



HAL
open science

Bovine colostrum as substrate for the preparation of growth factor-enriched protein extracts: Identifying the optimal collection period during lactation

Alicia Montoni, Sylvie F. Gauthier, Caroline Richard, Patrice E. Poubelle,
Yvan Chouinard, Yves Pouliot

► **To cite this version:**

Alicia Montoni, Sylvie F. Gauthier, Caroline Richard, Patrice E. Poubelle, Yvan Chouinard, et al.. Bovine colostrum as substrate for the preparation of growth factor-enriched protein extracts: Identifying the optimal collection period during lactation. *Dairy Science & Technology*, 2009, 89 (5), 10.1051/dst/2009028 . hal-00895709

HAL Id: hal-00895709

<https://hal.science/hal-00895709>

Submitted on 11 May 2020

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Bovine colostrum as substrate for the preparation of growth factor-enriched protein extracts: Identifying the optimal collection period during lactation

Alicia MONTONI¹, Sylvie F. GAUTHIER^{1*}, Caroline RICHARD¹, Patrice E. POUBELLE²,
Yvan CHOUINARD¹, Yves POULIOT¹

¹ STELA Dairy Research Center and Institute of Nutraceuticals and Functional Foods (INAF),
Université Laval, Quebec City, QC, G1V 0A6, Canada

² Centre de recherche en Rhumatologie et Immunologie (CRCHUL), Quebec City, QC,
G1V 4G2, Canada

Received 28 November 2008 – Accepted 9 June 2009

Published online 25 July 2009

Abstract – A growing number of growth factor-enriched bioactive protein extracts derived from colostrum are proposed for the treatment of inflammatory and cutaneous diseases. However, growth factor concentrations peak in the first secretions after parturition and vary widely among cows. The aim of this study was to identify the optimal collection period for colostrum used to prepare growth factor-enriched protein extracts and to characterize the relationship between growth factor and protein contents in colostrum and in milk collected during early lactation. Lactal secretions from five Holstein cows from the first milking (< 12 h) to the 21st day after parturition were analysed for total protein and growth factor (TGF- β 2, TGF- β 1 and IGF-I) contents. Although concentrations varied widely between cows, statistically significant decreases were obtained for all components during the early stages of lactation. The concentrations of each growth factor during lactation were highly correlated ($R^2 > 0.93$) and were also correlated with the total protein content ($R^2 > 0.90$). The greatest decreases in the concentrations of the three growth factors (78–84%) were observed within 24–48 h after parturition, while values stabilized in mature milk. However, relative to protein, these decreases appeared much smaller (37–45%). Overall, these results suggest that the first 48 h after parturition would be optimal for harvesting colostrum for the production of growth factor-enriched protein extracts.

bovine colostrum / bovine milk / growth factor / IFG-I / TGF- β 1 / TGF- β 2 / growth factor-enriched ingredient

摘要 – 制备富含生长因子的牛初乳蛋白提取物：泌乳期内最佳收乳时间的确定。源于牛初乳的富含生长因子的活性蛋白提取物在治疗炎症和皮肤疾病中的显著疗效引起了人们广泛关注。然而，在泌乳初期分泌生长因子浓度的高峰值在奶牛个体之间变化幅度非常大。本研究目的是确定用于制备富含生长因子的牛初乳的最佳采乳时间，并且确定生长因子与初乳和泌乳初期原料乳中蛋白质含量之间的关系。分析了5头荷斯坦奶牛（分娩后12 h到21天）乳样中生长因子（TGF-2, TGF-1, IGF-I）和蛋白质含量。在5头

*Corresponding author (通讯作者): Sylvie.Gauthier@fsaa.ulaval.ca

奶牛之间，所有分析指标的变化幅度较大，但是，所有指标在整个泌乳初期内随着时间的增加而显著降低。而且在整个泌乳期内，每种生长因子的浓度之间 ($R^2 > 0.90$) 及与蛋白质浓度之间 ($R^2 > 0.90$) 具有非常高的相关性。在泌乳后的 24 h ~ 48 h 内三种生长因子浓度降低的幅度非常大 (78% ~ 84%)，但是在常乳中生长因子浓度变化不大。蛋白质浓度的变化幅度较小 (37% ~ 45%)。综上所述，如果生产富含生长因子的蛋白提取物，泌乳后 48 h 内是采集牛初乳的最佳时间。

牛初乳 / 牛乳 / 生长因子 / IFG-I / TGF- β 1 / TGF- β 2 / 富含生长因子的配料

Résumé – Le colostrum bovin comme substrat pour la préparation d’extraits protéiques enrichis en facteurs de croissance : identification de la période optimale de collecte au cours de la lactation. Un nombre croissant d’ingrédients protéiques bioactifs à base de facteurs de croissance issus du colostrum sont proposés pour le traitement des maladies inflammatoires et cutanées. Cependant, l’utilisation du colostrum comme matière première est limitée par l’apparition d’une forte concentration de ces molécules dans les premières sécrétions suivant le vêlage et par la grande variabilité de leur concentration entre les animaux. Le but de la présente étude était d’identifier la période optimale de collecte du colostrum pour la préparation d’extraits protéiques enrichis en facteurs de croissance et de caractériser la relation entre le contenu en facteurs de croissance et en protéines d’échantillons de colostrum et de lait collectés en début de lactation. Les sécrétions lactées de cinq vaches Holstein ont été collectées de la première traite (< 12 h) jusqu’au 21^e jour suivant le vêlage, puis analysées pour leur teneur en protéines totales et en facteurs de croissance (TGF- β 2, TGF- β 1, IGF-I). Malgré la grande variabilité de la concentration de ces composantes observée entre les vaches, des différences statistiquement significatives ont été observées dès le début de la lactation. Les diminutions dans la concentration en facteurs de croissance des sécrétions lactées en cours de lactation étaient fortement inter-reliées ($R^2 > 0,93$), mais aussi corrélées ($R^2 > 0,90$) avec la teneur en protéines totales des échantillons. Les diminutions les plus élevées dans la concentration des trois facteurs de croissance (78–84 %) ont été observées après 24–48 h suivant le vêlage, pour atteindre des valeurs stables dans le lait mature. Cependant, lorsque les concentrations en facteurs de croissance étaient exprimées sur base protéique, ces diminutions étaient considérablement réduites (37–45 %) au cours de la même période. Les résultats suggèrent que les premières 48 h suivant le vêlage constituent la période optimale pour la collecte du colostrum en vue de la production d’ingrédients protéiques enrichis en facteurs de croissance.

colostrum bovin / lait bovin / facteurs de croissance / IFG-I / TGF- β 1 / TGF- β 2 / ingrédients enrichis en facteurs de croissance

1. INTRODUCTION

Colostrum, the first lacteal secretion of the mammary glands following calving, is known to provide passive immunity and all the nutrients essential to the development of the newborn calf [3]. It contains immunoglobulins and other antimicrobial factors, growth factors, cytokines, hormones and other biologically active molecules [10, 20]. The growth factors found in significant amounts in colostrum are transforming growth factors-beta 2 (TGF- β 2) and insulin-like growth factor-I (IGF-I), for which

wide concentration variations (150–1500 and 49–2000 ng·mL⁻¹, respectively) have been reported [8]. The physiological effects of TGF- β -related growth factors and IGFs in colostrum and milk have been reviewed by Pakkanen and Aalto [15] and Gauthier et al. [8]. Several health-related applications of milk growth factors have been proposed such as for skin disorders, gut health, bone health and others [17]. Characterization of the numerous protein components in bovine colostrum, milk and whey has been done recently using the proteomic approach [7, 19], confirming the diversity of

molecules present in these products. These new data are likely to stimulate the development of new colostrum-based bioactive protein ingredients. However, the known variability of colostrum composition during the first hours of lactation still represents a major challenge in the commercial-scale production of colostrum-based ingredients.

Technological approaches to prepare bioactive extracts containing growth factors found in bovine colostrum, milk or whey are growing in number [17]. Maubois et al. [13] proposed a combination of acidification/heat treatment of native (unheated) whey to precipitate a TGF- β 2-rich fraction that was further recovered by tangential microfiltration. This method also enabled to fractionate whey proteins but was not applicable to bovine colostrum due to the presence of caseins that precipitate at their isoelectric point. Piot et al. [16] showed the potential of microfiltration for the removal of caseins from colostrum, while Lachkar et al. [11] proposed complexing with polysaccharides as an alternative approach of preparing a TGF- β 2-enriched fraction from bovine colostrum. Recent work by Akbache et al. [1] has suggested that ultrafiltration/microfiltration membranes can be used to concentrate TGF- β 2 and IGF-I from whey.

Despite the recent technological advances, only a few studies have focussed on the variability of colostrum composition as a function of time during early lactation. Elfstrand et al. [6] investigated compositional changes in growth factors in pooled colostrum collected 0–80 h postpartum from Swedish Friesian cows, demonstrating a rapid decrease in TGF- β 2 and IGF-I within the first 24 h. In a recent study, Purup et al. [18] showed that the concentration of these growth factors decreased during lactation in Holstein Friesian cows. These authors also showed that whey separated from colostrum and milk stimulated growth of a human intestinal cell line to different extents depending on the stage of lactation. Changes in growth-promoting activity with

stage of lactation were also observed and were attributed to changes in concentrations of minor bioactive compounds such as growth factors.

The aim of this study was to identify the optimal collection period for obtaining the starting material suitable for the preparation of growth factor-enriched protein extracts from colostrum. Although it is known that growth factor concentrations decrease during lactation, values published for bovine colostrum and milk vary to a large extent, probably because they are never expressed relative to protein content [8]. The aim of this study was therefore also to characterize the relationship between the growth factors and the total protein content of colostrum and milk collected during early lactation.

2. MATERIALS AND METHODS

2.1. Animals and sampling

Colostrum samples (total lacteal secretion) collected within 24 h from 14 Holstein cows aged 23–110 months (primipara or 2–7-calf multipara heifers) at the Laval University Animal Sciences Research Center (Deschambault, QC, Canada) were analysed for total protein, TGF- β 2, TGF- β 1 and IGF-I contents. In a second phase, five cows were selected for a 21-day sampling experiment. These were 24–81 months old and multipara (2–4 calves) or primipara in one case. They were housed in a tie-stall barn and given individually a single feeding per day ad libitum in the morning. The diet consisted of a total mixed ratio formulated to meet the National Research Council recommendations for transition cows [14], containing alfalfa silage, corn silage, grass hay, cracked corn, rolled barley, beet pulp and commercial energy (Energina, Agribands Purina Canada Inc., Saint-Hubert, QC, Canada) and protein supplements (B-Top 45, Aliments Breton Inc., Saint-Bernard, QC, Canada).

The first colostrum sample was collected as soon as possible after parturition (< 12 h), followed by twice-daily collections (approximately at 8 and 17 h) for the next six days. Additional samples were collected from each cow on the 7th, 14th and 21st days postpartum once in the morning. All samples were immediately frozen and kept at -18°C .

2.2. Defatting of colostrum and milk

Colostrum samples (1 L) were thawed overnight at 4°C and then for 2–3 h at room temperature until entirely liquid. They were skimmed by centrifugation at $10\,000\times g$ for 30 min at 4°C (Sorvall model RC-5B, GSA rotor, DuPont Instruments, Boston, MA, USA) and the solid fat layer was then carefully removed manually. The defatted colostrum samples were refrozen without delay at -20°C until analysis.

2.3. Protein analysis

The total protein concentration was determined by the Dumas combustion method [9] using a FP-528 LECO apparatus (LECO Corp., Saint Joseph, MI, USA). The calibration standard curve was prepared using ammonium sulphate (99.99% w/w, Sigma-Aldrich, Oakville, ON, Canada) and the nitrogen content was converted to protein using the milk conversion factor (6.38).

2.4. Growth factors quantification

IGF-I, TGF- β 1 and TGF- β 2 contents were determined by enzyme-linked immunosorbent assay (ELISA) using the methodology described by Ben Ounis et al. [2], except for adjusting the sample dilutions to the range of growth factor concentrations in colostrum and milk.

Absorbance readings were performed at 450 nm for the three growth factors using

a spectrophotometer plate reader (Multi Skan Spectrum model, Thermolabsystems, Brussels, Belgium). The assays were performed in triplicate (three wells per sample). The detection limit (DL) of the ELISA tests was established at $0.031\text{ ng}\cdot\text{mL}^{-1}$ for TGF- β 1 and TGF- β 2, and at $0.023\text{ ng}\cdot\text{mL}^{-1}$ for IGF-I.

2.5. Statistical analysis

The concentrations of total proteins and the three growth factors in colostrum and milk collected during the lactation period were submitted to statistical analysis using one-way analysis of variance and paired *t* tests between two consecutive samplings using SAS version 9.1. Also, linear regression analysis was performed on the total protein, TGF- β 2, TGF- β 1 and IGF-I concentrations for each colostrum and milk sample over the lactation period in order to establish their mutual coefficients of determination (R^2).

3. RESULTS

The concentrations of total proteins, TGF- β 2, TGF- β 1 and IGF-I in colostrum collected during the first day postpartum (< 24 h) were first determined for 14 individual cows. A wide range of values were obtained for all components. Total proteins ranged from 80.3 to $220.1\text{ mg}\cdot\text{mL}^{-1}$ (mean \pm SD of 155.2 ± 40.9), whereas TGF- β 2, TGF- β 1 and IGF-I ranged from 73 to $436\text{ ng}\cdot\text{mL}^{-1}$ (mean \pm SD of 213.9 ± 21.0), from 9 to $56\text{ ng}\cdot\text{mL}^{-1}$ (mean \pm SD of 20.6 ± 2.8) and from 4 to $247\text{ ng}\cdot\text{mL}^{-1}$ (mean \pm SD of 90.9 ± 11.8), respectively. No significant statistical correlation could be established (data not shown) between the concentration of these components in the colostrum and the cow age or number of calvings.

Given the high variability observed in the colostrums collected from the 14 cows during the first day postpartum (< 24 h),

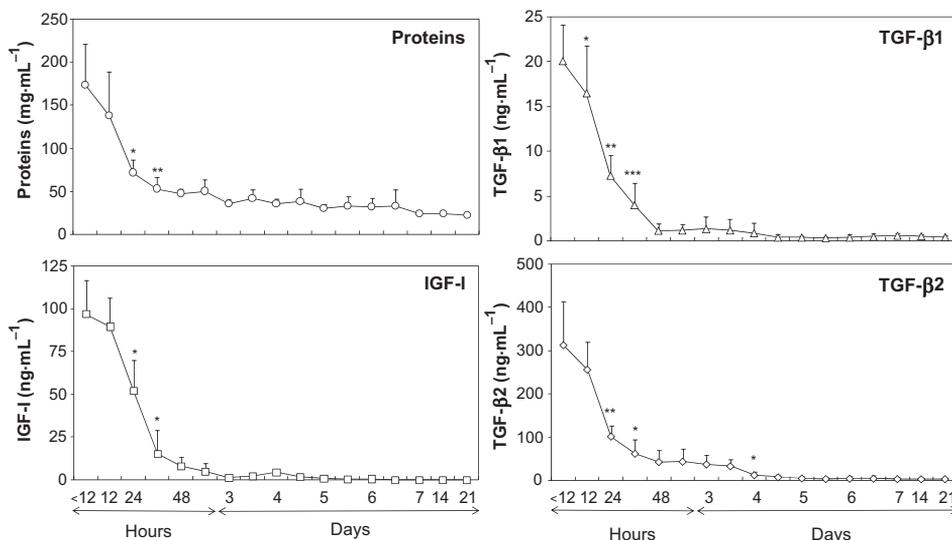


Figure 1. Evolution of total protein, IGF-I, TGF- β 1 and TGF- β 2 concentrations in the first milking (< 12 h) colostrum and milk collected from five Holstein cows over a 21-day post-parturition period. Values are reported as mean \pm SD of the mean and significant differences between two consecutive values are indicated (* $P < 0.05$; ** $P < 0.01$; and *** $P < 0.001$).

a second experimental phase including a smaller number of cows ($n = 5$) was designed in order to characterize the changes in total proteins, TGF- β 2, TGF- β 1 and IGF-I from the first milking after parturition (< 12 h) to the 21st day postpartum. As shown in Figure 1, wide variability was observed among the cows for all components analysed, especially during the first 48 h period, as indicated by the error bars. However, a paired t test statistical analysis of the data showed significant decreases already at 12 h ($P < 0.05$) and 24 h (3rd milking, $P < 0.01$; 4th milking, $P < 0.001$) for TGF- β 1, while significant decreases in total proteins (3rd milking, $P < 0.05$; 4th milking, $P < 0.01$), IGF-I (3rd and 4th milkings, $P < 0.05$) and TGF- β 2 (3rd milking, $P < 0.01$; 4th milking, $P < 0.05$) occurred only at 24 h. A smaller but significant ($P < 0.05$) decrease also occurred for TGF- β 2 on the 4th day. This figure shows that the first 48 h after parturition represent the

most critical period with respect to the growth factor content of colostrum.

In view of the similarities of the time-dependent decreases for the components investigated (Fig. 1), coefficients of determination (R^2) were calculated by linear regression between the growth factors and also between each growth factor and the total protein content for each cow during the lactation period (data not shown). The calculated R^2 values were higher than 0.90 for all three growth factors (0.93–0.95) and for their comparisons with total proteins (0.90–0.92), indicating that the growth factor contents of the lacteal secretions for 21 days postpartum were directly related to total protein content, even though the amount of growth factor was proportionally very low compared to protein (Fig. 1).

From the trends observed in Figure 1, the compositional data were grouped in five stages of lactation, namely first (< 12 h) and second milkings (12 h), 3rd to 6th milkings

Table I. Growth factor concentrations relative to protein and ratios of protein extracts at different stages of lactation.

Growth factors	Stage of lactation				
	< 12 h	12 h	24–48 h	3–6 days	7–21 days
TGF- β 2 (ng·g protein ⁻¹)	1799	1855	1131	414	169
TGF- β 1 (ng·g protein ⁻¹)	115	119	63	20	22
IGF-I (ng·g protein ⁻¹)	568	647	321	48	< DL ¹
TGF- β 2/TGF- β 1	15.6	15.6	18.0	20.7	7.7
TGF- β 1 + TGF- β 2/IGF-I	3.4	3.1	3.7	9.0	< DL

¹ Below the detection limit of ELISA test for IGF-I.

(24–48 h), transition milk (3–6 days) and mature milk (7–21 days). The relative changes (%) in total proteins and growth factors for each stage of lactation (data not shown) indicated that the TGF- β 2, TGF- β 1 and IGF-I concentrations in colostrum decreased by 78–84% from the initial value within the first 48 h. Although a major decrease (67%) in total protein concentration was also observed during this period, the subsequent decrease during the lactation period (3–21 days) was much less than for the three growth factors.

Table I presents growth factor contents relative to total protein for colostrum or milk collected during the five stages of lactation, along with the TGF- β 2/TGF- β 1 and TGF- β 1 + TGF- β 2/IGF-I concentration ratios. These values are important from the perspective of industrial production of protein extracts rich in growth factors. The data show that TGF- β 2, TGF- β 1 and IGF-I contents decreased, respectively, from 1799 to 1131 ng·g protein⁻¹, from 115 to 63 ng·g protein⁻¹ and from 568 to 321 ng·g protein⁻¹ during the < 12–48 h period, corresponding to a reduction of 37–45% in growth factor contents of the protein extract. This reduction is much lower than those based on liquid volume (78–84%). The TGF- β 2/TGF- β 1 ratio varied between 15.6 and 18.0 during the same period (< 12–48 h), while TGF- β 1 + TGF- β 2/IGF-I varied between 3.1 and 3.7,

indicating that the protein-based composition did not vary to a great extent during this period. However, the TGF- β 1 + TGF- β 2/IGF-I ratio increased to 9.0 during the 3–6 days period, mainly as a result of a major decrease in IGF-I.

4. DISCUSSION

The compositional data obtained in the present study are in good agreement with those published by Purup et al. [18] and Elfstrand et al. [6] on the early changes postpartum in total protein and TGF- β 2 contents, despite the difference in animal breed. The main discrepancy was for IGF-I, which was 5- to 10-fold lower in this study, compared to Elfstrand et al. [6] who used radioimmunoassay. TGF- β 1 was not determined by Elfstrand et al. [6], but our data are very close to those of Purup et al. [18].

The changes in total protein concentrations reported in this study are also in good agreement with those reported by Božanić [5], Blum and Hammon [4] and Levieux and Ollier [12]. As noted by Gauthier et al. [8], the marked decrease in total protein concentration during the first four days of lactation is generally not taken into account in published growth factor concentration data. The strong correlations

($R^2 > 0.90$) found between the three growth factors and the total protein contents in our study suggest that growth factor concentrations in colostrum and milk should be expressed per gram of protein and that this would probably reduce some of the discrepancies between the published data and thus facilitate their comparison.

The growth factor contents found in colostrum and milk collected during the first month postpartum suggest that the first 12 h should be targeted to maximize concentration in growth factor-enriched ingredients, although the compositional data are still variable. In fact, the first milking postpartum would represent the ideal condition. However, as seen in [Table I](#), when expressed relative to protein, the decreases in TGF- β 2, TGF- β 1 and IGF-I during the first 48 h are 37%, 45% and 43%, respectively, compared to the 78–84% range previously found when expressed per volume of liquid ([Fig. 1](#)). Moreover, the relative proportions of TGF- β 2, TGF- β 1 and IGF-I in colostrum protein-based extracts would not vary to a large extent within the first 48 h of lactation. It is of interest that should the purpose of the collection be to produce a TGF- β -enriched ingredient with the lowest amount of IGF-I, the optimal collection period might be days 3–6, when the TGF- β 1 + TGF- β 2/IGF-I ratio reaches 9.0 ([Tab. I](#)). This results from the fact that the IGF-I concentration is lowest, while TGF- β 2 and TGF- β 1 contents are low but still found at a useful concentration.

5. CONCLUSION

The results from this study underline the technological and economical challenges of using colostrum as a starting material for the production of growth factor-enriched protein extracts. Collecting and processing separately the first colostrum postpartum (< 12 h) would be difficult to achieve in practice because of the low volumes of

colostrum produced. As a consequence, most commercial-scale colostrum producers are currently pooling milkings up to 24 h and those up to 48 h separately. However, our results suggest that, expressed relative to protein, the growth factor content of colostrum would be optimal during the first 48 h after parturition.

Acknowledgments: This work was supported by a Cooperative Research and Development (CRD) grant from the Natural Sciences and Engineering Research Council of Canada (NSERC) and Advitech Inc. The authors thank the Deschambault Research Center for the collaboration of their staff with this work and L. Troquier for her technical assistance with the analysis of samples.

REFERENCES

- [1] Akbache A., Lamiot E., Moroni O., Turgeon S.L., Gauthier S.F., Pouliot Y., Use of membrane processing to concentrate TGF- β 2 and IGF-I from bovine milk and whey, *J. Membr. Sci.* 326 (2009) 435–440.
- [2] Ben Ounis W., Gauthier S.F., Turgeon S.L., Roufik S., Pouliot Y., Separation of minor protein components from whey protein isolates by heparin affinity chromatography, *Int. Dairy J.* 18 (2008) 1043–1050.
- [3] Blum J.W., Nutritional physiology of neonatal calves, *J. Anim. Physiol. Anim. Nutr.* 90 (2006) 1–11.
- [4] Blum J.W., Hammon H., Colostrum effects on the gastrointestinal tract, and on nutritional, endocrine and metabolic parameters in neonatal calves, *Livest. Prod. Sci.* 66 (2000) 151–159.
- [5] Božanić R., [Importance and utilization of colostrum], *Mljekarstvo* 54 (2004) 209–224 (in Croatian).
- [6] Elfstrand L., Lindmark-Månsson H., Paulsson M., Nyberg L., Åkesson B., Immunoglobulins, growth factors and growth hormone in bovine colostrum and the effects of processing, *Int. Dairy J.* 12 (2002) 879–887.
- [7] Fong Y.B., Norris C.S., Palmano K.P., Fractionation of bovine whey proteins and

- characterisation by proteomic techniques, *Int. Dairy J.* 18 (2008) 23–46.
- [8] Gauthier S.F., Pouliot Y., Maubois J.L., Growth factors from bovine milk and colostrum: composition, extraction and biological activities, *Lait* 86 (2006) 99–126.
- [9] IDF, Determination of nitrogen content – routine method using combustion according to the Dumas principle, *Int. Dairy Fed.*, Brussels, Belgium 186 (2002).
- [10] Korhonen H., Pihlanto A., Technological options for the production of health-promoting proteins and peptides derived from milk and colostrum, *Curr. Pharm. Des.* 13 (2007) 829–843.
- [11] Lachkar D., Lamiot E., Turgeon S.L., Gauthier S.F., Paquin P., Pouliot Y., An experimental approach for removing caseins from bovine colostrum using anionic polysaccharides, *Int. J. Dairy Sci.* 61 (2008) 43–50.
- [12] Levieux D., Ollier A., Bovine immunoglobulin G, β -lactoglobulin, α -lactalbumin and serum albumin in colostrum and milk during the early post partum period, *J. Dairy Res.* 66 (1999) 421–430.
- [13] Maubois J.L., Fauquant J., Jouan P., Bourtourault M., Method for obtaining a TGF-beta enriched protein fraction activated form, protein fraction and therapeutic applications, PCT Pat. No. WO 03/006500 (2003).
- [14] National Research Council, Nutrient Requirements of Dairy Cattle, 7th revised edn., National Academy Press, Washington, DC, USA (2001) 184–213.
- [15] Pakkanen R., Aalto J., Growth factors and antimicrobial factors of bovine colostrums, *Int. Dairy J.* 7 (1997) 285–297.
- [16] Piot M., Fauquant J., Madec M.N., Maubois J.L., Preparation of serocolostrum by membrane microfiltration, *Lait* 84 (2004) 333–341.
- [17] Pouliot Y., Gauthier S.F., Milk growth factors as health products: some technological aspects, *Int. Dairy J.* 16 (2006) 1415–1420.
- [18] Purup S., Vestergaard M., Pedersen L.O., Sejrsen K., Biological activity of bovine milk on proliferation of human intestinal cells, *J. Dairy Res.* 74 (2007) 58–65.
- [19] Smolenski G., Haines S., Kwan F.Y., Bond J., Farr V., Davis S.R., Stelwagen K., Wheeler T.T., Characterisation of host defence proteins in milk using a proteomic approach, *J. Proteome Res.* 6 (2007) 207–215.
- [20] Stelwagen K., Carpenter E., Haigh B., Hodgkinson A., Wheeler T.T., Immune components of bovine colostrum and milk, *J. Anim. Sci.* 87 (Suppl. 1) (2009) 3–9.