



HAL
open science

Impact of water on the cis-trans photoisomerization of hydroxychalcones.

Yoann Leydet, Pinar Batat, Gediminas Jonusauskas, Sergey A. Denisov, João Carlos Lima, A. Jorge Parola, Nathan D. Mcclenaghan, Fernando Pina

► **To cite this version:**

Yoann Leydet, Pinar Batat, Gediminas Jonusauskas, Sergey A. Denisov, João Carlos Lima, et al.. Impact of water on the cis-trans photoisomerization of hydroxychalcones.. *Journal of Physical Chemistry A*, 2013, 117 (20), pp.4167-4173. 10.1021/jp402761j . hal-00845493

HAL Id: hal-00845493

<https://hal.science/hal-00845493>

Submitted on 6 Mar 2018

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial - ShareAlike 4.0 International License

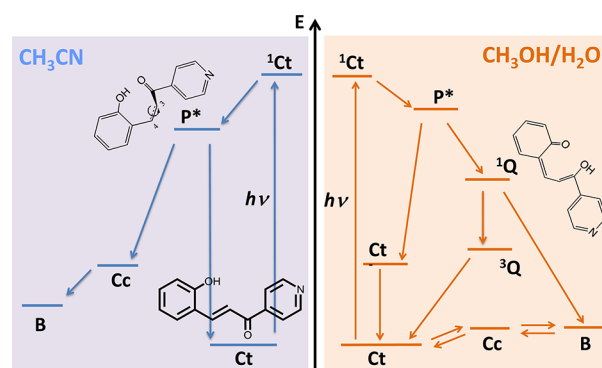
Impact of Water on the Cis–Trans Photoisomerization of Hydroxychalcones

Yoann Leydet,^{*,†} Pinar Batat,^{‡,§} Gediminas Jonusauskas,[§] Sergey Denisov,^{‡,§} João Carlos Lima,^{*,†}
A. Jorge Parola,[†] Nathan D. McClenaghan,[‡] and Fernando Pina[†]

[†]REQUIMTE, Departamento de Química, Faculdade de Ciências e Tecnologia, Universidade Nova de Lisboa, 2829 516 Caparica, Portugal

[‡]Institut des Sciences Moléculaires (ISM), CNRS UMR 5255, and [§]Laboratoire Ondes et Matière d'Aquitaine (LOMA), CNRS UMR 5798, Université de Bordeaux, F 33400 Talence, France

ABSTRACT: The photochromism of a 2 hydroxychalcone has been studied in CH₃CN and H₂O/CH₃OH (1/1, v/v), as well as in analogous deuterated solvents using steady state (UV–vis absorption, ¹H and ¹³C NMR) and time resolved (ultrafast transient absorption and nanosecond flow flash photolysis) spectroscopies. Whereas the irradiation of *trans* chalcone (Ct) under neutral pH conditions leads to the formation of the same final chromene derivative (B) in both media, two distinct photochemical mechanisms are proposed in agreement with thermodynamic and kinetic properties of the chemical reaction network at the ground state. Following light excitation, the first steps are identical in acetonitrile and aqueous solution: the Franck–Condon excited state rapidly populates the *trans* chalcone singlet excited state ¹Ct* (LE), which evolves into a twisted state ¹P*. This excited state is directly responsible for the photochemistry in acetonitrile in the nanosecond time scale (16 ns) leading to the formation of *cis* chalcone (Cc) through a simple isomerization process. The resulting *cis* chalcone evolves into the chromene B through a tautomerization process in the ground state ($\tau = 10$ ms). Unlike in acetonitrile, in H₂O/CH₃OH (1/1, v/v), the P* state becomes unstable and evolves into a new state attributed to the tautomer ¹Q*. This state directly evolves into B in one photochemical step through a consecutive ultrafast tautomerization process followed by electrocyclization. This last case represents a new hypothesis in the photochromism of 2 hydroxychalcone derivatives.



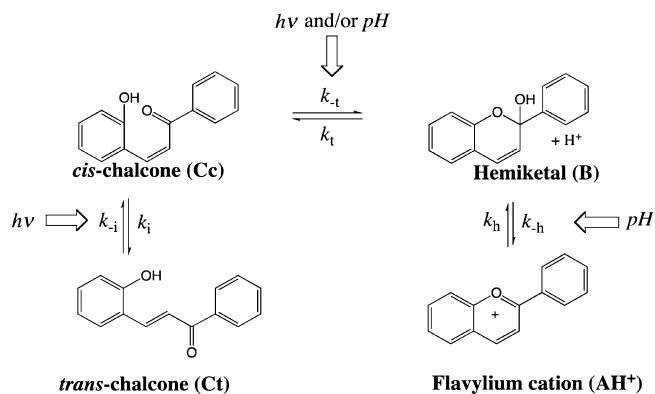
INTRODUCTION

Chalcones constitute an important class of compounds with applications in medical sciences,¹ biotechnology,² and photochemistry, because of their photochromism based on a *trans*–*cis* isomerization process.³ Within this family of compounds, the natural 2 hydroxyderivatives are particularly interesting because they can promote the formation of the flavylium cation and the quinoidal base of anthocyanins, a family of pigments responsible for most of the red and blue colors appearing in flowers and fruits.⁴ Anthocyanins are involved in a network of chemical reactions including proton transfer, hydration, tautomerization, and *cis*–*trans* isomerization.⁵ The complexity of this natural network has been reproduced by flavylium salts, synthetic analogues of anthocyanins, leading to efficient and versatile photochromic systems depending on pH and light inputs.⁶

The network of chemical reactions occurring in flavylium compounds is shown in Scheme 1 for 2 phenyl 1 benzopyrylium (structurally, the simplest flavylium). In aqueous solution, at sufficiently acidic pH values (pH < 1), the flavylium cation, AH⁺, is the more stable and dominant species. As the pH is

increased (pH > 2), the flavylium cation undergoes hydration to give the hemiketal B form, which, in turn, can tautomerize to form a *cis* chalcone (Cc). This species can further undergo isomerization to the *trans* chalcone (Ct) form. The system can proceed forward and backward by the action of pH and light and has been used as a “write–lock–read–unlock–erase” molecular switching system⁷ and to mimic some elementary properties of neurons.⁸ The photoisomerization observed for 2 hydroxychalcone derivatives has been widely studied, revealing a relationship between the quantum yield of isomerization and the chemical structure, viscosity, and polarity of the medium.⁶ Despite the apparent simplicity of photochemical processes, the ubiquity of systems displaying such reactions (e.g., olefins, stilbenes, azobenzenes, amides, enols) has led to a large variety of mechanisms accounting for the elementary processes after photoexcitation.⁹ Some parallels can be drawn between chalcones¹⁰ and other *trans*–*cis* photoisomerization model compounds (such as stilbene)¹¹ and perhaps even more so with

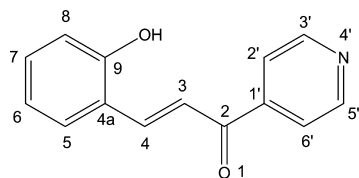
Scheme 1. Network of Chemical Reactions in Acidic/Neutral Medium Connecting *trans* 2 Hydroxychalcone (Ct) and 2 Phenyl 1 benzopyrylium (AH⁺)



compounds, such as dimethylaminobenzonitrile (DMABN),¹² that contain donor/acceptor substituents across the double bond, which confers charge transfer character to the first excited state. However, unlike in the aforementioned photochromic families, the photoisomerization reaction represents only one of the several processes taking place in the excited state, and competition between these processes can occur. Other photophysical processes have already been identified, such as the formation of various triplet states, electrocyclization, or photoinduced proton transfer especially in aqueous medium.¹³ To the best of our knowledge, a comprehensive study including the effect of the solvent on the deactivation pathways of the first excited state and the identification of the first intermediate species after excitation leading to photoisomerization and other competing channels has never been published.

We recently demonstrated that the formation of a flavylium salt can be excluded or limited to extremely acidic conditions when an electron withdrawing substituent is inserted in the structure of the 2 hydroxychalcone skeleton.^{14,15} In this study, the absence of a flavylium cation facilitates not only the attribution of the processes involved in the photophysics of 2 hydroxychalcones but also the determination of the photo product of the reaction. We report a detailed study on the photophysics and photochemistry of a 2 hydroxypyridinechalcone (compound 1, Scheme 2). The aim of this work was to

Scheme 2. Structure and Numbering of Compound 1^a



^aNotation of the flavylium cation instead of the chalcone ions was used to simplify the discussion.

characterize the photochromism of 2 hydroxychalcones, from the formation of the excited state responsible for the isomerization process to the identification of the photoproduct. The solvent dependence of the photochemistry was studied first in an organic solvent (CH₃CN) and then in an aqueous solution under neutral conditions (H₂O/CH₃OH, 1/1, v/v), as

well as in the analogous deuterated mixture (D₂O/CD₃OD, 1/1, v/v).

EXPERIMENTAL SECTION

Instrumentation and Measurements. Compound 1 was available from previous studies.¹⁴ Solutions were prepared using Millipore water and spectroscopic grade methanol and acetonitrile. Electronic absorption spectra were recorded on a Varian Cary 100 Bio or Shimadzu VC2501 PC spectrophotometer. Photochemical transformations were carried out using a medium pressure xenon/mercury arc lamp, and the excitation bands (254 and 365 nm) were isolated with interference filters (Oriol). The incident light intensity was measured by ferrioxalate actinometry.¹⁶ NMR spectra at 298.0 K were obtained on a Bruker AMX400 spectrometer operating at 400.13 MHz (¹H) and 100 MHz (¹³C). Nanosecond laser flash photolysis experiments were run on an LKS.60 laser photolysis spectrometer from Applied Photophysics, using an Applied Photophysics SX20 stopped flow spectrometer to guarantee that all of the transient signals were reproducibly obtained with fresh solution and to avoid accumulation of photoproducts competing for light excitation.

Picosecond transient absorption spectra were recorded on a setup that was built as follows: A frequency tripled Nd:YAG amplified laser system (30 ps, 30 mJ at 1064 nm, 20 Hz, Ekspla model PL 2143) output was used to pump an optical parametric generator (Ekspla model PG 401), producing tunable excitation pulses in the range of 410–2300 nm. The residual fundamental laser radiation was focused in a high pressure Xe filled breakdown cell where a white light pulse for sample probing was produced. All light signals were analyzed by a spectrograph (Princeton Instruments Acton model SP2300) coupled with a high dynamic range streak camera (Hamamatsu C7700). Accumulated sequences (sample emission, probe without and with excitation) of pulses were recorded and treated with HPDTA (Hamamatsu) software to produce two dimensional maps (wavelength versus delay) of transient absorption intensity in the range of 300–800 nm. Typical measurement error was better than 10⁻³ of the optical density (OD). Samples were equally studied on the subpicosecond time scale: This experiment was based on a femtosecond 1 kHz Ti:sapphire system producing 30 fs, 0.8 mJ, laser pulses centered at 800 nm (Femtopower Compact Pro) coupled with an optical parametric generator (Light Conversion Topas C) and frequency mixers to excite samples at the maximum of the steady state absorption band. White light continuum (360–1000 nm) pulses generated in a 2 mm D₂O cell were used as a probe. A variable delay time between excitation and probe pulses was obtained using a delay line with 0.66 fs resolution. The solutions were placed in a 1 mm circulating cell. White light signal and reference spectra were recorded using a two channel fiber spectrometer (Avantes Avaspec 2048 2). A self written acquisition and experiment control program in LabVIEW enabled the recording of transient spectra with an average error of less than 10⁻³ of the optical density for all wavelengths. The temporal resolution of the setup was better than 50 fs. A temporal chirp of the probe pulse was corrected by a computer program with respect to a Lorentzian fit of a Kerr signal generated in a 0.2 mm glass plate used in place of a sample.

RESULTS AND DISCUSSION

Study in Acetonitrile. Irradiation of an equilibrated solution of compound **1** in acetonitrile at 365 nm was followed by UV-vis absorption spectroscopy (Figure 1A). The spectrum

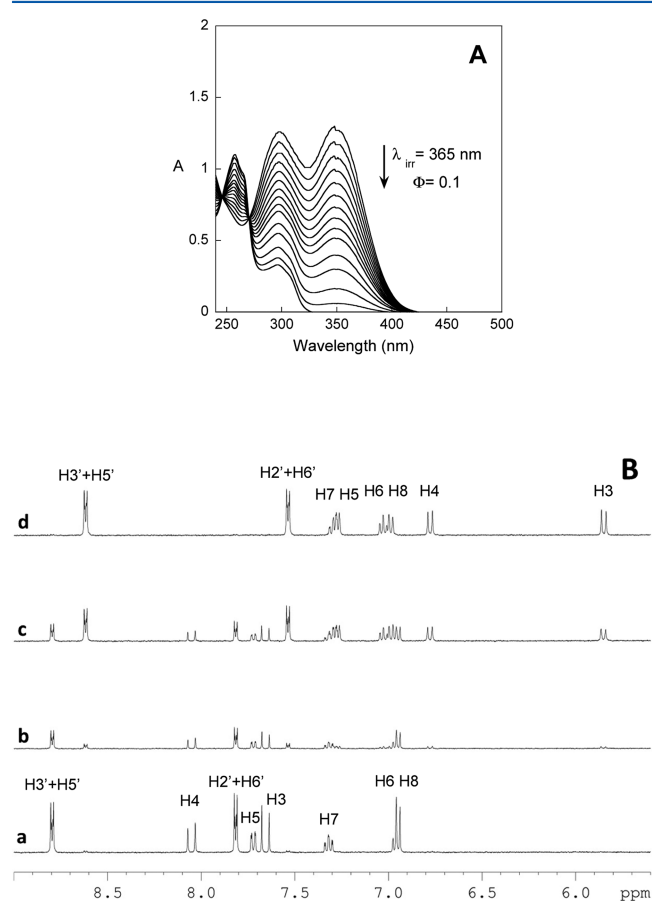


Figure 1. (A) Spectral variations upon continuous irradiation of compound **1** ($C_0 = 1.1 \times 10^{-4}$ M) in acetonitrile at 365 nm ($\Phi = 0.1$). (B) ^1H NMR spectra (400 MHz, 300 K) of **1** in CD_3CN (a) at thermal equilibrium (>95% Ct, <5% B), (b,c) upon irradiation at 365 nm, and (d) after reaching the photostationary state upon irradiation at 365 nm (100% B). Proton assignments for Ct and B given in traces a and d, respectively (according to the numbering of Ct in Scheme 2 and assuming the same numbering for B).

of the initial solution (thermally equilibrated) is dominated by a broad band at 350 nm assigned to the absorption of the Ct form. This band decreases upon irradiation until a photostationary state is reached, displaying two absorption bands with maxima at 254 and 300 nm. The spectral variations are consistent with the disappearance of Ct and concomitant formation of a photoproduct with a global quantum yield of 0.1. The well defined isosbestic points (246 and 270 nm) support the conclusion that Ct is being converted into a single photoproduct.

^1H NMR spectra, obtained before, during, and after reaching the photostationary state (traces a–d in Figure 1B), show that the initially (thermally equilibrated) solution comprises two different species, one of which is predominant (>95%). The predominant species displays a large coupling constant between protons H3 ($\delta = 7.65$ ppm) and H4 ($\delta = 8.04$ ppm) [$^3J(\text{H3}, \text{H4}) = 16.0$ Hz], typical of the *trans* chalcone isomer, whereas the minor species (<5%) is characterized by a smaller coupling constant [$^3J(\text{H3}, \text{H4}) = 9.8$ Hz] (spectrum a). The initial Ct is converted progressively into the minority species, which is also present in the initial solution (spectra b–d). Formation of additional species was not observed during irradiation or at the photostationary state. The nature of the photoproduct was unambiguously confirmed by NMR spectroscopy (HMBC experiments), where the C2 carbon presents a chemical shift in agreement with that of an sp^3 carbon ($\delta = 97.4$ ppm, Figure S1, Supporting Information), which is only present in the hemiketal species, B. In contrast to aqueous solutions, for which complete recovery of Ct is observed when solutions are allowed to equilibrate in the dark, species B is stable in acetonitrile, and only traces of Ct can be found even after 3 days. Nevertheless, the photochemistry of **1** in CH_3CN allows it to be described as a photochromic system because the recovery of Ct can be achieved upon irradiation of B at 254 nm (data not shown).

Ultrafast transient absorption spectroscopy was employed to follow the evolution of the excited *trans* chalcone. The overall spectral evolution between 400 and 750 nm in CH_3CN following 355 nm excitation is divided into two sequential steps (Figure 2).

Following excitation of **1**, two distinct signals can be observed on a picosecond time scale (Figure 2, left). An initial positive absorption signal, assigned to a locally excited state

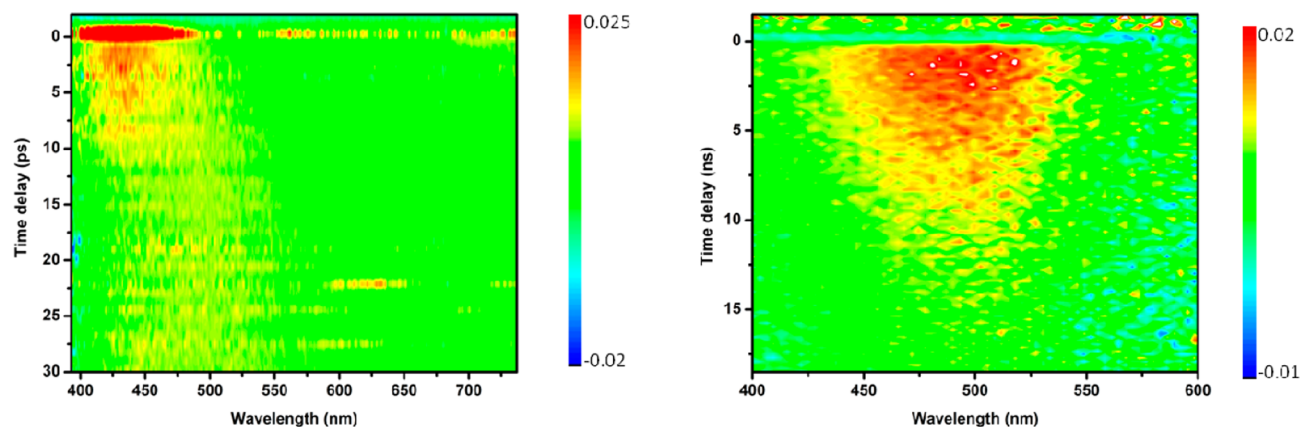


Figure 2. Ultrafast dynamics of **1** in CH_3CN . Transient absorption spectra with time delays of 0–30 ps (left) and 0–20 ns (right). Transient absorption bands were observed between 425 and 550 nm and decayed with a lifetime of 16 ns. Colors correspond to ΔOD values according to the included scales.

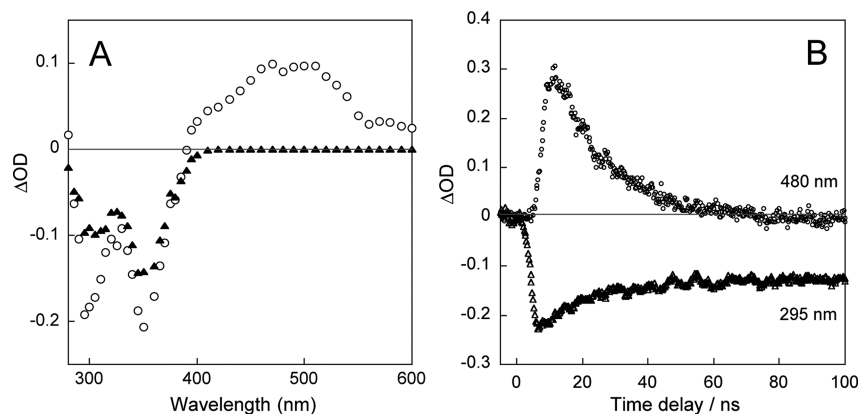


Figure 3. (A) Transient absorption spectra of **1** in CH_3CN obtained by flash photolysis in air equilibrated solution (\circ) immediately and (\blacktriangle) 100 ns after the pulse. (B) Kinetic traces at (\circ) 480 and (Δ) 295 nm.

(LE) absorbing at 420 nm, appears immediately after the pulse and subsequently relaxes to a second state (**R**), which presents a broad red shifted band extending between 400 and 550 nm. The LE state decay time is identical to the formation time of the relaxed state **R** ($\tau_1 = 2$ ps). The spectrum of **R** does not shift further, and the relaxed state decays on the nanosecond time scale ($\tau_2 = 16$ ns) (see Figure 2, right), at the same rate over the whole spectral region, indicating the presence of a single transient species. The absorption of **R** cannot be attributed to any molecule of the network in the ground state (**Cc**, **B**) because such species do not absorb at these wavelengths. Nanosecond flash photolysis ($\lambda_{\text{exc}} = 355$ nm) was employed to further follow the decay of the relaxed state **R** and characterize its relation with the ground state photo product formation. The time resolved absorption spectra immediately after the flash and after complete disappearance of **R** (100 ns) are represented between 280 and 600 nm in Figure 3.

Whereas the spectrum recorded immediately after the nanosecond laser pulse is composed of the sum of **Ct** bleaching (negative ΔOD) and transient absorption of **R** (positive ΔOD), the spectrum after 100 ns presents only a negative signal that is stable on the microsecond time scale. The recovery of **Ct** is not complete, in agreement with the photoconversion of *trans* chalcone into photoproducts. Also, the final spectrum is different from that of pure **Ct**, confirming the presence of a photoproduct absorbing in the same region. The quantum yield for photoproduct formation can be crudely estimated to be 0.5, based on the intensity of the persistent negative signal with respect to the initial intensity, immediately after excitation (see Figure 3b); however, the presence of a photoproduct absorbing in the same region can introduce a significant error in this value.

The pure **R** absorption can be obtained by subtracting the contribution of **Ct** to the initial spectrum (Figure S2, Supporting Information), and the photoproduct can be obtained on subtracting the contribution of **Ct** to the final spectrum (Figure S3, Supporting Information). It can be seen that the spectrum obtained for the photoproduct in this time window corresponds to a photoproduct different from **B** (see Figure S3, Supporting Information), the final product of the irradiation identified by ^1H NMR spectroscopy. The photo product absorption in this time window resembles the features of the absorption spectrum of the **Cc** isomer (Figure S3, Supporting Information); unfortunately, it is not possible to

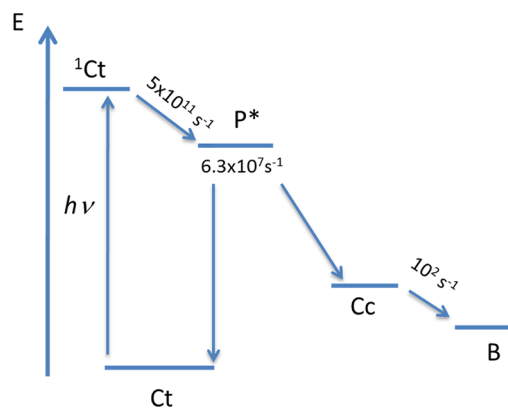
obtain the spectrum of short lived **Cc** in acetonitrile, and the comparison must be made with the spectrum of **Cc** in water.¹⁴ For the stated reasons, the photoproduct is attributed to the formation of **Cc** through deactivation of **R** together with **Ct**. On this basis, the most reliable alternative is the attribution of the transient absorption **R** to a twisted excited state **P*** corresponding to a rotation of 90° around the isomerizing double bond, which, in 16 ns, yields both **Cc** and **Ct** with equal relaxation rates, as deduced from the bleaching intensities immediately and 100 ns after excitation (Figure 3b).

This interpretation is complemented by a kinetic analysis performed on the millisecond time scale at 330 nm, showing that the resulting *cis* chalcone evolves to the chromene **B** through a fast tautomerization process ($\tau = 10$ ms), in agreement with similar kinetics recorded previously¹⁷ for this type of system (Figure S4, Supporting Information).

The thermal stability of the final photoproduct, **B**, in acetonitrile accounts for the absence of **Cc** in the final solution; that is, the **Cc** isomer initially formed from **P*** is completely converted to **B**. The **Cc** species could not be obtained, either thermally or upon irradiation.

Pertinent photophysical events, with kinetic parameters, leading to the formation of **B** are summarized in Scheme 3. Following light excitation, the Franck–Condon excited state rapidly populates the *trans* chalcone singlet excited state $^1\text{Ct}^*$ (LE), which evolves into the twisted state $^1\text{P}^*$ ($\tau = 2$ ps). This

Scheme 3. Jablonski Diagram Showing the Species at Both the Excited and Ground States Intervening in the Photochemistry of Compound 1 (Ct) in CH_3CN Solution



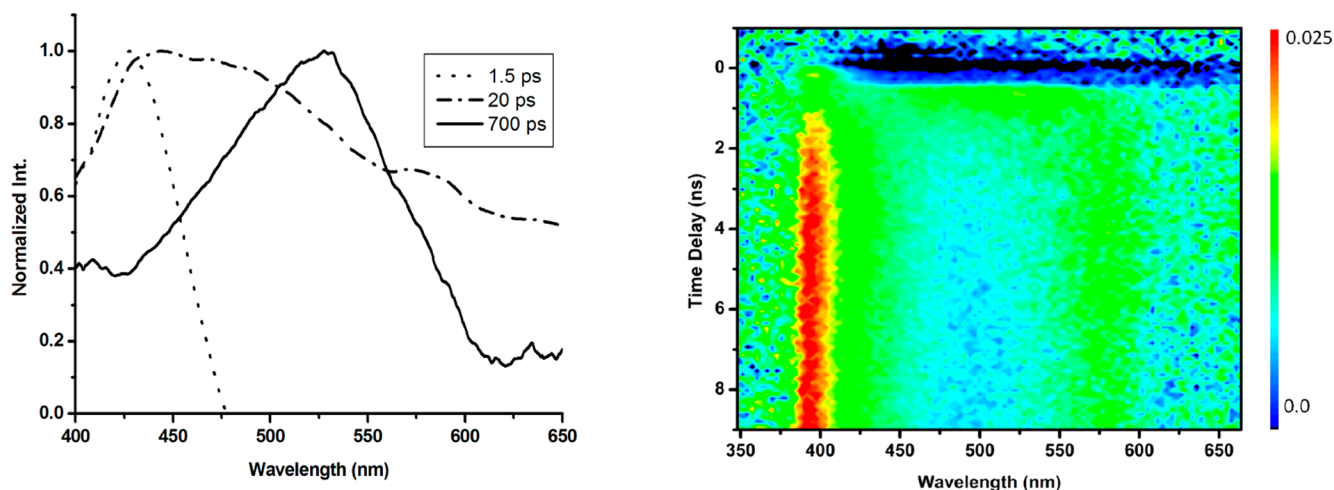


Figure 4. Ultrafast dynamics of **1** in $\text{H}_2\text{O}/\text{CH}_3\text{OH}$ at neutral pH. Transient absorption spectra (left) on the picosecond time scale after 1.5, 40, and 700 ps and (right) on the longer nanosecond time scale after 0–9 ns. Colors correspond to ΔOD values according to the included scale.

excited state is responsible for the formation of both **Ct** and **Cc** in the ground state.

The final product observed in the NMR experiment was formed through a tautomerization process in the ground state ($\tau = 10$ ms) with a quantum yield of 0.1 (Figure 1 and Figure S4, Supporting Information).

Photochemistry in $\text{H}_2\text{O}/\text{CH}_3\text{OH}$ (1/1, v/v). The photochemistry of **1** in the aqueous medium $\text{H}_2\text{O}/\text{CH}_3\text{OH}$ (1/1, v/v) was previously studied.¹⁴ Irradiation of non equilibrated solutions (100% **Ct**) also leads to the exclusive formation of **B**. The results from pump–probe experiments of **1** in $\text{H}_2\text{O}/\text{CH}_3\text{OH}$ (1/1, v/v) solution are presented in Figure 4 for two different temporal scales: picosecond and nanosecond.

An initial positive absorption at 420 nm appeared immediately after the excitation pulse and subsequently relaxed to a second red shifted broad band in the visible region with a maximum at 480 nm. The initial spectral evolution is similar to that observed in previous experiments in CH_3CN and can be assigned to the formation of the twisted $^1\text{P}^*$ state from $^1\text{Ct}^*$ (LE state) with a lower kinetic rate constant ($\tau = 5$ ps) (Figure S5, Supporting Information). The initial spectra of $^1\text{P}^*$ in the two media are identical. This signature is constant over time in acetonitrile solution, until the disappearance of the twisted excited state after 16 ns.

However, in aqueous solution, the maximum of the broad band further shifts from 480 to 520 nm (Figure 4, left). Unfortunately, the rate of disappearance of the P^* state could not be obtained because the signal was too weak in the ultrafast experiment, but on the other hand, no shift was observed in the longer time scale experiment, meaning that this process is probably faster than 50 ps. The aforementioned stabilization of the $\text{S}_n \leftarrow \text{S}_1$ transition of the twisted $^1\text{P}^*$ state from 480 to 520 nm (ca. 1600 cm^{-1}), could correspond to water mediated tautomerization (enolization). We refer to the state absorbing at 520 nm as the Q^* state.

To confirm the involvement of proton transfer in the formation of the Q^* state, pump–probe experiments of **1** in $\text{D}_2\text{O}/\text{CD}_3\text{OD}$ (1/1, v/v) solution were performed (Figure S6, Supporting Information). In $\text{D}_2\text{O}/\text{CD}_3\text{OD}$, the P^* state evolves into the Q^* state after ca. 100 ps, and even though the signal is still too weak in the ultrafast experiment, the spectral evolution can be clearly seen on the longer time scales (see Figure S6, Supporting Information). The decrease in the

Q^* formation rate in the deuterated solvent mixture is a clear indication that proton migration is involved in the $\text{P}^* \rightarrow \text{Q}^*$ conversion, supporting the hypothesis of water mediated tautomerization. The Q^* state, which has a short lifetime (900 ps in $\text{H}_2\text{O}/\text{CH}_3\text{OH}$, 3 ns in $\text{D}_2\text{O}/\text{CD}_3\text{OD}$) evolves into a transient absorption extending over the whole visible region with two different maxima at 395 and 580 nm that does not decay in this time window (Figure 4, right). The kinetic traces collected at 580 nm showed a single exponential formation with a rate identical to the decay rate of the Q^* state.

To clarify the subsequent process(es), flash photolysis experiments were performed between 280 and 700 nm (Figure 5).

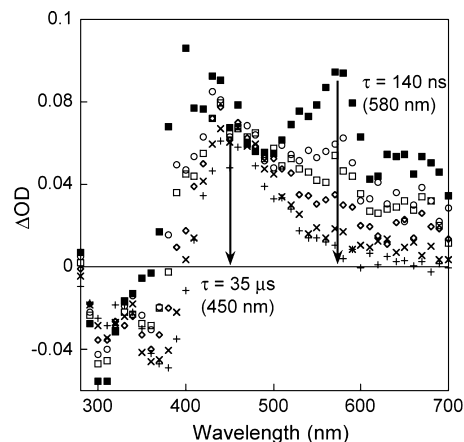


Figure 5. Transient absorption spectra of compound **1** in $\text{H}_2\text{O}/\text{CH}_3\text{OH}$ (1/1, v/v) at neutral pH on the nano to microsecond time scale from 15 ns to 0.9 μs . The evolution of the remaining signal is not shown ($\tau = 35\ \mu\text{s}$). (■) 15, (○) 100, (□) 200, (◇) 300, (×) 600, and (+) 900 ns.

The evolution on the nano /microsecond time scale allowed identification of two different contributions in addition to the negative signal clearly identified as the photobleaching of **Ct**. The previously observed transient at 580 and 395 nm decays according to a single exponential process with a rate equal to $7.3 \times 10^6\ \text{s}^{-1}$ ($\tau = 140$ ns), which could be affected by molecular oxygen as expected for a triplet state. After the triplet

decayed, the remaining spectrum could be fitted by superimposing the spectra of Ct and Ct⁻ (Figure S7, Supporting Information, and Figure 5). Ct⁻ (transient absorbing at 450 nm) further decays as a single exponential with a rate of $2.9 \times 10^4 \text{ s}^{-1}$ ($\tau = 35 \text{ }\mu\text{s}$). The assignment to Ct⁻ is further corroborated by an observed dependence of the decay rate on the pH (and independence of the presence of oxygen). Whereas it is clear that part of the Ct recovery is linked to the triplet decay, it is not straightforward, because of the spectral overlap, to elucidate whether Ct⁻ is formed initially from the deprotonation of Q* or later from the deprotonation of the triplet state. However, the isotope effect observed in the rate of disappearance of Q* (900 ps in H₂O/CH₃OH, 3 ns in D₂O/CD₃OD) is compatible with a deprotonation reaction and, as a consequence, with the formation of Ct⁻ from Q*. Inspection of Figure 5 shows that the transient corresponding to Ct⁻ does not change during this time scale, which is also compatible with its formation from the deprotonation of the Q* state. In fact, the subtraction of Ct and Ct⁻ contributions at 2 μs from the transient spectra obtained at successive times (Figure S8, Supporting Information) yields the disappearance of the same spectrum that we assigned to the pure triplet state.

Nevertheless, the final photoproduct (Cc or B) is produced in a very small amount, either from Q* or from the triplet. Also, the photochemical quantum yield in methanol/water (0.02) is much lower than that in acetonitrile (0.1), and this probably implies that formation of Ct⁻ and the triplet compete with the photoproduct formation pathway. From the flash photolysis data, after the disappearance of Ct⁻, only the presence of Ct can be observed. To identify the nature of the photoproducts, photoradiations (at 365 nm with a continuous source) were carried out in equilibrated solutions of **1** in H₂O/CH₃OH (1/1, v/v) whose composition is a mixture of Ct, Cc, and B in a ratio 8:1:1, as followed by ¹H NMR spectroscopy (Figure 6).

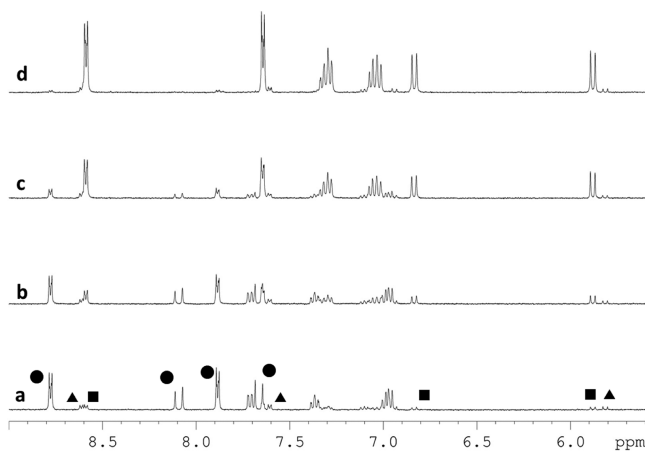


Figure 6. ¹H NMR spectra (400 MHz, 298 K) of **1** in D₂O/CD₃OD (1:1, v/v) (a) at thermal equilibrium [(●) 80% Ct, (▲) 10% Cc, (■) 10% B], (b,c) during the course of the irradiation at 365 nm, and (d) after reaching the photostationary state upon irradiation at 365 nm (90% B, 10% Cc).

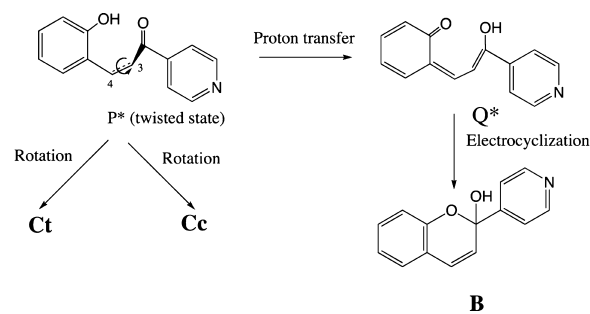
¹H NMR spectra were run before, during, and after reaching the photostationary state (Figure 6). The initial solution composed of 80% Ct, 10% Cc, and 10% B is converted into a solution composed by 90% B and 10% Cc. The molar fraction of Cc remains stable during the irradiation. The disappearance of Ct and concomitant formation of B has a global quantum

yield of 0.02, which why it was not observed during flash photolysis.

If the photoproduct of the irradiation of Ct were Cc, which, in a later step, would equilibrate with B, then the ratio between B and Cc would be constant. The ¹H NMR data clearly show that the photoproduct of the irradiation of Ct is B and that the equilibrium between B and Cc is not attained during the time scale of the experiment.¹⁴ In other words, Q* yields B (in competition with triplet and Ct⁻ formation) whereas P* yields Cc plus Ct. The formation of B in methanol/water solutions (and not Cc, as in acetonitrile) must be related with the nature of the new intermediary state Q*, which is present only in this solvent.

The formation of hydrogen bonds with water can facilitate the proton transfer between the two oxygen atoms, and the most straightforward assignment of Q* is the quinoidal tautomer derived from proton transfer (Scheme 4).

Scheme 4. Representation of the Two Distinct Solvent Dependent Mechanisms for **1**^a



^aIn CH₃CN, the excited state P* directly induces the formation of Ct or Cc, whereas in H₂O/CH₃OH, B is promoted from the Q* state formed after a solvent assisted proton transfer.

Whereas P* can yield Ct or Cc through rotation around the C3–C4 bond, after the proton transfer, the most efficient reaction pathway for the quinone is electrocyclization to form B. The assignment of Q* as a quinone is also in agreement with very efficient formation of the triplet, which was not previously observed in acetonitrile, where P* was the only state observed.

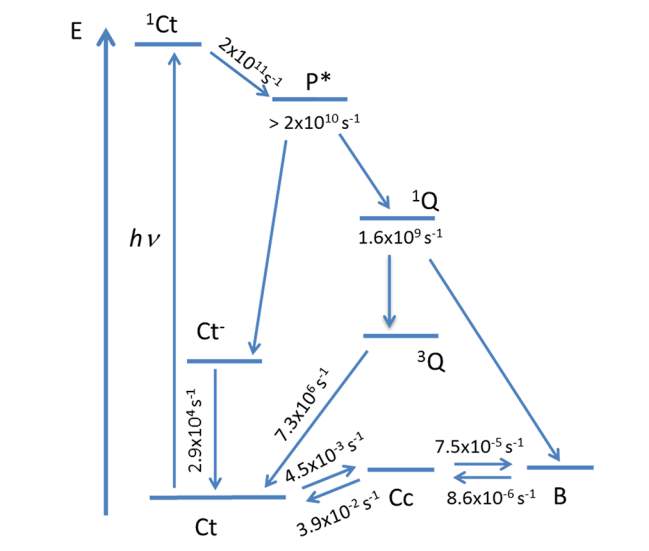
A summary of the photophysical events with kinetic parameters leading to the formation of B in methanol/water is shown in Scheme 5. Following light excitation, the first steps are identical in acetonitrile and in water: The Franck–Condon excited state rapidly populates the *trans* chalcone singlet excited state ¹Ct (LE), which evolves into the twisted state P* ($\tau = 5 \text{ ps}$). In D₂O/CD₃OD, this value reaches 5.4 ps. The state P* evolves into Ct⁻ and into the transient Q*, which leads to B and triplets.

CONCLUSIONS

The photochromism of a 2 hydroxychalcone has been studied in CH₃CN and in H₂O/CH₃OH (1/1, v/v) by steady state (UV–vis absorption, ¹H and ¹³C NMR) and time resolved (ultrafast transient absorption and nanosecond flow flash photolysis) spectroscopies.

In the two types of media, the first step after excitation is the same and consists of the relaxation of the LE state to a twisted ¹P* state in a few picoseconds. In acetonitrile, this state deactivates to form Cc and Ct in identical proportions, the classic situation of double bond photoisomerization. The final

Scheme 5. Jablonski Diagram Showing the Species at Both the Excited and Ground States Intervening in the Photochemistry of Compound 1 (Ct) in H₂O/CH₃OH (1/1, v/v) Solution



product, B, results from tautomerization of Cc with an observed quantum yield of 0.1

The presence of water, on the other hand, dramatically affects the photochemical pathway by promoting the formation of a tautomeric intermediate, between the ¹P* state and the photoproducts. The main photochemical pathway from this tautomer is electrocyclization to form chromene B, whereas all of the other pathways lead to Ct (through Ct⁻ and ³Q*); that is, no *cis* isomer is formed photochemically. However, the weight of the pathways leading back to Ct is much larger than that of the pathways yielding B ($\phi = 0.02$).

AUTHOR INFORMATION

Corresponding Author

*E mail: yoann.leydet@gmail.com (Y.L.), lima@fct.unl.pt (J.C.L.).

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

This work was supported by the Fundação para a Ciência e Tecnologia, National NMR Network, through Projects PTDC/QUI QUI/117996/2010 and SFRH/BPD/44230/2008 (Y.L.); the University of Bordeaux, Région Aquitaine; and the PHC Pessoa Franco Portuguese Programme.

(1) Dimmock, J. R.; Elias, D. W.; Beazely, M. A.; Kandepu, N. M. *Curr. Med. Chem.* **1999**, *6*, 1125–1149.

(2) Ono, M.; Watanabe, R.; Kawashima, H.; Yan, C.; Kimura, H.; Watanabe, H.; Haratake, M.; Saji, H.; Nakayama, M. *J. Med. Chem.* **2001**, *44*, 6401.

(3) Bouas Laurent, H.; Dürr, H. *Pure Appl. Chem.* **2001**, *73*, 639–665.

(4) (a) Swain, T. In *The Flavonoids*; Harborne, J. B., Mabry, T. J., Mabry, H., Eds.; Chapman and Hall/CRC Press: London, 1975; pp 1096–1129. (b) *Anthocyanins As Food Colors*; Markakis, P., Ed.; Academic Press: New York, 1982. (c) Brouillard, R. O.; Dangles, O. In *The Flavonoids: Advances in Research Since 1986*; Harborne, J. B., Ed.; Chapman and Hall/CRC Press: New York, 1994; pp 565–588.

(5) (a) Yoshida, K.; Mori, M.; Kondo, T. *Nat. Prod. Rep.* **2009**, *26*, 884–915. (b) Goto, T.; Kondo, T. *Angew. Chem., Int. Ed. Engl.* **1991**, *30*, 17–33. (c) Yoshida, K.; Kondo, T.; Okazaki, Y.; Katou, K. *Nature* **1995**, *373*, 291. (d) Kondo, T.; Yoshida, K.; Nakagawa, K. A.; Kawai, T.; Tamura, H.; Goto, T. *Nature* **1992**, *358*, 515–518. (e) Brouillard, R.; Delaporte, B. *J. Am. Chem. Soc.* **1977**, *99*, 8461–8468. (f) Brouillard, R.; Delaporte, B.; Dubois, J. E. *J. Am. Chem. Soc.* **1978**, *100*, 6202–6205. (g) Brouillard, R.; Lang, J. *Can. J. Chem.* **1990**, *68*, 755–761. (h) McClelland, R. A.; Gedge, S. *J. Am. Chem. Soc.* **1980**, *102*, 5838–5848. (i) McClelland, R. A.; McGall, G. H. *J. Org. Chem.* **1982**, *47*, 3730–3736.

(6) Pina, F.; Melo, M. J.; Laia, C. A. T.; Parola, A. J.; Lima, J. C. *Chem. Soc. Rev.* **2012**, *41*, 869–908.

(7) Pina, F.; Melo, M. J.; Maestri, M.; Ballardini, R.; Balzani, V. *J. Am. Chem. Soc.* **1997**, *119*, 5556–5561.

(8) Pina, F.; Melo, M. J.; Maestri, M.; Passaniti, P.; Balzani, V. *J. Am. Chem. Soc.* **2000**, *122*, 4496–4498.

(9) Fedorova, O. A.; Gulakova, E. N.; Fedorov, Y. V.; Lobazova, I. E.; Alfimov, M. V.; Jonusauskas, G. *J. Photochem. Photobiol. A* **2008**, *196*, 239–243.

(10) (a) Jiang, Y. B.; Wang, X. J.; Lin, L. *J. Phys. Chem.* **1994**, *98*, 12367–12372. (b) Rurack, K.; Dekhtyar, M. L.; Bricks, J. L.; Resch Genger, U.; Rettig, W. *J. Phys. Chem. A* **1999**, *103*, 9626–9635. (c) Rurack, K.; Bricks, J. L.; Reck, G.; Radeaglia, R.; Resch Genger, U. *J. Phys. Chem. A* **2000**, *104*, 3087–3109. (d) Fayed, T. A.; Awad, M. K. *Chem. Phys.* **2004**, *303*, 317–326.

(11) Waldeck, D. H. *Chem. Rev.* **1991**, *91*, 415–436.

(12) Grabowski, Z. R.; Rotkiewicz, K.; Rettig, W. *Chem. Rev.* **2003**, *103*, 3899–4032.

(13) (a) Diniz, M.; Gomes, R.; Parola, A. J.; Laia, C. A. T.; Pina, F. *J. Phys. Chem. B* **2009**, *113*, 719–727. (b) Jiang, Y. B.; Wang, X. J.; Lin, L. *J. Phys. Chem.* **1994**, *98*, 12367–12372. (c) Rurack, K.; Dekhtyar, M. L.; Bricks, J. L.; Resch Genger, U.; Rettig, W. *J. Phys. Chem. A* **1999**, *103*, 9626–9635. (d) Rurack, K.; Bricks, J. L.; Reck, G.; Radeaglia, R.; Resch Genger, U. *J. Phys. Chem. A* **2000**, *104*, 3087–3109. (e) Fayed, T. A.; Awad, M. K. *Chem. Phys.* **2004**, *303*, 317–326.

(14) Leydet, Y.; Parola, A. J.; Pina, F. *Chem.—Eur. J.* **2010**, *16*, 545–555.

(15) Leydet, Y.; Gavara, R.; Cunha Silva, L.; Parola, A. J.; Pina, F. *Chem.—Eur. J.* **2011**, *17*, 3663–3671.

(16) Hatchard, C. G.; Parker, C. A. *Proc. R. Soc. London, Ser. A* **1956**, *235*, 518–536.

(17) Pina, F.; Maestri, M.; Balzani, V. In *Handbook of Photochemistry and Photobiology*; Nalwa, H. S., Ed.; American Science Publishers: Los Angeles, CA, 2003; Vol. 3, pp 411–449.