



HAL
open science

In situ application of stir bar sorptive extraction as a passive sampling technique for the monitoring of agricultural pesticides in surface waters

A. Assoumani, S. Lissalde, C. Margoum, Nicolas Mazzella, Marina Coquery

► To cite this version:

A. Assoumani, S. Lissalde, C. Margoum, Nicolas Mazzella, Marina Coquery. In situ application of stir bar sorptive extraction as a passive sampling technique for the monitoring of agricultural pesticides in surface waters. *Science of the Total Environment*, 2013, 463-464, p. 829 - p. 835. 10.1016/j.scitotenv.2013.06.025 . hal-00873137

HAL Id: hal-00873137

<https://hal.science/hal-00873137>

Submitted on 15 Oct 2013

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

In situ application of stir bar sorptive extraction as a passive sampling technique for the monitoring of agricultural pesticides in surface waters

Azziz Assoumani^{1*}, Sophie Lissalde², Christelle Margoum¹, Nicolas Mazzella², Marina Coquery¹

¹Irstea, UR MALY, 5 rue de la Doua, CS70077, F-69626, Villeurbanne Cedex, France

²Irstea, UR REBX, 50 Avenue de Verdun, Gazinet, 33612 Cestas Cedex, France

*Corresponding author: Tel: 00 33 4 72 20 10 86; Fax: 00 33 4 78 47 78 75; Email address: azziz.assoumani@irstea.fr

Abstract

Grab sampling and automated sampling are not suitable or logistically too constraining for the monitoring of pesticides in dynamic streams located in agricultural watersheds. In this work, we applied stir bar sorptive extraction (SBSE) Twisters® directly in two small rivers of a French vineyard (herein referred to as “passive SBSE”), for periods of one or two weeks during a month, for the passive sampling of 19 agricultural pesticides. We performed qualitative and semi-quantitative comparisons of the performances of passive SBSE firstly to automated sampling coupled to analytical SBSE, and secondly to the polar organic chemical integrative sampler (POCIS), a well-known passive sampler for hydrophilic micropollutants. Applying passive SBSE in river waters allowed the quantification of more pesticides and in greater amounts than analytical SBSE as shown for samples collected concurrently. Also, passive SBSE and POCIS proved to be complementary techniques in terms of detected molecules; but only passive SBSE was able to integrate a concentration peak triggered by a quick flood event that lasted 5 hours. Passive SBSE could be an interesting tool for the monitoring of moderately hydrophobic to hydrophobic organic micropollutants in changing hydrosystems. In this purpose, further studies will focus on the accumulation kinetics of target pesticides and the determination of their sampling rates.

Keywords: Passive sampling, Field study, SBSE, POCIS, Pesticides, Water monitoring

30 **1. Introduction**

31

32 Monitoring of organic micropollutant contamination in surface waters has become a
33 challenging issue in Europe since the Water Framework Directive (European Commission,
34 2000), which general aim is to improve and protect European water quality. The evaluation of
35 the chemical quality of surface waters requires reliable measurements of concentrations for
36 priority pollutants including several pesticides (Coquery et al., 2005). Floods are a major
37 pathway for the transport of pesticides in surface waters located in agricultural watersheds and
38 can induce concentration peaks that can vary over several orders of magnitude (Rabiet et al.,
39 2010). In dynamic hydrosystems like small rivers located in agricultural watersheds, flood-
40 induced concentration peaks can be very quick (down to a few hours); therefore low
41 frequency grab sampling is not suitable for the monitoring of the pesticide contamination.
42 Moreover, this sampling technique does not allow the determination of ultra trace levels of
43 some fungicides, which may have an impact on aquatic microbial life (Artigas et al., 2012).
44 Numerous grab samples or automated samples are necessary to assess the time variability of
45 the contamination. Moreover, these sampling techniques would trigger a large number of
46 analyses, and the use and maintenance of an automated sampler are costly.

47 Passive (or integrative) sampling has recently been developed as an alternative to grab or
48 automated sampling in order to obtain, at lower cost, more realistic estimates of the average
49 concentrations of micropollutants in surface waters (Greenwood et al., 2007; Namieśnik et al.,
50 2005; Vrana et al., 2005). In addition, the passive accumulation of chemicals from large
51 volumes of water results in ultra trace level detection and smoothed integrative sampling over
52 periods ranging from days to months. Passive samplers can monitor a broad range of
53 micropollutants, depending on their physical chemical properties. The polar organic chemical
54 integrative sampler (POCIS) is one of the main devices used for the passive sampling of the

55 moderately polar organic compounds (Alvarez et al., 2004). Its efficiency for the
56 determination of time-weighted average (TWA) concentrations of hydrophilic pesticides in
57 natural waters has been reported in the literature (Greenwood et al., 2007; Vrana et al., 2005).
58 Moreover, numerous studies have been dedicated to the passive sampling of hydrophobic
59 organic micropollutants such as polycyclic aromatic hydrocarbons (PAH),
60 polychlorobiphenyl (PCB) congeners, and highly hydrophobic organochlorine and
61 organophosphate pesticides (Allan et al., 2012; Booij et al., 2002; Jahnke et al., 2008; Paschke
62 et al., 2006; Prokeš et al., 2012; Stuer-Lauridsen, 2005; Vrana et al., 2001). To our
63 knowledge, however, the sampling of moderately hydrophobic to hydrophobic pesticides ($2 <$
64 $\log K_{ow} < 5$) is poorly documented.

65 Stir bar sorptive extraction (SBSE) is a solvent free sample preparation technique dedicated to
66 moderately hydrophobic to hydrophobic compounds in aqueous and gaseous samples. The
67 extraction device, named Twister®, is composed of a magnet enclosed in a glass tube coated
68 with a thick film of polydimethylsiloxane (PDMS) (Baltussen et al., 1999a). The extraction is
69 performed with a Twister by immersion in the aqueous sample (SBSE) or by headspace
70 sampling (headspace sorptive extraction, HSSE) (Baltussen et al., 1999a; Tienpont et al.,
71 2000). This novel sample treatment technique has been successfully used for the analytical
72 extraction of several compounds, such as hormones, pesticides, PAH and PCB in air, soil, and
73 various liquid matrices (David and Sandra, 2007; Prieto et al., 2010). The application of
74 SBSE on site has been reported in the literature, but only for the analysis of PAH (Roy et al.,
75 2005).

76 In this work, we applied SBSE directly *in situ* as a passive sampling technique for the
77 monitoring of fugacious agricultural pesticides in dynamic streams (herein named “passive
78 SBSE”). For this purpose, we first compared the performances of passive SBSE and
79 automated sampling coupled with analytical SBSE, i.e., the extraction with Twisters of water

80 samples collected concurrently in a French river located in an agricultural watershed and
81 performed in the laboratory. Secondly, we compared the qualitative and semi-quantitative
82 performances of the passive SBSE and the POCIS during base flow and a flood event of a
83 second dynamic stream located in the same watershed.

84

85 **2. Experimental**

86

87 **2.1 Chemicals and materials**

88

89 The 19 pesticides selected for this study belong to different chemical classes (herbicides,
90 insecticides, and fungicides) and have different physical chemical properties, such as their
91 octanol-water partitioning coefficient $\log K_{ow}$ (Table 1). They were provided by Dr.
92 Ehrenstorfer GmbH (Augsburg, Germany): acetochlor, atrazine, azoxystrobin,
93 chlorfenvinphos, chlorpyrifos-ethyl, diflufenican, dimethomorph, diuron, 3,4-dichloroaniline
94 (metabolite of diuron), 1-(3,4-dichlorophenyl)-3-methyl urea (metabolite of diuron),
95 fenitrothion, flufenoxuron, isoproturon, metolachlor, norflurazon, procymidon, simazine,
96 spiroxamine, and tebuconazole (purity $\geq 92.5\%$). For chemical analyses, atrazine-d5,
97 chlorpyrifos-ethyl-d10, diuron-d6, isoproturon-d6, and metolachlor-d6, used as internal
98 standard or surrogate, were also provided by Dr. Ehrenstorfer (purity $\geq 98.5\%$).

99 For both passive SBSE and analytical SBSE techniques, LC-MS grade acetonitrile and
100 methanol, and dichloromethane for pesticide residue analysis were purchased from VWR
101 (Strasbourg, France). Formic acid (purity = 98%) for LC-MS analysis was provided by
102 Fischer Bioblock (Illkirch, France). Ultrapure water was produced by a MilliQ water
103 purification system purchased from Millipore (Billerica, MA, USA). The Twisters (20 mm x

104 1-mm thick PDMS film, with an external surface area of 2.1 cm² and a PDMS phase volume
105 of 126 µL) were purchased from Gerstel (Mülheim a/d Ruhr, Germany).
106 For the POCIS technique, all solvents (HPLC grade) were obtained from Sharlau (Sentmenat,
107 Spain) except ethyl acetate, which was purchased from Fluka (St. Louis, MO, USA).
108 Ultrapure water was produced by a Synergy UV system from Millipore (Billerica, MA, USA).
109 All eluents were filtered through 0.45 µm regenerated cellulose filters from Whatman
110 (Versailles, France). Ammonium acetate was purchased from Fluka (St Louis, MO, USA).
111 POCIS (Alvarez et al., 2004; Mazzella et al., 2007) contains about 200 mg of Oasis HLB
112 sorbent, purchased from Waters (St Quentin-en-Yvelines, France), weighted with accuracy
113 and enclosed between two hydrophilic polyethersulfone (PES) SUPOR 100 membrane disc
114 filters (0.1 µm, 90 mm membrane diameter), purchased from Pall (Saint-Germain-en-Laye,
115 France). The total exchanging surface area of the membrane (both sides) is approximately 41
116 cm² and the surface area per mass of sorbent ratio is approximately 200 cm² g⁻¹.

117

118 2.2 Field experiments

119

120 The passive samplers were deployed in two rivers of a French vineyard watershed located
121 about 70 km north of Lyon in the Beaujolais region, the Ardières and the Morcille Rivers.
122 Two deployment sites -one per river- were selected for a one-month exposition campaign. For
123 the comparison of the performances of passive SBSE and analytical SBSE, Twisters were
124 immersed in triplicates for 4 periods of one week in the Morcille River (herein named
125 “passive Twisters”). During the same period, passive Twisters and POCIS were deployed,
126 both in triplicates, for two periods of two weeks in the Ardières River. The passive Twisters
127 were placed in deployment bags, made of two pieces of plastic mesh, in order to expose the
128 PDMS phase directly to the aquatic medium, and protect it from small rocks, pieces of wood

129 or coarse sand. The POCIS orientation was vertical with the PES membranes perpendicular to
130 the water surface and the flow (Mazzella et al., 2010). The two passive samplers were placed
131 in the same cages for deployment in the rivers. Field blanks for passive Twisters and POCIS
132 were systematically used.

133 Simultaneously, at both sites, weekly time-averaged water samples were collected with a
134 refrigerated automated sampler (Bühler 4010, Hach-Lange) in amber glass bottles. The water
135 samples and the passive Twisters were brought to the laboratory in Lyon for chemical
136 analysis, whereas the POCIS were sent in an isothermal case to the laboratory in Bordeaux for
137 the determination of the pesticide concentrations.

138

139 2.3 Chemical analysis of water samples and passive Twisters

140

141 The pesticide concentrations of the Ardières River water samples were determined by solid
142 phase extraction (6-mL Oasis HLB cartridges, Waters) followed by liquid chromatography
143 coupled with tandem mass spectrometry (SPE-LC-MS/MS). For the Morcille River water
144 samples, pesticide concentrations were determined by analytical SBSE followed by liquid
145 desorption and liquid chromatography coupled with tandem mass spectrometry (SBSE-LD-
146 LC-MS/MS). The development and the validation of the extraction of the selected pesticides
147 by SBSE and analysis by LC-MS/MS have been published elsewhere (Margoum et al., 2013).
148 Briefly, the extraction was performed at 800 rpm for 3 hours on 20 mL of the weekly
149 averaged water samples filtered with 0.7 μm GF/F glass fiber membranes. The Twisters
150 (herein named “analytical Twisters”) were then placed in 200 μL of methanol/acetonitrile
151 (50/50, v/v), and the pesticides were desorbed under sonication for 15 min. Finally, 150 μL of
152 ultrapure water and 10 μL of diuron-d6 at 200 $\mu\text{g L}^{-1}$, in acetone, were added to 40 μL of the
153 desorbate to constitute the sample for LC-MS/MS analysis.

154 After exposure, the passive Twisters were taken out of their deployment bags, gently rinsed
155 and dried, then placed overnight at -18 °C. Afterwards, the pesticides sorbed in the passive
156 Twisters were extracted the same way as for the analytical Twisters.

157 The chemical analyses were performed with an LC 1100 Series apparatus from Agilent
158 (Massy, France) coupled with a MS triple quadrupole API 4000 from AB Sciex (Les Ulis,
159 France), equipped with an electrospray ionization source (ESI) that was operated in the
160 positive ionization mode. An Atlantis T3 (2.1 mm x 100 mm; $d_p = 3 \mu\text{m}$) purchased from
161 Waters (St Quentin-en-Yvelines, France) was used for the chromatographic separation of the
162 analytes. Acetonitrile and ultrapure water both with formic acid (0.1%) were used in an
163 analytical gradient of 15 min.

164

165 2.4 Recovery from POCIS and extract analysis

166

167 Full details of the treatment of the POCIS and the analysis of pesticides performed after
168 exposure can be found elsewhere (Lissalde et al., 2011; Mazzella et al., 2010). Briefly, the
169 POCIS was open and the sorbent was transferred into a 3-mL empty SPE tube with a PE frit
170 and packed under vacuum by using a Visiprep SPE Manifold. Analytes were eluted with 3
171 mL of methanol, then with 3 mL of a methanol:ethyl acetate mix (75:25, v/v). The solvent
172 was then evaporated under a gentle stream of nitrogen for 30 min. Finally, the dried extract
173 was dissolved in 1 mL of the injection solvent (ultrapure water:acetonitrile, 90:10, v/v) for
174 LC-MS/MS analysis.

175 An HPLC Ultimate 3000 apparatus from Dionex (Voisin Le Bretonneux, France) was used
176 (solvent rack SRD-3600 6 degasser channels, DGP-3600M pump, WPS-3000 TSL Micro
177 autosampler, TCC-3100 HP 1xRH 2P-6P thermostated column oven). Acetonitrile and 5 mM
178 ammonium acetate solution were used with an analytical gradient of 15 min.

179 Chromatographic separation was performed with a Gemini-NX C18 3 μm , 110 \AA , 2.0 mm x
180 100 mm with a SecurityGuard cartridge Gemini-NX C18 2.0 mm x 4 mm, both from
181 Phenomenex (Le Pecq, France). The detector was an API 2000 triple quadrupole mass
182 spectrometer from AB Sciex. It was equipped with an ESI source operated in the positive
183 ionization mode.

184

185 3. Theory and modeling

186

187 3.1 Stir Bar Sorptive Extraction

188

189 Stir Bar Sorptive Extraction is a sampling technique governed by diffusion of the analytes.
190 This technique relies on equilibrium, and the extraction of solutes from the aqueous samples
191 into the extraction phase is controlled by the partitioning coefficient of the solutes between
192 the PDMS phase and the aqueous phase (K_{sw}) (Baltussen et al., 2002; David and Sandra,
193 2007). Baltussen et al. (1999a, 1999b) have correlated the partitioning coefficient of
194 hydrophobic analytes (K_{sw}) with their octanol-water distribution coefficient (K_{ow}). The
195 partitioning coefficient of a compound is linked to the concentration in the Twister and the
196 concentration in the water sample (Eq. 1):

$$K_{ow} \approx K_{sw} = \frac{C_s}{C_w} = \frac{m_s}{m_w} \times \frac{V_w}{V_s} = \frac{m_s}{m_w} \times \beta \quad (1)$$

197

198 where C_w ($\mu\text{g L}^{-1}$) is the concentration of analyte in the water sample at equilibrium; C_s ($\mu\text{g L}^{-1}$)
199 is the concentration of analyte in the extraction phase at equilibrium; m_w (μg) is the mass of
200 analyte remaining in the water sample; m_s (μg) is the mass of analyte in the extraction phase;
201 V_w (L) is the volume of water sample; V_s (L) is the volume of the extraction phase; β
202 (adimensional) is the phase ratio.

203 The extraction recovery η is expressed as the ratio of the mass of analyte in the extraction
204 phase (m_s) over the initial mass of analyte in the water sample ($m_0 = m_s + m_w$). It is
205 determined by the partitioning coefficient K_{sw} and by the phase ratio β , as described in Eq. 2.

$$\eta = \frac{m_s}{m_0} = \frac{K_{sw}/\beta}{1 + (K_{sw}/\beta)} \quad (2)$$

206
207 From Eq. 2, it is easily deduced that the extraction recovery increases with K_{sw} . Since K_{sw} is
208 approached by K_{ow} (Eq. 1), extraction recovery on PDMS, in general, decreases with
209 increasing polarity. Moreover, the phase ratio β can also affect the extraction recovery. When
210 the volume of the PDMS extraction phase is increased, β decreases, and the extraction
211 efficiency increases.

212

213 3.2 Passive sampling

214

215 The mass transfer of an analyte in a sampler includes several diffusion and interfacial
216 transport steps across all barriers, i.e., the stagnant aqueous boundary layer, possible biofilm
217 layer, the membrane and then, the receiving phase. Assuming isotropic exchange, the
218 corresponding uptake in the sampler over time with constant ambient concentration can be
219 described as follows (Eq. 3):

$$N(t) = M_s K'_{sw} C_w (1 - \exp(-k_e \cdot t)) \quad (3)$$

220

221 where N (μg) is the mass of analytes accumulated in the receiving phase; M_s (g) is the mass of
222 the receiving phase; K'_{sw} (mL g^{-1}), described by the ratio of the concentration of analytes in
223 the sampler C'_s ($\mu\text{g g}^{-1}$) and the concentration of the analytes in the water phase C_w ($\mu\text{g mL}^{-1}$),
224 is the receiving phase/water partitioning coefficient; and t (d) equals time.

225 The elimination constant k_e (d^{-1}) is defined as follows (Eq. 4):

$$k_e = \frac{R_s}{K'_{sw} M_s} = \frac{\lambda A}{K'_{sw} M_s} \quad (4)$$

226

227 where R_s is the sampling rate (mL d^{-1}) and A (cm^2) is the sampler surface area.

228 The overall mass transfer coefficient λ (cm d^{-1}) describes the movement of the analytes out of
229 the bulk solution, across multiple barriers, to the receiving phase. The overall resistance ($1/\lambda$)
230 is given by sum of all particular barrier resistances:

$$\frac{1}{\lambda} = \frac{\delta_w}{D_w} + \frac{\delta_b}{D_b K_{bw}} + \frac{\delta_m}{D_m K_{mw}} + \frac{\delta_s}{D_s K_{sw}} \quad (5)$$

231

232 with δ the thickness of the particular barrier, D_w the diffusion coefficient of the analyte in
233 water (i.e., stagnant aqueous boundary layer), D_i the diffusion coefficient of the analyte in the
234 i th barrier (i.e., biofilm, membrane, etc.), and K_{iw} the partitioning coefficient between water
235 and the i th barrier or receiving phase (in subscripts, w stands for the water, b the possible
236 biofouling, m the membrane, and s the receiving phase).

237 In passive SBSE, the receiving phase/water partitioning coefficient K_{sw} is adimensional,
238 since it is described by the ratio of the concentration in the Twisters C_s ($\mu\text{g mL}^{-1}$) and the
239 concentration in the water phase C_w ($\mu\text{g mL}^{-1}$). With POCIS, K'_{sw} (mL g^{-1}) is used, since the
240 receiving phase is a powder and the concentration of analytes in the sampler C'_s is expressed
241 in $\mu\text{g g}^{-1}$. For comparison of passive SBSE with the POCIS, one can convert K_{sw} into K'_{sw}
242 with the mass density of PDMS ρ_s , which is 1.15 g mL^{-1} according to Rusina et al. (2007), as
243 follows (Eq. 6):

$$K'_{sw} = \frac{K_{sw}}{\rho_s} = \frac{K_{sw}}{M_s} V_s \quad (6)$$

244

245 For POCIS, the overall mass transfer coefficient highly depends on δ_w as it is assumed that
246 the analyte uptakes are mainly under aqueous boundary layer control (Alvarez et al., 2004;
247 Mazzella et al., 2008; Vrana et al., 2005). In passive SBSE, no membrane separates the

248 Twister from the aqueous medium. Hence, the analyte uptakes are limited either by the
249 resistance in the water boundary layer or by the resistance in the receiving phase (PDMS)
250 (Vrana et al., 2006). Eq. 5 shows that the resistance in the receiving phase decreases with
251 increasing K_{sw} value for substances having similar diffusion coefficient D_s in this material.
252 Nevertheless, since K_{sw} is approached by K_{ow} (Eq. 1), there will be a critical K_{ow} value where
253 the analyte uptakes will turn to be controlled by the water boundary layer, likely due to
254 decreasing diffusivity of more hydrophobic molecules, with increasing size/volume. Studies
255 have suggested that uptake control switches from membrane to water boundary layer for
256 compounds with $\log K_{ow}$ values in the range of 4.5 to 5.0, for non polar compound passive
257 samplers with membrane such as semipermeable membrane device (SPMD) (Huckins et al.,
258 2006), membrane enclosed sorptive coating (MESCO) and Chemcatcher, as well as for
259 samplers without membrane such as low density polyethylene (LDPE) membranes and silicon
260 strips (Allan et al., 2009). Therefore, for passive SBSE, we refer to the two kinetic limitations
261 of the compound uptakes as membrane and water boundary layer controls, and we assume
262 that the transition from one mass transfer control to the other occurs for compounds with \log
263 K_{ow} between 4.5 and 5.0.

264

265 **4. Results and discussion**

266

267 4.1 Comparison of the passive SBSE and automated sampling coupled with analytical 268 SBSE

269

270 We first compared the accumulation of 19 target pesticides in the Twisters deployed in the
271 Morcille River (passive Twisters) and in those used for the analytical extraction of weekly
272 averaged river water samples (analytical Twisters), collected at the same site. The masses of

273 pesticides accumulated in either the passive Twisters or analytical Twisters for 4 consecutive
274 periods of one week are presented in Table 1. Firstly, the two techniques showed similar
275 repeatability; the relative standard deviations (RSD) calculated (with $n = 3$) ranged from 4.0
276 to 57.5% for passive SBSE and from 5.7 to 48.9% for analytical SBSE. Secondly, over the 4
277 weeks of the study, passive Twisters accumulated 6 pesticides more frequently than analytical
278 Twisters, including atrazine, diflufenican and chlorpyrifos-ethyl, which were accumulated
279 only by passive Twisters. This was not caused by a difference in limits of quantification
280 (LOQ) since the two techniques reached similar LOQ in ng, estimated with signal-to-noise
281 ratios. For most pesticides quantified by both techniques, accumulation in the passive
282 Twisters was 1.3 to 8 times higher in average. In contrast, dimethomorph, norflurazon and
283 simazine were accumulated in passive Twisters to a lower extent (about 1.6 times less in
284 average). Considering the lower $\log K_{ow}$ values for these compounds (Table 1), we can
285 assume lower $\log K_{sw}$ values (Eq. 1), thus a lower affinity for the PDMS phase. As a result,
286 lower masses of these 3 pesticides in the passive Twisters in comparison with analytical
287 Twisters could be due to desorption phenomena during the one-week exposition periods.
288 Nevertheless, for the most hydrophobic pesticides, our results imply that lower LOQ could be
289 reached by passive SBSE, in agreement with the theory of passive sampling (Greenwood et
290 al., 2007; Huckins et al., 2006).

291

292 4.2 Comparison of passive SBSE and POCIS

293

294 During both two-week exposition periods, passive Twisters and POCIS were deployed
295 simultaneously at the same site, thus they were exposed to the same concentrations of
296 pesticides. After recovery of both samplers, no biofilm was observed. Consequently, we
297 assume that the determination of the masses of pesticides accumulated in the samplers was

298 not biased by biofouling. Moreover, concentrations of the target pesticides in the river water
299 were monitored for both exposition periods by means of automated samplings and SPE-LC-
300 MS/MS analyses. After recovery of the samplers and chemical analyses, we performed
301 qualitative and semi-quantitative comparisons of the two devices. For the qualitative
302 comparison, we focused only on the 7 pesticides quantified in the two passive samplers and in
303 the weekly averaged water samples, i.e., simazine, azoxystrobin, dimethomorph, diuron, 1-
304 (3,4-dichlorophenyl)-3-methyl urea, metolachlor, and chlorpyrifos-ethyl. As shown in Figure
305 1, the most polar compounds (simazine, dimethomorph, azoxystrobin, diuron, and 1-(3,4-
306 dichlorophenyl)-3-methyl urea) were either accumulated only by the POCIS or sorbed by the
307 POCIS to a higher extent than by the passive Twisters. In contrast, the most hydrophobic
308 pesticides (metolachlor and chlorpyrifos-ethyl) were accumulated only by the passive
309 Twisters. Similar results were obtained from passive Twisters and POCIS deployed
310 concurrently in 2 other sites on the same river and one site on the Morcille River (data not
311 shown). Thus, passive SBSE and POCIS could be used as two complementary techniques for
312 the monitoring of a broad range of pesticides in natural waters. This conclusion was quite
313 expected since SBSE is originally an analytical sample preparation technique for moderately
314 hydrophobic to hydrophobic contaminants ($\log K_{ow} > 3$) (Baltussen et al., 1999a, 1999b;
315 David and Sandra, 2007), and POCIS are known to target more hydrophilic contaminants (\log
316 $K_{ow} < 4$) (Alvarez et al., 2007, 2004; Mazzella et al., 2007; Morin et al., 2012). Over the 7
317 pesticides presented in Figure 1, azoxystrobin, simazine and dimethomorph were accumulated
318 in both passive samplers. Interestingly, in the case of simazine, $\log K'_{sw}$ values for SBSE
319 (1.90) deduced from Eqs. 1 and 6, and for POCIS (4.68) available in the literature (Mazzella
320 et al., 2010), indicate the higher affinity of this polar compound for the receiving phase of the
321 POCIS. This comparison is possible only for simazine, because it is the only pesticide for
322 which we have $\log K'_{sw}$ values for both samplers.

323 For semi-quantitative comparison, we chose to normalize the masses of accumulated analytes
324 to the respective device surface areas (2.1 cm^2 for passive Twisters and 41 cm^2 for POCIS), as
325 shown in Figure 1. The normalization of the mass of a pesticide accumulated in the sampler
326 by its surface area is a useful way to approach its overall mass transfer coefficient λ (Eq. 4).
327 The weekly averaged pesticide concentrations, obtained from the automated sampler, were
328 similar during the two exposition periods (Figure 1). But average normalized masses of
329 accumulated pesticides and RSD ($n = 3$) varied between samplers and from one exposition
330 period to the other. Larger RSD for the pesticides accumulated in the POCIS exposed during
331 the first two-week exposition period compared to the second exposition period (7 to 27 times
332 larger) may be attributed to a 5-hour flood event that occurred two days before retrieval of the
333 samplers (on day 12). In other words, high turbulences and a short and brutal change in flow
334 velocity probably made each POCIS of the triplicates accumulate pesticides with a different
335 accumulation rate. Indeed, measurements of the flow rate revealed a two hundred-fold
336 increase at the peak of the flood event (from 11.5 to 2210 L s^{-1} in less than 5 hours). For the
337 same reasons, larger normalized masses of pesticides accumulated in POCIS during the
338 second exposition period (from 1.5 to 2.8 larger) could be attributed to a two-fold increase of
339 the flow rate of the river between the first and the second two-week exposition period
340 (average flow rate increased from 18 L s^{-1} to 37 L s^{-1}). Hence, we assume that the increased
341 flow velocity probably triggered faster chemical accumulations in the POCIS. These
342 observations suggest two distinct behaviors for the devices. Indeed, δ_s , D_s and K'_{sw} of the
343 pesticides studied are different for the two devices (Eq. 5); also, the mass transfer of the solute
344 into the POCIS is controlled by the aqueous boundary layer, i.e., it depends on the
345 hydrodynamic conditions during the exposition (Alvarez et al., 2004; Mazzella et al., 2008).
346 On the other hand, 1.5 to 11 times more simazine, azoxystrobin, dimethomorph, metolachlor
347 and chlorpyrifos-ethyl were quantified in passive Twisters exposed during the first exposition

348 period in comparison to those deployed for the second exposition period. This could be
349 attributed to a quick concentration peak which occurred during the flood event (on day 12)
350 (Rabiet et al., 2010). Response times of the passive Twisters -without membranes- with
351 respect to pesticide accumulation were probably short enough to integrate the concentration
352 peak, which was not integrated by the automated sampler. Moreover, as mentioned in the
353 section 3.2, the mass transfer of simazine, azoxystrobin, dimethomorph, diuron, 1-(3,4-
354 dichlorophenyl)-3-methyl urea, metolachlor, chlorfenvinphos and chlorpyrifos-ethyl into the
355 passive Twisters was probably controlled by the membrane, since their log K_{ow} values are
356 below 5.0. Therefore, unlike the POCIS, the hydrodynamic conditions during this flood event
357 had little impact on the repeatability of the accumulation of the target pesticides into the
358 passive Twisters. For instance, for the first exposition period, the RSD for the normalized
359 masses of pesticides (simazine, azoxystrobin, dimethomorph, metolachlor and chlorpyrifos-
360 ethyl) accumulated in the passive Twisters ranged from 4.1 to 10.5% whereas, for the
361 pesticides accumulated in POCIS (simazine, azoxystrobin, dimethomorph, diuron, and 1-(3,4-
362 dichlorophenyl)-3-methyl urea), RSD ranged from 34.7 to 56.2%. Moreover as a comparison
363 of the two samplers, the RSD of the accumulation of simazine, azoxystrobin and
364 dimethomorph in the passive Twisters were 6 to 8 times lower than those for the same
365 pesticides accumulated in the POCIS.

366 In this study, we focused on the performances of passive SBSE for the sampling of a broad
367 range of pesticides ($2.18 < \log K_{ow} < 5.11$). Several pesticides among those have been
368 targeted by POCIS via laboratory or *in situ* studies: for instance, acetochlor, 1-(3,4-
369 dichlorophenyl)-3-methyl urea, diuron, simazine, atrazine, isoproturon, metolachlor,
370 chlorfenvinphos, fenitrothion, and chlorpyrifos-ethyl (Alvarez et al., 2007, 2005, 2004;
371 Mazzella et al., 2010). Moreover, performances of passive samplers for hydrophobic
372 organochlorine or organophosphate pesticides, such as SPMD, LDPE, silicon rubbers, PDMS

373 membranes or MESCO have been reported in several studies (Allan et al., 2012; Booij et al.,
374 2002; Jahnke et al., 2008; Namieśnik et al., 2005; Paschke et al., 2006; Prokeš et al., 2012;
375 Stuer-Lauridsen, 2005; Vrana et al., 2005, 2001). Nevertheless, to our knowledge, the passive
376 sampling of moderately hydrophobic to hydrophobic pesticides targeted in our study such as
377 spiroxamine, flufenoxuron, diflufenican, tebuconazole, procymidon, dimethomorph,
378 azoxystrobin and norflurazon has not been reported in the literature.

379

380 **5. Conclusion**

381

382 This study focused on the passive sampling of 19 moderately hydrophobic to hydrophobic
383 pesticides in surface waters by passive SBSE. Firstly, results showed that this technique could
384 allow to reach lower LOQ than automated sampling coupled with analytical SBSE for most
385 hydrophobic studied pesticides. Secondly, passive SBSE and POCIS were shown
386 complementary regarding the ranges of polarity for the chemicals targeted. The two
387 techniques, when the masses of accumulated pesticides are normalized to the respective
388 surface areas of the devices, showed, however, different accumulation performances. Thirdly,
389 the passive SBSE and POCIS revealed two different behaviors in changing hydrodynamics,
390 due to different analyte uptake controls. Flow velocity seemed to impact the accumulation of
391 the target pesticides in POCIS only. An additional way to compare these two samplers would
392 be the calculation of time-weighted averaged concentrations using the sampling rates of the
393 target pesticides obtained from kinetic studies and laboratory calibration of the passive
394 Twisters.

395 One of the advantages of the passive SBSE technique is the simple handling, preparation
396 before deployment, *in situ* deployment and sample treatment after exposition of the Twisters.
397 Moreover, in case of analysis by liquid chromatography, passive SBSE is environmentally

398 friendly and cost-effective with respect to solvent consumption, since small solvent volumes
399 (50 to 200 μL) are generally used for the desorption of the chemicals accumulated in the
400 Twisters. In case of thermal desorption and analysis by gas chromatography, passive SBSE
401 seems even more promising regarding the reduction of the use of organic solvents and the
402 improvement of LOQ. Finally, since no membrane separates the Twister from the aquatic
403 medium in passive SBSE, response times to concentration peaks may be shorter than most
404 samplers equipped with a membrane, such as SPMD, POCIS and MESCO. Hence, this first
405 study proves that passive SBSE is an interesting technique for monitoring chemicals in
406 hydrosystems with high concentration variations and needs further work for the determination
407 of sampling rates in order to calculate time-weighted averaged concentrations.

408

409 **Acknowledgements**

410

411 The authors thank C. Guillemain for analytical support, L. Liger, O. Garcia and B. Motte for
412 technical and logistical support; and the French National Research Agency (ANR) through
413 PoToMAC project (ANR 2011 CESA 022 02) and the French National Agency for Water and
414 Aquatic Environments (ONEMA) for financial support.

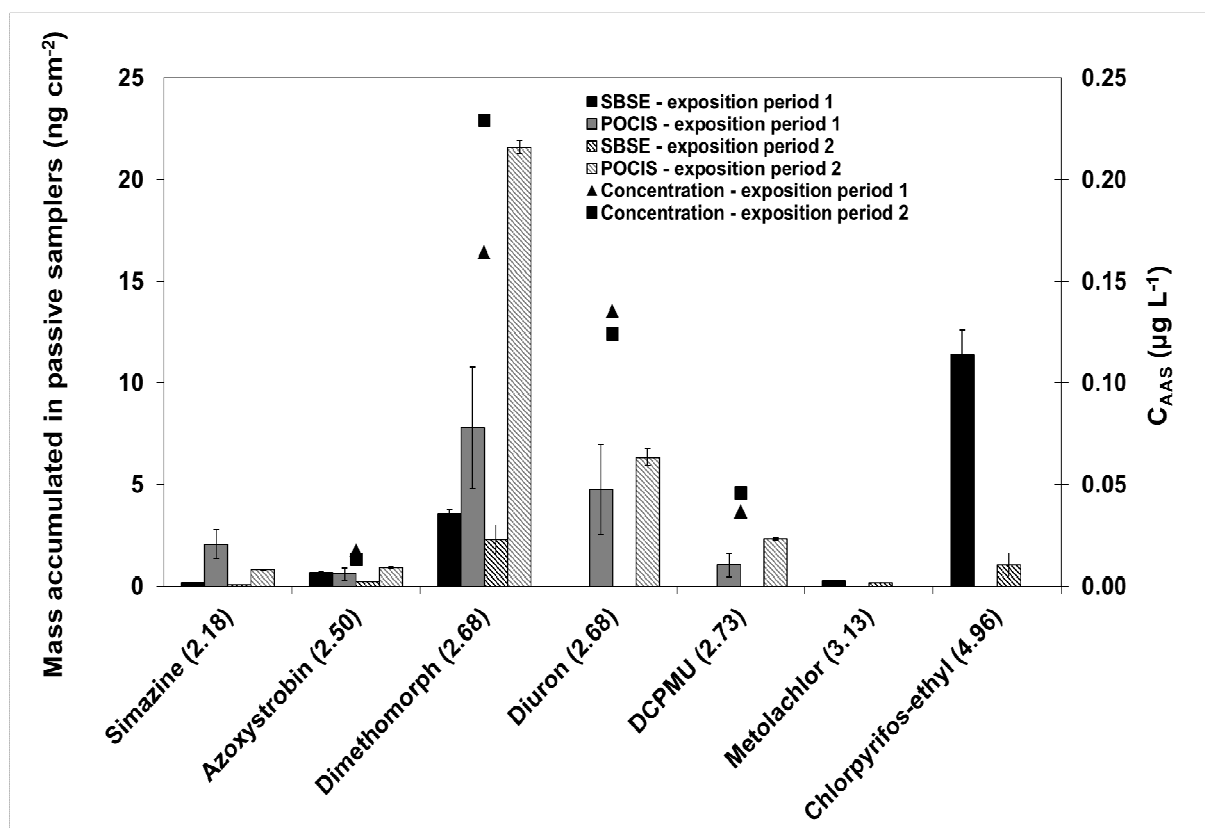
415

416 **References**

- 417 Allan IJ, Booij K, Paschke A, Vrana B, Mills GA, Greenwood R. Field Performance of Seven
418 Passive Sampling Devices for Monitoring of Hydrophobic Substances. *Environ Sci*
419 *Technol* 2009;43:5383–90.
- 420 Allan IJ, Ruus A, Schaanning, MT, Macrae, KJ, Næs, K. Measuring nonpolar organic
421 contaminant partitioning in three Norwegian sediments using polyethylene passive
422 samplers. *Sci Total Environ* 2012;423:125–31.
- 423 Alvarez DA, Huckins JN, Petty JD, Jones-Lepp T, Stuer-Lauridsen F, Getting DT, Goddard
424 JP, Gravell A. Tool for monitoring hydrophilic contaminants in water: polar organic
425 chemical integrative sampler (POCIS). In: Greenwood R, Mills G, Vrana B (ed).
426 *Comprehensive Analytical Chemistry 48: Passive Sampling Techniques in*
427 *Environmental Monitoring*. Amsterdam: Elsevier Science; 2007. p. 171-97.
- 428 Alvarez DA, Petty JD, Huckins JN, Jones-Lepp TL, Getting DT, Goddard JP, Manahan SE.
429 Development of a passive, in situ, integrative sampler for hydrophilic organic
430 contaminants in aquatic environments. *Environ Toxicol Chem* 2004;23:1640–8.
- 431 Alvarez DA, Stackelberg PE, Petty JD, Huckins JN, Furlong ET, Zaugg SD, Meyer MT.
432 Comparison of a novel passive sampler to standard water-column sampling for
433 organic contaminants associated with wastewater effluents entering a New Jersey
434 stream. *Chemosphere* 2005;61:610–22.
- 435 Artigas J, Majerholc J, Foulquier A, Margoum C, Volat B, Neyra M, Pesce S. Effects of the
436 fungicide tebuconazole on microbial capacities for litter breakdown in streams. *Aquat*
437 *Toxicol* 2012;122–123:197–205.
- 438 Baltussen E, Sandra P, David F, Cramers C. Stir bar sorptive extraction (SBSE), a novel
439 extraction technique for aqueous samples: Theory and principles. *J Microcolumn Sep*
440 1999;11:737–47.
- 441 Baltussen E, Sandra P, David F, Janssen H-G, Cramers C. Study into the Equilibrium
442 Mechanism between Water and Poly(dimethylsiloxane) for Very Apolar Solutes:
443 Adsorption or Sorption? *Anal Chem* 1999;71:5213–6.
- 444 Baltussen E, Cramers C, Sandra P. Sorptive sample preparation - a review. *Anal Bioanal*
445 *Chem* 2002;373:3–22.
- 446 Booij K, Smedes F, Van Weerlee E. Spiking of performance reference compounds in low
447 density polyethylene and silicone passive water samplers. *Chemosphere*
448 2002;46:1157–61.
- 449 Coquery M, Morin A, Bécue A, Lepot B. Priority substances of the European Water
450 Framework Directive: analytical challenges in monitoring water quality. *TrAC Trends*
451 *in Analytical Chemistry* 2005;24:117–27.
- 452 David F, Sandra P. Stir bar sorptive extraction for trace analysis. *J Chromatogr A*
453 2007;1152:54–69.

- 454 European Commission (EC), (2000) Directive 2000/60/EC of the European Parliament and of
455 the Council of 23 October 2000 establishing the framework for Community action in
456 the field of water policy. OJ L 327, 22.12.2000, p. 73
- 457 Greenwood R, Mills G, Vrana B (ed). *Comprehensive Analytical Chemistry 48: Passive*
458 *Sampling Techniques in Environmental Monitoring*. 1st ed. Amsterdam: Elsevier
459 Science; 2007.
- 460 Huckins JN, Petty JD, Booij, K. *Monitors of Organic Chemicals in the Environment:*
461 *Semipermeable Membrane Devices*. 1st ed. New York:Springer Science+Business
462 Media;2006
- 463 Jahnke A, Mclachlan M, Mayer P. Equilibrium sampling: Partitioning of organochlorine
464 compounds from lipids into polydimethylsiloxane. *Chemosphere* 2008;73:1575–81.
- 465 Lissalde S, Mazzella N, Fauvelle V, Delmas F, Mazellier P, Legube B. Liquid
466 chromatography coupled with tandem mass spectrometry method for thirty-three
467 pesticides in natural water and comparison of performance between classical solid
468 phase extraction and passive sampling approaches. *J Chromatogr A* 2011;1218:1492–
469 502.
- 470 Margoum C, Guillemain C, Yang X, Coquery M. Stir bar sorptive extraction coupled to liquid
471 chromatography-tandem mass spectrometry for the determination of pesticides in
472 water samples: Method validation and measurement uncertainty. *Talanta* 2013;116:1-
473 7.
- 474 Mazzella N, Debenest T, Delmas F. Comparison between the polar organic chemical
475 integrative sampler and the solid-phase extraction for estimating herbicide time-
476 weighted average concentrations during a microcosm experiment. *Chemosphere*
477 2008;73:545–50.
- 478 Mazzella N, Dubernet J-F, Delmas F. Determination of kinetic and equilibrium regimes in the
479 operation of polar organic chemical integrative samplers: Application to the passive
480 sampling of the polar herbicides in aquatic environments. *J Chromatogr A*
481 2007;1154:42–51.
- 482 Mazzella N, Lissalde S, Moreira S, Delmas F, Mazellier P, Huckins JN. Evaluation of the Use
483 of Performance Reference Compounds in an Oasis-HLB Adsorbent Based Passive
484 Sampler for Improving Water Concentration Estimates of Polar Herbicides in
485 Freshwater. *Environ Sci Technol* 2010;44:1713–9.
- 486 Morin N, Miège C, Coquery M, Randon J. Chemical calibration, performance, validation and
487 applications of the polar organic chemical integrative sampler (POCIS) in aquatic
488 environments. *TrAC* 2012;36:144–75.
- 489 Namieśnik J, Zabiegała B, Kot-Wasik A, Partyka M, Wasik A. Passive sampling and/or
490 extraction techniques in environmental analysis: a review. *Anal Bioanal Chem*
491 2005;381:279–301.
- 492 Paschke A, Schwab K, Brümmer J, Schüürmann G, Paschke H, Popp P. Rapid semi-
493 continuous calibration and field test of membrane-enclosed silicone collector as
494 passive water sampler. *J Chromatogr A* 2006;1124:187–95.

- 495 Prieto A, Basauri O, Rodil R, Usobiaga A, Fernandez L A, Etxebarria N, Zuloaga O. Stir-bar
496 sorptive extraction: A view on method optimisation, novel applications, limitations
497 and potential solutions. *J Chromatogr A* 2010;1217:2642–66.
- 498 Prokeš R, Vrana B, Klánová, J. Levels and distribution of dissolved hydrophobic organic
499 contaminants in the Morava river in Zlín district, Czech Republic as derived from their
500 accumulation in silicone rubber passive samplers. *Environ Pollut* 2012;166:157–66.
- 501 Rabiet M, Margoum C, Gouy V, Carluer N, Coquery M. Assessing pesticide concentrations
502 and fluxes in the stream of a small vineyard catchment - Effect of sampling frequency.
503 *Environ Pollut* 2010;158:737–48.
- 504 Roy G, Vuillemin R, Guyomarch J. On-site determination of polynuclear aromatic
505 hydrocarbons in seawater by stir bar sorptive extraction (SBSE) and thermal
506 desorption GC-MS. *Talanta* 2005;66:540–6.
- 507 Rusina TP, Smedes F, Klanova J, Booij K, Holoubek I. Polymer selection for passive
508 sampling: A comparison of critical properties. *Chemosphere* 2007;68:1344–51.
- 509 Stuer-Lauridsen F. Review of passive accumulation devices for monitoring organic
510 micropollutants in the aquatic environment. *Environ Pollut* 2005;136:503–24.
- 511 Tienpont B, David F, Bicchi C, Sandra P. High capacity headspace sorptive extraction. *J*
512 *Microcolumn Sep* 2000;12:577–84.
- 513 Vrana B, Mills G, Allan IJ, Dominiak E, Svensson K, Knutsson J, Morrison G, Greenwood R.
514 *TrAC* 2005;24:845–68
- 515 Vrana B, Paschke A, Popp, P. Calibration and field performance of membrane-enclosed
516 sorptive coating for integrative passive sampling of persistent organic pollutants in
517 water. *Environ Pollut* 2006;144:296–307.
- 518 Vrana B, Paschke A, Popp P, Schüürmann G. Use of semipermeable membrane devices
519 (SPMDs). *Environ Sci Pollut R* 2001;8:27–34.
- 520



521
522 Figure 1. Comparison of masses of pesticides accumulated in POCIS and passive Twisters
523 normalized to the respective surface area of the devices, for the two exposition periods of two
524 weeks; and weekly average water concentration obtained via automated sampler (C_{AAS}).
525 Numbers in brackets on the x axis are the log K_{ow} of the pesticides, sorted by increasing
526 values. DCPMU stands for 1-(3,4-dichlorophenyl)-3-methyl urea. Error bars represent ±
527 standard deviation, n = 3 for passive samplers.

528 Table 1. Masses of pesticides accumulated in Twisters for comparison of passive SBSE and automated sampling coupled with analytical SBSE.
 529 Passive Twisters were exposed in triplicates (n = 3) for 4 periods of 1 week. Analytical Twisters were used in triplicates (n = 3) for the extraction
 530 of 4 weekly-averaged river water samples, collected during the same period.

Pesticides	log K _{ow}	LOQ (ng)	Passive SBSE					Analytical SBSE				
			Week 1 (ng) RSD (%)	Week 2 (ng) RSD (%)	Week 3 (ng) RSD (%)	Week 4 (ng) RSD (%)	Average ^a (ng)	Week 1 (ng) RSD (%)	Week 2 (ng) RSD (%)	Week 3 (ng) RSD (%)	Week 4 (ng) RSD (%)	Average ^a (ng)
Simazine	2.18	0.2	0.4 (9.7)	2.3 (22.1)	0.8 (11.6)	0.5 (31.6)	1.0	0.3 -	0.6 (5.9)	8.2 (17.7)	2.1 (18.7)	2.8
Norflurazon	2.30	4.0	4.1 -	4.5 (12.7)	nq -	4.1 -	4.2	nq -	nq -	6.6 (18.6)	9.7 (48.9)	8.2
Azoxystrobin	2.50	0.4	1.5 (15.7)	3.4 (28.3)	1.5 (35.8)	1.4 (57.5)	1.9	1.3 (14.4)	1.4 (30.4)	1.5 (7.7)	1.4 (14.3)	1.4
Atrazine	2.61	0.2	0.3 (5.1)	nq -	nq -	nq -	0.3	nq -	nq -	nq -	nq -	nq
Dimethomorph	2.68	2.0	4.3 (7.5)	6.8 (17.6)	2.2 -	2.1 -	3.9	8.9 (28.8)	15.8 (22.7)	7.2 (11.8)	4.7 (11.1)	9.2
Diuron	2.68	20	nq -	nq -	nq -	nq -	nq	nq -	nq -	nq -	nq -	nq
3,4-dichloroaniline	2.69	1.0	nq -	nq -	nq -	nq -	nq	nq -	nq -	nq -	nq -	nq
1-(3,4-dichlorophenyl)-3-methyl urea	2.73	20	nq -	nq -	nq -	nq -	nq	nq -	nq -	nq -	nq -	nq
Isoproturon	2.87	2.0	nq -	nq -	nq -	nq -	nq	nq -	nq -	nq -	nq -	nq
Spiroxamine	2.89	0.4	4.8 (4.3)	9.8 (47.1)	13.1 (15.6)	10.7 (13.7)	9.6	0.3 (13.5)	2.8 (21.7)	1.0 (16.8)	0.5 (5.7)	1.2
Procymidon	3.08	4.0	9.7 (11.9)	23.7 (48.3)	14.0 (21.5)	17.9 (36.5)	16.3	15.0 (26.0)	10.6 (20.3)	nq -	nq -	12.8
Metolachlor	3.13	0.2	0.4 (11.4)	0.3 (8.1)	0.2 -	0.2 (12.3)	0.3	0.2 -	nq -	nq -	nq -	0.2
Fenitrothion	3.32	10	nq -	nq -	nq -	nq -	nq	nq -	nq -	nq -	nq -	nq
Tebuconazole	3.70	2.0	3.7 (19.8)	9.9 (18.1)	8.3 (26.2)	8.5 (41.8)	7.6	2.8 (10.3)	5.7 (39.3)	2.3 (7.2)	2.5 (10.6)	3.3
Chlorfenvinphos	3.81	2.0	3.2 (4.0)	nq -	nq -	nq -	3.2	2.2 -	nq -	nq -	nq -	2.2
Acetochlor	4.14	2.0	nq -	nq -	nq -	nq -	nq	nq -	nq -	nq -	nq -	nq
Diflufenican	4.20	4.0	6.6 (10.3)	5.3 (5.8)	nq -	nq -	6.0	nq -	nq -	nq -	nq -	nq
Chlorpyrifos-ethyl	4.96	1.0	nq -	2.8 (33.8)	nq -	nq -	2.8	nq -	nq -	nq -	nq -	nq
Flufenoxuron	5.11	4.0	nq -	nq -	nq -	nq -	nq	nq -	nq -	nq -	nq -	nq

531 ^a: Average masses of pesticides were calculated with only the values above the LOQ; nq: not quantified