcellular fractions with those in other animal species, to identify enzyme distributions in pig livers and to compare enzyme velocity in pig subcellular fractions with those of other animal species, including man.

In conclusion, the present experiments with the MCF-7 cell bioassay demonstrated the higher oestrogenic activity of α -ZOL when compared to that displayed by the parent toxin or the β -hydroxylated metabolite. Moreover, the *in vitro* experiments with pig liver subcellular fractions showed the formation of α -ZOL to be prevalent in this animal species. Taken together, the bioconversion to α -ZOL has to be considered as a bio-activation step, which may together with the known enterohepatic recirculation

of ZEA provide the mechanistic explanation for the high sensitivity of pigs towards feed materials, contaminated with the mycotoxin zearalenone.

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