



**HAL**  
open science

## Pharmacokinetic and residue studies of quinolone compounds and olaquinox in poultry

A Anadón, Mr Martinez-Larrañaga, Mj Diaz, C Velez, P Bringas

► **To cite this version:**

A Anadón, Mr Martinez-Larrañaga, Mj Diaz, C Velez, P Bringas. Pharmacokinetic and residue studies of quinolone compounds and olaquinox in poultry. *Annales de Recherches Vétérinaires*, 1990, 21 (suppl1), pp.137s-144s. hal-00902002

**HAL Id: hal-00902002**

**<https://hal.science/hal-00902002>**

Submitted on 11 May 2020

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

## Pharmacokinetic and residue studies of quinolone compounds and olaquinox in poultry

A Anadón, MR Martinez-Larrañaga, MJ Diaz, C Velez, P Bringas

*Department of Pharmacology, Institute of Pharmacology and Toxicology,  
CSIC, Faculty of Medicine, Complutense University, 28040 Madrid, Spain*

(Pharmacokinetics of Veterinary Drugs, 11–12 October 1989, Fougères, France)

**Summary** — Nalidixic acid and similar antimicrobial agents have been available for more than 20 years, mainly for treating infections caused by Gram-negative enterobacteria. Recently, several chemically related drugs, including oxolinic acid, pipemidic acid, piromidic acid and flumequine, have been developed. They are either naphthyridine-carboxylic acid or quinoline-carboxylic acid derivatives and, with nalidixic acid, are so-called quinolones. A major advance in antimicrobial chemotherapy was the synthesis of newer quinolones containing at least 1 fluorine atom and a piperazinyl group. These new fluoroquinolones have an extended antimicrobial spectrum compared to the first quinolone generation, and are highly active against most Gram-negative pathogens including the Enterobacteriaceae and *Pseudomonas aeruginosa*. The pharmacokinetic properties and residue levels of these quinolones and fluoroquinolones for which clinical experience or experimental information exists in poultry are reviewed here. On the other hand, administration of the quinoxaline-di-N-oxide, olaquinox, for medical purposes raises questions concerning the pharmacokinetic disposition of the drug and the risk of its residues in poultry. This paper presents information about the pharmacokinetic profile of olaquinox and the presence of its residues in chickens.

**quinolone / poultry / pharmacokinetics / residues**

**Résumé** — Étude pharmacocinétique des quinolones et de l'olaquinox chez la volaille. L'acide nalidixique et ses analogues structuraux sont utilisés dans le traitement des infections causées par des entérobactéries Gram négatif. Ces dernières années, des médicaments chimiquement voisins ont été développés : acide oxolinique, acide pipémidique, acide piromidique et fluméquine. Ces agents, appelés collectivement quinolones, dérivent de l'acide carboxylique-naphtyridine, de l'acide carboxylique-quinoline ou de l'acide nalidixique. Un progrès dans la chimiothérapie antimicrobienne a été réalisé avec la synthèse de nouvelles quinolones contenant au moins un atome de fluor et un groupe pipérazinyle. Ces fluoroquinolones ont un plus large spectre d'action antimicrobien; elles sont très actives sur la plupart des agents pathogènes à Gram négatif notamment les Enterobacteriaceae et *Pseudomonas aeruginosa*. Les propriétés pharmacocinétiques et les niveaux des résidus liés à l'utilisation des quinolones et fluoroquinolones, chez la volaille, sont présentées dans ce travail. Une attention particulière a été portée à l'olaquinox.

**quinolone / volaille / pharmacocinétique / résidus**

## INTRODUCTION

Among oral anti-bacterial agents, the quinolone class has been demonstrated to be effective in the treatment of *Escherichia coli* infections in poultry, especially colibacillosis in broilers. Nalidixic acid (a 1,8-naphthyridine derivative), the first agent in this series, and a number of other chemically related drugs (oxolinic acid, pipemidic acid, piromidic acid, flumequine), are active *in vitro* against a wide range of Gram-negative bacilli (with the exception of *Pseudomonas aeruginosa*) but inactive against Gram-positive organisms. In addition, the clinical use of first generation quinolones is often associated with the rapid emergence of resistant mutants (Fass, 1985). The development of new fluoroquinolone agents (norfloxacin, enoxacin, ciprofloxacin, ofloxacin, enrofloxacin, pefloxacin) with good systemic bioavailability and improved intrinsic antimicrobial activity especially against *P. aeruginosa* and Gram-positive organisms, has renewed interest in this class of antimicrobial agents.

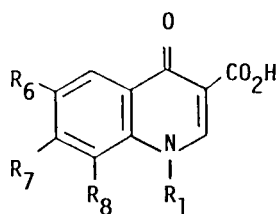
The primary target of nalidixic acid and oxolinic acid and probably all the other fluoroquinolones is DNA gyrase (topoisomerase II), an essential bacterial enzyme that maintains superhelical twists in DNA (Cozzarelli, 1980; Drlica, 1984; Gellert, 1981).

The theoretical advantage of fluoroquinolones led to the evaluation of their pharmacokinetic parameters in poultry and to assess their therapeutic potential and their residue levels in food-producing animals. The encouraging results obtained in the preliminary trials prompted us to review these agents. Other quinoxaline-di-N-oxide compounds, such as olaquinox, will also be discussed. The use of this drug in poultry for medical purpose (anti-bacterial activity) may or may not have a practical relevance.

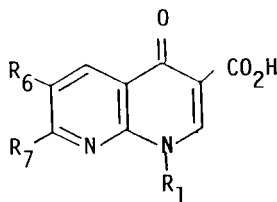
## STRUCTURE OF QUINOLONES

The general structures of two of the most studied classes of quinolones are shown in figure 1. Molecular modifications of the parent structures have been carried out in order to develop agents with higher potency and broader bacterial spectra. The results of structure-activity studies performed to date can be summarized as follows: maximum *in vitro* potency (expressed as MICs) and *in vivo* efficacy occur with a fluorine substituent at C-6 and with the concomitant presence of an amino functionality of optimal size at C-7 (fig 2).

Fortunately, information on the dissociation, solubility and solubility-pH relationship for nalidixic acid, a model for the newer quinolones, is available in the literature (Grubb, 1979; Staroscik and Sulkowska, 1971; Sulkowska and Staroscik, 1975). Nalidixic acid has two  $pK_a$ s which have been determined spectrophotometrically

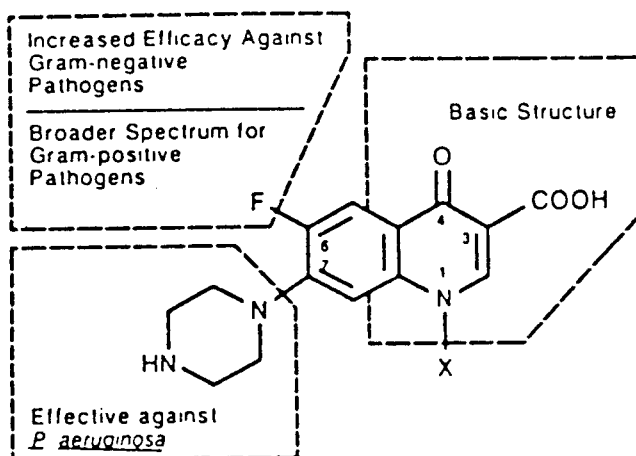


Quinolines



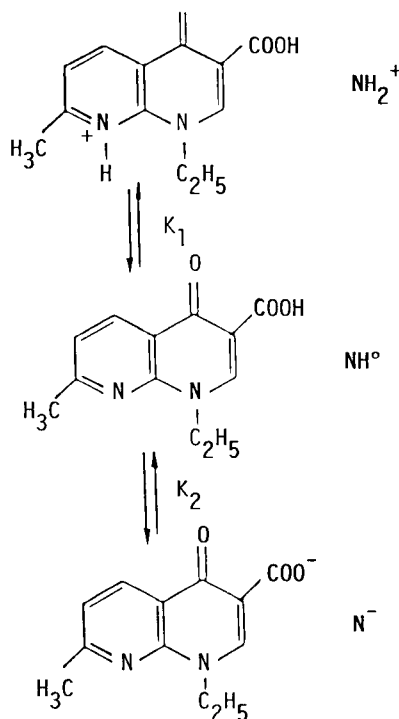
1,8-Naphthyridines

**Fig 1.** General structures of quinolone anti-bacterial agents.



**Fig 2.** Basic structural modifications of quinolones that contribute to their antimicrobial activity. X = a variety of possible substitutions (from Neer, 1988).

(Staroscik and Sulkowska, 1971) by solubility measurements (Sulkowska and Staroscik, 1975) and by partition studies (Grubb, 1979). The spectrophotometric  $pK_{a1}$  of 0.94 corresponds to the dissociation of a protonated heterocyclic nitrogen of nalidixic acid, while the spectrophotometric  $pK_{a2}$  value of 6.02 corresponds to the dissociation of the carboxylic acid group (Staroscik and Sulkowska, 1971). The dissociation scheme for nalidixic acid is given in figure 3. Their solubility-pH and partition-pH profiles have also been studied by Ogata *et al* (1984a) and Ismail and Gadalla (1983). The  $pK_a$ s determined by solubility were  $1.03 \pm 0.13$  and  $6.12 \pm 0.03$ , whereas those determined by partition measurements were  $0.86 \pm 0.07$  and  $5.99 \pm 0.03$ , respectively. In its neutral form ( $NH^0$ ), between pH values of 2 and 5, nalidixic acid has a solubility of  $8.3 \times 10^{-6}$  M (19  $\mu\text{g/ml}$ ) (Staroscik and Sulkowska, 1971). Most of the new quinolones have dissociation constants for their carboxyl group which are very similar to that of nali-



**Fig 3.** The dissociation scheme of nalidixic acid.

dixic acid. Substitution at the 7-position appears to have little electronic or steric effect on the dissociation of the carboxyl group. In contrast to nalidixic acid, the new quinolone antimicrobials have a basic functional group in the 7-position which has a much higher  $pK_a$  than the heterocyclic nitrogen. This has a profound effect on their solubility and partitioning properties which in turn significantly influence their pharmacological and biopharmacological properties. The  $pK_a$  values of several quinolone antimicrobials have been determined (Ogata *et al*, 1984a, b). It appears that the  $pK_a$  corresponding to the carboxylic group is around  $6.0 \pm 0.3$  and is relatively independent of substitution at the 7-position. On the other hand, the basic amine  $pK_a$  can vary between 5 and 9, depending upon the chemical nature of the side chain.

The structures of the six quinolones considered in the present report and

the structure of olaquinox are given in figure 4.

## PHARMACOKINETICS

The pharmacokinetic characteristics of 7 agents are shown in table I. After oral administration, these agents are more or less rapidly absorbed with peak plasma concentrations reached within 3 h. Piromidic acid, ciprofloxacin and olaquinox are the most rapidly absorbed, reaching a maximum level ( $T_{max}$ ) after 0.19–0.22 h following administration. Norfloxacin is absorbed more slowly ( $T_{max}$  0.30 h), but enrofloxacin, flumequine and oxolinic acid have the slowest rates of absorption with  $T_{max}$  of 1–2, 2 and 2.72 h, respectively. The peak plasma levels ( $C_{max}$ ) reached are also different and dose-dependent. Single oral doses of each of the 7 drugs considered in the present review are able to reach peak

**Table I.** Pharmacokinetic parameters for a single oral dose (mg/kg) of oxolinic acid (OXO), piromidic acid (PIRO), flumequine (FLUM), enrofloxacin (ENRO), norfloxacin (NOR), ciprofloxacin (CIP) or olaquinox (OLAQ) in chickens.

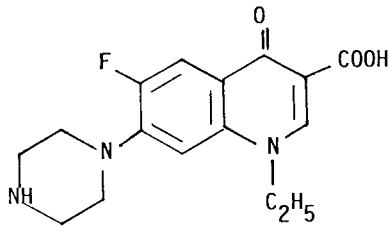
Plasma parameters	OXO <sup>a</sup>	PIRO <sup>b</sup>	FLUM <sup>c</sup>	NRO <sup>d</sup> (mg/kg)			NOR <sup>e</sup>	CIP <sup>f</sup>	OLAQ <sup>g</sup>
	15 mg/kg	10 mg/kg	12 mg/kg	2.5	5	10	8 mg/kg	8 mg/kg	20 mg/kg
$T_{max}$ (h)	2.72 (0.14)	0.19 (0.03)	2	1	1	2	0.30 (0.07)	0.22 (0.05)	0.22 (0.01)
$C_{max}$ (µg/ml)	11.93 (0.29)	1.65 (0.09)	5	0.5	0.6	1.4	1.95 (0.16)	3.54 (0.46)	29.96 (1.41)
$t_{1/2\beta}$ (h)	33.54 (3.88)	11.21 (0.77)	8	2–3.5 (0.4)			13.06 (1.18)	9.13 (0.30)	5.13 (0.19)
AUC (mg·h/l)	478.10 (33.10)	9.04 (0.74)	NA	NA			12.62 (0.69)	14.70 (1.36)	55.32 (2.28)

a, b, e, f, g Anadón *et al* (unpublished data). Mean data from 8 experiments are presented, with the SEM between parentheses. 40 d old male broiler chickens (*Hubbard x Hubbard*) weighing 2.5 kg were used. Oxolinic acid, piromidic acid, norfloxacin, ciprofloxacin and olaquinox were administered orally, directly into the crop. The drug plasma concentrations were determined by HPLC as previously described (Groeneveld and Brouwers, 1986; Horii *et al*, 1987; Boies, 1979). Plasma levels were fitted to a 2-compartment open model. The half-life of the  $\beta$  phase ( $t_{1/2\beta}$ ), the area under the concentration–time curve (AUC), the peak plasma level ( $C_{max}$ ), and the time needed to reach  $C_{max}$  ( $T_{max}$ ) were calculated (Wagner, 1976).

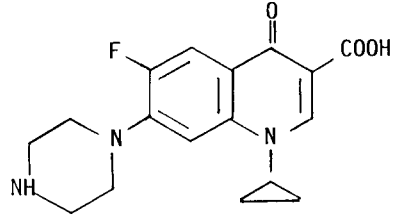
c Chevalier *et al* (1982). Flumequine dissolved in gum arabic was administered orally to Warren hens. The flumequine concentrations in plasma were determined using a microbiological assay.

d Sheer (1987). Eighteen 3–6 week old broiler chickens received enrofloxacin in drinking water. The enrofloxacin plasma concentrations were determined using a microbiological assay.

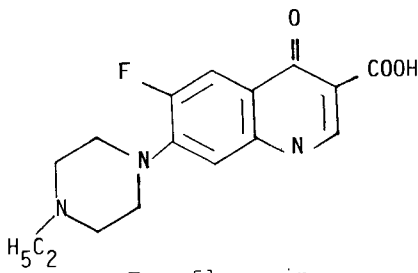
NA, data not available.



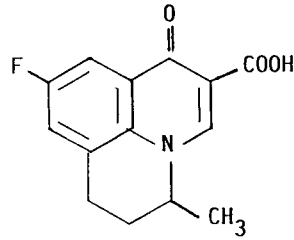
Norfloxacin



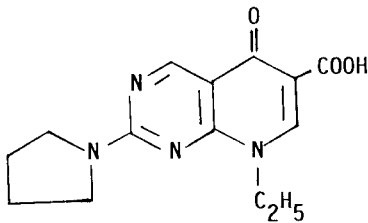
Ciprofloxacin



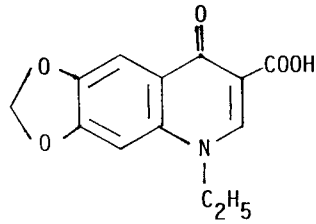
Enrofloxacin



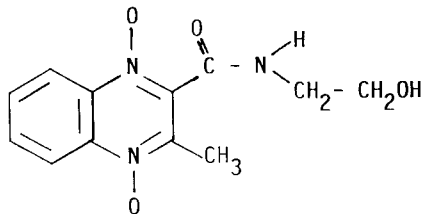
Flumequine



Piromidic acid



Oxolinic acid



Olaquinox

Fig 4. Structural formulae of quinolone and quinoxaline anti-bacterial drugs.

plasma levels above 1  $\mu\text{g/ml}$  (and thus above the *MIC* for many organisms). The differences in  $C_{\text{max}}$  values among different agents probably reflect variability in gastrointestinal absorption. When given orally at an equivalent dose, ciprofloxacin produces higher values of  $C_{\text{max}}$  and of the area under the curve (*AUC*) of the plasma level plotted *versus* time (*AUC* is related to bioavailability). The  $C_{\text{max}}$  of 3.54  $\mu\text{g/ml}$  following the administration of 8 mg/kg of ciprofloxacin is higher than that observed for the same dose of norfloxacin (1.95  $\mu\text{g/ml}$ ). The drugs considered, with the exception of enrofloxacin, persisted in the body of chickens for a long time. The mean terminal plasma elimination half-life ( $t_{1/2\beta}$ ) was 5.13 h for olaquinox, 8 h for flumequine, 9.13 h for ciprofloxacin, 11.21 h for piromidic acid, 13.06 h for norfloxacin and 33.54 for oxolinic acid. The mean  $t_{1/2\beta}$  of enrofloxacin is about 2–3.5 h.

The relatively long plasma elimination half-lives of norfloxacin and ciprofloxacin

that we observed in chickens are not in agreement with values reported by Sheer (1987) for enrofloxacin. These differences may result from the use of different analytical methods (microbiological assay for enrofloxacin vs HPLC assay for norfloxacin and ciprofloxacin).

## RESIDUE LEVELS

Few studies of tissue distribution and residue levels of quinolone and quinoxaline compounds in poultry have been reported. Table II summarizes the available results (Anadón *et al*, unpublished data). In poultry, the quinolone, oxolinic acid, piromidic acid and the quinoxaline compound, olaquinox, were widely distributed throughout the body with tissue concentrations exceeding 1  $\mu\text{g/ml}$  24 h after administration (table II). This good tissue penetration and high drug concentrations well above the

**Table II.** Tissue concentrations of oxolinic acid, piromidic acid and olaquinox after oral administration to chickens.

Treatment	Tissue	Drug concentration ( $\mu\text{g/g}$ ) on				
		day 1	day 3	day 6	day 8	day 14
Oxilinic acid 200 mg/kg single dose	muscle	1.46 (0.15)	0.57 (0.15)	0.02 (0.01)	ND	–
	liver	2.16 (0.15)	0.49 (0.06)	0.05 (0.01)	ND	–
	kidney	2.38 (0.35)	0.91 (0.45)	0.16 (0.05)	0.05 (0.01)	ND
Piromidic acid 10 mg/kg/d for 3 d	muscle	1.03 (0.16)	0.42 (0.09)	0.08 (0.01)	0.05 (0.01)	ND
	liver	1.19 (0.10)	0.71 (0.13)	0.28 (0.07)	0.05 (0.01)	ND
	kidney	2.06 (0.28)	1.18 (0.10)	0.21 (0.02)	0.06 (0.01)	ND
Olaquinox 20 mg/kg/d for 3 d	muscle	3.33 (0.84)	1.69 (0.51)	0.38 (0.08)	0.18 (0.04)	0.03 (0.01)
	liver	3.69 (0.50)	2.93 (0.38)	1.49 (0.33)	0.88 (0.22)	0.11 (0.01)
	kidney	1.43 (0.23)	2.23 (0.65)	1.92 (0.28)	1.34 (0.18)	0.12 (0.01)

Values are presented as means ( $\mu\text{g/g}$  of tissue) from 6 chickens with the SEM between parentheses. ND = not detected. 40 d old male broiler chickens (*Hubbard x Hubbard*) weighing 2.5 kg were used. Oxolinic acid, piromidic acid or olaquinox was administered orally, directly into the crop. The drug concentrations in tissues were determined by HPLC as previously described (Groeneveld and Brouwers, 1986; Horii *et al*, 1987; Bories, 1979).

*MIC* for most bacterial pathogens (Barry, 1989) suggest the potential clinical use to treat bacterial infections in poultry. In most species of Enterobacteriaceae, nalidixic acid and piperidic acid have similar activities (median *MICs* of 1.0–4.0  $\mu\text{g/ml}$ ); piperidic acid is less active against Gram-negative bacteria (median *MICs* of 1.0–10  $\mu\text{g/ml}$ ). The first compound in the quinolone series (oxolinic acid) is much more active; the median *MICs* for different species are  $\leq 0.5$   $\mu\text{g/ml}$ . Norfloxacin is 10–100-times more active than nalidixic acid. Other fluoroquinolones are also extremely active against the enteric bacilli; ciprofloxacin is the most potent quinolone reported (Barry, 1989). Among various quinoxaline-1,4-di-*N*-oxides synthesized to date, the growth promoter olaquinox is also used in the treatment and prevention of infectious diseases caused by diverse bacteria, such as *Salmonella* and *E coli* (Bertschinger, 1976) (median *MICs* of 8–16  $\mu\text{g/ml}$ ).

However, on the basis of detected residue levels, specific requirements might include a preslaughter withdrawal time. Assays of drug concentrations in muscle, liver and kidney showed that oxolinic acid, piperidic acid and olaquinox persisted for a long time in the body of chickens. Olaquinox was found to be the slowest drug eliminated from the body with an unchanged drug concentration of 0.03, 0.11 and 0.12  $\mu\text{g/g}$  of muscle, liver and kidney tissues, respectively, 14 d after oral drug administration.

No data have yet been published on the tissue penetration of new fluoroquinolones (norfloxacin, ciprofloxacin, ofloxacin, enoxacin and pefloxacin) in poultry. However, taking into account data obtained in other animal species and in man (Gilfillan *et al*, 1984; Hooper and Wolfson, 1985; Walker *et al*, 1989), they are also likely to have good tissue penetration. A study on the physiological disposition of enrofloxacin

has been reported (Sheer, 1987). After a single oral dose of 10 mg/kg to chickens, enrofloxacin was rapidly and widely distributed to tissues. Concentrations above 1  $\mu\text{g/g}$  were found 4–6 h after drug administration in lung, heart, liver, spleen, kidney and muscle. However, the drug was also eliminated quickly from tissues. Twenty-four hours after drug administration, the residue levels of enrofloxacin fell below 0.02–0.05  $\mu\text{g/g}$  of tissue. This indicates that the duration of the antimicrobial effect is shorter, since it depends upon the time during which the free drug concentration exceeds the *MIC* of susceptible pathogens (Baggot, 1980).

## CONCLUSIONS

The common pharmacokinetic properties of the quinolones are: 1) a rapid oral absorption, 2) attainable serum and tissue concentrations above the *MIC* for most Gram-negative and many Gram-positive organisms, and 3) relatively long half-lives in plasma, allowing dose intervals of at least 12 h. These features suggest possible clinical applications.

Although many investigations remain to be done, since olaquinox persists in the body of chickens for at least 14 d, this drug may pose a toxicological risk for man if a preslaughter withdrawal time is not observed. Currently, available data on oxolinic acid and piperidic acid residues recommend a preslaughter withdrawal time of 8 d.

## ACKNOWLEDGMENTS

This work was supported by the Comision Interministerial de Ciencia y Tecnologia, Programa Nacional de Investigacion y Desarrollo Farmaceutico, project FAR 88-0478, Spain.



## REFERENCES

- Baggot JD (1980) Distribution of antimicrobial agents in normal and diseased animals. *J Am Vet Med Assoc* 176, 1085-1090
- Barry AL (1989) *In vitro* activities of the quinolone antimicrobial agents. In: *International Telesymposium on Quinolones*. (Fernandes PB, ed) JR Prous Barcelona, 237-254
- Bertschinger HU (1976) Die chemotherapeutische wirksamkeit von olaquinox bei ferkein mit experimenteller colidiarrohoe und colienterotaxämie. *Schweiz Arch Tierheilkd* 118, 397-408
- Bories GF (1979) High-performance liquid chromatographic determination of olaquinox in feeds. *J Chromatogr* 172, 505-508
- Chevalier R, Oudar J, van Haverbeke G (1982) Pharmacologie et toxicologie de la fluméquine chez le poulet d'élevage et la poule pondeuse. In: *Pharmacologie et Toxicologie Vétérinaires. Colloq INRA* 8, INRA Publ, Paris, 99-104
- Cozzarelli NR (1980) DNA gyrase and the supercoiling of DNA. *Science* 207, 953-960
- Drlca K (1984) Biology of bacterial deoxyribonucleic acid topoisomerases. *Microbiol Rev* 48, 273-289
- Fass RJ (1985) The quinolones. *Ann Intern Med* 102, 400-402
- Gellert M (1981) DNA topoisomerases. *Annu Rev Biochem* 50, 879-910
- Gilfillan EC, Pelak BA, Bland JA, Malatesta PF, Gadebusch HH (1984) Pharmacokinetic studies of norfloxacin in laboratory animals. *Chemotherapy* 30, 288-296
- Groeneveld AJN, Brouwers JRB (1986) Quantitative determination of ofloxacin, ciprofloxacin, norfloxacin and pefloxacin in serum by high-pressure liquid chromatography. *Pharm Weekbl (Sci)* 8, 79-84
- Grubb PE (1979) Nalidixic acid. In: *Analytical Profiles of Drug Substances*, vol 8 (Florey K, ed) Academic Press, New York, 371-397
- Hooper DC, Wolfson JS (1985) The fluoroquinolones: pharmacology, clinical uses, and toxicities in humans. *Antimicrob Agents Chemother* 28, 716-721
- Horii S, Yasuoka C, Matsumoto M (1987) High-performance liquid chromatographic method for the simultaneous determination of oxolinic, nalidixic and piroimidic acids in cultured fish. *J Chromatogr* 388, 459-461
- Ismail AA, Gadalla MAF (1983) Effect of selected surfactants on nalidixic acid solubility. *Pharmazie* 38, 733-736
- Neer TM (1988) Clinical pharmacologic features of fluoroquinolone antimicrobial drugs. *J Am Vet Med Assoc* 193, 577-580
- Ogata H, Aoyagi N, Kaniwa N, Shibazaki T, Ejima A, Takasugi N, Mafune E, Hayashi T, Suwa K (1984a) Bioavailability of nalidixic acid from uncoated tablets in humans. Part II. Bioavailability in beagles and its correlation with bioavailability in humans and *in vitro* dissolution rates. *Int J Clin Pharm Ther Toxicol* 22, 240-245
- Ogata H, Aoyagi N, Kaniwa N, Ejima A (1984b) Effect of food on bioavailability of nalidixic acid from uncoated tablets having different dissolution rates. *J Pharmacobiodyn* 7, 760-767
- Sheer M (1987) Concentrations of active ingredient in the serum and in tissues after oral and parenteral administration of baytril. *Vet Med Rev* 2, 104-118
- Staroscik R, Sulkowska J (1971) Studies on acid-base equilibria of nalidixic acid. *Acta Polon Pharm* 28, 603-609
- Sulkowska J, Staroscik R (1975) Study on distribution equilibria of nalidixic acid. *Pharmazie* 30, 405-406
- Wagner JG (1976) Linear pharmacokinetic equations allowing direct calculation of many needed pharmacokinetic parameters from the coefficients and exponents of polyexponential equations which have been fitted to the data. *J Pharmacokinet Biopharm* 4, 443-467
- Walker RD, Stein GE, Budsberg SC, Rosser EJ, MacDonald KH (1989) Serum and tissue fluid norfloxacin concentrations after oral administration of the drug to healthy dogs. *Am J Vet Res* 50, 154-157