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Selecting honey bees for resistance to *Varroa jacobsoni*

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Abstract – This report describes a systematic approach to selecting honey bees (*Apis mellifera* L.) for resistance to *Varroa jacobsoni* Oudemans. The equation $(P_1(ab)^n = P_2)$ describes the growth of the mite population in a colony of honey bees that has a constant supply of worker brood. P_1 and P_2 are the initial and final mite populations, a is population change while mites are in brood cells, b is population change outside brood cells and n is the number of reproductive cycles of the mite. By comparing the growth of mite populations in each colony (P_2/P_1), one can determine which bees are more resistant to mites. The values of a , b and n provide details about the growth of the mite population by identifying which portion of the mite's reproductive cycle was affected. Selection should be based on specific characteristics of bees rather than on general changes in mite populations. When specific characteristics of bees affect different components of the reproductive cycle of the mite, it may be possible to combine the characteristics to produce bees that are more resistant to mites.
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Apis mellifera / mite / *Varroa jacobsoni* / resistant population / selective breeding

1. INTRODUCTION

This report is a review of our approach to select honey bees (*Apis mellifera* L.) for resistance to *Varroa jacobsoni* Oudemans (Acari: Varroidae). We outline a systematic approach which should be effective in any selection program that involves characteristics of honey bees that are measured at the colony level.

We define mite resistance as the ability of a colony of honey bees to impede the growth of a population of *V. jacobsoni*. With this definition, a highly resistant colony of bees would cause a mite population to decline and then to either disappear or be maintained at a very low level. This is the breeding objective. However, during the breeding process, especially at the beginning, a breeder may select colonies that measur-

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ably slow the growth of their mite populations but would eventually die from the mite infestation. These susceptible colonies with resistant characteristics can be important in the selection process, especially in difficult selection projects (such as resistance to *V. jacobsoni*), and accurate measurements are needed to identify the colonies that are slightly better than others. Therefore, selective breeding of bees for resistance to *V. jacobsoni* relies on our ability to accurately measure and describe the growth of mite populations within colonies of bees.

The first step in selective breeding is to choose a population of bees within which to work. It is best if this population contains some colonies with the desired phenotype (in this case resistance to mites), but the population may express only low levels of resistance, or even undetectable levels. For example, the importation of bees from far eastern Russia [11, 51] is the beginning of a breeding program that began with stock that already contained some of the desirable qualities.

In bee breeding, man can do things that nature cannot do. These are: 1) special mating schemes (e.g. single drone inseminations or inseminating many queens with the same mixture of semen); 2) the ability to focus selection on one characteristic at a time; and 3) the ability to select susceptible colonies that contain characteristics of resistance (colonies that would eventually die in nature). There is no need for colonies to die in a breeding program that selects bees for resistance to mites.

The use of single-drone inseminations may be important at the beginning of a selection program, especially when selecting for colony traits that are present at low frequencies. When a queen is mated to semen from a single drone, worker bees in the colony all have identical genetic material from their father, who is represented by identical spermatozoa that are now in the spermatheca of the queen. Thus the worker bees in such colonies are more closely

related than normal sisters and have a relatedness of 0.75. The use of single-drone matings makes it easier to detect colony characteristics that may be masked by multiple mating, and it amplifies the differences among colonies [26, 55].

Advantage may shift to multiple-drone inseminations during the mid and latter stages of selective breeding [28]. The reasons are that: 1) queens survive longer when inseminated with multiple drones [8, 26]; 2) more effective selection schemes can be used (such as mating a group of queens with the same mixture of semen); and 3) daughter queens from a multiply mated queen are more variable and would therefore reduce the rate of inbreeding at a time when the specific characteristic of selection is well-established.

We used the following sequence in our approach to breed bees for resistance to *V. jacobsoni*:

- 1) develop techniques for measuring populations of bees and mites and for measuring characteristics that are associated with resistance;
- 2) identify specific characteristics that are related to the growth of mite populations;
- 3) determine if these characteristics are heritable;
- 4) enhance heritable characteristics with selective breeding;
- 5) assemble resistant components into productive, mite-resistant bees.

2. EVALUATION PROCEDURE

2.1. Establishing heritability

It is important to calculate heritability (h^2) of a desirable characteristic before beginning a program of selective breeding. Heritability (h^2) is the proportion of the observed variance (among a group of bee colonies in this case) for which differences in heredity are responsible [35]. The estimate of h^2 is a pragmatic measurement that

predicts breeding success. If a characteristic has an h^2 close to 1, then the characteristic can be rapidly changed with selective breeding. If h^2 approaches 0, selective breeding will probably fail. As a general rule, it is reasonable to attempt selective breeding if $h^2 > 0.25$.

We used sibling analysis [10] to estimate heritability of various characteristics of bees that may be associated with the growth of mite populations (table 1, [27]). The heritability test consisted of 28 colonies with unrelated queens that had not been selected for resistance to *V. jacobsoni*. The relatedness of the colonies was established by inseminating groups of four queens with a single mixture of semen that had been collected from the drones produced by one queen. Seven such queens that served as drone mothers for the experiment were unrelated to each other and were unrelated to

the queens that were inseminated. This produced seven groups of four colonies, with each colony related as a full sister to the other three colonies in its group and unrelated to the other 24 colonies [27].

2.2. Three levels of evaluation and measurement

We always measured the growth of a mite population within a colony of bees that was being evaluated for resistance. A field test is the core of the evaluation with the honey bee colony serving as the experimental unit in the analyses. The evaluation requires at least 10 weeks and measures growth of mite populations in a group of honey bee colonies and provides a framework for measuring other characteristics at appropriate times during the course of the

Table 1. Heritability of colony characteristics based on sibling analysis of 28 colonies during a 70-day test period in Baton Rouge, Louisiana (15 May–24 July) [27]. Uniform, broodless colonies were established on 15 May from a large cage of mite-infested bees.

Characteristic	Range of data ^a	$h^2 \pm SE$
Suppression of mite reproduction, June 2 ^b	8–52 %	0.38 \pm 0.58
Suppression of mite reproduction, June 23 ^b	15–48 %	0.06 \pm 0.48
Suppression of mite reproduction, July 24 ^b	9–48 %	0.46 \pm 0.59
Hygienic behavior	4–91 %	0.65 \pm 0.61
Physical damage to mites (total)	4–35 %	0.00 \pm 0.45
Dents in body	1–11 %	0.00 \pm 0.45
Broken legs or bodies	0–26 %	0.17 \pm 0.52
Capped period (h)	268–290	0.89 \pm 0.59
Proportion mites in brood	39–82 %	1.24 \pm 0.49
Mites per 100 cells of brood (24 July) ^c	4–30.5	0.28 \pm 0.56
Final mite population ^d	534–3 389	0.17 \pm 0.52
Mites per 1 000 bees (24 July) ^c	26–198	0.01 \pm 0.46

^a The range of data from the 28 colonies in the test. This experiment was not designed to produce colonies that were highly resistant to mites.

^b Suppression of mite reproduction combines the following three components: 1) dead foundress mite in a cell with no progeny, 2) live foundress mite with no progeny, and 3) live foundress mite with progeny produced too late to mature.

^c The number of adult foundress female mites, based on counts of 200 cells of capped brood per colony.

^d The mite population refers to the total number of adult female mites in the brood and on adult bees. Since all colonies started with the same number of mites, this is an expression of the growth of the mite populations (final population/original population).

^e As above, mites refers to all of the adult female mites in the colony (including the foundress females in the brood cells). Bees refers to all adult bees in a colony.

Table II. Three field tests conducted in 1997 and 1998 in Baton Rouge, Louisiana. The colonies within each group started with a population of bees and mites that were collected from a single cage, combs with no brood, and a test queen. Colonies in group 1 had queens that were not selected for resistance, colonies in group 2 had queens selected for resistance that were outcrossed to non-resistant drones, colonies in group 3 had mite-resistant stock.

Experiment	Initial mite population (P_1) ^a	Population change per reproduction cycle (ab)	Population change in cell (a)	Population change outside cell (b)	Duration (in days) of reproduction cycle (t)	Final mite population (P_2)
Group 1						
(70 days) $n = 28$ colonies						
group mean ^b	784	1.22 ± 0.13	2.41 ± 0.22	0.50 ± 0.06	19.2 ± 3.7	1 649 ± 618
best colony ^c	784	0.888	2.181	0.41	18.8	520
worst colony ^c	784	1.452	2.437	0.60	16.2	3 288
Group 2						
(60 days) $n = 25$ colonies						
group mean	748	1.08 ± 0.25	1.86 ± 0.64	0.58 ± 0.20	19.4 ± 5.8	1 203 ± 781
best colony	748	0.718	0.874	0.82	16.1	221
worst colony ^c	748	1.430	2.779	0.51	15.0	3 044
Group 3						
(111 days) $n = 23$ colonies						
group mean	527	0.83 ± 0.16	1.17 ± 0.47	0.80 ± 0.30	23.1 ± 6.3	273 ± 214
best colony	527	0.422	1.171	0.36	37.9	43
worst colony	527	1.057	2.323	0.46	20.5	710

^a Groups 2 and 3 began 23 days earlier and thus had one brood cycle prior to the beginning of the test period. This was because both of these groups had been selected for suppression of mite reproduction and this characteristic (as we measured it) was not expressed in the brood from the first brood cycle. Details are in the text.

^b Group means ± standard deviation.

^c Best and worst report the equations for the colonies that had the most and fewest mites at the end of the test. It does not necessarily reflect the lowest *a*, *b* or *ab*.

(t), which is converted to n , the number of reproductive cycles during the evaluation period. All three factors are colony averages. Therefore, ab (a times b) is the average population change per reproductive cycle, and multiplying $(ab)^n$ times the initial population (P_1) equals the final mite population (P_2). If any four of the five values are known, the remaining one can be calculated.

The product ab is the average rate of population growth per reproductive cycle for a short test. During our short field tests (10–15 wk) in which colonies begin with relatively low mite populations, the growth of the mite population probably follows the classic exponential growth curve, $P_2 = P_1 e^{rt}$. In this equation r is the intrinsic rate of increase per unit time and t is the duration of the growth period. Exponential growth of *V. jacobsoni* populations occurs only for short periods of time and before the population reaches a carrying capacity. If r is defined as ‘per reproductive cycle’ and $t = n$ (which is the number of cycles in a test), then $r = \ln(ab)$.

The third level of evaluation describes a specific characteristic that is measurable and heritable. We concluded [27] that this is the level at which one should select for resistance to *V. jacobsoni*. Each specific characteristic (described below) affects one of the three components described above (a , b or t). When specific characteristics affect the same component (for example suppression of mite reproduction and hygienic behavior both affect a), their combined effect may not be additive and may not be beneficial. However, when specific characteristics affect different components, they would probably combine to produce a colony with increased resistance to the mite.

2.3. Specific characteristics of resistance to *V. jacobsoni*

2.3.1. Duration of capped period

A brood cell is normally capped ≈ 8 days after a fertilized egg is laid, and it remains

capped for ≈ 12 days until the worker bee emerges as an adult. However, among a group of colonies, there is significant variation in the average duration of the capped period, and this length of time is a heritable characteristic [2, 24, 27, 42–44]. Biensfeld [2] also showed that the duration of the precapping period was affected by the genotype of the nurse bees. However, the duration of the capped period was affected by the genotype of the adult bees in the colony during the capped period but not by the genotype of the bees that had nursed them as larvae.

Büchler and Drescher [6] found a positive relationship between the duration of the capped period and the mite population in field colonies. Their data showed that about 25 % of the variation in mite populations in their colonies could be explained by differences in the duration of the capped brood.

2.3.2. Suppression of mite reproduction

Mites that do not reproduce in the brood cell are found in nearly every colony. However, the frequency of non-reproducing mites in European honey bees is normally below 40 % [9, 14, 16, 21, 32–34, 36, 37, 40, 47]. We define non-reproducing mites as mites that enter the cell to reproduce but 1) produce no progeny, 2) produce males only, 3) produce progeny too late to mature, or 4) die in the cell before they can reproduce. Because non-reproduction of mites was found to be a heritable characteristic of bees [27], we call it suppression of mite reproduction. This characteristic is directly related to a and has been linked to resistance to *V. jacobsoni* by many investigators [1, 9, 15–17, 23, 38, 52, 53, 58, 62].

Data suggest that there may be two components that suppress mite reproduction. We (*table 1*) evaluated both the immediate brood effects of this characteristic reported by Camazine [9] and a delayed expression of this characteristic. In the delayed expression, suppression of mite reproduction was not evident when mites went through their

first reproductive cycle in colonies that would ultimately suppress mite reproduction about 2 months later. Fuchs [19] attributed this delayed effect to attributes that the mite attained before it entered a brood cell. Based on sibling analysis in *table I*, mite reproduction was heritable during our first observation. Therefore, larvae and/or pupae suppressed mite reproduction and this immediate effect, described by Camazine, was a heritable characteristic of bees. Mite reproduction had low heritability during the transition period (early July measurement in *table I*) but was again heritable at the third measurement. This third measurement was the delayed effect described by Fuchs [19] and the characteristic that we used as the basis of our selection (*table II*).

2.3.3. Entrapped mites

We recently observed a higher frequency of foundress mites that were dead between the cocoon and the cell wall. When these were observed in cells with spinning bee larvae, we noticed that the entrapped mites were often alive. In most cases, the mites are found at the bottom of the cell, ventral side up, the same posture that they take while they are inactive in the brood food. In a 1997 field test, 27 % of the foundress mites in the brood cells were entrapped [30]. However, we do not know if this characteristic is heritable. Martin [37] reports that about 1 % of the foundress mites become entrapped, and we found similar levels of entrapment (0.4 % or 11/2 930 observations of foundress mites) in unselected colonies of bees in *table II*, group 1.

This characteristic is more effective in reducing mite populations than suppression of mite reproduction because a non-reproducing mite that survives has an *ab* of 1, while an entrapped mite has an *ab* of zero.

2.3.4. Hygienic behavior

Hygienic behavior is the rate at which adult bees remove dead or diseased brood

from capped cells [56]. Spivak [60] has shown that this characteristic is related to resistance to *V. jacobsoni*, and *table I* [27] shows that hygienic behavior (when measured with freeze-killed brood) is heritable ($h^2 = 0.65$). Hygienic behavior is an important characteristic for mite resistance in *Apis cerana* [50, 54] and resistance to other diseases of bees such as chalkbrood [22, 61] and American foulbrood [56, 57].

2.3.5. Grooming behavior

Physical damage to mites may be caused by the activities of adult bees [59], and this grooming behavior is a heritable characteristic of bees ($h^2 = 0.71$) [41]. Caution should be used in deciding what damage to mites is actually caused by grooming bees. For example, we found that dents in the dorsal surface of the idiosoma often occurred while young adult mites were still in their brood cells [27], and not by the mandibles of grooming bees. Although self grooming and nestmate grooming are important mechanisms of resistance to *V. jacobsoni* for *Apis cerana* F., it is a less important component of resistance for our western honey bee *Apis mellifera* L. [3, 4, 7, 13, 18, 48, 49].

2.3.6. Proportion of mites in brood

This is a measure of the duration of the reproductive cycle of the mite [25, 46], which is explained in more detail in section 2.4.1. Mites are either on adult bees (in a phoretic stage) or in the brood cells (in a potentially reproductive stage).

The proportion of mites in brood was found to be a highly heritable characteristic in bees ($h^2 = 1.24$) [27]. This is sometimes called invasion of brood cells [5, 20, 39]. Perhaps mites with a low proportion of mites in brood and therefore those with longer reproductive cycles are less attracted to bee larvae and therefore are slower to enter brood cells. This may be best explained by the work of Trouiller et al. [63] who describe a chemical signal produced by bee larvae

that attracts *V. jacobsoni* to the brood. Because of a direct effect on t , this characteristic may serve well in combination with characteristics that affect a such as suppression of mite reproduction or hygienic behavior.

2.4. Describing the reproduction of mites

The most basic calculation is the overall growth of the mite population in each colony. This is simply the final mite population divided by the initial mite population (P_2/P_1). By rearranging the equation ($P_1(ab)^n = P_2$), the overall growth of the mite population (P_2/P_1) equals $(ab)^n$. The next step is to solve for ab by calculating n .

2.4.1. Measuring n

Before we calculated the number of reproductive cycles (n), we needed to calculate the duration of the reproductive cycle of the mites in each colony. The duration of a reproductive cycle is the average time from when a mite enters a cell until it enters another cell (time spent in the cell plus time spent on adult bees). Not all mites will have the same time for their reproductive cycle and the length may change with the age of the mite. However, only the group average is important.

The duration of the reproductive cycle of the mite is almost entirely related to the variability of the time that the mites spend outside the brood cell. There is significant variation in the duration of the capped period of bee brood [2, 24, 42] but this variation is only slightly greater than ± 1 day in European honey bees. Mites enter the cell about 1/2 day before the cell is capped and remain until the capping is removed by the teneral bee in the cell. Therefore, the reproductive stage of the mite is relatively constant at $\approx 12.5 \pm 1$ days or 1/2 day longer than the capped period. In contrast, the time spent outside the cells can vary by more than a week (summarized by Fries et al. [17]). For

example, if 65 % of the mite population is in the brood (typical), then the average duration of the mite reproductive cycle in that colony is $12.5/0.65$ or 19 days. If 40 % are in brood cells, the reproductive cycle extends to 31 days. A longer reproductive cycle slows the growth of mite populations.

We estimated the length of the reproductive cycle of mites (t) for each colony by comparing the number of mites on adult bees with the number of mites in brood cells (both were measured at the end of the test period). The number of reproductive cycles in the test was then calculated from the estimate of the length of the reproductive cycle for mites in a colony (divide the duration of the experiment by the length of the reproductive cycle). However, most of our tests were initiated in bee colonies with no brood. Since mites cannot begin their reproductive cycle until bees have capped brood, the mites in a colony that begins with no brood were each assigned 23 days for their first reproductive cycle (7 + 12.5 + 3.5, which was 7 broodless days + 12.5 days in brood + half of a normal phoretic period).

The worst colony in *table II* (group 1) is used as an example of calculating n . The test ran for 70 days and based on proportion mites in brood, the colony had a reproductive cycle (t) equal to 16.2 days. Therefore, $n = 1 + (70 - 23)/16.2 = 3.9$ reproductive cycles.

2.4.2. Calculating ab

As shown in *figure 1*, ab is the population growth of mites per reproductive cycle. An ab of 1 equals no change in the mite population; numbers > 1 indicate population increase and < 1 indicate population decline.

For calculating ab the equation becomes: $ab = \sqrt[n]{(P_2/P_1)}$. Again using the worst colony in *table II*, group 1, as the example where n was calculated as 3.9: $ab = \sqrt[3.9]{4.19} = 1.45$. These data and the calculations can be handled on nearly any spreadsheet program. However, when using a computer it is simpler to write $\sqrt[3.9]{(P_2/P_1)}$ as $(P_2/P_1)^{(1/3.9)}$.

However, we know that *ab* is not always constant. Fuchs [19] showed that the major control of mite reproduction depended on events that occurred before the mite enters the cell. Harris and Harbo [27] have shown that colonies that suppress mite reproduction at a high rate may not express that characteristic in their first brood cycle when they have started with package bees. Thus, we not only assign the first brood cycle a standard length, we assign a standard *ab* of 1.22 (the group average for unselected bees, group 1 in *table II*) when evaluating colonies for suppression of mite reproduction.

2.4.3. Calculating *a* and *b*

When *ab* is already estimated, only one of the factors needs to be measured and the other can be calculated. We suggest measuring *a*, the rate of mite increase from the cell. We define *a* as the sum of mature daughters and surviving mothers that exit the cells divided by the number of foundress females that entered those cells. Our *a* differs from the rate of reproduction, which is the total number of progeny produced per mother mite [15, 31].

For *table II*, we calculated *a* by observing mites in brood cells. We examined mite-infested cells when the bees were at the tan body stage (about 16–17 days after the bee egg was laid). In each cell we counted the number of foundresses and the number of female progeny that were deutonymphs or adults. We allowed each adult daughter and immobile deutonymph to have a 100 % chance of reaching maturity and emerging from the cell, while each mobile deutonymph only had a 39 % chance of doing so [21]. Therefore, our calculation of *a* was (the number of live foundress females + the number of female progeny that were immobile deutonymphs or adults + [0.39 * the number of mobile female deutonymphs]) / (the number of live and dead foundress females). Harbo [25] estimated *a* by three different methods, and others have calculated *a* by various techniques [17, 38].

There are also various ways to report *a*. Most report the number of female progeny that mature in a cell. Some do not include cells with mites that produce no progeny or cells with multiple foundresses. We include the contents of all infested cells, even those with dead foundresses. As described in the paragraph above, our *a* included the surviving foundress. Since 98 % of the foundress mites usually survive the reproductive phase, our estimates of *a* are about 1 greater than the reproductive rate reported by many others.

An example of measuring *a* and then separating *a* and *b* from *ab* is again taken from the worst colony in *table II*, group 1, where *ab* had been calculated as 1.45. We counted 20 mobile female deutonymphs, 38 immobile female deutonymphs, 28 adult daughters, 48 live foundresses and two dead foundresses. Therefore, 121.8/50 equals an *a* of 2.44. Thus $b = ab/a$ or $1.45/2.44 = 0.60$.

What we call *b* is not strictly survivorship outside the cell. It also includes mites that transfer into or out of the colony. When mite populations are relatively uniform, perhaps one can assume that immigration and emigration are equal. However, when adjacent colonies have highly variable populations of mites, drifting or robbing bees will probably tend to move mites into the colonies with fewer mites. This is a good argument for evaluating colonies that begin with uniform populations of bees and mites and for ending a field test as soon as possible.

3. CONCLUSIONS

It is certainly possible to breed bees that are resistant to *V. jacobsoni*. *Table II* (group 3) is an example of a group of 23 colonies that were selected for their ability to suppress the reproduction of mites in brood cells. Those colonies (as a group) averaged fewer mites at the end of the test than at the beginning [30]. However, breeding for resistance to *V. jacobsoni* has proven to be more dif-

ficult than most other selection programs with bees, so it requires more precise measurements and a more systematic approach if one demands quick results.

There are presently many mechanisms of resistance that show promise. We think that the suppression of mite reproduction, hygienic behavior and proportion of mites in brood (the tendency for adult mites to not enter brood cells) are presently the three most promising of the specific characteristics that are heritable and should respond to selection.

It is important to note that there are locations around the world where honey bees survive without a program to control mites. Brazil is probably the best example [45]. Moreover, the levels of *V. jacobsoni* infestation in both European and Africanized bees in Brazil have further declined with time [12]. This suggests that when a population of bees is able to coexist with *V. jacobsoni*, natural selection will work on mites to make them less virulent, or selection will work on bees to make them more resistant. Both possibilities are likely. However, data suggest that mite populations in Brazil are less virulent than in most other parts of the world and that this virulence may be decreasing. Perhaps we can use this to our advantage. If we select bees to a point where they can survive reasonably well in the presence of mites, mites may respond by becoming less virulent.

Our strategy is to select bees that will retain an acceptable level of resistance when outcrossed to drones from non-resistant colonies. This would provide a broader genetic base for resistant bees, would preserve the genetic diversity of our honey bee populations, and would enable natural selection to operate more effectively. In this situation, a bee population would retain most of its genetic diversity as it gradually incorporates genes for resistance to mites.

Table II (group 2) is an example of queens from a mite-resistant stock that were out-

crossed. The mite-resistant queens, selected for suppression of mite reproduction, were instrumentally inseminated with drones collected from colonies that had not been selected for resistance to mites. Some colonies were very good and expressed a high level of resistance, but others were highly susceptible. The group was variable. If a second specific characteristic of resistance was incorporated into this stock (such as a low proportion of mites in brood), a group of colonies with outcrossed queens may be more uniform and may be able to hold mite growth to an *ab* of 1.0 or less.

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Résumé – La sélection d'abeilles mellifères (*Apis mellifera* L.) résistantes à *V. jacobsoni* Oud. Cet article examine les méthodes de sélection d'abeilles mellifères résistantes à l'acarien *V. jacobsoni*. Un plan de sélection est décrit ci-après, qui devrait être mis en œuvre dans tout programme de sélection prenant en compte les caractères des abeilles mesurés au niveau de la colonie.

- 1) Développer des méthodes pour mesurer les populations d'abeilles et d'acariens et pour mesurer les caractères associés à la résistance.
- 2) Identifier les caractères spécifiques en relation avec la croissance des populations d'acariens.
- 3) Déterminer si ces caractères sont héréditaires.
- 4) Renforcer ces caractères par un élevage sélectif.
- 5) Regrouper les composantes de la résistance dans des abeilles productives et résistantes aux acariens.

Dans cet article on entend par résistance à *V. jacobsoni* la sélection chez les abeilles de caractères qui vont retarder la croissance de la population d'acariens établie dans la colonie. Lors du criblage initial des caractères, nous avons utilisé des reines fécondées par un seul mâle. Nous pensons que cela a permis de détecter aussi des caractères peu fréquents. Les caractères au niveau de la colonie auraient pu être cachés si les reines avaient été fécondées par plusieurs mâles. Notre approche décrit la croissance de la population d'acariens au sein d'une colonie d'abeilles par l'équation : $P_1(ab)^n = P_2$, où P_1 et P_2 sont les populations initiale et finale d'acariens, a la variation de la population tant que les acariens sont dans les cellules de couvain, b la variation quand ils sont hors des cellules de couvain et n le nombre de cycles reproducteurs de l'acarien. L'équation est valable pour une colonie qui a un apport régulier en couvain d'ouvrières de façon à permettre la reproduction de l'acarien. Seules les femelles adultes d'acariens sont comptées et toutes les mesures sont des moyennes pour la colonie. En comparant les taux de croissance des populations d'acariens de chaque colonie (P_2 / P_1), on peut déterminer la croissance réelle dans chaque colonie. Les valeurs a , b et n peuvent alors expliquer comment la population d'acariens est passée de P_1 à P_2 en termes de composantes mesurables du cycle reproducteur de l'acarien. Les variations de la population d'acariens doivent se refléter dans une ou plusieurs de ces composantes. Les caractères spécifiques des abeilles (qui sont la base de la sélection) peuvent alors être associés à l'une de ces trois composantes. Quand plusieurs caractères spécifiques agissent sur une même composante (par exemple, la suppression de la reproduction de l'acarien et le comportement hygiénique), leur effet combiné n'est pas nécessairement additif ni bénéfique. Si en revanche un même caractère spécifique agit en même temps sur différentes composantes, ceux-ci se combineraient probable-

ment pour donner une colonie ayant une résistance accrue à *V. jacobsoni*.

Jusqu'ici notre sélection s'est concentrée sur un caractère, la suppression de la reproduction de l'acarien, qui provoque la non reproduction des acariens qui pénètrent dans les cellules de couvain. Sur la base de ce seul caractère héréditaire, nous avons produit des colonies dans lesquelles la population d'acariens décline durant une période test de 70 j. Deux autres caractères héréditaires, le comportement hygiénique et la proportion d'acariens sur le couvain, peuvent avoir aussi une importance pour la sélection d'abeilles résistantes. © Inra/DIB/AGIB/Elsevier, Paris

Apis mellifera / *Varroa jacobsoni* / population résistante / élevage sélectif

Zusammenfassung – Selektion von Honigbienen auf Resistenz gegen *Varroa jacobsoni*. Dieser Beitrag befaßt sich mit Methoden zur Selektion von Honigbienen (*Apis mellifera*) auf Resistenz gegen den Befall durch *Varroa jacobsoni* Oudemans. In der Folge wird ein Zuchtplan beschrieben, der innerhalb jedes auf der Messung von Volkseigenschaften fußenden Selektionsprogramms wirkungsvoll angewendet werden kann.

- 1) Entwicklung von Methoden zur Erfassung der Populationen von Bienen und Milben und zur Messung der mit Resistenz verbundenen Eigenschaften.
- 2) Identifizierung spezifischer mit dem Anwachsen der Varroapopulationen in Beziehung stehender Eigenschaften.
- 3) Ermittlung, ob diese Eigenschaften vererbbar sind.
- 4) Verstärkung dieser Eigenschaften durch selektive Zucht.
- 5) Vereinigung der Resistenzkomponenten in produktiven, milbenresistenten Bienenlinien.

In diesem Bericht wird Varroaresistenzzucht als Selektion der Honigbienen auf Eigenschaften verstanden, die das Anwachsen

von Varroapopulationen in den Völkern verlangsamen. In den anfänglichen Untersuchungen zum Auffinden von Resistenzeigenschaften benutzten wir Königinnen, die von nur einem Drohn besamt waren. Wir gehen davon aus, daß hierdurch auch weniger häufige Eigenschaften ermittelt werden können. Diese Volkseigenschaften könnten verdeckt werden, wenn die Königinnen mit mehreren Drohnen verpaart sind. Unser Ansatz beschreibt das Wachstum von Varroapopulationen in den Bienenvölkern unter Verwendung der Formel $P_1 (ab)^n = P_2$. Hierbei sind P_1 und P_2 die Milbenpopulation zu Anfang und am Ende, a die Änderung der Population solange die Milben in den Brutzellen sind, b die Änderung außerhalb der Brutzellen und n die Anzahl von Reproduktionszyklen der Milben. Diese Gleichung beschreibt das Anwachsen einer Milbenpopulation mit einem konstanten Angebot an Brutzellen zur Vermehrung. Es werden nur die erwachsenen Milben gezählt, und alle Messungen sind Mittelwerte für die Völker. Durch Ermittlung des tatsächlichen Anwachsens der Milbenpopulation in jedem Volk kann die Zuwachsrates der Milbenpopulationen (P_2/P_1) ermittelt werden. Die Werte von a , b , und n können dann erklären, auf welche Weise die Milbenpopulation sich in Bezug auf meßbare Komponenten des Reproduktionszyklus der Varroamilben von P_1 nach P_2 entwickelt hat. Änderungen der Varroapopulation müssen auf eine oder mehrere dieser Komponenten zurückzuführen sein. Spezifische Eigenschaften der Bienen (die Grundlage der Selektion) können dann mit einer dieser drei Komponenten in Verbindung gebracht werden. Falls mehrere Eigenschaften eine dieser Komponenten beeinflussen (beispielsweise wirken sich eine Unterdrückung der Milbenreproduktion und das hygienische Verhalten beide auf a aus) muß sich ihr kombinierter Effekt nicht notwendigerweise additiv oder günstig verhalten. Falls dagegen eine einzelne Eigenschaft verschiedene dieser Komponenten gleichzeitig beeinflußt, würden diese kombiniert mit hoher Wahrscheinlichkeit

zu erhöhter Resistenz gegenüber *Varroa jacobsoni* führen.

Zur Zeit hat sich unsere Selektion auf eine einzelne Eigenschaft konzentriert, die Unterdrückung der Vermehrung der Milben. Dies führt dazu, daß Milben nicht reproduzieren, nachdem sie eine Brutzelle befallen haben. Auf Grundlage dieser einzelnen erblichen Eigenschaft haben wir Völker erzeugt, in denen die Milbenpopulation über einen Zeitraum von 70 Tagen abnimmt. Zwei weitere Eigenschaften, das hygienische Verhalten sowie der Anteil der in der Brut befindlichen Milbenpopulation könnten ebenfalls wichtig sein, um Honigbienen auf Resistenz gegen Varroamilben zu selektieren. © Inra/DIB/AGIB/Elsevier, Paris

Apis mellifera / Milben / *Varroa jacobsoni* / Populationen / Selektionszucht

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