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Food Additives and Contaminants



Detection of illegal treatment of barrows with nandrolone ester; effects on growth, histology and residue levels in urine and hair

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Methods/Techniques:	Chromatography - LC/MS, Bioassay, Chromatography - GC/MS
Additives/Contaminants:	Drug residues - hormones
Food Types:	Animal products – meat
Abstract:	Abstract To evaluate the effects of 17β -19-nortestosterone (17β NT) treatment of barrows on residue levels and on growth results, an animal experiment was conducted. Five barrows were treated 3 times during the fattening period with 17β NT phenylpropionate (Nandrosol, nandrolone phenylpropionate 50 mg/ml,1 mg/kg BW). Another 5 barrows were not treated, and 5 boars (not treated) were kept as a positive control. Boars and treated barrows showed 13 % and 9 % better growth than not-treated barrows, leading to mean final body weights of 121.6, 117.8 and 109.0 kg respectively. Moreover the bulbourethral glands of the treated barrows were 3 times heavier than the not-treated barrows. The histology of the prostate and bulbourethral gland of the treated barrows was comparable to the boars, whereas the control barrows showed atrophic glands. Levels of 17β NT ester in hair from treated barrows were high, whereas

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3	
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13	
14	Abstract
15	To evaluate the effects of 17β -19-nortestosterone (17 β NT) treatment of barrows on
16	residue levels and on growth results, an animal experiment was conducted. Five
17	barrows were treated 3 times during the fattening period with $17\beta NT$
18	phenylpropionate (Nandrosol, nandrolone phenylpropionate 50 mg/ml,1 mg/kg BW).
19	Another 5 barrows were not treated, and 5 boars (not treated) were kept as a positive
20	control. Boars and treated barrows showed 13 % and 9 % better growth than not-
21	treated barrows, leading to mean final body weights of 121.6, 117.8 and 109.0 kg
22	respectively. Moreover the bulbourethral glands of the treated barrows were 3 times
23	heavier than the not-treated barrows. The histology of the prostate and bulbourethral
24	gland of the treated barrows was comparable to the boars, whereas the control
25	barrows showed atrophic glands. Levels of $17\beta NT$ ester in hair from treated barrows

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1	were high, whereas boars and non-treated barrows did not show levels above LLQ. It
2	is concluded that analysis of hair can provide a method for detection of illegal
3	treatment with $17\beta NT$ ester in barrows. The size of the bulbourethral gland can be
4	used for screening in the slaughterhouse.
5	
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7	+31(0) 317 417717
8	
9	Key words: 17β -19-nortestosterone, barrows, hair analysis, histology, urine analysis
10	
11	Introduction
12	In male pigs 17 β -19-nortestosterone (17 β NT) is a naturally occurring compound in
13	urine (Poelmans et al., 2005). In boars high levels can be measured but in barrows
14	and gilts only low levels occur. In practice however sometimes high levels of $17\beta NT$
15	are found in urine from barrows. This can be due to cryptorchid animals, having one
16	testicle in the abdomen that produces hormones. But illegal treatment with $17\beta NT$ for
17	growth promotion is also a possibility, since 19-nortestosterone (nandrolone) has a
18	long history for abuse as growth promoter in livestock (Rico, A.G., 1983).
19	The use of hormonal substances for fattening purposes in livestock is banned in the
20	EU since 1988 according to Council Directive 96/22 EC (Offic. J. Eu. Comm., 1996).
21	Steroids having estrogenic, androgenic, progestagenic action are forbidden as well as
22	thyreostatics and β -agonists. Residue analysis on target compounds is carried out by
23	control laboratories to enforce the ban.

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1	Some hormones that were considered as exogenic appeared to occur naturally in some
2	species (Scarth <i>et al.</i> , 2009). This applies to 17β NT which occurs in urine of stallions
3	(Houghton et al., 1994) and in urine from boars (Debruyckere et al., 1990).
4	Boars grow faster and show a better feed conversion with 20 to 30% less fat than
5	barrows (Field, 1971; Mulvaney et al., 1988; Kudson et al., 1985). Gonadally intact
6	males also have 8 to 15% greater muscle mass than castrated males (Knudson et al.,
7	1885; Prescott & Lamming, 1967). In boars, testis and accessory genital glands are
8	normally developed, whereas barrows are castrated at a young age and have atrophied
9	accessory genital glands. Treatment with androgens leads to increase in growth of the
10	seminal vesicles and bulbourethral glands in barrows (Lauwers et al., 1983; Lauwers,
11	1984).
12	
13	In recent years, apart from urine analysis, hair has been analyzed for the presence of
14	esters of anabolic steroids (Sauer et al., 1998; Duffy et al., 2009; Nielen et al, 2006,
15	Stolker et al., 2009). For growth promoting purposes steroid hormones are usually
16	administered with an ester side-chain attached in order to facilitate the steroid
17	reaching the site of activity within the animal's body. Because steroid hormones do
18	not occur naturally in an ester form, detection of intact steroid esters demonstrates
19	illegal administration of these drugs.
20	
21	To investigate if treatment with $17\beta NT$ is anabolic in barrows can lead to high levels
22	of 17 β NT in urine and hair we treated barrows with 17 β NT and compared them with
23	untreated barrows and untreated boars.
24	
25	Material and methods

1	Experimental
2	The animal experiment was performed by Schothorst Feed Research BV in Lelystad,
3	Netherlands. The trial comprised three treatments according to sex and Nandrosol
4	injection: 1) non-injected boars, 2) non-injected barrows and 3) barrows injected with
5	17βNT phenylpropionate (nandrolone phenylpropionate 50 mg/ml, 1 mg/kg BW,
6	Nandrosol, REG NL 10067, Alfasan, The Netherlands). A total of 15 pigs were
7	distributed in 5 replicates per treatment group.
8	
9	Experimental animals were grower pigs (Tempo x (Great Yorkshire x Finnish
10	Landrace)) of 60-70 days of age at the start of the experiment and approximately 25
11	kg live body weight. Pigs were housed in three separate pens (one pen/treatment) of
12	1.80 x 1.00 m. One spare pig per treatment was also allocated within the experimental
13	pigs and the control barrows and was fed and weighed in case a pen mate dropped out.
14	The animals were fed ad libitum and had free access to water. The animals were fed
15	commercial growing/finishing diets (Arkevaart-Twente Coöperatie, Nijkerk, The
16	Netherlands) commonly used in Schothorst Feed Research in practical conditions.
17	The animals were injected at 19, 47 and 75 days after the start of the experiment and
18	were slaughtered on day 93 of the experiment. Once a month urine and hair was
19	sampled and every two weeks the animals were weighed. Sampling times for hair and
20	urine were day 12, 33, 61 and 81, respectively time points 1, 2, 3 and 4. All pigs were
21	delivered to a commercial slaughterhouse at a body weight around 100-115 kg.
22	Treated animals were condemned for destruction. This study was approved by the
23	Ethical Committee of Animal Science Group Lelystad, part of Wageningen
24	University Research Centre, Netherlands.
25	

1 Histology

2	At slaughter the animals were inspected for gross pathology. Testes (only in boars),
3	prostate and bulbourethral gland were sampled for histological investigation. After
4	weighing the bulbourethral glands, samples of the prostate and bulbourethral gland
5	were immediately fixed in 10 % phosphate buffered formalin, whereas samples of the
6	testis were fixed in Bouin Hollandais fixative, which gives a better morphology. After
7	the usual dehydration series with alcohols and xylol, the tissues were embedded in
8	paraffin. Sections, 5 μ m thick, were made using a rotating microtome. Sections were
9	stained with heamatoxylin-eosin (HE) according to Mayer (Bancroft & Stevens, 1990)
10	for routine staining and the prostate and bulbourethral gland were also stained for
11	mucins with the combined Alcian Blue/periodic Acid Schiff (ABPAS) technique
12	staining both acidic (blue) and neutral mucins (red) (Bancroft & Stevens, 1990).
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14	Chemicals
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1 Sample preparation

2	The hair samples were washed with water and dried for 24 hours in an oven at 60°C.
3	Then the hair samples were cut in 0.5 cm pieces using a pair of scissors and 500 mg
4	was pulverized in a Sartorius (Goettingen, Germany) model Mikro-dismembrator S
5	ball mill. From each sample two test portions of two hundred milligrams of the
6	pulverized hair were weighed into a plastic tube. Both test portions were spiked with
7	the internal standard d ₃ -testosterone phenylpropionate at 10 ng/g. In addition one of
8	the test portions was spiked with 17 β NT phenylpropionate at 5 ng/g. Blanks and
9	controls were spiked with the internal standard d_3 -testosterone phenylpropionate at
10	10 ng/g. Control hair samples were prepared by spiking blank hair with $17\beta NT$
11	phenylpropionate at 0, 2.5, 5.0, 10 and 25 ng/g. Digestion of the hair was performed
12	by addition of 2 ml of 25 mM tris(2-carboxyethyl)phosphine (TCEP) and incubation
13	at room temperature for 1 hour. The tube was shaken by using a 'head-over-head'
14	shaker. After addition of 4 ml of methanol, the tube was centrifuged for 5 min at
15	1700g. Next, 4 ml of water was added and the mixture was applied to a C18 solid-
16	phase extraction (SPE) cartridge, pre-activated with acetonitrile, methanol and water.
17	The SPE-cartridge was washed with 2 ml of methanol/water (60/40 v/v) and eluted
18	with 2 ml of acetonitrile followed by 2 ml of ethyl acetate. The combined eluate was
19	evaporated to dryness under a gentle stream of nitrogen gas at 40°C and re-dissolved
20	in 200 μ l of water-acetonitrile-methanol-formic acid (300/350/350/20, v/v/v/v). 40 μ L
21	was injected into LC-MS/MS system 1.
22	

23 From each urine sample two test portions (2 ml) were transferred into a glass tube.

24 Both test portions were spiked with the internal standard d_3 -17 β -19-nortestosterone at

25 2 ng/ml. In addition one of these portions was spiked with 17B-19-nortestosterone at 2

1	ng/ml. Blanks and controls were spiked with the internal standard d_3 -17 β -19-
2	nortestosterone at 2 ng/ml. The control urine samples were prepared by spiking blank
3	urine with 17ß-19-nortestosterone at 0, 0.5, 1.0, 2.0, 3.0 and 5.0 ng/ml.
4	Prior to hydrolysis, the urine samples were adjusted to pH 4.8. Following the addition
5	of 10µL ß-glucuronidase/arylsulfatase (ready to use, Merck), enzymatic
6	deconjugation was carried out in a water bath at 37 °C for 16 hr. The hydrolyzed
7	sample was applied to a C18 solid-phase extraction (SPE) cartridge, pre-activated
8	with methanol and sodium acetate buffer. The vacuum dried SPE column was eluted
9	with 4 ml of methanol. The eluate was applied to a NH_2 SPE cartridge and the eluate
10	was evaporated to dryness under a gentle stream of nitrogen. The residue obtained
11	was re-dissolved in 225 μ L (methanol/water 20/80 v/v) and 200 μ L was injected into
12	a HPLC system consisting of a vacuum degasser, an autosampler, a binary pump, an
13	UV detector and a fraction collector from Agilent (Santa Clara, California). The
14	function where 170, 10, as starts starts and the damage shift damage shift damages
14	fraction where 17ß-19-nortestosterone eluted was collected and evaporated to dryness
14	under a gentle stream of nitrogen and re-dissolved in 300 μ l methanol/formic acid
15	under a gentle stream of nitrogen and re-dissolved in 300 μ l methanol/formic acid
15 16	under a gentle stream of nitrogen and re-dissolved in 300 μ l methanol/formic acid
15 16 17	under a gentle stream of nitrogen and re-dissolved in 300 μ l methanol/formic acid (0.5%) (50/250 v/v). 40 μ l was injected into LC-MS/MS system 2.
15 16 17 18	under a gentle stream of nitrogen and re-dissolved in 300 µl methanol/formic acid (0.5%) (50/250 v/v). 40 µl was injected into LC-MS/MS system 2.
15 16 17 18 19	under a gentle stream of nitrogen and re-dissolved in 300 µl methanol/formic acid (0.5%) (50/250 v/v). 40 µl was injected into LC-MS/MS system 2. <i>LC-MS/MS systems</i> System 1: The analysis of the hair samples was carried out using a UFLC liquid
15 16 17 18 19 20	under a gentle stream of nitrogen and re-dissolved in 300 μl methanol/formic acid (0.5%) (50/250 v/v). 40 μl was injected into LC-MS/MS system 2. <i>LC-MS/MS systems</i> System 1: The analysis of the hair samples was carried out using a UFLC liquid chromatography system, consisting of a vacuum degasser, an auto sampler and a
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15 16 17 18 19 20 21 22 23	under a gentle stream of nitrogen and re-dissolved in 300 μl methanol/formic acid (0.5%) (50/250 v/v). 40 μl was injected into LC-MS/MS system 2. <i>LC-MS/MS systems</i> System 1: The analysis of the hair samples was carried out using a UFLC liquid chromatography system, consisting of a vacuum degasser, an auto sampler and a binary pump from Shimadzu Scientific Instruments (Colombia, MD, USA) equipped with a reversed phase Waters Acquity UPLC BEH C18 analytical column of 100*2.1 mm and 1.7 μm particle size kept in a column oven at 50 °C. The LC gradient

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1	ml/min was: 0 min 10% B; 0.1-2 min linear increase to 70% B; 2-8 min linear
2	increase to 100%B; 8-9.5 min 100% B. Injection volume was 40 μ L. The UFLC
3	system was connected to a AB Sciex (Concord, Ontario, Canada) mass spectrometer
4	model QTRAP 5500 equipped with an electrospray interface operating in the positive
5	ionization mode. IonSpray voltage was set at 5500V and the source temperature was
6	set at 250°C. Curtain gas, gas 1 and gas 2 were 20, 55 and 60 psi respectively. The
7	desolvation gas and the collision-induced dissociation (CID) gas was nitrogen.
8	Entrance potential (EP) for all analytes was 10V. For 17ß-19-nortestosterone
9	phenylpropionate ester the most abundant MRM transition (used for the
10	quantification) was acquired using the following conditions: precursor ion mass (m/z)
11	407.1, Product ion mass (m/z) 105.2 with a Collision Energy of 33 (eV). A second
12	MRM transition, used for the confirmation of the identity of 17ß-19-nortestosterone
13	phenylpropionate ester, was acquired using the following conditions: precursor ion
14	mass (m/z) 407.1, Product ion mass (m/z) 133.1 with a Collision energy of 25 (eV).
15	For d ₃ -17β-testosterone phenylpropionate the most abundant MRM transition was
16	acquired using the following conditions: precursor ion mass (m/z) 424.2, product ion
17	mass (m/z) 105.1 with a collision energy of 30 (eV).
18	
19	The concentrations of 17ß-19-nortestosteronephenylpropionate in hair samples were
20	calculated using the isotope dilution method and a standard addition of 5 ng/g 17B-19-
21	nortestosterone phenylpropionate to each individual sample.
22	
23	System 2: The LC-MS/MS system consisted of a Waters (Manchester, UK) Acquity
24	UPLC model LC system equipped with a Waters model Quattro Premier XE triple-
25	quadrupole mass spectrometer. The mass spectrometer was operated in the positive

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1	electrospray ionization (ESI) mode at a capillary voltage of 2.7 kV, a desolvation
2	temperature of 450 °C, a source temperature of 120 °C and a cone voltage of 30V.
3	The desolvation gas was nitrogen and the collision-induced dissociation (CID) gas
4	was argon. A Waters Acquity UPLC BEH C18 analytical column of 50*2.1 mm and
5	1.7 μ m particle size was used and kept in a column oven at 40 °C. The gradient
6	(solvent A, water/formic acid 980/20 v/v); solvent B, water-acetonitrile-formic acid
7	(80/900/20; v/v/v)) with a flow of 0.4 ml/min was: 0 – 1 min 30% B;1-5 min linear
8	increase to 50% B; 5-5.1 min linear increase to 100%B; 5.1-7 min 100% B. Injection
9	volume was 40 μ L. For 17β-19-nortestosterone the most abundant MRM transition
10	(used for the quantification) was acquired using the following conditions: precursor
11	ion mass (m/z) 275.1, Product ion mass (m/z) 109.2 with a collision energy of 25
12	(eV). A second MRM transition, used for the confirmation of the identity of 17ß-19-
13	nortestosterone, was acquired using the following conditions: precursor ion mass
14	(m/z) 275.1, product ion mass (m/z) 145.1 with a collision energy of 20 (eV). For d_3 -
15	17BNT the most abundant MRM transition was acquired using the following
16	conditions: precursor ion mass (m/z) 278.2, product ion mass (m/z) 109.0 with a
17	collision energy of 30 (eV).
18	
19	The concentrations of 17BNT in urine samples were estimated using the isotope
20	dilution method and a linear calibration curve were constructed for 17BNT by
21	fortifying blank urine aliquots at six different concentration levels (matrix-matched
22	standards, MMS).
23	
24	Results and discussion
25	

Food Additives and Contaminants

1	The controlled treatment experiment was performed without problems, the animals
2	were in good health and all animals were suitable for slaughter at the end of the
3	experimental period. Including the spare animals we ended up with 6 treated barrows,
4	6 control barrows and 5 boars for investigation.
5	
6	An overview of the live weights of the animals is listed in Table 1.
7	Mean live weight at slaughter was 121.6 kg for boars, 108.9 kg for barrows and 117.8
8	kg for the 17 β NT treated barrows. The treated barrows showed a significant (p <0.05)
9	better growth performance than the control barrows, and were not significantly
10	different from the boars. At the facility where the animals were housed it appeared
11	that the last weeks before slaughter there was an increased growth of the treated
12	barrows as compared to the control animals (fig. 1).
10	Income Table 1 and Discuss 1
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14 15 16 17	These data confirmed our suggestion that 17β NT has a growth promoting effect in barrows. In literature the effects of anabolic steroids in pigs are not conclusive: In a series of experiments with implants in pigs, the combination of trenbolone and 17β -
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14 15 16 17 18 19 20 21	These data confirmed our suggestion that 17 β NT has a growth promoting effect in barrows. In literature the effects of anabolic steroids in pigs are not conclusive: In a series of experiments with implants in pigs, the combination of trenbolone and 17 β - estradiol or 17 β -estradiol and 17 β -testosterone in barrows adversely affected growth rate and feed intake, but the implanted barrows had less fat than the controls (De Wilde & Lauwers, 1984). Other experiments with castrated males (Van Weerden & Grandadam, 1976) using implants with trenbolone and 17 β -estradiol showed
14 15 16 17 18 19 20 21 22	These data confirmed our suggestion that 17β NT has a growth promoting effect in barrows. In literature the effects of anabolic steroids in pigs are not conclusive: In a series of experiments with implants in pigs, the combination of trenbolone and 17β - estradiol or 17β -estradiol and 17β -testosterone in barrows adversely affected growth rate and feed intake, but the implanted barrows had less fat than the controls (De Wilde & Lauwers, 1984). Other experiments with castrated males (Van Weerden & Grandadam, 1976) using implants with trenbolone and 17β -estradiol showed increased protein deposition and decreased fat deposition in the treated animals.

		Food Additives and Contaminants
	1	characteristics were not investigated because the aim of our experiment was to
	2	examine the effects of $17\beta NT$ on histology and residue levels in hair and urine.
	3	At slaughter there appeared a striking difference in the weight and size of the
	4	bulbourethral glands (Table 2), not only between barrows and boars, but also between
	5	control and treated barrows. Treated barrows had larger and significant $(p < 1*10^{-7})$
	6	almost 3 times heavier bulbourethral glands than the controls (fig. 2). Increase of the
	7	weight of male accessory glands is a well-known effect of androgens in castrated
	8	males (Brandes, D, 1994; Booth, W.D., 1890). This increased size could be used as a
	9	screening method for illegal treatment of barrows with androgenic hormones in the
1	0	slaughterhouse.
1	1	Insert Table 2 and figure 2
1	2	Histology
1	3	Histological investigation of the testes of the boars revealed normal spermatogenesis
1	4	as was expected in these animals. The prostate showed abundant glandular tissue and
1	5	the bulbourethral glands showed mature glandular tissue (fig. 3) consisting of
1	6	mucinous acini with secretion of Alcian Blue positive (acidic)secreta. The barrows
1	17	showed atrophied glandular tissue in the prostate and bulbourethral gland showing
1	8	immature glandular tissue presenting with rosette shaped alveoli in abundant stroma
1	9	(fig. 4) and scarce PAS positive secretion. The treated barrows showed an increased
2	20	amount of glandular tissue in the prostate. The bulbourethral gland presented with
2	21	mature glandular tissue (fig. 5), which stained blue with Alcian Blue PAS staining.
2	22	This means that treatment with $17\beta NT$ induced maturation of the glandular tissue of
2	23	the prostate and bulbourethral glands in barrows, causing histology resembling that of
2	24	boars. The results of the histological findings are in accordance with what Lauwers
2	25	(1984) described as androgen like changes, which he found in barrows treated with

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1	allyltrenbolone. In treated barrows he observed development of the accessory glands
2	in the physiological direction with more glandular tissue in the prostate and increased
3	secretion in the bulbourethral gland and seminal vesicals. In barrows implanted with
4	trenbolone acetate (López-Bote et al., 1994), no changes in carcass weight were
5	observed, but the implanted animals had less fat. They found enlargement of the
6	prostate and bulbourethral gland with increased secretion and cyst formation. Cyst
7	formation was not observed in our experiment, maybe because the animals were not
8	continually exposed to hormones and the difference in androgenic and anabolic action
9	between trenbolone and 17β NT.
10	Insert figures 3-5
11	Residue analysis in urine
12	All urines were analyzed for the presence of 17β NT. The urine was sampled at 4 time
13	points, corresponding with day 12, 33, 61 and 81 of the trial. Levels of $17\beta NT$ were
14	generally low (< 10 μ g/L) in barrows and treated barrows, except for one control
15	barrow that had a level of 300 μ g/L. It was not clear why this animal had such a high
16	level only at the last time point. No difference was observed between the $17\beta NT$
17	levels in the urine of the treated and control barrows. This may be due to the sampling
10	schodyle which was appear a month and approximately two washs after the day of

- 18 schedule which was once a month and approximately two weeks after the day of
- 19 treatment. In practice occasionally barrows and gilts are observed with high levels of
- 20 urinary 17β NT, but possibly after treatment levels are only high in urine for a few
- 21 days. In future experiments we have planned to investigate the kinetics of 17β NT in
- 22 urine from hour to hour and from day to day after treatment. Another possibility is to
- 23 examine the possible differences in metabolite profiles in treated and non-treated
- animals.

1	In boars high levels up to 400 μ g/L of 17 β NT were observed in urine, especially in
2	older animals (table 3). Analysis of urine was in this trial not able to differentiate
3	between treated and non-treated barrows, but most boars showed much higher levels
4	than barrows.
5	Insert table 3
6	Residue analysis in hair
7	In hair we measured the intact ester of $17\beta NT$ phenylpropionate. Hair was also
8	sampled at 4 time points. For this method a calibration series was used from
9	0 to 25 μ g/kg. All values above this series are estimated (table 4).
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11	In all non-treated barrows the levels were low <llq. <llq.<="" all="" boars="" levels="" showed="" th=""></llq.>
12	In non-treated animals no esters were expected in the hair, these results confirmed this
13	expectation. The treated animals showed from the start of the trial high levels $17\beta NT$
14	phenyl propionate in the hair, ranging from 15 to 220 μ g/kg, which increased in time
15	reaching a maximum level of 600 µg/kg (fig. 6).
16	
17	We conclude that analysis of 17β NT phenyl propionate in hair is a good method to
18	detect treated animals.
19	Insert table 4 and figure 6
20	
21	Conclusion
22	In this experiment we showed that $17\beta NT$ phenyl propionate is anabolic in barrows in
23	the dosage used and that this treatment yields increased size and weight of the
24	bulbourethral gland and profound histological alterations in the prostate en
25	bulbourethral gland making them resemble the glands of boars. Inspection of the

1	weight and histology of bulbourethral glands could be used as a screening method for
2	illegal treatment of barrows with androgenic hormones in the slaughterhouse.
3	Treatment of barrows with $17\beta NT$ phenyl propionate can be detected by hair analysis
4	for the intact $17\beta NT$ esters. More investigation is needed to evaluate the excretion of
5	17 β NT phenyl propionate in urine after treatment and the incorporation of 17 β NT
6	phenyl propionate in hair after treatment in barrows as well as in gilts.
7	
8	Acknowledgement
9	This project was financially supported by the Dutch Ministry of Economic Affairs,
10	Agriculture and Innovation.
11	
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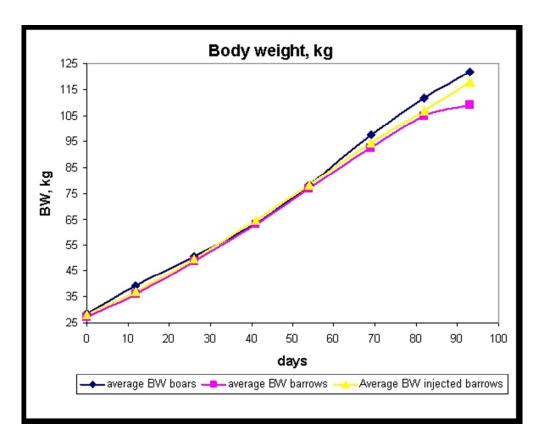
Table 1. Body weights at slaughter

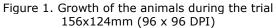
Body weight (kg)	Boar	Barrow	Barrow17β19NT
	116.4	103.2	113.4
	123.8	116.7	119.5
	119.2	114	120.1
	114.3	108.3	119.2
0	134.1	114.1	122.5
		97.5	112.3
mean	121.6*	109.0	117.8**
s.d.	7.8	7.4	4.0

* Boars significant heavier than barrows (p < 0.05)

** Barrow17βNT significant heavier (p <0.05) than control barrows

erent than boars Weights barrow17BNT not significant different than boars

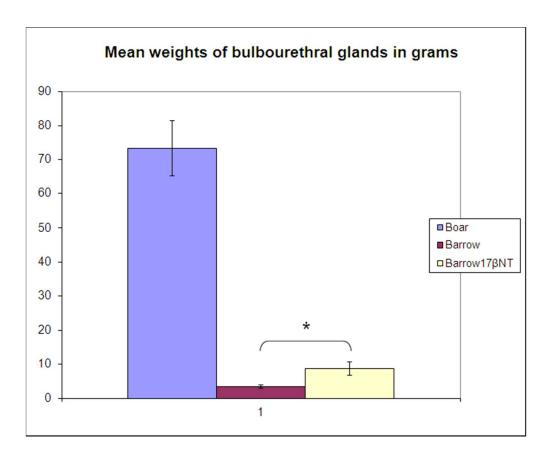




	Bulbourethral gland 1 (grams)	Bulbourethral gland 2
Control boar	69.9	65.5
Control boar	71.7	69.2
Control boar	84.4	90.7
Control boar	69.0	66.3
Control boar	75.7	70.6
Mean	73.3 ** ± 8.1	
Control barrow	*	*
Control barrow	4.1	3.9
Control barrow	2.3	3.3
Control barrow	3.6	3.4
Control barrow	3.3	2.8
Control barrow	3.9	3.2
mean	$3.4^{**} \pm 0.5$	
barrow17β19NT	9.5	10.5
barrow17β19NT	6.4	6.2
barrow17β19NT	9.6	10.3
barrow17β19NT	6.5	7.9
barrow17β19NT	7.0	8.2
barrow17β19NT	11.6	11.6
mean	8. 8** ± 2.0	
* means of the paired or	gans	

Table 2. Weights of the bulbourethral glands

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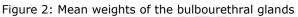


Table 3. Levels of 17β -19-nortestosterone in urine per animal

Concentration 17 β -19-nortestosterone (μ g/L)

(LLQ = $0.5 \mu g/L$ for pig urine)

Animal	Sex/	Time point 1	Time point 2	Time point 3	Time point 4
number	treatment	(prior to			
		treatment)			
1087	Barrow	35	1	5	300
6504	Barrow	2	2	8	3
7630	Barrow	n.d.*	1	20	10
7702	Barrow	n.d.	1	13	7
9975	Barrow	n.d.	3	5	10
9950	Barrow	n.d.	2	6	No sample
9373	Barrow+**	12	2	4	4
8111	Barrow+	n.d.	1	2	3
5133	Barrow+	n.d.	0.5	1	3
6185	Barrow +	n.d.	2	4	5
6559	Barrow+	n.d.	2	1	5
7177	Barrow+	7	3	3	2
1210	Boar	15	100	125	175
4473	Boar	185	165	150	2
7805	Boar	-	130	120	40
8775	Boar	55	140	130	100
9631	Boar	50	80	190	400

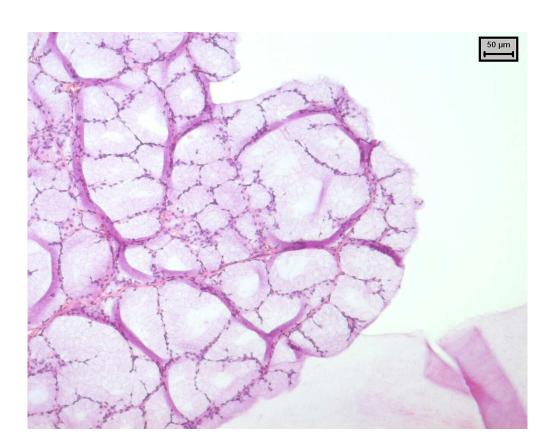
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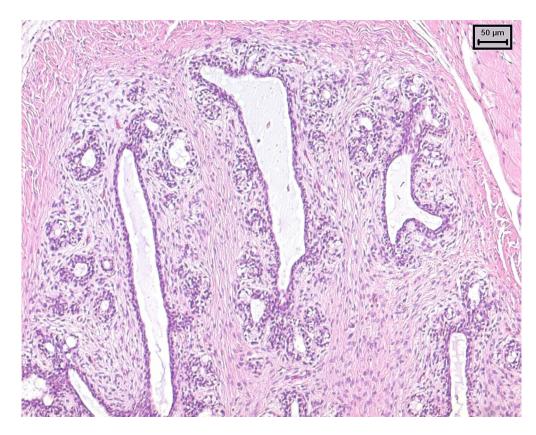
1	Table 4. Levels of 17β -19-nortestosterone phenylpropionate in hair per animal							
			Concentration 17β-19nortestosterone phenylpropionate					
			$(\mu g/kg)$, (LLQ = 2.5 $\mu g/kg$ for pig hair)					
	Animal	treatment	Time point 1	Time point 2	Time point 3	Time point 4		
	number		Prior or					
			treatment					
	1087	Barrow	< LLQ	< LLQ	< LLQ	< LLQ		
	6504	Barrow	< LLQ	< LLQ	< LLQ	< LLQ		
	7630	Barrow	< LLQ	< LLQ	< LLQ	< LLQ		
	7702	Barrow	< LLQ	< LLQ	< LLQ	< LLQ		
	9950	Barrow	< LLQ	< LLQ	< LLQ	< LLQ		
	9975	Barrow	< LLQ	< LLQ	< LLQ	< LLQ		
	8111	Barrow+*	< LLQ	17	240	330		
	9373	Barrow+	< LLQ	220	15	340		
	5133	Barrow+	< LLQ	15	60	200		
	6185	Barrow+	< LLQ	365	125	600		
	6559	Barrow+	< LLQ	13	175	265		
	7177	Barrow+	< LLQ	120	20	90		
	7805	Boar	< LLQ	< LLQ	< LLQ	< LLQ		
	8775	Boar	< LLQ	< LLQ	<llq< th=""><th>< LLQ</th></llq<>	< LLQ		
	9631	Boar	< LLQ	< LLQ	< LLQ	< LLQ		
	1210	Boar	< LLQ	< LLQ	< LLQ	< LLQ		
	4473	Boar	< LLQ	< LLQ	< LLQ	< LLQ		



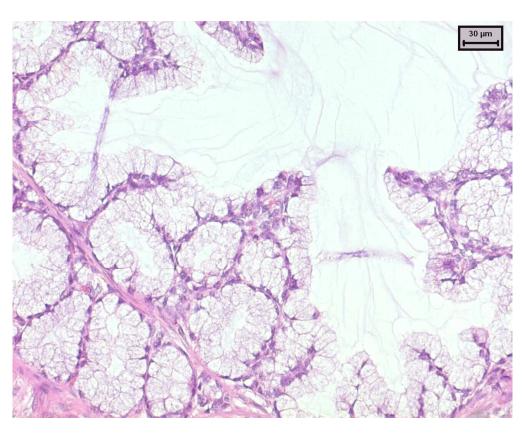
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Bulbourethral gland boar (HE), showing mature glandular tissue 343x272mm (96 x 96 DPI)



Bulbourethral gland barrow, showing immature glandular tissue (HE) 343x272mm (96 x 96 DPI)



Bulbourethral gland of a treated barrow, showing mature glandular tissue (HE). 343x272mm (96 x 96 DPI)

