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Food Additives and Contaminants



Identification and quantification of migration of chemicals from plastics baby bottles used as substitutes for polycarbonate

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Abstract:	This report presents the results of the study on the analytical identification and quantification of migration of chemicals from plastics baby bottles found in the EU market made of materials that are now present as substitutes for polycarbonate. 449 baby bottles with a focus on 1st age or sets of bottles were purchased from 26 EU countries, Canada, Switzerland, and USA. From this collection, which contained several duplicates, a total of 277 baby bottles were analysed. The materials included different types of plastic such as polycarbonate (PC), polyamide (PA), polyethersulfone (PES), polypropylene (PP), but also silicone, and from the USA a copolyester marketed under the trade name Tritan [™] . The bottles were subjected to the conventional migration test for hot fill conditions i.e. 2 hours at 70°C. The simulant used was that of specified in the EU legislation 2007/19/EC for milk, i.e. 50% EtOH. In a first phase one migration was conducted since the scope of this investigation was a screening rather than a true compliance testing check. Second and third migrations were performed on selected articles when migrated substances exceeded limits specified in the legislation. In order to verify some materials, a portion of the bottle was cut to run a FT-IR fingerprint to confirm the nature of the polymer. The migration solutions in general showed low release of substances. Results showed that bottles made of polypropylene and silicones showed a

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greater number of substances in the migration solutions and in greater quantity. Chemicals from polypropylene included alkanes, which could be found in >65% of the bottles at levels up to 3500 μ g/kg, and benzene derivatives in 17% of the baby bottles and found at levels up to 113 μ g/kg. Some substances were found on a regular basis such as plasticisers, esters, and antioxidants (e.g.tris(2,4-di-tert-butylphenyl)phosphate, known as Irgafos 168. Some substances found were not included in the Community positive list, which means that those should not be found even in the first migration. Such substances included 2,6-diisopropylnaphthalene (DIPN), found in 4% of the bottles at levels up to 25µg/kg, 2,4-di-tert-butyl phenol (in 90% of the bottles at levels up 400 µg/kg.). Moreover, Bisphenol A was detected and quantified in baby bottles made of polyamide, but limited to one brand and model specific (but labelled BPA-free). Results for baby bottles made of silicone also indicated the presence of components for example potentially coming from inks (benzophenone, diisopropyl naphtahalene - DIPN, which could come for example from and could be from the presence of instruction leaflets in the bottles). In the case of silicone, phthalates were also found in relevant concentrations, with levels for DiBP and DBP from the first migration test of 50-150 μ g/kg. and DEHP at levels 25-50 μ g/kg.

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Identification and quantification of migration of chemicals from plastics baby bottles used as substitutes for polycarbonate

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Abstract

The results of a study on the analytical identification and quantification of migration of chemicals from plastics baby bottles found in the EU market made of materials that are now present as substitutes for polycarbonate are reported. Four hundred and forty-nine baby bottles with a focus on 1st age or sets of bottles were purchased from 26 EU countries, Canada, Switzerland, and USA. From this collection, which contained several duplicates, a total of 277 baby bottles were analysed. The materials included different types of plastic such as polycarbonate (PC), polyamide (PA), polyethersulfone (PES), polypropylene (PP), but also silicone, and from the USA a co-polyester marketed under the trade name TritanTM. The bottles were subjected to the conventional migration test for hot fill conditions i.e. 2 hours at 70°C. The simulant used was that of specified in the EU legislation 2007/19/EC for milk, i.e. 50% ethanol. In a first phase one migration was conducted since the scope of this investigation was a screening rather than a true compliance testing check. Second and third migrations were performed on selected articles when migrated substances exceeded limits specified in the legislation. In order to verify some materials, a portion of the bottle was cut to run a FT-IR fingerprint to confirm the nature of the polymer. The migration solutions in general showed low release of substances. Results showed that bottles made of polypropylene and silicones showed a greater number of substances in the migration solutions and in greater quantity. Chemicals from polypropylene included alkanes, which could be found in >65% of the bottles at levels up to $3500 \,\mu g/kg$, and benzene derivatives in 17% of the baby bottles and found at levels up to 113 μ g/kg. Some substances were found on a regular basis such as plasticisers, esters, and antioxidants (e.g. tris(2,4-di-tert-butylphenyl)phosphate, known as Irgafos 168. Some substances found were not included in the Community positive list, which means that those should not be found even in the first migration. Such substances included

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2,6-di-isopropylnaphthalene (DIPN), found in 4% of the bottles at levels up to 25 μ g/kg, 2,4di-tert-butyl phenol (in 90% of the bottles at levels up 400 μ g/kg.). Moreover, bisphenol A (BPA) was detected and quantified in baby bottles made of polyamide, but limited to one brand and model specific (but labelled BPA-free). Results for baby bottles made of silicone also indicated the presence of components for example potentially coming from inks (benzophenone, diisopropyl naphtahalene – DIPN, which could come for example from from the presence of instruction leaflets in the bottles). In the case of silicone, phthalates were also found in relevant concentrations, with levels for DiBP and DBP from the first migration test of 50-150 μ g/kg. and DEHP at levels 25-50 μ g/kg.

Introduction

Baby bottles are widely used to feed newborns and infants. Baby bottles made of many more plastics materials than in the past have appeared on the market as a direct consequence of the ban of the use of Bisphenol A for the manufacture placing on the market of polycarbonate infant feeding bottles (EU 2011a, EU 2011b).

Plastic substitutes which have massively appeared on the EU market include for example polypropylene (PP), silicone, polyamide (PA), a new co-polyester under the trade name 'TritanTM', and polyether sulphone (PES). Yet for these materials, chemical substances such as monomers or additives may migrate from the plastic into the foodstuffs. Since infants have a lower bodyweight compared to adults, their exposure to the food in contact with the plastic material is higher and it is therefore particularly important to verify the absence of possible causes for health concern

Although there have been many studies on release from polycarbonate baby bottles from mild to harsh conditions as well as ageing (Hoekstra and Simoneau, 2011; Simoneau et al, 2011, Santillana et al, 2011; Kubwabo, et al, 2009; Maia et al, 2009; Biedermann-Brem et al, 2009; Biedermann-Brem et al, 2008; Cao et al, 2008; Maragou, et al, 2008; ; Ehlert et al, 2008; Brede et al, 2003; Biles et al, 1997; Mountfort et al, 1997), there is a major lack of data on the release of substances from the substitutes plastics now in use (Aschberger et al, 2010).

The question becomes whether these substitutes are also safe to use. The objective of this study was therefore to assess the nature of materials, chemicals, and potential release of

chemicals from baby bottles found in the EU market with the aim of informing risk managers on trends of evolution of the market and potential issues based on experimental data. Our study focused first on developing specific methods measuring all possible components migrating from the different plastics. The methods were then applied to the migration testing of baby bottles from different materials and different countries of the EU.

Materials and methods

Samples

 449 baby bottles or sets of bottles with a focus on first age were purchased or sent by the National Reference Laboratories (NRLs) mostly in EU Member States, as well as from Canada and the USA during the first half of 2010. From this collection, which contained several duplicates as well as baby cups, a total of 277 baby bottles were analysed. The materials of these baby bottles included different types of plastic such as Polypropylene (PP, n=149), Polyamide (PA, n=28), Polycarbonate (PC, n=58), Polyethersulfone (PES, n=30), a new co-polyester marketed under the trade name 'Tritan^{TM'} (n=7), silicone (n=5).

Chemicals

Ultrapure water was prepared from a Milli-Q purification system (Millipore); methanol, acetonitrile, HPLC grade quality \geq 99.95%, were purchased from Sigma Aldrich (Italy). Ammonium acetate, \geq 99.99%, was purchased by Aldrich (Italy). 2,2,4-Trimethyl pentane (isooctane) and dichloromethane for high-performance liquid chromatography were purchased by Sigma Aldrich (Italy). The following substances were used for GC-MS quantification: 1,2,4-trimethyl benzene (Fluka, \geq 99.5%), 1,2-dimethoxy benzene (Aldrich, >99%), 2,2,4-trimethyl-1,3-pentanediol-diisobutyrate (Eastman), 2,2-bis(4-hydroxyphenyl) propane (Aldrich, >99%), 2,4-di-tert-butylphenol (Aldrich, 99%), 2,6-di-tert-butyl-1,4benzoquinione (Aldrich, 99%), 2-butoxyethyl acetate (Aldrich, 99%), 2-ethyl-1-hexanol (Fluka, $\geq 99\%$), 2-methylnaphthalene (Fluka, analytical grade), 3,4-dimethylbenzaldehyde (Acros, 97%), benzophenone (Fluka, \geq 99%), benzothiazole (Aldrich, 96%), bis(2ethylhexyl)phthalate (Fluka, ≥97%), 2,6-bis(1,1-dimethylethyl)-4-methylphenol (Aldrich, \geq 99%), 2,6-bis(1,1-dimethylethyl)-4-methylphenol -D21 (CIL Inc, 98%), dibutyl phthalate (Aldrich, ≥99%), cis-9,10-epoxystearic acid (Sigma,99%), diethyl phthalate (Sigma Aldrich, 99.5%), diisobutyl phthalate (Fluka, ≥98%), 2,6- diisopropyl naphthalene (Acros, 99%), ndodecane (Sigma, 99%), erucamide (Aldrich, >97%), naphthalene (Fluka, \geq 97%), nonanal

(Acros, 95%), tris(2,4-di-tert-butylphenyl)phosphite (Aldrich, 98%), methyl tridecanoate (Sigma, \geq 97%).

Confirmation of the materials

FT-IR was used to confirm the nature of the material of the baby bottles since it was not always stated on the package or on the bottle. A Perkin Elmer FT-IR (Spectrum One) was applied using the attenuated total reflectance mode. ATR spectra were measured with a resolution of 4 cm-1 in the range of 4000-650 cm-1, each generated from four scans. Identification of the polymer was achieved based on comparisons with reference spectra.

Migration test

Since most manufacturers recommended sterilisation of the baby bottle prior to use (specified on the leaflet) the bottles were sterilised. The common treatment was selected as boiling in water for 5 min. The bottles were then subjected to the conventional migration test for hot fill conditions i.e. 2 hours at 70°C. The simulant used for milk was 50% ethanol as specified in the EU legislation 2007/19/EC. This was identified as the worst case simulant for this study since for juices, which would be modelled by acetic acid 3%, the age group of the infant would be older than the most sensitive population of newborn where the simulant for milk would apply. The simulant was pre-heated to 70°C, and a volume corresponding to the specified volume of the baby bottles was added to the item. The test specimens were covered with a glass plate weighted to ensure absence of evaporation and placed in a convection oven for 2 hours at 70 °C. The loss of simulant was measured in preliminary tests and was shown not to exceed 3%. Calibration curves were done in the 50% ethanol simulant in order to take recoveries into account. After treatment the migration solution was transferred from the baby bottle into a glass bottle with PTFE lined cap and left to cool down prior to analysis. An aliquot was then used in glass GC vial with PTFE lined cap for analysis. A second and third migration tests were performed according to the repeated use conditions for food contact material (82/711/EEC as amended). The bottles were quickly rinsed with deionised water between two migration tests. In a first phase only the first migration was analyzed to assess the nature and amount of migrants. In cases where substances from the first migration exceeded limits specified in the legislation, third migration solutions where also analysed to assess the final compliance of the articles. Blank samples were also prepared from a portion of the simulant used for the migration tests.

Analysis of migration solutions

The determination and quantification of the migrants in the simulant was carried out by GC-MS. Ultra performance liquid chromatography-mass spectrometry (UPLC-MS) and HPLC with fluorescence (FLD) detection were carried out for the detection of targeted compounds such as BPA. For GC-MS a change of solvent was necessary. In this case, 10 ml of the migration solution was extracted with 1 ml isooctane prior to injection.

GC-MS analysis

The isooctane extraction solutions were analysed by GC-MS (Hewlett Packard 5890 GC) coupled with a Hewlett-Packard mass selective detector (HP 5973 MSD). The instrument was operated with a HP-5MS phase column 30 m x 0.25 mm x 0.25µm using Helium as a carrier gas at a flow rate of 1.3 mL/min and a splitless injection. Injection volume consisted of 1 µL and injector block operated at 300°C. The oven programme was set at 50°C for 5 min, increased by 15°C/min to 300°C (1 min), and subsequently increased 25°C/min to 340°C and kept to this temperature to the end of the analysis. After each sequence a blank isooctane was run to clean the GC column between successive injections of samples and verify the absence of contamination or memory effect. GC-MS peak identification was performed by the Hewlett-Packard Enhanced ChemStation Software, mass spectra of the unknown compounds were matched with Wiley and NIST mass spectra libraries.

Deuterated 2,6-bis(1,1-dimethylethyl)-4-methylphenol-D21 (3.4 μ g/ml) was added to the isooctane used for the extraction of the food simulant, as internal standard, to potentially correct variations in the instrumental response. Quantification was done using different calibration curves of external standards that were prepared in 50% ethanol and subsequently extracted with isooctane to mimic the performance of the correspondent substances since not all the substances would have a 100% recovery (i.e. polar substance are poorly extracted by isooctane).

Over 70 compounds were recognised by GC-MS therefore individual quantification would be time consuming. Since the objective of this work was a screening of the potential release of different types of baby bottles, a selected number of standards were used for the quantification of either the target substance itself or substances of a similar molecular class and/or functional group.

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Substances were quantified using the following standards: erucamide, tris(2,4-di-tertbutylphenyl)phosphite (Irgafos 168),1,2,4-trimethyl benzene, nonanal, dodecane, 2-ethyl-1hexanol, 2-butoxyethyl acetate, 3,4-dimethyl benzaldehyde, benzothiazole, 2-methyl naphthalene, 1,2-dimethoxy benzene, 2,6-di-tert-butyl -1,4-benzoquinone, tridecanoic acid methyl ester, 2-ethylhexanoic acid (free fatty acid).

In addition, alkanes, alcohols, aldehydes, and aromatic compounds. were quantified as groups. For example, alkanes were calculated as sum of alkanes and were quantified on the basis of the response factor of dodecane, (as they went from C7 to C20). Alcohols were calculated as sum of alcohols and were quantified on the basis of the response factor of 2-ethyl-1-hexanol. Aldehydes were calculated as sum of aldehydes (except for 3,4-dimethyl benzaldehyde) and were quantified on the basis of the response factor of nonanal (or 3,4-dimethyl benzaldehyde). Benzene derivatives were calculated as sum of benzene substances, quantified on the basis of the response factor of 1,2,4-trimethyl benzene. The respective recoveries could vary with the different polarities of substances (higher recovery for non-polar substances).

Specific substances, which were detected on a regular basis and had a relevant SML, were quantified with the corresponding standard in Selected Ion Monitoring mode (SIM). These substances were: 2,4-di-tert-butylphenol, 2,2,4-trimethyl-1,3-pentanediol diisobutyrate (TXIB), 2,6-diisopropyl naphthalene, benzophenone, naphthalene, butylated hydroxytoluene (BHT).

For the analysis of phthalates the quantitative in SIM was used only as a screening to select samples in which the level of phthalates was above an expected random ambient contamination. In these cases (about 30 samples) the quantification was done in LC-MS.

HPLC and UPLC-MS

BPA was analysed from migration solutions using an HPLC (Hewlett Packard 1100) coupled with Fluorescence detector. The instrument was operated with a column phenomenex Kinetex C18, 150 mm x 3 mm x 2.6µm particle size, operated at a temperature of 30°C, using a flow rate of 0.27 ml/min and an injection volume of 20 µl. The fluorescence detection was operated with the excitation wavelength of λ exc = 235 nm and an emission wavelength of λ ems = 317 nm. The mobile phase was composed of methanol water in proportion (68:32) held for 10 minutes in isocratic conditions. The limit of detection (LOD) was 0.1 µg/kg and limit of quantification (LOQ) was 0.3 μ g/kg. The LOD was calculated based on a signal to noise ratio of 3, whereas the LOQ was based on 3 times the LOD.

The migration solutions were analysed for BPA and for phthalates by UPLC-MS (Waters Xevo) coupled with an electrospray interface in multiple reaction monitoring mode (MRM) using a column Waters Acquity BEH C18 (100 mm x 2.1 mm x 1.7 μ m particle size) operated at 30°C with a flow rate of 0.3 ml/min and an injection volume of 7.5 μ l.

BPA was analysed with electrospray ionization in negative ion mode (ES-) with a mobile phase of water ammonium acetate 1mM /methanol in proportion of (60:40) held for 0.3 min, followed by a gradient to 20:80 in 2.7 min., and held at the ratio of 20:80 for 0.5 min. The analysis was performed in multiple reaction mode (MRM) by monitoring the transition reactions m/z 227 \rightarrow 133 and m/z 227 \rightarrow 212 with a Cone voltage of 39. The LOD was 0.1 µg/kg and LOQ was 0.3 µg/kg.

For the detection of the group of phthalates, electrospray ionization in positive ion mode (ES+) was applied with a mobile phase of water ammonium acetate 1mM/methanol was applied in proportion of 37:63 held for 3.5 min, followed by a gradient to 23:77 in 2.5 min and subsequently another gradient to 5:95 in 1 min (held for 1.5 min). The analysis was performed in multiple reaction mode (MRM) by monitoring the transition reactions m/z $223 \rightarrow 149$ and m/z $223 \rightarrow 177$ with a Cone voltage of 17 for diethyl phthalate, m/z $279 \rightarrow 149$ and m/z $391 \rightarrow 149$ with a Cone voltage of 17 for diethyl phthalate.

Polymer dissolution in solvents

 Solvent dissolution was carried out on polypropylene baby bottles to verify the QM limits for phthalates. A sub-sample of the polymer (approximately 30mg), cut from the neck of the baby bottle, was added with 1 ml of xylene in a closed glass vial and heated in an oven at 135°C until the complete dissolution of the polymer. The vial was then cooled down to room temperature and 3ml of hexane were added to allow the precipitation of the polymer. The vial was centrifuged at 2500 rpm for 5 min and an aliquot of the solvent transferred into a 2 ml vial for the GC-MS analysis.

Results and discussion

The migration solutions in general showed low release of substances and differed not only between materials but also between brands of the same material. An overview of the amount of substances detected in the first migration from different types of baby bottles is given in Table 1. This table lists the migrants identified by GC-MS together with the quality index of recognition compared to a known spectra, and their specific migration limit (SML) when specified in the legislation. The quality index gives an estimation of the confidence that can be placed in the identification. Table 1 shows that all the detected compounds migrated in concentrations far below their corresponding SML. Additionally, a number of unregulated substances appeared in some cases in the first migration. Since these substances are not in the approved list, they should not be found migrating from the material in the first migration itself. The levels are however fairly low. The substances detected that were on the Community list were found mostly for baby bottles made of silicone, which are not classed as plastics and so this was not unexpected.

[Table 1 about here please]

Bottles made of polypropylene (PP)

The largest proportion of the EU market seems to consist of bottles made of polypropylene. In total, 149 bottles were investigated in this survey. These bottles tended to release a higher number of substances, such as alkanes, which could be found in >65% of the bottles at levels up to 3500 μ g/kg, and benzene derivatives in 17% of the baby bottles and found at levels up to 113 μ g/kg. Some substances were found on a regular basis such as plasticisers, esters, and antioxidants (e.g. tris-(2,4-di-tert-butylphenyl)phosphate, known as Irgafos 168. The migrated amounts of these compounds were compliant with the limits set in the EU food contact legislation. However, some substances found were not included in the EC positive list, such as 2,6-di-isopropylnaphthalene (DIPN)- a compound which is more typical for ink and/or recycled paper and board. This substance was found migrating in >45 % of the baby bottles at levels up to 25 µg/kg. Another substance found was 2,4-di-tert-butyl phenol. This substance was found fairly systematically (90%) in polypropylene baby bottles at levels up 400 μ g/kg. This could be a degradation product from tris-(2,4-di-tert-butylphenyl)phosphate, i.e. Irgafos 168. An interesting observation in some brands was the coupled migration of a group of fatty acid esters: octanoic acid, 2-ethylhexyl ester, decanoic acid, 2-ethylhexyl ester, dodecanoic acid, 2-ethylhexyl ester and tetradecanoic acid, 2-ethylhexyl ester. Figure 1 shows a chromatogram of a GC-MS analysis of the migration solution of a polypropylene baby bottle.

The migrated esters were semi-quantified using 1,2,4-trimethyl benzene as an external standard. The detected concentrations varied between 20 and 970 μ g/kg in the first migration round. Since all four esters are represented in the EC list with the maximum migration limit of 60000 μ g/kg (60 mg/kg), the obtained results regarding these esters are compliant with the European regulations.

[Figure 1 about here please]

However, polypropylene bottles also showed the presence of substances not present in the Community list. In particular, 2,2,4-trimethyl-1,3-pentanediol diisobutyrate was found in 72 samples out of 149 (close to 50%), in the range of 1-62 μ g/kg with an average of 5 μ g/kg. This substance is present in the EU plastics legislation with an SML of 5000 μ g/kg, but only for single-use gloves. Other substances identified and not present on the Community list included 2-propenamide, 2-methyl-N-phenyl, present in 7 out of 149 polypropylene bottles at levels of 90-132 μ g/kg with an average of 105 μ g/kg and N-butyl benzenesulfonamide, present in 3 out 149 bottles in range of 38-108 μ g/kg with an average of 64 μ g/kg.

A targeted analysis was performed for the phthalates specifically regulated in the food contact legislation (diethyl phthalate DEP, disobutyl phthalate DIBP, dibutyl phthalate DBP, and diethylhexyl phthalate DEHP) on bottles that had shown traces in the migration solutions. The reason was that for phthalates, the EU legislation imposes a limit based on a residual content know as Quantity in Material (QM) for compliance testing. The results from a dissolution reprecipitation of the polymer and analysis of the extract showed the levels obtained were much below the respective QMs of 0.1% mg substance per kg of material (DEP, DiBP, DEHP) and 0.05% mg substance per kg of material (DBP).

Bottles made of a copolymer marketed under the name TritanTM

This material has become popular in the USA and all bottles investigated in this study were purchased in the USA. It has been now authorised by Regulation EU No 10/2011 (EU 2011a) as of 1 May 2011. Some baby bottles were obtained from the USA for scoping purposes in foresight of the foreseen market expansion to Europe upon authorisation. The migration testing from the 7 bottles tested of this material did not reveal detectable substances.

Bottles made of Polyamide (PA)

28 PA baby bottles were analysed, of which 27 were from one brand. The most surprising finding of the analysis of migration solutions from PA baby bottles was the identification and subsequent quantification of BPA on bottles labelled as "no Bisphenol A". BPA was detected and quantified (both by HPLC and UPLC-MS) in 20 out of 27 bottles analysed, i.e. 77% (Table 2) in the specific brand.

[Table 2 about here please]

Six bottles released BPA in the range of 0.5-4 μ g/kg, seven in the range of 15-70 μ g/kg, six in the range of 100-250 μ g/kg and one bottle released even more than 1000 μ g/kg in the first migration. In the third migration test significant amounts of BPA could still be found in these bottles (1-330 μ g/kg). These values are much higher than those found in the same survey from polycarbonate bottles (Simoneau et a, 2011).

The recognition of BPA as the migrated substance from polyamide baby bottles was confirmed by LC-MS fragmentation spectra. Daughters ions of the precursor at m/z 227 [M-H]- corresponded to the ions obtained from the fragmentation of a pure standard of bisphenol A (Fig. 2). The fragmentation pathway of BPA confirmed the results in literature (Gallart-Ayala et al, 2007).

[Figure 2 about here please]

Since it was surprising to detect BPA in an article labelled as polyamide a FT-IR analysis was also performed to verify that the material was not polycarbonate (Fig. 3)

[Figure 3 about here please]

The spectra illustrated that the material was not polycarbonate. According to Commission Regulation (EU) No 10/2011 of 14 January 2011 and its amendments (EU 2011a; EU 2011b) BPA can no longer be used as an additive or as a monomer for the manufacture of polycarbonate infant feeding bottles. However when used as a monomer in materials other than polycarbonate it is still subjected to a specific migration limit of 600 μ g/kg. In the context of this study it is not possible to determine whether BPA was used as an additive (not

 allowed) or a monomer (subjected to the SML), although some studies in other context (RA, 2003, Brede et al, 2003) could suggest that its presence in PA would be more likely to result as that of an additive and the release seen is relatively higher than the one seen for example from PC when used as monomer (300 - 600 times higher).

In order to eliminate doubts concerning identification and quantification, a blind confirmation was conducted with two independent laboratories. Samples of CH001A, CH001B, CH004, CH009A, CH009B, CH009C, CH010C, CH015A, CH017, NL003A, NL008A, NL008B were sent blind to both ThermoFisher Scientific and the UK Food & Environment research Agency (FERA). In addition, baby bottles already tested for migration CH009A and CH015A and virgin bottle CH015B were sent to FERA to conduct independent migration. The results from the simulants solutions confirmed our results, with non-detectable found for samples CH010C and NL008A, and concentrations within a range of 25% of those we found. The migration test conducted on bottles CH009A gave a migration of 500 μ g/kg, whereas the one conducted on bottle CH015A gave 22 µg/kg. These results were higher than those we found in the third migration (but lower than those we found in our first migration), suggested that the bottles had recuperated a capacity for release in the lag time of about 4 months. The migration test of the virgin bottle CH015B) gave 70 and 12 μ g/kg respectively in the first and third migration. These results were also close to another of our results for bottle CH016 (same brand and model) giving 74 and 14 μ/kg in the first and third migration respectively (table 2). Overall, the results for the PA baby bottles suggested not only a seemingly abnormal presence of BPA for this type of polymer but also an illicit one as the bottles were labelled 'NO Bisphenol-A". Table 2 shows that baby bottles containing BPA were of the same brand, but in particular 2 models mostly available in Switzerland and the Netherlands, with larger migration seen on the smaller volume (125ml) bottles.

It should also be noted that other substances migrated from the investigated bottles made from this type of polyamide. In particular, cyclododecene which is not included in the EC positive list, was found in nearly all (85%) of the bottles investigated, in concentrations from 10-60 μ g/kg. It should be noted that this substance was not found in other materials. Studies indicate that cyclododecene can be an intermediate for the synthesis of monomers used in the manufacture of polyamide, co-polyamides and polyesters (Adams et al, 2007). DIPN (and naphthalene) were also found in respectively 6 out of 28 bottles (21%) for DIPN and 7 out 28 bottles for naphthalene, but in very low levels 1-2 μ g/kg. Again for these substances more

 common to paper and board, the cause could be from the presence of leaflets inside the bottles since benzophenone was also present in some of the bottles under consideration.

Bottles made of Silicones

Although silicone articles are not regarded as "plastic materials and articles" as laid down by the EU Framework Regulation 1935/2004 of plastic materials in contact with foodstuffs, preliminary results were obtained from 5 samples purchased and analysed since this material can be found in the EU market. This material generally showed migration of a greater number and extent substances than polypropylene, as shown as Figure 3.

[Figure 3 about here please]

The most striking finding was the presence of phthalates in the migration solutions, more specifically diisobutyl phthalate (DiBP), dibutyl phthalate (DBP) and diethylhexylphthalate (DEHP). The levels for DiBP and DBP from the first migration test were in the range 50-150 μ g/kg. DEHP was found to a lesser extent at levels 25-50 μ g/kg.

All bottles also released what could be considered ink or cardboard related substances such as DIPN, benzophenone, and naphthalene. DIPN in particular (not regulated for plastics) could be found in all the baby bottles analysed, at levels of 5-20 μ g/kg. Finally all the bottles also released a significant number of aldehydes (not found in any of the other bottles investigated with an exception of a few bottles made from polypropylene), to level up to 700 μ g/kg in the first migration which remained to 60 μ g/kg at the third migration.

Due to the ban of polycarbonate bottles it may be expected that more silicone bottles could be one of the choices for baby bottles on the European market. There is therefore a need to further investigate this material, especially in view of the suspected endocrine disrupting characteristics of phthalates.

Bottles made of Polyethersulphone (PES)

The migration solutions of 30 polyethersulphone bottles were analysed to determine the presence of other migrants. Generally, migration solutions showed a very low release (less than $150 \ \mu g/kg$) of mostly one or two compounds but the majority of the analysis indicated no migration at all. However, one brand released more compounds than the average

polyethersulfone article. From these articles a noticeable amount of benzophenone migrated. Since these concentrations were higher than the SML (600 μ g/kg) a second and third migration test were performed. Analysis of the third migration indicated the level of benzophenone had decreased within regulatory limits. Remarkably, these articles contained a piece of cardboard when purchased. This implies the benzophenone could have migrated from the polymer as well as from the cardboard. The bottle was dissolved with DCM and reprecipitated with methanol but the resulting extract showed only trace amounts of benzophenone, which seems to confirm the hypothesis of a contamination by the board. Furthermore, the migration from these typical polyethersulfone bottles seemed to indicate the presence of a substance identified as close to decahydro-4,4,8,9,10-pentamethylnaphthalene at levels between 12 and 42 μ g/kg in the simulant after the third migration. As the quality index was below 60, the confirmation of identity of the substance needs further investigations since as such, it is not listed in the EC positive list.

In a previous study (Simoneau et al, 2011) migration of diphenyl sulphone, 4,4'dihydroxydiphenyl sulphone (Bisphenol S) and 4,4'-dichlorodiphenyl sulphone was specifically measured by UPLCS-MS and HPLCPLHP. The data obtained showed that the migration of all 30 bottles remained much below their migration limits of 3000 μ g/kg for diphenyl sulphone and 50 μ g/kg for Bisphenol S and 4,4'-dichlorodiphenyl sulphone after the first migration test. According to our results, all the analyzed baby bottles made of PES were therefore compliant with the European regulations concerning the migration of Bisphenol S, diphenyl sulphone and 4,4'-dichlorodiphenyl sulphone.

Bottles made of Polycarbonate (PC)

In total, 58 baby bottles of Polycarbonate were investigated to verify the presence of BPA and other migrated compounds. The results of the release of BPA from polycarbonate bottles are presented in a previous study (Simoneau et al, 2011). Another 18 bottles were also analysed by GC-MS to investigate migration of other compounds. These results did not show any noticeable release of compounds from polycarbonate baby bottles.

Conclusion

 Overall, the findings with respect to the different materials led to the following conclusions: No issue was found for bottles made of polyethersulphone nor for the polymer TritanTM in view of release of chemical substances. For the latter, the survey sample size was limited and

may need further monitoring once reaching the EU market. A significant issue for polyamide bottles was identified as the migration of BPA in relevant amounts. Bottles made from polypropylene showed substances which are not on the positive list, and in particular DIPN present in about 45 % of the 149 bottles; a significant issue might arise from the presence of silicone baby bottles on the EU market due to the migration of phthalates

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 Table 1: Migrants from different type of baby bottles detected by GC-MS (with index of recognition >80%)

				Polypropylene (n=149)			Polyamide (n=28)			Silicone (n=5)		
Migrant	CAS	Quality	SML (mg/kg)	n	range (µg/kg)	average (µg/kg)	n	range (µg/kg)	average (µg/kg)	n	range (µg/kg)	average (µg/kg)
2,2,4-trimethyl-1,3-pentanediol diisobutyrate (TXIB)	6846-50-0	100	5 ¹	72	1-62	5	3	2	2	5	14-574	159
2,2-bis(4-hydroxyphenyl)propane (BPA)	80-05-7	100	0.6 ²				17 ³	1-1005	106			
2,4,6-trimethyl benzaldehyde	487-68-3	90	n/a							1	94	
2,4-dimethyl benzaldehyde	15764-16-6	87	60	6	21-62	47				1	94	
2,4-di-tert-butyl phenol	96-76-4	100	60	133	1-419	49	1	3		4	26-221	103
2,6-diisopropylnaphthalene (DIPN)	24157-81-1; 38640-62-9	100	n/a	69	1-23	5	6	1-2	1	5	4-23	12
2,6-di-tert-butyl-p-benzoquinone	719-22-2	97	60							2	341-382	361
2-butoxyethyl acetate	112-07-2	86	60	6	139-778	354						
2-methylnaphthalene	91-57-6	80	60	1	3		1	3				
2-propenamide,2-methyl-N-phenyl	1611-83-2	86	n/a	7	90-132	105						
2-propenoic acid,3-(-4-methoxyphenyl)- 2-ethylhexyl ester	5466-77-3	95	n/a							1	175	
2-propenoic acid-2-ethylhexyl ester	103-11-7	86	0,05							3	24-72	76
3,5,5-trimethyl-2-cyclohexen-1-one	78-59-1	91	60							1	88	
4-ethylbenzaldehyde	4748-78-1	91	60							1	92	
4-tert-butylcyclohexyl acetate	32210-23-4	90	n/a							1	145	
9-octadecenamide (oleamide)	301-02-0	93	60	2	227-2487	1357						
9-octadecenoic acid, methyl ester	112-62-9	99	60	12	18-294	65						
acetic acid, 2-ethylhexyl ester	103-09-3	86	60							4	23-76	51
alpha-methylstyrene	98-83-9	86	0.05									
benzaldehyde-4-methylthio	3446-89-7	90	n/a							1	33	
benzoic acid, 4-ethoxy-ethyl ester	23676-09-7	91	3,6							1	23	
benzophenone	119-61-9	100	0.6	39	1-286	43	2	2	2	5	11-637	184
butylated hydroxytoluene (BHT)	128-37-0	100	3	14	1-156	44				3	1-3	2
camphor	76-22-2	96	60^{4}							3	18-147	87
cyclododecene	1501-82-2	97	n/a				17	6-63	29			
cyclohexanone	108-94-1	93	60							1	109	

Migrant cyclohexanone-5-methyl-2- (1-methylethyl) 104: decanoic acid 2-ethylhexyl ester 739: dodecanoic acid 1-methylethyl ester 102: dodecanoic acid 2-ethylhexyl ester 202:	CAS 458-14-7 947-30-5 233-13-3 292-08-4	Quality 86 83	SML (mg/kg) 60	n	range (µg/kg)	average (µg/kg)	n	range (µg/kg)	average (µg/kg)	n	range (µg/kg)	average (µg/kg)
cyclohexanone-5-methyl-2- (1-methylethyl)104:decanoic acid 2-ethylhexyl ester739-dodecanoic acid 1-methylethyl ester102:dodecanoic acid 2-ethylhexyl ester2029	458-14-7 947-30-5 233-13-3 292-08-4	86 83	60									
decanoic acid 2-ethylhexyl ester 739- dodecanoic acid 1-methylethyl ester 102- dodecanoic acid 2-ethylhexyl ester 2029-	947-30-5 233-13-3 292-08-4	86 83	60							2	20-128	74
dodecanoic acid 1-methylethyl ester102dodecanoic acid 2-ethylhexyl ester202	233-13-3 292-08-4	83	00	8	28-170	113						
dodecanoic acid 2-ethylhexyl ester 2029	292-08-4		60	16	29-3131	570				3	356-1464	739
		86	60	9	15-1220	692						
dodecanoic acid, methyl ester 111-	1-82-0	91	60							1	45	
erucamide 112-	2-84-5	100	60	9	138-791	303				2	73-160	116
eucalyptol 470-	0-82-6	91	60	2	237-391	314				2	149-589	369
hexadecanoic acid 57-1	-10-3	99	60	7	1590-2965	2347				1	9540	
hexadecanoic acid methyl ester 112-	2-39-0	97	60	7	19-98	49						
hexadecanoic acid,2-hydroxy-1- (hydroxymethyl) ethyl ester 234	470-00-0	91	60	8	21-208	49						
methyl 3(3,5-di-tert-butyl-4- hydroxyphenyl)propionate 6380	86-38-5		60	38	15-34	23				3	29-52	37
napthalene 91-2	-20-3	100	60	27	1-11	2	7	1-5	2	4	2-4	3
N-butyl benzenesulfonamide 362	22-84-2	94	n/a	3	38-108	64						
octadecanoic acid 57-1	-11-4	99	60	5	792-10416	3449				1	2256	
octadecanoic acid, ethyl ester 1111-	1-61-5	97	60	8	17-47	25						
octanoic acid,2-ethylhexyl ester 6332	321-70-0	90	60	8	38-235	138						
tetradecanoic acid 2-ethylhexyl ester 298	806-75-5		60	4	42-59	53						
tris(2,4-di-tert-butylphenyl)phosphite 315' (Irgafos 168)	570-04-4	100	60	132	4-1884	144				1	36	
benzenes				26	1 1 1 2	26				5		
alcohols				5	72 1030	20 450					303 1563	668
alconois				5	16 140	4J9 54				4	34 676	123
				9	10-140	34				5	54-070	123

1: To be used in single-usegloves only; 2: Not to be used for the manufacture of polycarbonate infant(*) feeding bottles(**); with: (*) Infant as defined in Article 2 of Directive 2006/141/EC and (**) This restriction is applicable from 1 May 2011 as regards the manufacture and from 1 June 2011 as regards the placing on the market and importation into the Union. SML applicable for the use as monomer only; 3: Total of 28 polyamide baby bottles were analysed by HLPC to verify the presence of BPA; 4: Warning: there is a risk that the migration of the substance deteriorates the organoleptic characteristics of the food in contact; *: in this case the standard used was that of the substance itself, making the identification 100%.

n/a: not applicable: "substances not included in the european legislation)"

Table 2: identification	of BPA i	n nolvamide using	HPLC and	UPLC-MS.
Table 2. Inclution	ULDI / L	i poryannuc using		ULC-MD.

product name	labelling	volume	sample	ppb (UPLC) 1st migration	ppb (HPLC) 1st migration	ppb (HPLC) 3rd migration
Brand A	no bisphenol A	125	AT003	nd	nd	nd
	no bisphenol A	250	NL009	nd	nd	nd
Brand B model	no bisphenol A	150	CH010 A	nd	nd	nd
1	no bisphenol A	150	CH010 B	nd	nd	nd
	no bisphenol A	150	CH010 C	nd	nd	nd
Brand B model 2	no bisphenol A	150	BE009	nd	nd	nd
Brand B model 3	no bisphenol A	250	CH017	nd	3	nd
~	no bisphenol A	250	CH004	7	3	nd
Brand B model	no bisphenol A	250	CH013 A	nd	nd	nd
	no bisphenol A	125	CH014	29	29	6
· ·	no bisphenol A	125	CH015A	36	30	7
	no bisphenol A	125	CH016	83	74	14
	no bisphenol A	250	CH011 A	nd	1	nd
	no bisphenol A	250	CH011 B	nd	1	na
	no bisphenol A	250	CH011 C	nd	1	na
	no bisphenol A	250	NL003 A	31	26	nd
_	no bisphenol A	250	NL003 B	2	1	nd
-	no bisphenol A	250	NL008 A	5	nd	nd
Durand Durandal	no bisphenol A	250	NL008 B	36	29	6
Brand B model	no bisphenol A	250	NL011 A	2	4	1
э - -	no bisphenol A	125		102	119	27
		125	CH001 R	103	129	29
		125	CH001 C	23	125	<u> </u>
	no bisphenol A	125	CH009 A	1000	1005	329
	no bisphenol A	125	CH009 B	268	262	127
	no bisphenol A	125	CH009 C	164	147	83
	no bisphenol A	125	CH018	167	136	31
Note: nd: non de	etectable; na: not analy	vsed				



Figure 2: MS-MS spectra of BPA from baby bottle CH009A-PA (upper spectrum) and BPA standard (lower spectrum)



