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An algorithm for detecting *Trichodesmium* surface blooms in the South Western Tropical Pacific

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1
2 **Abstract**
3

4 *Trichodesmium*, a major colonial cyanobacterial nitrogen fixer, forms large blooms in NO₃-
5 depleted tropical oceans and enhances CO₂ sequestration by the ocean due to its ability to fix
6 dissolved dinitrogen. Thus, its importance in C and N cycles requires better estimates of its
7 distribution at basin to global scales. However, existing algorithms to detect them from
8 satellite have not yet been successful in the South Western Tropical Pacific (SP). Here, a
9 novel algorithm (TRICHODESMIUM SATellite) based on radiance anomaly spectra (RAS)
10 observed in SeaWiFS imagery, is used to detect *Trichodesmium* during the austral
11 summertime in the SP (5°S-25°S 160°E-170°W). Selected pixels are characterized by a
12 restricted range of parameters quantifying RAS spectra (e.g. slope, intercept, curvature). The
13 fraction of valid (non-cloudy) pixels identified as *Trichodesmium* surface blooms in the region
14 is low (between 0.01 and 0.2 %), but is about 100 times higher than deduced from previous
15 algorithms. At daily scale and in the SP, this fraction represents a total surface oceanic area
16 varying from 16 to 48 km² in Winter and from 200 to 1000 km² in Summer (and at monthly
17 scale, from 500 to 1000 km² in Winter and from 3100 to 10890 km² in Summer with a
18 maximum of 26432 km² in January 1999. The daily distribution of *Trichodesmium* surface
19 accumulations in the SP detected by TRICHOSAT is presented for the period 1998-2010
20 which demonstrates that the number of selected pixels peaks in November-February each
21 year, consistent with field observations. This approach was validated with in situ observations
22 of *Trichodesmium* surface accumulations in the Melanesian archipelago around New
23 Caledonia, Vanuatu and Fiji Islands for the same period.

24
25 **Keywords:** *Trichodesmium*, surface blooms, radiance anomaly spectra, algorithm, SeaWiFS,
26 ocean color, New Caledonia, Vanuatu, Fiji Islands, Tropical Pacific Ocean, Melanesian
27 archipelago.
28
29

1

2 **1. INTRODUCTION**

3 The balance between oceanic N₂ fixation and nitrogen losses (denitrification) in the ocean has
4 been postulated to regulate atmospheric CO₂ over geological time via the enhancement of
5 biological sequestration of CO₂ (Falkowski, 1997; Gruber and Sarmiento, 1997; Deutsch et
6 al., 2007; Capone and Knapp, 2007). Unicellular (Zehr et al., 2001; Montoya et al., 2004;
7 Church et al., 2008; 2009; Zehr et al., 2011) and filamentous cyanobacteria (Carpenter, 1983;
8 Capone et al., 1997; Capone et al., 2005; LaRoche and Breitbarth, 2005; Bonnet et al., 2009;
9 Moisander et al., 2010) incorporate this form of “new” nitrogen (N) into the marine food web
10 of tropical and subtropical oceans (Berman-Frank et al., 2004; Mahaffey et al., 2005;
11 Mulholland, 2007). N₂ fixation is considered to be the major source of new N in stratified,
12 oligotrophic tropical oceans (Capone et al., 1997; Karl et al., 2002). Future change in sea
13 surface temperature (Breitbarth et al., 2006) or/and CO₂ concentration are expected to
14 stimulate photosynthesis (C fixation) and N₂ fixation by filamentous cyanobacteria,
15 particularly by *Trichodesmium* spp. (Barcelos et al., 2007; Hutchins et al., 2007; Kranz et al.,
16 2009; Levitan et al., 2010). This enhancement of *Trichodesmium* growth could compensate
17 the decreased growth of other phytoplankton owing to a presumed decrease of nitrate supply.

18 *Trichodesmium* spp. can form extensive blooms which have been observed for a long time
19 in the South Western Pacific Ocean (SP), particularly in austral summer (Dandonneau and
20 Gohin, 1984). The presence of three major archipelagos (New Caledonia, Vanuatu and Fiji-
21 Tonga in the SP region ([5°S- 25°S, 150°E- 170°W], Figure 1a) and their potential for oceanic
22 iron enrichment from land may trigger these cyanobacterial blooms (Bowman and Lancaster,
23 1965; Mantas et al., 2011) as *Trichodesmium* blooms require large quantities of iron (Rubin et
24 al., 2011). The blooms appear as brown or orange meandering patterns around those
25 archipelagos (Dupouy et al., 1988; Dupouy, 1990; Tenório, 2006; Hashihama et al., 2010), are
26 clearly detected from the International Space Shuttle (December 2001 around Tonga Islands),
27 and were recently highlighted by the NASA Ocean Color Website (Feldman et al, 2010).
28 Blooms are also regularly observed in waters of the Dampier Archipelago, the Arafura Sea
29 (Neveux et al., 2006) and off the Great Barrier Reef (Kuchler and Jupp, 1988; Furnas, 1989;
30 Bell et al., 1999). *Trichodesmium* was reported in the Western North Pacific (Shiozaki et al.,
31 2009; Kitajima et al., 2009; Konno et al., 2010). Nevertheless, observations in the SP
32 contradict the recently published global map of *Trichodesmium* analogs based on ecosystem

1 model results that indicate a predominance of higher densities in the North Pacific than in the
2 South Western Pacific Ocean (Monteiro et al., 2010).

3 Estimating the occurrence of *Trichodesmium* surface blooms from satellite is a major
4 challenge, but will be required for large-scale estimates of nitrogen fixation (e.g., Westberry et
5 al., 2005; Westberry and Siegel, 2006). Regional algorithms have been successfully applied
6 on the Atlantic continental shelf (Subramaniam et al., 2002), off Canary Islands (Ramos et al.,
7 2005) as well as the Indian coast (Sarangi et al., 2004). Global algorithms have also been
8 recently developed for estimating phytoplankton community structure in the surface oceans.
9 PHYtoplankton SATellite (PHYSAT) algorithm was successful in identifying *Synechococcus*-
10 like cyanobacteria with maxima in the tropics, and found that they are particularly abundant in
11 the tropical ocean (Alvain et al., 2005). The Scanning Imaging Absorption Spectrometer for
12 Atmospheric Cartography (SCHIAMACHY) sensor also detects a cyanobacterial signal
13 within the same latitudinal band (Bracher et al., 2008). However, all algorithms generally fail
14 at identifying seasonality of *Trichodesmium* blooms in the SP and none retrieve the relatively
15 high abundance of *Trichodesmium* surface blooms expected and observed in situ during
16 austral summer (November to March). Here we develop an algorithm to detect
17 *Trichodesmium* surface blooms in the SP which is based on SeaWiFS radiance anomalies
18 (similar to PHYSAT) and apply it to SeaWiFS data from 1997-2010.

20 **2. MATERIAL AND METHODS**

21 **2.1. DATA**

22 **2.1.1. In situ observations**

23 Bloom observations (Figure 2) included these done during the maritime survey of the
24 Economic Exclusive Zone of New Caledonia by the French Navy (aerial and shipboard
25 observations) and these obtained during scientific cruises on the R/V *Alis* between New
26 Caledonia, Vanuatu, Fiji and Wallis and Futuna Islands (15°S to 23°S, 160°E to 180°E;
27 Dupouy et al., 2004a). At most of the observation by the French Navy and the R/V *Alis*,
28 surface water samples was collected with bucket and preserved on board in a 4% formalin
29 solution. Then, identification of diazotroph morphological groups was made under a Zeiss
30 microscope at IRD Nouméa (LOCEAN laboratory). Furthermore, between October 2001 and
31 October 2003 (nine Diapalis cruises as part of the DIAzotrophy in a Pacific ZONE program),
32 more detailed identification and abundance distribution of filamentous diazotrophs was

1 obtained from inverted microscopy (Tenório, 2006). In this case, sampling was done with 8 L
2 Niskin bottles (sometimes with bucket sampling for comparison). During the Diapalis cruises,
3 chlorophyll and phycoerythrin measurements were obtained from spectrofluorometry
4 (Lantoiné and Neveux, 1997; Neveux et al., 1999; Neveux et al., 2006). A slightly higher
5 concentration of chlorophyll a (Chl *a*) was observed from bucket samples compared to those
6 from Niskin samples.

7

8 **2.1.2 Satellite ocean color data**

9 For the development of the new algorithm, representative global SeaWiFS Level3 data
10 (R2009) were selected at summer and winter seasons, and included normalized water leaving
11 radiance, nLw (λ), at 6 channels (412, 443, 490, 510, 555, and 670 nm) as well as SeaWiFS
12 chlorophyll and the diffuse attenuation coefficient at 490 nm (K490 product). From these
13 data, a Look Up Table (LUT) relating K490 to remote sensing reflectance was created (see
14 Section 2.2). However, to avoid compositing artifacts upon application of the LUT, daily
15 SeaWiFS Level-2 GAC (R2009) between 1998-2010 and covering the Western Pacific Ocean
16 (160°E-160°W/25°N-25°S area), were used.

17

18 **2-2. METHODS**

19 The general approach was to define a spectral radiance anomaly from SeaWiFS nLw(λ) that
20 was specifically related to *Trichodesmium* surface blooms. It aimed at removing the first order
21 variability in ocean color caused by chlorophyll concentration while preserving the variability
22 that may be specifically caused by individual phytoplankton species or other optically active
23 components. This objective is similar to the PHYSAT classification method (Alvain et al.,
24 2005) that was initially developed for discrimination of major phytoplankton groups in Case 1
25 waters. Waters dominated by diatoms, *Prochlorococcus*, *Synechococcus*-like cyanobacteria or
26 haptophytes could thus be classified according to their radiance anomaly spectra (RAS):

27

$$28 \quad nL^*w(\lambda) = nLw(\lambda) / \langle nLw(\lambda) \rangle \quad (1)$$

29

30 where $\langle nLw(\lambda) \rangle$ is the expectation of nLw at a given chlorophyll concentration, computed as
31 the average of a large global SeaWiFS dataset, and nL*w is the radiance anomaly relative to
32 this average. The main advantage of PHYSAT is to provide thresholds allowing

1 characterization of RAS at each pixel. PHYSAT uses LUT of $\langle nLw(\lambda) \rangle$ as a function of
2 chlorophyll.

3 Here, we employed a LUT based on the diffuse attenuation coefficient at 490 nm
4 (SeaWiFS “K490”). K490 has the advantage over chlorophyll of being computed
5 straightforwardly while operational SeaWiFS chlorophyll estimates result from switching
6 between three wavelength ratios. This new LUT was built using four daily, global Level-3
7 images from February 15, May 15, August 15 and November 15 2002, in order to encompass
8 a seasonal cycle. The LUT contains the likelihood of SeaWiFS radiances, noted $\langle nLw(\lambda)_{K490} \rangle$
9 \rangle , for all K490 values in the 0.0186 m^{-1} to 0.2499 m^{-1} interval. Radiance anomaly spectra
10 (RAS) are then computed as

$$11 \quad nL*w_{K490}(\lambda) = nLw(\lambda) / \langle nLw(\lambda)_{K490} \rangle \quad (2)$$

12 where $nLw(\lambda)$ are the SeaWiFS radiance estimates. At the end, we checked that results are
13 strictly equivalent than when Chlor_a is used as in PHYSAT. Radiance anomaly spectra
14 $nL*w_{K490}$, hereafter denoted RAS, can thus be considered as equivalent than PHYSAT
15 nLw^* .

16 There is another significant difference between the approach used here compared with the
17 previous PHYSAT effort. Instead of defining RAS thresholds and associating them with
18 pigment classes (and thus, different phytoplankton groups), the new algorithm defines
19 quantitative shape and magnitude criteria of the RAS itself. Eighteen summer scenes around
20 New Caledonia in 2003 and 2004 were selected as they correspond to periods where slicks
21 and high abundance ($> 5000 \text{ trichomes.L}^{-1}$) of *Trichodesmium* was observed in the surface
22 ocean (Tables 1 and 2). In 2003, SeaWiFS level2-GAC of January 6, 13, 18, February 1, 4, 9,
23 13, 16, 18, 21, 28, March 2, 6; Yeardays: 6, 13, 18, 32, 35, 40, 44, 47, 49, 52, 59, 62, 66)
24 corresponded to slicks observations by the French Navy from December 2002 to February and
25 March 2003 (Table 1). The gap between December 2002 and February 2003 may be due to
26 the January 2003 cyclone “Beni” that prevented sea observations. In 2004, SeaWiFS level 2-
27 GAC of February 10, 18, 23, and March 3, 17; Yeardays 41, 48, 53, 63, 77), corresponded to
28 slicks observations from January to March 2004 (Table 1). The definition of the RAS of
29 *Trichodesmium* surface blooms on the SeaWiFS satellite imagery was generated for pixels
30 around New Caledonia and Fiji from these 18 scenes.

31 The criteria have been chosen in such a way that they permit to rebuild RAS spectra. The
32 RAS was calculated and the “shape” and magnitude criteria of this RAS examined. RAS

1 spectra could be described by the coefficients of a polynomial fit (degree 2) (Figure 3a). We
2 thus retained the following criteria for an objective description of the *Trichodesmium* RAS 1)
3 slope of the linear fit of RAS vs wavelength (S), and 2) the ordinate of the polynomial fit for
4 which the tangent is parallel to linear fit (Yt). Additional criteria related to the shape of the
5 RAS were defined as the major positive and major negative deviations relative to the second
6 degree polynomial fit (largest “bump” or largest “trough”, respectively).

7

8 **3. RESULTS**

9

10 **3.1 FIELD OBSERVATIONS**

11 Aerial and shipboard visual observations of *Trichodesmium* surface blooms (Figure 1b, c)
12 around New Caledonia between 1998 and 2004 are presented in Table 1 and average cruise
13 and transect surface biomass and abundance are presented in Table 2.

14 Generally, slick sizes were a few tens of meters wide and two to three nautical miles long
15 (shipboard or aircrafts photographs, Figure 2). Twice in the series (November 1998,
16 November 1999) large areas of 30 km² were covered by slicks, and once in February 2004,
17 the whole area between New Caledonia and Vanuatu (300 km²) was covered by numerous
18 elongated slicks well detected by longline aircrafts. *Trichodesmium* was responsible of the
19 majority of the slicks observations. Only one formalined sample (12 December 2002)
20 contained small pumices originated from Vanuatu or Tonga volcanoes which were mixed with
21 the *Trichodesmium* colonies. Sea-rafted pumices can drift to the west and reach New
22 Caledonia. Some of these rafts reached the Australian coast in October 2002, approximately
23 7-8 months after a submarine eruption in the Tonga Trench (Bryan et al., 2004). None of the
24 slick observations corresponded to coral spawning which is responsible of fugitive coastal red
25 waters once a year in New Caledonia. The abundance of *Trichodesmium* spp. in the SP
26 exhibited a strong seasonality as previously pointed out (Dupouy et al., 2004a; Moutin et al.,
27 2005) with the highest number of visual observations (65% or 61 over 93 total observations)
28 over the study period (November 1998 to June 2010) occurring between December and
29 February (Table 1). Surface blooms were never detected during winter (only one observation
30 of slicks was available in July 2002, at the west of Fiji Islands, Table 1). Nearly every year,
31 surface blooms developed between New Caledonia and Vanuatu and one of these blooms was
32 exceptionally well tracked (February 2004, Table 1). Slicks were first observed in November
33 2003 near Vanuatu (17.65°S, 167.56°E), in December 2003 at the north and south of New

1 Caledonia, in January (on 26 at 22°S and on 29-30 at 20.33°S/ 166.12°E) then in February
2 2004 (on 1st around New Caledonia and on 12-16 near Vanuatu and again North of New
3 Caledonia at 19.49.9°S, 169. 54°E). The surface bloom persisted after heavy rains and wind-
4 mixing by Category 4 cyclone Ivy on February 24. It was detected near New Caledonia (on
5 February, 26), and under calm meteorological and sea conditions, its slick was sampled on
6 February 28 showing an orange surface scum (around 21°S, New Caledonia) or large flocks
7 of dead colonies (at 18.55°S, 166.05°E, Vanuatu) which disappeared in the evening (17.38°S,
8 166.07°E) (Motevas cruise, Table 2). In March, white senescent *T. erythraeum* (March 1-4, at
9 22°S, 167°E and 20.16°S/168.71°E) and remnant coastal slicks on (March 29) were seen near
10 Vanuatu (17°S, 167°E). In April, remnant slicks were seen nearby Lifou island at the east of
11 the "Grande terre" of New Caledonia (Table 1). Surface slicks were also observed in summer
12 during a water-column observational program in the Loyalty Channel (12 October 2001, 5
13 February 2003 and 2004, see Table 1 and Table 2).

14 Among the pelagic species described in the Pacific region (Revelante and Gilmartin, 1982;
15 Carpenter et al., 1993), a high morphological diversity of filamentous cyanobacteria as
16 described by Lundgren et al. (2005) and Tenório (2006) was observed in our surface samples
17 (Figure 1c). Colonies were essentially composed of small rafts from 10 to 50 filaments, small
18 in length (noted *T. erythraeum*), or long and twisted rafts (noted *T. thiebautii*) of 50-100
19 filaments. Long and curved filaments composed of cells larger than long, and some thinner
20 filaments with cells 5-6 fold longer than wide were noted *K. pelagica* and *T. tenue*,
21 respectively. The distribution of these four morphotypes is shown in Figure 1c. There is no
22 evidence that the same *Trichodesmium* genus or species is widespread all over the tropics
23 except for the *Katagnymene* form (Lundgren et al., 2005). We tended to find *T. erythraeum*
24 near the New Caledonia mainland but elsewhere most of the morphotypes were mixed.
25 During the peak observed *Trichodesmium* abundance in 2004, and along a transect from New
26 Caledonia (20-21°S) to Vanuatu (19°S -17°S), short tufts of *T. erythraeum* were observed at
27 the beginning of the transect (NC) and a mix of *T. thiebautii* (50%), *T. tenue* (25%),
28 *erythraeum* (15%), and *Katagnymene pelagica* and *spiralis* (10%) dominated populations at
29 the end (Vanuatu). This population change was also seen in fluorescence excitation spectra of
30 phycoerythrin (change in the phycourobilin/phycoerythrobilin ratio, as in Neveux et al.,
31 2006). Mixed assemblages of *T. erythraeum*, *T. thiebautii*, and *T. contortum* filaments were
32 also observed on the 2nd and 9th of February 2005 around Fiji Islands (Hashihama et al.,
33 2010), while floating puffs and tufts of mixed taxonomy were observed in March 2007

1 (Biegala, pers. com.). Only *K. pelagica* + *T. tenue* colonies were found isolated near the Niue
2 Island at 19° 2' S 169° 52' W (Table 1, Figure 1c).

3 Surface *Trichodesmium* abundance and associated biomass (Chl *a* > 10 µm) in the Loyalty
4 Channel and along transects between New Caledonia, Vanuatu or Wallis and Futuna allowed
5 to determine a seasonal variability (Table 2). Low filament densities characterized the end of
6 the bloom season in April 1998 and *Trichodesmium* contribution to biomass was reduced
7 (Dupouy *et al.*, 2000; Campbell *et al.*, 2006), as in April 2002 and 2003 (Table 2). High
8 abundance was never observed in May, except in 2002 along the transect 1, where
9 phycoerythrin spectral characteristics confirmed the dominance of *Trichodesmium* between
10 New Caledonia and Vanuatu. At the beginning of the summer season (October), densities
11 were high in 2001 but nul in 2003. *Trichodesmium* abundance was always higher in summer,
12 except during two 2001-2002 Diapalis cruises (December, January). During Motevas in
13 February 2004 (Dupouy *et al.*, 2004b; Dupouy *et al.*, 2008), the beam attenuation was high
14 (0.1 m^{-1}), and a maximal *Trichodesmium* contribution to Chl *a* of 50-70% was observed with
15 a high density of 5000 filaments.L⁻¹ and maximal biomass of 1.9 and 14 mg.m⁻³ for Chl_ *a* and
16 phycoerythrin, respectively. Densities were similar to those measured during February 2003
17 during Diapalis 07 (Tenório, 2006; Neveux *et al.*, 2006; Masotti *et al.*, 2007). The
18 determination of biomass in surface slicks was difficult and will need a specific thin-layer
19 surface sampler. Nevertheless, Chl *a* concentrations up to 2 µg L⁻¹ in the ocean and 3 mg L⁻¹
20 in the New Caledonia lagoon were observed (Tenório, 2006). In winter, *Trichodesmium*
21 abundance was always low and relatively high chlorophyll values were associated with small
22 size algae (low values of the Chl_ *a* fraction above 10 µm).

24 3.2 DETECTION OF TRICHODESMIUM IN SATELLITE DATA

25 In summer 2003 and 2004, the majority of pixels for the region around New Caledonia
26 exhibited the same RAS. These RAS characteristics were examined under the assumption that
27 instances where Chlor_ *a* >0.2 mg.m⁻³ were dominated by *Trichodesmium* while those with
28 <0.2 mg.m⁻³ were not, similar to in situ observations. For Chl_ *a* >0.2 mg.m⁻³, linear slopes of
29 the RAS varied between -0.005 and 0.008, and the tangent ordinate Yt varied within a range
30 of 0.8 to 1.4. For Chl_ *a* < 0.2 mg.m-3, the RAS slope varied within -0.005 to 0.012, while Yt
31 also varied between 0.8 to 1.4. A narrow range of slopes S (S between -0.0019 and -0.0017)
32 and intercepts Yt (Yt from 0.9725 to 1.0175) characterized *Trichodesmium* pixels (Figure 1b).
33 These ranges corresponded to low but not minimal values of the total slope range [-0.005 to +

1 0.012], and to a narrow interval of values of the total intercept Y_t range [0.8 to 1.4]) (Figure
2 3b). Moreover, it was found that RAS of pixels corresponding to *Trichodesmium* surface
3 blooms had a specific shape. These RAS spectra were characterized by: 1) lack of a “bump” at
4 555 nm (flat RAS or trough at 555 nm) and 2) always a “bump” at 670 nm and never a trough
5 at 670 nm (Figure 3b). Criteria for 412, 443, 490 and 510 nm were rather neutral. No
6 weighting of these criteria was necessary for the selection of pixels which approach this ideal
7 shape. The criteria of the RAS for the 555 nm and 670 nm SeaWiFS channels were essential
8 while the criteria for other wavelengths were less useful. On the contrary, quantitative criteria
9 of the RAS defined as in Figure 3b were discriminant. Ultimately, it appeared that the special
10 RAS shape and quantitative criteria was independent of the Chl_a of the pixels.

11 The application of *Trichodesmium* RAS criteria (TRICHODESMIUM by SATellite
12 algorithm: TRICHOSAT) to all SeaWiFS Level-2 data from 1997 to 2010 within the whole
13 Western Pacific ocean region [25°N- 25°S and 160°E -160°W, noted WP] is shown as
14 composites in Figure 4 for the interseason (April-May, October). The application of the shape
15 criteria and the quantitative criteria separately were not sufficient to discriminate
16 *Trichodesmium* surface bloom pixels (Figure 4b and 4c). The TRICHOSAT algorithm finally
17 selected the pixels which satisfy both criteria (Figure 4d), i.e. the intersection of the two
18 groups selected by the shape criteria or the quantitative criteria. The resulting composite
19 Figure 4d shows that the majority of selected *Trichodesmium* pixels is in the SP. Note that the
20 algorithm selection differs markedly from the distribution of pixels with Chl_a concentration
21 greater than $0.2 \text{ mg}\cdot\text{m}^{-3}$ (Figure 4a) as there are no *Trichodesmium* pixels in the huge
22 equatorial upwelling maximum, and only few pixels in the northern part of the Pacific Ocean.
23 *Trichodesmium* flagged pixels are not necessarily associated with high chlorophyll content
24 pixels, similar to [Westberry and Siegel \(2006\)](#).

25 In order to get statistically coherent results for the comparison of the percentage of
26 *Trichodesmium* pixels identified for each year and season, it was verified that a sufficient
27 number of processed SeaWiFS Level-2 GAC images (5×10^4 - 1×10^6 pixels with quasi
28 equivalent numbers of non-cloudy valid pixels for each season) were analyzed (Figure 5).

29 *Trichodesmium* bloom distributions by TRICHOSAT for the WP are shown as
30 composites over 12 years for each of the three seasons (winter, interseason, and summer) on
31 Figure 6abc. The increase in selected *Trichodesmium* pixels from austral winter (Figure 6a) to
32 the interseason (Figure 6b) to austral summer (Figure 6c) in the SP is visually evident.
33 Although there is a relatively high response of TRICHOSAT between 1998 and 1999 during
34 boreal winter in the northern Pacific (November-December 1998 (in yellow); January-March

1 1999 (in black)) around 160°W, no *Tricho* pixels were retrieved in the 2000-2010 years at
2 that season.

3 Figure 7a shows the temporal evolution of the percentage of *Trichodesmium* bloom pixels
4 identified by TRICHOSAT on a monthly basis. The algorithm was applied to the WP and to
5 the SP boxes. Regular peaks were observed from Dec-Feb nearly every year over the 12-year
6 period examined, albeit with strong inter-annual variations in the monthly maximum
7 percentage of retrieved *Trichodesmium* pixels exhibited strong inter-annual variations. For the
8 SP box, the percentage was above 0.2 % in 1999 and in 2004-2005 and dropped to 0.01% in
9 winter. At monthly scale, this fraction represents a total number of *Trichodesmium* bloom
10 pixels varying from 31 to 62 in Winter and from 193 to 680 in summer compared to a number
11 of valid (non-cloudy) pixels varying between 200×10^3 and 10^6 pixels with a maximum of
12 1652 in January 1999. This is equivalent to monthly surface areas varying between 500 to
13 1000 km^2 in Winter and from 3100 to 10890 km^2 in Summer with a maximum of 26432 km^2
14 in January 1999. For the WP box, the percentage was much reduced with a maximum of
15 0.03%. Indeed, the percentage was comparatively greater for SP box because there were much
16 less *Trichodesmium* pixels detected in the northern half of the domain. The mean monthly
17 chlorophyll concentration extracted from Giovanni for the SP is also shown. Note that only a
18 few *Trichodesmium* peaks correspond to a secondary chlorophyll maximum, which further
19 illustrates the independence of bloom occurrence and chlorophyll concentration.

20 The mean seasonal cycle of the *Trichodesmium* bloom pixels monthly percentage detected
21 for the 1997-2010 period in the SP is shown at Figure 8ab. Over 12 years, *Trichodesmium*
22 bloom pixels maximum were observed in January and February with relatively high inter-
23 annual variation. Similarly, the 12-y average monthly percentage of slick observations (from
24 Table 1, and [Moutin et al., 2005](#) for observations before 2004), peaks between January and
25 February.

26 TRICHOSAT was also applied to single daily level 2-GAC images around New Caledonia
27 and Vanuatu and for short periods representative of summer and winter months (Figure 9ab).
28 From the 10th to the 22th of February 2003, TRICHOSAT selected pixels spread out between
29 Vanuatu and the northwestern part of New Caledonia and Fiji (Figure 9a). Two observations
30 from the French Navy coincide with TRICHOSAT pixels. From the 9th to the 15th of June
31 2003, no pixels were detected around New Caledonia (Figure 9b). The few *Trichodesmium*
32 pixels at the southern limit of the equatorial upwelling between 7°S and 5°S could correspond
33 to observations of *Trichodesmium* blooms in the southern and northern convergence zones of
34 the upwelling ([Lebouteiller et al., 1992](#); [Blanchot, pers.com.](#)).

1 The comparison between in situ observations of slicks (summed by month from Table 1) and
2 the percentage of *Trichodesmium* bloom pixels detected on each single level-2 GAC by
3 TRICHOSAT in the SP for the total 1998-2010 period is shown at Figure 10. The striking
4 coincidence between the number of observations of surface slicks (from 1 to 6 between
5 October 2003 and February 2004) and the peak of percentage of pixels screened by
6 TRICHOSAT is the result of favorable observation conditions in the SP. Despite the reduced
7 amount of observations out of the 2003-2004 period, the maximum percentage of
8 *Trichodesmium* bloom pixels generally corresponds to a significant number (> 2) of observed
9 slicks (Figure 10).

10

11 **4. DISCUSSION**

12 **4.1- GENERAL CONSIDERATIONS OF THE ALGORITHM**

13 **A rather linear RAS spectrum**

14 TRICHOSAT identifies pixels for which the RAS is characterized by a narrow range of S
15 (small negative numbers) and of Yt (values near 1) which implies a relatively linear RAS
16 spectrum compared with the total range of S and Yt found within the whole Pacific Ocean.
17 Shape criteria (bumps and troughs), of the RAS are also essential as the intersection between
18 shape and RAS quantitative criteria is required for the successful selection of *Trichodesmium*
19 bloom pixels (Figure 4bcd). This means that the RAS spectra of *Trichodesmium* blooms are
20 very similar to what is expected at a given chlorophyll concentration contrary to other
21 phytoplankton groups for which the PHYSAT approach is based specifically on low or high
22 reflectance relatively to the detected chlorophyll content (Alvain et al., 2005). Westberry et
23 al. (2005) also found remarkably similar bulk reflectance spectra for cases which contained
24 moderate amounts of *Trichodesmium* compared to those where it was absent.

25

26 **A rather weak relationship with biomass**

27 The RAS is by definition, independent of chlorophyll concentration (second order anomaly).
28 Therefore, it is not surprising that selected pixels correspond to a large range of Chl_a values,
29 from 0.07 to 0.3 mg.m⁻³ in single level2-GAC images. It has already been shown that
30 chlorophyll associated with surface accumulations of *Trichodesmium* is highly variable. Also,
31 a well-known underestimation of chlorophyll by SeaWiFS due to a strong package effect in
32 filaments and colonies (Subramaniam et al., 1999; Dupouy et al., 2008) was observed but this

1 would concern only living colonies. As TRICHOSAT probably screens pixels containing high
2 *Trichodesmium* concentrations mixed in the upper oceanic layer or forming surface
3 accumulations, this might alter the relationship between reflectance and chlorophyll
4 concentration.

5
6

7 **Comparison with previous algorithms**

8 The percentage of TRICHOSAT selected pixels in the New Caledonia region (Figure 7, 8, 10)
9 is low (maximum 0.2% of total valid pixels). TRICHOSAT was set to detect a specific case of
10 *Trichodesmium* bloom. This configuration is when *Trichodesmium* is concentrated on 1 mm
11 or maybe more, and visible by eye. Densities are then between 17000 trichomes.L⁻¹ to 39 10⁶
12 trichomes.L⁻¹ (Devassy et al., 1978). Assuming a strong accumulation which contains about 3
13 mg.L⁻¹ Chl *a* distributed on a 1 mm thickness, this is equivalent to an integrated concentration
14 of 3 mg.m⁻² of Chl *a* related to *Trichodesmium*. If the bloom occupies an area of 100 m², i.e.
15 the 1/160000e of the pixel size, the mean Chl *a* concentration linked to *Trichodesmium* in the
16 pixel is 0.018 mg m⁻². Supposing that the rest of the phytoplankton represents on average 0.1
17 mg.m⁻³, we need about 500 m² of concentrated accumulation within the pixel to double the
18 chlorophyll concentration. The probability of finding *Trichodesmium* bloom pixels is low as
19 the probability to observe a 500 m² concentrated patch within a pixel, is also weak.
20 Nevertheless, the algorithms of [Subramaniam et al. \(2002\)](#) and [Westberry et al. \(2005\)](#)
21 (hereafter denoted as W05) detect even fewer pixels (100 times less). In the SP, the W05
22 algorithm retrieves only highly reflectant pixels, most of which correspond to the extremely
23 high reflectance of New Caledonia or Fijian sandy lagoons or Islands. Furthermore, apart
24 from the coastal lagoon pixels, only 15 points are identified as *Trichodesmium* by W05 and
25 there is no indication of seasonal variation. Recall that the W05 algorithm was built to detect
26 *Trichodesmium* at bloom concentrations (Chl_ *a* threshold for a bloom was set to 0.8 mg.m⁻³)
27 while TRICHOSAT detection does not imply Chl_ *a* concentration threshold. The rather weak
28 spectral anomalies of *Trichodesmium* pixels may explain why W05 does not detect more
29 blooms in the SP. The first published set of criteria for screening pixels containing
30 *Trichodesmium* overemphasized backscattering (for $\lambda > 500$ nm) and Colored Dissolved
31 Organic Matter (CDOM) absorption (for $\lambda < 440$ nm) so that finally a refined model using
32 subtle variations in reflectances between phytoplankton and *Trichodesmium* spectra had to be
33 used ([Westberry and Siegel, 2006](#)). In the SP, *Trichodesmium* bloom signatures may not be

1 strong enough to be detected by W05, or blooms occur in filaments whose signatures may not
2 be strong enough to affect 16 km² pixels. Finally, living colonies of *Trichodesmium* are often
3 associated with other phytoplankton as observed during the Diapalis 7 cruise (Tenório, 2006)
4 and serve to create a mixed optical signal obfuscating detection. *Trichodesmium* biomass
5 could then be well below the W05 detection threshold, and/or the other phytoplankton
6 biomass would be higher in proportion in the SP than in the northern hemisphere. This may
7 also explain why the W05 algorithm detects similar *Trichodesmium* pixel numbers in winter
8 and in summer. Overall, TRICHOSAT was tuned to detect the *Trichodesmium* blooms that we
9 have observed in the field in summer, and the W05 algorithm fails to detect these blooms.

10

11 **4.2 OPTICAL VALIDATION OF THE ALGORITHM**

12 Ideally, one would validate the RAS of TRICHOSAT detected *Trichodesmium* pixels with *in*
13 *situ* radiometric measurements made in *Trichodesmium* surface blooms. Unfortunately, such a
14 task is difficult as there is no direct measurement of the RAS which would require accurate
15 measurements of above/in water reflectance. Past work has recognized this as problematic
16 (Subramaniam et al., 1999; Kutzer, 2009). Recall that the *Trichodesmium* bloom RAS is
17 defined by a very small range of S and Yt. This implies that SeaWiFS spectra are not so
18 different from what they are expected at a given K₄₉₀ (i.e. at a given Chl_a) despite the fact
19 that filamentous cyanobacteria blooms appear brighter than surrounding water areas due to
20 high backscattering associated with gas vesicle and to a microbiotope (bacteria, detritus)
21 (Subramaniam et al., 1999).

22 One of the TRICHOSAT shape criterion is that the RAS must never show a bump at 555
23 nm. It is well known that reflectance spectra of colonies, assembled on filters (Dupouy et al.,
24 2008) are characterized by a succession of troughs, each trough corresponding to different
25 pigment absorption maxima (e.g., chlorophyll, phycourobilin and phycoerythrobilin).
26 Phycoerythrin absorption in green wavelengths are observed in *Trichodesmium* suspension
27 (PSICAM measurements, Dupouy and Röttgers, 2010), in raw H6-backscattering spectra
28 (absorption at 550 nm along the pathway of backscattered light (Dupouy et al., 2008), and
29 observed *in situ* during the 2004 bloom (unp. data)).

30 The second robust criterion is that the RAS must never show a trough at 670 nm (rather a
31 bump at 670 nm). It has been shown that high near-infrared reflectance is observed if colonies
32 are accumulated on top of the water (Subramaniam et al., 1999; Dupouy et al., 2008). RAS
33 would then depend on the proportion of colonies lying above the water surface which will

1 depends on thickness, or/and on the physiological characteristics of colonies, or age of the
2 bloom (Dupouy et al., 1990; 1992; Dupouy et al., 2008). The high reflectance of blooms at
3 670 nm which was observed with the Coastal Zone Colour Scanner (CZCS) could provide an
4 approach to detecting slicks for MODIS (Hu et al., 2010; McKinna et al., 2011) or IRS-P4
5 OCM (Sarangi et al., 2004).

6 There was no robust criteria for the RAS at 412 nm even though a significant RAS feature
7 was expected at this wavelength as it has been shown that Dissolved Organic Carbon (DOC)
8 and CDOM is released from dead or living colonies (Subramaniam et al., 1999). Optical
9 characteristics of surrounding waters may also be important for defining RAS of
10 *Trichodesmium* blooms. As seen in Figure 7b, the dissolved matter + detritus absorption
11 coefficient, ACDM (derived from the GSM optical inversion model at 412 nm; Maritorena et
12 al, 2002), for the SP box experiences a strong seasonal variation due to the well documented
13 annual cycle of solar bleaching and photolysis in the South Pacific (Siegel et al., 2002; 2005).
14 *Trichodesmium* surface blooms correspond to the period of minimum CDOM concentration in
15 summer. In the SP, some peaks are associated with a secondary CDOM maximum which
16 could be produced by blooms (e.g., February 1999, 2003, 2004 and 2006). In contrast, the
17 particulate backscattering coefficient, b_{bp} (also derived from Maritorena et al. (2002) was
18 rather constant though noisy over the 12-year period (Figure 7b). *Trichodesmium* blooms
19 correspond to the period of minimum backscattering, as a weak maximum appears in June-
20 September (more or less in phase with chlorophyll concentration). The b_{bp} cycle in the SP
21 could be linked to small-sized detritus as suggested by Loisel et al. (2006) rather than to
22 *Trichodesmium* blooms.

23 The relatively high response by TRICHOSAT in the northern Pacific around 160°W
24 between 1998 and 1999 during boreal winter (November-December 1998 (in yellow);
25 January-March 1999 (in black)) do not correspond to visual observations of blooms in this
26 area (Dore et al., 2008). From 2000 to 2010, no *Trichodesmium* pixels were retrieved during
27 boreal winter in the Northern Pacific area by TRICHOSAT. We have no explanation for what
28 happened during the 98-99 boreal winter period near Hawaii. We conclude that there is some
29 signal there picked up by the TRICHOSAT algorithm. Blooms of unknown origin were
30 already reported using CZCS observations in December (Dore et al, 2008). This has to be
31 caused by floating living material similar in reflectance to *Trichodesmium* and therefore
32 having the same SeaWiFS RAS. Also, there may be a different relation between reflectance
33 and K_{490} in SeaWiFS data for these five months than for the rest of the studied period. A new

1 research on this zone should be done to determine the nature of this optical signal in the
2 Northern Pacific.

3 **4.3 RELATIONSHIPS WITH OTHER CYANOBACTERIA IN THE** 4 **WATER COLUMN**

5 The austral summer maxima of surface blooms detected with TRICHOSAT in SP (Figure
6 7a) corresponds with the inter-annual variations of *Trichodesmium* abundances in the surface
7 layer (2 meters, Table 2). Lower *Trichodesmium* densities and slick observations were
8 observed in situ in April-May 2002-2003 and October 2001-2003 (Table 1 and 2) which also
9 appear as lower surface bloom extent identified by TRICHOSAT. A surprisingly low
10 *Trichodesmium* concentration (200-400 trichomes.L⁻¹) was observed in the water column in
11 December 2001-January 2002 compared to 2002 and 2003 February months. In June-August
12 2003 (winter season), *Trichodesmium* was rare and total chlorophyll in the > 10 µm fraction
13 was less than 10% (Table 2). Thus, the community was dominated by picoplankton
14 (cyanobacteria and picoeucaryotes) and surface bloom extent detected by TRICHOSAT was
15 minimum.

16 The number of selected pixels by TRICHOSAT is however small. It corresponds to
17 surface blooms, occurring in summer, which occupy a large area according to ocean color
18 maps and to field observations. These surface blooms correspond to a physiological state
19 where colonies become buoyant because they cannot synthesize ballast anymore due to
20 phosphate limitation in; colonies thus float as dead material and accumulate at the surface
21 (Moutin et al., 2005). Such conditions may be spatially and temporally de-coupled from sub-
22 bloom *Trichodesmium* concentrations and may not correspond to actively growing
23 *Trichodesmium* colonies. So, the global estimation of dinitrogen fixation by TRICHOSAT
24 remains to be estimated.

25 Last, the relative abundance of *Trichodesmium* and other nitrogen-fixing cyanobacteria
26 needs to be determined. Coccoid cyanobacteria (*Chrocosphaera*, *Cyanothece*) have been
27 detected from their phycoerythrin signature (Neveux et al., 2006) and flow cytometric
28 properties in the SP (Neveux, unpublished res.; Campbell et al., 2006; Moisander et al., 2010;
29 Sato et al., 2010; Biegala, pers. com.). Nitrogen-fixing heterotrophic bacteria have also been
30 identified in the SP (Rieman et al., 2010). During summer, nitrogen fixation both from
31 filamentous and coccoid cyanobacteria (Garcia et al., 2007; Hynes et al., 2009) experienced
32 high rates (151-703 µM N₂ m⁻² d⁻¹) compared to rates measured at the North of Papua-New

1 Guinea (Bonnet et al., 2009). Late 2007 field observations confirmed the dominance of
2 *Trichodesmium* in association with *Crocosphaera* in the South Pacific, spatially decoupled
3 from unicellular cyanobacteria (Hewson et al., 2009; Moisander et al., 2010; Sato et al.,
4 2010). *Crocosphaera*, *Cyanothece* and picocyanobacteria populations may form a high
5 fraction of total nitrogen fixation which will not be detected by TRICHOSAT. Applications of
6 the algorithm for an estimate of potential nitrogen fixation would require a better
7 understanding of the relationship between the development of *Trichodesmium* over the whole
8 water column and surface accumulations. Furthermore, a better knowledge of the spatial and
9 temporal association of coccoid cyanobacteria with *Trichodesmium* (Campbell et al., 2006;
10 Sato et al., 2010; Moisander et al., 2010), would help in getting a global view of all nitrogen
11 fixing organisms.

12

13 **5. CONCLUSION**

14 The TRICHOSAT algorithm is efficient at discriminating *Trichodesmium* surface
15 accumulations in the Western tropical Pacific Ocean. Its results can complement the
16 identification of major functional groups provided by PHYSAT. TRICHOSAT was developed
17 for the SP and is even able to follow the daily evolution of surface blooms over the whole
18 1998-2010 period (Figure 10). Its application at a global scale requires that the RAS criteria
19 in TRICHOSAT is representative of *Trichodesmium* blooms in other regions, and also that
20 surrounding waters characteristics resemble those in the SP. Reasons why detection of
21 *Trichodesmium* with other algorithms was much less successful (100 times less pixels and no
22 seasonality) in the Western Tropical Pacific Ocean may be, 1) low *Trichodesmium*
23 concentrations, 2) a weak discriminating optical signal such as that due to a mixed optical
24 assemblage, and 3) a significantly different *Trichodesmium* optical signature in the SP.

25 *Trichodesmium* experienced a high inter-annual variability in the region with maxima in
26 1999, 2003, 2004 and 2005, while 2001 and 2002 were less favourable. This inter-annual
27 variability might be related to large scale circulation dynamics. For example, the bifurcation
28 latitude of the South Equatorial Current is found to move southward from about 15°S near the
29 surface to south of 22°S in the intermediate layers (Qu and Lindstrom, 2002) associated with
30 large scale changes in the phosphate pool (Dyrhman et al., 2006; Tadokoro et al., 2009). Also,
31 iron-rich dust deposition patterns (Gao et al., 2001) may be highly variable. Whether other
32 phytoplankton blooms are triggered by *Trichodesmium* decomposition of organic matter is

1 also of interest (Chen et al., 2008; 2011). Late summer chlorophyll blooms in the oligotrophic
2 North Pacific subtropical gyre may be fuelled by *Trichodesmium* (Wilson and Qiu, 2008).

3 TRICHOSAT demonstrated that *Trichodesmium* blooms are a common feature in the SP
4 during austral summer (October to March). They are temporally and spatially linked to
5 diverse nitrogen-fixing populations and can be identified as a major potential carbon sink in
6 the SP. A large-scale physical and biogeochemical modeling of the distribution of
7 *Trichodesmium* is required in the SP.

8

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7 REFERENCES

8 Acker, J. G. and Leptoukh, G.: Online Analysis Enhances Use of NASA Earth Science Data,
9 Eos, Trans. AGU, Vol. 88(2), p. 14, p. 17, 2007.

10 Alvain, S., Moulin, C., Dandonneau, Y., and Breon, F. M.: Remote sensing of phytoplankton
11 group in case 1 waters from global SeaWiFS imagery, Deep-Sea Res. Pt I, 52, 1989-2004,
12 2005.

13 Barcelos e Ramos, J., Biswas, H., Schulz, K.G., LaRoche, J., and Riebesell, U.: Effect of
14 rising atmospheric carbon dioxide on the marine nitrogen fixer *Trichodesmium*. Global
15 Biogeochemical Cy., 21, GB2028, doi:10.1029/2006GB002898, 2007.

16 Bell, P. R. F., Elmetri, I., and Uwins, P.: Nitrogen fixation by *Trichodesmium* spp. in the
17 Central and Northern Great Barrier Reef Lagoon: relative importance of the fixed-nitrogen
18 load. Mar. Ecol. -Prog. Ser., 186, 119-126, 1999.

19 Berman-Frank, I., Bidle, K. D., Haramaty, L., and Falkowski, P. G.: The demise of the marine
20 cyanobacterium, *Trichodesmium* spp., via an autocatalyzed cell death pathway. Limnol.
21 Oceanogr., 49, 997-1005, 2004.

22 Bonnet, S., Biegala, I. C., Dutrieux, P., Slemmons, L. O., and Capone, D. G.: Nitrogen fixation
23 in the western equatorial Pacific: rates, diazotrophic cyanobacterial size class distribution,
24 and biogeochemical significance. Global Biogeochem. Cy., 23, 1-13, 2009.

25 Bowman, T. E., and Lancaster, L. J.: A bloom of the planktonic blue-green alga,
26 *Trichodesmium erythraeum*, in the Tonga Islands. Limnol. Oceanogr., 10, 291-293, 1965.

27 Bracher, A., Vountas M., Dinter, T., Burrows, J. P., Röttgers, R., and Peeken, I.: Quantitative
28 observation of cyanobacteria and diatoms from space using PhytoDOAS on
29 SCIAMACHY data, Biogeosciences, 6, 751-754, doi:10.5194/bg-6-751-2009, 2009.

30 Breitbarth, E., Oschlies, A., and LaRoche, J.: Physiological constraints on the global
31 distribution of *Trichodesmium*: effect of temperature on diazotrophy, Biogeosciences, 4,
32 53-61, doi:10.5194/bg-4-53-2007, 2007.

- 1 Bryan, S. E., Cook, A., Evans, J. P., Colls, P. W., Wells, M. G., Lawrence, M. G., Jell, J. S.,
2 Greig, A., Leslie, E.: Pumice rafting and faunal dispersion during the 2001-2002 in the
3 Southwest Pacific: record of a dacitic submarine explosive eruption from Tonga. *Earth*
4 *Plan. Sci. Lett.*, 227, 135-154, 2004.
- 5 Campbell, L., Carpenter, E. J., Montoya, J. P., Kustka, A. B., and Capone, D.G.: Picoplankton
6 community structure within and outside a *Trichodesmium* bloom in the southwestern
7 Pacific Ocean. *Vie Milieu*, 55, 185-195, 2006.
- 8 Capone, D. G. and Knapp, A. N.: Oceanography, A marine nitrogen cycle fix? *Nature*, 445,
9 159-160, doi:10.1038/445159A, 2007.
- 10 Capone, D. G., Zehr, J. P., Paerl, H. W. , Berman, B., and Carpenter, E. J.: *Trichodesmium*, a
11 globally significant marine cyanobacterium. *Science*, 276, 1221-1229, 1997.
- 12 Capone, D. G., Burns, J.A., Montoya, J. P., Subramaniam, A., Mahaffey, C., Gunderson, T.,
13 Michaels, A.F., and Carpenter E. J.: Nitrogen fixation by *Trichodesmium* spp.: an
14 important source of new nitrogen to the tropical and subtropical North Atlantic Ocean.
15 *Global Biogeochem. Cy.*, 19, GB2024, doi:10.1029/2004GB002331, 2005.
- 16 Carpenter, E. J.: Nitrogen fixation by marine Oscillatoria *Trichodesmium* in the world's
17 ocean. In: *Nitrogen in the Marine Environment*, edited by: Carpenter, E. J. and Capone, D.
18 J., Academic Press, New-York, 65-103, 1983.
- 19 Carpenter, E. J., O'Neil, J. M., Dawson, R., Capone, D. G., Siddiqui, P. J. A., Roenneberg, T.,
20 and Bergman, B.: The tropical diazotrophic phytoplankter *Trichodesmium*: biological
21 characteristics of two common species. *Mar. Ecol.-Prog. Ser.*, 95, 295-304, 1993.
- 22 Chen, Y. L. L., Chen H. Y., Tuo S., and Ohki, K.: Seasonal dynamics of new production from
23 *Trichodesmium* N₂ fixation and nitrate uptake in the upstream Kuroshio and South China
24 Sea basin. *Limnol. Oceanogr.*, 53, 1705–1721, 2008.
- 25 Chen, Y. L. L., Tuo, S., and Chen, H.Y.: Co-occurrence and transfer of fixed nitrogen from
26 *Trichodesmium* spp. to diatoms in the low-latitude Kuroshio Current in the NW Pacific.
27 *Mar. Ecol.-Prog. Ser.*, 421, 25-38, 2011.
- 28 Church, M. J., Bjorkman, K. M., Karl, D. M., Saito, M. A., and Zehr, J. P.: Regional
29 distributions of nitrogen fixing bacteria in the Pacific Ocean, *Limnol. Oceanogr.*, 53(1),
30 63-77, 2008.
- 31 Church, M. J., Mahaffey, C., Letelier, R. M., Lukas, R., Zehr, J. P., and Karl, D. M.: Physical
32 forcing of nitrogen fixation and diazotroph community structure in the North Pacific

1 Subtropical Gyre, Global Biogeochem. Cy., 23, GB2020, doi:10.1029/2008GB003418,
2 2009.

3 Dandonneau, Y. and Gohin, F.: Meridional and seasonal variations of the sea surface
4 chlorophyll concentration in the South western tropical Pacific ocean. Deep-Sea Res., 31,
5 137-139, 1984.

6 Deutsch, C., Sarmiento, J. L., Sigman, D. M., Gruber, N., and Dunne, J. P.: Spatial coupling
7 of nitrogen inputs and losses in the ocean, Nature, 445, doi:10.1038/nature05392, 2007.

8 Devassy, V. P., Bhattathiri, P. M. A., and Qasim, S. Z.: *Trichodesmium* phenomenon, Ind. J.
9 Mar. Sci., 7, 168–186, 1978.

10 Dore, J. E., Letelier, R. M., Church, M. J., Lukas, R., and Karl, D.: Summer phytoplankton
11 blooms in the oligotrophic North Pacific Subtropical gyre: Historical perspective and
12 recent observations, Prog. Oceanogr., 76, 2-38, 2008.

13 Dupouy, C.: La chlorophylle de surface observée par le satellite NIMBUS-7 CZCS autour de
14 la Nouvelle Calédonie et de ses dépendances. Une première analyse. Bulletin de l'Institut
15 Océanographique de Monaco, p. 125-148. Colloque Scientifique Franco-Japonais;
16 Colloque d'Océanographie, 5: 2, Tokyo; Shimizu J. P. N, 3-13 October 1998, 1990.

17 Dupouy, C.: Discoloured waters in the Melanesian archipelago New Caledonia and Vanuatu.
18 The value of the Nimbus-7 Coastal Zone Colour Scanner observations, in: Marine Pelagic
19 Cyanobacteria: *Trichodesmium* and other diazotrophs, edited by Carpenter, E. J., Capone,
20 D. G. and Rueter J. G., Kluwer Academic Press, NATO Adv. Sci. I. C-Mat., 362: 177-
21 191, 1992.

22 Dupouy, C. and Röttgers, R.: Absorption by different components during a high freshwater
23 event of the 2008 La Nina episode in a tropical lagoon. Poster Session “Bio-optics and
24 biogeochemistry”, Ocean Optics XX, Anchorage (Alaska), 25-30 september 2010.

25 Dupouy, C., Petit, M., and Dandonneau, Y.: Satellite detected cyanobacteria bloom in the
26 southwestern tropical Pacific. Implication for oceanic nitrogen fixation. Int. J. Rem.
27 Sens., 9, 389-396, 1988.

28 Dupouy, C., Neveux, J., Subramaniam, A., Mulholland, M., Campbell, L., Montoya, J.,
29 Carpenter, E., and Capone, D.: Satellite captures *Trichodesmium* blooms in the
30 Southwestern Tropical Pacific. EOS Trans. AGU, 81, 13, 14-16, 2000.

31 Dupouy, C., Dirgerg, G., Tenório, M. M. B., Neveux, J., and Le Bouteiller, A.: Surveillance
32 des *Trichodesmium* autour de la Nouvelle-Calédonie, du Vanuatu, de Fidji et de Tonga
33 1998-2004. Archives Sciences de la Mer, 7, 51, 2004a.

- 1 Dupouy, C., Neveux, J., and Le Bouteiller, A.: Spatial and temporal analysis of SeaWiFS sea
2 surface chlorophyll, temperature, winds and sea level anomalies in the South Tropical
3 Pacific Ocean (10°S-25°S, 150°E-180°E). In : Proceedings “6ème conférence PORSEC,
4 Pan Ocean Remote Sensing Conference”, 29 November- 3 December 2004, Guyana,
5 Conception (Chili), Guyana 68(2) Suppl. I. Proc. 161-166, 2004 ISSN 0717-652X, doi:
6 10.4067/S0717-65382004000200030, 2004b.
- 7 Dupouy, C., Neveux J., Dirberg, G., Röttgers, R., Tenório, M. M. B., and Ouillon, S.: Bio-
8 optical properties of the marine cyanobacteria *Trichodesmium* spp., J. Appl. Remote
9 Sens., 2, 1-17. doi:10.1117/1.2839036, 2008.
- 10 Dyrhman, S. T., Chappell, P. D., Haley, S. T., Moffett, J.W., Orchard, E. D., Waterbury, J. B.,
11 and Webb, E. A.: Phosphonate utilization by the globally important marine diazotroph
12 *Trichodesmium*. Nature, 439, 68-71, 2006.
- 13 Falkowski, P.G.: Evolution of the nitrogen cycle and its influence on the biological
14 sequestration of CO₂ in the ocean, Nature, 387, 272-275, 1997.
- 15 Feldman, G. C. and McClain C. R.: Ocean Color Web, edited by: Kuring, N., Bailey, S. W.,
16 Franz, B. F., Meister, G., Werdell, P. J., and Eplee, R. E., NASA Goddard Space Flight
17 Center, 2010.
- 18 Furnas, M. J.: Cyclonic disturbance and a phytoplankton bloom in a tropical shelf ecosystem,
19 in: Red tides: Environmental Science and Toxicology, edited by: Okaichi, T., Anderson, D.
20 M., and Nemoto, T., Elsevier, Amsterdam, 271-274, 1989.
- 21 Gao, Y., Kaufman, Y.J., Tanre, D., Kolber, D., and Falkowski, P.: Seasonal distributions of
22 aeolian iron fluxes to the global ocean. Geophys. Res. Lett. 28: 29-32, 2001.
- 23 Garcia, N., Raimbault, P., and Sandroni, V.: Seasonal nitrogen fixation and primary
24 production in the southwest Pacific: nanoplankton diazotrophy and transfer of nitrogen to
25 picoplankton organisms. Mar. Ecol.-Prog. Ser. 343:25-33, 2007.
- 26 Gruber, N. and Sarmiento, J. L.: Global patterns of marine nitrogen fixation and
27 denitrification. Global Biogeochem. Cy., 11, 235-266, 1997.
- 28 Hashihama, F., Sato, M., Takeda, S., Kanda, J., and Furuya, K.: Mesoscale decrease of
29 surface phosphate and associated phytoplanktonic dynamics in the vicinity of the
30 subtropical South Pacific Islands. Deep-Sea Res. Pt I, 57, 338-350, 2010.
- 31 Hewson, I., Poretsky, R. S., Dyrhman, S. T., Zielinski, B., White, A. E., Tripp, H. J.,
32 Montoya, J., and Zehr, J. P.: Microbial community gene expression within colonies of the
33 diazotroph, *Trichodesmium*, from the Southwest Pacific Ocean, ISME, 1-15, 2009.

- 1 Hu, C., Cannizzaro, J., Carder, K. L., Muller-Karger, F. E., and Hardy, R.: Remote detection
2 of *Trichodesmium* blooms in optically complex coastal waters: Examples with MODIS
3 full-spectral data, *Remote Sens. Environ.*, 114, 2048-2058, 2010.
- 4 Hutchins, D.A., Fu, F.-X., Zhang, Y., Warner, M.E., Portune, K., Bernhardt, P.W., and
5 Mulholland, M.R.: CO₂ control of *Trichodesmium* N₂ fixation, photosynthesis, growth
6 rates, and elemental ratios: implications for past, present, and future ocean
7 biogeochemistry. *Limnol. Oceanogr.*, 52, 1293–1304, 2007.
- 8 Hynes, A. M., Chappell, P. D., Dyhrman, S. T., Doney, S. C., and Webb, E. A.: Cross-basin
9 comparison of phosphorus stress and nitrogen fixation in *Trichodesmium*. *Limnol.*
10 *Oceanogr.*, 54, 5, 1438-1448, 2009.
- 11 Karl, D. M., Michaels, A., Bergman, B., Capone, D., Carpenter, E., Letelier, R., Lipschultz,
12 F., Paerl, H., Sigman, D., and Stal, L.: Dinitrogen fixation in the world's oceans.
13 *Biogeochemistry*, 57/58, 47-98, 2002.
- 14 Kitajima, S., Furuya, K., Hashihama, F., and Takeda, S.: Latitudinal distribution of
15 diazotrophs and their nitrogen fixation in the tropical and subtropical western North
16 Pacific, *Limnol. Oceanogr.*, 54, 537-547, 2009.
- 17 Konno, U., Tsunogai, U., Komatsu, D. D., Daita, S., Nakagawa, F., Tsuda, A., Matsui, T.,
18 Eum, Y.-J., and Suzuki, K., 2010. Determination of total N₂ fixation rates in the ocean
19 taking into account both the particulate and filtrate fractions, *Biogeosciences*, 7, 2369–
20 2377, doi:10.5194/bg-7-2369-2010, 2010.
- 21 Kranz, S., Sültemeyer, D., Richter, K. U., and Rost, B.: Carbon acquisition by
22 *Trichodesmium*: the effect of pCO₂ and diurnal changes. *Limnol. Oceanogr.*, 54, 548-559,
23 2009.
- 24 Kuchler, D. and Jupp, D. L. B.: Shuttle photograph captures massive phytoplankton bloom in
25 the Great Barrier Reef, *Int. J. Remote Sens.*, 9(8), 1299-1301, 1988.
- 26 Kutzer, T.: Passive optical remote sensing of cyanobacteria and other intense phytoplankton
27 blooms in coastal and inland waters, *Int. J. Remote Sens.*, 30(17), 4401-4425, 2009.
- 28 Lantoiné, F. and Neveux, J.: Spatial and seasonal variations in abundance and spectral
29 characteristics of phycoerythrins in the Tropical Northeastern Atlantic Ocean. *Deep-Sea*
30 *Res. Pt I*, 44: 223-246, 1997.
- 31 LaRoche J. and Breitbarth, E.: Importance of the diazotrophs as a source of new nitrogen in
32 the ocean. *J. Sea Res.*, 53, 67-69, 2005.
- 33 Levitan, O., Brown, C. M., B., Sudhaus, S., Campbell, D., LaRoche, J., and Berman-Frank, I.:
34 Regulation of nitrogen metabolism in the marine diazotroph *Trichodesmium* IMS101

1 under varying temperatures and atmospheric CO₂ concentrations, *Environ. Microb.*, 12,
2 1899-1912, 2010.

3 Loisel, H., Nicolas, J. M. Sciandra, A., and Stramski, D.: Spectral dependency of optical
4 backscattering by marine particles from satellite remote sensing of the global ocean”, *J.*
5 *Geophys. Res.* 111, C09024, doi:10.1029/2005JC003367, 2006.

6 Lundgren, P., Janson, S., Jonasson, S., Singer, A., and Bergman B.: Unveiling of Novel
7 Radiations within *Trichodesmium* Cluster by *hetR* Gene Sequence Analysis, *App.*
8 *Environm. Microb.*, 71, 190-196, 2005.

9 Mahaffey, C., Michaels, A.F., and Capone, D.G.: The conundrum of marine N₂ fixation.
10 *Amer. J. Sci.*, 305: 546-595, 2005.

11 Mantas, V. M., Pereira, A.J.S.C., Morais, P.V.: P.V. Plumes of discolored water of volcanic
12 origin and possible implications for algal communities. The case of the Home Reef
13 eruption of 2006 (Tonga, Southwest Pacific Ocean). *Remote Sens. Environ.*, 115, 1341–
14 1352, 2011.

15 Maritorena, S. and Siegel, D. A.: Consistent merging of satellite ocean color data sets using a
16 bio-optical model. *Remote Sensing Environ.*, 94, 429-440, 2005.

17 Masotti I., Ruiz-Pino, D., and Le Bouteiller, A.: Photosynthetic characteristics of
18 *Trichodesmium* in the southwest Pacific Ocean: importance and significance. *Mar. Ecol.-*
19 *Prog. Ser.*, 338, 37-49, 2007.

20 McKinna, L. I.W., Furnas, M. J., and Ridd, P. V.: A simple, binary classification algorithm
21 for the detection of *Trichodesmium* spp. within the Great Barrier Reef using MODIS
22 imagery, *Limnol. Oceanogr. Methods*, 9:50-66 (2011), doi: 10.4319/lom.2010.9.50, 2011.

23 Moisander, P. H., Beinart, R. A., Hewson, I. White, A. E., Johnson, K. S., and Carlson, C. A.:
24 Unicellular Cyanobacterial Distributions Broaden the Oceanic N₂ Fixation Domain,
25 *Science*, 327, 1512-1514, doi:10.1126/science.1185468, 2010.

26 Monteiro, F. M., Follows, M. J., and Dutkiewicz, S.: Distribution of diverse nitrogen fixers in
27 the global ocean, *Global Biogeochem. Cy.*, 24, GB3017, doi:10.1029/2009GB003731,
28 2010.

29 Montoya, J., Holl, C. M., Zehr, J. P., Hansen, A., Villareal, T. A., and Capone, D. G.: High
30 Rates of N₂ Fixation by Unicellular Diazotrophs in the Oligotrophic Pacific. *Nature*, 430,
31 1027-1032, 2004.

32 Moutin, T., Van den Broeck, N., Beker, B., Dupouy, C., Rimmelin, P., and Le Bouteiller, A.:
33 Phosphate availability controls *Trichodesmium* spp. biomass in the SW Pacific Ocean.
34 *Mar. Ecol.-Prog. Ser.*, 207, 15-21, 2005.

- 1 Mulholland, M.: The fate of nitrogen fixed by diazotrophs in the ocean, *Biogeosciences*, 4,
2 37-51, doi.10.5194/bg-4-37-2007, 2007.
- 3 Neveux, J., Lantoiné, F., Vaultot, D., Marie, D. and Blanchot. J.: Phycoerythrins in the
4 southern tropical and equatorial Pacific Ocean: Evidence for new cyanobacterial types. *J.*
5 *Geophys. Res.*, 104(C2): 3311–3321, 1999.
- 6 Neveux, J., Tenório, M.M.B., Dupouy, C., and Villareal, T.: Spectral diversity of
7 phycoerythrins and diazotrophs abundance in tropical South Pacific. *Limnol. Oceanogr.*,
8 51, 4, 1689-1698, 2006.
- 9 Qu T. and Lindstrom, E. J.: A climatological interpretation of the circulation in the western
10 south Pacific. *J. Phys. Oceanogr.*, 32, 2492-2508, 2002.
- 11 Ramos, A.G.R., Martel, A., Codd, G.A., Soler, E., Coca, J., Redondo, A., Morrison, L.F.,
12 Metcalf, J.S., Ojeda, A., Duarez, S., and Petit, M.: Bloom of the marine diazotrophic
13 cyanobacterium *Trichodesmium erythraeum* in the Northwest African Upwelling, *Mar.*
14 *Ecol.-Prog. Ser.*, 301, 303-305, 2005.
- 15 Revelante, N. and Gilmartin, M.: Dynamics of phytoplankton in the Great Barrier Reef
16 lagoon. *J. Plankton Res.*, 4, 47-76, 198, 1982.
- 17 Riemann, L., Farnelid, H., and Steward, G.F.: Nitrogenase genes in non-cyanobacterial
18 plankton: prevalence, diversity and regulation in marine waters, *Aquat. Microb. Ecol.*, 61,
19 235–247, doi:10.3354/ame01431, 2010.
- 20 Rubin, M., Berman-Frank, I. and Shaked H. : Dust- and mineral-iron utilization by the marine
21 dinitrogen-fixer *Trichodesmium*. *Nature Geosci.*, 4, 529–534, doi:10.1038/ngeo1181, 2011.
- 22 Sarangi, R.K., Chauhan, P., and Nayak, S. R.: Detection and monitoring of *Trichodesmium*
23 blooms in the coastal waters off Saurashtra coast, India using IRS-P4 OCM data, *Curr.*
24 *Sci.*, 86, 12, 1636-1641, 2004.
- 25 Sato, M., Hashihama, F., Kitajima, S., Takeda, S., and Furuya, K.: Distribution of nano-sized
26 cyanobacteria in the western and central Pacific Ocean, *Aquatic Microb. Ecol.*, 59, 273-
27 282, 2010.
- 28 Shiozaki, T., Furuya, K., Kodama, T., and Takeda, S.: Contribution of N₂ fixation to new
29 production in the western North Pacific Ocean along 155°E, *Mar. Ecol.-Prog. Ser.*, 377,
30 19-32, 2009.
- 31 Siegel, D. A., S. Maritorena, N. B. Nelson, D. A. Hansell, and Lorenzi-Kayser, M.: Global
32 distribution and dynamics of colored dissolved and detrital organic materials, *J. Geophys.*
33 *Res.*, 107(C12), 3228, doi:10.1029/2001JC000965, 2002.

- 1 Siegel, D. A., S. Maritorena, N. B. Nelson, and Behrenfeld, M. J.: Independence and
2 interdependencies of global ocean color properties: Reassessing the bio-optical
3 assumption, *J. Geophys. Res.*, 110, C07011, doi:10.1029/2004JC002527, 2005.
- 4 Subramaniam, A., E. J., Carpenter, D., Karentz, and Falkowski, P.G.: Bio-optical properties
5 of the diazotrophic cyanobacteria *Trichodesmium* spp. I. Absorption and photosynthetic
6 action spectra. *Limnol. Oceanogr.*, 44, 608-617, 1999.
- 7 Subramaniam, A., Brown, C. W., Hood, R. R., Carpenter, E. J., and Capone, D. G.: Detecting
8 *Trichodesmium* blooms in SeaWiFS imagery. *Deep-Sea Res. Pt I*, 49, 107-121, 2002.
- 9 Tadokoro, K., Ono, T., Yasuda, I., Osafune, S., Shiimoto, A., and Sugisaki, H.: Possible
10 mechanisms of decadal scale variation in PO₄ concentration in the western North Pacific,
11 *Geophys. Res. Lett.*, 36, L08606, doi:10.1029/2009GL037327, 2009.
- 12 Tenório, M. M. B. : Les cyanobactéries en milieu tropical: occurrence, distribution, écologie
13 et dynamique, PhD Thesis, Université Paris VI, 2006.
- 14 Westberry T., Subramaniam, A., and Siegel, D.: An improved bio-optical algorithm for the
15 remote sensing of *Trichodesmium* spp. blooms, *J. Geophys. Res.*, 110, C06012,
16 doi10.1029/2004JC, 2005.
- 17 Westberry, T. K. and Siegel, D. A.: Spatial and temporal distribution of *Trichodesmium*
18 blooms in the world's oceans. *Global Biogeochemical Cy.*, 20, 4016,
19 doi10.1029/2005GB002673, 2006.
- 20 Wilson, C. and X. Qiu, X.: Global distribution of summer chlorophyll blooms in the
21 oligotrophic gyres, *Prog. Oceanogr.*, 78, 107-134, doi:10.1016/j.pocean.2008.05.002,
22 2008.
- 23 Zehr, J. P. and Kudela, R. M.: Nitrogen cycle of the open ocean: from genes to ecosystems,
24 *Annu. Rev. Marine Sci.*, 3, 197-225, 2011.
- 25 Zehr, J. P., Waterbury, J. B., Turner, P. J., Montoya, J. P., Omoregie, E., Steward, G. F.,
26 Hansen, A., and Karl, D. M.: Unicellular cyanobacteria fix N₂ in the subtropical North
27 Pacific Ocean. *Nature*, 412, 635-638: doi:10.1038/35088063, 2001.
- 28 .

1

2 **Table 1.** In situ *Trichodesmium* bloom observations for the period 1998-2010 in the South Western Tropical Pacific (NC, New Caledonia). Data
3 from aerial observations, and/or bucket sampling from cruises and various transects made by the French Navy ships and the R/V *Alis*.
4 Abbreviations: T. e., T. t. and T. ten., K., for *Trichodesmium erythraeum*, *thiebautii*, and *tenue*, *Katagnymene* spp., respectively. Col. = colonies;
5 fil. = filaments. Nq s.: nautical miles slicks, shipb.: Shipboard observations. * Ground-truthed (microscopy) aerial or shipboard observations are
6 indicated by an asterisk, ** E. J. Carpenter, personal communication

| Yearday/Year | Date | Latitude | Longitude | Observed species | Observed form | Estimated Slick size | Location in the Tropical Pacific | Observation Mean |
|--------------|-------------|----------|-----------|---------------------|------------------|-------------------------|-------------------------------------|-------------------------------------|
| 101/1998** | 11 Apr 1998 | 17° S | 179° E | T. t. | large col. | und. | Fiji Isl. | R/V <i>Revelle</i> , NSF |
| 101/1998* | 11 Apr 1998 | 18° S | 188° E | T. t. | large col. | und. | Vavau Isl. | F. Navy, <i>J. Cartier</i> |
| 108/1998 | 18 Nov 1998 | 22°35' S | 168°58' E | und. | und. | nq s. | Walpole Isl. | F. Navy aerial |
| 108/1998 | 18 Nov 1998 | 24°21' S | 170°40' E | und. | und. | nq s. | SE NC | F. Navy aerial |
| 329/1998 | 25 Nov 1998 | 22°15' S | 163°35' E | und. | und. | nq s. | W NC | F. Navy aerial |
| 329/1998 | 25 Nov 1998 | 21°20' S | 164°10' E | und. | und. | nq s. | W NC | F. Navy aerial |
| 329/1998 | 25 Nov 1998 | 19°35' S | 160°55' E | und. | und. | 30 km ² | NW NC | F. Navy aerial |
| 352/1998 | 18 Dec 1998 | 22°40' S | 165°50' E | und. | und. | nq s. | SW NC | F. Navy aerial |
| 008/1999 | 8 Jan 1999 | 23°S | 166°E | und. | und. | nq s. | SW NC | F. Navy aerial |
| 008/1999 | 8 Jan 1999 | 24°10' S | 166°15' E | und. | und. | nq s. | SW NC | F. Navy aerial |
| 008/1999 | 8 Jan 1999 | 23°50' S | 164°20' E | und. | und. | nq s. | SW NC | F. Navy aerial |
| 313/1999 | 9 Nov 1999 | 20° S | 170° E | und. | und. | 30 km ² | Vanuatu to NC | R/V <i>Alis</i> (<i>Wespalis</i>) |
| 007/2000 | 7 Jan 2000 | 21°49' S | 165°11' E | und. | und. | nq s. | W NC | F. Navy aerial |
| 007/2000 | 7 Jan 2000 | 20°30' S | 161°52' E | und. | und. | nq s. | W NC | F. Navy aerial |
| 363/2000 | 28 Dec 2000 | 19°57' S | 162°58' E | und. | und. | nq s. | Belep | F. Navy aerial |
| 363/2000 | 28 Dec 2000 | 21°20' S | 164°12' E | und. | und. | nq s. | Poum | F. Navy aerial |
| 363/2000 | 28 Dec 2000 | 21°48' S | 163°35' E | und. | und. | nq s. | Poum | F. Navy aerial |
| 008/2001 | 8 Jan 2001 | 22°20' S | 172° E | und. | und. | nq s. | Hunter Isl. | F. Navy aerial |

| | | | | | | | | |
|-----------|-------------|----------|-----------|-----------------------|--------------------------|--------|--------------------|------------------------------|
| 018/2001 | 18 Jan 2001 | 21°11' S | 164°17' E | und. | und. | nq s. | NW Kone, NC | F. Navy aerial |
| 018/2001* | 18 Jan 2001 | 18°31' S | 164°28' E | T. e., T. t., T. Ten. | large col., green | nq s. | Petri Reefs | F. Navy aerial |
| 23/2001* | 23 Jan 2001 | 22°11' S | 161°79' E | T. e. | Isolated fil. | nq s. | Coral Sea | F. Navy aerial |
| 23/2001* | 23 Jan 2001 | 22°10' S | 161°79' E | T. e. | flocks | Shipb. | Coral Sea | F. Navy, <i>La Moqueuse</i> |
| 29/2001 | 29 Jan 2001 | 24°53' S | 161°40' E | und. | flocks | Shipb. | Coral Sea | F. Navy, <i>La Moqueuse</i> |
| 29/2001 | 29 Jan 2001 | 24°47' S | 162°37' E | und. | flocks | Shipb. | Coral Sea | F. Navy, <i>La Moqueuse</i> |
| 125/2001* | 7 May 2001 | 20°12' S | 163°29' E | T. e. | green col. | nq s. | Belep | F. Navy aerial |
| 248/2001 | 5 Sep 2001 | 20°08' S | 170°45' E | und. | und. | nq s. | Anatom | F. Navy aerial |
| 248/2001 | 5 Sep 2001 | 20°41' S | 170°16' E | und. | und. | nq s. | Anatom | F. Navy aerial |
| 295/2001* | 22 Oct 2001 | 22°16' S | 167°27' E | T. e., T. t., K. | flocks | Shipb. | E NC | R/V <i>Alis</i> (Diapalis 1) |
| 285/2001* | 3 Nov 2001 | 20°47' S | 167°03' E | T. e. | grey col. | nq s. | Lifou Isl. | F. Navy aerial |
| 317/2001* | 13 Nov 2001 | 22°09' S | 167°19' E | T. e., T. t., K. | und. | nq s. | East Coast | F. Navy, <i>La Glorieuse</i> |
| 317/2001* | 13 Nov 2001 | 22' S | 167°29' E | T. e., T. t., K. | und. | nq s. | Loyalty Channel | F. Navy, <i>La Moqueuse</i> |
| 030/2002* | 30 Jan 2002 | 22°22' S | 166°50' E | T. e. | green col. | nq s. | Uitoe Pass. | F. Navy aerial |
| 030/2002 | 30 Jan 2002 | 22°12' S | 166°01' E | und. | und. | nq s. | Uitoe Pass. | F. Navy aerial |
| 039/2002 | 8 Feb 2002 | 21°26' S | 162°32' E | und. | und. | nq s. | Chesterfields Isl. | R/V <i>Alis</i> |
| 111/2002* | 21 Apr 2002 | 23°29' S | 162°53' E | T. e. | orange col. | Shipb. | SW NC | F. Navy, J. Cartier |
| 111/2002* | 24 Apr 2002 | 22°28' S | 169°12' E | T. e. | pink col. | Shipb. | E Walpole Isl. | F. Navy, J. Cartier |
| 189/2002* | 8 Jul 2002 | 19°08' S | 176°41' W | T. e. | grey col. | Shipb. | Fiji Isl. | F. Navy, J. Cartier |
| 197/2002 | 16 Jul 2002 | 23°03' S | 164°42' E | und. | und. | Shipb. | W NC | F. Navy, <i>J. Cartier</i> |
| 344/2002* | 10 Dec 2002 | 18°40' S | 168°59' E | K. + T. e. + T. t. | large col. | nq s. | Erromango Isl. | F. Navy, J. Cartier |
| 345/2002 | 11 Dec 2002 | 20°05' S | 167°30' E | und. | und. | Shipb. | E Lifou Bay | visual obs |
| 346/2002* | 12 Dec 2002 | 19°19' S | 164°51' E | T. e. + K. | green, grey, pummices | Shipb. | Surprises Isl. | F. Navy, <i>La Glorieuse</i> |
| 347/2002* | 13 Dec 2002 | 19°07' S | 167°20' E | K. + T. e. + T. t. | large col. | Shipb. | Erromango Isl. | F. Navy, <i>J. Cartier</i> |
| 350/2002* | 16 Dec 2002 | 20°48' S | 170°09' W | T. ten., K. | grey, long thin fil. | Shipb. | Niue Isl. | R/V <i>Alis</i> |
| 350/2002* | 16 Dec 2002 | 20°51' S | 170°33' W | T. ten., K. | grey, long thin fil. | Shipb. | Niue Isl. | R/V <i>Alis</i> |

| | | | | | | | | |
|-----------|-------------|----------|-----------|-------------|---|---------------------|----------------------|-------------------------------|
| 352/2002* | 18 Dec 2002 | 21°11' S | 174°08' W | T. ten., K. | grey, long thin fil. | Shipb. | Niue Isl. | R/V <i>Alis</i> |
| 352/2002* | 18 Dec 2002 | 21°11' S | 173°57' W | T. ten., K. | grey, long thin fil. | Shipb. | Niue Isl. | R/V <i>Alis</i> |
| 359/2002 | 25 Dec 2002 | 20°11' S | 169°04' E | und. | und. | und. | W Tanna Isl. | shipboard |
| 038/2003* | 5 Feb 2003 | 21 °S | 166°30' E | T.e. | Brown col. | Shipb. | E NC Loyalty Channel | R/V <i>Alis</i> (Diapalis 07) |
| 048/2003 | 17 Feb 2003 | 23°14' S | 165°29' E | und. | und. | nq s. | Santo Isl. | F. Navy aerial |
| 051/2003 | 20 Feb 2003 | 15°04' S | 166°30' E | und. | und. | nq s. | W NC | F. Navy aerial |
| 087/2003* | 28 Mar 2003 | 24°44' S | 163°21' E | T. e. | pink col. | Shipb. | S NC | F. Navy, J. Cartier |
| 115/2003 | 25 Apr 2003 | 17°57' S | 168°10' E | und. | und. | Shipb. | Efate Isl. | F. Navy, <i>J. Cartier</i> |
| 303/2003 | 30 Oct 2003 | 14°50' S | 168° E | und. | und. | Shipb. | Bank Isl. | ship of opportunity |
| 308/2003* | 4 Nov 2003 | 17°39' S | 167°33' E | T. e. | red col. | Shipb. | Vanuatu | F. Navy, <i>La Moqueuse</i> |
| 309/2003 | 5 Nov 2003 | 20° S | 166°30' E | und. | und. | Shipb. | Ouvea Isl. | visual obs |
| 343/2003 | 9 Dec 2003 | 20°24' S | 167°51' E | und. | und. | 300 km ² | 500 km Lifou-Vanuatu | R/V <i>Alis</i> |
| 352/2003 | 18 Dec 2003 | 24°03' S | 168°02' E | und. | und. | Shipb. | SE NC | F. Navy, <i>La Glorieuse</i> |
| 017/2004 | 17 Jan 2004 | 23°20' S | 168°20' E | und. | und. | Shipb. | SE NC | F. Navy |
| 023/2004 | 23 Jan 2004 | 20°16' S | 161° E | und. | und. | Shipb. | NW NC | F. Navy |
| 026/2004* | 26 Jan 2004 | 22°11' S | 167°22' E | T. e. | small orange col. | Shipb. | Loyalty Channel | F. Navy, <i>La Glorieuse</i> |
| 029/2004* | 29 Jan 2004 | 20°20' S | 166°07' E | T. e. | Large pink flocks | nq s. | Beautemps-Baupré | R/V <i>Alis</i> (PIL) |
| 030/2004* | 30 Jan 2004 | 20°17' S | 166°17' E | T. e. | Large pink flocks | nq s. | NE NC | R/V <i>Alis</i> (PIL) |
| 032/2004* | 1 Feb 2004 | 21°20' S | 167°35' E | T. e. | Large pink flocks | nq s. | NE NC, Oua Isl. | R/V <i>Alis</i> (PIL) |
| 043/2004 | 12 Feb 2004 | 18°30' S | 166°30' E | und. | und. | nq s. | W Vanuatu | F. Navy aerial |
| 044/2004 | 13 Feb 2004 | 19°50' S | 169°54' E | und. | und. | Shipb. | N NC Balabio Isl. | French Navy |
| 047/2004 | 16 Feb 2004 | 18°49' S | 168°30' E | und. | und. | km s. | 500 km NC-Vanuatu | Air Vanuatu |
| 059/2004* | 28 Feb 2004 | 21°10' S | 168°30' E | T. e. | orange col. | Shipb. | NE NC coast | R/V <i>Alis</i> (Motevas) |
| 059/2004* | 28 Feb 2004 | 20°52' S | 165°57' E | T.e. | brown col. | Shipb. | NE NC coast | R/V <i>Alis</i> (Motevas) |
| 060/2004* | 29 Feb 2004 | 18°33' S | 166°03' E | T. t. + K. | floating flocks green tufts long fil. | Shipb. | Vanuatu | R/V <i>Alis</i> (Motevas) |
| 060/2004* | 29 Feb 2004 | 18°20' S | 166°02' E | T. t. + K. | fil. | Shipb. | Vanuatu | R/V <i>Alis</i> (Motevas) |

| | | | | | | | | |
|-----------|-------------|-----------|------------|----------------------------|------------------------------|--------|--------------------------------|------------------------------|
| 060/2004* | 29 Feb 2004 | 17°23' S | 166°04' E | T. t. + K. | large puffs/tufts, flocks | Shipb. | Vanuatu | R/V <i>Alis</i> (Motevas) |
| 061/2004* | 1 Mar 2004 | 22° S | 167° E | T. e. | White col. | Shipb. | Yate Barrier Reef | F. Navy |
| 064/2004 | 4 Mar 2004 | 20°09' S | 168°43' E | und. | und. | nq s. | Vanuatu Masquilignes Isl. N | F. Navy aerial |
| 089/2004 | 29 Mar 2004 | 17° S | 167° E | und. | und. | nq s. | Vanuatu | Air Vanuatu |
| 117/2004 | 26 Apr 2004 | 20°09' S | 163°01' E | und. | und. | nq s. | Aircraft | F. Navy aerial |
| 275/2004 | 1 Oct 2004 | 21°30' S | 165' E | und. | und. | nq s. | Lifou | F. Navy aerial |
| 316/2004 | 11 Nov 2004 | 18°30' S | 168' E | und. | und. | nq s. | Vanuatu | F. Navy aerial |
| 347/2004 | 12 Dec 2004 | 22°29' S | 169°50' E | und. | und. | nq s. | E Pine Isl. , Walpole Isl. | F. Navy aerial |
| 033/2005* | 2 Feb 2005 | 20° 07' S | 170° W | T. e. +T. t. + T. Cont. | und. | und. | East Tonga Isl. | Hashihima et al., 2010 |
| 040/2005* | 9 Feb 2005 | 18° 02' S | 175° E | T. e. +T. t. + T. Cont. | und. | und. | W Fiji Isl. | Hashihima et al., 2010 |
| 090/2007* | 30 Mar 2007 | 22° 49' S | 169° 59' E | T. e. + K. + T. C. | Floating puffs | Shipb. | Walpole Isl. | R/V <i>Kilo Moana</i> cruise |
| 089/2007* | 8 Apr 2007 | 15° S | 169° 59' W | T. e. + K. + T. C. | Floating puffs | Shipb. | N. Fiji Isl. | R/V <i>Kilo Moana</i> cruise |
| 102/2007* | 12 Apr 2007 | 14° 59' S | 178° 45' E | T. e. + K. + T. C. | Surface Bloom | Shipb. | North East Fiji | R/V <i>Kilo Moana</i> cruise |
| 062/2010* | 3 Mar 2010 | 19°11' S | 166°57' E | T. e. | orange col. | nq s. | Beautemps- Beaupré Reef | F. Navy aerial |
| 062/2010* | 3 Mar 2010 | 19°06' S | 166°07' E | T. e. | orange col. | nq s. | Beautemps- Beaupré Reef | F. Navy, <i>J. Cartier</i> |
| 102/2010 | 12 Apr 2010 | 22°48' S | 165°17' S | und. | und. | Shipb. | W NC | Ship obs |
| 102/2010 | 12 Apr 2010 | 22°06' S | 165°26' E | und. | und. | Shipb. | idem | Ship obs |
| 141/2010* | 21 May 2010 | 15°19' S | 166°30' E | T. e. | orange col. | Shipb. | Sabine Bank | R/V <i>Alis</i> (Geodeva4) |
| 300/2010 | 27 Oct 2010 | 19° S | 165°30' E | und. | und. | Shipb. | W NC | Ship obs |
| 301/2010 | 28 Oct 2010 | 22° S | 167° E | und. | und. | Shipb. | W NC | Ship obs |
| 307/2010* | 3 Nov 2010 | 19°15' S | 166°47' E | T. e. | orange col. | Shipb. | SE Vanuatu | F. Navy, R/V <i>Prairial</i> |
| 307/2010* | 3 Nov 2010 | 19°27' S | 166°52' E | T. e. | orange col. | Shipb. | SE Vanuatu | F. Navy, R/V <i>Prairial</i> |
| 339/2010 | 4 Dec 2010 | 22°12' S | 169°42' E | und. | und. | nq s. | SE NC | F. Navy aerial |
| 339/2010 | 4 Dec 2010 | 21°26' S | 167°19' E | und. | und. | nq s. | W NC | F. Navy aerial |
| 339/2010 | 4 Dec 2010 | 22°12' S | 167°42' E | und. | und. | nq s. | W NC | F. Navy aerial |

1 **Table 2.** Average in situ observations at the surface layer (3 meters Niskin bottle) of filamentous cyanobacteria during the DIAPAZON Program
2 from Tenório (2006), the Motevas cruise, and in surface bucket samples during the transects made on the French Navy ships from New
3 Caledonia to Tr1: Vanuatu, Tr2: Walpole, Tr 3: Vanuatu, Tr 4: Vanuatu, Tr 5: Fiji, Vanuatu, Tr6: Wallis and Futuna and Vanuatu. Tchl *a* =
4 chlorophyll *a* + divinyl chlorophyll *a*; PE = phycoerythrin; chl > 10 = percentage of Tchl *a* associated with the > 10 µm fraction, Cp= attenuation
5 coefficient (m⁻¹). Und: undetermined. * E. J. Carpenter, personal communication, ** 1000 in a slick North of Lifou.

| Yearday | date | cruise | Tchl- <i>a</i> | PE | Chl > 10 | Beam attenuation | Trich. Ab. | Bloom obs |
|--------------|---------------|-------------|-----------------------|-----------------------|----------|---------------------|------------------------------|-----------|
| | DD/MM/YY | name | (mg.m ⁻³) | (mg.m ⁻³) | (%) | (m ⁻¹) | (Trichomes.L ⁻¹) | |
| 105/1998 | 15-Apr-98 | Trichonesia | 0.14 | und. | 20 | und. | 298 (10000 Fiji Isl.)* | Shipboard |
| 295-304/2001 | 22-31 Oct 01 | Diapalis01 | 0.14 | 0.16 | 16 | und. | 411 | Shipboard |
| 344-356/2001 | 10-22 Dec 01 | Diapalis02 | 0.13 | 0.15 | 10 | 0.072 | 474 | no |
| 15-22/2002 | 15-22 Jan 02 | Diapalis03 | 0.1 | 0.09 | 8.8 | 0.062 | 401 | no |
| 92-99/2002 | 2-9 Apr 02 | Diapalis04 | 0.11 | 0.1 | 9 | 0.059 | 347 | no |
| 133-144/2002 | 13-14 May 02 | Transect 1 | 0.26 | 0.83 | und. | und. | und. | no |
| 141-148/2002 | 21-28 May 02 | Diapalis05 | 0.11 | 0.23 | 9.3 | 0.046 | 300 (1000 **) | no |
| 217-224/2002 | 5-12 Aug 02 | Diapalis06 | 0.22 | 0.19 | 9 | 0.069 | 76 | no |
| 34-43/2003 | 3-12 Feb 03 | Diapalis07 | 0.23 | 0.6 | 53 | 0.1 | > 6000 | Shipboard |
| 59/2003 | 28-Feb-03 | Transect 2 | 0.29 | 0.45 | und. | und. | und. | und. |
| 71-80/2003 | 12-21 Mar 03 | Transect 3 | 0.29 | 0.33 | und. | und. | und. | und. |
| 107-115/2003 | 17-25 Apr 03 | Transect 4 | 0.21 | 0.53 | und. | und. | und. | no |
| 112-115/2003 | 22-25 Apr 03 | Transect 5 | 0.18 | 0.44 | und. | und. | und. | no |
| 161-165/2003 | 9-14 Jun 2003 | Diapalis08 | 0.2 | 0.25 | 7 | 0.051 | 300 | no |
| 188-208/2003 | 7-27 Jul 03 | Transect 6 | 0.25 | 0.09 | und. | und. | und. | no |
| 281-289/2003 | 8-16 Oct 03 | Diapalis09 | 0.099 | 0.08 | 9 | 0.057 | 9 | no |
| | 28-29 Feb 04 | Motevas | 0.48 | 0.37 | 70 | und. | 4000 | Shipboard |
| 59-60/2004 | (Niskin) | | | | | | | |
| | 28-29 Feb 04 | Motevas | 0.84 | 3.4 | 70 | und. | 6500 | Shipboard |
| 59-60/2004 | (Bucket) | | | | | | | |

6

FIGURE LEGENDS

Fig. 1. (a) Situation map showing limits of the Western Pacific ocean (WP: 25°N-25°S/160°E-160°W) and of the Southern Pacific Ocean box (SP: 5°S-25°S/150°E-190°E/170°W). (b) Mean SeaWiFS composite image of chlorophyll a (mg m^{-3}) in the South Western Tropical Pacific Ocean in February 2003 (austral Summer) with observations of *Trichodesmium* surface accumulations for December 2002-March 2003. Main lands are indicated in yellow (New Caledonia, 20-22°S, 165°E) , Vanuatu (15-20°S, 168°E), Fiji (17°S, 180°E) and South Tonga Islands (21°S, 175°W). (c) French Navy observations of *Trichodesmium* surface blooms (1998-2010) from Table 1 as empty black circles, with: light blue - aerial observations; red - small form filaments (named *T. erythraeum* spp. after Tenório, 2006); magenta - mix of small *T. erythraeum* and long forms (filaments of *Trichodesmium thiebautii* and *T. tenue*); yellow - mix of *T. tenue* and *Katagnymene* sp.

Fig. 2. Selected pictures of the January-February 2004 *Trichodesmium* slick as seen from long line airplanes (aerial observation), and from the ship of the French Navy La Glorieuse (shipboard observation with bucket sample) gathered at Table 1.

Fig. 3 (a) Descriptive parameters to characterize the SeaWiFS radiance anomaly spectra relative to K490 for *Trichodesmium* detection in surface oceanic waters: S = slope of the linear fit; Yt = ordinate of the second degree fit for which tangent is parallel to linear fit; “Bump” and “Trough” are the wavelengths of the major positive and major negative deviations relative to the second degree polynomial fit (largest “bump” or largest “trough”, respectively) (b) Ranges of S and Yt for all RAS of all pixels (black). In orange, specific S and Yt and positions of bumps and troughs schematically shown for RAS of a *Trichodesmium* bloom.

Fig. 4. Application of the TRICHOSAT algorithm to satellite data (1997 to 2010) over the large box corresponding to the Western Pacific Ocean (WP: 25°N-25°S/160°E-160°W) for the interseason (April-May and October) (a) all pixels for which $\text{Chl}_a > 0.2 \text{ mg m}^{-3}$; (b) pixels satisfying the slope and intercept criteria of the RAS; (c) pixels satisfying the shape criteria, (d) pixels satisfying all criteria. The main islands of New Caledonia at 20°S, Vanuatu at 15°S, Fiji Islands at 17°S, Hawaii Islands at 20°N are indicated in black Fig. 4d.

Fig. 5. Statistics of the number of SeaWiFS Level_2 GAC images used to determine the spatio-temporal distribution of *Trichodesmium* accumulations in the SP between 1997 and 2010 by year and season. Seasons are defined as: austral summer (November to March); austral winter (June to September); interseason (April-May and October);

Fig. 6. Application of the TRICHOSAT algorithm to satellite data (1997 to 2010) over the large box corresponding to the Western Pacific Ocean (WP: 25°N-25°S/160°E-160°W); **a**): austral winter (June to September); **b**) interseason (April-May and October); **c**) austral summer (November to March). The main islands of New Caledonia at 20°S, Vanuatu at 15°S, Fiji Islands at 17°S, Hawaii Islands at 20°N are indicated in grey.

Fig. 7. (top) Monthly percentage of valid pixels identified as *Trichodesmium* when applying the TRICHOSAT algorithm to SeaWiFS Level-2 imagery (4 km x 4 km). Results are shown for the Southern Pacific Ocean box (SP: 5°S-25°S, 150°E-190°E/170°W) in red and for the WP (WP: 25°N-25°S/160°E-160°W) as black. *Trichodesmium* blooms occur regularly during the austral summer and peak in February. SeaWiFS chlorophyll-*a* for SP also shown as green line (note that the monthly chlorophyll is smoothed compared to chlorophyll concentrations in pixels of individual level2-GAC images) (bottom) GSM-derived ACDM absorption coefficient (m^{-1}) (grey line) and scaled particulate backscattering coefficient (m^{-1}) (dark blue line).

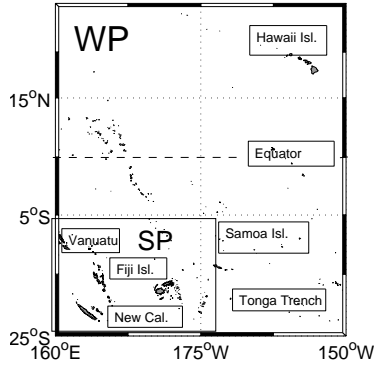
Fig. 8. **a**) Histogram representing the mean annual cycle of *Trichodesmium* bloom pixels percentage (with standard deviation) over the 12 years (1998-2010) of the SeaWiFS acquisition period; **b**) Histogram representing the mean annual number of in situ slicks observations (Nb, from Table 1).

Fig. 9. Spatial distributions of *Trichodesmium* bloom pixels retrieved with TRICHOSAT in the SP box (5°S- 25°S 160°-170°W) for short periods **(a)** during Austral summer, 10- 22 February 2003 (In Yellow: 2003, 40 (9 Feb), in Magenta: 2003, 42 - 43 (11-12 Feb), in Blue: 2003, 44 - 45 (13-14 Feb), in Red : 2003, 47 - 48 (16-17 Feb), in Green: 2003, 49 - 50 (18-19 Feb) Black: 2003, 52 (21 Feb) and **(b)** during Austral winter, 9 - 15 June, 2003 (Magenta: 2003, 160 (June 9) Blue: 2003, 162 (June 11) Red: 2003, 163 (June 13) Green: 2003, 165 (June 15). The two aerial observations from Table 1 are added. Each pixel represents a surface

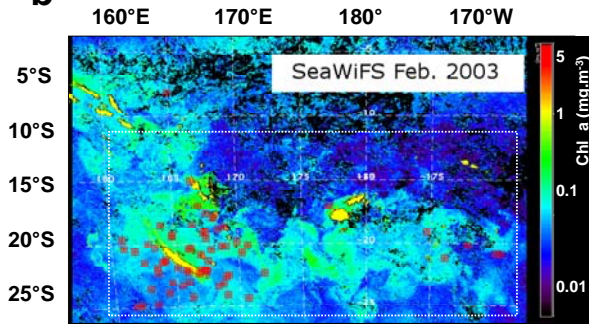
of 16 km² (the size of colored squares is equivalent to a surface of 256 km², i.e. 16"real" SeaWiFS pixels of 4x4 km). Also shown are results obtained from [Westberry et al. \(2005\)](#) (black crosses) for the same time periods. In grey, the main lands of New Caledonia at 20°S, Vanuatu at 15°S and Fiji Islands at 17°S.

Fig. 10. Daily evolution of the TRICHOSAT retrievals percentage shown as red bars (in % pixels, over the SP domain) from individual SeaWiFS Level_2 GAC images, and number of surface slicks observed per month (Nb issued from Table 1) as black open circles in the SP area for the period 1998-2010.

a Figure 1



b



c

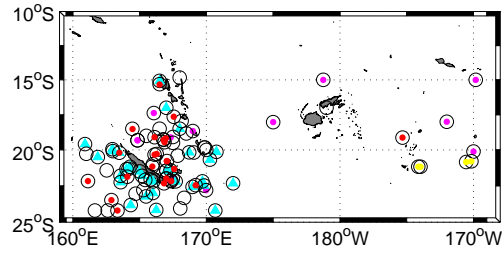


Figure 2

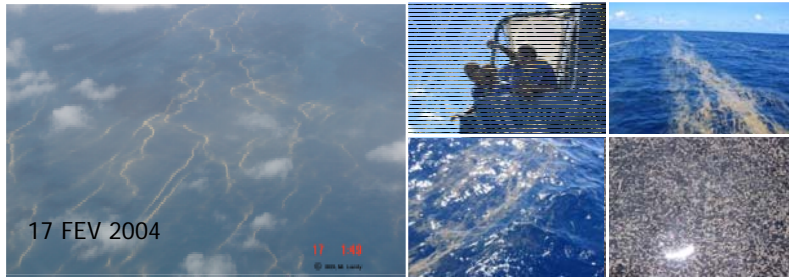


Figure 3ab

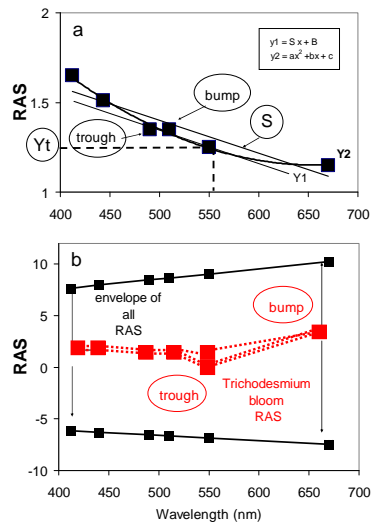


Figure 4abcd

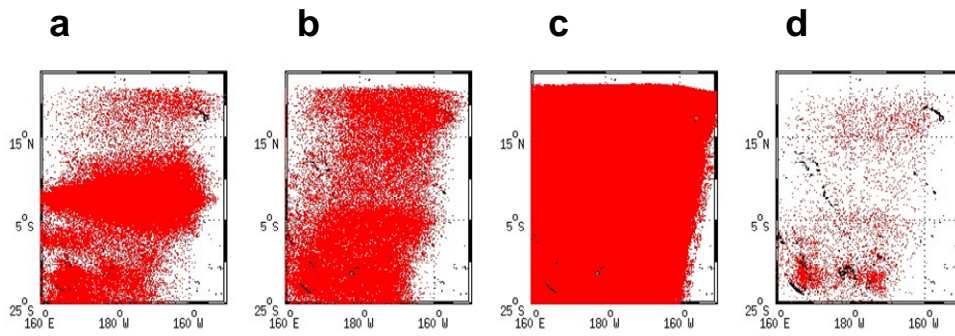


Figure 5

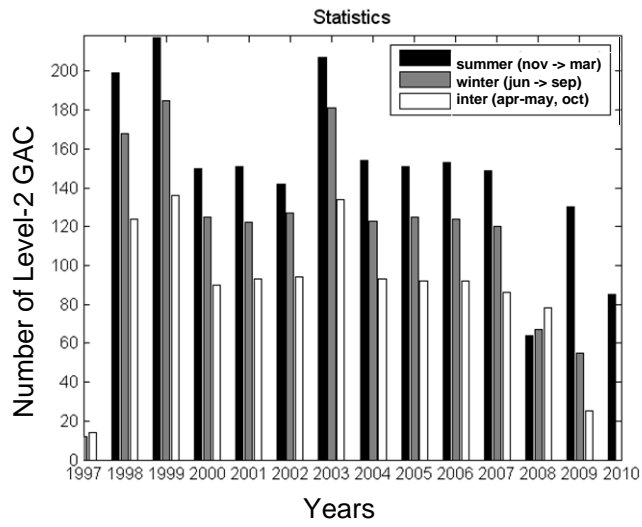


Figure 6

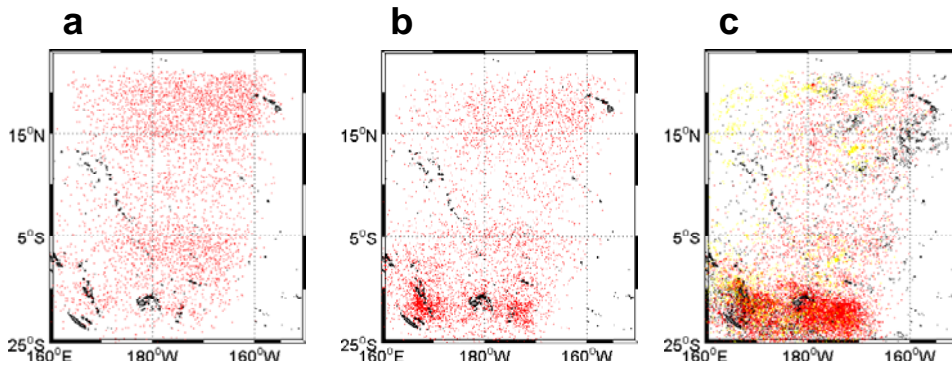
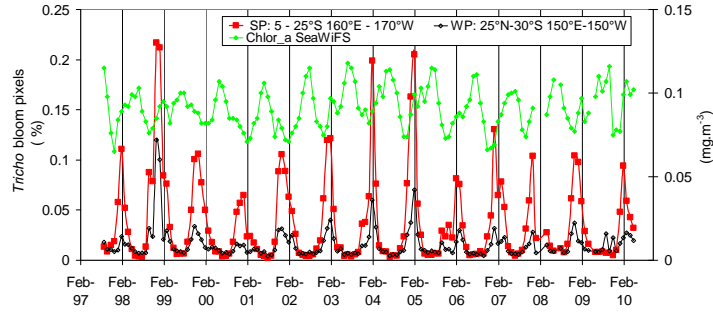


Figure 7

(top)



(bottom)

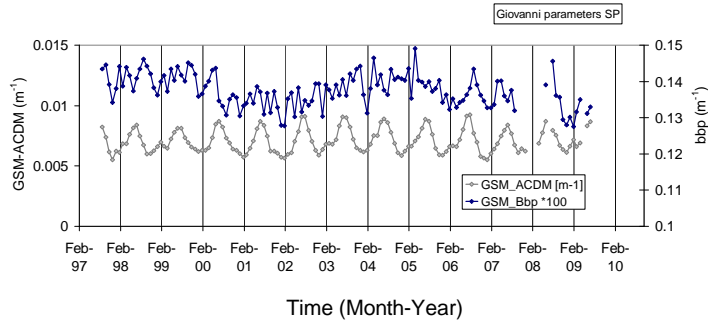
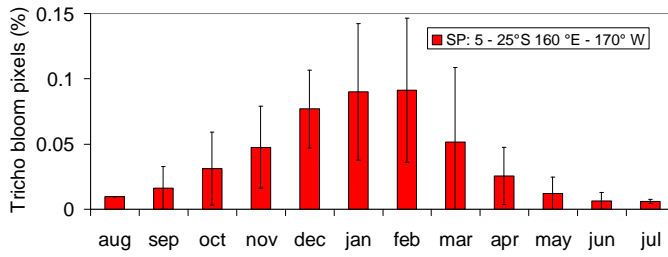
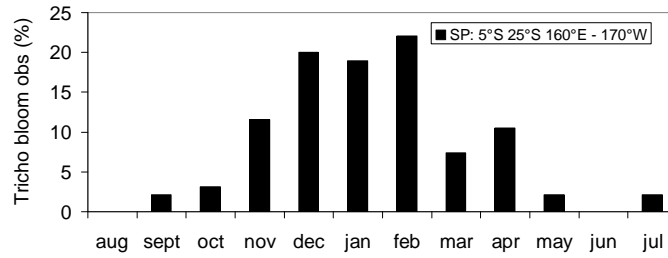


Figure 8

a



b



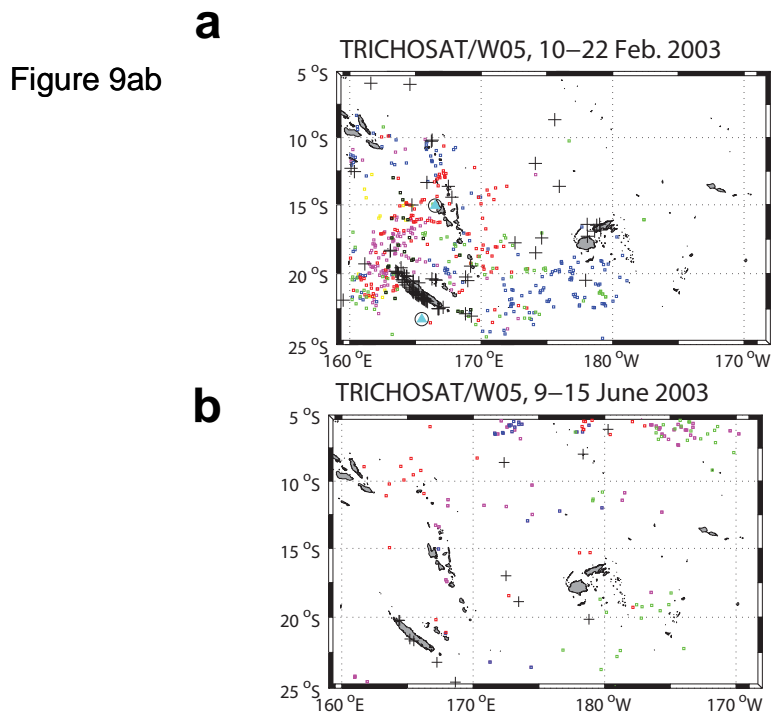


Figure 9ab

