

# Metal immobilization and soil amendment efficiency at a contaminated sediment landfill site: a field study focusing on plants, springtails, and bacteria

Valérie Bert, Christine Lors, Jean-François Ponge, Lucie Caron, Asmaa Biaz, Marc Dazy, Jean-François Masfaraud

# ▶ To cite this version:

Valérie Bert, Christine Lors, Jean-François Ponge, Lucie Caron, Asmaa Biaz, et al.. Metal immobilization and soil amendment efficiency at a contaminated sediment landfill site: a field study focusing on plants, springtails, and bacteria. Environmental Pollution, 2012, 169, pp.1-11. 10.1016/j.envpol.2012.04.021. hal-00704382

# HAL Id: hal-00704382 https://hal.science/hal-00704382v1

Submitted on 5 Jun2012

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Public Domain

1 Environmental Pollution
---------------------------

2	Full Research Pap	er
---	-------------------	----

- 3 **Title:** Metal immobilization and soil amendment efficiency at a contaminated sediment
- 4 landfill site: a field study focusing on plants, springtails, and bacteria

## 5 Author names and affiliations:

- 6 Valerie Bert<sup>a, b,\*</sup>, Christine Lors<sup>b, c, d</sup>, Jean-François Ponge<sup>e</sup>, Lucie Caron<sup>a</sup>, Asmaa Biaz
- <sup>e</sup>, Marc Dazy<sup>f</sup>, Jean-François Masfaraud <sup>f</sup>
- <sup>a</sup> INERIS, RISK, DRC, Technologies et Procédés Propres et Durables, Parc
- 9 Technologique Alata, BP2, F-60550 Verneuil en Halatte, France
- <sup>b</sup> Centre National de Recherche sur les Sites et Sols Pollués, BP 537, 59505 Douai
- 11 Cedex, France
- <sup>c</sup> Université Lille Nord de France, 1 bis rue Georges Lefèvre, 59044 Lille Cedex, France
- <sup>d</sup> Ecole des Mines de Douai, LGCgE-GCE, 941, rue Charles Bourseul, 59500 Douai
   Cedex, France
- <sup>e</sup> Muséum National d'Histoire Naturelle, CNRS UMR 7179, 4 avenue du Petit-Château,
- 17 91800 Brunoy, France
- <sup>f</sup> Université Paul Verlaine-Metz, Laboratoire "Interactions Ecotoxicologie, Biodiversité,
- 19 Ecosystèmes", CNRS UMR 7146, Campus Bridoux, rue du général Delestraint, 57070
- 20 Metz, France

21

- **\*Corresponding author.** Tel.: +33 3 44 55 63 82; fax: +33 3 44 55 65 56.
- 23 E-mail address: <u>valerie.bert@ineris.fr</u> (V. Bert).

24 Capsule

25 In-situ incorporation of Thomas Basic Slag into a landfilled metal-contaminated

sediment decreases metal mobility and ecotoxicity and increases bacterial activity.

27

#### 28 Abstract

29 Metal immobilization may contribute to the environmental management strategy of

30 dredged sediment landfill sites contaminated by metals. In a field experiment,

31 amendment effects and efficiency were investigated, focusing on plants, springtails and

32 bacteria colonisation, metal extractability and sediment ecotoxicity. Conversely to

33 hydroxylapatite (HA, 3 % DW), the addition of Thomas Basic Slag (TBS, 5 % DW) to a

34 5-yr deposited sediment contaminated with Zn, Cd, Cu, Pb and As resulted in a decrease

in the 0.01M Ca(NO<sub>3</sub>)<sub>2</sub> extractable concentrations of Cd and Zn. Shoot Cd and Zn

36 concentration in *Calamagrostis epigejos*, the dominant plant species, also decreased in

the presence of TBS. The addition of TBS and HA reduced sediment ecotoxicity and

improved the growth of the total bacterial population. Hydroxylapatite improved plant

39 species richness and diversity and decreased antioxidant enzymes in *C. Epigejos and* 

40 Urtica dioica. Collembolan communities did not differ in abundance and diversity

41 between the different treatments.

42

Keywords : basic slag; *Calamagrostis epigejos*; dredged sediment; hydroxylapatite;
ecotoxicity

45

#### 47 **1. Introduction**

Human activities during the last decades have contaminated canal sediments 48 49 with various organic and inorganic pollutants. Those of most concern are metal(loid)s, 50 polyaromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs), and mineral 51 oils. Polluted sediments in canals may be disposed on land. Indeed, the maintenance of 52 waterways requires dredging on a regular basis to prevent flooding, facilitate 53 navigation, and allow for use of a given water system. European Community policy encourages sediment recovery (Directive 2008/98/EC). Nevertheless, due to the high 54 55 concentration of pollutants and their potential toxicity, contaminated dredged sediments cannot be used in civil engineering as raw material or the deposit cannot be used to 56 produce valuable biomass. Many treatments are available for contaminated sediments, 57 but relatively few are applicable to metal(loid) pollution. Currently, treatment and reuse 58 of heavily contaminated dredged materials is not a cost-effective alternative to disposal 59 60 landfill sites (Bert et al., 2009). In the Nord-Pas-de-Calais region (France), which is 61 affected by intensive industrial activities, local authorities are required to manage contaminated landfill sites where large volumes of polluted dredged materials were 62 63 deposited. The regional division of Voies Navigables de France (VNF) developed a management strategy for its disposal sites. This strategy includes the implementation of 64 65 an environmental management system which aims to meet best practices and comply with environmental regulation in the field of human health and environment. The VNF 66 is involved in the reclamation of its disposal sites into 'natural' and 'green' zones 67 68 (Prevost, 2008). Metal immobilization in metal-contaminated sediments at some landfill sites may contribute to this environmental management strategy. 69

70 Metal immobilization aims at (1) changing speciation of trace elements (TE) in the substrate to reduce their soluble and exchangeable fractions, (2) limiting TE-uptake 71 72 by plants, (3) reducing the direct exposure through soil by reducing metal availability to heterotrophic organisms, resulting in enhanced biodiversity (Vassilev et al., 2004). In 73 situ immobilization of metal(loid)s is achieved by incorporating amendments into the 74 75 soil, promoting their sorption and precipitation. In addition to immobilization effects, soil amendment can improve soil fertility by increasing pH, organic matter content, 76 77 microbial activity, and moisture retention, and reducing soil compaction (Vangronsveld 78 et al., 1995, 1996).

The efficiency of amendments is site-specific, depending on various factors 79 80 among which pollutant types and soil properties (Kumpiene et al., 2008). Numerous soil amendments have been tested in pots, at pilot and field scale, for remediation purposes 81 (Bes and Mench, 2008; Knox et al., 2001; Kumpiene et al., 2008). The effects of 82 Thomas Basic Slag (TBS) and hydroxylapatite (HA) incorporation have only been 83 investigated in batch and pot experiments (Bes and Mench, 2008; Boisson et al., 1999; 84 85 Friesl et al., 2006; Mench et al., 1994a, 1994b; Mench et al., 2000; Misra and 86 Chaturvedi, 2007; Negim et al., 2010; Panfili et al., 2005). Hydroxylapatite, a mineral from the phosphate group, is the major component of tooth enamel and bone mineral. It 87 88 is mainly used for medical purposes. Hydroxyapatite powders can be synthesised via numerous production routes, using a range of different reactants. Some processing 89 90 techniques include wet chemical methods (precipitation), hydrothermal techniques, 91 hydrolysis of other calcium phosphates and sol-gel. Due to its particular properties 92 including the sorption of metallic ions, HA can be useful for the management of contaminated groundwater and soil. The application of HA [Ca<sub>10</sub>(PO<sub>4</sub>)<sub>6</sub>(OH)<sub>2</sub>] to 93

contaminated soil immobilizes dissolved Pb (Ma et al., 1993). Other studies on metal 94 stabilization by phosphorous compounds focused on reduction of plant uptake of 95 metals, as well as reduction of their solubility and mobility (Knox et al., 2001). After 96 HA incorporation into a contaminated soil, the concentrations of exchangeable metals 97 decreased and plant uptake of these elements was reduced (Boisson et al., 1999). 98 However, arsenic uptake by plants increased and there were nutrient deficiencies. 99 Therefore it was concluded that HA application could be effective to immobilize Zn, 100 101 Pb, Cu and Cd but was inappropriate in the case of mixed metal-arsenic pollution and 102 when potential nutrient deficiencies may occur. Thomas Basic Slag (TBS) is an alkaline by-product of steel industry used as a fertilizer by farmers. It usually increases soil pH 103 104 (Bes and Mench, 2008) and contains P, Mn, Mg, and Fe (Panfili et al., 2005). Soil treatment with TBS decreased Cd and Zn-soluble and  $(Ca(NO_3)_2)$  exchangeable 105 fractions and reduced Cd concentrations in tobacco shoots (Mench et al., 1994a, 1994b). 106 Zinc availability decreased after TBS addition and this persisted over 5 months without 107 any phytotoxicity (Mench et al., 2000). Bes and Mench (2008) studied the incorporation 108 109 of 0.25% and 3.9% (DW) TBS into Cu-contaminated soils. Based on soil phytotoxicity, 110 3.9% TBS was one of the most efficient amendments even though high levels of Cu remained in the soil solution. Panfili et al. (2005) studied the effect of HA and TBS on 111 112 Zn in a metal-contaminated sediment and suggested that the formation of Zn phosphate contributed to Zn immobilization. 113

114

115

Plants, bacteria and soil fauna, notably springtails (Collembola), can develop tolerant populations on metal-contaminated soils (Bes et al., 2010; Escarré et al., 2011;

116 Gillet and Ponge, 2003; Lors et al., 2005; Ryan et al., 2004; Tyler et al., 1989;

117 Vangronsveld et al., 1995, 1996). Vegetation reduces contaminant mobility by reducing

118 leaching, soil erosion, and improves the aesthetic value of formerly barren areas (Ruttens et al., 2006). The vegetation itself may contribute to metal immobilization in 119 120 the rhizosphere and through the production of litter (Bouwman and Vangronsveld, 2004). When metal bioavailability and exposure decrease, due to vegetation or 121 amendment or both, sensitive plants, bacteria and soil fauna can develop and participate 122 in turn to the restoration of an ecosystem on contaminated soils (Bouwman et al., 2001; 123 Lock et al., 2003; Lock and Janssen, 2003, 2005; Vangronsveld et al., 1995, 1996). 124 125 This field-scale work was performed at an experimental metal-contaminated 126 sediment landfill site. The aim was to evaluate the efficiency of TBS and HA incorporation on metal(oid) immobilization. Efficient soil amendments may decrease 127 128 metal(oid) extractability and shoot metal concentrations. In parallel, the potential adverse or beneficial effects of TBS and HA incorporation were investigated on an 129 130 array of biological parameters related to plant and springtail communities composition and diversity, total and metal specific bacterial populations (anaerobic sulphate-131 reducing bacteria and aerobic sulphur-oxidising bacteria), springtail trophic status (gut 132 133 contents), sediment ecotoxicity and sublethal effects in plants (maximum photochemical 134 efficiency of photosystem II and antioxidant enzymes activity levels). This battery might be relevant to assess the efficiency of soil additives. 135

This is the first time that a full-scale study of metal immobilization combined
with ecosystem monitoring is reported on a contaminated dredged sediment disposal
site.

139

140 2. Materials and methods

141 2.1. Description of experimental site and sediment treatment

142 In May 2002, a field trial was set up in an agricultural area in Lallaing (North of France, 50°23'17''N and 3°11'59''E). Three plots of 60 m<sup>2</sup> and 40 cm depth were dug 143 144 in an uncontaminated soil. These plots were filled with freshly dredged sediments from 145 the nearby Scarpe canal (Pont de Râches). These sediments were contaminated due to 146 past and present non-ferrous metal processing and smelting activities. Two plots were used to assess soil additives. Thomas Basic Slag (TBS) and a synthetic hydroxylapatite 147 (HA), both in powdered forms, were singly incorporated into the sediment at a rate of 148 5% and 3% DW, respectively. TBS was obtained from Cedest® (Mâcon, France) and 149 HA was from Brenntag<sup>®</sup> (Mülheim/Ruhr, Germany). The purity of HA was certified to 150 99% by the supplier. The third plot remained untreated (NT). After TBS and HA 151 152 incorporation, the treated and untreated plots were mechanically homogenized with a 153 crane shovel for two hours to ensure that the mixture was homogeneous. The plots were 154 air dried for two months to reduce the sediment water content. The three plots were further subdivided into 9 sub-plots of 20 m<sup>2</sup> each, resulting in 3 untreated plots (NT), 3 155 HA-treated plots and 3 TBS-treated plots. Six of these plots were further sown with 156 157 grasses (Deschampsia cespitosa and Festuca rubra) while the 3 remaining plots remained unplanted. This study reported the work performed on these unsown plots. 158 159 From mid-spring 2003, spontaneous vegetation started to develop on these plots. 160 Vegetation management (i.e. removal of selected species, harvest, fertilizer addition, and irrigation) was not carried out from this time to March 2007. 161

162 2.2. Collection of sediment and soil samples

In March 2007, top-sediments (0-20 cm depth) of the three tested plots (NT, HA,
TBS) were sampled. Three surface sediment samples per plot were randomly collected
with a hand auger to determine (pseudo)-total metal(oid) and extractable concentrations,

166 pH<sub>water</sub> and sediment ecotoxicity. Six additional randomly collected samples were taken on each of the plots at a depth of about 20 cm using a hand auger for bacterial analyses 167 168 (3 samples/plot) and a corer of 5 cm diameter for collembolan analyses (3 samples / plot). One composite sample per plot was formed with 5 sub-samples randomly 169 collected to determine the other physico-chemical characteristics (particle size, organic 170 171 carbon, total nitrogen, carbonates and cation exchange capacity). In parallel, an uncontaminated soil (0-20 cm depth), located outside the plots, was collected. This 172 173 control area (CA) was selected because it had similar spontaneous vegetation to the 174 plots. Physicochemical properties were determined on a composite sample made of 5 sub-samples collected below the vegetation of interest. Three additional soil samples 175 176 were collected for collembolan analyses.

# 177

#### 2.3. Sediment and soil characteristics

All samples were air dried until constant weight and sieved through 2 mm mesh
size. All analyses on composites have been performed at the Laboratoire Départemental
d'Analyses et de Recherche (LDAR), Laon, France using standard methods (particle
size (NF X 31 – 107, 2003), organic carbon (NF ISO 14 235, oxidation method, 1998),
total N (NF ISO 11261, colorimetric method, 1995), carbonates (NF ISO 10693, 2004),
cation exchange capacity (CEC, NF X31-130, 1999).

A sample of 0.5 g of air-dried and sieved sediment and soil was weighed into 70

mL teflon microwave tubes .Eight mL of *aqua regia* containing 2 mL 67% concentrated

186  $HNO_3$  and 6 mL 36% concentrated HCl (NF EN 13657, 2003) was added. Samples

187 were heated in a microwave digester (MARS Xpress, CEM Corporation<sup>®</sup>, Matthews,

NC) to 180 °C for 20 min, with a 30 minute ramp time. After filtration through a 0.45

189  $\mu$ m Whatman filter, the pseudo-total metal(oid) concentrations (As, Cd, Pb, Zn, Cu)

190	were determined using an Inductively Coupled Plasma Atomic Emission Spectrometer
191	ICP-AES (Jobin Yvon <sup>®</sup> , Longjumeau, France). To assess the analytical quality, a
192	standard reference sediment material SRM 2704 (Buffalo River Sediment, Standard
193	Reference Material) was subjected to the same protocol. Recoveries were: 100 % for
194	Cd, 101% for Zn, 104% for Pb, 90% for Cu and 94% for As.
195	Soil pH was measured in 1:5 sediment/soil:water suspension using a glass
196	electrode pH meter (NF ISO 10390, 1994).
197	Table 1 lists physico-chemical properties of contaminated sediments (NT, HA,
198	TBS) and the uncontaminated soil (CA).

- 199 *2.4. Extractable trace element analysis and sediment ecotoxicity*
- Extractable sediment metal(oid) concentrations of treated and untreated plots were determined after extraction with 0.01 M Ca(NO<sub>3</sub>)<sub>2</sub>. Prior to analysis, 20 g of airdried and sieved sediment were shaken for 48 h with 40 mL of 0.01 M Ca(NO<sub>3</sub>)<sub>2</sub> solution. Extracts were filtered though a 0.45  $\mu$ m cellulose membrane and metal concentrations were measured using ICP-AES.

Ecotoxicity was assessed by two standardized bioassays: the bacterial Microtox® 205 206 assay and the algal test, using Vibrio fischeri and Pseudokirchneriella subcapitata as test organisms, respectively. Those two assays were previously shown to be highly 207 208 sensitive to leachates obtained from dredged materials (Piou et al., 2009). Prior to the 209 bioassays, soil leachates were prepared according to the French standard AFNOR X31-210 (1991) and diluted (10 to 90%) with demineralised water. Ecotoxicity assays were 210 run according to AFNOR T90-320 (1991) and ISO 8692 (1989) standards for 211 Microtox<sup>®</sup> and algal bioassays, respectively. The half maximum effective concentration 212

213	(EC <sub>50</sub> , Microtox <sup>®</sup> bioassay) and the 50% inhibitory concentration (IC <sub>50</sub> , algal bioassay)
214	were derived from dilution-effect curves. When the toxicity of leachates was not
215	sufficient to reach 50% of effect, EC50% or $IC_{50}$ values were indeterminable and the %
216	of effect observed at the 90% dilution was then reported.
217	2.5. Plant survey, biodiversity indices and plant analysis
218	2.5.1. Plant survey and similarity index
219	Vascular plants were surveyed in May and June 2007 in all plots using
220	Lambinon et al. (2004) as a standard reference book for plant taxonomy. The total
221	vegetation cover and the cover rate of mosses, herbaceous plants, shrubs and trees were
222	assessed on each plot. Individuals per species and per plot were counted and numbers
223	were related to plot area. Species richness corresponded to the number of species per
224	plot.

A similarity index (Sørensen, 1948) was calculated to compare the composition of vegetation between (i) experimental plots (influence of treatments) and (ii) experimental plots and surrounding biotopes (identification of putative biotopes that may contribute to plot colonization).

229

2.5.2. Biodiversity indices

230 The diversity of vascular plants was assessed using the Shannon-Weaver index
231 H' (Shannon and Weaver, 1949), calculated as

232  $H' = -\sum_{i=1}^{S} pi * \log_2 pi$ 

233	where $pi$ is the species proportion in a community composed of $s$ species. The
234	homogeneity of species distribution was assessed through the regularity index J of
235	Piélou (1966).

236 2.5.3. Plant analyses

*Calamagrostis epigejos*, a grass, and *Urtica dioica*, a forb, were chosen for
phytotoxicity and metal content analyses as both herb species were present in all plots
and in the control area in sufficient abundance to sample 5 individuals per species of
approximately similar size and sufficient biomass to perform analyses. Plant sampling
occurred in June 2007.

242

#### 2.5.3.1 Chlorophyll fluorescence

The fluorescence of chlorophyll a was measured on dark-adapted leaves of *C*. *epigejos* and *U. dioica* submitted to a saturating light pulse using a portable chlorophyll
fluorimeter (Handy-PEA, Hansatech Instruments<sup>®</sup>, Norfolk, UK). Basal fluorescence
(F0) and maximum fluorescence (Fm) values were used to derive Fv/Fm ratio
(maximum photochemical efficiency of PSII), with the variable fluorescence Fv=FmF0.

249

## 2.5.3.2 Antioxidant enzymes

Leaves from both plant species were collected from CA, NT, HA and TBS plots and immediately transferred to polypropylene tubes then frozen in liquid nitrogen. At the laboratory, leaf samples were crushed manually in a 125 mM phosphate buffer at pH 7.8 by using a porcelain mortar placed on ice. After centrifugation (15 000 g for 10 min at 4° C), the supernatant was used to determine antioxidant enzyme activities.

Superoxide dismutase (SOD), catalase (CAT), ascorbate peroxidase (APX), guaiacol peroxidase (GPX), and glutathione reductase (GRD) activities were measured according to methods described by Dazy et al. (2008). The protein concentration was determined in plant extracts (Bradford, 1976); enzyme activity levels were expressed in  $\mu$ moles/min/mg protein excepted for SOD for which one unit of activity corresponded to the amount of enzyme causing 50% inhibition of absorbance readings by comparison to control tubes lacking enzymes.

#### 262 2.5.3.3 Elemental concentrations in plant shoots

263 Plant shoots were carefully washed (3 times) with distilled water and oven-dried 264 at 40°C until constant weight. Weighed aliquots (0.5 g DW) were wet digested in 10 mL 265 HNO<sub>3</sub> (65%) using a microwave digester (Mars Xpress) according to the following 266 program: 3 min to reach 100 °C, 2 min to reach 140 °C, 2 min to reach 160 °C, 2 min to 267 reach 180 °C and 20 min at 180 °C. Solutions were filtered to <0.45  $\mu$ m (Whatman filter) and concentrations of Cd, Zn, Pb, Cu, As, Ca, Fe, Mg, Mn, P, Na, K and Al were 268 analyzed using ICP- AES (VARIAN 720 AES-ICP, Ulys<sup>®</sup>, France). Blanks and 269 certified reference material (trace elements in hay, IAEA V10, International Atomic 270 271 Energy Agency, Vienna, Austria) were included for quality control. Recoveries were: 272 100% for Ca, 91% for Cu, 79% for Fe, 113% for Mg, 91% for As, 89% for Mn and Pb, 105% for K, 89% for Na, 118% for P, 87% for Zn, 93% for Cd and 67% for Al. 273 274 275 2.6 *Collembola identification and gut content observation* 

After sampling at a depth of 20 cm, considering that the whole collembolan
community was sampled on a surface of ~20 cm<sup>2</sup>, sediments (NT, HA, TBS) and soil

278 (CA) samples were embedded in polythene bags then transported to the laboratory. The 279 extraction of arthropods started on the same day using the dry funnel (Berlese) method 280 (Edwards and Fletcher, 1971) and was completed within 7-10 days. The daytime maximum temperature averaged 25-30 °C. Animals were collected and preserved in 281 282 ethyl alcohol until sorting and identification. Sorting was done in alcohol under a 283 dissecting microscope at x 20 magnification. All springtails were mounted in chloral-284 lactophenol then observed in a light microscope under phase contrast at x 400 285 magnification. Identification was done at the species level using keys by Bretfeld (1999), Fjellberg (1998), Gisin (1960), Hopkin (2000), Jordana et al. (1997), Potapov 286 (2001) and Zimdars and Dunger (1994), Gut contents were observed by transparency 287 288 and classified in 8 categories using morphological features observable in phase contrast light microscopy at x 400 magnification, and were counted according to the method 289 290 devised by (Gillet and Ponge, 2003, 2005): algae, animal remains, bacteria, melanised 291 and hyaline hyphae, fungal spores, holorganic humus, hemorganic humus, to which empty guts were added. The examination of gut contents may elucidate the composition 292 293 of horizons in which the animals were living (Ponge, 2000). We sought to determine 294 when toxicity forced the organisms to move to horizons in which they do not normally live or delete some food resources from their current diet (Gillet and Ponge, 2003). 295

296

2.7 Bacterial analyses

Sediment samples (NT, HA and TBS) were collected in polythene bags then
transported to the laboratory. After homogenization, samples were maintained at 4°C
before performing bacterial analyses. The enumeration of total sulphur-oxidising
bacteria (SOB) was carried out on Nunc<sup>®</sup> Delta (Nalge Nunc International<sup>®</sup>, Roskilde,
Denmark) microplates of 96 wells (8 lines x 12 columns), on which each well contained

302	250 $\mu$ L of specific medium. Nutrient Broth <sup>®</sup> (Difco <sup>®</sup> , Detroit, USA) medium (diluted to
303	1/10), added to fungicidal cyclohexamide solution (0.2 g L <sup>-1</sup> ), was used for total
304	bacterial count. The medium used to enumerate neutrophilic SOB contained 5 g
305	Na <sub>2</sub> SO <sub>3</sub> .5H <sub>2</sub> O, 2 g K <sub>2</sub> HPO <sub>4</sub> , 0.1 g (NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub> , 0.1 g MgSO <sub>4</sub> .7H <sub>2</sub> O, 0.1 g CaCl <sub>2</sub> , 0.002 g
306	FeCl <sub>3</sub> .6H <sub>2</sub> O, 0.005 g MnSO <sub>4</sub> .4H <sub>2</sub> O and 3 ml bromocresol purple (c = 0.02 g $L^{-1}$ ). The
307	pH of the medium was initially adjusted to pH 6.5 with NaOH. For acidophilic SOB,
308	the medium, which contained 1.5 g KH <sub>2</sub> PO <sub>4</sub> , 0.1 g CaCl <sub>2</sub> , 0.1 g MgCl <sub>2</sub> .H <sub>2</sub> O, and 0.1 g
309	NH <sub>4</sub> Cl, was combined with 3 ml of bromocresol green ( $c = 0.02 \text{ g L}^{-1}$ ). The pH was
310	initially adjusted to 4.5 with HCl. A suspension containing 10 g of dry soil and 50 ml of
311	Ringer solution was mixed in a Waring blender for 1.5 min at high speed. After
312	decantation, the solution was successively diluted with Ringer solution to $10^{-1} - 10^{-8}$
313	(Ramsay, 1984). Each well was inoculated with 25 $\mu$ l of solution. The wells of the eight
314	lines corresponded to the eight tested dilutions. Each sample was inoculated in the wells
315	of three columns and a column was not inoculated to control the sterility of the medium.
316	As each microplate had twelve columns, only one microplate was used for the three
317	samples of a given plot. The inoculated plates were incubated at 30°C for 15 days and
318	then the coloured products were measured. Bacteria were counted using the Most
319	Probable Numbers (MPN) method (De Man, 1977). Sulphate-reducing bacteria (SRB)
320	were counted in tubes containing a solid medium specific to SRB (American Petroleum
321	Institute, Washington, DC) (Lors et al., 2004).
222	

2.8 Statistical analyses

A one-way ANOVA was used to detect treatment effects on metal

324 concentrations, Fv/Fm ratio, antioxidant enzyme activities, species distribution,

biodiversity indices, distribution of gut contents and bacterial numbers. When ANOVA

326 was significant, mean values were compared with post-hoc multiple-comparison procedures. When the sample weight was too low and variance homogeneity was not 327 328 fulfilled, a non-parametric statistical test followed by a multiple-comparison procedure was performed. Correlation analysis, using Pearson product-moment correlation 329 coefficient, was performed between metal concentrations and antioxidant enzymes 330 activities in leaves. Statistica v7 (StatSoft Inc.) was used for bacterial calculations 331 whereas XLSTAT<sup>®</sup> (Addinsoft<sup>®</sup>, Paris, France) was used for springtail calculations. All 332 333 other statistics were made with PASW statistics 18.

334 A semi-quantitative method was specially devised to make an overall assessment of beneficial or adverse effects of soil amendment (HA and TBS) addition on dredged 335 336 sediments. For this purpose, the treatments (NT, HA, TBS) were ranked 1 to 3, a value 337 of 1 being attributed to the treatment that maximized variables describing optimal features of plant, faunal and microbial communities (i.e. vegetation succession state, 338 species richness, species diversity, equitability, abundance) and plant Fv/Fm ratio, or 339 minimized those describing metal mobility, ecotoxicity, plant metal concentration and 340 341 antioxidant enzyme activity,. Some variables cannot be taken into account in these 342 calculations, either because they may respond to other factors than soil toxicity (acid vs 343 neutral sulphur-oxidizing bacteria, collembolan gut contents, vegetation composition, pH), or because they do not differ significantly among treatments (plant Fv/Fm ratio for 344 345 C. epigejos). Ranks were then averaged among the different variables which had been kept for the calculations. Overall differences between the three treatments NT, HA and 346 347 TBS were then tested by a sign-test, using ranks as unit values.

348 **3 Results** 

# 3.1. Metal(oid) mobility and sediment ecotoxicity

350	Extractable Pb and As concentrations in all plots were below quantification
351	limits (0.07 and 0.1 mg kg <sup>-1</sup> DW for Pb and As, respectively). Cadmium and Zn showed
352	a similar pattern with higher 0.01 M $Ca(NO_3)_2$ -extractable concentrations in the NT plot
353	than in the TBS-treated plot, while intermediate values were found in the HA-treated
354	plot (Table 2). Expressed as percentages of the total metal concentrations, extractable
355	Cd and Zn represented 0.14 and 0.07%, respectively, in the TBS-treated plot, whereas
356	they represented 0.37 and 0.76% in the NT plot and 0.31 and 0.61% in the HA-treated
357	plot. For Cu, no significant differences occurred across the plots.
358	Sediment leachates were moderately toxic to the bacteria V. fischeri since no
359	$EC_{50}$ could be determined. Considering the % of effect measured at the 90% dilution,
360	the ecotoxicity of sediment leachates could be ranked in the following order $NT > HA >$
361	TBS (Table 2). Sediment leachates were more toxic to Algae, especially those prepared
362	from NT and HA (IC $_{50}$ values being 12.8 and 19.2 %, respectively). TBS sediment
363	leachate had only a slight inhibitory effect on Algae.
364	
365	3.2. Plant survey, plot similarity, source of colonization and element
366	concentrations
367	The composition of the plant community was influenced by the treatments of the
368	plots as shown by plant survey and the number of individuals per species and per
369	surface (Table 3). The TBS plot was mainly colonized by herbaceous plants while HA
370	and NT plots were dominated by shrubs. The maximum height of shrubs was higher in
371	HA plot than in NT plot. The TBS plot had the lowest plant species richness (n=14).

Based on Shannon-Weaver diversity and Piélou equitability indices, the HA plot had the
highest plant diversity with a more homogeneous species distribution while TBS and
NT plots showed a lower diversity and at least one species that dominated the overall
community (Table 3).

In all plots, the dominant family was Poaceae with 5 species for HA, 4 species 376 377 for TBS and 3 species for NT. Among Poaceae, only C. epigejos was present on all 378 plots. Calamagrostis epigejos was more present on the NT plot (46.9%), than on TBS 379 (30%) and HA plots (19.9%). On all plots, other plant species were abundant: Carex 380 hirta (11.5%) and Glechoma hederacea (11.7%) on the NT plot, Betula pendula 381 (12.7%) and G. hederacea (22.8%) on the HA plot, and Phalaris arundinacea (32.1%), 382 Galium aparine (16.4%) and Epilobium angustifolium (11.7%) on the TBS plot. The 383 second most abundant family was Asteraceae with three species. At least two Asteraceae were present on all plots. Perennial species dominated the three group 384 385 communities. Plant species were native to the Nord-Pas-de-Calais region with no clear specificity linked to sediment traits except for some plants showing hygrophilic 386 characteristics (e.g. Eupatorium cannabinum, C. hirta, Carex cuprina, P. arundinacea, 387 388 Salix alba).

The Sørensen index, comparing vegetation composition between plots and between plots and surrounding biotopes, indicated that 85% of plant species were common between NT and HA plots whereas the TBS plot showed 60% and 57% species similarity with HA and NT plots, respectively (data not shown). The surrounding biotopes consisted of a slag heap, a ruderal zone, a canal bank, and an agricultural land. A similarity index indicated that the plant colonizers of the NT and HA plots mainly originated from the slag heap, with 45 and 41% of species similarity.

396	This result agreed with the direction of the south-easterly prevailing wind and the seed
397	shadow colonization hypothesis (Clark et al., 2005), as the slag heap was situated south-
398	east from the plots. The TBS plot showed less similarity with the slag heap (33%).
399	The concentrations of the chemical elements were measured in C. epigejos and
400	U. dioica as both species were sampled in all plots (Table 4). Their shoot Pb and As
401	concentrations were below quantification limits (1 and 0.5 mg kg <sup>-1</sup> for Pb and As,
402	respectively). Shoot Cd and Zn concentrations for both species and Cu concentration for
403	U. dioica were lower in CA than in the three treatments. The shoot Cd concentration
404	was the lowest in C. epigejos grown on TBS whereas for U. dioica it did not differ
405	across treatments. Shoot Zn concentration was the lowest in C. epigejos grown on TBS.
406	The highest shoot P and K concentrations were measured in U. dioica grown on TBS.
407	No other significant effect of TBS treatment on metal concentration was observed. The
408	HA treatment did not decrease shoot Cd and Zn concentrations of C. epigejos and U.
409	dioica compared to NT but increased shoot Cu in C. epigejos. Shoot Mg, Na, and Al
410	concentrations in C. epigejos and shoot Fe concentration in U. dioica were higher in the
411	HA plot than in the NP plot. The HA treatment did not influence other shoot element
412	concentrations.

414

# 3.3. Antioxidant enzyme activities and chlorophyll fluorescence of leaves

Unstressed plants typically exhibit Fv/Fm ratios of 0.8-0.83 (Laposi et al., 2009),
which was the case for *C. epigejos* on all plots (Fig. 1). For all antioxidant enzymes,
lowest activities were determined in CA plants. In untreated sediment (NT), *C. epigejos*enzymes were significantly increased 1.2 to 3 fold depending on the enzyme considered.

419 For plants collected on HA plots, the activity of SOD, CAT, and GRD were similar to 420 CA values, except APX which remained higher. For GPX, the HA treatment had no 421 significant effect. This enzyme was less influenced than the others by the composition 422 of the NT sediment since its activity only increased by a factor of 1.2 compared to plants harvested on CA plots. Plant enzymes responded differently to the TBS 423 treatment. For SOD, CAT and APX, activities levels were not influenced since their 424 values did not differ significantly from corresponding values in NT plants. GPX and 425 426 GRD were both affected by TBS treatment, but in opposite ways, since GPX increased 427 by comparison to NT values while GRD decreased.

Urtica dioica collected from CA and HA plots, and to a lesser extend NT plots, 428 had all Fv/Fm ratios around 0.8 (Fig. 2). For TBS-plants, the Fv/Fm ratios were 429 430 significantly lower. As noted for C. epigejos, for all antioxidant enzymes, the lowest activities were found for CA plants. For the untreated sediment (NT), U. dioica 431 432 enzymes significantly increased by a factor 3 to 5. The HA and TBS treatments had opposite effects on antioxidant enzymes. All tested enzyme activities in U. dioica leaves 433 434 decreased for HA compared to NT and tended to recover CA values. However, 435 differences between HA and CA plants remained significant for SOD, APX and GPX activities. In contrast, TBS plants still exhibited high enzyme activities, similar to NT 436 values (e.g. APX and CAT) or even higher (e.g. SOD, GPX, and GRD). 437 3.4. Correlations between shoot element concentrations and antioxidant 438 439 enzyme activities

A correlation analysis was performed, between shoot element concentrations in *C. epigejos* and *U. dioica* and their antioxidant enzyme activities in leaves, to reveal
significant linear relationships (Table 5). All enzyme activities negatively correlated

with shoot Cd concentration in *C. epigejos*. Manganese concentrations were positively
correlated with 4 out of the 5 enzyme activities whereas Cu, Zn, Mg, and Na
concentrations were correlated with GRD. For *U. dioica*, no relationship was found
between shoot Cd concentration and any of the enzyme activities, but all enzyme
activities were positively correlated with shoot Cu, Zn, P, and K concentrations.

448

# 3.5. Collembolan diversity and gut contents

A total of fourteen species were found in the four areas (Table 6). Neither total abundance nor biodiversity indices (species richness, Shannon index, equitability) showed significant differences between treatments. However, the TBS plot tended to harbour more specimens and species than NT and HA plots and CA. In the NT plot, there was no decrease in any of the four biodiversity indices when compared to CA, with a mean abundance of 20 specimens per core sample, shared between four species. At the species level, *Friesea truncata* was only present in the TBS samples.

The only significant effect on the gut contents of Collembola (Fig. 3) was an increase in the percent of specimens with hyaline fungal hyphae, which increased in all cores sampled in the HA plot, reaching an average of 16% compared to less than 3% for the other treatments. Hemorganic humus (amorphous organo-mineral matter) was dominant everywhere, indicating that the sediments (treated or not) were consumed by springtails, in particular by the dominant species *Mesaphorura florae*.

462

## *3.6. Total and specific bacterial numbers*

463 The total bacterial population in all plots varied from  $10^7$  to  $10^8$  bacteria g<sup>-1</sup> dry 464 sediment, and was similar to that usually found in unpolluted topsoils (Robert, 1996;

465	Taylor et al., 2002) (Table 7). However, in the TBS plot, the total bacterial population
466	(in $10^8$ bacteria.g <sup>-1</sup> dry soil) was significantly higher (1.7) than in the NT plot (0.21).
467	In all plots, neutrophilic sulphur-oxidising bacteria (NSOB) were the most
468	abundant group, with about $10^5$ bacteria g <sup>-1</sup> dry sediment, whereas acidophilic sulphur-
469	oxidising bacteria (ASOB) were the least abundant group with about $10^3$ - $10^4$ bacteria g
470	<sup>1</sup> dry sediment. The amendments did not modify the size of these bacterial populations.
471	The sulphate-reducing bacterial microflora (SRB) was about $10^{1}$ - $10^{2}$ bacteria g <sup>-1</sup>
472	dry sediment and was of the same order of magnitude in all plots.

## 474 *3.7. Integration of data from the ecotoxicity battery*

Averaging ranks of all variables selected for the synthetic assessment of treatment effect (see Material and methods) indicated that TBS and HA exhibited similar favourable effects on the ecosystem when compared to the untreated sediment (NT) (Table 8). HA and TBS treatments ranked significantly above NT (one-sided ttests, P = 0.001 and 0.011, respectively) while they did not differ from each other (twosided t-test, P = 0.797).

## 481 **4. Discussion**

Although the TBS application rate and the soil type used in this study differed from those investigated in other works, our results are broadly similar for Zn and Cd. The incorporation of TBS (5% DW) into the metal contaminated sediment decreased the 0.01 M Ca(NO<sub>3</sub>)<sub>2</sub>-extractable Cd and Zn fractions and shoot Cd and Zn concentrations of *C. Epigejos* (Tables 2 and 4), confirming previous findings (Mench et al., 1994a, 1994b, 2000). The TBS incorporation into a Cd-Zn-contaminated soil, reduced both the

488	$Ca(NO_3)_2$ extractability of Cd and Zn and their contents in tobacco shoots (Mench et al.,
489	1994a, 1994b). In addition, TBS added into a Zn-contaminated soil increased soil pH,
490	decreased extractable Zn and shoot Zn concentration in ryegrass, 3 and 5 months after
491	soil treatment (Mench et al., 2000). Contrary to TBS, HA incorporation (3% DW) into
492	the sediment did not decrease significantly Cd and Zn extractability nor Cd and Zn
493	concentrations in plants (Tables 2 and 4). These results were not consistent with those
494	of Boisson et al. (1999) who found a decrease of extractable Cd and Zn and metal
495	concentrations in maize when compared to the untreated soil and with increasing HA
496	application rate (0.5, 1 and 5% DW).

In contrast to other works (Boisson et al., 1999; Friesl et al., 2006), Asmobilization was not observed either in the presence of HA or TBS (Table 2). No
amendment effect was recorded for Cu (Tables 2 and 4) whereas TBS incorporation
(0.25 and 3.9% DW) into a Cu contaminated soil decreased Cu concentrations in the
primary leaves of bean (Bes and Mench, 2008).

502 In our field study, TBS was the most efficient amendment to decrease Zn and Cd mobility and transfer to the plants. We hypothesise that Zn- and Cd-phosphates were 503 504 precipitated, thereby immobilizing these elements in a non-available form as suggested 505 by Panfili et al. (2005) who studied Zn speciation in the same metal-contaminated 506 sediment treated with TBS. In our field experiment, the low efficiency of HA compared 507 to TBS may be explained by the low solubility of HA at neutral pH (Table 1). As 508 suggested by Miretzky and Fernandez-Cirelli (2007), the solubility of the phosphate phase is necessary for successful in situ treatment, metal immobilization by phosphorus 509 510 being limited or inefficient when the matrix is neutral or alkaline.

511 As the essential nutrient concentrations did not decrease in plant shoots (Table 512 4), our results suggested that neither TBS nor HA led to deficiency problems in the 513 tested plant species. In the presence of HA (5% DW), Boisson et al. (1999) reported a 514 Mn-deficiency in maize. Neither treatment promoted the uptake of essential nutrients such as Ca except in the case of U. dioica where shoot P and K concentrations were 515 516 increased by TBS addition. In primary leaves of bean, Ca concentration was enhanced in the presence of TBS whereas P concentration was not promoted (Bes and Mench, 517 518 2008; Negim et al., 2010).

519 In our study, sediment ecotoxicity was reduced by the incorporation of TBS and520 HA, TBS being the most efficient amendment.

521 At excessive concentrations in the soil solution, Cd, Cu, Pb and Zn exert toxic effects on plants, including alterations in photosynthetic and respiration processes or 522 523 inhibition of plant growth. They may also stimulate the formation of reactive oxygen 524 species (Dazy et al., 2009). To prevent oxidative damage, plant cells usually use enzymatic protection mechanisms, such as superoxide dismutase, catalase, and 525 526 peroxidases, and metabolites such as ascorbate and glutathione (Dazy et al., 2008). 527 Following amendment of the soil, a decrease in phytotoxicity and a recovery in plant 528 antioxidant defence to control values are expected as a result of metal-immobilization 529 processes (Ruttens et al., 2006; Vangronsveld et al., 1995, 1996). In the NT plot, all antioxidant enzyme activities of both plant species were increased compared to the CA 530 531 plot, indicating that the plants suffered from oxidative stress. The increase in shoot Zn and Cu concentrations were consistent with enzymatic activity increases, suggesting 532 533 that these metals accounted for the stress response recorded in plants growing on 534 untreated dredged sediment. This was not the case for Cd, as no correlation for U.

535 dioica and no positive correlation for C. epigejos were found. The addition of HA permitted an almost total recovery of antioxidant enzyme activities to CA values, 536 537 whereas the addition of TBS did not. Thus, HA was a better amendment for keeping the stress responses of plants at a low level. Levels of chlorophyll fluorescence (Fig. 1 and 538 Fig. 2) indicated that the plants were not stressed on NT, HA and TBS plots. There are 539 few studies that report the effect of TBS on phytotoxicity and an absence of studies on 540 the effect of HA on phytotoxicity. After the incorporation of TBS (0.25% and 3.9%) 541 542 into a Cu-contaminated soil, the chlorophyll density of primary leaves of bean 543 decreased and guaiacol-peroxidase activity in roots fell to control level (Bes and Mench, 2008). In another experiment, TBS added to a Zn-contaminated soil restored guaiacol-544 545 peroxidase activity of the primary leaves in beans to control values (Mench et al., 2000).

546 Most of the plant species present on the NT plot were also observed on the surrounding uncontaminated biotopes (slag heap and ruderal zone), which were 547 548 similarly firstly devoid of vegetation. Thus, colonization of the NT plot was not limited by the total metal concentration in the sediment, which is consistent with previous 549 findings (Escarré et al., 2011). Madejon et al. (2006) showed that the application of 550 551 amendments to a metal contaminated soil improved spontaneous colonization and establishment of early-successional plants. In our study, all plots were covered by dense 552 vegetation (close to 100%) but differed in plant community composition and 553 biodiversity descriptors. This may indicate that the amendment application was not a 554 key factor for colonization and establishment of spontaneous vegetation but an 555 556 important factor that could modify vegetation composition and dynamics. The addition of TBS or HA to the sediment influenced the dynamic process of plant colonization, the 557 three plots showing differences in composition of the plant community. The large 558

559 increase in pH following addition of TBS to the sediment (7.4 to 10.2 in May 2002), 560 compared to the modest increase (7.4 to 7.7 in May 2002) in the presence of HA, may 561 be responsible for the main changes in plant community in the TBS plot compared to 562 other plots. Only spontaneous vegetation that is able to tolerate alkaline pHs could colonize this plot. Elevated pH favours the establishment of herbaceous rather than 563 564 woody species (Skousen et al., 1994). Five years after the single application, the pH in the TBS-treated sediment decreased to 7.7 while pHs of NT and HA plots were 7.0 and 565 566 7.3, respectively (Table 1). This decrease may be sufficient to favour the establishment 567 of *B. pendula* in the next few years, as two seedlings were noticed in the TBS plot. *Calamagrostis epigejos* is known as a competitive social grass able to arrest plant 568 569 succession by forming extended dense areas (Prach, 2003). At maturity, it can reach 1.5 570 m in height thereby shading smaller or slowly-growing plant species (Rebele and 571 Lehmann, 2001). As the higher rate of C. epigejos was recorded on the TBS plot, this plant would contribute to the maintenance of early stage succession vegetation on this 572 573 plot.

The highest level of plant species richness was found in the presence of HA (26 species out of 16 families) compared to the NT plot (21 species out of 14 families). In contrast, TBS did not favour species richness as it accounted for the least value (14 species out of 9 families). This is paradoxical since the TBS plot had the lowest concentrations of extractable Zn and Cd and the lowest sediment ecotoxicity, but the competitive exclusion of plants by *C. epigejos* possibly outweighed effects of decreased toxicity.

581 Concerning diversity indices, our H' values were much higher than those given
582 by Conesa et al. (2007) and Dazy et al. (2008) who studied plant communities growing

respectively on Zn-Pb-contaminated tailings and experimental plots filled with a
multicontaminated soil. However, such comparisons of H' values may be confounded
by different soil and pollution parameters as well as the time elapsed after colonization..
For Dazy et al. (2008), the survey was performed after 14 months of colonization
whereas in this study the survey was done 5 years after colonization.

588 The influence of metal contamination on natural springtail communities has 589 been documented (Bengtsson and Rundgren, 1988; Chauvat and Ponge, 2002; Gillet 590 and Ponge, 2002, 2003; Lock et al. 2003; Strojan, 1978). In these works, no major 591 adverse effects of metal contamination were recorded. Springtails were abundant even 592 in highly contaminated sites, which indicates the resistance of this group to toxic effect of metals and other environmental stresses (Hopkin, 1997), exemplified by high  $EC_{50}$ 593 values in *Folsomia candida* reproduction test at 20 °C for Cd (590 µg.g<sup>-1</sup>), Cu (700 594  $\mu g.g^{-1}$ ), Zn (900  $\mu g.g^{-1}$ ) and Pb (2790  $\mu g.g^{-1}$ ) (Sandifer and Hopkin, 1997). In our study, 595 neither total abundance nor biodiversity indices showed significant differences between 596 597 NT plot and CA, showing the normal colonization by springtails of the NT plot. The addition of TBS and HA did not change the abundance or biodiversity indices, showing 598 599 no effect of these amendments on the springtail community. Some effect may have been 600 expected on the TBS plot as Lock et al. (2003) reported a significant negative relationship between the metal extractable fraction (0.01 M CaCl<sub>2</sub> solution) and the 601 602 number of springtail species. In our study, the gut content analyses showed that untreated and amended dredged sediments were consumed by springtails (Fig. 3). 603 In soils that are highly contaminated with metal(loid)s, a lower number of 604

bacteria than in uncontaminated soil could be expected, as a result of the harmful effects
 of the contaminants on soil bacterial activity, especially bacterial respiration and soil

bacterial biomass (Renella et al., 2005; Sobolev and Begonia, 2008). In our study, the
total bacterial population in the NT plot was similar to that usually found in unpolluted
topsoils (Taylor et al., 2002).

610 The addition of TBS to the soil improved the total bacterial population, probably611 as a consequence of the low Cd-Zn-extractable fraction on this plot.

612 Bacteria play an important role in increasing or decreasing metal availability, notably in sediments (Bert et al., 2009). Bioleaching of metals is caused by sulphur-613 614 oxidising bacteria (SOB) whereas anaerobic sulphate-reducing bacteria (SRB) can precipitate metals. Neutrophilic and acidophilic SOB were found in similar numbers in 615 all plots, showing no amendment effect. In all sediment plots, neutrophilic SOB were 616 617 more numerous than acidophilic SOB, due to soil pH close to neutrality (Table 1). The 618 moderate development of acidophilic SOB was consistent with the results of Lors et al. (2004) who reported an acidophilic sulphur-oxidising population of about  $10^4$  bacteria 619 620  $g^{-1}$  dry soil in a similar sediment. These results indicated that metals might be released in the case of metal sulphide availability. 621

Sulphate-reducing bacteria were present in the same low range in the three
sediment plots, showing no amendment effect (Table 7). The low amount of SRB may
be explained by the sampling performed on the first 20 cm depth layer corresponding to
aerobic conditions, not favourable to the growth of SRB that are strictly anaerobic.
However, during wet periods, the saturation of the soil pore system might induce anoxic
conditions that would promote the size of the SRB community, leading to increased
metal sulphide content.

629

## 630 **5.** Conclusions

631	In this paper, TBS and HA were investigated in an experimental metal-
632	contaminated sediment landfill site to evaluate their effects on metal extractability,
633	sediment ecotoxicity and on living organisms such as plants, bacteria and springtails.
634	The incorporation of TBS into the metal contaminated sediment decreased the
635	extractable fraction of Cd and Zn. In contrast, HA did not decrease Cd and Zn
636	extractability. TBS at a rate of 5% DW was effective in Cd and Zn immobilization, did
637	not cause any nutrient imbalance or sediment ecotoxicity, and improved the bacterial
638	activity.
639	However, TBS application had either no influence or a negative effect, on

maximum photochemical efficiency of PSII and antioxidant enzymes in plants. The
bioavailability of the compound(s) responsible for those biological responses was not
decresead by TBS treatment. Even if plants successfully colonized the TBS plot, they
remained under stress.

644 Our study revealed (i) the favourable influence of both HA and TBS compared to NT, (ii) the better influence of HA on vegetation development and physiological 645 646 welfare (as measured by PSII measurements and antioxidant enzymes), (iii) the better influence of TBS on sediment ecotoxicity, metal immobilization, microbial population 647 and Cd-Zn accumulation in C. epigejos. The choice of using metal immobilization with 648 TBS rather than HA as a tool in the management of dredged sediment landfill sites will 649 depend on the objectives of such strategy, i.e. to limit metal transfer and sediment 650 ecotoxicity or to favour plant diversity and welfare. Realising both objectives requires a 651 study of a mixture of both soil amendments. 652

653

# 654 Acknowledgements

655	This study was financially supported through the Programme de Recherches
656	Concertées (PRC) Environnement-Ecotoxicité: Meresbio, étude des mécanismes de la
657	restauration de la biodiversité sur un site contaminé par les métaux, by the Contrat de Plan
658	Etat/Région Nord-Pas-de-Calais and the European FEDER (ANR - ex.FRT 2005 - action
659	Régionale – CPER DT n°15006 Décision n°2005 – CPER – Nord - 176). We thank the
660	anonymous reviewers for improving the manuscript and B. Robinson for providing
661	language assistance.
662	
663	References
664	AFNOR T90-320, 1991. Testing water-determination of the inhibition of the light
665	emission of Photobacterium phosphoreum. Association Française de
666	Normalisation, Paris.
667	AFNOR X31-210, 1992. Waste – Leaching of waste. Association Française de
668	Normalisation, Paris.
669	Bengtsson, G., Rundgren, S., 1988. The Gusum case : a brass mill and the distribution
670	of soil Collembola. Canadian Journal of Zoology 66, 1518-1526.
671	Bert, V., Seuntjens, P., Dejonghe, W., Lacherez, S., Thi Thanh Thuy, H.,
672	Vandecasteele, B., 2009. Phytoremediation as a management option for
673	contaminated sediments in tidal marshes, flood control areas and dredged sediment
674	landfill sites. Environmental Science and Pollution Research 16, 745–764.
675	Bes, C., Mench, M., 2008. Remediation of copper-contaminated topsoils from a wood
676	treatment facility using in situ stabilization. Environmental Pollution 156, 1128-
677	1138.
	20

678	Bes, M.C., Mench, M., Aulen, M., Gaste, H., Taberly, J., 2010. Spatial variation of
679	plant communities and shoot Cu concentrations of plant species at a timber
680	treatment site. Plant and Soil 330, 267-280.
681	Boisson, J., Ruttens, A., Mench, M., Vangronsveld, J., 1999. Evaluation of
682	hydroxyapatite as a metal immobilizing soil additive for the remediation of
683	polluted soils. I. Influence of hydroxyapatite on metal exchangeability in soil,
684	plant growth and plant metal accumulation. Environmental Pollution 104, 225-
685	233.
686	Bouwman, L.A., Bloem, J., Römkens, P.F.A.M., Boon, G.T., Vangronsveld, J., 2001.
687	Beneficial effects of the growth of metal tolerant grasses on biological and
688	chemical parameters in copper and zinc contaminated sandy soils. Minerva
689	Biotecnologica 13, 19-26.
690	Bouwman, L.A., Vangronsveld, J., 2004. Rehabilitation of the nematode fauna in a
691	phytostabilized heavily zinc-contaminated, sandy soil. Journal of Soils and
692	Sediments 4, 17-23.
693	Bradford, M.M., 1976. A rapid and sensitive method for the quantification of
694	microgram quantities of protein utilizing the principle of protein-dye binding.
695	Analytical Biochemistry 72, 248-254.
696	Bretfeld, G., 1999. Synopses on Palaearctic Collembola. II. Symphypleona.
697	Abhandlungen und Berichte des Naturkundemuseums Görlitz 71, 1-318.
698	Chapin, F.S. III, Walker, L.R., Fastie, C.L., Sharman, L.C., 1994. Mechanisms of
699	primary succession following deglaciation at Glacier Bay, Alaska. Ecological
700	Monographs 64, 149-175.

701	Chauvat, M., Ponge, J.F., 2002. Colonization of heavy metal-polluted soils by
702	collembola : preliminary experiments in compartmented boxes. Applied Soil
703	Ecology 21, 91-106.
704	Clark, C.J., Poulsen, J.R., Bolker, B.M., Connor, E.F., Parker, V.T., 2005. Comparative
705	seed shadows of bird-, monkey-, and wind-dispersed trees. Ecology 86, 2684-
706	2694.
707	Conesa, H., M., Garcia, G., Faz, A., Arnaldos, R., 2007. Dynamics of metal tolerant
708	plant communities' development in mine tailings from the Cartagena-La Union
709	Mining District (SE Spain) and their interest for further revegetation purposes.
710	Chemosphere 68, 1180–1185.
711	Dazy, M., Jung, V., Férard, J.F., Masfaraud, J.F., 2008. Ecological recovery of
712	vegetation at a former coke-factory industrial wasteland: role of plant
713	antioxidant enzymes and possible implications in site restoration. Chemosphere
714	74, 57–63.
715	Dazy, M., Masfaraud, J.F., Férard J.F., 2009. Induction of oxidative stress biomarkers
716	associated with heavy metal stress in Fontinalis antipyretica Hedw.
717	Chemosphere 75, 297–302.
718	De Man, J.C., 1977. MPN tables for more than one test. European Journal of Applied
719	Microbiology 4, 307-316.
720	Edwards, C.A., Fletcher, K.E., 1971. A comparison of extraction methods for terrestrial
721	arthropods. In: Phillipson, J. (Ed.), Methods of Study in Quantitative Soil
722	ecology: Population, Production and Energy Flow. Blackwell, Oxford, pp. 150-
723	185.

724	Escarré, J., Lefèbvre Cl., Raboyau, S., Dossantos, A., Gruber, W., Cleyet Marel, J., Cl.,
725	Frérot, H., Noret, N., Mahieu., S., Collin, Ch., Van Oort F., 2011. Heavy metal
726	concentration survey in soils and plants of the Les Malines mining district
727	(Southern France): implications for soil restoration. Water, Air and Soil
728	Pollution 216, 485-504.
729	Fjellberg, A., 1998. The Collembola of Fennoscandia and Denmark. I. Poduromorpha.
730	Brill, Leiden.
731	Friesl, W., Friedl, J., Platzer, K., Horak, O., Gerzabek, M.H., 2006. Remediation of
732	contaminated agricultural soils near a former Pb/Zn smelter in Austria: Batch,
733	pot and field experiments. Environmental Pollution 144, 40-50.
734	Gillet, S., Ponge, J.F., 2002. Humus forms and metal pollution in soil. European Journal
735	of Soil Science 53, 529-539.
736	Gillet, S., Ponge, J.F., 2003. Changes in species assemblages and diets of Collembola
737	along a gradient of metal pollution. Applied Soil Ecology 22, 127-138.
738	Gillet, S., Ponge, J.F., 2005. Species assemblages and diets of Collembola in the organic
739	matter accumulated over an old tar deposit. European Journal of Soil Biology
740	41, 39-44.
741	Gisin, H., 1960. Collembolenfauna Europas. Muséum d'Histoire Naturelle, Geneva.
742	Hopkin, S.P., 1997. Biology of the Springtails. Oxford University Press, Oxford.
743	Hopkin, S.P., 2000. A key to the Collembola (springtails) of Britain and Ireland. Field
744	Studies Council AIDGAP Guides 111, 1-245.

745	ISO 8692, 1989. Water quality – Freshwater algal growth inhibition test with
746	Scenedesmus subspicatus and Selenastrum capricornutum. International
747	Organization for Standardization, Geneva.
748	Jordana, R., Arbea, J.I., Simón, C., Luciáñez, M.J., 1997. Fauna Iberica. VIII.
749	Collembola Poduromorpha. CSIC, Madrid.
750	Knox, A.S., Seaman, J.C., Mench, M., J., Vangronsveld, J., 2001. Remediation of
751	metal- and radionuclides-contaminated soils by in situ stabilization techniques,
752	in: Iskandar, I.K. (Ed), Environmental Restoration of Metal-contaminated Soils.
753	Lewis Publishers, Boca Raton, pp. 21-60.
754	Kumpiene, J., Lagerkvist, A., Maurice, C., 2008. Stabilization of As, Cr, Cu, Pb and Zn
755	in soil using amendments – a review. Waste Management 28, 215-225.
756	Lambinon, J., Delvosalle, L., Duvignaud, J., 2004. Nouvelle Flore de la Belgique, du
757	Grand-Duché de Luxembourg, du Nord de la France et des Régions Voisines
758	(Ptéridophytes et Spermatophytes), 5th éd. Jardin Botanique de Belgique, Meise.
759	Laposi, R., Veres, S, Lakatos, G., Olah, V., Fieldsend, A., Meszaros, I., 2009.
760	Responses of leaf traits of European beech (Fagus sylvatica L.) saplings to
761	supplemental UV-B radiation and UV-B exclusion. Agricultural and Forest
762	Meteorology 149, 745-755.
763	Lock, K., Janssen, C.R., 2003. Effect of new soil immobilizing agents on metal toxicity
764	to terrestrial invertebrates. Environmental Pollution 121, 123-127.
765	Lock, K., Janssens, F., Janssen, C.R., 2003. Effects of metal contamination on the
766	activity and diversity of springtails in an ancient Pb-Zn mining area at
767	Plombières, Belgium. European Journal of Soil Biology 39, 25-29.

768	Lock, K., Janssen, C.R., 2005. Influence of soil zinc concentration on zinc sensitivity
769	and functional diversity of microbial communities. Environmental Pollution 136,
770	275-281.
771	Lors, C., Tiffreau, C., Laboudigue, A., 2004. Effects of bacterial activities on the release
772	of heavy metals from contaminated dredged sediments. Chemosphere 56, 619-
773	630.
774	Ma, Q.Y., Traina, S.J., Logan, T.J., 1993. In situ lead immobilization by apatite.
775	Environmental Science and Technology 27, 1803-1810.
776	Madejon, E., Perez de Mora, A., Felipe, E., Burgos, P., Cabrera, F., 2006. Soil
777	amendments reduce trace element solubility in a contaminated soil and allow
778	regrowth of natural vegetation. Environmental Pollution 139, 40-52.
779	Mench, M., Vangronsveld, J., Didier, V., Clijsters, H., 1994a. Evaluation of metal
780	mobility, plant availability and immobilization by chemical agents in a limed-
781	silty soil. Environmental Pollution 86, 279-286.
782	Mench, M.J., Didier V.L., Löffler M., Gomez, A., Masson, P., 1994b. A mimicked in-
783	situ remediation study of metal-contaminated soils with emphasis on cadmium
784	and lead. Journal of Environmental Quality 23, 58-63.
785	Mench, M.J., Manceau, A., Vangronsveld, J., Clijsters, H., Mocquot, B., 2000. Capacity
786	of soil amendments in lowering the phytoavailability of sludge-borne zinc.
787	Agronomie 20, 383-397.
788	Miretzky, P., Fernandez-Cirelli, A., 2007. Phosphates for Pb immobilization in soils: a
789	review Environmental and Chemical Letters 6, 121-133.

<ul> <li>mineral composition of beans in a Cu-contaminated soil. Journal of Soil and</li> <li>Sediment Contamination 19, 174-187.</li> <li>NF EN 13657, 2003. Characterization of waste - Digestion for subsequent</li> <li>determination of aqua regia soluble portion of elements. AFNOR, Association</li> <li>française de normalisation, Paris.</li> <li>NF ISO 10693, 1995. Soil quality - Determination of carbonate content - Volumetric</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 11261, 1995. Soil quality - Determination of total nitrogen - Modified Kjeldahl</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> </ul>	790	Misra, V., Chaturvedi, P. K., 2007. Plant uptake/bioavailability of heavy metals from
<ul> <li>Negim, O, Eloifi, B., Mench, M., Bes, C., Gaste, H., Motelica-Heino, M., Le</li> <li>Coustumer, P., 2010. Effect of basic slag addition on soil properties, growth and leaf</li> <li>mineral composition of beans in a Cu-contaminated soil. Journal of Soil and</li> <li>Sediment Contamination 19, 174-187.</li> <li>NF EN 13657, 2003. Characterization of waste - Digestion for subsequent</li> <li>determination of aqua regia soluble portion of elements. AFNOR, Association</li> <li>française de normalisation, Paris.</li> <li>NF ISO 10693, 1995. Soil quality - Determination of carbonate content - Volumetric</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 11261, 1995. Soil quality - Determination of total nitrogen - Modified Kjeldahl</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-107, 2003. Soil Quality - Particle size determination by sedimentation - Pipette</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-130, 1999. Soil quality - Chemical methods - Determination of cathionic</li> <li>exchange capacity (CEC) and extractible cations. AFNOR, Association</li> <li>Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic</li> <li>oxidation. AFNOR, Association Française de Normalisation, Paris.</li> </ul>	791	the contaminated soil after treatment with humus soil and hydroxyapatite
<ul> <li>Coustumer, P., 2010. Effect of basic slag addition on soil properties, growth and leaf</li> <li>mineral composition of beans in a Cu-contaminated soil. Journal of Soil and</li> <li>Sediment Contamination 19, 174-187.</li> <li>NF EN 13657, 2003. Characterization of waste - Digestion for subsequent</li> <li>determination of aqua regia soluble portion of elements. AFNOR, Association</li> <li>française de normalisation, Paris.</li> <li>NF ISO 10693, 1995. Soil quality - Determination of carbonate content - Volumetric</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 11261, 1995. Soil quality - Determination of total nitrogen - Modified Kjeldahl</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-107, 2003. Soil quality - Particle size determination by sedimentation - Pipette</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-130, 1999. Soil quality - Chemical methods - Determination of cathionic</li> <li>exchange capacity (CEC) and extractible cations. AFNOR, Association</li> <li>Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic</li> <li>oxidation. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 10390, 1994. Soil quality - Determination of pH. AFNOR, Association</li> </ul>	792	Environmental and Monitoring Assessment 133, 169–176.
<ul> <li>mineral composition of beans in a Cu-contaminated soil. Journal of Soil and</li> <li>Sediment Contamination 19, 174-187.</li> <li>NF EN 13657, 2003. Characterization of waste - Digestion for subsequent</li> <li>determination of aqua regia soluble portion of elements. AFNOR, Association</li> <li>française de normalisation, Paris.</li> <li>NF ISO 10693, 1995. Soil quality - Determination of carbonate content - Volumetric</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 11261, 1995. Soil quality - Determination of total nitrogen - Modified Kjeldahl</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-107, 2003. Soil quality - Particle size determination by sedimentation - Pipette</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-130, 1999. Soil quality - Chemical methods - Determination of cathonic</li> <li>exchange capacity (CEC) and extractible cations. AFNOR, Association</li> <li>Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic</li> <li>oxidation. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 10390, 1994. Soil quality - Determination of pH. AFNOR, Association</li> </ul>	793	Negim, O, Eloifi, B., Mench, M., Bes, C., Gaste, H., Motelica-Heino, M., Le
<ul> <li>Sediment Contamination 19, 174-187.</li> <li>NF EN 13657, 2003. Characterization of waste - Digestion for subsequent</li> <li>determination of aqua regia soluble portion of elements. AFNOR, Association</li> <li>française de normalisation, Paris.</li> <li>NF ISO 10693, 1995. Soil quality - Determination of carbonate content - Volumetric</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 11261, 1995. Soil quality - Determination of total nitrogen - Modified Kjeldahl</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-107, 2003. Soil quality - Particle size determination by sedimentation - Pipette</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-130, 1999. Soil quality - Chemical methods - Determination of cathionic</li> <li>exchange capacity (CEC) and extractible cations. AFNOR, Association</li> <li>Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic</li> <li>oxidation. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of pH. AFNOR, Association</li> </ul>	794	Coustumer, P., 2010. Effect of basic slag addition on soil properties, growth and leaf
<ul> <li>NF EN 13657, 2003. Characterization of waste - Digestion for subsequent</li> <li>determination of aqua regia soluble portion of elements. AFNOR, Association</li> <li>française de normalisation, Paris.</li> <li>NF ISO 10693, 1995. Soil quality - Determination of carbonate content - Volumetric</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 11261, 1995. Soil quality - Determination of total nitrogen - Modified Kjeldahl</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-107, 2003. Soil Quality - Particle size determination by sedimentation - Pipette</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-130, 1999. Soil quality - Chemical methods - Determination of cathionic</li> <li>exchange capacity (CEC) and extractible cations. AFNOR, Association</li> <li>Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic</li> <li>oxidation. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic</li> <li>oxidation. AFNOR, Association Française de Normalisation, Paris.</li> </ul>	795	mineral composition of beans in a Cu-contaminated soil. Journal of Soil and
<ul> <li>determination of aqua regia soluble portion of elements. AFNOR, Association</li> <li>française de normalisation, Paris.</li> <li>NF ISO 10693, 1995. Soil quality - Determination of carbonate content - Volumetric</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 11261, 1995. Soil quality - Determination of total nitrogen - Modified Kjeldahl</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-107, 2003. Soil Quality - Particle size determination by sedimentation - Pipette</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-107, 2003. Soil quality - Chemical methods - Determination of cathionic</li> <li>exchange capacity (CEC) and extractible cations. AFNOR, Association</li> <li>Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic</li> <li>oxidation. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic</li> <li>oxidation. AFNOR, Association Française de Normalisation, Paris.</li> </ul>	796	Sediment Contamination 19, 174-187.
<ul> <li>française de normalisation, Paris.</li> <li>NF ISO 10693, 1995. Soil quality - Determination of carbonate content - Volumetric</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 11261, 1995. Soil quality - Determination of total nitrogen - Modified Kjeldahl</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-107, 2003. Soil Quality - Particle size determination by sedimentation - Pipette</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-107, 2003. Soil quality - Chemical methods - Determination of cathionic</li> <li>exchange capacity (CEC) and extractible cations. AFNOR, Association</li> <li>Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic</li> <li>oxidation. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 10390, 1994. Soil quality - Determination of pH. AFNOR, Association</li> </ul>	797	NF EN 13657, 2003. Characterization of waste - Digestion for subsequent
<ul> <li>NF ISO 10693, 1995. Soil quality - Determination of carbonate content - Volumetric method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 11261, 1995. Soil quality - Determination of total nitrogen - Modified Kjeldahl method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-107, 2003. Soil Quality - Particle size determination by sedimentation - Pipette method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-107, 2003. Soil Quality - Particle size determination by sedimentation - Pipette method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-130, 1999. Soil quality - Chemical methods - Determination of cathionic exchange capacity (CEC) and extractible cations. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic oxidation. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic not oxidation. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 10390, 1994. Soil quality - Determination of pH. AFNOR, Association</li> </ul>	798	determination of aqua regia soluble portion of elements. AFNOR, Association
<ul> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 11261, 1995. Soil quality - Determination of total nitrogen - Modified Kjeldahl</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-107, 2003. Soil Quality - Particle size determination by sedimentation - Pipette</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-130, 1999. Soil quality - Chemical methods - Determination of cathionic</li> <li>exchange capacity (CEC) and extractible cations. AFNOR, Association</li> <li>Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic</li> <li>oxidation. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 10390, 1994. Soil quality - Determination of pH. AFNOR, Association</li> </ul>	799	française de normalisation, Paris.
<ul> <li>NF ISO 11261, 1995. Soil quality - Determination of total nitrogen - Modified Kjeldahl</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-107, 2003. Soil Quality - Particle size determination by sedimentation - Pipette</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-130, 1999. Soil quality - Chemical methods - Determination of cathionic</li> <li>exchange capacity (CEC) and extractible cations. AFNOR, Association</li> <li>Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic</li> <li>oxidation. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 10390, 1994. Soil quality - Determination of pH. AFNOR, Association</li> </ul>	800	NF ISO 10693, 1995. Soil quality - Determination of carbonate content - Volumetric
<ul> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-107, 2003. Soil Quality - Particle size determination by sedimentation - Pipette</li> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-130, 1999. Soil quality - Chemical methods - Determination of cathionic</li> <li>exchange capacity (CEC) and extractible cations. AFNOR, Association</li> <li>Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic</li> <li>oxidation. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 10390, 1994. Soil quality - Determination of pH. AFNOR, Association</li> </ul>	801	method. AFNOR, Association Française de Normalisation, Paris.
<ul> <li>NF X 31-107, 2003. Soil Quality - Particle size determination by sedimentation - Pipette method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-130, 1999. Soil quality - Chemical methods - Determination of cathionic exchange capacity (CEC) and extractible cations. AFNOR, Association</li> <li>Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic oxidation. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 10390, 1994. Soil quality - Determination of pH. AFNOR, Association</li> </ul>	802	NF ISO 11261, 1995. Soil quality - Determination of total nitrogen - Modified Kjeldahl
<ul> <li>method. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF X 31-130, 1999. Soil quality - Chemical methods - Determination of cathionic</li> <li>exchange capacity (CEC) and extractible cations. AFNOR, Association</li> <li>Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic</li> <li>oxidation. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 10390, 1994. Soil quality - Determination of pH. AFNOR, Association</li> </ul>	803	method. AFNOR, Association Française de Normalisation, Paris.
<ul> <li>NF X 31-130, 1999. Soil quality - Chemical methods - Determination of cathionic</li> <li>exchange capacity (CEC) and extractible cations. AFNOR, Association</li> <li>Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic</li> <li>oxidation. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 10390, 1994. Soil quality - Determination of pH. AFNOR, Association</li> </ul>	804	NF X 31-107, 2003. Soil Quality - Particle size determination by sedimentation - Pipette
<ul> <li>exchange capacity (CEC) and extractible cations. AFNOR, Association</li> <li>Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic</li> <li>oxidation. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 10390, 1994. Soil quality - Determination of pH. AFNOR, Association</li> </ul>	805	method. AFNOR, Association Française de Normalisation, Paris.
<ul> <li>Française de Normalisation, Paris.</li> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic</li> <li>oxidation. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 10390, 1994. Soil quality - Determination of pH. AFNOR, Association</li> </ul>	806	NF X 31-130, 1999. Soil quality - Chemical methods - Determination of cathionic
<ul> <li>NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic</li> <li>oxidation. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 10390, 1994. Soil quality - Determination of pH. AFNOR, Association</li> </ul>	807	exchange capacity (CEC) and extractible cations. AFNOR, Association
<ul> <li>oxidation. AFNOR, Association Française de Normalisation, Paris.</li> <li>NF ISO 10390, 1994. Soil quality - Determination of pH. AFNOR, Association</li> </ul>	808	Française de Normalisation, Paris.
811 NF ISO 10390, 1994. Soil quality - Determination of pH. AFNOR, Association	809	NF ISO 14235, 1998. Soil quality - Determination of organic carbon by sulfochromic
	810	oxidation. AFNOR, Association Française de Normalisation, Paris.
812 Française de Normalisation, Paris.	811	NF ISO 10390, 1994. Soil quality - Determination of pH. AFNOR, Association
	812	Française de Normalisation, Paris.

813	Panfili, F.R., Manceau, A., Sarret, G., Spadini, L., Kirpichtchikova, T., Bert, V.,
814	Laboudigue, A., Marcus, M.A., Ahamdach, N., Libert, M.F., 2005. The effect of
815	phytostabilization on Zn speciation in a dredged contaminated sediment using
816	scanning electron microscopy, X-ray fluorescence, EXAFS spectroscopy, and
817	principal components analysis. Geochimica et Cosmochimica Acta 69, 2265-2284.
818	Piélou, E.C. 1966. The measures of diversity in different types of biological collections.
819	Journal of Theoretical Biology, 13, 131-144.
820	Piou, S., Bataillard, P., Laboudigue, A., Férard, J.F., Masfaraud, J.F., 2009. Changes in
821	the geochemistry and ecotoxicity of a Zn and Cd contaminated dredged
822	sediment over time after land disposal. Environmental Research 109, 712–720.
823	Ponge, J.F., 2000. Vertical distribution of Collembola (Hexapoda) and their food
824	resources in organic horizons of beech forests. Biology and Fertility of Soils, 32,
825	508-522.
826	Potapov, M., 2001. Synopses on Palaearctic Collembola. III. Isotomidae. Abhandlungen
827	und Berichte des Naturkundemuseums Görlitz 73, 1-603.
828	Prach, K., 2003. Spontaneaous succession in Central-European man-mad habitats: what
829	information can be used in restoration practice? Applied Vegetation Science 6,
830	125-129.
831	Prevost, O., 2008. Destination des sédiments fluviaux contaminés retirés des canaux:
832	optimisation des filières et possibilités de valorisation. Techniques Sciences
833	Méthodes 9, 71-79.

834	Ramsay, A.J., 1984. Extraction of bacteria from soil: efficiency of shaking or
835	ultrasonification as indicated by direct counts and autoradiography. Soil Biology
836	and Biochemistry 16, 475-481.
837	Rebele, F., Lehmann, C., 2001. Biological flora of Central Europe, Calamagrostis
838	epigejos (L.) Roth. Flora, 196, 325–344.
839	Renella, G., Mench, M., Gelsomino, A., Landi, L., Nannipieri, P., 2005. Functional
840	activity and microbial community structure in soils amended with bimetallic
841	sludges. Soil Biology and Biochemistry 37, 1498-1506.
842	Robert, M., 1996. Soil: Interface in the Environment, Resource for the Development
843	Masson, Paris (in French).
844	Ruttens, A., Mench, M., Colpaert, J., V., Boisson, J., Carleer, R., Vangronsveld, J.,
845	2006. Phytostabilization of a metal contaminated sandy soil. I. Influence of
846	compost and/or inorganic metal immobilizing soil amendments on phytotoxicity
847	and plant availability of metals. Environmental Pollution 144, 524-532.
848	Ryan, R.P., Ryan, D.J., Dowling, D.N., 2005. Multiple metal resistant transferable
849	phenotypes in bacteria as indicators of soil contamination with heavy metals.
850	Journal of Soils and Sediments 5, 95-100.
851	Sandifer, R.D., Hopkin, S.P., 1997. Effects of temperature on the relative toxicities of
852	Cd, Cu, Pb, and Zn to Folsomia candida (Collembola). Ecotoxicology and
853	Environmental Safety, 37, 125-130.
854	Shannon, C.E., Weaver, W., 1949. The Mathematical Theory of Communication.
855	University of Illinois Press, Urbana.

856	Skousen, J.G., Johnson, C.D., Garbutt, K., 1994. Natural revegetation of 15 abandoned
857	mine land sites in West Virginia. Journal of Environmental Quality 23, 1224-
858	1230.
859	Sobolev, D.,. Begonia, M., F., T., 2008. Effects of heavy metal contamination upon soil
860	microbes: Lead-induced changes in general and denitrifying microbial
861	communities as evidenced by molecular markers. International Journal of
862	Environmental Research and Public Health 5, 450-456.
863	Sørensen, T., 1948. A method of establishing groups of equal amplitude in plant
864	sociology based on similarity of species and its application to analyses of the
865	vegetation on Danish commons. Biologiske Skrifter 5, 1-34.
866	Strojan, C.L., 1978. The impact of zinc smelter emissions on forest litter arthropods.
867	Oikos 31, 41-46.
868	Tyler, G., Balsberg Pahlsson, M., Bengtsson, G., Baath, E., Tranvik, L., 1989. Heavy-
869	metal ecology of terrestrial plants, microorganisms and invertebrates – a review.
870	Water, Air and Soil Pollution 47, 189-215.
871	Taylor, J.P., Wilson, B., Mills, M.S., Burns, R.G., 2002. Comparison of microbial
872	numbers and enzymatic activities in surface soils and subsoils using various
873	techniques. Soil Biology and Biochemistry 34, 387-401.
874	Vangronsveld, J., Van Assche, F., Clijsters, H., 1995. Reclamation of a bare industrial
875	area contaminated by non-ferrous metals: in situ metal immobilization and
876	revegetation. Environmental Pollution 87, 51-59.
877	Vangronsveld, J., Colpaert, J., Van Tichelen, K., 1996. Reclamation of a bare industrial
878	area contaminated by non-ferrous metals: physico-chemical and biological

- 879 evaluation of the durability of soil treatment and revegetation. Environmental880 Pollution 94, 131-140.
- 881 Vassilev, A., Schwitzguébel, J., P., Thewys, T., van der Lelie, D., Vangronsveld, J.,
- 882 2004. The use of plants for remediation of metal-contaminated soils.
- ScientificWorld Journal 4, 9-34.
- Zimdars, B., Dunger, W., 1994. Synopses on Palaearctic Collembola. I. Tullbergiinae.
- Abhandlungen und Berichte des Naturkundemuseums Görlitz 68, 1-71.

## 887 **\_Figure captions**

888 Figure 1.

- 889 Fv/Fm ratios and antioxidant enzyme activities (μmoles/min/mg proteins excepted for
- 890 SOD expressed as U/mg proteins) in *C. epigejos* leaves collected from control area
- 891 (CA), untreated plot (NT), plots treated with hydroxylapatite (HA) or Thomas basic slag
- 892 (TBS). Different letters indicate significant differences between plots according to post-
- 893 hoc Tukey HSD tests run when ANOVA was significant.

894 Figure 2.

895 Fv/Fm ratios and antioxidant enzyme activities (µmoles/min/mg proteins excepted for

896 SOD expressed as U/mg proteins) in U. dioica leaves collected from control area (CA),

untreated plot (NT), plots treated with hydroxylapatite (HA) or Thomas basic slag

898 (TBS). Different letters indicate significant differences between plots according to post-

899 hoc Tukey HSD tests run when ANOVA was significant.

900 Figure 3.

901 Distribution of gut contents among the 403 collembolan specimens collected in three

902 treatments and in the control area (CA). Letters indicate significant differences among

903 means at 0.05 level (ANOVA followed by SNK procedure).

906 Main characteristics and (pseudo)-total metal concentrations of sediments (NT,

907 untreated; HA, hydroxylapatite; TBS, Thomas Basic Slag) and the control area (CA).

908 For trace elements (except for CA) and pH, values are means of triplicate measurements

and their standard deviation.

	CA	NT	HA	TBS
Particle size distribution (%)				
Sand	13.2	69.6	60.1	65.4
Silt	70.3	14	21.1	19.1
Clay	16.5	16.4	18.8	15.5
Organic C (%)	1.64	3.2	4.1	3.7
N (g kg <sup>-1</sup> )	1.6	1.62	1.67	2.04
CaCO <sub>3</sub> (g kg <sup>-1</sup> )	63	76	76	105
CEC (cmol kg <sup>-1</sup> )	13.3	7.2	8.3	11.3
pH <sub>water</sub>	$7.6 \pm 0.1$	$7.0\pm0,1$	$7.3\pm0,1$	7.7 ± 0,3
Zn (mg kg <sup>-1</sup> DW)	75	3250 ± 319	$2521 \pm 461$	3029 ± 217
$Cd (mg kg^{-1} DW)$	0.4	$84 \pm 2$	$67 \pm 11$	$86\pm8$
Pb (mg kg <sup>-1</sup> DW)	27	$448\pm82$	$389\pm96$	$413\pm57$
Cu (mg kg <sup>-1</sup> DW)	17	93 ± 13	$94 \pm 11$	$123\pm29$
As (mg kg <sup>-1</sup> DW)	8	53 ± 4	$48 \pm 4$	$45 \pm 2$

- 911 **Table 2**
- 912 Concentrations of trace elements extracted from soils by 0.01 M Ca(NO<sub>3</sub>)<sub>2</sub> (means of
- 913 triplicate values  $\pm$  standard deviation) and corresponding leachate ecotoxicity.
- 914 Significant differences among means at 0.05 level were indicated by different letters. nr
- 915 = not reached

	NT	HA	TBS
Zn (mg kg <sup>-1</sup> DW)	$12.05a \pm 0.96$	7.77ab ± 0.33	$2.07b\pm2.73$
Cd	$0.64a \pm 0.12$	$0.41ab \pm 0.04$	$0.12b \pm 0.13$
Pb	< 0.07	< 0.07	< 0.07
Cu	$0.12a \pm 0.01$	$0.13a \pm 0.02$	$0.21a \pm 0.14$
As	< 0.1	<0.1	< 0.1
Microtox EC <sub>50</sub> (%)	nr	nr	nr
Microtox	44	20.5	6.3
(% luminescent inhibition at 90%)			
Algae test IC <sub>50</sub> (%)	12.8	19.2	nr
Algal test	100	100	36.4
(% inhibition at 90%)			

919	Composition of the plant community in the three treatment plots. Species richness is the

920	number of species. H	is the Shannon-Weaver index.	J is the Piélou equitability index.
-----	----------------------	------------------------------	-------------------------------------

Family	Species	Plant type	Plant life	NT	HA	TBS
			cycle			
Apiaceae	Pastinaca sativa	Herbaceous	Annual	0.47	0.46	0
Asteraceae	Eupatorium cannabinum	Herbaceous	Perennial	0	3.15	0.29
	Achillea millefolium	Herbaceous	Annual	0.12	7.59	0
	Cirsium arvense	Herbaceous	Perennial	0.47	0	1.39
Brassicaceae	Cardamine hirsuta	Herbaceous	Annual	6.18	1,76	0.19
Boraginaceae	Myosotis arvense	Herbaceous	Annual	0.12	0.09	0
Convolvulaceae	Convolvulus arvensis	Herbaceous	Perennial	0	0.46	0
Cyperaceae	Carex hirta	Herbaceous	Perennial	11.55	1.85	0.33
	Carex cuprina	Herbaceous	Perennial	0.12	0.09	0
Lamiaceae	Glechoma hederacea	Herbaceous	Perennial	11.67	22.85	7.08
	Lycopus europaeus	Herbaceous	Perennial	0.70	1.57	0
Onagraceae	Epilobium angustifolium	Herbaceous	Annual	0.35	1.2	11.76
Poaceae	Deschampsia cespitosa	Herbaceous	Perennial	1.52	1.76	0
	Festuca rubra	Herbaceous	Perennial	3.73	6.85	0
	Phalaris arundinacea	Herbaceous	Perennial	0	0.93	33.09

	Calamagrostis epigejos	Herbaceous	Perennial	46.91	19.98	30.0
	Holcus lanatus	Herbaceous	Perennial	0	2.22	0.1
	Poa trivialis	Herbaceous	Perennial	0	0	0.14
Ranunculaceae	Ranunculus repens	Herbaceous	Perennial	0.12	1.11	0
Rubiaceae	Galium aparine	Herbaceous	Annual	2.22	2.13	16.4
Scrophulariaceae	Linaria vulgaris	Herbaceous	Perennial	0	1.67	0
Urticaceae	Urtica dioica	Herbaceous	Perennial	2.33	4.63	0.8
Betulaceae	Betula pendula	Tree	Perennial	8.05	12.67	0.0
	Betula pubescens	Tree	Perennial	1.87	2.31	0.0
Fagaceae	Quercus robur	Tree	Perennial	0.12	0.28	0
Salicaceae	Salix sp.	Tree	Perennial	0	0.46	0
	Salix alba	Tree	Perennial	0.12	1.02	0
	Salix capraea	Tree	Perennial	1.40	0.93	0
Species richness H' J				21 2.7 0.62	26 3.6 0.77	14 2.35 0.62

surface.

	CA	NT	HA	TBS
Calar	nagrostis epigejos			
Cd	< 0.05	2.16 (a) ± 1.4	2.51 (a) ± 1.1	$0.34$ (b) $\pm 0.18$
Zn	23 (a) ± 9	121 (c) ± 58	98 (bc) $\pm$ 32.6	44 (ab) ± 20
Pb	<1	<1	<1	<1
Cu	4.5 (ab) ± 0,9	$3.5(a) \pm 0.3$	$5.1 (b) \pm 0.7$	$4.3 (ab) \pm 0.2$
As	< 0.5	< 0.5	< 0.5	< 0.5
Ca	$405 \pm 331$	$617 \pm 115$	$769\pm273$	$791 \pm 243$
Fe	$23 \pm 2$	$52 \pm 56$	$33 \pm 5$	$23 \pm 1$
Mg	453 (b) ± 121	251 (a) ± 83	532 (b)± 147	446 (ab) ± 62
Mn	16 (a) ± 5	51 (b) ± 8	13 (a) ± 3	$15(a) \pm 1$
Р	$1570\pm247$	$1530\pm189$	$1863\pm875$	$1175\pm235$
Na	123 (ab) ± 4	115 (a) ± 4	$131 (b) \pm 12$	119 (ab) ± 13
Κ	12407 (a) ± 1329	14523 (ab) ± 1543	16648 (b) ± 2620	13074 (a) ± 1288
Al	n.d	5 (a) ± 2	8.5 (b) ± 1.6	24 (c) ± 1
Urtice	a dioica			
Cd	$0.06(a) \pm 0.01$	$1.5 (b) \pm 1.3$	$0.9 (ab) \pm 0.3$	$0.36 (ab) \pm 0.18$
Zn	12 (a) ± 1	$210 (b) \pm 104$	$184 (b) \pm 28$	145 (b) ± 21
Pb	<1	<1	<1	<1
Cu	$4.0(a) \pm 0.5$	$6.9(b) \pm 1.9$	6.8 (b) ± 1.9	$7.4 (b) \pm 0.9$
As	< 0.5	< 0.5	< 0.5	< 0.5
Ca	$19719\pm8227$	$20945 \pm 3292$	$24398 \pm 2407$	$23901 \pm 3334$
Fe	35 (a) ± 6	51 (ab) ± 10	60 (b) ± 12	43 (ab) ± 11
Mg	$1726 \pm 1753$	$1278\pm433$	$2176\pm285$	$1722 \pm 241$
Mn	$13 \pm 3$	$12 \pm 6$	$11 \pm 2$	$12 \pm 2$
Р	2314 (a) ± 267	4229 (b) ± 465	4310 (b) ± 841	$6096(c) \pm 563$
Na	$136\pm38$	$177 \pm 41$	$162 \pm 20$	$149 \pm 12$
K	17355 (a) ± 4593	19029 (ab) ± 1434	23699 (bc) ±	27963 (c) ± 2117
			1721	
Al	n.d	$20 \pm 8$	$32 \pm 10$	$28 \pm 12$

925 Concentrations (mg/kg DW) in shoots of *Calamagrostis epigejos* and *Urtica dioica*.

926 Mean values and standard deviations (n=5; mg kg<sup>-1</sup> DW). Different letters indicate

927 significant differences among plots for $\alpha = 0.05$	5.
---	----

	SOD	CAT	APX	GPX	GRD
Cd	-0.68**	-0.67**	-0.79***	-0.61**	-0.58**
Zn	0.31ns	0.35ns	0.56**	0.01ns	0.52*
Cu	-0.27ns	-0.48*	-0.38ns	0.00ns	-0.56*
Mn	0.55*	0.60**	0.53*	-0.11ns	0.80***
Р	0.09ns	-0.02ns	-0.18ns	0.13ns	-0.23ns
Ca	0.37ns	0.18ns	0.38ns	0.44ns	0.08ns
K	0.06ns	-0.01ns	0.22ns	-0.03ns	0.14ns
Mg	-0.30ns	-0.43ns	-0.52*	-0.32ns	-0.56*
Na	-0.33ns	-0.54*	-0.19ns	-0.21ns	-0.50*
U. dioica					
	SOD	CAT	APX	GPX	GRD
Cd	0.22ns	0.24ns	0.30ns	0.32ns	0.11ns
Zn	0.48*	0.47*	0.56**	0.61**	0.45*
Cu	0.51*	0.49*	0.56*	0.60**	0.70**
Mn	-0.07ns	-0.02ns	-0.08ns	-0.04ns	0.08ns
Р	0.79***	0.74***	0.79***	0.85***	0.82***
Ca	0.10ns	0.12ns	0.18ns	0.21ns	0.26ns
Κ	0.54*	0.45*	0.48*	0.56*	0.60**
Mg	-0.20ns	-0.21ns	-0.10ns	-0.07ns	-0.11ns
Mg Na ns: not signit	-0.20ns 0.22ns ficant, ***,**,*,	-0.21ns 0.24ns	0.28ns	0.22ns	0.02ns
Mg Na	-0.20ns 0.22ns ficant, ***,**,*,	-0.21ns 0.24ns	0.28ns	0.22ns	0.02ns
Mg Na ns: not signit	-0.20ns 0.22ns ficant, ***,**,*,	-0.21ns 0.24ns	0.28ns	0.22ns	0.02ns
Mg Na ns: not signit	-0.20ns 0.22ns ficant, ***,**,*,	-0.21ns 0.24ns	0.28ns	0.22ns	0.02ns
Mg Na ns: not signit	-0.20ns 0.22ns ficant, ***,**,*,	-0.21ns 0.24ns	0.28ns	0.22ns	0.02ns
Mg Na ns: not signit	-0.20ns 0.22ns ficant, ***,**,*,	-0.21ns 0.24ns	0.28ns	0.22ns	0.02ns
Mg Na ns: not signit	-0.20ns 0.22ns ficant, ***,**,*,	-0.21ns 0.24ns	0.28ns	0.22ns	0.02ns
Mg Na ns: not signit	-0.20ns 0.22ns ficant, ***,**,*,	-0.21ns 0.24ns	0.28ns	0.22ns	0.02ns
Mg Na ns: not signit	-0.20ns 0.22ns ficant, ***,**,*,	-0.21ns 0.24ns	0.28ns	0.22ns	0.02ns

**Table 5.** Correlation analysis between shoot element concentrations and antioxidant
enzyme activities in leaves (n=20).

Table 6. Species distribution and biodiversity indices of collembolan species.

ANOVA: NS = not significant; \*\* P < 0.01. Post-hoc tests (SNK procedure):

significant differences indicated by different letters

	CA	NT	HA	TBS	F value
Dicyrtomina minuta	0	0	0	0.33±0.27	1 NS
Entomobrya sp.	0	$1.33 \pm 1.09$	0	0	1 NS
Folsomia candida	0.33±0.27	$0.33 \pm 0.27$	0	$1\pm0.82$	0.58 NS
Friesea truncata	$0^{\mathrm{b}}$	$0^{b}$	$0^{\mathrm{b}}$	$4\pm1.25^{a}$	6.86**
Hererosminthurus claviger	0.33±0.27	0	0	0	1 NS
Isotomodes productus	$0.67 \pm 0.54$	0	0	2.67±1.78	1.22 NS
Lepidocyrtus lignorum	0	0	0	0.33±0.27	1 NS
Mesaphorura florae	1.33±0.72	6.33±2.13	21.67±7.92	25.33±8.57	2.56 NS
Mesaphorura macrochaeta	3.33±2.72	0	$0.67 \pm 0.27$	$0.67 \pm 0.54$	0.75 NS
Mesaphorura pongei	0	0	0	1.33±1.09	1 NS
Orchesella sp.	0	$0.33 \pm 0.27$	0.33±0.27	0	0.67 NS
Parisotoma notabilis	6.67±1.96	4.67±1.66	5.67±2.13	0.33±0.27	1.85 NS
Proisotoma minima	7±2.45	9±2.94	$0.67 \pm 0.54$	7±2.36	1.71 NS
Tomocerus vulgaris	0	0	0.33±0.27	0	1 NS
Total abundance	19.67±3.07	22±5.44	29.33±6.4	43±10.6	1.53 NS
Species richness	$4\pm0.47$	4	3.67±0.27	5.67±1.19	1.28 NS
Shannon Index	$1.44 \pm 0.21$	$1.65 \pm 0.07$	$1.09\pm0.32$	$1.78 \pm 0.26$	1.09 NS
Equitability Index	$0.72 \pm 0.04$	$0.82 \pm 0.04$	0.56±0.15	0.75±0.03	1.25 NS

943

## 946 **Table 7. Total and specific bacterial numbers**

947 Counts of total bacteria, acidophilic sulphur-oxidising bacteria (ASOB), neutrophilic 948 sulphur-oxidising bacteria (NSOB) and sulphate-reducing bacteria (SRB) (expressed as 949 bacteria  $g^{-1}$  dry soil) collected from untreated plot (NT), plot treated with 950 hydroxylapatite (HA) or Thomas basic slag (TBS). Values are expressed as means ± 951 standard deviation in triplicate samples. Values with the same letters are not 952 significantly different according to ANOVA used Newman-Keuls (SNK) test at P ≤ 953 0.05.

	NT	HA	TBS	
Total bacteria	2.12 10 <sup>7</sup>	5.58 10 <sup>7</sup>	1.75 10 <sup>8</sup>	
	$\pm 5.09 \ 10^5 \ c$	$\pm 7.90 \ 10^{6} \ b$	$\pm 1.8610^{6}$ a	
ASOB	$3.69 \ 10^3$	$7.36\ 10^3$	$2.93 \ 10^3$	
	$\pm 2.31 \ 10^1 \ a$	$\pm 3.19 \ 10^3 \ a$	$\pm 1.29 \ 10^3 \ a$	
NSOB	$1.00 \ 10^5$	$3.05 \ 10^5$	$1.45 \ 10^5$	
	$\pm 6.78 \ 10^4 \ a$	$\pm 1.15 \ 10^5 \ a$	$\pm 4.62 \ 10^4 \ a$	
SRB	$6.97 \ 10^1$	$5.97 \ 10^1$	$5.97 \ 10^{1}$	
	$\pm 4.04 \ 10^1 \ a$	$\pm 2.8910^{1}$ a	$\pm 2.89 \ 10^1 \ a$	

954

957	Treatments (NT = Untreated, HA = Hydroxyapatite, TBS = Thomas Basic Slag) ranked
958	according to 24 variables describing optimal features of plant, faunal and microbial
959	communities and sediment toxicity. Average ranks are indicated by mean $\pm$ standard
960	deviation.

Variable	NT	HA	TBS			
Plant, faunal and microbial communities good						
health						
Vegetation succession state	2	1	3			
Plant species richness	2	1	3			
Plant species diversity	2	1	3			
Plant species equitability	2.5	1	2.5			
Fv/Fm in U. dioica	2	1	3			
Collembola abundance	3	2	1			
Collembola species richness	2	3	1			
Collembola species diversity	2	3	1			
Collembola species equitability	1	3	2			
Total bacteria abundance	3	2	1			
Sediment toxicity						
Cd mobility	3	2	1			
Zn mobility	3	2	1			
Microtox	3	2	1			
Algal test	2.5	2.5	1			
Cd in <i>C. epigejos</i>	2	3	1			
Zn in <i>C. epigejos</i>	3	2	1			
Cd in Urtica	3	2	1			
Zn in Urtica	3	2	1			
SOD in <i>C. epigejos</i>	3	1	2			
CAT in <i>C. epigejos</i>	3	1	2			
APX in <i>C. epigejos</i>	3	1	2			
SOD in <i>U. dioica</i>	2	1	3			
CAT in U. dioica	2	1	3			
APX in U. dioica	2	1	3			
Average rank	2.46±0.57	1.73±0.76	1.81±0.9			





